

Article

Not peer-reviewed version

# Isavuconazole Therapy for Patients with Hematologic Diseases and Hematopoietic Stem Cell Transplantation with and without Breakthrough Invasive Fungal Infections

Fabián Herrera <sup>\*</sup> , [Diego Roberto Torres](#) , [Gustavo Mendez](#) , [Noelia Soledad Mañez](#) , [Rosana Jordán](#) , [Adriana Manzur](#) , [Myrna Cabral](#) , [Manuel Alderete](#) , [Natalia García Allende](#) , [José Benso](#) , [Claudia Salgueira](#) , [María Laura Pereyra](#) , [Hugo Peretti](#) , [Carla Niveyro](#) , [Maximiliano Gabriel Castro](#) , [Federico Pollastrelli](#) , [Silvina García Rojas](#) , [Juan Dapás](#) , [Agustina Risso Patrón](#) , [Verónica Fernández](#) , [Rocío Gago](#) , [Javier Afeltra](#)

Posted Date: 28 August 2025

doi: [10.20944/preprints202508.2030.v1](https://doi.org/10.20944/preprints202508.2030.v1)

Keywords: Isavuconazole therapy; hematologic diseases; hematopoietic stem cell transplantation; breakthrough invasive fungal infections



Preprints.org is a free multidisciplinary platform providing preprint service that is dedicated to making early versions of research outputs permanently available and citable. Preprints posted at Preprints.org appear in Web of Science, Crossref, Google Scholar, Scilit, Europe PMC.

Copyright: This open access article is published under a Creative Commons CC BY 4.0 license, which permit the free download, distribution, and reuse, provided that the author and preprint are cited in any reuse.

Disclaimer/Publisher's Note: The statements, opinions, and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions, or products referred to in the content.

## Article

# Isavuconazole Therapy for Patients with Hematologic Diseases and Hematopoietic Cell Transplantation with and without Breakthrough Invasive Fungal Infections

Fabián Herrera <sup>1</sup>, Diego Torres <sup>1</sup>, Gustavo Mendez <sup>2</sup>, Noelia Mañez <sup>3</sup>, Rosana Jordán <sup>4</sup>, Adriana Manzur <sup>5</sup>, Myrna Cabral <sup>6</sup>, Manuel Alderete <sup>7</sup>, Natalia García Allende <sup>8</sup>, José Benso <sup>9</sup>, Claudia Salgueira <sup>10</sup>, María Laura Pereyra <sup>11</sup>, Hugo Peretti <sup>12</sup>, Carla Niveyro <sup>2</sup>, Maximiliano Castro <sup>1</sup>, Federico Pollastrelli <sup>4</sup>, Silvina García Rojas <sup>4</sup>, Juan Dapás <sup>6</sup>, Agustina Risso Patrón <sup>8</sup>, Verónica Fernández <sup>9</sup>, Rocío Gago <sup>11</sup> and Javier Afeltra <sup>13</sup> on behalf of Multicenter study on Isavuconazol for Treatment and Prophylaxis of Invasive Fungal Infections in Patients with Hematologic Malignancies and Hematopoietic Cell Transplantation (EMISA)

<sup>1</sup> Infectious Diseases Section, Internal Medicine Department, Centro de Educación Médica e Investigaciones Clínicas, CEMIC, Buenos Aires, Argentina

<sup>2</sup> Infectious Diseases Service, Hospital Dr. Ramón Madariaga, Misiones, Argentina

<sup>3</sup> Infectious Diseases Section, Internal Medicine Department, Hospital Italiano de Buenos Aires, Buenos Aires, Argentina

<sup>4</sup> Infectious Diseases Service, Hospital Británico de Buenos Aires, Buenos Aires, Argentina

<sup>5</sup> Infectious Diseases Service, Hospital Rawson, San Juan, Argentina

<sup>6</sup> Infectious Diseases Service, Hospital Central, Mendoza, Argentina

<sup>7</sup> Instituto Alexander Fleming, Buenos Aires, Argentina

<sup>8</sup> Infectious Diseases Service, Hospital Alemán, Buenos Aires, Argentina

<sup>9</sup> Infectious Diseases Section, Internal Medicine Department, Hospital Italiano de San Justo, Buenos Aires, Argentina

<sup>10</sup> Infectious Diseases Service, Sanatorio Anchorena, Buenos Aires, Argentina

<sup>11</sup> Infectious Diseases Service, Hospital Universitario Austral, Buenos Aires, Argentina

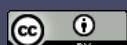
<sup>12</sup> Hematopoietic Stem Cell Transplant Unit, Sanatorio Británico, Rosario, Argentina

<sup>13</sup> Microbiology Laboratory, Hospital Dr. José María Ramos Mejía, Buenos Aires, Argentina

\* Correspondence: to Fabián Herrera: fabian1961@gmail.com

## Abstract

There are no data available on the effectiveness and safety of isavuconazole (ISA) for treating breakthrough invasive fungal infections (bIFIs). A retrospective and prospective cohort study was conducted between January 2020 and March 2025 in 13 centers in Argentina. Hematologic diseases (HD) and hematopoietic cell transplantation (HCT) patients who received ISA for IFI were included and followed for 12 weeks. Patients with proven and probable bIFIs and non-bIFIs were compared. One hundred and sixty-three patients were included. IFIs were classified as proven (13.5%), probable (26.9%) and possible (59.5%). Among 66 proven and probable IFIs, 53% were bIFIs, with aspergillosis and mucormycosis being the most common. Twenty-three (34.8%) patients had acute myelogenous leukemia, and 40.9% had received HCT. Forty-eight (72.7%) patients experienced neutropenia, with a median duration of 26 days (interquartile range [IQR] 16-44). Fluconazole and posaconazole were the antifungal prophylaxis most frequently received. ISA was prescribed as first-line therapy in 31 (46.9%) patients. The other 35 received ISA as a continuation therapy, mainly as a step-down therapy after liposomal amphotericin B. Four (6.1%) patients developed adverse effects, and one discontinued ISA. The 90-day overall clinical response between patients with bIFI vs. non-bIFI was 91.4% vs. 70.9% ( $p=0.052$ ). The 90-day overall and IFI-related mortality rates were, respectively, 11.4% vs. 32.3% ( $p=0.001$ ).



0.068) and 5.7% vs. 9.7% ( $p = 0.659$ ). The study data evidence ISA effectiveness and safety for the treatment of HD and HCT patients with and without bIFIs.

**Keywords:** Isavuconazole therapy; hematologic diseases; hematopoietic cell transplantation; breakthrough invasive fungal infections

## 1. Introduction

Invasive fungal infections (IFIs) are a frequent complication in patients with hematologic diseases (HD) and hematopoietic cell transplantation (HCT), with significant morbidity and mortality rate, as well as high healthcare costs [1-4].

Patients with acute leukemia and prolonged neutropenia, as well as those with allogeneic HCT with high doses of corticosteroids, have an IFI incidence of 7-13.2% and 8.8-16%, respectively (1,2,3,5). The epidemiology of IFIs has evolved worldwide, with a significant predominance of mold in recent decades, largely *Aspergillus* spp. *Mucorales* and *Fusarium* spp. [1,2]. Two multicenter studies have addressed this issue. Data from the TRANSNET surveillance study in the United States identified 983 IFIs among 875 HCT recipients, with invasive aspergillosis being the most common (43%) [6]. According to the Prospective Antifungal Therapy (PATH) Alliance registry, invasive aspergillosis was the most common IFI (59.2%) among 234 adult HCT patients [7]. More recently, a study carried out in Switzerland in 515 allogeneic HCT recipients showed that 48 (9.3%) patients developed 51 proven/probable IFI, with invasive aspergillosis (67%) and mucormycosis (18%) being the most frequent [8]. This is largely due to primary antifungal prophylaxis strategies, which are highly active against *Candida* spp. [9,10]. In this regard, all high-risk patients currently receive antifungal prophylaxis, even before this epidemiology change [11]. IDSA, ESCMID, ASTCT, ECIL, and AGIHO/DGHO guidelines recommend posaconazole (POSA) use as the first choice, followed by voriconazole (VORI). Equinocandins and fluconazole are recommended as alternative drugs due to their narrow spectrum. Some guidelines consider isavuconazole (ISA) as an alternative antifungal prophylaxis in those cases where POSA and VORI are not appropriate (prolonged QTc, patients who receive QTc-prolong medications, or drug-drug interaction) [12-16]. Therefore, most of the IFIs generally developed are breakthrough IFIs (bIFIs) [17]. In this sense, a systematic review and meta-analysis that identified 991 patients who received ISA prophylaxis found an incidence of bIFIs of 7% [18]. They pose a significant challenge for diagnosis; furthermore, no randomized studies have been conducted to determine the optimal treatment option [19,20].

