

Review

Not peer-reviewed version

Food Hypersensitivity: Distinguishing Allergy from Intolerance, Main Characteristics, and Symptoms; A Systematic Review

[Gregory Marcel Hage](#) , [Yonna Sacre](#) , [Marcel Anoir Hajj](#) , [Joanne Antoine Haddad](#) , [Lea Nicole Sayegh](#) , [Nicole Fakhoury Sayegh](#) *

Posted Date: 9 April 2025

doi: 10.20944/preprints202503.1317.v2

Keywords: food hypersensitivity; food allergy; food intolerance; clinical manifestations; IBS



Preprints.org is a free multidisciplinary platform providing preprint service that is dedicated to making early versions of research outputs permanently available and citable. Preprints posted at Preprints.org appear in Web of Science, Crossref, Google Scholar, Scilit, Europe PMC.

Copyright: This open access article is published under a Creative Commons CC BY 4.0 license, which permit the free download, distribution, and reuse, provided that the author and preprint are cited in any reuse.

Systematic Review

Food Hypersensitivity: Distinguishing Allergy from Intolerance, Main Characteristics, and Symptoms: A Systematic Review

Gregory Hage ^{1,2}, Yonna Sacre ¹, Marcel Hajj ², Joanne Haddad ^{2,3}, Lea Nicole Sayegh ⁴ and Nicole Fakhoury-Sayegh ^{1,5,*}

¹ Department of Nutrition and Food Science, Faculty of Arts and Sciences, Holy Spirit University of Kaslik, P.O. Box 446 Jounieh, Mount Lebanon – LEBANON

² Hajj Medical Center-Medical & Dental Clinics, Naccache, Mount Lebanon, Green Zone A Building 71 Ground Floor-Lebanon

³ Graduate in Dental Surgery, and Student at the Faculty of Dental Medicine, Saint Joseph University of Beirut, Medical Sciences Campus, Damascus Road, P.O.B. 11-5076, Riad Solh Beirut 1107 2180, Lebanon

⁴ Internal medicine resident, Yale New Haven Hospital, 20 York Street, New Haven, CT 06510, United States

⁵ Faculty of Pharmacy, Department of Nutrition, Saint Joseph University of Beirut, Medical Sciences Campus, Damascus Road, P.O.B. 11-5076, Riad Solh Beirut 1107 2180, Lebanon

* Correspondence: nicolaysayegh@usek.edu.lb

Abstract: Food hypersensitivity remains an understudied and overlooked subject globally. It is characterized by adverse reactions to dietary substances potentially triggered by various mechanisms. Food allergy, a subset of food hypersensitivity, denotes an immune response to food proteins categorized into immunoglobulin IgE-mediated or non-IgE-mediated reactions. Conversely, food intolerance, another facet of food hypersensitivity, refers to non-immunological reactions, in which the human body cannot properly digest certain foods or components, leading to gastrointestinal discomfort and other non-immune-related symptoms. The main objective of this study was to determine and differentiate the differences, characteristics, and types of food hypersensitivities. This study involved a comprehensive review of key research from 1990 onward, including review articles, prospective studies, nested case-control studies, and meta-analyses. Recognizing these differences is essential for healthcare professionals to ensure accurate diagnosis, effective management, and improved patient outcomes, while also aiding dietitians in providing optimal nutritional and dietary guidance. In conclusion, there are big differences between the main characteristics such as symptoms, complications, and treatments between allergies, and food intolerances. Commonly reported trigger foods include cow milk, gluten, eggs, nuts, and seafood.

Keywords: food hypersensitivity; food allergy; food intolerance; clinical manifestations; IBS

1. Introduction

Food hypersensitivity is a prevalent condition affecting individuals worldwide, characterized by adverse reactions to specific food components [1]. It encompasses both food allergies and immune-mediated reactions and intolerances (non-immune mediated reactions), which can manifest with a variety of symptoms, ranging from mild discomfort to severe life-threatening reactions [1] such as skin manifestations (rash, dermatitis, rosacea, angioedema), gastrointestinal symptoms (bloating, diarrhea, constipation, gas, reflux, macro and micronutrients malabsorption), and respiratory symptoms (anaphylactic shock, shortness of breath, nasal congestion, rhinorrhea, sneezing, itching of the nose and throat, coughing, and wheezing) [1].

The prevalence of food hypersensitivity has been steadily increasing, with food allergies affecting approximately 2–10% of the population, varying by region and age group [2]. Studies

indicate that immunoglobulin E (IgE)-mediated food allergies are the most common in children, whereas non-IgE-mediated hypersensitivities and intolerances are more frequently observed in adults [3]. Lactose intolerance, for instance, is a well-documented form of non-immune-mediated hypersensitivity, affecting over 65% of the global population due to lactase enzyme deficiency [4]. Similarly, non-celiac gluten sensitivity has gained recognition as a distinct condition, though its pathophysiology remains under investigation [5].

Distinguishing between food allergy and intolerance is crucial for appropriate management and treatment strategies [6] (Figure 1). This systematic review aimed to elucidate the differences between food allergies and intolerances, including their epidemiology, pathophysiology, clinical manifestations, diagnostic approaches, and management strategies. By understanding the nuances of these conditions, healthcare professionals can provide better care, treatment, prevention, and nutritional management, and improve the quality of life for individuals suffering from food hypersensitivities.

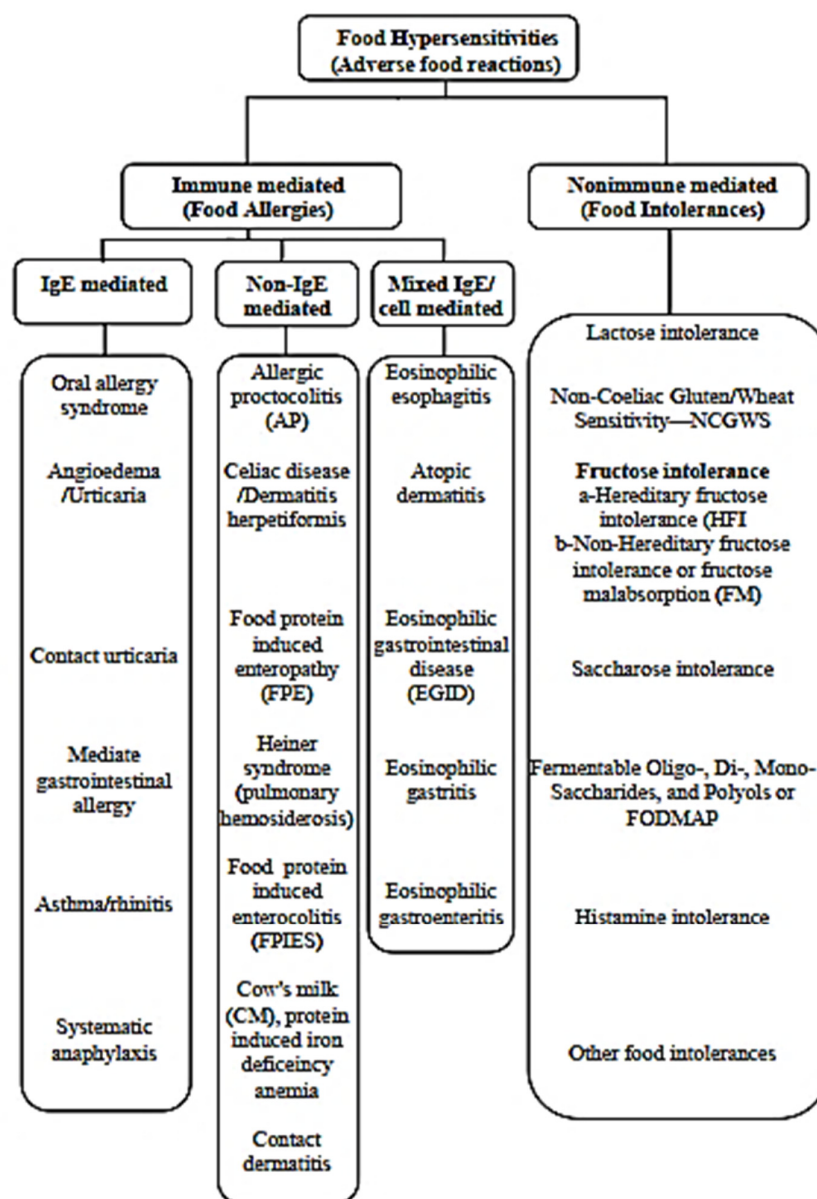


Figure 1. Food hypersensitivities classification, subclassification and infographic [3,4].

2. Material and Methods

This research utilized the Medline database, accessed via its search engine, PubMed, and Google Scholar databases. The following Medical Subject Headings (MeSH) terms or text words were used: “Food hypersensitivity” or “food allergies” or “Food intolerances”

Studies included in this review focused on human studies published in English, with publication dates from 1990 onwards. Specifically, we prioritized prospective studies, meta-analyses, systematic reviews, narrative reviews, and nested case-control studies.

Studies excluded: Studies published before 1990, animal studies, non-English language articles.

For precision purposes, two researchers reviewed all relevant articles independently to check for any discrepancies and to ensure rigorous data extraction. For each article, we focused on the key study characteristics, including publication year, country of origin, study design, sample size, and participant characteristics.

This systematic review followed the PRISMA (Preferred Reporting Items for Systematic Reviews and Meta-Analyses) guidelines to ensure a transparent and rigorous reporting process [9]. The PRISMA framework was used to structure the identification, screening, eligibility, and inclusion of relevant studies.

1,534 records were identified through database searches. After removing 321 duplicates, 1,213 records remained for screening. Of these, 954 records were excluded for not meeting inclusion criteria, leaving 259 full-text articles for eligibility assessment. This resulted in 152 studies being included in the final narrative synthesis.

Details of the study selection process are illustrated in Figure 2.

The review is divided into 3 sections, highlighting the 2 main components of Food Hypersensitivity, which are food allergies and food intolerance, with the third part being about the relation of Food Hypersensitivity with IBS and the role of microbiota.

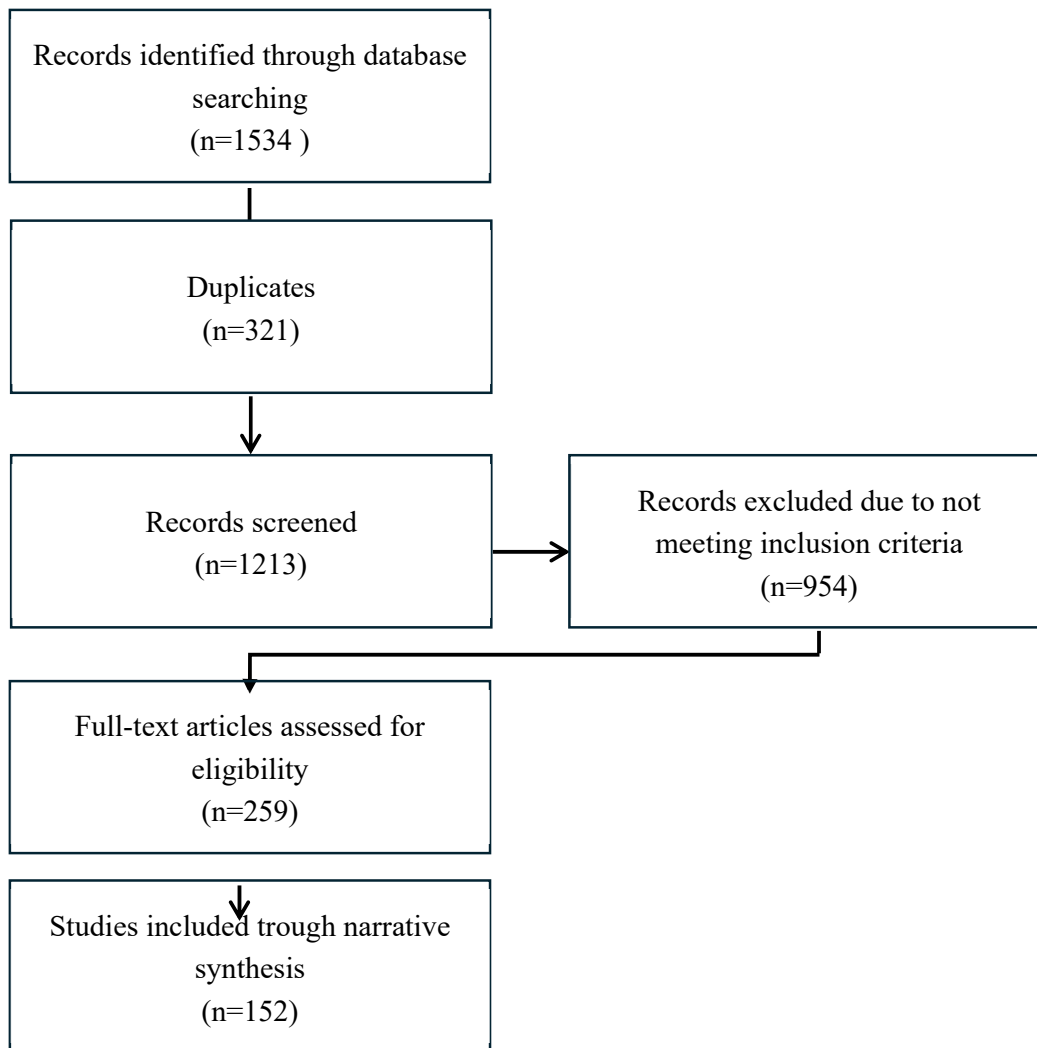


Figure 2. Flowchart of studies selection.

3. Results

3.1. Food Allergies

Table 1. Main characteristics of included studies.

Author	Year	Study Type	Country	Sample Size	Duration of Intervention	Summary of Findings
Vassilopoulou et al. [10]	2022	Retrospective, observational, multicenter case-control study	Greece	96 mothers of infants with and 141 mothers of infants without a history of Food Protein-Induced Allergic Proctocolitis (FPIAP).	From May 2018 to November 2020	Identified cow milk (83%), eggs (7.3%), wheat (6.4%), and beef (6.4%) as the main triggers for allergic proctocolitis (AP) in infants through the maternal diet.

