

Review

Not peer-reviewed version

Lifestyle and Environmental Factors Affecting Male Fertility, Individual Predisposition, Prevention and Intervention

[Jan Tesarik](#)*

Posted Date: 18 March 2025

doi: 10.20944/preprints202503.1302.v1

Keywords: male fertility; smoking; alcohol; stress; physical activity; obesity; electronic devices; air pollution; heat; harmful chemicals; genetics; epigenetics; systemic disease; infection; prevention; treatment



Preprints.org is a free multidisciplinary platform providing preprint service that is dedicated to making early versions of research outputs permanently available and citable. Preprints posted at Preprints.org appear in Web of Science, Crossref, Google Scholar, Scilit, Europe PMC.

Copyright: This open access article is published under a Creative Commons CC BY 4.0 license, which permit the free download, distribution, and reuse, provided that the author and preprint are cited in any reuse.

Review

Lifestyle and Environmental Factors Affecting Male Fertility, Individual Predisposition, Prevention and Intervention

Jan Tesarik

MARGen (Molecular Assisted Reproduction and Genetics) Clinic, Calle Gracia 36, 18002 Granada, Spain; jantesarik13@gmail.com; Tel.: +34 606376992

Abstract: Current lifestyle brings about increasing prevalence of unhealthy habits that can negatively affect male fertility. Cigarette smoking, alcohol intake, stress, inadequate physical activity, unbalanced diet leading to obesity, and use of mobile telephones and portable electronic devices can affect male reproductive system through multiple mechanisms. Moreover the modern man is often exposed to environmental factors independent of his will, such as air pollution, exposure to heat or toxicants in his workplace, or the presence of harmful chemicals in food, beverages, agricultural and industrial products, etc. The susceptibility to these factors depends on genetic and epigenetic predisposition, potentially present systemic disease and medication, and local affections of genitourinary system. The multifaceted nature of both the causative factors and the susceptibility background makes the resulting fertility disturbance highly individual and variable among different men exposed to the same conditions. This paper critically reviews current knowledge of different causative and susceptibility factors with a special attention to molecular mechanisms of their action. Finally, strategies for prevention of abnormalities due to lifestyle and environmental factors and available treatment modalities for already present abnormalities are exposed.

Keywords: male fertility; smoking; alcohol; stress; physical activity; obesity; electronic devices; air pollution; heat; harmful chemicals; genetics; epigenetics; systemic disease; infection; prevention; treatment

1. Introduction

Lifestyle factors, also referred to as behavioural factors, are those related to the change in people's behaviours and the way they live their life. Together with environmental factors, they can be at the origin of a wide range of noncommunicable diseases (NCDs), such as cardiovascular diseases, type-2 diabetes, chronic obstructive pulmonary disease and some types of cancer, or risk factors behind them [1]. The prevalence and incidence of NCDs shows an increasing trend in different parts of the world [2–5]. In addition to the above systemic NCDs, lifestyle and environmental factors were also reported to affect male fertility [6,7]. Environmental factors implicated in the pathogenesis of male subfertility and infertility act on the background of each individual's susceptibility defined by genetic predisposition, occasional presence of a systemic disease, local affections of genitourinary system and others. This review deals with the most important lifestyle and susceptibility factors, their molecular mechanisms of action, and the current possibilities of prevention and intervention.

2. Contemporary Lifestyle and Male Fertility

There has been a continuous decline of semen quality, including a decrease in semen volume, sperm count, concentration and the percentage of normal forms, over the last decades [8–10]. This trend is supposed to be due to the negative influence of widespread lifestyle-related habits to which the modern man is exposed during his reproductive period [6]. The main negative lifestyle habits

and environmental conditions affecting male fertility include inadequate diet leading to obesity, air pollution, exposure to harmful chemicals (food and drinks, agricultural and industrial products), exposure to heat, cigarette smoking, alcohol intake, stress, inadequate physical activity, and use of mobile telephones and portable computers [6,7,11]. Apart from possible effects on erectile function, most of the lifestyle and environmental factors affecting male fertility impact different aspects of sperm structure and function.