In this complex scenario, several antifungal drugs have proved effective for treating IFIs. ISA was approved for the treatment of invasive aspergillosis and mucormycosis, based on the SECURE and VITAL trials [21,22]. After the implementation of these studies ISA was approved by key regulatory agencies, including the US Food and Drug Administration (FDA) and the European Medicines Agency (EMA) [23,24]. Given its efficacy and safety, several guidelines recommend ISA as a first-line treatment for invasive aspergillosis and as a step-down therapy or first-line therapy for mucormycosis [12,25]. In addition, many real-life studies are consistent with the data obtained in pivotal studies [26-30].

ISA has several advantages over VORI, including *Mucorales* coverage, stable drug levels with low intra- and interpatient variability, lower CYP3A4 inhibition, resulting in significantly fewer drug-drug interactions, a highly safe profile, and less drug discontinuation, among others [31-36]. In terms of adverse effects, clinical trial data show an overall tolerability profile comparable to voriconazole, with nausea, vomiting, and diarrhea being the most common events. However, ISA is associated with a lower incidence of hepatotoxicity and visual disturbances, with lack of QTc prolongation being a key differentiating safety feature. On the other hand, it causes mild concentration-dependent QTc interval shortening. It is also associated with fewer severe skin reactions compared to other azoles [21,22]. These characteristics are crucial in patients with HD and HCT, as they receive a large number of medications, especially immunosuppressants, which interact

with potent CYP3A4 inhibitors, such as VORI. These inhibitors can also have a high rate of adverse effects and are therefore unsuitable for use in these patients.

To the best of our knowledge, no multicenter studies have been published despite all the above mentioned advantages of ISA therapy for bIFIs.

This study aimed to outline the use of ISA for the treatment of IFIs in patients with HD and HCT. We further aimed to describe and compare those who have proven and probable bIFI and non-bIFI in terms of ISA effectiveness and safety, and patients' outcomes.

## 2. Materials and Methods

### 2.1. Setting, Patients and Study Design

A retrospective and prospective observational multicenter study was performed in 13 referral teaching centers (8 private and 5 public) specialized in the management of patients with HD and HCT in Argentina.

Adult patients ( $\geq 18$  years of age) treated with ISA for IFIs and managed as inpatients or outpatients were included. The retrospective cohort comprised patients included from 1 January 2020 to 31 March 2024; however, all of them were treated and followed up by the Infectious Diseases physicians conducting the study. Patients from the prospective cohort were recruited from 1 April 2024 to 31 March 2025. For the total cohort, the following criteria were met: (a) patients presenting with HD or autologous and allogeneic HCT; (b) those treated with ISA for IFIs for at least 7 days; and (c) those followed until day 90 since the beginning of ISA or until the patient's death, whichever occurred first.

Patients were excluded in case of missing data that precluded the assessment of baseline, clinical, microbiological, treatment characteristics, and outcomes.

Patients were identified through data files from the Infectious Diseases Services, which treat and follow up all patients with HD and HCT at each center. Data were obtained from direct patient care, medical records, and data from laboratory, microbiology, and pathology databases. Clinical, microbiological, diagnostic, treatment, and outcome variables from the total cohort were evaluated. In addition, these variables were compared between patients with proven and probable bIFIs and non-bIFIs.

Patient data were recorded with RedCap (Research Electronic Data Capture) software (RedCap version 13.7.19) and the server hosting was provided by the Argentine Society of Infectious Diseases.

The study was approved by the Ethics Committees from the different participating institutions, and patient informed consent was waived.

### 2.2. Definitions

Proven, probable, and possible IFIs were defined according to the revised and updated European Organization for Research and Treatment of Cancer and Mycoses Study Group EORTC/MSG criteria [37]. Proven IFI was defined as histopathologic, cytopathologic, direct microscopic examination, or culture of a biopsy or other specimen obtained by a sterile procedure from a normal sterile site. Probable mold infection was defined as that occurring in patients with a) one host factor: recent history of neutropenia ( $< 500$  neutrophils/mm $^3$ ) for  $> 10$  days, allogeneic HCT, prolonged use of corticosteroids, treatment with other T-cell or B-cell-immunosuppressants; b) at least one clinical feature: pulmonary CT-scan showing nodules with or without a halo sign, air crescent sign, cavity, consolidation or a reverse halo sign; evidence of tracheobronchitis, sino-nasal or central nervous system infection; and c) microbiological evidence: microscopic detection or culture of any mold from bronchoalveolar lavage (BAL) or sinus aspirates, galactomannan (GM) antigen test  $\geq 1.0$  from serum or BAL, or single serum or plasma  $\geq 0.7$  and BAL fluid  $\geq 0.8$  or *Histoplasma* urinary antigen. A possible mold infection was defined as that occurring in patients with one host factor and at least one pulmonary imaging on CT scan.

bIFIs were defined according to the Mycoses Study Group Education and Research Consortium (MSG-ERC) and the European Confederation of Medical Mycology (ECMM) [38]. bIFI was defined to occur during exposure to an antifungal drug, including fungi outside the spectrum of activity of an antifungal. bIFI time point is the first clinical sign or symptom, mycological finding, or radiological feature attributable to it. bIFI time point begins when each antifungal reaches plasma steady state and finishes during the last dose interval upon drug discontinuation.

Risk factors for IFIs were considered and defined as follows: a) neutropenia  $< 500$  neutrophils/mm $^3$  for  $> 10$  days, and profound and prolonged neutropenia  $< 100$  neutrophils/mm $^3$  for  $> 14$  days prior to the diagnosis of IFI; b) high doses of corticosteroids, such as prednisone (or equivalent) at doses  $\geq 20$  mg/day for a period  $\geq 2$  weeks prior to IFI, and the use of biological agents and/or anti-lymphocyte therapies within three months prior to IFI; c) recent chemotherapy, such as the cycle of immunosuppressant drugs within one month prior to the diagnosis of IFI; d) T-cell depletion, such as antithymocyte globulin or alemtuzumab for conditioning regimen of allogeneic HCT; e) graft-versus-host disease (GvHD) and grading consistent with consensus guidelines [39]; f) cytomegalovirus infection or disease occurring within 15 days prior to the diagnosis of IFI; g) iron overload as ferritin serum level  $> 2000$  ng/ml [40]; and h) no HEPA filter system in the isolation room for induction chemotherapy in acute leukemia.

The first-line antifungal treatment was selected by the investigator based on the suspicion or diagnosis of IFI according to published guidelines [12–15,25]. Continuation treatment with ISA was considered either as a step-down therapy, or when the first-line drug could not be prescribed.

Preemptive therapy was given to patients whose diagnosis was based on positive GM and/or pulmonary CT-scan imaging, while targeted therapy implied that the diagnosis was made by microscopic detection or culture of any mold or yeast in clinical samples.

Favorable response to treatment was defined as absence of fever, improved signs and symptoms of the initial infectious source, decrease in GM index, and/or improved results in CT-scan imaging. Partial remission: improved signs and symptoms and imaging, though without resolution. Stable disease: improved signs and symptoms, though with no changes in imaging. Cure: clinical and imaging resolution.

Unfavorable outcome was defined as the patient's death during the follow-up. Attributable mortality was considered to be the patient's death with no response to treatment and documented clinical, radiological, microbiological, or histological findings suggestive of active IFI.

### 2.3. Statistical Analysis

The study population was characterized by descriptive statistics. For continuous variables, centrality (median) and dispersion (IQR) measures were used according to the distribution of variables. Categorical variables were analyzed using absolute frequency and percentage. Groups were compared using the U Mann-Whitney test for continuous variables and the Fisher exact test or the chi-square test for categorical variables. For all tests, a 95% level of statistical significance was used. Analyses were performed with the SPSS (Statistics for Windows, Version 22.0, Armonk, NY, USA) software packages.