Ruffner et al. [11]	2013	Retrospective chart review	USA	462 cases were identified from the hospital patients	From 2007 until 2012	Food Protein-Induced Enterocolitis Syndrome (FPIES) reactions were observed more frequently than previously reported, though their presentation and clinical characteristics remained consistent with earlier findings. Milk- and soy-induced FPIES were prevalent, with 43.5% of patients who reacted to milk also experiencing a reaction to soy.
Pinto-Sánchez et al. [12]	2021	Prospective study	Canada	prospective study of 50 patients with Irritable Bowel Syndrome (IBS) (ROME III, all subtypes), with and without serologic reactivity to gluten (antigliadin IgG and IgA), and 25 healthy subjects (controls)	Between 2012 and 2016	Evaluated the effectiveness of a gluten-free diet in achieving mucosal healing for celiac patients.
Ford et al. [13]	2014	Cross Sectional	Canada	4224 patients recruited	Between January 2008 and December 2014	Functional bowel disorders (FBDs) showed significant demographic and psychological

						<p>differences among patients. The Rome III classification system did not clearly distinguish between different FBD subtypes. There was considerable symptom overlap among irritable Bowel syndrome (IBS), functional diarrhea, and chronic idiopathic constipation (CIC). The findings suggest a need for improved diagnostic criteria to differentiate FBDs more effectively.</p>
Schink et al. [14]	2018	Cross-sectional observational study	Germany	64 participants 8 with histamine intolerance (HIT), 25 with food hypersensitivity (FH), 21 with food allergy and 10 healthy controls (HC)	12 months	Suggested dietary modifications and DAO supplements for histamine intolerance.
Halmos et al. [15]	2014	Randomized, controlled, cross-over trial	Australia	30 patients with IBS and 8 healthy individuals (controls, matched for demographics and diet)	Between April 2009 and June 2011	Confirmed the efficacy of the low FODMAP diet in IBS symptom reduction.

Nwaru et al. [16]	2014	Systematic review and meta-analysis	Europe	Not Applicable	Between 1 January 2000 and 30 September 2012	Highlighted that early introduction of allergenic foods may reduce the risk of developing IgE-mediated food allergies.
West et al. [17]	2014	Observational population-based study	UK	57 million	Between 1990 and 2011	Found that the incidence of celiac disease is increasing, estimating 19.1 per 100,000 cases annually.

3.1.1. Definition

A food allergy, categorized under food hypersensitivity, refers to an immunological reaction to food proteins [18]. This reaction can involve immunoglobulin IgE-mediated responses, mixed IgE and non-IgE mediated reactions, or non-IgE-mediated responses [4,5]. IgE-mediated food allergies affect millions worldwide, significantly impacting individuals' daily lives [6]. The prevalence of food allergies is estimated to affect 10-25% of the global population, with an upward trend observed over the last two decades [19]. The classification of food allergies is summarized in the Figure 1 [7].

3.1.2. Symptoms

Symptoms may affect various systems including the respiratory tract, the gastrointestinal tract, skin, and the cardiovascular system [21]. Respiratory symptoms encompass sneezing, congestion, rhinorrhea, wheezing, and laryngeal edema [21]. Gastrointestinal manifestations consist of nausea, vomiting, abdominal pain, and diarrhea [21]. Cutaneous presentations may include urticaria, angioedema, flushing, or pruritus [21]. Cardiovascular signs may involve tachycardia, hypotension, or syncope [21]. Typically, symptoms manifest within minutes of food ingestion, although they may delay up to 2 hours [21]. The severity ranges from pruritus alone to anaphylactic shock [21]. The symptoms for each food allergy disorder can be found in Table 2 [7].

Table 2. Food allergies disorders and main features.

Pathology	Disorder	Key features	Most common causal foods
IgE-mediated (acute onset)	Acute urticaria/angioedema	Food commonly causes acute (20%) but rarely chronic urticaria	Cow milk, gluten, eggs, wheat, beans, soybean, nuts, and seafood
	Contact urticaria	Direct skin contact results in lesions. Histamine release, in rare cases, can cause urticaria.	Multiple
	Anaphylaxis	Rapidly progressive, multiple organs system reactions can include cardiovascular collapse	Any but more commonly peanut, Tree nuts, shellfish, fish, milk, and egg
	Food-associated, exercise-induced anaphylaxis	Food triggers anaphylaxis only if ingestion is followed temporally by exercise	Wheat, shellfish, and celery are most often described
	Oral allergy syndrome (pollen-associated food allergy syndrome)	Pruritus and mild edema are confined to the oral cavity and uncommonly progress beyond the mouth (w7%) and rarely to anaphylaxis (1% to 2%). It might increase after the pollen season.	Raw fruit/vegetables; cooked forms tolerated; examples of relationships: birch (apple, peach, pear, carrot), ragweed (melons)
	Immediate gastrointestinal hypersensitivity	Immediate vomiting, pain	Cow milk, gluten, eggs, wheat, beans, soybean, nuts, and seafood
Combined IgE and cell-mediated (delayed onset/chronic)	Atopic dermatitis	Associated with food allergy in 35% of children with moderate-to-severe rash	Major allergens, particularly egg, milk
	Eosinophilic esophagitis	Symptoms might include feeding disorders, reflux symptoms, vomiting, dysphagia, and food impaction.	Multiple

	Eosinophilic gastroenteritis	Vary on site(s)/degree of eosinophilic inflammation; might include ascites, weight loss, edema, obstruction	Multiple
Cell-mediated (delayed onset/chronic)	Food protein-induced enterocolitis syndrome Cow's milk, soy, rice, oat, meat	Primarily affects infants; chronic exposure: emesis, diarrhea, poor growth, lethargy; re-exposure after restriction: emesis, diarrhea, hypotension (15%) 2 hours after ingestion	Cow's milk, soy, rice, oat, meat
	Food protein induced allergic proctocolitis	Mucus-laden, bloody stools in infants Milk (through breast-feeding)	Milk (through breast-feeding)
	Allergic contact dermatitis	Often occupational because of chemical moieties, oleoresins. Systemic contact dermatitis is a rare variant because of ingestion	Spices, fruits, vegetables
	Heiner syndrome	Pulmonary infiltrates, failure to thrive, iron deficiency anemia	Cow's milk

3.1.3. IgE-Mediated Food Allergies

Regarding IgE-mediated food allergies, they are categorized as type I hypersensitivity reactions. Symptoms typically manifest rapidly, occurring within minutes to a few hours after ingestion. Diagnosis involves a detailed assessment of clinical symptoms and various testing methods, including oral food challenges, food allergy skin prick testing, and specific food IgE testing [20]. Furthermore, IgE-mediated food allergies are characterized by the immune system's production of IgE antibodies in response to specific food proteins, leading to rapid allergic reactions that can range from mild symptoms to anaphylaxis. Several studies have examined the prevalence and mechanisms of these allergies. A study by Sicherer and Sampson, (2016) highlighted that IgE-mediated food allergies are increasing globally, particularly among children, with common allergens including peanuts, tree nuts, and shellfish [22]. Another study by Savage et al., compared different diagnostic approaches, such as oral food challenges and specific IgE testing, noting that while specific IgE testing is useful, it can sometimes result in false positives, making oral food challenges the gold standard for diagnosis [23]. Furthermore, research by Nwaru et al. emphasized that the early introduction of allergenic foods might reduce the risk of developing IgE-mediated food allergies, a finding that has influenced recent guidelines on allergy prevention [16]. These studies collectively underscore the complexity of diagnosing and managing IgE-mediated food allergies and the need for tailored approaches based on individual patient profiles.

Diagnosis methods

The skin prick test (SPT) serves as a common diagnostic tool for identifying type I hypersensitivity reactions. It involves applying an allergen extract onto the skin, typically on the forearm or back, followed by pricking the skin with a lancet [24]. This minimally invasive test offers the advantage of assessing multiple allergens, including inhalants, foods, drugs, venom, latex, and

occupational allergens, within a short timeframe of 15-20 minutes, with a high sensitivity of 95-100% [24].

Skin Prick Testing (SPT) is a widely used method for diagnosing IgE-mediated allergic conditions by exposing the skin to small amounts of suspected allergens and observing for a localized allergic reaction. This method is often compared with other diagnostic approaches like specific IgE (sIgE) testing. A study by Heinzerling et al. found that SPT is highly sensitive, particularly to respiratory allergens such as pollen, mold, and dust mites, and provides rapid results, making it a first-line test in many allergy clinics. However, the study also noted variability in results depending on factors like the allergen extract quality, the technique used, and the patient's skin condition at the time of testing [25].

Another study by Bousquet et al., explored the correlation between SPT results and clinical symptoms and found a strong correlation between positive SPT results and clinical manifestations of allergic rhinitis and asthma, though the strength of the correlation can vary depending on the allergen tested [26].

Despite its advantages, SPT has limitations. For instance, as highlighted by Nelson et al., there can be a risk of false negatives, particularly in cases of recent antihistamine use or improper technique [14]. Additionally, false positives can occur due to irritant reactions rather than true allergic sensitizations [14]. Consequently, while SPT remains a cornerstone in allergy diagnostics, especially in respiratory and food allergies, it is often used in conjunction with sIgE testing and a thorough patient history to confirm diagnoses and guide treatment plans [14]. These studies collectively underscore that while SPT is a valuable and widely used diagnostic tool, its results must be interpreted within the broader clinical context, with consideration given to the method's limitations.

In vitro, total IgE testing measures the overall levels of IgE in the bloodstream and lacks specificity. On the other hand, specific IgE testing targets particular allergens, both food and inhalants, through a blood sample collection from the individual [24]. While specific IgE testing provides high sensitivity and specificity, it requires a longer turnaround time, typically around 10-20 days, to obtain results [27].

Specific IgE (sIgE) testing is a widely used diagnostic tool in allergy testing, designed to detect IgE antibodies specific to particular allergens in a patient's blood. Numerous studies have compared sIgE testing with other diagnostic methods such as skin prick tests (SPT) and component-resolved diagnostics (CRD). For example, Wood et al. found that sIgE testing is particularly sensitive in detecting sensitization to common allergens like pollen, house dust mites, and pet dander [28]. However, the study noted variability in specificity depending on the allergen and the chosen cutoff values [16]. Another study by Matricardi et al., compared sIgE testing to CRD and concluded that CRD offers a more refined identification of allergen sensitization, particularly in distinguishing primary sensitization from cross-reactivity, which is critical in complex allergy cases [29]. Additionally, a study by Zuberbier et al. underscored that while sIgE testing is a powerful tool for detecting allergic sensitizations, it should not be used in isolation due to the potential for false positives, particularly in individuals with high total IgE levels [30]. This analysis highlighted the importance of interpreting sIgE results within the broader context of a patient's clinical history and other diagnostic tests. Ewan and Dugue also discussed the possibility of false positives in sIgE testing, especially in patients with elevated total IgE levels, further emphasizing the need for a comprehensive approach to allergy diagnosis [31]. These studies collectively suggest that while sIgE testing is valuable in allergy diagnostics, it should be integrated with other diagnostic methods to achieve the most accurate results.

Common allergens and their prevalences according to diagnosis techniques

Food allergies are a growing concern globally, with significant variations in prevalence across different populations and age groups. Studies have consistently identified certain foods as the most common triggers of IgE-mediated allergic reactions, including cow milk, gluten, eggs, wheat, beans, soybean, nuts, and seafood as the primary culprits behind food allergies [32]. A landmark study by Sicherer and Sampson in the United States highlighted that peanut allergy, affecting approximately

1-2% of children, is the most prevalent food allergy in Western countries, with a rising incidence over the past two decades [33]. Another critical review by Gupta et al., explored the burden of food allergies across the globe, noting that while peanut and tree nut allergies are particularly prevalent in Western nations, rice and sesame are more common allergens in Asia and the Middle East, [34]. Moreover, the Australian Health Nuts study emphasized the significant role of environmental and genetic factors in the development of food allergies, with peanut and egg allergies being the most common among Australian children, affecting nearly 10% of infants by the age of one [35]. These studies collectively underscore the importance of geographic and demographic factors in determining the prevalence and types of food allergies, and they highlight the need for region-specific public health strategies to manage and prevent food allergies effectively.

3.1.4. Mixed IgE/Non IgE Mediated Food Allergies

Eosinophilic esophagitis

Eosinophilic esophagitis (EoE) is a chronic, immune-mediated esophageal disease characterized by the infiltration of eosinophils in the esophageal epithelium, leading to symptoms such as dysphagia and food impaction [36]. The rising incidence of EoE has prompted numerous studies aimed at understanding its pathophysiology, prevalence, and treatment options. A pivotal study by Dellon et al., highlighted the increasing prevalence of EoE in both pediatric and adult populations in the United States, noting a significant rise in diagnoses over the past two decades [37]. This study underscored the role of environmental factors, such as food allergens and aeroallergens, in the pathogenesis of EoE [37].

Another key study by Liacouras et al. provided a comprehensive review of diagnostic criteria and treatment guidelines for EoE, emphasizing the importance of dietary management, including elimination diets and the use of proton pump inhibitors (PPIs), in managing the condition [38]. The study compared the effectiveness of different therapeutic approaches, including dietary elimination and pharmacologic treatments, concluding that while both strategies are effective, the choice of the treatment should be individualized based on patient characteristics and preferences [38].

Further research by Hirano et al. explored the long-term outcomes of patients with EoE, revealing that while the disease is chronic and relapsing, early diagnosis and consistent treatment can significantly improve quality of life and reduce the risk of complications such as esophageal strictures) [39]. This study also compared the effectiveness of topical corticosteroids, such as fluticasone and budesonide, with dietary interventions, finding that while corticosteroids are effective in reducing inflammation, dietary changes are crucial in managing symptoms long-term) [39].

Finally, a meta-analysis by Lucendo et al. compared the outcomes of various treatments for EoE, including elimination diets, PPIs, and corticosteroids [40]. The study concluded that while elimination diets are effective in inducing histologic remission, PPIs and corticosteroids also play a vital role, particularly in patients who do not respond to dietary changes [40].

These studies collectively highlight the complex and multifactorial nature of EoE, emphasizing the need for individualized treatment plans that consider both dietary and pharmacologic interventions to manage this chronic condition effectively.