Seminal oxidative stress [12–14] is the main endpoint to which the molecular effects of different harmful factors converge. Oxidative stress arises from conditions in which the production of reactive oxygen species (ROS), needed for normal sperm development and function, exceeds the ROS-scavenging capacity of inherent antioxidative systems. The resulting excessive ROS produce a chain of events leading to damage of sperm lipids, proteins and DNA, ultimately affecting male fertility [15,16]. Basically, a spermatozoon consists of sperm head and sperm tail. The head contains sperm nucleus with its DNA and is covered by the acrosome. The main functional parts of the sperm tail are the midpiece and the principal piece. Mitochondria, contained in the midpiece, produce energy in the form of adenosine triphosphate (ATP) which is mainly used to sustain propulsive forces for sperm cell movement, generated in the axoneme of the principal piece. In addition, ATP is used to power other energy-consuming sperm functions, such as the exocytosis of sperm acrosome (acrosome reaction) required for sperm penetration into the oocyte at fertilization. Harmful external factors can act directly on sperm acrosome and nuclear DNA. But they also can disturb mitochondrial function, leading to oxidative stress (see above) which, in its turn, feeds back into mitochondrial integrity and aggravates nuclear and acrosomal damage (Figure 1).

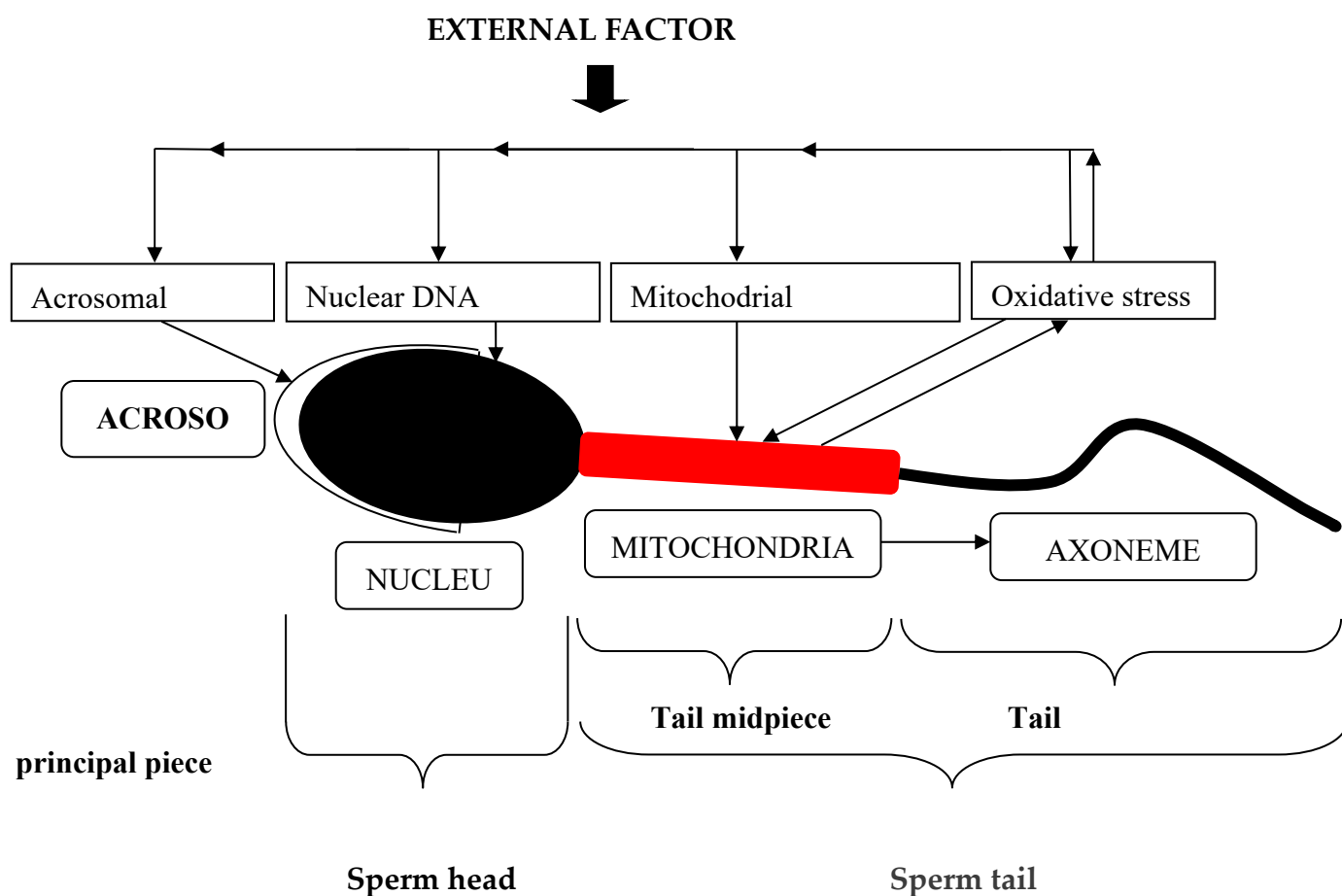


Figure 1. Schematic representation of a human spermatozoon showing how it can be affected by the action of external factors.

2.1. Obesity

Obesity and overweight prevalence is increasing globally and contributes to various human health problems including male infertility [11,17]. Obesity was reported to impair both conventional and biofunctional sperm parameters, and to induce epigenetic changes that can be transferred to offspring [11,17,18]. There are several potential biological mechanisms linking central obesity to reduced semen quality. They include a decrease in serum sex hormone-binding globulin (SHBG), total and free testosterone (T) and inhibin B levels, and increased conversion of T into 17 β -estradiol (E2) [19]. Reduced hepatic insulin clearance, due to increased delivery of free fatty acids released from the adipose tissue to the liver, leads to hyperinsulinemia and insulin resistance [18]. Hyperinsulinemia dysregulates the endocrine activity of hypertrophied adipocytes by stimulating the production of non-esterified fatty