## 3. Results

### 3.1. Characteristics and Outcomes of Patients' Cohort

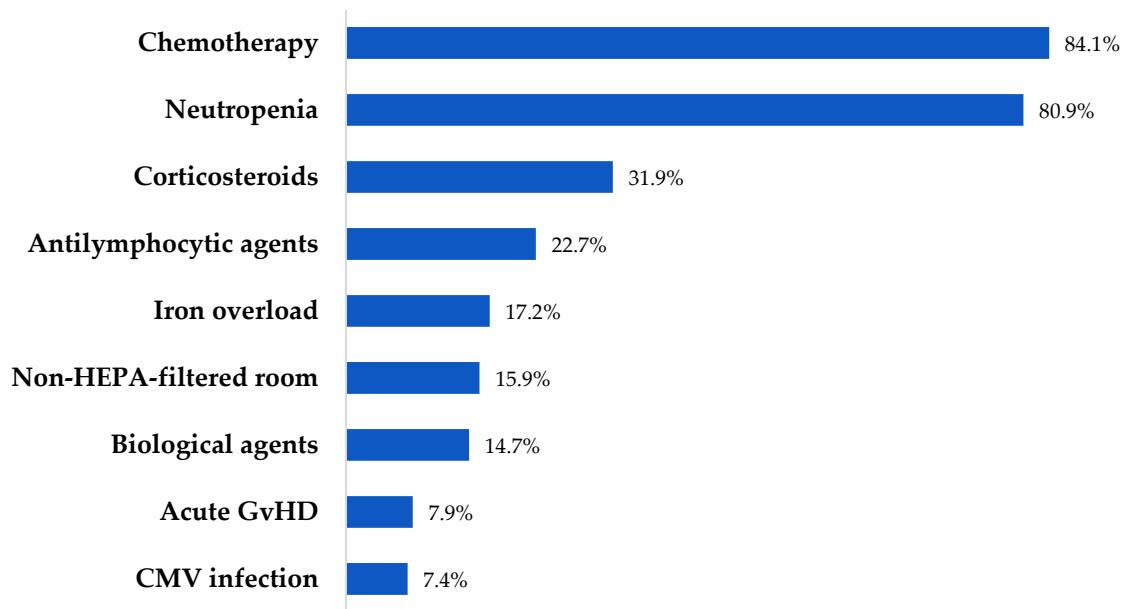
A total of 163 patients (126 retrospective and 37 prospective) diagnosed with IFI were included. Acute myelogenous leukemia (AML) and acute lymphoblastic leukemia (ALL) were the most frequent underlying diseases (96, 58.9%), and were active in 125 (76.6%). Forty-nine (30.1%) had undergone HCT, with allogeneic being the most common type. Thirteen (43.3%) and 7 (23.3%) of allogeneic HCT developed acute and chronic GvHD, respectively. From the total cohort, 63 (38.6%) patients presented with oral mucositis, and 10 (6.1%) HCT patients developed acute or chronic GvHD affecting the gastrointestinal tract. One hundred and thirty-two (80.9%) patients were neutropenic at

the onset of IFI. Of them, 115 (87.1%) presented profound and prolonged neutropenia, with a median duration of 27 days (IQR: 16-47). The cohort comprised patients with increased risk factors for IFIs, with a median of 2 (IQR: 2-3). Patients' baseline characteristics and risk factors are outlined in Table 1 and Figure 1.

**Table 1.** Baseline characteristics of patients diagnosed with IFIs.

Variables	Patients (n=163)
	N (%)
Age (years) median (IQR)	50 (38-63)
Sex, male	95 (58.3)
Charlson comorbidity index score ≥ 3	73 (44.8)
Underlying disease	
Acute myelogenous leukemia	76 (46.6)
Acute lymphoblastic leukemia	20 (12.3)
Non-Hodgkin lymphoma	16 (9.8)
Myelodysplastic syndrome	8 (4.9)
Hodgkin lymphoma	11 (6.7)
Aplastic anemia	9 (5.5)
Chronic myelogenous leukemia	6 (3.7)
Chronic lymphoblastic leukemia	4 (2.4)
Multiple myeloma	7 (4.3)
Others	6 (3.7)
Disease status	
Complete remission	38 (23.3)
Partial remission	8 (4.9)
Relapsed	33 (20.2)
Refractory	18 (11)
Recently diagnosed	66 (40.5)
HCT	49 (30.1)
Allogeneic	30 (18.4)
HLA matching and donor type	
Haploidentical	15 (9.2)
Matched unrelated donor	5 (3.1)
Matched related donor	10 (6.1)
T-cell depletion	9 (5.5)
Acute GvHD	13 (7.9)
Grade I	2 (1.2)
Grade II	4 (2.4)
Grade III	7 (4.3)
Grade IV	0 (0)
Chronic GvHD	7 (4.3)

Abbreviations: HCT= Hematopoietic Cell Transplantation; GvHD= Graft-versus-Host Disease. Other underlying diseases include chronic myeloid leukemia, myelofibrosis, hairy cell leukemia, Richter's syndrome, and Sézary syndrome.



**Figure 1.** Risk factors for IFIs.

Proven and probable IFIs were diagnosed in 66 (40.5%) patients and possible IFIs in 97 (59.5%). Ninety-two (56.4%) were bIFIs. The most common locations were lungs (147, 90.2%) and paranasal sinuses (23, 14.1%). Lung CT scan showed nodules in 89 (54.6%) and halo sign in 37 (22.7%). IFIs characteristics are depicted in Table 2.

**Table 2.** IFIs classification, location and radiological characteristics.

Variables	Patients (n=163)	N (%)
IFI classification		
Proven	22 (13.5)	
Probable	44 (26.9)	
Possible	97 (59.5)	
IFI location		
Lungs	147 (90.2)	
Paranasal sinuses	23 (14.1)	
Liver	5 (3.1)	
Skin and soft tissue	5 (3.1)	
Central nervous system	3 (1.8)	
Disseminated	3 (1.8)	
Lung CT scan		
Nodules	89 (54.6)	
Ground glass appearance	59 (36.2)	
Halo sign	37 (22.7)	
Tree in bud	22 (13.5)	
Alveolar infiltrate	14 (8.6)	
Cavity	3 (1.8)	
Air crescent sign	1 (0.6)	
Reverse halo sign	1 (0.6)	

ISA treatment was prescribed as preemptive therapy in 136 (83.4%) and as targeted therapy in 29 (17.8%) patients. It was used as first-line therapy in 73 (44.8%) of all the patients, and 153 (93.8%) received it as monotherapy. Ten (6.1%) patients received ISA in combination with L-AmB. The other antifungal drugs used as first-line therapy (90 patients) were L-AmB (65, 72.2%), VORI (13, 14.4%), POSA (6, 6.7%), LC-AmB (4, 4.4%), caspofungin (1, 1.1%), and fluconazole (FLUCO) (1, 1.1%). ISA administration route was oral (77, 47.2%), intravenous (IV) followed by oral (57, 34.9%), IV (27, 16.6%), and oral followed by IV (2, 1.2%), with a median duration of 90 days (IQR: 59-113). Five patients undergoing treatment with medication that interacts with POSA and VORI (venetoclax 2, sirolimus 2 and ponatinib 1) were therefore administered ISA.

Seven (4.3%) patients developed related adverse effects (nausea 1, hepatobiliary abnormalities 6, rash 1, and shortened QTc 1), and only 1 (0.6%) had to discontinue ISA. One hundred and thirty (79.7%) patients achieved a favorable response at week 12 (cure 54.6%, partial remission 19.6%, and stable disease 5.5%). Overall mortality was 22.7%, and IFI-related mortality was 4.9%. Ninety-day overall mortality in patients with pulmonary vs. non-pulmonary locations was 23.2% vs. 18.4% ( $p=0.53$ ).

### 3.2. Characteristics and Outcomes of Patients with Probable and Proven IFIs

A total of 66 patients were diagnosed with proven and probable IFIs, and 35 (53%) of them were bIFIs. Both bIFIs and non-bIFIs patients had similar baseline characteristics regarding sex, Charlson score comorbidity index, underlying diseases, and disease status. Forty-eight (72.7%) patients had neutropenia, mostly profound and prolonged, with a median duration of 26 days (IQR: 16-44).

The most common primary antifungal prophylaxis administered to bIFIs patients was FLUCO (13, 37.1%), followed by POSA (10, 28.6%). Baseline characteristics and the antifungal prophylaxis prescribed are outlined in Table 3.

**Table 3.** Baseline characteristics and primary antifungal prophylaxis of patients with proven and probable non-bIFIs and bIFIs.

Variable	Total (n = 66)	non-bIFI (n = 31)	bIFI (n = 35)	p- value
Age, median (IQR)	47 (39-61)	54 (43-67)	44 (36-52)	0.014
Male sex – n (%)	40 (60.6)	16 (51.6)	24 (68.6)	0.159
Charlson comorbidity index $\geq 3$ – n (%)	29 (43.9)	17 (54.8)	12 (34.3)	0.093
Underlying disease – n (%)				
Acute myelogenous leukemia	23 (34.8)	11 (35.5)	12 (34.3)	0.918
Acute lymphoblastic leukemia	7 (10.6)	1 (3.2)	6 (17.1)	0.41
Myelodysplastic syndrome	7 (10.6)	2 (6.4)	5 (14.3)	0.433
Non-Hodgkin lymphoma	4 (6.1)	4 (12.9)	0 (0)	0.043
Hodgkin lymphoma	10 (15.1)	4 (12.9)	6 (17.1)	0.738
Multiple myeloma	7 (10.6)	5 (16.1)	2 (5.7)	0.239
Disease status – n (%)				
Complete remission	16 (24.2)	5 (16.1)	11 (31.4)	0.165
Partial remission	4 (6.1)	1 (3.2)	3 (8.6)	0.616
Relapsed	17 (25.8)	9 (29)	8 (22.9)	0.567
Refractory	9 (13.6)	7 (22.6)	2 (5.7)	0.071
Recently diagnosed	20 (30.3)	9 (29)	11 (31.4)	0.832
HCT – n (%)	27 (40.9)	9 (29)	18 (51.4)	0.064
Allogeneic	13 (19.7)	4 (12.9)	9 (25.7)	0.228
Corticosteroid use – n (%)	24 (364)	11 (35.5)	13 (37.1)	0.888
Biological agents – n (%)	15 (22.7)	7 (22.6)	8 (22.9)	0.978
Antilymphocyte drugs – n (%)	12 (18.2)	3 (9.7)	9 (25.7)	0.117
Neutropenia – n (%)	48 (72.7)	21 (67.7)	27 (77.1)	0.392

