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Article

# Assessment of Biodegradable Films as Protective Barriers Toward Sustainable Protection of Coastal Archaeological Sites

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## Abstract

Saltwater Intrusion (SWI) is threatening coastal archaeological sites, particularly in Crotona, southern Italy. The study area has been experiencing notable SWI due to over-pumping of groundwater, rising land subsidence, and climate change. Consequently, this study examines the applicability of polycaprolactone (PCL), commonly a biodegradable polymer, as a protective barrier for archaeological conservation. PCL films were synthesized via solvent casting and dried under controlled conditions. Physicochemical properties of the films were evaluated using six analytical techniques: (1) contact angle measurements for surface hydrophobicity, (2) Fourier-Transform Infrared Spectroscopy (FTIR) for chemical stability, (3) Scanning Electron Microscopy (SEM) for morphological characterization, (4) permeability testing for evaluating saltwater diffusion, (5) mechanical testing for tensile properties, and (6) biodegradability assays for degradation rates. All samples were evaluated at 0, 30, 60, and 90 days in natural seawater. Results from these tests indicate that unmodified PCL films exhibit moderate hydrophobicity, low chemical stability, moderate permeability, declining mechanical strength, and limited biodegradability over the testing period. Therefore, their suitability as long-term protective barriers requires further optimization. Nevertheless, material enhancement using nanoparticles or surface functionalization is highly recommended to improve hydrophobicity, enhance surface stability, reduce permeability, decrease biodegradability, and increase durability in saline environments.

**Keywords:** Saltwater Intrusion (SWI); polycaprolactone (PCL); biodegradable polymer; saline conditions; coastal archaeological sites

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## 1. Introduction

Coastal archaeological sites are consistently experiencing impacts from many probable threats: erosion, rising sea level, groundwater pollution, salinization, and climate change. The action underlain by the increased human pressures related to urbanization, agriculture, and development contributes to the existing impacts on coastal archaeological heritage.

It is noted that the expected amount of at-risk coastal archaeological sites will likely increase substantially by the years 2050 and 2100 if greenhouse gas emissions choose to remain high [1,2,3]. As a result of natural or human-induced erosion, archaeological features have already been lost in areas such as Libya, New Zealand, and the Mediterranean [4,5]. The implications of saltwater intrusion and agricultural or urban contamination are also contributing factors for damaging subsurface remains in places such as Florida and Italy [6,7]. Moreover, the growth of urban centers, as well as poor land management and unsustainable agricultural practices, are compounding the risk

factors noted previously, along with the increased extraction of potable groundwater, land subsidence, and the destruction of heritage sites [3,7,8,9]. Submerged and buried archaeological sites (as in the current study area of Crotona) can be especially vulnerable to as the preservation of deposits largely depends on the stability of their depositional environment [10,11]. For example, the ancient Greek temple of Hera Lacinia at Capo Colonna (near Crotona) may ultimately be impacted by its proximity to the coast, and the current threat of SWI which could endanger the integrity of the site.

The traditional protective materials used in archaeological sites such as geomembranes and concrete are non-biodegradable materials and would frequently conflict with ethical conservation considerations because of the environmental damage incurred and the need for manual extraction in a non-destructive manner. To address these issues, researchers have begun investigating biodegradable, eco-compatible polymers that can perform protective functions and promote environmental responsibility [12,13].

Polycaprolactone (PCL) is emerging as a promising candidate which is semi-crystalline, hydrophobic, and processable into thin films with relatively low water sorption [14]. On the other hand, PCL exhibits greater durability compared to other biodegradable polymers such as Polylactic Acid (PLA) and Polybutylene Succinate (PBS) [15,16].

The barrier qualities of the biodegradable polymers depend on the ambient conditions. For instance, extreme salinity will likely increase hydrolysis and mass loss, which could compromise the protective qualities of the biodegradable polymers [17]. Therefore, it is vital to examine the long-term survivability of biodegradable polymers in adverse saline, sub-surface conditions going forward [15]. Functional additives could reduce the negative impact of salinity and improve the durability and effectiveness of these polymers. Addressing these gaps is crucial for the adoption of the protective barriers in heritage conservation and for advancing global sustainability goals.