acids which increase the secretion of tumour necrosis factor α (TNF α) by the adipocytes. TNF α , in turn, potentiates the secretion of adipokines (mainly leptin) and other pro-inflammatory cytokines. The pro-inflammatory cytokines, mainly interleukin 1 β (IL1 β), interleukin 6 (IL6), as well as acute phase proteins and chemokines, mainly C-C motif chemokine ligand-2 (CCL2) and monocyte chemoattractant protein-1 (MCP-1), attract more monocytes/macrophages within adipose tissue [20]. The resulting vicious cycle promotes the inflammation of the adipose tissue first and then a systematic state of low-grade inflammation [21]. The increased production of adipokines and pro-inflammatory cytokines significantly influences insulin signaling in insulin-responsive tissues, leading to systemic insulin resistance and consequent disturbance of gonadotropin-releasing hormone (GnRH) secretion by hypothalamic neurons [22]. In addition to acting through endocrine imbalance, insulin resistance also inhibits spermatogenesis by increasing nuclear and mitochondrial DNA damage [23]. A pivotal role is also played by heat-induced damage due to fat accumulation in the suprapubic region and around the pampiniform plexus. The resulting increase in scrotal temperature reduces sperm concentration and motility and enhances oxidative stress and sperm DNA fragmentation [24].

As to the epigenetic changes that can be transferred to offspring, epidemiological studies have shown that children born from obese fathers are more likely to be obese [25]. Among the most relevant epigenetic mechanisms that are involved in gene activity regulation (DNA methylation, histone modifications, and non-coding RNAs), DNA methylation was most widely studied in the context of obesity [18,26]. Significantly altered DNA methylation in the regulatory region of many imprinted genes has been reported in spermatozoa from overweight and obese men as compared to normal-weight men [27]. The main genes that undergo abnormal epigenetic modification (hypomethylation) of differentially methylated regions in obese males include maternally expressed gene 3 (*MEG3*), *neccin (NDN)*, small nuclear ribonucleoprotein polypeptide N (*SNRPN*), and sarcoglycan epsilon (*SGCE*)/paternally expressed gene 10 (*PEG10*), while a slight increase in DNA methylation has been detected in *H19* gene [28]. The hypomethylation of imprinted genes was associated with higher levels of seminal oxidative stress, sperm DNA fragmentation and decreased pregnancy rates [29,30].