Antifungal prophylaxis – n (%)	35 (53)	0 (0)	35 (100)	–
Fluconazole	13 (19.7)	0 (0)	13 (37.1)	–
Posaconazole	10 (15.1)	0 (0)	10 (28.6)	–
Voriconazole	2 (3)	0 (0)	2 (5.7)	–
L-AmB	5 (7.6)	0 (0)	5 (14.3)	–
LC-AmB	2 (3)	0 (0)	2 (5.7)	–
Caspofungin	3 (4.5)	0 (0)	3 (8.6)	–

Abbreviations: IFI, Invasive Fungal Infection; non-bIFI, non-breakthrough IFI; bIFI, breakthrough IFI; IQR, interquartile range; HCT, hematopoietic cell transplantation; L-AmB, Liposomal Amphotericin B; LC-AmB, Amphotericin B lipid complex. *P*-values were obtained using the chi-square or Fisher's exact test for categorical variables and the Mann-Whitney U test for continuous variables.

Invasive aspergillosis and mucormycosis were the most frequent IFIs in both groups. We compared and contrasted IFI locations in non-bIFI and bIFI patients and found lungs in 26 (83.9%) vs. 27 (77.1%), *p* = 0.492, and paranasal sinuses in 7 (22.6%) vs. 8 (22.9%), *p* = 0.97, respectively. A few patients had other locations. The most common CT-scan findings were nodules in 32 (48.5%) patients, halo sign in 17 (25.8%), and a ground glass appearance in 20 (30.3%), with no differences between groups. IFIs were diagnosed by microscopic detection in 16 (24.2%) patients, culture in 29 (43.9%), GM test in 35 (53%), and histopathology in 17 (25.8%).

The etiology and methodology used for the diagnosis of proven and probable IFIs are described in Table 4.

**Table 4.** Type of mycoses, etiology, diagnostic methods, and treatment of proven and probable non-bIFIs and bIFIs.

Variable	Total (n = 66)	non-bIFI (n = 31)	bIFI (n = 35)	p- value
Type of IFI – n (%)				
Aspergillosis	43 (65.1)	22 (70.9)	21 (60)	0.351
Mucormycosis	8 (12.1)	2 (6.4)	6 (17.1)	0.265
Fusariosis	1 (1.5)	0 (0)	1 (2.9)	1
Other hyalo or phaeohyphomycosis	5 (7.6)	2 (6.4)	3 (8.6)	1
Histoplasmosis	1 (1.5)	1 (2.9)	0 (0)	0.470
Cryptococcosis	1 (1.5)	1 (2.9)	0 (0)	0.470
Unidentified hyphae	8 (12.1)	3 (9.7)	5 (14.3)	0.713
Microscopic detection – n (%)				
Septate branched hyphae	16 (24.2)	9 (29)	7 (20)	0.245
Coenocytic hyphae	10 (15.1)	7 (22.6)	3 (8.6)	0.170
<i>Cryptococcus</i> yeast	5 (7.6)	1 (3.2)	4 (11.4)	0.360
Culture isolates – n (%)				
<i>Alternaria</i> sp.	1 (1.5)	0 (0)	1 (2.9)	1
<i>Aspergillus</i> sp.	3 (4.5)	1 (3.2)	2 (5.7)	1
<i>Aspergillus flavus</i> complex	5 (7.6)	2 (6.4)	3 (8.6)	1
<i>Aspergillus fumigatus</i> complex	5 (7.6)	4 (12.9)	1 (2.9)	0.178
<i>Aspergillus niger</i>	3 (4.5)	2 (6.4)	1 (2.9)	0.596
<i>Cryptococcus neoformans</i> var. <i>neoformans</i>	1 (1.5)	1 (2.9)	0 (0)	0.470
<i>Cunninghamella</i> sp.	1 (1.5)	0 (0)	1 (2.9)	1
<i>Curvularia</i> sp.	2 (3)	2 (6.4)	0 (0)	0.216
<i>Fusarium</i> sp.	1 (1.5)	0 (0)	1 (2.9)	1
<i>Penicillium</i> sp.	2 (3)	0 (0)	2 (5.7)	1
<i>Rhizopus</i> sp.	2 (3)	0 (0)	2 (5.7)	1
<i>Rhizopus microsporum</i>	1 (1.5)	0 (0)	1 (2.9)	1
<i>Rhizopus oryzae</i>	1 (1.5)	0 (0)	1 (2.9)	1

<i>Rhizopus arrhizus</i>	1 (1.5)	0 (0)	1 (2.9)	1
Aspergillus GM – n (%)				
Positive in serum	12 (18.2)	5 (16.1)	7 (20)	0.684
Positive GM in BAL	17 (25.8)	9 (29)	8 (22.9)	0.567
Positive serum + BAL GM	6 (9.1)	3 (9.7)	3 (8.6)	1
Histoplasma urinary antigen – n (%)	1 (1.5)	1 (2.9)	0 (0)	0.470
Histopathology – n (%)				
Hyphal invasion of blood vessels	7 (10.6)	3 (9.7)	3 (8.6)	1
Septate branched hyphae	5 (7.6)	3 (9.7)	2 (5.7)	0.659
Coenocytic hyphae	6 (9.1)	2 (6.4)	4 (11.4)	0.676
Yeast	1 (1.5)	1 (3.2)	0 (0)	0.470

Abbreviations: IFI, Invasive Fungal Infection; non-bIFI, non-breakthrough IFI; bIFI, breakthrough IFI; GM, galactomannan; BAL, bronchoalveolar lavage. *P*-values were obtained using the chi-square or Fisher's exact test for categorical variables.

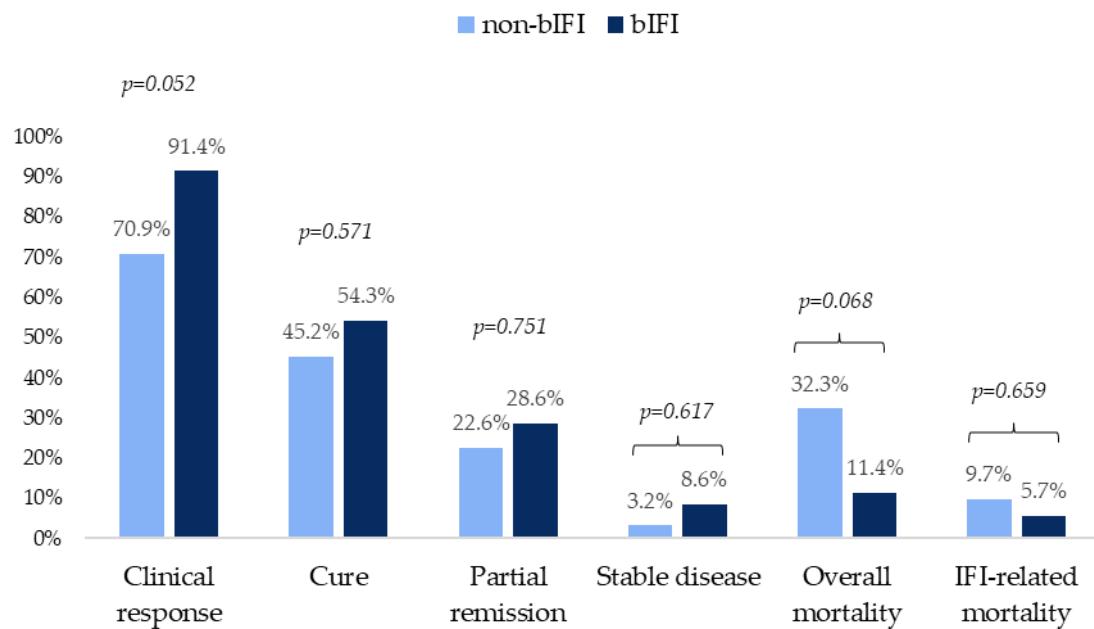
ISA was prescribed as monotherapy in 62 (93.9%) patients and as first-line treatment in 31 (46.9%). In the bIFI group, 15 patients (42.9%) received ISA as first-line treatment. Ten (66.7%) were under prophylaxis with FLUCO, 2 (13.3%) with VORI, 2 (13.3%) with L-AmB, and 1 (6.7%) with LC-AmB. As a continuation treatment, it was used in 20 patients (57.1%). The reasons for prescribing ISA were step-down therapy after L-AmB in 11 (55%), (10 had undergone POSA prophylaxis), and VORI in 1 (5%); related-adverse effects with L-AmB in 2 (10%) or VORI in 2 (10%); and combination treatment with L-AmB in 3 (15%), or caspofungin in 1 (5%). The median duration of treatment in bIFI vs. non-bIFI patients was 90 days (IQR: 62.5-119.5) vs. 85 days (IQR: 61.5-149), *p*=0.908. Source control was performed in 18 (27.3%) patients, and mostly consisted of paranasal sinus endoscopic surgery (7 patients in each group). Treatment of proven and probable characteristics is described in Table 5.