Non IgE mediated food allergies

Non-IgE cell-mediated food allergies pose a greater diagnostic challenge and are categorized into various disorders, including Allergic proctocolitis (AP), Celiac disease/dermatitis herpetiformis, food protein-induced enteropathy (FPE), Heiner syndrome (pulmonary hemosiderosis), food protein-induced enterocolitis syndrome (FPIES), and Cow's milk (CM) protein-induced iron deficiency anemia [4,5,7].

These allergies are mostly diagnosed during early childhood, except for celiac disease with the main allergens being identified as either wheat, soy and cow milk. [41].

Allergic Proctocolitis

Formerly recognized as allergic colitis, this benign condition primarily affects young children [41]. It is characterized by the presence of bright red blood in the stool (hematochezia), often accompanied by diarrhea, and has been primarily linked to the consumption of cow and soy milk in children [12,13]. Diagnosis of allergic proctocolitis (AP) involves clinical evaluation, laboratory tests, stool examination (fecal calprotectin), endoscopic procedures, and allergy assessments (specific IgE and skin prick testing) [42]. It is marked by mild anemia, elevated eosinophil count (eosinophilia), higher-than-normal total IgE levels, presence of eczema, and hypoalbuminemia [12–14]. Treatment typically entails eliminating the offending food trigger, leading to symptom resolution within a maximum of ninety-six hours [41]. According to Elizur et al. the prevalence of AP in young children was 1.6 per 1000 infants [44].

Another study done by Mennini et al. a review study has found that 0.16% of healthy children and 64% of children suffering from blood in stool are in fact suffering from Allergic proctocolitis [45].

Furthermore, Vassilopoulou et al. has found similar results as Mennini et al. [34] in terms of the prevalence of the disease, with cow milk (83%), eggs (7.3%), wheat (6.4%), and beef (6.4%) causing the symptoms of allergic proctocolitis by the ingestion of these food proteins from the mother's diet, which feeds the infant breastfeeding [35].

A 2019 study by Nowak-Węgrzyn et al. provided a comprehensive overview of the condition, emphasizing that while allergic proctocolitis is typically benign and self-limited, occurring in infants within the first few months of life, it requires careful dietary management to avoid complications [46]. This study stressed the importance of eliminating the offending protein from the maternal diet in breastfeeding infants or switching to a hypoallergenic formula for formula-fed infants [10].

A more recent study by Ruffner et al. explored the long-term outcomes of infants diagnosed with allergic proctocolitis, revealing that most children outgrow the condition by the age of one, though a small percentage may develop other atopic conditions later in life. This study highlighted the need for ongoing monitoring, particularly in infants with a family history of atopic diseases [11].

Further research by Caubet et al. emphasized the role of maternal diet during pregnancy and breastfeeding in the prevention and management of allergic proctocolitis, suggesting that maternal dietary restrictions can significantly reduce the incidence and severity of symptoms in infants at risk [47]. This study provided evidence supporting the early identification and elimination of food allergens as a key strategy in managing allergic proctocolitis effectively [47].

These studies collectively highlight the evolving understanding of allergic proctocolitis, emphasizing the importance of personalized dietary management strategies, ongoing monitoring, and the potential benefits of maternal dietary modifications. They underline the significance of early diagnosis and intervention to prevent long-term complications and ensure optimal growth and development in affected infants.

Celiac disease/dermatitis herpetiformis

Celiac disease (CD) is an immune-mediated disorder characterized by a permanent immune response triggered by consuming gluten-containing foods such as wheat, barley, and rye [48]. Classified as an enteropathy, it leads to severe malabsorption of several vitamins (D, B12, B6), minerals (iron), and macronutrients due to the atrophy of intestinal villi, primarily in the duodenum, rendering individuals with CD susceptible to various nutritional deficiencies [17,18]. Most of CD patients are reported to carry haplotype HLA DR3-DQ2 and/or DR4-DQ8, which serves as a susceptibility indicator for predicting celiac disease [51]. Endoscopic duodenal intestinal biopsy is considered the gold standard for diagnosing celiac disease [49]. Serologic testing, including tissue transglutaminase (TTG), endomysia antibody (EMA), deamidated gliadin peptide (DGP), and antigliadin antibodies (AGAs), can help in suspecting CD, with AGAs being less specific and gradually replaced by newer tests like EMA, TTG, and DPG [50]. Treatment involves completely eliminating gluten from the diet for life, which resolves intestinal damage [50], and 1.4% of the global population is estimated to be suffering from celiac disease [52].

According to West et Al., the incidence of celiac disease was estimated to be around 19.1 per 100,000 which has been increasing since the early 2000s [17].

A 2020 study by Lebwohl et al., investigated the global prevalence of celiac disease, finding significant geographical variation, with higher rates in Europe and the United States compared to Asia and Africa [53]. This study underscored the role of genetic predisposition and environmental factors in the development of the disease [53].

In contrast, a 2019 study by Ludvigsson et al. focused on the diagnostic approaches, comparing serological tests such as anti-tissue transglutaminase (tTG) antibodies with biopsy findings [54]. The study emphasized the high sensitivity and specificity of tTG testing, making it a cornerstone in the non-invasive diagnosis of celiac disease, though biopsies remain the gold standard for confirmation [54].

A 2021 systematic review by Pinto-Sánchez et al. evaluated various dietary management strategies, particularly the effectiveness of a strict gluten-free diet (GFD) in achieving mucosal healing and reducing symptoms [12]. The review concluded that while most patients benefit from a GFD, there is a subset of patients with non-responsive celiac disease who may require additional interventions, such as the exclusion of trace gluten or refractory celiac disease therapies [12].

Further research by Rubio-Tapia et al. explored the long-term outcomes of patients with celiac disease, particularly focusing on the risks of complications such as enteropathy-associated T-cell lymphoma (EATL) [55]. This study highlighted the importance of early diagnosis and strict adherence to a GFD to reduce the risk of such severe complications [55].

These studies collectively contribute to a nuanced understanding of celiac disease, highlighting the importance of early and accurate diagnosis, effective management through a strict gluten-free diet, and ongoing monitoring to prevent complications and improve quality of life.

Similarly, Dermatitis herpetiformis (DH) is an autoimmune skin condition characterized by the formation of small blisters or papules, rash, and urticaria, typically appearing on the elbows, knees, and buttocks [56]. DH shares a similar genetic background with CD, with some researchers considering it a subtype of CD [57]. The incidence of DH is decreasing globally, while CD is on the rise, with DH more prevalent in males and CD in females [20,21]. Diagnosis of DH primarily involves examining clinical signs and symptoms and confirming them through direct immunofluorescence examination of perilesional skin, revealing granular immunoglobulin A (IgA) in the papillary dermis [20,21]. Patients with DH may also experience gastrointestinal symptoms and nutritional deficiencies related to CD, which resolve upon complete gluten avoidance [20]. Although adopting a gluten-free diet resolves urticaria and blisters, it may take months to years for skin symptoms to completely clear [20,21].

DH's incidence is estimated at 0.8 per 100,000 people [22]. The prevalence is thought to be between 11.2 to 75.3 per 100 000 individuals [58]. Another study conducted by Antiga et al. [59] has shown the same prevalence number in the population while additionally showing that the disease is almost absent in African and Asian populations while being frequent in the Caucasian population. This is mainly due to the absence of the human leukocyte haplotypes (HLA) DQ2 and DQ8 from the African and Asian populations and low wheat consumption in these regions [59].

Reunala et al. confirmed that DH is a specific manifestation of celiac disease, with nearly all patients exhibiting some degree of intestinal involvement, even if asymptomatic [60]. This study highlighted the importance of adhering to a strict gluten-free diet (GFD) as the primary treatment for DH, alleviates the skin symptoms and addresses the underlying intestinal inflammation associated with celiac disease [60].

Further research by Mansikka et al., provided insights into the epidemiology of DH, showing that its incidence has decreased over the past few decades, likely due to better recognition and earlier diagnosis of celiac disease [61]. This population-based study from Finland reported a decrease in the prevalence of DH, which the authors attributed to the widespread adoption of gluten-free diets among individuals with celiac disease and the earlier initiation of treatment before the development of skin symptoms [61].

Additionally, Collin et al. examined the long-term outcomes of DH patients, particularly focusing on the risk of associated autoimmune diseases and malignancies [62]. Their findings suggested that while DH patients on a strict GFD had a similar overall mortality risk as the general population, those who did not strictly adhere to the diet had an increased risk of developing other autoimmune conditions, particularly thyroid disease, and certain cancers, such as non-Hodgkin lymphoma [62].

Moreover, a review by Caproni et al. summarized the immunopathogenesis of DH, emphasizing the role of IgA deposits in the skin and the cross-reactivity between epidermal transglutaminase (the autoantigen in DH) and tissue transglutaminase (the autoantigen in celiac disease) [63]. This understanding has informed the diagnostic approaches, with skin biopsies for direct immunofluorescence being the gold standard for diagnosing DH, and serological tests for IgA antibodies used to support the diagnosis of celiac disease [63].

Celiac disease and DH are nonetheless diseases that can cause many serious health problems if not diagnosed early. Following a lifetime avoidance of gluten is currently the only solution to diseases.

Food protein-induced enteropathy (FPE)

Food protein-induced enteropathy (FPE) is a chronic, non-IgE-mediated gastrointestinal disorder primarily affecting infants, characterized by persistent diarrhea, malabsorption, and failure to thrive due to intolerance to specific dietary proteins, particularly cow's milk and soy [64]. Comparative studies on FPE and related conditions, such as food protein-induced enterocolitis syndrome (FPIES), have highlighted distinct differences in clinical presentation and progression [64]. Jenkins et al. established cow's milk as a predominant trigger in FPE, with symptoms typically emerging in the first few months of life [65]. The review of 462 patients with FPE provided insights into the variable prognosis, where some children outgrow the condition, while others continue to experience symptoms into later childhood [11]. Caubet et al. further underscored the challenge of managing FPE in cases of multiple food sensitivities, stressing the need for personalized dietary interventions [66]. Moreover, Fernandes et al. explored the potential for FPE to persist into adulthood, which remains a growing area of interest [67]. These studies suggest that while FPE shares certain clinical characteristics with other non-IgE-mediated disorders, its management requires a more nuanced approach, particularly in children with complex dietary protein intolerances. The evolving body of research advocates continuous monitoring and reevaluation of dietary strategies as children age, ensuring both symptom control and optimal growth and development.

Heiner syndrome (pulmonary hemosiderosis)

Cow milk has been identified as the primary trigger in Heiner syndrome (pulmonary hemosiderosis), a non-IgE-mediated allergy characterized by pulmonary diseases in young infants [68]. This condition is considered rare and often requires time for diagnosis [69]. It typically presents gastrointestinal symptoms, poor growth, anemia, pulmonary hemosiderosis (PH), and symptoms resembling pulmonary infections, which resolve after removing cow milk from the diet [28]. Diagnosis involves clinical evaluation, laboratory tests (cow milk IgE and IgG antibodies), and radiography [70].

Despite its rarity, Heiner syndrome remains an important differential diagnosis in infants with recurrent pulmonary infections and anemia, especially when symptoms do not respond to conventional treatments [71]. Misdiagnosis is common, as symptoms overlap with chronic lung diseases such as cystic fibrosis and recurrent aspiration pneumonia [72]. This highlights the need for increased awareness among pediatricians and pulmonologists to ensure timely diagnosis and intervention.

While cow milk elimination remains the primary treatment, studies suggest that delayed diagnosis can lead to long-term complications, including chronic lung damage and persistent iron deficiency anemia [73]. This raises concerns about whether early nutritional interventions, such as alternative hypoallergenic formulas or hydrolyzed protein-based diets, should be implemented in high-risk infants to prevent disease progression [74].

Emerging research also suggests that genetic and environmental factors may play a role in the development of Heiner syndrome. Some studies have hypothesized that certain genetic predispositions may influence immune responses to food proteins, leading to an exaggerated inflammatory reaction in the lungs [75]. However, further research is needed to confirm these associations and explore potential biomarkers for early detection.

Overall, while Heiner syndrome is still relatively under-researched, the available studies underscore the importance of awareness and early intervention to prevent long-term complications in affected children. Future studies should focus on identifying risk factors, improving diagnostic accuracy, and evaluating the effectiveness of novel therapeutic strategies, such as immunomodulatory treatments or microbiome-based interventions, to better manage the condition.

Food protein-induced enterocolitis syndrome (FPIES)

Food protein-induced enterocolitis syndrome (FPIES) represents another form of non-IgE-mediated food allergy, characterized by severe delayed gastrointestinal symptoms typically occurring within the first year of life. Symptoms include repeated vomiting, hypotension, blood in the stool, and diarrhea following ingestion of the offending food [76]. Diagnosis of FPIES relies primarily on clinical symptoms, elimination diets, oral food challenges, and assessment of symptom progression in patients [77]. Cow milk is the most common trigger food for this syndrome [7]. According to Katz et al., 0.34% is the current prevalence of the disease in pediatric patients up to 3 years of age with 90% of patients recovering at the age of 3, with the disease being diagnosed in the majority of patients in the 6 first months of life [66].

In Australia, it is estimated that up to 90 patients suffering of FPIES visit allergy clinic for their condition with an estimation of 1 in 10000 Australian infants less than 2 years of age having the disease [78].

A recent study by Ruffner et al. delved into the immune mechanisms underlying FPIES, proposing that T-cell responses may play a pivotal role in its pathogenesis, contrasting the traditional focus on innate immune responses [11]. These studies emphasize the need for tailored approaches in managing FPIES, including individualized elimination diets and careful food reintroduction protocols. Given the severity of FPIES reactions and the potential for misdiagnosis, there is a growing call for the development of standardized diagnostic criteria and improved awareness among healthcare providers [79]. Delayed diagnosis can lead to unnecessary medical interventions and prolonged dietary restrictions, affecting a child's nutritional status and growth [80]. Moreover, research suggests that the gut microbiome may play a role in disease progression, with dysbiosis potentially influencing immune responses to trigger foods [81]. This raises questions about whether probiotic interventions could help modulate immune tolerance in FPIES patients, an area that warrants further investigation.