2.2. Air Pollution

Air pollution, mainly arising from motor vehicle exhaust, factories, fire, household, agriculture, waste treatment, oil refineries, and natural sources, such as volcanic eruptions, is increasing steadily [31]. In addition to causing a number of unrelated diseases, it also affects male fertility [32–34]. The main air pollutants include particulate matter, volatile organic compounds, ozone, nitrogen oxides, sulfur dioxide, carbon monoxide, polycyclic aromatic hydrocarbons (PAHs), and various types of radiation (such as X-ray exposure) [35]. Impairment of several parameters of male reproductive function in response to various air pollutants has been reported. In 2018, a critical review of 22 studies dealing with effects of various air pollutants on semen quality, has suggested that all of the pollutants studied (airborne particular matter, nitric oxides, sulfur dioxide, PAHs, ozone, carbon monoxide, heavy metals) can affect sperm motility, morphology and DNA integrity [36]. Exposure to air pollutants is often occupational, for instance in men working as toll collectors at motorways. Actually,

one study has reported a significant decrease in total sperm count, total and progressive motility, the percentage of spermatozoa with normal morphology and that of spermatozoa with normal chromatin and intact DNA in toll collectors as compared to unexposed healthy men [37]. A significant direct correlation was found between spermatozoa with damaged chromatin or fragmented DNA and the length of occupational exposure, suggesting a time-dependent relationship [37].

The mechanisms by which air pollutants affect male fertility are not completely clear and have only been studied for some of them (PAHs, heavy metals, particular matter, sulfur dioxide, carbon monoxide, nitric oxides). PAHs and heavy metals (lead, zinc, copper) from car exhaust can have estrogenic, antiestrogenic and androgenic actions, and they may also cause hormonal disruption leading to abnormal gonadal steroidogenesis and spermatogenesis [38,39]. Furthermore, exposure to PAHs can also result in changes in gene expression and DNA methylation [40]. Particular matter, sulfur dioxide, carbon monoxide and nitric oxides stimulate the production of reactive oxygen species which cause oxidative stress leading to lipid peroxidation and sperm DNA fragmentation [39]. Further research into the molecular mechanisms underlying the effects of air pollutants on male fertility is warranted.

2.3. Harmful Chemicals (Other Than Air Pollutants)

Food, drink, and various agricultural and industrial products are the main sources of potentially harmful chemicals to which the contemporary man can be exposed. The main harmful chemicals and their sources have been reviewed recently [31]. They include dioxins, bisphenols, pesticides and herbicides, phthalates, and heavy metals (Table 1).

Dioxins are highly persistent by-products of several industrial processes, such as smelting, chlorine bleaching of paper, production of some pesticides, or biomedical and plastic waste incineration [35]. Their effects were best studied in residents of Seveso (Italy) exposed to 2,3,7,8-tetrachlorodibenzo-*p*-dioxin released accidentally from a trichlorophenol manufacturing plant in 1976 [41]. Negative effects on sperm quality (count, motility, progressive motility) were observed mainly after exposure of children. Dioxins act as endocrine disruptors, and their toxic effects are mediated by binding to the aryl hydrocarbon receptor (AHR)/aryl hydrocarbon receptor nuclear translocator (ARNT) receptor complex present in human testicular cells [42].

Bisphenols are a group of industrial chemical compounds related to diphenylmethane, commonly used in the creation of plastics and epoxy resins; they are released into the environment during the process of production, use, or disposal of plastics and result from breakdown of plastic-related wastes [35]. In particular, bisphenol A (BPA) was shown to affect male fertility as an endocrine disruptor. It impairs spermatogenesis by antagonizing the androgen receptor, and it also displays estrogenic and antithyroid actions [43,44]. Other studies reported BPA to disturb sperm function through inducing premature sperm acrosome reaction and to decrease sperm motility by reducing adenosine triphosphate (ATP) levels in spermatozoa [45]. Moreover, BPA causes activation of proapoptotic signaling pathways, including mitogen-activated protein kinase (MAPK), Fas/FasL, Caspases 3 and 9, and Bax, leading to diminished proliferation, increased reactive oxygen species-mediated damage and enhanced apoptosis of male germ cells [46]. Exposure to BPA was also shown to lead to sperm DNA damage, mitochondrial dysfunction and degeneration, and increased risk of aneuploidies in spermatozoa [44,47].