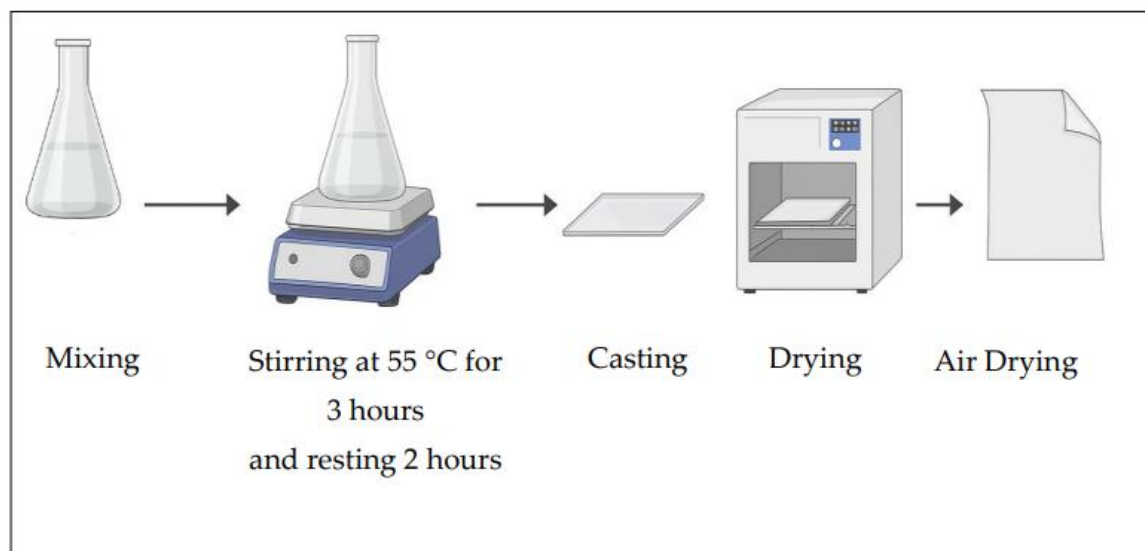
## 2. Materials and Methods

### 2.1. Materials

Polycaprolactone (PCL) with an average molecular weight ( $M_n \approx 80,000$  g/mol) was acquired from Sigma-Aldrich without further purification. Tri-ethyl phosphate (TEP) was selected as a green solvent which supports our sustainable goals. Seawater samples from the Ionian Sea from the study location (Crotona city) were collected and analysed to confirm usability of experiments.

### 2.2. Film Preparation

PCL films were made with TEP as the green solvent as shown in **(Figure 1)**. 7 g of TEP was added to a glass flask, followed by 1 g of PCL which was added in three equal portions of 0.33 g each. Each portion was dissolved completely before the next portion was added. Once mixed, the TEP/PCL was stirring at 55 °C for 3 hrs to produce a uniform solution. The solution was left to rest for 2 hours before pouring the solution onto a flat glass plate. The films were placed in an oven at 55 °C to evaporate the solvent. Once the solvent evaporated, films were detached from the glass by immersing them in deionized water until the films were released from the glass and were then air-dried overnight at room temperature for testing.



**Figure 1.** Preparation of PCL films.

The films were cold cut in to strips of 3cm × 3cm for weight loss experiment, and strips of 10cm × 2cm for mechanical strength tests. The average thickness of the films measured between 0.10 mm to 0.22 mm using a digital micrometer. All tests were performed in triplicate.

### 2.3. Ionian Seawater Immersion

For simulating condition burial of coastal artifacts, PCL films were placed in glass containers with natural seawater. Ionian seawater was used because of the similar saline conditions in the other coastal archaeological site of Crotona and similar effects to the seawater intrusion. The containers were placed in an environmental chamber at 15 °C which was selected to simulate coastal artifacts that would typically be buried beneath the earth especially in cooler stable environmental conditions. The measured temperature will provide for stabilized laboratory observations but also keep in mind the conditions with static immersion, and some water dynamic water moving effect conditions in the field of real environments.

### 2.4. Surface Hydrophobicity: Contact Angle Measurements

To determine the wettability of the surface, a goniometer was used, and the sessile drop method was completed. Each film surface had a droplet of two microliters of deionized water deposited, and the contact angle was measured after waiting 20 seconds. Each sample was measured three times, and the average value was calculated. Any changes in contact angle allows for interpretation of increased risk of deterioration of artifacts. A hydrophilic surface will contribute to less-desirable interactions of water with cultural heritage materials.

### 2.5. Fourier-Transform Infrared Spectroscopy (FTIR)

The chemical structural changes were followed by attenuated total reflectance Fourier-transform infrared (ATR-FTIR) spectroscopy (4000 - 500  $\text{cm}^{-1}$ , 4  $\text{cm}^{-1}$ ). All spectra are the means of thirty-two scans. The observation of a diminishment in the absorption peaks related to ester carbonyl (approximately 1720  $\text{cm}^{-1}$ ) and new peaks related to hydroxyl (approximately 3300  $\text{cm}^{-1}$ ) or the arousal of new peaks for carboxylate groups (approximately 1600  $\text{cm}^{-1}$ ) can be attributed to hydrolytic degradation. FTIR data assists to demonstrate the chemical instability risk and impacts on the stability of the artifact.

### 2.6. Scanning Electron Microscopy (SEM)

The morphologies of the films were assessed at 0, 30, 60, and 90 days using a ZEISS Scanning Electron Microscope. All the samples were sputter coated in gold to improve conductivity of the samples. Micrographs taken at 500× to 2000× magnification were documented to record surface roughness, pitting, and cracking during the degradation of the surface films. The PM was analysed via SEM to determine how the microstructure impacted any physical vulnerabilities that might jeopardize archaeological artifacts.