Furthermore, the variability in triggers and the immune mechanisms involved suggest that a one-size-fits-all approach may not be adequate.

Future research should focus on identifying potential biomarkers for early detection, as well as refining food challenge protocols to minimize risk while ensuring accurate diagnosis. Additionally, the psychological impact on parents and caregivers should not be overlooked, as managing FPIES requires strict vigilance in food selection and preparation, which can contribute to anxiety and stress.

3.2. Food Intolerances

Food intolerances, another facet of food hypersensitivity, refer to non-immunological reactions triggered by a food or food component typically tolerated in certain doses [82]. It is estimated that up to 20% of the global population experiences food intolerance [33]. However, diagnosing this condition often requires understanding various clinical presentations, including the intensity and timing of symptom onset. Complicating matters further are the diverse modes of action of food intolerance, which may include pharmacological effects (such as with coffee), enzyme deficiencies (like lactose malabsorption), and nonspecific gastrointestinal functioning [33]. Lactose, gluten/wheat, histamine-rich foods, and FODMAPs are among the most commonly implicated triggers for food

intolerances [8]. Gastrointestinal symptoms like bloating, gas, diarrhea, abdominal pain, or nausea are typical presentations of food intolerance, with life-threatening reactions being rare [7].

3.2.1. Lactose Intolerance

Lactose intolerance (LI) occurs when individuals with lactose malabsorption (LM) experience symptoms like diarrhea, bloating, nausea, and abdominal pain after consuming lactose-containing foods [83]. It is estimated to affect between 57 to 65 percent of the global population [84]. Lactose, a disaccharide sugar found in most dairy products, is broken down into glucose and galactose by the enzyme lactase [8]. LM is a prerequisite for LI and can stem from various causes, including primary lactase deficiency (a gradual decline in lactase levels as individuals age) [85], secondary lactase deficiency (resulting from intestinal epithelium injury due to conditions like AIDS, chemotherapy, or gastrointestinal infections, which is reversible upon treatment of the underlying cause) [86], congenital lactase deficiency (a rare pediatric genetic disorder characterized by severe symptoms and failure to thrive) [87], and developmental lactase deficiency (occurring in premature infants with immature gastrointestinal systems) [8]. Diagnosis of LM typically relies on non-invasive methods like hydrogen breath tests [88], although other approaches such as genetic testing and enzymatic assays exist, with enzymatic assay measurements in bowel biopsies considered the gold standard [41,47,48]. Treatment options for LM include avoiding lactose-containing foods, using oral lactase enzyme replacements, and incorporating probiotics like *Lactobacillus* spp., *Bifidobacterium longum*, or *Bifidobacterium* into the diet. These probiotics have been demonstrated to stimulate the production of lactase when consumed [8].

A pivotal study by Swallow identified genetic polymorphisms associated with lactase persistence in populations of European descent, shedding light on the evolutionary aspects of lactose tolerance [91]. The study demonstrated that the ability to digest lactose into adulthood is primarily due to genetic mutations that allowed for the persistence of lactase production beyond infancy [91]. This genetic advantage likely provided a nutritional benefit in pastoral societies, where dairy was a major food source [91].

Further exploring dietary management, a study by Shaukat et al. systematically reviewed the efficacy of different interventions for lactose intolerance [92]. The review found that while lactase enzyme supplements can help reduce symptoms, many individuals benefit from gradually introducing small amounts of dairy into their diet to build tolerance [92]. This approach, known as lactose adaptation, takes advantage of the fact that some individuals with lactose intolerance can tolerate up to 12 grams of lactose (the amount in one cup of milk) without significant symptoms [92].

The variation in symptoms and tolerance levels underscores the need for individualized dietary recommendations based on genetic background and symptom severity.

3.2.2. Non-Coeliac Gluten/Wheat Sensitivity (NCGWS)

Non-coeliac gluten/Wheat Sensitivity (NCGWS) refers to individuals sensitive to gluten or wheat but lacking immune serological coeliac antibodies or allergic biomarkers, with a higher prevalence observed among females [93]. The Salerno Experts' Criteria, relying on exclusion diagnosis, is currently the only reliable diagnostic tool for NCGWS [5]. Manifestations of NCGWS include both intestinal (bloating, diarrhea, constipation, nausea, etc.) and non-intestinal/extraintestinal symptoms (headache, anxiety, weight loss, anemia) [94]. NCGWS shares some similarities and differences with wheat allergy and celiac disease, as summarized in Table 3 [93]. However, the exact mechanism underlying NCGWS remains poorly understood, leading some individuals to be prescribed a gluten-free diet by healthcare practitioners despite negative results on other tests for gluten-containing foods [5].

Several studies have aimed to define the characteristics, prevalence, and management of NCGWS, often highlighting the complexity and controversies surrounding its diagnosis. A 2015 study by Catassi et al. reviewed the diagnostic criteria and noted that while NCGWS shares symptoms with celiac disease, such as bloating, diarrhea, and fatigue, it lacks the serological markers

and histological changes seen in celiac disease [5]. The study also highlighted the placebo effect in gluten challenge trials, which complicates the diagnosis further, suggesting that some cases of NCGWS might be due to non-gluten components like FODMAPs [5].

In contrast, a 2016 study by Uhde et al. explored the immunological response in NCGWS patients and found evidence of systemic immune activation in response to gluten, without the intestinal damage typical of celiac disease [95]. This study suggested that NCGWS might involve an innate immune response rather than the adaptive immune response seen in celiac disease [95]. However, the study also acknowledged the heterogeneity of the condition, indicating that different patients might react to different components of wheat, including gluten or other proteins [95].

Further research by Skodje et al. compared the effects of gluten, FODMAPs, and placebo in NCGWS patients and concluded that many individuals who believed they were sensitive to gluten were reacting to FODMAPs, which are poorly absorbed carbohydrates found in wheat and other foods [96]. This study emphasized the importance of a structured dietary approach to diagnose NCGWS, as misdiagnosis could lead to unnecessary dietary restrictions and nutritional deficiencies [96].

These studies highlight the complexities in diagnosing and managing NCGWS, emphasizing that while some individuals may indeed react to gluten, others might be sensitive to other components in wheat or even experience a placebo effect. The variability in immune responses and symptoms underscores the need for personalized approaches in both diagnosis and treatment.

Table 3. Comparison of prevalence, pathogenic, and diagnostic features of gluten-related disorders.

	Celiac Disease	NCGWS	Wheat Allergy
Prevalence	0.5–1.7%	0.6-10%	0.5–9% in children
Pathogenesis	Autoimmune	Non-specific immune response	IgE mediated response
DQ2-DQ8 HLA haplotypes	Positive in 95% cases	Positive in 50% cases	Negative
Serological markers	IgA anti-EMA, IgA anti-tTG, IgG anti-DGP, IgA anti-gliadin	IgA/IgG anti-gliadin in 50% cases	specific IgE antibodies against wheat and gliadin
Duodenal biopsy *	Marsh I to IV with domination of Marsh III and IV	Marsh 0-II, but according to some experts Marsh III might also be in NCGS	Marsh 0-II
Duodenal villi atrophy	Present	Absent	Might be present or absent

* Marsh classification (histological grading system used to evaluate and classify the degree of intestinal damage, particularly in the small intestine, in individuals with celiac disease or other gluten-sensitive enteropathies [97]).

Non-celiac gluten/wheat sensitivity (NCGWS), IgA anti-EMA (IgA antibodies against endomysium), IgA anti-tTG (IgA antibodies against transglutaminase), IgG anti-DGP (IgG antibodies against deamidated gliadin peptides).

3.2.3. Fructose Intolerance

Hereditary Fructose Intolerance

Hereditary Fructose Intolerance (HFI) is a rare autosomal hereditary disorder characterized by the inability to metabolize fructose directly or indirectly through sucrose or sorbitol, as noted by Singh & Sarma in 2022 [98]. Individuals with fructose intolerance may experience symptoms such as abdominal pain, diarrhea, nausea, and flatulence upon consuming fructose-rich foods like honey,

fruits, or vegetables, [99]. The mutation of Aldolase B, the primary enzyme responsible for fructose metabolism in the liver, located on chromosome 9q22.3, is implicated in HFI [98]. Diagnostic criteria for HFI typically involve Benedicts test, glucose dipstick test in urine, and serum carbohydrate-deficient transferrin (CDT), complemented by clinical correlations and sometimes requiring liver biopsies to assess Aldolase B enzyme activity [100]. Treatment for HFI primarily revolves around adopting a diet low in fructose, sucrose, and sorbitol (FSS). In acute cases, patients may require admission to an intensive care unit for intravenous glucose administration to manage metabolic acidosis, and it's crucial for individuals with HFI to avoid medications and vaccines containing sucrose, such as the rotavirus oral vaccine [98].

A study by Ali et al. identified several mutations in the *ALDOB* gene responsible for HFI, including the common A149P mutation, which is prevalent in European populations [101]. This research provided insight into the genetic basis of HFI and established the importance of genetic testing for accurate diagnosis [101].

In a more recent study, Tolan reviewed the pathophysiology of HFI and discussed the clinical symptoms that arise due to the accumulation of toxic metabolites, such as fructose-1-phosphate, in the liver, kidneys, and intestines [102]. The study highlighted the importance of early diagnosis and strict dietary management to avoid severe complications like hypoglycemia, liver dysfunction, and failure to thrive in infants [102]. Tolan also discussed the role of genetic counseling for families with a history of HFI and the potential for prenatal diagnosis [102].

These studies underline the genetic complexity of HFI and the critical importance of early diagnosis and dietary management in preventing serious health issues. The advancements in genetic testing have greatly improved the accuracy of HFI diagnosis, enabling better patient outcomes.

Non-Hereditary Fructose Intolerance

Non-Hereditary Fructose Intolerance, or fructose malabsorption (FM), presents as a syndrome where the uptake of fructose in the small intestine is minimal in some individuals, leading to fermentation of unabsorbed fructose in the colon, akin to lactose intolerance and HFI, according [103]. Diagnosis of FM typically involves a specifically designed hydrogen breath test [104]. Treatment options primarily involve the elimination of fructose-containing foods from the diet, with xylose isomerase proposed as an oral treatment to convert fructose to glucose, resulting in a favorable hydrogen breath test in FM patients [105].

A study by Gibson et al. investigated the role of the low FODMAP diet in managing fructose malabsorption [106]. This study highlighted that fructose, along with other fermentable carbohydrates, contribute to symptoms like bloating and diarrhea in susceptible individuals [106]. The low FODMAP diet was shown to be effective in reducing these symptoms, indicating that dietary management is a critical component of treatment for non-hereditary fructose intolerance [106].

Further research by Tuck et al. focused on the diagnostic challenges associated with fructose malabsorption [107]. The study noted that breath hydrogen testing is commonly used to diagnose this condition, but the results can be inconsistent due to variations in individual gut microbiota and other factors [107]. Tuck et al. advocated for a more comprehensive approach to diagnosis, combining dietary history with symptom tracking and possibly genetic testing, although the latter is more relevant to distinguishing between HFI and fructose malabsorption [107].

These studies suggest that non-hereditary fructose intolerance is a complex and multifaceted condition that requires careful dietary management and accurate diagnosis. Unlike HFI, which is caused by a genetic mutation, fructose malabsorption often arises from dietary factors and gut microbiota composition, making individualized treatment plans essential.

3.2.4. Saccharose Intolerance

Saccharose intolerance arises from deficiencies in sucrase-isomaltase enzyme function, caused by congenital sucrase-isomaltase deficiency (CSID) or secondary factors such as celiac disease or Crohn's disease [105]. Symptoms typically involve gastrointestinal issues such as cramps, bloating, gas, and diarrhea [108]. Diagnosis of saccharose intolerance often involves duodenal endoscopic

biopsies or breath tests (hydrogen, C-sucrose) [58]. Treatment may include using sacrosidase enzyme supplementation, which has shown promising results in alleviating symptoms [58].

A 2020 study by Treem et al., published in *Journal of Pediatric Gastroenterology and Nutrition*, highlighted the various genetic mutations affecting the SI gene and discussed enzyme replacement therapies like sacrosidase as an effective treatment [109]. The study also pointed out the importance of dietary modifications, emphasizing low-sucrose and low-starch diets to manage the symptoms effectively [95]. Furthermore, a study by Robayo-Torres et al. highlighted the importance of genetic screening and enzyme activity testing in diagnosing CSID, emphasizing that many cases remain undiagnosed due to the overlap of symptoms with other gastrointestinal disorders like irritable bowel syndrome (IBS) [110]. Misdiagnosis remains a challenge, as individuals with CSID often experience symptoms similar to functional gastrointestinal disorders, leading to unnecessary dietary restrictions or ineffective treatments [111]. This underscores the need for heightened clinical awareness and improved diagnostic protocols, particularly in patients with persistent unexplained gastrointestinal symptoms [112]. This study also pointed to enzyme replacement therapy, particularly with sacrosidase, as an effective treatment option for managing symptoms, alongside dietary modifications that limit sucrose and starch intake [96]. However, long-term adherence to dietary changes and enzyme therapy requires patient education and continuous monitoring, as variations in food processing and enzyme activity levels can affect symptom control [113]. Future research should explore individualized treatment approaches, including personalized enzyme dosing and probiotic interventions, to optimize therapeutic outcomes [114].