**Table 5.** Treatment of proven and probable non-bIFIs and bIFIs.

Variable	Total (n = 66)	non-bIFI (n = 31)	bIFI (n = 35)	p-value
ISA treatment modality-n (%)				
Preemptive therapy	39 (59.1)	17 (54.8)	22 (62.9)	0.508
Targeted therapy	29 (43.9)	14 (45.2)	15 (42.8)	0.851
Treatment with ISA – n (%)				
As first-line therapy	31 (46.9)	16 (51.6)	15 (42.9)	0.477
As continuation therapy	35 (53%)	15 (48.4)	20 (57.1)	0.477
Other first-line therapy – n (%)				
L-AmB	23 (34.8)	8 (25.8)	15 (42.9)	0.147
LC-AmB	1 (1.5)	1 (3.2)	0 (0)	0.470
Voriconazole	11 (16.7)	8 (25.8)	3 (8.6)	0.097
Posaconazole	1 (1.5)	1 (3.2)	0 (0)	0.470
Fluconazole	1 (1.5)	1 (3.2)	0 (0)	0.470
Caspofungin	1 (1.5)	0 (0)	1 (2.9)	1

Abbreviations: IFI, Invasive Fungal Infection; non-bIFI, non-breakthrough IFI; bIFI, breakthrough IFI; ISA, isavuconazole; L-AmB, Liposomal Amphotericin B; LC-AmB, Amphotericin B lipid complex. *P*-values were obtained using the chi-square or Fisher's exact test for categorical variables.

Regarding outcomes, a large number of patients achieved a favorable response, being higher in those with bIFI. Overall and IFI-related mortality in this group were 11.4% and 5.7%, respectively. Outcome variables are shown in Figure 2.



**Figure 2.** Clinical outcomes at 12 weeks in patients with probable and proven non-bIFIs and bIFIs.

#### 4. Discussion

The study assessed the effectiveness and safety of ISA for treating HD and HCT patients with IFIs. Patients with several risk factors for IFIs, most of them neutropenic, were included. Patients with proven and probable IFIs, either non-bIFIs or bIFIs, were analyzed separately. Aspergillosis and mucormycoses were the IFIs most commonly diagnosed, largely located in the lungs and paranasal sinuses. Almost half of the cases received ISA as a first-line treatment, mainly as monotherapy. Among bIFI patients, those who received ISA as a first-line therapy were mainly undergoing FLUCO or VORI prophylaxis. Half of the patients treated with ISA as a continuation therapy after L-AmB were receiving POSA prophylaxis. In a large proportion of patients with bIFIs, a clinical response was observed, with low overall and IFI-related 90-day mortality rates.

These are the major findings of the present study: first, in terms of IFI epidemiology, our cohort had similarities and differences with other real-life studies. In a multicenter study including centers from the USA, Europe, and Brazil, aspergillosis was the leading cause of IFI (79%), followed by fusariosis (8%) [41]. In contrast, in our cohort fusariosis only represents 1.5%. Second, approximately 50% of the patients had bIFI. Aspergillosis and mucormycoses were the most frequent molds in those with proven and probable IFI. In this respect, a multicenter study from Spain reported bIFI in 121 cases, with aspergillosis in 59% and mucormycosis in 7% being the most common among 94 patients with proven or probable bIFI [42]. Third, as reported in the SECURE trial and many real-life studies, most IFI locations were the lungs [21,26,27,29]. Fourth, bIFIs are currently and frequently a major concern in clinical practice among HD and HCT patients, as it is difficult to identify the type of fungus involved. The sensitivity of GM in BAL proved to be lower in patients that receive antifungal prophylaxis compared to those that do not: 52% vs. 81%. Likewise, the sensitivity of serum GM and culture in BAL is 31.3% and 18.8%. A combination of these methods can increase diagnostic efficacy [43,44]. Based on these findings, in our cohort bIFI could be diagnosed using several diagnostic methods. In this regard, the ECMM consensus status recommends the use of all available methods to diagnose bIFI [45]. Fifth, almost half of the patients received ISA as a first-line treatment, mostly as monotherapy. This differs from the literature, which recommends using L-AmB as a first-line therapy [19,20,46]. Sixth, 10 patients under continuation ISA therapy after L-AmB were receiving primary prophylaxis with POSA. Interestingly, ISA and POSA have the same antifungal spectrum, and both proved effective for aspergillosis and mucormycosis [21,22,47]. Antifungal cross-resistance between

these two azoles does not necessarily occur. Cross-resistance between POSA (used for prophylaxis) and ISA (used for treatment) is complex and not fully deterministic. While a theoretical risk exists due to their shared drug class, several factors mitigate absolute cross-resistance. Thus, ISA could be considered a viable therapeutic option even after POSA prophylaxis failure. In this sense, the binding affinity of ISA to the fungal Cyp51A enzyme target is different from that of POSA. The environmentally driven tandem repeats (TR34/L98H and TR46/Y121F/T289A) are the most relevant resistance mechanisms [48]. These mutations often confer panazole resistance to both POSA and ISA. However, their prevalence is not yet universal, and breakthrough infections in patients on prophylaxis can still be caused by wild-type or susceptible isolates with other resistance mechanisms. Key studies support this lack of absolute cross-resistance. The phase 3 SECURE trial and subsequent analyses have documented successful outcomes with ISA in patients who had received prior azole prophylaxis, including POSA [21]. Moreover, one of the several reasons for developing bIFI is that the antifungal agent does not achieve enough serum levels. This is frequently observed in VORI, but has also been described with POSA, even in tablet formulations. A study in patients with acute leukemia and HCT receiving POSA prophylaxis found that 18% of them had subtherapeutic serum levels (<700 ng/mL). Factors such as having diarrhea, receiving proton pump inhibitors, and weighing more than 90 kg were associated with subtherapeutic serum levels, with the first two being common in HD and HCT patients [49]. Nonetheless, as POSA serum levels were not available, we cannot state that this could occur in our patients. On the other hand, ISA levels are adequate, even in patients with mucositis and gastrointestinal GvHD, as could often be the case with many of our patients [50,51]. Seventh, the cohort had a low rate of related adverse effects and drug discontinuation, which is consistent with the literature [29,30]. Given its safety, ISA is suitable for treating IFIs in severely ill patients. Eighth, our patients with bIFI had higher clinical response and lower mortality rate than those in other real-life studies, which report a mortality rate of 35% [17]. Apart from the effectiveness of ISA, 62.9% of our patients received preemptive therapy, which means that they were treated early. Several studies have demonstrated that this strategy is consistent with higher survival rates [52–54]. We also consider that other factors could have contributed to the high clinical response and low mortality rate, particularly in patients with bIFI. They were younger than non-bIFI patients and had a higher rate of underlying disease in complete or partial remission. Moreover, since all the cohorts were diagnosed with IFI and were followed up by the ID physicians participating in the study, we assume that the diagnostic and therapeutic approach has been timely and appropriate.

Our study has some limitations that should be considered. First, data on the azoles serum levels or susceptibility testing were not available. Patients' good outcomes could only be partly explained by these factors. However, there are no clinical susceptibility cut-off values for molds, except for *Aspergillus fumigatus* complex. In addition, the association between exposure to and efficacy of ISA treatment has not been proven [55]. Second, the sample size of patients with bIFI was small. Therefore, a larger population is required to evaluate the outcome variables. Third, several patients were retrospectively included, which may have led to some biased results. Notwithstanding that, all those patients were treated and prospectively followed by the Infectious Diseases physician conducting the study and missing data were not allowed.

The strengths of our study rely on its multicenter design. It was carried out in healthcare facilities specialized in the treatment of patients with HD and HCT. In addition, all of them were evaluated, treated, and followed up by the investigators. Therefore, our results accurately reveal IFIs complex scenario. Moreover, our study comprised the largest cohort from Latin America.

In conclusion, the study data evidenced ISA effectiveness and safety for the treatment of HD and HCT patients with IFI. It further showed a suitable option for treating patients with bIFIs. However, larger studies should be conducted to confirm this finding.

**Author Contributions:** Conceptualization, F.H., D.T. and J.A.; methodology, F.H., D.T. and J.A.; software, D.T. and J.A.; formal analysis, D.T.; investigation, G.M., N.M., R.J., A.M., M.C., M.A., N.G.A., J.B., C.S., J.B., M.L.P.,

H.P., C.N., M.C., F.P., S.G.R., J.D., A.R.P., V.F. and R.G.; data curation, D.T. and J.A.; writing: original draft preparation, F.H.; writing: review and editing, D.T. and J.A.; funding acquisition, F.H. All authors have read and agreed to the published version of the manuscript.