### 2.7. Seawater Permeability Testing

Seawater permeability tests are considerable methods of evaluating the performance of films to inhibit saline infiltration, which ultimately affects the amount of protection offered to the artifacts. Water permeation tests on the films used in the study were conducted using a dead-end filtration cell (UHP-25, Strelitech, Japan) with a feed pressure of 0.5 bar and medium speed of magnetic stirrer (190m) at room temperature. As time went on, mass loss was recorded as the system was stirred at continuous periods. Conductivity was also measured in the receiving chamber over a 6-hour period. The higher the permeability the greater the likelihood of salt damage occurring in the artifacts.

$$J = \frac{Q}{A.t} \quad (1)$$

Where: J: Permeate flux (L/m<sup>2</sup>.h)

Q: Volume of permeate collected (L)

A: Membrane area (m<sup>2</sup>)

t: Time (hr)

$$P = \frac{J}{\Delta P} \quad (2)$$

Where: P: Permeability (L/m<sup>2</sup>.h.bar)

J: Permeate flux (L/m<sup>2</sup>.h)

ΔP: Applied pressure difference (bar)

### 2.8. Mechanical Strength Testing

The mechanical integrity of samples was conducted using a Zwick/Roell Z2.5 testing unit (BTC-FR2.5TN-D09, Zwick Roell Group, Ulm, Germany). Film strips were allowed to stretch, and at a constant rate of elongation (5 mm/min), were tested to rupture. The values of tensile strength (MPa), Young's modulus (MPa), and elongation at break (%) were produced, which could indicate long-term structural integrity of the polymer. An increase in loss of those mechanical properties could increase the susceptibility of the archaeological object to physical stress.

### 2.9. Biodegradability Assessment

Biodegradation evaluation was completed by tracking the loss of mass of samples (using the sample's dry mass) after submersion in Crotonese seawater. After weighing the samples, they were submerged in the seawater for an assigned time, rinsed with freshwater, dried at ambient temperature, and weighed again. The mass loss was finally estimated as a percentage to approximate the percent of hydrolytic biodegradation during the treatment. Changes to the sample colour also provide observable qualitative data. The conceptualizations of biodegradability as a property guarantee that the selected protective materials would not produce excessive long-term detriment to the environment while preserving archaeological integrity through time.

$$\text{Weight loss (\%)} = \frac{W_0 - W_t}{W_0} \times 100 \quad (3)$$

Where: W<sub>0</sub>: the initial weight of the sample before biodegradation (g)

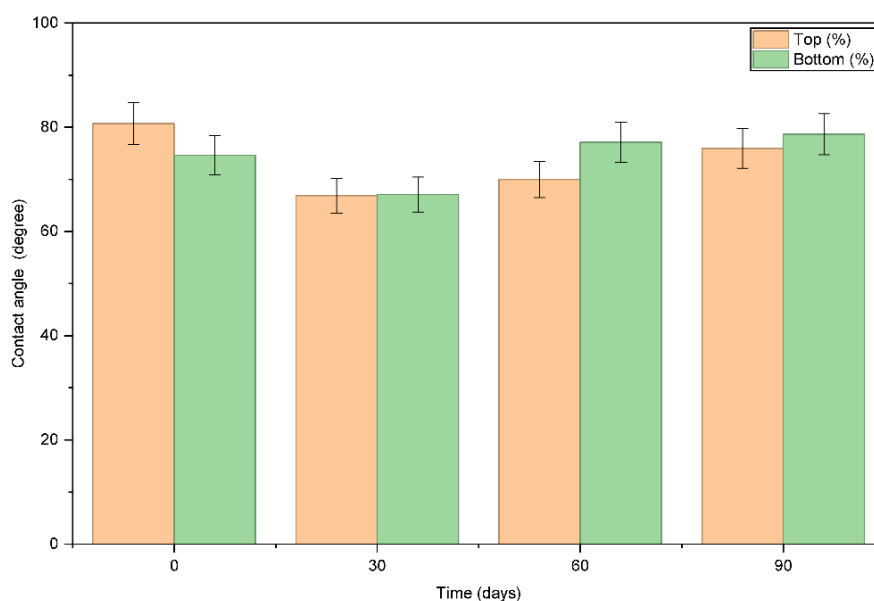
W<sub>t</sub>: the weight of the sample after biodegradation (g)

### 3. Results and Discussion

The degradation behavior of unmodified polycaprolactone (PCL) films sub-merged in actual seawater for 90 days was described utilizing different analytical techniques to determine changes in physical, chemical, mechanical and permeability. The following section describes the changes observed in surface hydrophobicity, chemical structure, micro-morphology, saltwater permeability, mechanical strength, and total mass loss.

#### 3.1. Surface Hydrophobicity

Contact angle analysis was achieved for both upper and lower surfaces of PCL films over the 90-day simulated exposure period to the salinity of seawater, demonstrating in **(Figure 2)**. Zero day had average contact angles of 80.69° (top) and 74.64° (bottom), denoting moderate hydrophobicity. At day 30, contact angles decreased to 66.86° (top) and 67.10° (bottom) indicating increased surface wettability due to the interaction with saline. At day 60, the upper surface partially recovered to 69.99°, while the lower surface increased to 87.11°, indicating partial surface reorganization and uneven salt deposition between the two surfaces. An increase in surface polarity will elevate the affinity for water for the material and could catalyze degradation behavior. Day 90 was stable at 75.96° (top) and 78.67° (bottom) lined in the table below, indicating partial retention of hydrophobicity, while some altered surface dynamics persist.



**Figure 2.** The water contact angle of pure PCL films measured on top and bottom surfaces after immersion in natural seawater (15 °C) for 0, 30, 60, and 90 days.