3.2.5. Histamine Intolerance

Histamine Intolerance (HIT) refers to a non-immunological condition believed to result from elevated histamine levels in the blood due to the ingestion of histamine-rich foods, potentially causing adverse effects [115]. Histamine intoxication, on the other hand, occurs following the ingestion of histamine-rich foods, with symptoms ranging from gastrointestinal symptoms to skin reactions, low blood pressure, headaches, and palpitations [116]. Suspected causes of histamine intolerance include a lack of oxidative degradation by diamine oxidase (DAO) activity or reduced levels of methylation by histamine N-methyltransferase (HNMT) [117]. This deficiency may be congenital or acquired, with factors such as gastrointestinal disorders (e.g., inflammatory bowel disease, celiac disease), chronic alcohol consumption, and certain medications (e.g., NSAIDs, histamine-releasing drugs) contributing to reduced DAO activity [118]. Studies suggest that DAO activity is predominantly localized in the intestinal mucosa, meaning that any disruption in gut health, such as increased intestinal permeability or dysbiosis, may exacerbate symptoms in susceptible individuals [14]. Common symptoms of HIT predominantly affect the gastrointestinal tract, with constipation, diarrhea, abdominal pain, and postprandial fullness being most prevalent [119]. However, the variability in symptoms between individuals suggests that other factors, such as genetic predisposition, gut microbiota composition, and cumulative histamine load from endogenous and exogenous sources, may play a role in symptom severity [14]. This variability also makes HIT difficult to diagnose, as symptoms overlap with those of other functional gastrointestinal disorders like irritable bowel syndrome (IBS) and non-celiac gluten sensitivity [120]. Treatment for HIT typically involves adhering to a low-histamine diet, with antihistamines used to alleviate symptoms [121]. However, managing dietary intake can be challenging due to the fluctuating histamine content in foods, storage conditions, and individual tolerance thresholds [122]. Fermented foods, aged cheeses, cured meats, and alcohol are well-documented triggers, yet some individuals may tolerate small amounts without symptoms, complicating dietary recommendations [122]. Mast cell stabilizers may also be utilized, although further research is needed to determine their efficacy in HIT [123]. Oral supplementation with exogenous DAO has shown promise in reducing symptoms, but additional studies with larger sample sizes are required to ascertain its effectiveness in HIT patients [124].

Histamine intolerance (HI) is an emerging area of research with various studies examining its prevalence, pathophysiology, and management strategies, that explored the physiological mechanisms of histamine metabolism and its relation to HIT [125]. They emphasize that histamine intolerance results from an imbalance between histamine release and its degradation due to enzyme deficiencies, particularly diamine oxidase (DAO). Another significant contribution is by Schink, et al., who provide insights into diagnostic approaches and clinical management of HIT [14]. They suggested that HIT can be managed through dietary modifications and the use of DAO supplements. These studies underscore the multifaceted nature of HIT and highlight the importance of a comprehensive approach in both diagnosis and management [14]. Histamine intolerance remains an under-researched condition, with ongoing debates regarding its true prevalence, diagnostic criteria, and pathophysiological mechanisms. While histamine skin prick tests, plasma histamine levels, and DAO activity measurements have been suggested as potential diagnostic tools, none have been universally accepted due to inconsistencies in sensitivity and specificity [126]. Given the overlap of symptoms with other conditions, a comprehensive diagnostic approach—including elimination diets, symptom tracking, and laboratory assessments—is crucial for accurately identifying HIT. Future research should focus on refining diagnostic criteria, understanding individual variations in histamine metabolism, and exploring novel treatment approaches, such as targeted enzyme therapy or gut microbiota modulation, to improve patient outcomes [127].

3.2.6. FODMAP

FODMAPs, or Fermentable Oligo-, Di-, Mono-Saccharides, and Polyols, are short-chain carbohydrates found abundantly in fruits, vegetables, dairy, cereals, and sweeteners [106]. Lactose, fructose, sorbitol, mannitol, fructans, stachyose, and raffinose are all classified as FODMAPs [128]. Consumption of more than 4g of lactose, more than 0.3g of mannitol, sorbitol, galacto-oligosaccharides, or fructans is considered a high FODMAP diet [129]. FODMAPs are poorly digested by intestinal bacteria, leading to the production of short-chain fatty acids (SCFAs) and gas, which can cause symptoms such as abdominal pain, flatulence, diarrhea, and indigestion in susceptible individuals, particularly those with Irritable Bowel Syndrome (IBS) [130]. A low FODMAP diet (LFD) has been shown to effectively reduce symptoms and improve the quality of life in IBS patients [129]. The LFD typically follows a three-phase approach [15]. Phase 1 involves eliminating all FODMAPs from the diet for 4 to 6 weeks. Phase 2 assesses each patient's tolerance to FODMAP subgroups by reintroducing one food at a time over three days. Phase 3, building upon the findings of Phase 2, customizes a long-term FODMAP diet tailored to each patient's tolerance [131]. However, concerns have been raised regarding potential nutritional deficiencies, constipation, eating disorders, and alterations in gut microbiota among individuals on a long-term LFD [111–113].

One seminal paper by Gibson et al. in *Gastroenterology* introduced the low FODMAP diet and demonstrated its effectiveness in reducing IBS symptoms through a controlled trial [106]. Their research provided the foundation for subsequent studies exploring dietary interventions [106]. Another comprehensive review by Halmos et al. supports the low FODMAP diet, showing that it significantly alleviates IBS symptoms compared to a standard diet [15]. This study is crucial for understanding the clinical application of the diet [15]. Additionally, a more recent study by Staudacher et al. confirms the diet's effectiveness across various populations, emphasizing its role in symptom management [135]. These studies emphasize the low FODMAP diet's role in managing IBS and other functional gastrointestinal disorders, providing robust evidence for its clinical use.

3.3. Irritable Bowel Syndrome IBS

Irritable bowel syndrome (IBS) affects approximately 20% of the global population [136]. Its primary symptoms include abdominal pain, bloating, and irregularities in stool form and frequency [137]. Currently, there are no globally recognized biomarkers for diagnosing IBS; instead, diagnostic criteria based on symptoms have been established by the scientific community, known as the Rome IV criteria [138].

Patients are categorized into subtypes based on the Bristol stool form: IBS with constipation (IBS-C), IBS with diarrhea (IBS-D), IBS with mixed stool pattern (IBS-M), and IBS unclassified (IBS-U) [80,81]. IBS is a complex syndrome influenced by multiple factors and often overlaps with various comorbidities, even among its own subtypes [139].

Common gastrointestinal disorders that overlap with IBS include gastroesophageal reflux (GERD), nausea, constipation, diarrhea, heartburn, dyspepsia, and incontinence [83,84]. Non-gastrointestinal syndromes also frequently overlap with IBS [80,81]. These include psychiatric conditions such as depression, anxiety, and somatization, as well as premenstrual syndrome (PMS), overactive bladder, fibromyalgia, eating disorders, and food hypersensitivities (intolerances and allergies) [142].

This overlap has prompted discussion within the scientific community about whether IBS should be considered part of these syndromes rather than as a distinct syndrome on its own [142]. Additionally, IBS and its associated functional gastrointestinal and non-gastrointestinal disorders are now classified under the somatic symptom disorders in the Diagnostic and Statistical Manual of Mental Disorders, 5th Edition (DSM-5). This categorization reflects a historical trend where patients with gastrointestinal symptoms treated by psychiatrists often experienced inadequate treatment and care [143].

Numerous risk factors associated with the development of IBS have been identified, including personal factors such as being female and having a low body mass index (BMI) [144].

Psychological factors like anxiety, depression, and low quality of life also increase somatic issues such as diverticulosis, antibiotic use, gastrointestinal infections, and endometriosis, as well as social conditions like a family history of mental illness, childhood socioeconomic status, and marital status [144].

Moreover, many individuals diagnosed with IBS also report food allergies or intolerances, particularly to gluten, dairy (cow milk protein and lactose), and FODMAPs, which can trigger adverse reactions [53,88,89].

The pathophysiology of IBS involves various mechanisms. These include alterations in gut microbiota, changes in the epithelial barrier, immune system responses to food antigens, and bile acids, and interactions within the brain-gut axis, enteric nervous system, and hypothalamus-pituitary-adrenal axis, which are increasingly considered potential biomarkers of IBS [147].

Additionally, psychological factors such as depression, anxiety, and stress contribute to the pathophysiology of IBS by influencing intestinal motility [147].

As mentioned previously, the composition of gut microbiota in patients with IBS varies depending on the subtype of the condition, as detailed in Table [147]. Studies have demonstrated that culturing these microbiota types present in IBS patients, compared to those in individuals without IBS, can lead to lower intestinal motility, induce visceral hypersensitivity, and alter transit time. These findings on microbiota dysbiosis have been recognized by the ROME foundation as seen in Table 4 [147].

One pivotal study by Longstreth et al., provides a comprehensive review of IBS epidemiology, diagnostic criteria, and management options, emphasizing the heterogeneity of the disorder and the importance of a tailored treatment approach [148]. Another influential study by Ford et al. examines the efficacy of pharmacological treatments for IBS, including antispasmodics, laxatives, and antidiarrheals, and finds that while some medications are effective, their benefits vary among patients [13]. A more recent randomized controlled trial by Drossman et al. investigates the effectiveness of a novel treatment approach, such as a combination of dietary interventions and pharmacological agents, and showed significant improvement in IBS symptoms compared to traditional therapies [149]. Additionally, a meta-analysis by Lacy et al. provided an overview of the impact of the low FODMAP diet on IBS symptoms, confirming its efficacy in symptom relief and highlighting the need for personalized dietary strategies [150]. Furthermore, a review study done by Pasta et al. provides an in-depth analysis of the relationship between irritable bowel syndrome (IBS), food allergies, and food intolerances, highlighting their overlapping symptoms and distinct mechanisms. It emphasizes

that while food intolerances, particularly fermentable carbohydrates (FODMAPs) and lactose, are common in IBS, true food allergies involving immune system activation are rare in IBS patients. The study underscores that many IBS symptoms, such as bloating, abdominal pain, and altered bowel habits, are frequently mistaken for allergic reactions, leading to unnecessary dietary restrictions. The review also discusses the importance of accurate diagnostic methods to differentiate between IBS-related food intolerances and immune-mediated allergies, ensuring effective dietary management and symptom relief [151]. Lastly, a review by Quigley et al. explored the role of probiotics and prebiotics in managing IBS, revealing that while some evidence supports their use, more research is needed to establish their effectiveness conclusively [152]. Together, these studies underscored the complexity of IBS and the need for a multifaceted approach to its management.

4. Conclusions

Based on the current review, understanding the key differences between food allergies and intolerances is crucial for effective management and treatment strategies. While both conditions can lead to uncomfortable symptoms, they arise from distinct mechanisms within the body.

Food allergies involve an immune system response to specific proteins in food, often resulting in rapid and potentially life-threatening reactions. On the other hand, food intolerances stem from difficulty digesting certain foods or components, leading to gastrointestinal discomfort or other symptoms. These reactions are typically less severe and do not involve the immune system.

Recognizing the symptoms and triggers of each condition is essential for accurate diagnosis and appropriate management. Food allergies may require strict avoidance of triggering foods and the availability of emergency medication, such as epinephrine, in case of severe reactions. Meanwhile, managing food intolerances often involves identifying and eliminating problematic foods from the diet, as well as considering enzyme supplements or other supportive measures.

Overall, by understanding the nuances between food allergies and intolerances, individuals can take proactive steps to minimize their symptoms and maintain their overall health and well-being. Consulting with healthcare professionals and registered dietitians can provide personalized guidance and support in navigating these dietary challenges.

Author Contributions: G. H., as the principal investigator, developed the idea, performed the literature review, and wrote and edited the manuscript. G.H., Y.S., J.H., M.H, L.S., and N.S. contributed also to the literature review, wrote and edited the manuscript, and acted as lead reviewers. J. H. was responsible for reviewing and editing tables and the figures. All authors participated in the preparation of the manuscript and approved its final version.

Funding: This research received no external funding

Conflicts of Interest: The authors declare no conflict of interest

Acknowledgments: This work was supported by the Higher Center for Research (HCR) at the Holy Spirit University of Kaslik (USEK), Lebanon.

References

1. Gargano D, Appanna R, Santonicola A, De Bartolomeis F, Stellato C, Cianferoni A, Casolaro V, Iovino P. Food Allergy and Intolerance: A Narrative Review on Nutritional Concerns. *Nutrients* (2021) 13:1638. doi: 10.3390/nu13051638
2. Muraro A, Halken S, Arshad SH, Beyer K, Dubois AEJ, Du Toit G, Eigenmann PA, Grimshaw KEC, Hoest A, Lack G, et al. EAACI Food Allergy and Anaphylaxis Guidelines. Primary prevention of food allergy. *Allergy* (2014) 69:590–601. doi: 10.1111/all.12398
3. Boyce JA, Assa'ad A, Burks AW, Jones SM, Sampson HA, Wood RA, Plaut M, Cooper SF, Fenton MJ, Arshad SH, et al. Guidelines for the Diagnosis and Management of Food Allergy in the United States:

- Summary of the NIAID-Sponsored Expert Panel Report. *Journal of Allergy and Clinical Immunology* (2010) 126:1105–1118. doi: 10.1016/j.jaci.2010.10.008
4. Daniel L. Swagerty J, Walling AD, Klein RM. Lactose Intolerance. *afp* (2002) 65:1845–1851.
 5. Catassi C, Elli L, Bonaz B, Bouma G, Carroccio A, Castillejo G, Cellier C, Cristofori F, de Magistris L, Dolinsek J, et al. Diagnosis of Non-Celiac Gluten Sensitivity (NCGS): The Salerno Experts' Criteria. *Nutrients* (2015) 7:4966–4977. doi: 10.3390/nu7064966
 6. Tedner SG, Asarnoj A, Thulin H, Westman M, Konradsen JR, Nilsson C. Food allergy and hypersensitivity reactions in children and adults—A review. *Journal of Internal Medicine* (2022) 291:283–302. doi: 10.1111/joim.13422
 7. Connors L, O'Keefe A, Rosenfield L, Kim H. Non-IgE-mediated food hypersensitivity. *Allergy, Asthma & Clinical Immunology* (2018) 14:56. doi: 10.1186/s13223-018-0285-2
 8. Zingone F, Bertin L, Maniero D, Palo M, Lorenzon G, Barberio B, Ciacci C, Savarino EV. Myths and Facts about Food Intolerance: A Narrative Review. *Nutrients* (2023) 15:4969. doi: 10.3390/nu15234969
 9. Moher D, Liberati A, Tetzlaff J, Altman DG, PRISMA Group. Preferred reporting items for systematic reviews and meta-analyses: the PRISMA statement. *PLoS Med* (2009) 6:e1000097. doi: 10.1371/journal.pmed.1000097
 10. Vassilopoulou E, Feketea G, Konstantinou GN, Zekakos Xypolias D, Valianatou M, Petrodimopoulou M, Vourga V, Tasios I, Papadopoulos NG. Food Protein-Induced Allergic Proctocolitis: The Effect of Maternal Diet During Pregnancy and Breastfeeding in a Mediterranean Population. *Front Nutr* (2022) 9: doi: 10.3389/fnut.2022.843437
 11. Ruffner MA, Ruymann K, Barni S, Cianferoni A, Brown-Whitehorn T, Spergel JM. Food protein-induced enterocolitis syndrome: insights from review of a large referral population. *J Allergy Clin Immunol Pract* (2013) 1:343–349. doi: 10.1016/j.jaip.2013.05.011
 12. Pinto-Sanchez MI, Nardelli A, Borojevic R, De Palma G, Calo NC, McCarville J, Caminero A, Basra D, Mordhorst A, Ignatova E, et al. Gluten-Free Diet Reduces Symptoms, Particularly Diarrhea, in Patients With Irritable Bowel Syndrome and Antigliadin IgG. *Clin Gastroenterol Hepatol* (2021) 19:2343-2352.e8. doi: 10.1016/j.cgh.2020.08.040
 13. Ford AC, Bercik P, Morgan DG, Bolino C, Pintos-Sanchez MI, Moayyedi P. Characteristics of functional bowel disorder patients: a cross-sectional survey using the Rome III criteria. *Aliment Pharmacol Ther* (2014) 39:312–321. doi: 10.1111/apt.12573
 14. Schink M, Konturek PC, Tietz E, Dieterich W, Pinzer TC, Wirtz S, Neurath MF, Zopf Y. Microbial patterns in patients with histamine intolerance. *J Physiol Pharmacol* (2018) 69: doi: 10.26402/jpp.2018.4.09
 15. Halmos EP, Power VA, Shepherd SJ, Gibson PR, Muir JG. A diet low in FODMAPs reduces symptoms of irritable bowel syndrome. *Gastroenterology* (2014) 146:67-75.e5. doi: 10.1053/j.gastro.2013.09.046
 16. Nwaru BI, Hickstein L, Panesar SS, Muraro A, Werfel T, Cardona V, Dubois AEJ, Halken S, Hoffmann-Sommergruber K, Poulsen LK, et al. The epidemiology of food allergy in Europe: a systematic review and meta-analysis. *Allergy* (2014) 69:62–75. doi: 10.1111/all.12305
 17. West J, Fleming KM, Tata LJ, Card TR, Crooks CJ. Incidence and Prevalence of Celiac Disease and Dermatitis Herpetiformis in the UK Over Two Decades: Population-Based Study. *Am J Gastroenterol* (2014) 109:757–768. doi: 10.1038/ajg.2014.55
 18. Sicherer SH, Sampson HA. Food allergy: A review and update on epidemiology, pathogenesis, diagnosis, prevention, and management. *Journal of Allergy and Clinical Immunology* (2018) 141:41–58. doi: 10.1016/j.jaci.2017.11.003
 19. Wasserman S, Bégin P, Watson W. IgE-mediated food allergy. *Allergy, Asthma & Clinical Immunology* (2018) 14:55. doi: 10.1186/s13223-018-0284-3
 20. Anvari S, Miller J, Yeh C-Y, Davis CM. IgE-Mediated Food Allergy. *Clinic Rev Allerg Immunol* (2019) 57:244–260. doi: 10.1007/s12016-018-8710-3
 21. Patel BY, Volcheck GW. Food Allergy: Common Causes, Diagnosis, and Treatment. *Mayo Clinic Proceedings* (2015) 90:1411–1419. doi: 10.1016/j.mayocp.2015.07.012

22. Sampson HA, Aceves S, Bock SA, James J, Jones S, Lang D, Nadeau K, Nowak-Wegrzyn A, Oppenheimer J, Perry TT, et al. Food allergy: a practice parameter update-2014. *J Allergy Clin Immunol* (2014) 134:1016-1025.e43. doi: 10.1016/j.jaci.2014.05.013
23. Savage J, Sicherer S, Wood R. The Natural History of Food Allergy. *J Allergy Clin Immunol Pract* (2016) 4:196–203; quiz 204. doi: 10.1016/j.jaip.2015.11.024
24. Heinzerling L, Mari A, Bergmann K-C, Bresciani M, Burbach G, Darsow U, Durham S, Fokkens W, Gjomarkaj M, Haahtela T, et al. The skin prick test – European standards. *Clin Transl Allergy* (2013) 3:3. doi: 10.1186/2045-7022-3-3
25. Heinzerling L, Mari A, Bergmann K-C, Bresciani M, Burbach G, Darsow U, Durham S, Fokkens W, Gjomarkaj M, Haahtela T, et al. The skin prick test – European standards. *Clinical and Translational Allergy* (2013) 3:3. doi: 10.1186/2045-7022-3-3
26. Bousquet J, Schünemann HJ, Samolinski B, Demoly P, Baena-Cagnani CE, Bachert C, Bonini S, Boulet LP, Bousquet PJ, Brozek JL, et al. Allergic Rhinitis and its Impact on Asthma (ARIA): achievements in 10 years and future needs. *J Allergy Clin Immunol* (2012) 130:1049–1062. doi: 10.1016/j.jaci.2012.07.053
27. Ansotegui IJ, Melioli G, Canonica GW, Caraballo L, Villa E, Ebisawa M, Passalacqua G, Savi E, Ebo D, Gómez RM, et al. IgE allergy diagnostics and other relevant tests in allergy, a World Allergy Organization position paper. *World Allergy Organ J* (2020) 13:100080. doi: 10.1016/j.waojou.2019.100080
28. Wood RA, Kim JS, Lindblad R, Nadeau K, Henning AK, Dawson P, Plaut M, Sampson HA. A randomized, double-blind, placebo-controlled study of omalizumab combined with oral immunotherapy for the treatment of cow's milk allergy. *J Allergy Clin Immunol* (2016) 137:1103-1110.e11. doi: 10.1016/j.jaci.2015.10.005
29. Matricardi PM, Kleine-Tebbe J, Hoffmann HJ, Valenta R, Hilger C, Hofmaier S, Aalberse RC, Agache I, Asero R, Ballmer-Weber B, et al. EAACI Molecular Allergology User's Guide. *Pediatr Allergy Immunol* (2016) 27 Suppl 23:1–250. doi: 10.1111/pai.12563
30. Zuberbier T, Aberer W, Asero R, Abdul Latiff AH, Baker D, Ballmer-Weber B, Bernstein JA, Bindslev-Jensen C, Brzoza Z, Buense Bedrikow R, et al. The EAACI/GA²LEN/EDF/WAO guideline for the definition, classification, diagnosis and management of urticaria. *Allergy* (2018) 73:1393–1414. doi: 10.1111/all.13397
31. Ewan PW, Dugué P, Mirakian R, Dixon TA, Harper JN, Nasser SM. BSACI guidelines for the investigation of suspected anaphylaxis during general anaesthesia. *Clinical & Experimental Allergy* (2010) 40:15–31. doi: 10.1111/j.1365-2222.2009.03404.x
32. Gargano D, Appanna R, Santonicola A, De Bartolomeis F, Stellato C, Cianferoni A, Casolaro V, Iovino P. Food Allergy and Intolerance: A Narrative Review on Nutritional Concerns. *Nutrients* (2021) 13:1638. doi: 10.3390/nu13051638
33. Sicherer SH, Sampson HA. Food allergy: Epidemiology, pathogenesis, diagnosis, and treatment. *J Allergy Clin Immunol* (2014) 133:291–307; quiz 308. doi: 10.1016/j.jaci.2013.11.020
34. Gupta RS, Warren CM, Smith BM, Jiang J, Blumenstock JA, Davis MM, Schleimer RP, Nadeau KC. Prevalence and Severity of Food Allergies Among US Adults. *JAMA Netw Open* (2019) 2:e185630. doi: 10.1001/jamanetworkopen.2018.5630
35. Osborne NJ, Koplin JJ, Martin PE, Gurrin LC, Thiele L, Tang ML, Ponsonby A-L, Dharmage SC, Allen KJ, HealthNuts Study Investigators. The HealthNuts population-based study of paediatric food allergy: validity, safety and acceptability. *Clin Exp Allergy* (2010) 40:1516–1522. doi: 10.1111/j.1365-2222.2010.03562.x
36. Muir A, Falk GW. Eosinophilic Esophagitis: A Review. *JAMA* (2021) 326:1310–1318. doi: 10.1001/jama.2021.14920
37. Dellon ES, Jensen ET, Martin CF, Shaheen NJ, Kappelman MD. Prevalence of eosinophilic esophagitis in the United States. *Clin Gastroenterol Hepatol* (2014) 12:589-596.e1. doi: 10.1016/j.cgh.2013.09.008
38. Liacouras CA, Furuta GT, Hirano I, Atkins D, Attwood SE, Bonis PA, Burks AW, Chehade M, Collins MH, Dellon ES, et al. Eosinophilic esophagitis: Updated consensus recommendations for children and adults. *Journal of Allergy and Clinical Immunology* (2011) 128:3-20.e6. doi: 10.1016/j.jaci.2011.02.040
39. Hirano I, Pandolfino JE, Boeckxstaens GE. Functional Lumen Imaging Probe for the Management of Esophageal Disorders: Expert Review From the Clinical Practice Updates Committee of the AGA Institute. *Clin Gastroenterol Hepatol* (2017) 15:325–334. doi: 10.1016/j.cgh.2016.10.022

40. Lucendo AJ, Arias Á, Molina-Infante J. Efficacy of Proton Pump Inhibitor Drugs for Inducing Clinical and Histologic Remission in Patients With Symptomatic Esophageal Eosinophilia: A Systematic Review and Meta-Analysis. *Clin Gastroenterol Hepatol* (2016) 14:13-22.e1. doi: 10.1016/j.cgh.2015.07.041
41. Barni S, Mori F, Giovannini M, Liotti L, Mastrorilli C, Pecoraro L, Saretta F, Castagnoli R, Arasi S, Caminiti L, et al. Allergic Proctocolitis: Literature Review and Proposal of a Diagnostic–Therapeutic Algorithm. *Life (Basel)* (2023) 13:1824. doi: 10.3390/life13091824
42. Zubeldia-Varela E, Barker-Tejeda TC, Blanco-Pérez F, Infante S, Zubeldia JM, Pérez-Gordo M. Non-IgE-Mediated Gastrointestinal Food Protein-Induced Allergic Disorders. Clinical Perspectives and Analytical Approaches. *Foods* (2021) 10: doi: 10.3390/foods10112662
43. Martin VM, Virkud YV, Seay H, Hickey A, Ndahayo R, Rosow R, Southwick C, Elkort M, Gupta B, Kramer E, et al. PROSPECTIVE ASSESSMENT OF PEDIATRICIAN-DIAGNOSED FOOD-PROTEIN INDUCED ALLERGIC PROCTOCOLITIS BY GROSS OR OCCULT BLOOD. *J Allergy Clin Immunol Pract* (2020) 8:1692-1699.e1. doi: 10.1016/j.jaip.2019.12.029
44. Elizur A, Cohen M, Goldberg MR, Rajuan N, Cohen A, Leshno M, Katz Y. Cow's milk associated rectal bleeding: a population based prospective study. *Pediatr Allergy Immunol* (2012) 23:766–770. doi: 10.1111/pai.12009
45. Mennini M, Fiocchi AG, Cafarotti A, Montesano M, Mauro A, Villa MP, Di Nardo G. Food protein-induced allergic proctocolitis in infants: Literature review and proposal of a management protocol. *World Allergy Organ J* (2020) 13:100471. doi: 10.1016/j.waojou.2020.100471
46. Nowak-Węgrzyn A, Warren CM, Brown-Whitehorn T, Cianferoni A, Schultz-Matney F, Gupta RS. Food protein-induced enterocolitis syndrome in the US population-based study. *J Allergy Clin Immunol* (2019) 144:1128–1130. doi: 10.1016/j.jaci.2019.06.032
47. Caubet J-C, Szajewska H, Shamir R, Nowak-Węgrzyn A. Non-IgE-mediated gastrointestinal food allergies in children. *Pediatr Allergy Immunol* (2017) 28:6–17. doi: 10.1111/pai.12009
48. Lebowitz B, Rubio-Tapia A. Epidemiology, Presentation, and Diagnosis of Celiac Disease. *Gastroenterology* (2021) 160:63–75. doi: 10.1053/j.gastro.2020.06.098
49. Al-Toma A, Volta U, Auricchio R, Castillejo G, Sanders DS, Cellier C, Mulder CJ, Lundin KEA. European Society for the Study of Coeliac Disease (ESsCD) guideline for coeliac disease and other gluten-related disorders. *United European Gastroenterol J* (2019) 7:583–613. doi: 10.1177/2050640619844125
50. Alkalay MJ. Nutrition in Patients with Lactose Malabsorption, Celiac Disease, and Related Disorders. *Nutrients* (2021) 14:2. doi: 10.3390/nu14010002
51. Hadithi M, von Blomberg BME, Crusius JBA, Bloemena E, Kostense PJ, Meijer JWR, Mulder CJJ, Stehouwer CDA, Peña AS. Accuracy of Serologic Tests and HLA-DQ Typing for Diagnosing Celiac Disease. *Ann Intern Med* (2007) 147:294–302. doi: 10.7326/0003-4819-147-5-200709040-00003
52. Singh P, Arora A, Strand TA, Leffler DA, Catassi C, Green PH, Kelly CP, Ahuja V, Makharia GK. Global Prevalence of Celiac Disease: Systematic Review and Meta-analysis. *Clinical Gastroenterology and Hepatology* (2018) 16:823-836.e2. doi: 10.1016/j.cgh.2017.06.037
53. Lebowitz B, Rubio-Tapia A. Epidemiology, Presentation, and Diagnosis of Celiac Disease. *Gastroenterology* (2021) 160:63–75. doi: 10.1053/j.gastro.2020.06.098
54. Ludvigsson JF, Murray JA. Epidemiology of Celiac Disease. *Gastroenterol Clin North Am* (2019) 48:1–18. doi: 10.1016/j.gtc.2018.09.004
55. Rubio-Tapia A, Hill ID, Semrad C, Kelly CP, Greer KB, Limketkai BN, Lebowitz B. American College of Gastroenterology Guidelines Update: Diagnosis and Management of Celiac Disease. *Official journal of the American College of Gastroenterology | ACG* (2023) 118:59. doi: 10.14309/ajg.0000000000002075
56. SALMI T, HERVONEN K. Current Concepts of Dermatitis Herpetiformis. *Acta Derm Venereol* (2020) 100:5664. doi: 10.2340/00015555-3401
57. Collin P, Salmi TT, Hervonen K, Kaukinen K, Reunala T. Dermatitis herpetiformis: a cutaneous manifestation of coeliac disease. *Annals of Medicine* (2017) 49:23–31. doi: 10.1080/07853890.2016.1222450
58. Nguyen CN, Kim S-J. Dermatitis Herpetiformis: An Update on Diagnosis, Disease Monitoring, and Management. *Medicina (Kaunas)* (2021) 57:843. doi: 10.3390/medicina57080843