**Funding:** This work was supported by a research grant from Knight Therapeutics. The Company has not participated in the study design, sample collection, analysis data interpretation, or in the decision to submit the manuscript for publication. Only the authors had full access to the study data files.

**Institutional Review Board Statement:** The study was conducted in accordance with the Declaration of Helsinki and was approved by CEMIC Ethics Committee on November 27, 2023 (Approval identification number 1751) and the ethics committees from the different participating institutions.

**Informed Consent Statement:** Patient consent was waived by the Ethics Committees according to Data Protection Law 25326, section 7, subsection 2.

**Data Availability Statement:** Data are available upon request. Contact the corresponding author.

**Acknowledgments:** We thank Valeria Melia, scientific translator at CEMIC Research Unit, for English edition of the manuscript.

**Potential Conflicts of Interest:** F.H. has participated in advisory boards and/or received speaker honoraria and grants from Gilead, Knight Therapeutics, Merck, Sharp & Dohme (MSD), SteinCares, Biomerieux, Rochem Biocare, TEVA, TAKEDA, and Pfizer. D.T. has participated in advisory boards and/or received speaker honoraria from Gilead, Knight Therapeutics, MSD, Pfizer and GlaxoSmithKline. G.M. has participated in advisory boards and/or received speaker honoraria from Knight Therapeutics. R.J. has participated in advisory boards and/or received speaker honoraria from Knight Therapeutics and Pfizer. J.A. has participated in advisory boards and/or received speaker honoraria from Knight Therapeutics, Gilead and Pfizer. All other authors report no potential conflicts of interest.

## References

1. Wasylyshyn, A.I.; Linder, K.A.; Kauffman, C.A.; Richards, B.J.; Maurer, S.M.; Sheffield, V.M.; Benitez Colon, L.; Miceli, M.H. Invasive Fungal Disease in Patients with Newly Diagnosed Acute Myeloid Leukemia. *J Fungi (Basel)* **2021**, *7*, 761. <https://doi.org/10.3390/jof7090761>.
2. Alkan, A.; Buyukasik, Y.; Uzun, O.; Demir, A.U.; Coplu, L. Invasive fungal infections in patients with acute leukemia: A retrospective cohort study at a tertiary-care hospital. *Medicine (Baltimore)* **2024**, *103*, 39959. <https://doi.org/10.1097/MD.00000000000039959>.
3. Girmenia, C.; Raiola, A.M.; Piocchetti, A.; Algarotti, A.; Stanzani, M.; Cudillo, L.; Pecoraro, C.; Guidi, S.; Iori, A.P.; Montante, B.; et al. Incidence and outcome of invasive fungal diseases after allogeneic stem cell transplantation: a prospective study of the Gruppo Italiano Trapianto Midollo Osseo (GITMO). *Biol Blood Marrow Transplant.* **2014**, *20*, 872-80. <https://doi.org/10.1016/j.bbmt.2014.03.004>.
4. Menzin, J.; Meyers, J.L.; Friedman, M.; Korn, J.R.; Perfect, J.R.; Langston, A.A.; Danna, R.P.; Papadopoulos, G. The economic costs to United States hospitals of invasive fungal infections in transplant patients. *Am J Infect Control* **2011**, *39*, 15-20. <https://doi.org/10.1016/j.ajic.2010.06.009>.
5. Busca, A.; Passera, R.; Maffini, E.; Festuccia, M.; Brunello, L.; Dellacasa, C.M.; Aydin, S.; Frairia, C.; Manetta, S.; et al. Hematopoietic cell transplantation comorbidity index and risk of developing invasive fungal infections after allografting. *Bone Marrow Transplant.* **2018**, *53*, 1304-1310. <https://doi.org/10.1038/s41409-018-0161-1>.
6. Kontoyiannis, D.P.; Marr, K.A.; Park, B.J.; Alexander, B.D.; Anaissie, E.J.; Walsh, T.J.; Ito, J.; Andes, D.R.; Baddley JW, J.W.; Brown, J.M., et al. Prospective surveillance for invasive fungal infections in hematopoietic stem cell transplant recipients, 2001-2006: overview of the Transplant-Associated Infection

Surveillance Network (TRANSNET) Database. *Clin Infect Dis.* **2010**, *50*, 1091-100.  
<https://doi.org/10.1086/651263>.

- 7. Neofytos, D.; Horn, D.; Anaissie, E.; Steinbach, W.; Olyaei, A.; Fishman, J.; Pfaller, M.; Chang, C.; Webster, K.; Marr, K. Epidemiology and outcome of invasive fungal infection in adult hematopoietic stem cell transplant recipients: analysis of Multicenter Prospective Antifungal Therapy (PATH) Alliance registry. *Clin Infect Dis.* **2009**, *48*, 265-73. <https://doi.org/10.1086/595846>.
- 8. Roth, R.S.; Masouridi-Levrat, S.; Chalandon, Y.; Mamez, A. C.; Giannotti, F.; Riat, A.; Fischer, A.; Poncet, A.; Glampedakis, E.; Van Delden, C.; et al. Invasive Mold Infections in Allogeneic Hematopoietic Cell Transplant Recipients in 2020: Have We Made Enough Progress? *Open Forum Infect Dis.* **2021**, *9*, ofab596. <https://doi.org/10.1093/ofid/ofab596>.
- 9. Robenshtok, E.; Gafter-Gvili, A.; Goldberg, E.; Weinberger, M.; Yeshurun, M.; Leibovici, L.; Paul, M. Antifungal prophylaxis in cancer patients after chemotherapy or hematopoietic stem-cell transplantation: systematic review and meta-analysis. *J Clin Oncol.* **2007**, *25*, 5471-89.  
<https://doi.org/10.1200/JCO.2007.12.3851>.
- 10. Zeng, H.; Wu, Z.; Yu, B.; Wang, B.; Wu, C.; Wu, J.; Lai, J.; Gao, X.; Chen, J. Network meta-analysis of triazole, polyene, and echinocandin antifungal agents in invasive fungal infection prophylaxis in patients with hematological malignancies. *BMC Cancer.* **2021**, *21*, 404. <https://doi.org/10.1186/s12885-021-07973-8>.
- 11. Young, J. H.; Andes, D. R.; Ardura, M.I.; Arrieta, A.; Bow, E. J.; Chandrasekar, P.H.; Chen, SCA; Hammond, S.P.; Husain, S.; Koo, S.; et al. Modeling Invasive Aspergillosis Risk for the Application of Prophylaxis Strategies. *Open Forum Infect Dis.* **2024**, *11*, ofae082. <https://doi.org/10.1093/ofid/ofae082>.
- 12. Ullmann, A.J.; Aguado, J.M.; Arikan-Akdagli, S.; Denning, D.W.; Groll, A.H.; Lagrou, K.; Lass-Flörl, C.; Lewis, R.E.; Munoz, P.; Verweij, P.E.; et al. Diagnosis and management of Aspergillus diseases: executive summary of the 2017 ESCMID-ECMM-ERS guideline. *Clin Microbiol Infect.* **2018**, *24*, 1-38.  
<https://doi.org/10.1016/j.cmi.2018.01.002>.
- 13. Dadwal, S.S.; Hohl, T.M.; Fisher, C.E.; Boeckh, M.; Papanicolaou, G.; Carpenter, P.A.; Fisher, B.T.; Slavin, M.A.; Kontoyiannis, D.P. American Society of Transplantation and Cellular Therapy Series, 2: Management and Prevention of Aspergillosis in Hematopoietic Cell Transplantation Recipients. *Transplant Cell Ther.* **2021**, *27*, 201-211. <https://doi.org/10.1016/j.jtct.2020.10.003>.
- 14. Stemler, J.; Mellinghoff, S.C; Khodamoradi, Y.; Sprute, R.; Classen, A.Y.; Zapke, S.E.; Hoenigl, M.; Krause, R.; Schmidt-Hieber, M.; Heinz, W.J.; et al. Primary prophylaxis of invasive fungal diseases in patients with haematological malignancies: 2022 update of the recommendations of the Infectious Diseases Working Party (AGIHO) of the German Society for Haematology and Medical Oncology (DGHO). *J Antimicrob Chemother.* **2023**, *78*, 1813-1826. <https://doi.org/10.1093/jac/dkad143>.
- 15. Patterson, T. F.; Thompson, G.R. 3<sup>rd</sup>; Denning, D. W.; Fishman, J. A.; Hadley, S.; Herbrecht, R.; Kontoyiannis, D. P.; Marr, K. A.; Morrison, V. A.; Nguyen, M. H.; et al. Practice Guidelines for the Diagnosis and Management of Aspergillosis: 2016 Update by the Infectious Diseases Society of America. *Clin Infect Dis.* **2016**, *63*, e1-e60. <https://doi.org/10.1093/cid/ciw326>.
- 16. Pagano, L.; Maschmeyer, G.; Lamoth, F.; Blennow, O.; Xhaard, A.; Spadea, M.; Busca, A.; Cordonnier, C.; Maertens, J. Primary antifungal prophylaxis in hematological malignancies. Updated clinical practice guidelines by the European Conference on Infections in Leukemia (ECIL). *Leukemia.* **2025**, *39*, 1547-1557. <https://doi.org/10.1038/s41375-025-02586-7>.
- 17. Boutin, C.A.; Durocher, F.; Beauchemin, S.; Ziegler, D.; Abou Chakra, C.N.; Dufresne, S.F. Breakthrough Invasive Fungal Infections in Patients With High-Risk Hematological Disorders Receiving Voriconazole

and Posaconazole Prophylaxis: A Systematic Review. *Clin Infect Dis.* **2024**, *79*, 151-160. <https://doi.org/10.1093/cid/ciae203>.