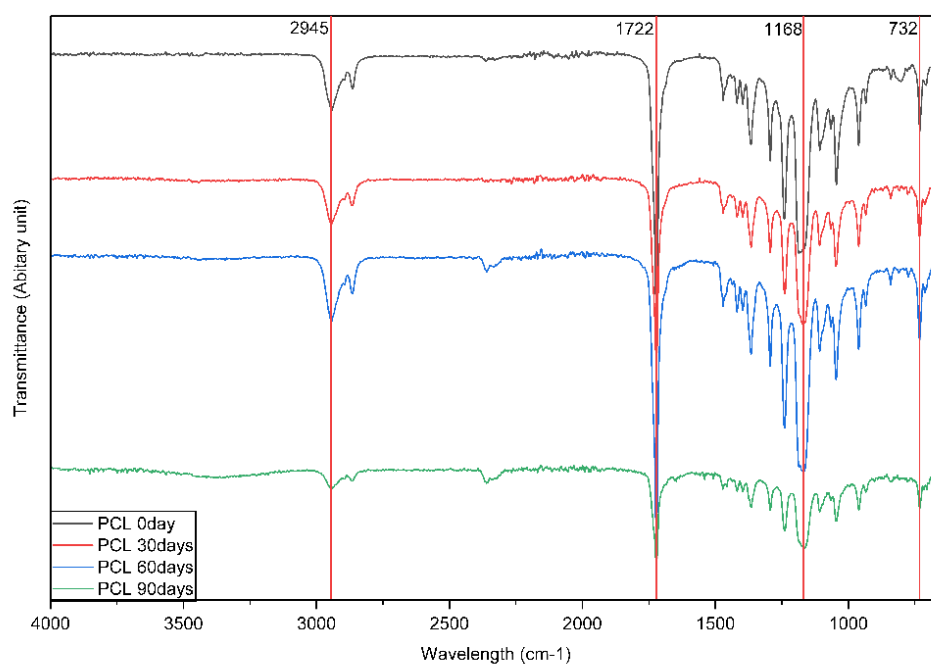
#### 3.2. FTIR Spectroscopy

Fourier Transform Infrared (FTIR) spectroscopy was performed to confirm the chemical structure and functional groups of the synthesized polycaprolactone (PCL) through the evaluation of the PCL films at 0, 30, 60 and 90 days in natural saline as shown in **(Figure 3)**. FTIR spectroscopy was able to demonstrate PCL specific absorption bands including CH<sub>2</sub> asymmetric stretching (2946 cm<sup>-1</sup>), a strong ester carbonyl (C=O) absorption band (1718 cm<sup>-1</sup>), C–O–C stretching (1169 cm<sup>-1</sup>), and CH<sub>2</sub> rocking (730 cm<sup>-1</sup>) over time. Although the absorption bands displayed in the FTIR spectra were relatively consistent over time, significantly lower absorbance intensity was noted over the same 4 time points, especially at 1718 cm<sup>-1</sup> and 1169 cm<sup>-1</sup>. The change in intensity indicated decreasing

chemical stability due to hydrolytic degradation of ester groups over time. No new absorption bands were produced beyond indicating the loss of chemical stability and change in structural integrity of the polymer backbone over time. The spectral analysis of PCL highlights the degradation in intensity following immersion at 30 days for carbonyl ( $1722\text{ cm}^{-1}$ ) and C–O–C ( $1168\text{ cm}^{-1}$ ) as time progressed to 60 and 90 days, respectively, confirming that the entire polymer suffered some extent from hydrolytic degradation of which has previously been reported necessary for the biodegradability of PCL. The absorption bands that would have normally confirmed the establishment of secondary functional groups were not demonstrated in the FTIR measurements following the application of the saline exposure.

The C=O absorption peak at around  $1725\text{ cm}^{-1}$  is often discussed as the main indicator for the presence PCL [18]. Additionally, the aliphatic C-H stretching vibrations identified in the range of  $2945\text{ cm}^{-1}$  and  $2865\text{ cm}^{-1}$  have also been shown to be indicative of PCL's presence in degraded composite materials [19]. Moreover, the peaks associated with C–O–C and C–C stretching vibrations at  $1240\text{ cm}^{-1}$  and  $1295\text{ cm}^{-1}$  have been similarly documented, verifying additional impact concerning the crystalline nature of the polymer [20].

Overall, the measured FTIR values verified continuous degradation of the chemical stability of PCL over 90 days but no alteration in its structural integrity and chemical stability was noted throughout the measurement periods.