59. Antiga E, Maglie R, Quintarelli L, Verdelli A, Bonciani D, Bonciolini V, Caproni M. Dermatitis Herpetiformis: Novel Perspectives. *Front Immunol* (2019) 10:1290. doi: 10.3389/fimmu.2019.01290
60. Reunala T, Hervonen K, Salmi T. Dermatitis Herpetiformis: An Update on Diagnosis and Management. *Am J Clin Dermatol* (2021) 22:329–338. doi: 10.1007/s40257-020-00584-2
61. Mansikka E, Hervonen K, Kaukinen K, Collin P, Huhtala H, Reunala T, Salmi T. Prognosis of Dermatitis Herpetiformis Patients with and without Villous Atrophy at Diagnosis. *Nutrients* (2018) 10:641. doi: 10.3390/nu10050641
62. Collin P, Salmi TT, Hervonen K, Kaukinen K, Reunala T. Dermatitis herpetiformis: a cutaneous manifestation of coeliac disease. *Ann Med* (2017) 49:23–31. doi: 10.1080/07853890.2016.1222450
63. Caproni M, Antiga E, Melani L, Fabbri P, Italian Group for Cutaneous Immunopathology. Guidelines for the diagnosis and treatment of dermatitis herpetiformis. *J Eur Acad Dermatol Venereol* (2009) 23:633–638. doi: 10.1111/j.1468-3083.2009.03188.x
64. Feuille E, Nowak-Węgrzyn A. Food Protein-Induced Enterocolitis Syndrome, Allergic Proctocolitis, and Enteropathy. *Curr Allergy Asthma Rep* (2015) 15:50. doi: 10.1007/s11882-015-0546-9
65. Jenkins HR, Pincott JR, Soothill JF, Milla PJ, Harries JT. Food allergy: the major cause of infantile colitis. *Arch Dis Child* (1984) 59:326–329.
66. Caubet JC, Ford LS, Sickles L, Järvinen KM, Sicherer SH, Sampson HA, Nowak-Węgrzyn A. Clinical features and resolution of food protein-induced enterocolitis syndrome: 10-year experience. *J Allergy Clin Immunol* (2014) 134:382–389. doi: 10.1016/j.jaci.2014.04.008
67. Fernandes BN, Boyle RJ, Gore C, Simpson A, Custovic A. Food protein-induced enterocolitis syndrome can occur in adults. *J Allergy Clin Immunol* (2012) 130:1199–1200. doi: 10.1016/j.jaci.2012.06.017
68. Koc AS, Sucu A, Celik U. A different clinical presentation of Heiner syndrome: The case of diffuse alveolar hemorrhage causing massive hemoptysis and hematemesis. *Respir Med Case Rep* (2019) 26:206–208. doi: 10.1016/j.rmcr.2019.01.019
69. Arasi S, Mastrorilli C, Pecoraro L, Giovannini M, Mori F, Barni S, Caminiti L, Castagnoli R, Liotti L, Saretta F, et al. Heiner Syndrome and Milk Hypersensitivity: An Updated Overview on the Current Evidence. *Nutrients* (2021) 13:1710. doi: 10.3390/nu13051710
70. Lee JY, Park M, Jung JH, Kim SY, Kim YH, Hahn SM, Kim S, Lee M-J, Shim HS, Sohn MH, et al. Children with Heiner Syndrome: A Single-Center Experience. *Children (Basel)* (2021) 8:1110. doi: 10.3390/children8121110
71. Moissidis I, Chaidaroon D, Vichyanond P, Bahna SL. Milk-induced pulmonary disease in infants (Heiner syndrome). *Pediatric Allergy and Immunology* (2005) 16:545–552. doi: 10.1111/j.1399-3038.2005.00291.x
72. Lee JY, Park M, Jung JH, Kim SY, Kim YH, Hahn SM, Kim S, Lee M-J, Shim HS, Sohn MH, et al. Children with Heiner Syndrome: A Single-Center Experience. *Children (Basel)* (2021) 8:1110. doi: 10.3390/children8121110
73. Arasi S, Mastrorilli C, Pecoraro L, Giovannini M, Mori F, Barni S, Caminiti L, Castagnoli R, Liotti L, Saretta F, et al. Heiner Syndrome and Milk Hypersensitivity: An Updated Overview on the Current Evidence. *Nutrients* (2021) 13:1710. doi: 10.3390/nu13051710
74. Exl BM, Vandenplas Y, Blecker U. Role of hydrolyzed formulas in nutritional allergy prevention in infants. *South Med J* (1997) 90:1170–1175. doi: 10.1097/00007611-199712000-00002
75. Labrosse R, Graham F, Caubet J-C. Non-IgE-Mediated Gastrointestinal Food Allergies in Children: An Update. *Nutrients* (2020) 12:2086. doi: 10.3390/nu12072086
76. Calvani M, Anania C, Bianchi A, D’Auria E, Cardinale F, Votto M, Martelli A, Tosca M, Chiappini E, Brambilla I, et al. Update on Food protein-induced enterocolitis syndrome (FPIES). *Acta Biomed* (2021) 92:e2021518. doi: 10.23750/abm.v92iS7.12394
77. Nowak-Węgrzyn A, Chehade M, Groetch ME, Spergel JM, Wood RA, Allen K, Atkins D, Bahna S, Barad AV, Berin C, et al. International consensus guidelines for the diagnosis and management of food protein-induced enterocolitis syndrome: Executive summary—Workgroup Report of the Adverse Reactions to Foods Committee, American Academy of Allergy, Asthma & Immunology. *Journal of Allergy and Clinical Immunology* (2017) 139:1111–1126.e4. doi: 10.1016/j.jaci.2016.12.966

78. Mehr S, Frith K, Campbell DE. Epidemiology of food protein-induced enterocolitis syndrome. *Curr Opin Allergy Clin Immunol* (2014) 14:208–216. doi: 10.1097/ACI.0000000000000056
79. Wang KY, Lee J, Cianferoni A, Ruffner MA, Dean A, Molleston JM, Pawlowski NA, Heimall J, Saltzman RW, Ram GS, et al. Food Protein–Induced Enterocolitis Syndrome Food Challenges: Experience from a Large Referral Center. *The Journal of Allergy and Clinical Immunology: In Practice* (2019) 7:444–450. doi: 10.1016/j.jaip.2018.09.009
80. Venter C, Mazzocchi A, Maslin K, Agostoni C. Impact of elimination diets on nutrition and growth in children with multiple food allergies. *Curr Opin Allergy Clin Immunol* (2017) 17:220–226. doi: 10.1097/ACI.0000000000000358
81. Taico Oliva C, Musa I, Kopulos D, Ardalani F, Maskey A, Wilson A, Yang N, Li X-M. The gut microbiome and cross-reactivity of food allergens: current understanding, insights, and future directions. *Front Allergy* (2025) 5: doi: 10.3389/falgy.2024.1503380
82. Tuck CJ, Biesiekierski JR, Schmid-Grendelmeier P, Pohl D. Food Intolerances. *Nutrients* (2019) 11:1684. doi: 10.3390/nu11071684
83. Misselwitz B, Butter M, Verbeke K, Fox MR. Update on lactose malabsorption and intolerance: pathogenesis, diagnosis and clinical management. *Gut* (2019) 68:2080–2091. doi: 10.1136/gutjnl-2019-318404
84. Catanzaro R, Sciuto M, Marotta F. Lactose intolerance: An update on its pathogenesis, diagnosis, and treatment. *Nutrition Research* (2021) 89:23–34. doi: 10.1016/j.nutres.2021.02.003
85. Bayless TM, Brown E, Paige DM. Lactase Non-persistence and Lactose Intolerance. *Curr Gastroenterol Rep* (2017) 19:23. doi: 10.1007/s11894-017-0558-9
86. Deng Y, Misselwitz B, Dai N, Fox M. Lactose Intolerance in Adults: Biological Mechanism and Dietary Management. *Nutrients* (2015) 7:8020–8035. doi: 10.3390/nu7095380
87. Enattah NS, Sahi T, Savilahti E, Terwilliger JD, Peltonen L, Järvelä I. Identification of a variant associated with adult-type hypolactasia. *Nat Genet* (2002) 30:233–237. doi: 10.1038/ng826
88. Simrén M, Stotzer P. Use and abuse of hydrogen breath tests. *Gut* (2006) 55:297–303. doi: 10.1136/gut.2005.075127
89. Simrén M, Barbara G, Flint HJ, Spiegel BMR, Spiller RC, Vanner S, Verdu EF, Whorwell PJ, Zoetendal EG. Intestinal microbiota in functional bowel disorders: a Rome foundation report. *Gut* (2013) 62:159–176. doi: 10.1136/gutjnl-2012-302167
90. Jo IH, Paik C-N, Kim Y-J, Lee JM, Choi SY, Hong KP. Lactase Deficiency Diagnosed by Endoscopic Biopsy-based Method is Associated With Positivity to Glucose Breath Test. *J Neurogastroenterol Motil* (2023) 29:85–93. doi: 10.5056/jnm22023
91. Swallow DM. Genetics of lactase persistence and lactose intolerance. *Annu Rev Genet* (2003) 37:197–219. doi: 10.1146/annurev.genet.37.110801.143820
92. Shaukat A, Levitt MD, Taylor BC, MacDonald R, Shamliyan TA, Kane RL, Wilt TJ. Systematic review: effective management strategies for lactose intolerance. *Ann Intern Med* (2010) 152:797–803. doi: 10.7326/0003-4819-152-12-201006150-00241
93. Roszkowska A, Pawlicka M, Mroczek A, Bałabuszek K, Nieradko-Iwanicka B. Non-Celiac Gluten Sensitivity: A Review. *Medicina (Kaunas)* (2019) 55:222. doi: 10.3390/medicina55060222
94. Carroccio A, Mansueto P, Iacono G, Soresi M, D’Alcamo A, Cavataio F, Brusca I, Florena AM, Ambrosiano G, Seidita A, et al. Non-Celiac Wheat Sensitivity Diagnosed by Double-Blind Placebo-Controlled Challenge: Exploring a New Clinical Entity. *Official journal of the American College of Gastroenterology | ACG* (2012) 107:1898. doi: 10.1038/ajg.2012.236
95. Uhde M, Ajamian M, Caio G, De Giorgio R, Indart A, Green PH, Verna EC, Volta U, Alaedini A. Intestinal cell damage and systemic immune activation in individuals reporting sensitivity to wheat in the absence of coeliac disease. *Gut* (2016) 65:1930–1937. doi: 10.1136/gutjnl-2016-311964
96. Skodje GI, Sarna VK, Minelle IH, Rolfsen KL, Muir JG, Gibson PR, Veierød MB, Henriksen C, Lundin KEA. Fructan, Rather Than Gluten, Induces Symptoms in Patients With Self-Reported Non-Celiac Gluten Sensitivity. *Gastroenterology* (2018) 154:529–539.e2. doi: 10.1053/j.gastro.2017.10.040
97. Ensari A, Marsh MN. Diagnosing celiac disease: A critical overview. *Turk J Gastroenterol* (2019) 30:389–397. doi: 10.5152/tjg.2018.18635