- 18. Ishida, K.; Haraguchi, M.; Kimura, M.; Araoka, H; Natori, A; Reynolds, J.M.; Raja, M; Natori, Y. Incidence of Breakthrough Fungal Infections in Patients With Isavuconazole Prophylaxis: A Systematic Review and Meta-analysis. *Open Forum Infect Dis.* **2025**, *12*, ofaf163. <https://doi.org/10.1093/ofid/ofaf163>.
- 19. Lionakis, M.S.; Lewis, R.E.; Kontoyiannis, D.P. Breakthrough Invasive Mold Infections in the Hematology Patient: Current Concepts and Future Directions. *Clin Infect Dis.* **2018**, *67*, 1621-1630. <https://doi.org/10.1093/cid/ciy473>.
- 20. Girmenia, C.; Busca, A.; Candoni, A.; Cesaro, S., Luppi, M.; Nosari, A.M.; Pagano, L.; Rossi, G.; Venditti, A.; Aversa, F. Breakthrough invasive fungal diseases in acute myeloid leukemia patients receiving mould active triazole primary prophylaxis after intensive chemotherapy: An Italian consensus agreement on definitions and management. *Med Mycol.* **2019**, *57*, S127-S137. <https://doi.org/10.1093/mmy/myy091>.
- 21. Maertens, J.A.; Raad, I.I.; Marr, K.A.; Patterson, T.F.; Kontoyiannis, D.P.; Cornely, O.A., Bow, E.J.; Rahav, G.; Neofytos, D.; Aoun, M. et al. Isavuconazole versus voriconazole for primary treatment of invasive mould disease caused by Aspergillus and other filamentous fungi (SECURE): a phase 3, randomised-controlled, non-inferiority trial. *Lancet.* **2016**, *387*, 760-9. [https://doi.org/10.1016/S0140-6736\(15\)01159-9](https://doi.org/10.1016/S0140-6736(15)01159-9).
- 22. Marty, F.M.; Ostrosky-Zeichner, L.; Cornely, O.A.; Mullane, K.M.; Perfect, J.R., Thompson, GR 3<sup>rd</sup>; Alangaden, G.J.; Brown, J.M.; Fredricks, D.N.; Heinz, W.J.; et al. Isavuconazole treatment for mucormycosis: a single-arm open-label trial and case-control analysis. *Lancet Infect Dis.* **2016**, *16*, 828-837. [https://doi.org/10.1016/S1473-3099\(16\)00071-2](https://doi.org/10.1016/S1473-3099(16)00071-2).
- 23. EMA (2015). Cresemba (isavuconazole). European Medicines Agency. EPAR summary for the public. Available online: <https://www.ema.europa.eu/en/medicines/human/EPAR/cresemba>
- 24. FDA (2015). FDA approves Cresemba (isavuconazonium sulfate) for invasive aspergillosis and invasive mucormycosis. FDA (2024). Drugs@FDA: FDA-Approved Drugs. Application Number: 207500. Available online: <https://www.accessdata.fda.gov/scripts/cder/daf/index.cfm?event=overview.process&AppN=207500>
- 25. Cornely, O.A.; Alastruey-Izquierdo, A.; Arenz, D., Chen, S.C.A.; Dannaoui, E.; Hochhegger, B.; Hoenigl, M.; Jensen, H.E.; Lagrou, K.; Lewis, R.E.; et al. Global guideline for the diagnosis and management of mucormycosis: an initiative of the European Confederation of Medical Mycology in cooperation with the Mycoses Study Group Education and Research Consortium. *Lancet Infect Dis.* **2019**, *19*, 405-421. [https://doi.org/10.1016/S1473-3099\(19\)30312-3](https://doi.org/10.1016/S1473-3099(19)30312-3).
- 26. Hassouna, H.; Athans, V.; Brizendine, K.D. Real-world use-Isavuconazole at a large academic medical center. *Mycoses* **2019**, *62*, 534-541. <https://doi.org/10.1111/myc.12910>.
- 27. Gow-Lee, V.; Abu Saleh, O.M.; Harris, C.E.; Gile, J.J.; Akhiyat, N.; Chedachai, S. Outcomes of Invasive Fungal Infections Treated with Isavuconazole: A Retrospective Review. *Pathogens* **2024**, *13*, 886. <https://doi.org/10.3390/pathogens13100886>.
- 28. Gunathilaka, S.S., Keragala, R.K.; Gunathilaka, K.M.; Wickramage, S.; Bandara, S.R.; Senevirathne, I.S.; Jayaweera, A.S. Use of isavuconazole in mucormycosis: a systematic review. *BMC Infect Dis.* **2025**, *25*, 25. <https://doi.org/10.1186/s12879-025-10439-y>.
- 29. Dagher, H.; Hachem, R.; Chaftari, A.M.; Jiang, Y.; Ali, S.; Deeba, R.; Shah, S.; Raad, I. Real-World Use of Isavuconazole as Primary Therapy for Invasive Fungal Infections in High-Risk Patients with Hematologic Malignancy or Stem Cell Transplant. *J Fungi (Basel)* **2022**, *8*, 74. <https://doi.org/10.3390/jof8010074>.

30. Weng, J.; Du, X.; Fang, B.; Li, Y.; Huang, L.; Ju, Y. Efficacy and safety of isavuconazole versus voriconazole for the treatment of invasive fungal infections: a meta-analysis with trial sequential analysis. *BMC Infect Dis.* **2025**, *25*, 230. <https://doi.org/10.1186/s12879-025-10627-w>.

31. Ellsworth, M.; Ostrosky-Zeichner, L. Isavuconazole: Mechanism of Action, Clinical Efficacy, and Resistance. *J Fungi (Basel)* **2020**, *6*, 324. <https://doi.org/10.3390/jof6040324>.

32. Lewis, J.S. 2<sup>nd</sup>; Wiederhold, N.P.; Hakki, M.; Thompson, G.R. 3<sup>rd</sup>. New Perspectives on Antimicrobial Agents: Isavuconazole. *Antimicrob Agents Chemother.* **2022**, *66*, 0017722. <https://doi.org/10.1128/aac.00177-22>.

33. Andes, D.; Kovanda, L.; Desai, A.; Kitt, T.; Zhao, M.; Walsh, T.J. Isavuconazole Concentration in Real-World Practice: Consistency with Results from Clinical Trials. *Antimicrob Agents Chemother.* **2018**, *62*, 00585-18. <https://doi.org/10.1128/AAC.00585-18>.

34. Risum, M.; Vestergaard, M.B.; Weinreich, U.M.; Helleberg, M.; Vissing, N.H.; Jørgensen, R. Therapeutic Drug Monitoring of Isavuconazole: Serum Concentration Variability and Success Rates for Reaching Target in Comparison with Voriconazole. *Antibiotics (Basel)* **2021**, *10*, 487. <https://doi.org/10.3390/antibiotics10050487>.

35. Lewis, R.; Niazi-Ali, S.; McIvor, A.; Kanj, S.S.; Maertens, J.; Bassetti, M.; Levine, D.; Groll, A.H.; Denning, D.W. Triazole antifungal drug interactions-practical considerations for excellent prescribing. *J Antimicrob Chemother.* **2024**, *79*, 1203-1217. <https://doi.org/10.1093/jac/dkae103>.

36. DiPippo, A.J.; Rausch, C.R.; Kontoyiannis, D.P. Tolerability of isavuconazole after posaconazole toxicity in leukaemia patients. *Mycoses* **2019**, *62*, 81-86. <https://doi.org/10.1111/myc.12851>.

37. Donnelly, J.P.; Chen, S.C.; Kauffman, C.A.; Steinbach, W.J.; Baddley, J.W.; Verweij, P.E.; Clancy, C.J.; Wingard, J.R.; Lockhart, S.R.; Groll, A.H.; et al. Revision and Update of the Consensus Definitions of Invasive Fungal Disease From the European Organization for Research and Treatment of Cancer and the Mycoses Study Group Education and Research Consortium. *Clin Infect Dis.* **2020**, *71*, 1367-1376. <https://doi.org/10.1093/cid/ciz1008>.