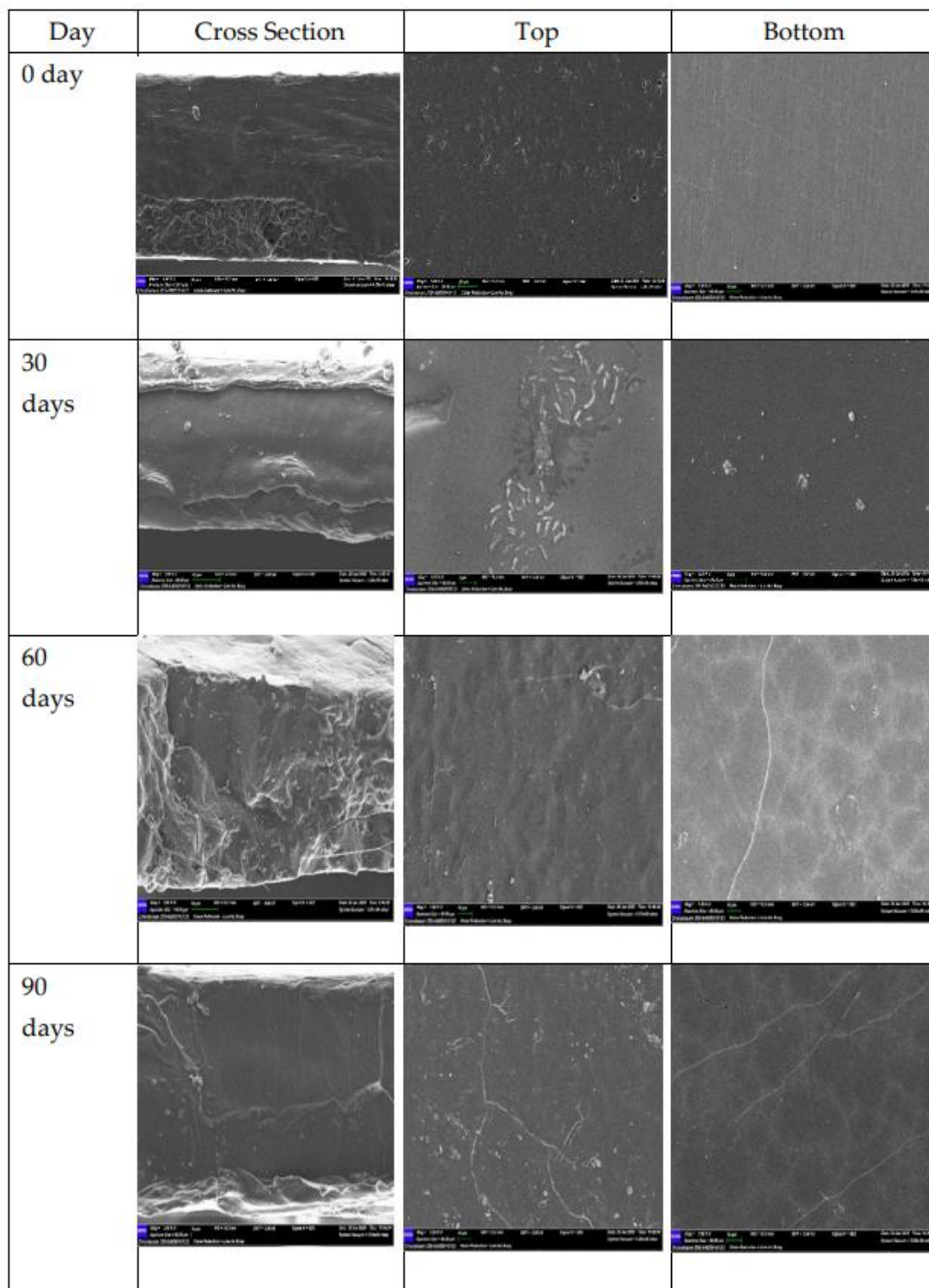


**Figure 3.** FTIR spectra of PCL films undergoing immersion at natural seawater ( $15\text{ }^{\circ}\text{C}$ ) at 0, 30, 60, and 90 days, indicating variations in peak heights and symmetry indicated continuous hydrolytic degradation of the polymer structure.

### 3.3. SEM Morphology

SEM images of the top, bottom, and cross-sections of PCL films (degradation time point of 0, 30, 60 days, and 90 days) are presented in (Figure 4). In the beginning, the surfaces of the films were smooth, homogeneously formed, and free of cracks or pores, both trimmed, and in cross-section appearing compact and uniform. After 30 days the cross-section began to show some roughness and slight delamination, while the top surface showed a few zealous aggregates and irregularities. The bottom surface exhibited small appearing scattered deposits which were likely due to salt crystallization. By 60 days, these features would become more pronounced. By 90 days the cross-

section exhibited large cracks and layered structures, indicating that the internal structural integrity was breaking down. These changes in morphology occurred simultaneously to the changes in hydrophobicity and chemical stability, demonstrating how length of time in saline environments negatively impacts the barrier property of unmodified PCL film.



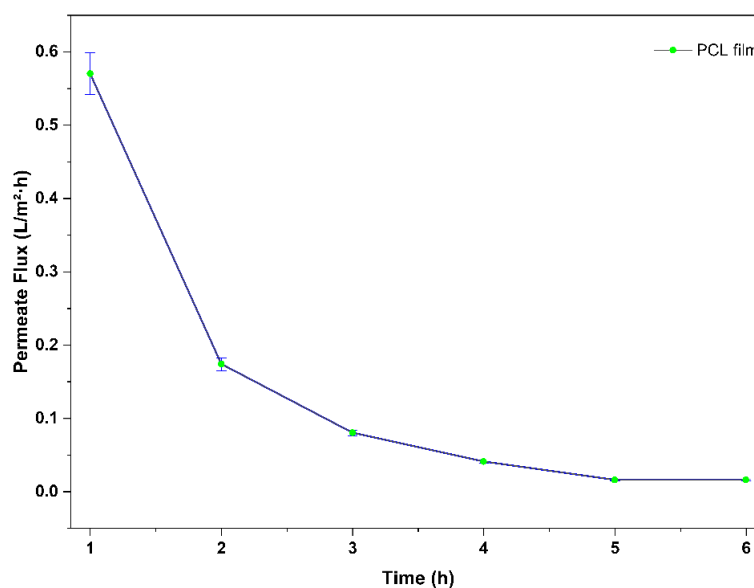
**Figure 4.** Representative SEM micrographs of PCL films testing in the natural seawater at 0, 30, 60, and 90 days with a temperature of 15 degrees.