98. Singh SK, Sarma MS. Hereditary fructose intolerance: A comprehensive review. *World J Clin Pediatr* (2022) 11:321–329. doi: 10.5409/wjcp.v11.i4.321
99. Debray F-G, Seyssel K, Fadeur M, Tappy L, Paquot N, Tran C. Effect of a high fructose diet on metabolic parameters in carriers for hereditary fructose intolerance. *Clinical Nutrition* (2021) 40:4246–4254. doi: 10.1016/j.clnu.2021.01.026
100. Adamowicz M, Płoski R, Rokicki D, Morava E, Gizewska M, Mierzewska H, Pollak A, Lefeber DJ, Wevers RA, Pronicka E. Transferrin hypoglycosylation in hereditary fructose intolerance: Using the clues and avoiding the pitfalls. *Journal of Inherited Metabolic Disease* (2007) 30:407. doi: 10.1007/s10545-007-0569-z
101. Ali M, Rellos P, Cox TM. Hereditary fructose intolerance. *J Med Genet* (1998) 35:353–365. doi: 10.1136/jmg.35.5.353
102. Tolan DR, Brooks CC. Molecular analysis of common aldolase B alleles for hereditary fructose intolerance in North Americans. *Biochem Med Metab Biol* (1992) 48:19–25. doi: 10.1016/0885-4505(92)90043-x
103. Ebert K, Witt H. Fructose malabsorption. *Mol Cell Pediatr* (2016) 3:10. doi: 10.1186/s40348-016-0035-9
104. Hammer HF, Fox MR, Keller J, Salvatore S, Basilisco G, Hammer J, Lopetuso L, Benninga M, Borrelli O, Dumitrascu D, et al. European guideline on indications, performance, and clinical impact of hydrogen and methane breath tests in adult and pediatric patients: European Association for Gastroenterology, Endoscopy and Nutrition, European Society of Neurogastroenterology and Motility, and European Society for Paediatric Gastroenterology Hepatology and Nutrition consensus. *United European Gastroenterol J* (2021) 10:15–40. doi: 10.1002/ueg2.12133
105. Komericki P, Akkilic-Materna M, Strimitzer T, Weyermair K, Hammer HF, Aberer W. Oral xylose isomerase decreases breath hydrogen excretion and improves gastrointestinal symptoms in fructose malabsorption – a double-blind, placebo-controlled study. *Alimentary Pharmacology & Therapeutics* (2012) 36:980–987. doi: 10.1111/apt.12057
106. Gibson PR, Shepherd SJ. Evidence-based dietary management of functional gastrointestinal symptoms: The FODMAP approach. *Journal of Gastroenterology and Hepatology* (2010) 25:252–258. doi: 10.1111/j.1440-1746.2009.06149.x
107. Tuck CJ, Biesiekierski JR, Schmid-Grendelmeier P, Pohl D. Food Intolerances. *Nutrients* (2019) 11:1684. doi: 10.3390/nu11071684
108. Frissora CL, Rao SSC. Sucrose intolerance in adults with common functional gastrointestinal symptoms. *Proc (Bayl Univ Med Cent)* (2022) 35:790–793. doi: 10.1080/08998280.2022.2114070
109. Treem WR, McAdams L, Stanford L, Kastoff G, Justinich C, Hyams J. Sacrosidase therapy for congenital sucrase-isomaltase deficiency. *J Pediatr Gastroenterol Nutr* (1999) 28:137–142. doi: 10.1097/00005176-199902000-00008
110. Robayo-Torres CC, Opekun AR, Quezada-Calvillo R, Villa X, Smith EO, Navarrete M, Baker SS, Nichols BL. 13C-breath tests for sucrose digestion in congenital sucrase isomaltase-deficient and sacrosidase-supplemented patients. *J Pediatr Gastroenterol Nutr* (2009) 48:412–418. doi: 10.1097/mpg.0b013e318180cd09
111. Aljaaly EA, Khatib MA. Exploring the Prevalence of Functional Gastrointestinal Diseases and the Accompanied Differences in Dietary and Lifestyle Patterns: A Two-Generational Study. *Diagnostics* (2024) 14:1630. doi: 10.3390/diagnostics14151630
112. Montoro-Huguet MA, Belloc B, Domínguez-Cajal M. Small and Large Intestine (I): Malabsorption of Nutrients. *Nutrients* (2021) 13:1254. doi: 10.3390/nu13041254
113. Puntis JW, Zamvar V. Congenital sucrase-isomaltase deficiency: diagnostic challenges and response to enzyme replacement therapy. (2015) doi: 10.1136/archdischild-2015-308388
114. Marcadier JL, Boland M, Scott CR, Issa K, Wu Z, McIntyre AD, Hegele RA, Geraghty MT, Lines MA. Congenital sucrase-isomaltase deficiency: identification of a common Inuit founder mutation. *CMAJ* (2015) 187:102–107. doi: 10.1503/cmaj.140657
115. Latorre-Moratalla ML, Comas-Basté O, Bover-Cid S, Vidal-Carou MC. Tyramine and histamine risk assessment related to consumption of dry fermented sausages by the Spanish population. *Food and Chemical Toxicology* (2017) 99:78–85. doi: 10.1016/j.fct.2016.11.011
116. Comas-Basté O, Sánchez-Pérez S, Veciana-Nogués MT, Latorre-Moratalla M, Vidal-Carou M del C. Histamine Intolerance: The Current State of the Art. *Biomolecules* (2020) 10:1181. doi: 10.3390/biom10081181

117. Schwelberger HG, Feurle J, Houen G. Mapping of the binding sites of human diamine oxidase (DAO) monoclonal antibodies. *Inflamm Res* (2018) 67:245–253. doi: 10.1007/s00011-017-1118-3
118. Comas-Basté O, Sánchez-Pérez S, Veciana-Nogués MT, Latorre-Moratalla M, Vidal-Carou M del C. Histamine Intolerance: The Current State of the Art. *Biomolecules* (2020) 10:1181. doi: 10.3390/biom10081181
119. Schnedl WJ, Lackner S, Enko D, Schenk M, Holasek SJ, Mangge H. Evaluation of symptoms and symptom combinations in histamine intolerance. *Intest Res* (2019) 17:427–433. doi: 10.5217/ir.2018.00152
120. Schnedl WJ, Lackner S, Enko D, Schenk M, Mangge H, Holasek SJ. Non-celiac gluten sensitivity: people without celiac disease avoiding gluten—is it due to histamine intolerance? (2017) doi: https://www.researchgate.net/publication/321310010_Non-celiac_gluten_sensitivity_People_without_celiac_disease_avoiding_gluten_-_is_it_due_to_histamine_intolerance
121. Reese I, Ballmer-Weber B, Beyer K, Dölle-Bierke S, Kleine-Tebbe J, Klimek L, Lämmel S, Lepp U, Saloga J, Schäfer C, et al. Guideline on management of suspected adverse reactions to ingested histamine: Guideline of the German Society for Allergology and Clinical Immunology (DGAKI), the Society for Pediatric Allergology and Environmental Medicine (GPA), the Medical Association of German Allergologists (AeDA) as well as the Swiss Society for Allergology and Immunology (SGAI) and the Austrian Society for Allergology and Immunology (ÖGAI). *Allergologie Select* (2021) 5:305. doi: 10.5414/ALX02269E
122. San Mauro Martin I, Brachero S, Garicano Vilar E. Histamine intolerance and dietary management: A complete review. *Allergol Immunopathol (Madr)* (2016) 44:475–483. doi: 10.1016/j.aller.2016.04.015
123. Hasler WL, Grabauskas G, Singh P, Owyang C. Mast cell mediation of visceral sensation and permeability in irritable bowel syndrome. *Neurogastroenterol Motil* (2022) 34:e14339. doi: 10.1111/nmo.14339
124. Schnedl WJ, Schenk M, Lackner S, Enko D, Mangge H, Forster F. Diamine oxidase supplementation improves symptoms in patients with histamine intolerance. *Food Sci Biotechnol* (2019) 28:1779–1784. doi: 10.1007/s10068-019-00627-3
125. Maintz L, Novak N. Histamine and histamine intolerance. *Am J Clin Nutr* (2007) 85:1185–1196. doi: 10.1093/ajcn/85.5.1185
126. Jochum C. Histamine Intolerance: Symptoms, Diagnosis, and Beyond. *Nutrients* (2024) 16:1219. doi: 10.3390/nu16081219
127. Sánchez-Pérez S, Comas-Basté O, Duelo A, Veciana-Nogués MT, Berlanga M, Vidal-Carou MC, Latorre-Moratalla ML. The dietary treatment of histamine intolerance reduces the abundance of some histamine-secreting bacteria of the gut microbiota in histamine intolerant women. A pilot study. *Front Nutr* (2022) 9: doi: 10.3389/fnut.2022.1018463
128. Ispiryan L, Zannini E, Arendt EK. FODMAP modulation as a dietary therapy for IBS: Scientific and market perspective. *Comprehensive Reviews in Food Science and Food Safety* (2022) 21:1491–1516. doi: 10.1111/1541-4337.12903
129. Morariu I-D, Avasilcai L, Vieriu M, Lupu VV, Morariu B-A, Lupu A, Morariu P-C, Pop O-L, Starcea IM, Trandafir L. Effects of a Low-FODMAP Diet on Irritable Bowel Syndrome in Both Children and Adults—A Narrative Review. *Nutrients* (2023) 15:2295. doi: 10.3390/nu15102295
130. Lenhart A, Chey WD. A Systematic Review of the Effects of Polyols on Gastrointestinal Health and Irritable Bowel Syndrome. *Adv Nutr* (2017) 8:587–596. doi: 10.3945/an.117.015560
131. Sultan N, Varney JE, Halmos EP, Biesiekierski JR, Yao CK, Muir JG, Gibson PR, Tuck CJ. How to Implement the 3-Phase FODMAP Diet Into Gastroenterological Practice. *J Neurogastroenterol Motil* (2022) 28:343–356. doi: 10.5056/jnm22035
132. Bellini M, Tonarelli S, Nagy AG, Pancetti A, Costa F, Ricchiuti A, de Bortoli N, Mosca M, Marchi S, Rossi A. Low FODMAP Diet: Evidence, Doubts, and Hopes. *Nutrients* (2020) 12:148. doi: 10.3390/nu12010148
133. Gwóźdź W, Krupa-Kotara K, Całyniuk B, Helisz P, Grajek M, Głogowska-Ligus J. Traditional, Vegetarian, or Low FODMAP Diets and Their Relation to Symptoms of Eating Disorders: A Cross-Sectional Study among Young Women in Poland. *Nutrients* (2022) 14:4125. doi: 10.3390/nu14194125
134. So D, Loughman A, Staudacher HM. Effects of a low FODMAP diet on the colonic microbiome in irritable bowel syndrome: a systematic review with meta-analysis. *Am J Clin Nutr* (2022) 116:943–952. doi: 10.1093/ajcn/nqac176

135. Staudacher HM, Lomer MCE, Farquharson FM, Louis P, Fava F, Franciosi E, Scholz M, Tuohy KM, Lindsay JO, Irving PM, et al. A Diet Low in FODMAPs Reduces Symptoms in Patients With Irritable Bowel Syndrome and a Probiotic Restores Bifidobacterium Species: A Randomized Controlled Trial. *Gastroenterology* (2017) 153:936–947. doi: 10.1053/j.gastro.2017.06.010
136. Oka P, Parr H, Barberio B, Black CJ, Savarino EV, Ford AC. Global prevalence of irritable bowel syndrome according to Rome III or IV criteria: a systematic review and meta-analysis. *The Lancet Gastroenterology & Hepatology* (2020) 5:908–917. doi: 10.1016/S2468-1253(20)30217-X
137. Lacy BE, Mearin F, Chang L, Chey WD, Lembo AJ, Simren M, Spiller R. Bowel Disorders. *Gastroenterology* (2016) 150:1393-1407.e5. doi: 10.1053/j.gastro.2016.02.031
138. Ford AC, Sperber AD, Corsetti M, Camilleri M. Irritable bowel syndrome. *The Lancet* (2020) 396:1675–1688. doi: 10.1016/S0140-6736(20)31548-8
139. Ford AC, Forman D, Bailey AG, Axon ATR, Moayyedi P. Irritable bowel syndrome: a 10-yr natural history of symptoms and factors that influence consultation behavior. *Am J Gastroenterol* (2008) 103:1229–1239; quiz 1240. doi: 10.1111/j.1572-0241.2007.01740.x
140. Yarandi SS, Nasser-Moghaddam S, Mostajabi P, Malekzadeh R. Overlapping gastroesophageal reflux disease and irritable bowel syndrome: Increased dysfunctional symptoms. *World J Gastroenterol* (2010) 16:1232–1238. doi: 10.3748/wjg.v16.i9.1232
141. Ford AC, Moayyedi P, Lacy BE, Lembo AJ, Saito YA, Schiller LR, Soffer EE, Spiegel BMR, Quigley EMM, Task Force on the Management of Functional Bowel Disorders. American College of Gastroenterology monograph on the management of irritable bowel syndrome and chronic idiopathic constipation. *Am J Gastroenterol* (2014) 109 Suppl 1:S2-26; quiz S27. doi: 10.1038/ajg.2014.187
142. Matheis A, Martens U, Kruse J, Enck P. Irritable bowel syndrome and chronic pelvic pain: A singular or two different clinical syndrome? *World J Gastroenterol* (2007) 13:3446–3455. doi: 10.3748/wjg.v13.i25.3446
143. Barsky AJ. Assessing the New DSM-5 Diagnosis of Somatic Symptom Disorder. *Psychosom Med* (2016) 78:2–4. doi: 10.1097/PSY.0000000000000287
144. Koloski NA, Jones M, Weltman M, Kalantar J, Bone C, Gowryshankar A, Walker MM, Talley NJ. Identification of early environmental risk factors for irritable bowel syndrome and dyspepsia. *Neurogastroenterol Motil* (2015) 27:1317–1325. doi: 10.1111/nmo.12626
145. Biesiekierski JR, Peters SL, Newnham ED, Rosella O, Muir JG, Gibson PR. No effects of gluten in patients with self-reported non-celiac gluten sensitivity after dietary reduction of fermentable, poorly absorbed, short-chain carbohydrates. *Gastroenterology* (2013) 145:320-328.e1–3. doi: 10.1053/j.gastro.2013.04.051
146. Czaja-Bulsa G. Non celiac gluten sensitivity - A new disease with gluten intolerance. *Clin Nutr* (2015) 34:189–194. doi: 10.1016/j.clnu.2014.08.012
147. Enck P, Aziz Q, Barbara G, Farmer AD, Fukudo S, Mayer EA, Niesler B, Quigley EMM, Rajilić-Stojanović M, Schemann M, et al. Irritable bowel syndrome. *Nat Rev Dis Primers* (2016) 2:16014. doi: 10.1038/nrdp.2016.14
148. Longstreth GF, Thompson WG, Chey WD, Houghton LA, Mearin F, Spiller RC. Functional bowel disorders. *Gastroenterology* (2006) 130:1480–1491. doi: 10.1053/j.gastro.2005.11.061
149. Drossman DA, Tack J, Ford AC, Szigethy E, Törnblom H, Van Oudenhove L. Neuromodulators for Functional Gastrointestinal Disorders (Disorders of Gut-Brain Interaction): A Rome Foundation Working Team Report. *Gastroenterology* (2018) 154:1140-1171.e1. doi: 10.1053/j.gastro.2017.11.279
150. Lacy BE, Mearin F, Chang L, Chey WD, Lembo AJ, Simren M, Spiller R. Bowel Disorders. *Gastroenterology* (2016) 150:1393-1407.e5. doi: 10.1053/j.gastro.2016.02.031
151. Pasta A, Formisano E, Calabrese F, Plaz Torres MC, Bodini G, Marabotto E, Pisciotta L, Giannini EG, Furnari M. Food Intolerances, Food Allergies and IBS: Lights and Shadows. *Nutrients* (2024) 16:265. doi: 10.3390/nu16020265
152. Quigley EMM. Prebiotics and Probiotics in Digestive Health. *Clin Gastroenterol Hepatol* (2019) 17:333–344. doi: 10.1016/j.cgh.2018.09.028

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s)

disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.