38. Cornely, O.A.; Hoenigl, M.; Lass-Flörl, C.; Chen, S.C.; Kontoyiannis, D.P.; Morrissey, C.O.; Thompson, G.R. 3<sup>rd</sup>. Mycoses Study Group Education and Research Consortium (MSG-ERC) and the European Confederation of Medical Mycology (ECMM). Defining breakthrough invasive fungal infection-Position paper of the mycoses study group education and research consortium and the European Confederation of Medical Mycology. *Mycoses* **2019**, *62*, 716-729. <https://doi.org/10.1111/myc.12960>.

39. Przepiorka, D.; Weisdorf, D.; Martin, P.; Klingemann, H.G.; Beatty, P.; Hows, J.; Thomas, E.D. 1994 Consensus Conference on Acute GVHD Grading. *Bone Marrow Transplant.* **1995**, *15*, 825-8.

40. Pagano, L.; Busca, A.; Candoni, A.; Cattaneo, C.; Cesaro, S.; Fanci, R.; Nadali, G.; Potenza, L.; Russo, D.; Tumbarello, M.; et al. Risk stratification for invasive fungal infections in patients with hematological malignancies: SEIFEM recommendations. *Blood Rev.* **2017**, *31*, 17-29. <https://doi.org/10.1016/j.blre.2016.09.002>.

41. Batista, M. V.; Ussetti, M. P.; Jiang, Y.; Neofytos, D.; Cortez, A. C.; Feriani, D.; Schmidt-Filho, J.; França-Silva, I. L. A.; Raad, I.; Hachem, R. Comparing the Real-World Use of Isavuconazole to Other Anti-Fungal Therapy for Invasive Fungal Infections in Patients with and without Underlying Disparities: A Multi-Center Retrospective Study. *J Fungi (Basel)* **2023**, *9*, 166. <https://doi.org/10.3390/jof9020166>.

42. Puerta-Alcalde, P.; Monzó-Gallo, P.; Aguilar-Guisado, M.; Ramos, J.C.; Laporte-Amargós, J.; Machado, M.; Martin-Davila, P.; Franch-Sarto, M.; Sánchez-Romero, I.; Badiola, J.; et al. Breakthrough invasive fungal infection among patients with hematologic malignancies: A national, prospective, and multicentre study. *J Infect.* **2023**, *87*, 46-53. <https://doi.org/10.1016/j.jinf.2023.05.005>.

43. Eigl, S.; Prattes, J.; Reinwald, M.; Thornton, C.R.; Reischies, F.; Sess, B.; Neumeister, P.; Zollner-Schwetz, I.; Raggam, R.B.; Flick, H.; Buchheidt, D.; et al. Influence of mould-active antifungal treatment on the performance of the *Aspergillus*-specific bronchoalveolar lavage fluid lateral-flow device test. *Int J Antimicrob Agents* **2015**, *46*, 401-5. <https://doi.org/10.1016/j.ijantimicag.2015.05.017>.

44. Eigl, S.; Hoenigl, M.; Spiess, B.; Heldt, S.; Prattes, J.; Neumeister, P.; Wolfler, A.; Rabensteiner, J.; Prueller, F.; Krause, R.; et al. Galactomannan testing and *Aspergillus* PCR in same-day bronchoalveolar lavage and blood samples for diagnosis of invasive aspergillosis. *Med Mycol.* **2017**, *55*, 528-534. <https://doi.org/10.1093/mmy/myw102>.

45. Jenks, J.D.; Gangneux, J.P.; Schwartz, I.S.; Alastruey-Izquierdo, A.; Lagrou, K.; Thompson Iii, G.R.; Lass-Flörl, C.; Hoenigl, M. European Confederation of Medical Mycology (ECMM) Council Investigators. Diagnosis of Breakthrough Fungal Infections in the Clinical Mycology Laboratory: An ECMM Consensus Statement. *J Fungi (Basel)* **2020**, *6*, 216. <https://doi.org/10.3390/jof6040216>.

46. Ruhnke, M.; Cornely, O.A.; Schmidt-Hieber, M.; Alakel, N.; Boell, B.; Buchheidt, D.; Christopeit, M.; Hasenkamp, J.; Heinz, W.J.; Henrich, M.; et al. Treatment of invasive fungal diseases in cancer patients- Revised 2019 Recommendations of the Infectious Diseases Working Party (AGIHO) of the German Society of Hematology and Oncology (DGHO). *Mycoses* **2020**, *63*, 653-682. <https://doi.org/10.1111/myc.13082>. Epub 2020 May 12.

47. Maertens, J.A.; Rahav, G.; Lee, D.G.; Ponce-de-León, A.; Ramírez Sánchez, I.C.; Klimko, N.; Sonet, A.; Haider, S.; Diego Vélez, J.; Raad I.; et al. study investigators. Posaconazole versus voriconazole for primary treatment of invasive aspergillosis: a phase 3, randomised, controlled, non-inferiority trial. *Lancet* **2021**, *397*, 499-509. [https://doi.org/10.1016/S0140-6736\(21\)00219-1](https://doi.org/10.1016/S0140-6736(21)00219-1).

48. Verweij, P.E.; Chowdhary, A.; Melchers, W.J.; Meis, J.F. Azole Resistance in *Aspergillus fumigatus*: Can We Retain the Clinical Use of Mold-Active Antifungal Azoles? *Clin Infect Dis.* **2016**, *62*, 362-8. <https://doi.org/10.1093/cid/civ885>.

49. Tang, L.A.; Marini, B.L.; Benitez, L.; Nagel, J.L.; Miceli, M.; Berglund, C.; Perissinotti, A.J. Risk factors for subtherapeutic levels of posaconazole tablet. *J Antimicrob Chemother.* **2017**, *72*, 2902-2905. <https://doi.org/10.1093/jac/dkx228>.

50. Kovanda, L.L.; Marty, F.M.; Maertens, J.; Desai, A.V.; Lademacher, C.; Engelhardt, M.; Lu, Q.; Hope, W.W.; Impact of Mucositis on Absorption and Systemic Drug Exposure of Isavuconazole. *Antimicrob Agents Chemother.* **2017**, *61*, e00101-17. <https://doi.org/10.1128/AAC.00101-17>.

51. Stern, A.; Su, Y.; Lee, Y.J.; Seo, S.; Shaffer, B.; Tamari, R.; Gyurkocza, B.; Barker, J.; Bogler, Y.; Giralt, S.; et al. A Single-Center, Open-Label Trial of Isavuconazole Prophylaxis against Invasive Fungal Infection in Patients Undergoing Allogeneic Hematopoietic Cell Transplantation. *Biol Blood Marrow Transplant.* **2020**, *26*, 1195-1202. <https://doi.org/10.1016/j.bbmt.2020.02.009>.

52. Greene, R.E.; Schlamm, H.T.; Oestmann, J.W.; Stark, P.; Durand, C.; Lortholary, O.; Wingard, J.R.; Herbrecht, R.; Ribaud, P.; Patterson, T.F.; et al. Imaging findings in acute invasive pulmonary aspergillosis: clinical significance of the halo sign. *Clin Infect Dis.* **2007**, *44*, 373-9. <https://doi.org/10.1086/509917>.

53. Aguado, J.M.; Vázquez, L.; Fernández-Ruiz, M.; Villaescusa, T.; Ruiz-Camps, I.; Barba, P.; Silva, J.T.; Batlle, M.; Solano, C.; Gallardo, D. PCRAGA Study Group; Spanish Stem Cell Transplantation Group; Study Group of Medical Mycology of the Spanish Society of Clinical Microbiology and Infectious Diseases; Spanish Network for Research in Infectious Diseases. Serum galactomannan versus a combination of galactomannan and polymerase chain reaction-based *Aspergillus* DNA detection for early

therapy of invasive aspergillosis in high-risk hematological patients: a randomized controlled trial. *Clin Infect Dis.* **2015**, *60*, 405–14. <https://doi.org/10.1093/cid/ciu833>.

54. Kontoyiannis, D.P.; Azie, N.; Franks, B.; Horn, D. L. Prospective antifungal therapy (PATH) alliance(®): focus on mucormycosis. *Mycoses* **2014**, *57*, 240–6. <https://doi.org/10.1111/myc.12149>.

55. Desai, A.V.; Kovanda, L.L.; Hope, W.W.; Andes, D.; Mouton, J.W.; Kowalski, D.L.; Townsend, R.W.; Mujais, S.; Bonate, P.L. Exposure-Response Relationships for Isavuconazole in Patients with Invasive Aspergillosis and Other Filamentous Fungi. *Antimicrob Agents Chemother.* **2017**, *61*, e01034–17. <https://doi.org/10.1128/AAC.01034-17>.

**Disclaimer/Publisher's Note:** The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.