### 3.4. Seawater Permeability Test

The water permeation performance of the PCL films was assessed with a dead-end filtration cell at a constant feeding pressure of 0.5 bar and room temperature. The permeate flux (J) was determined by measuring mass change over time, and the average data can be seen in (Figure 5) with a dramatic loss in flux over the 6-hour test. The initial flux was clocked at 0.57026 L/m<sup>2</sup>·h at 1 hour, then dropped dramatically to 0.17379 L/m<sup>2</sup>·h at 2 hours and continued to tail down to 0.01584 L/m<sup>2</sup>·h by 5 hours, stabilizing thee after at 6 hours.

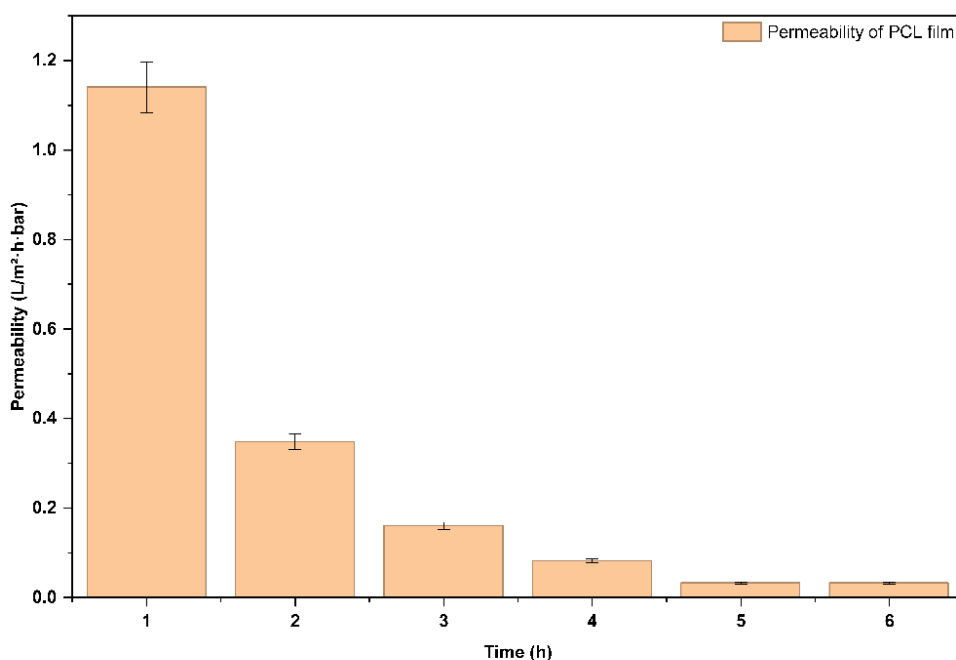
This trend is consistent with the behaviour of hydrophobic membranes in dead-end filtration systems, where fouling due to the accumulation of material on the membrane surface is a common challenge.

This pattern is consistent with typical behaviour of hydrophobic membranes in dead-end filtration systems, where fouling due to the accumulation of material on the membrane surface is a common challenge. For example, Wang et al., (2024), studied PCL membranes produced through electrospinning, in which hydrophobic membranes accrete and eventually block pores under dead-end pressure quickly, similar to the rapid loss of flux described in our work, as demonstrated in reference [21]. Similarly, Jo et al., (2024) observed similar initial fluxes to the ones observed in our study, plus also films produced with PCL decreases after time [22]. It is found that stable flux beyond the 5 hours time point demonstrates that there is a steady state whereby either incremental fouling or structural changes are negligible or not recognizable.



**Figure 5.** Permeate flux of pure PCL film as a function of filtration time.

Moreover, natural Seawater Permeability (P) starting at 1.14052 L/m<sup>2</sup>·h·bar at 1 hour and declining to 0.03168 L/m<sup>2</sup>·h·bar by 5 hours, remaining constant at 6 hours as shown in (Figure 6). This decline in both flux and permeability advocates a reduction in the film's water transport efficiency over time, due to fouling or compaction of the PCL film under the applied pressure. This support is also confirmed by Zhang et al., (2024), where the authors conclude that stable plateau region in permeability through ultrafiltration means that the cake layer is saturated[23].



**Figure 6.** Seawater permeability of PCL films measured at different filtration times, indicating pore blocking and fouling effects during prolonged exposure.

The initial elevated permeate flux and permeability demonstrate that the PCL film has reasonable water permeability for the beginning filtration period, due to its hydrophobicity and porous structure. The consistent permeate flux and permeability beyond 5 hours (0.01584 L/m<sup>2</sup>·h and 0.03168 L/m<sup>2</sup>·h·bar, respectively) indicates that a stable condition had occurred and thus further fouling or structure change was limited. These results capture the potential of the PCL film but also highlight the limitation of the PCL for extended filtration without antifouling procedures, which could include surface modification or periodic cleaning during filtration. Future work will also include studying using higher pressures or using crossflow filtration to improve PCL performance.

### 3.5. Mechanical Properties

The mechanical outcomes indicate an ongoing deterioration of both stiffness and tensile strength of PCL films over time in the natural seawater. Tensile properties of PCL films were determined after 0, 30, 60 and 90 days of immersion in natural seawater and summarized in **(Table 1)**. Initially, PCL films had a Young's modulus of 239.67 MPa, tensile strength of 15.3 MPa and elongation at break of 11.67% which demonstrate satisfactory mechanical integrity. After 30 days of immersion in saline (30 days), the Young's modulus dropped to 180.90 MPa, tensile strength dropped to 9.33 MPa, and elongation at break slightly decreased to 10.30%, all indices of mechanical integrity. At 60 days, Young's modulus dropped to 173.00 MPa, tensile strength dropped to 4.5 MPa and elongation at break dropped to 7.80%. At 90 days, Young's modulus lowered again to 145.00 MPa, tensile strength partially recovered to 9.91 MPa and elongation at break increased to 11.00%

**Table 1.** Mechanical properties of pure PCL films in natural seawater at different immersion times with constant temperature of 15 °C.

Film names	Time (days)	Average Young's modulus (MPa)	Average tensile strength (MPa)	Average Elongation break (%)
PCL films	0	239 ± 67	15 ± 30	11 ± 67
	30	180 ± 90	9 ± 33	10 ± 30
	60	173 ± 02	4 ± 50	7 ± 80

90

145 ± 03

9 ± 91

11 ± 01

The decline in tensile strength from 15.3 MPa (0 d) to 4.5 MPa over the 60 d indicates the breaking down of the polymer chains in saline conditions. The slight recovery in tensile strength to 9.91 MPa on day 90 may be attributed to structural reorganization or a change in crystallinity due to some of the degraded polymer chains being realigned. The indicator of the elongation at break dropped to (7.8%) 60 days, suggesting increased brittleness. Nonetheless, the high increase in tensile strength (11.0%) at 90 days may indicate plasticization effects due to water uptake that can improve ductility even if a degree of chemical degradation is occurring. Similar studies conducted by H. Tsuji (2002) indicated that when PCL is exposed to hydrolytic (aqueous) conditions, the hydrolysis of the amorphous portion of the polymer dominated how the overall properties change [24]. Further, for the seawater-degradable PCL films, G.X. Wang et al. (2021) provided evidence for a decline in mechanical strength due to time under immersion conditions and followed a parallel decline with both magnitudes and trends of mechanical strength observed [25]. Similarly, the research conducted by Lyu et al. (2019) demonstrated a comparable decline in mechanical properties whereby days post-immersion resulted in notable declines in the tensile strength of the PCL films test [26].

The overall trend of this study lines up with previous studies indicating that PCL immersed in water and saline environments undergo hydrolytic degradation and weakened mechanical properties after a certain period. The decline in average properties after 60 days and potential slight recoveries on day 90 would suggest a more complex relationship tending hydrolytic degradation, crystallinity levels, and water uptake. Therefore, the mechanical properties of film will still be limited for long-term viability unless designed.

### 3.6. Biodegradability of PCL Films

Gravimetric examination confirmed active biodegradation of the PCL films over the 90-day period of exposure to natural seawater at 15 °C, illustrating in **(Figure 7.A)**. As a starting point, at day 0 of the experiment, there was no weight loss at this time, indicative of polymer having good initial stability in the marine environment.

Following 30 days of immersion, the films revealed, very minimal loss of 0.02%, again, indicating limited degradation at the early stage under cold seawater conditions. The reduction of temperature (15 °C) led to reduced hydrolytic cleavage of the ester bonds within the PCL polymer and slowed microbiological activity, which can both be highly influential on the rate of biodegradation of aliphatic polyesters.

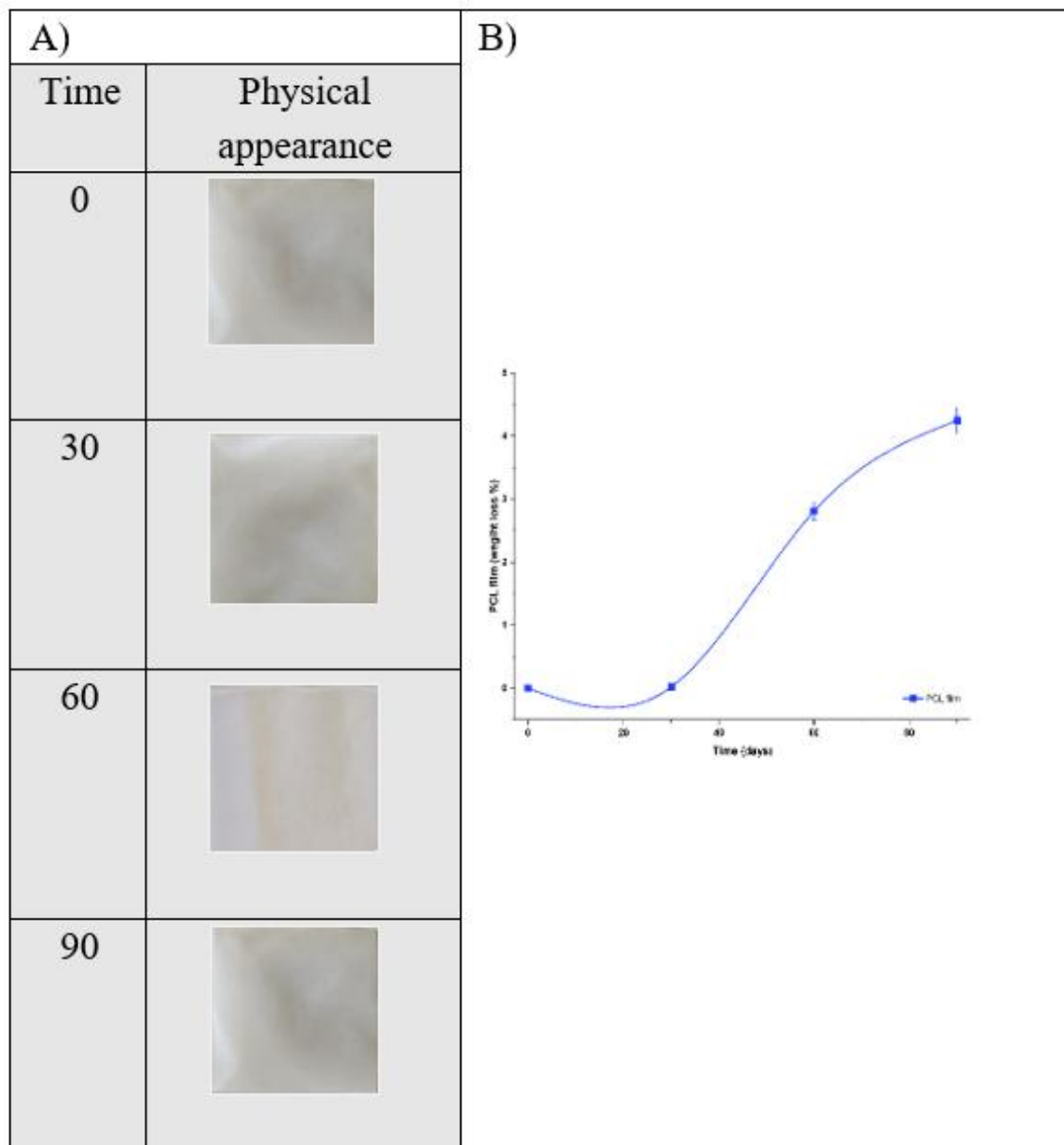
Following 60 days, mass loss had increased significantly to 2.81%, suggesting that hydrolytic processes had progressed beyond the surface of the films and started to influence the bulk polymer structure.

Finally, at day 90, the weight loss of the films indicated 4.25%, indicating that there had been some continuing breakdown of the films over the time of the test period with relatively low overall breakdown. See **(Figure 7.B)**.

In examining the studies from other researchers, the amount of mass loss reported in the current study (~4.25% after 90 days at 15 °C) is less than that of studies which reported the degradation of PCL at higher testing temperatures (25 – 30 °C) or in modified/seawater that are more biologically active [27]. Heimowska et al., (2017) recorded complete degradation of some films of PCL after 6 weeks of testing in Baltic Sea water. However, the degradation of PCL films was slow when the films were tested in less aggressive waterbodies [28]. Therefore, the results support that biodegradation of PCL takes place relatively slowly depending on temperature, microbial content, thickness of the films, and whether the films were crystalline or amorphous.

Overall, the results of our study illustrate that PCL films are degraded slowly in the influence of natural seawater at 15 °C. The slow degradation is due to the semi-crystalline structure and hydrophobic character of PCL, which prevents penetration of water and slows degradation.

However, an increase in mass loss indicates that after considerable time, films may ultimately degrade in marine conditions.



**Figure 7.** Biodegradability of pure PCL films in natural seawater at 15 °C. **(A)** Physical appearance of the films after 0, 30, 60, and 90 days of immersion. **(B)** Weight loss (%) of PCL films as a function of incubation time, showing gradual degradation with increasing exposure duration.

#### 4. Conclusions

This study assessed the performance of unmodified polycaprolactone (PCL) films as biodegradable barriers for protecting archaeological sites from saltwater intrusion. In total, the 90-day exposure period evaluated how PCL performed against saline conditions for hydrophobicity, mechanical properties, and long-term stability. The reduction in contact angle, FTIR showing hydrolytic degradation of the PCL, increases in saltwater permeability, and loss in tensile strength showed all together that unmodified PCL films can decline both structurally and functionally when exposed to saline conditions for a long duration. These results demonstrate the necessity for material

changes, including the addition of nanofillers for improving hydrophobicity, chemical stability, resistance to permeability, and durability over time.

The 90-day exposure condition may not be a long enough duration to assess the long-term performance of PCL films in situ, nor did we consider chemical interactions with specific soil types or artifacts. Future research should employ field validation to further support laboratory performance under authentic environmental conditions, including varying temperature, soil conditions, and water movement, which may also include ongoing biological activity in the archaeological site. It will be important to set benchmarks for future performance, such as establishing acceptable levels of loss - less than 5% mass loss of the polymerized PCL over 12 months to protect archaeological assets. This can help future optimization studies on archaeological site protection, while meeting the regulatory standards.

Despite these limitations, this study provides initial knowledge of PCL film performance in saline conditions and could be a foundation for the development of next generation biodegradable materials that allow for environmental sustainability and preservation of cultural heritage.

**Data Availability Statement:** The original contributions presented in this study are included in the article. Further inquiries can be directed to the corresponding author.

**Conflicts of Interest:** The authors declare no conflicts of interest.

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