Pesticides can cause direct sperm damage or disrupt the endocrine function needed for spermatogenesis at any stage of hormonal regulation (hormone synthesis, release, storage, transport and clearance, receptor recognition and binding); in addition to the reproductive system, they can also affect thyroid function and the central nervous system [48]. The endocrine-disrupting action of organochlorine pesticides, the most widely used ones, is well-documented [49]. The use of dichlorodiphenyltrichloroethane (DDT), the most notorious organochlorine pesticide, is currently restricted in most countries. However, due to its stability and its capacity to accumulate in adipose tissue, DDT can persist in the human organism even for decades after exposure, leading to the conclusion that "there is now not a single living organism on the planet that does not contain DDT"

[49]. Most of the endocrine-disrupting action of DDT is due to its main metabolite, *p,p'*-dichlorodiphenyl-dichloroethylene (*p,p'*-DDE), which binds to androgen receptors and thus inhibits the action of testosterone [50]. *p,p'*-DDE may have an additive or multiplicative effect with other endocrine-disrupting environmental pollutants, potentiating their adverse impacts on reproductive functions [49]. Exposure of spermatozoa to *p,p'*-DDE was also reported to promote mitochondrial Ca^{2+} overload, a decrease in mitochondrial membrane potential, an increase in reactive oxygen species (ROS) production and sperm DNA fragmentation, which collectively lead to a general mitochondrial dysfunction and cellular ATP depletion with a remarkable decrease in sperm motility and fertilizing ability [51,52]. DDT and other organochlorine chemicals enter the food chain and can be transported over long distances [53].

Table 1. Overview of the most important harmful chemicals, their sources, effects, and the mechanisms of action.[41,42]

Name	Sources	Effects	Mechanism of action	References
Dioxins	Industrial accidents	↓ Sperm count ↓ Sperm motility	Endocrine disruption	[41,42]
Bisphenols	Plastics	↓ Spermatogenesis ↓ Sperm motility ↓ Sperm function ↑ Sperm apoptosis ↑ Sperm aneuploidy	Endocrine disruption Oxidative stress ↑ MAPK ↑ Fas/FasL ↑ Caspases 3, 9 ↑ Bax ↑ DNA damage ↑ Mitochondrial damage	[44–47]
Pesticides and herbicides	Agricultural products	↓ Spermatogenesis ↓ Sperm motility ↓ Sperm count ↑ Sperm apoptosis	Endocrine disruption Oxidative stress ↑ DNA damage ↑ Mitochondrial dysfunction	[48–55]
Phthalates	Various industrial goods	↓ Spermatogenesis ↓ Sperm motility ↓ Sperm count ↑ Teratozoospermia	Endocrine disruption ↓ Gene transcription ↓ Androgene synthesis	[56–64]
Heavy metals	Various industrial goods	↓ Spermatogenesis ↓ Sperm motility	Endocrine disruption Oxidative stress	[65,66]

		↓ Sperm count	↑ DNA damage	
		↑ Teratozoospermia	↓ Androgene synthesis	
		↓ Sperm viability		

Herbicides have received less attention than pesticides in academic publications. In fact, published data only concern one herbicide named Roundup. Direct exposure of human spermatozoa to the active component of this herbicide (glyphosate), acting as endocrine disrupter, was reported to cause sperm mitochondrial dysfunction and a decrease in sperm motility [54]. Another study, carried out in the rat model, analyzed the molecular mechanisms of glyphosate action in the testis and isolated Sertoli cells [55]. It was shown that glycosate increases intracellular Ca^{2+} concentration by opening L-type voltage-dependent Ca^{2+} channels as well as endoplasmic reticulum inositol trisphosphate (IP3)- and ryanodine receptor-operated channels, leading to Ca^{2+} overload within the rat prepubertal Sertoli cells cells, which sets off oxidative stress and necrotic cell death. These effects were prevented by the antioxidants Trolox and ascorbic acid [55]. Activated protein kinase C, phosphatidylinositol 3-kinase, and the mitogen-activated protein kinases, such as ERK1/2 and p38MAPK, were shown to play a role in eliciting the glycosate-induced Ca^{2+} influx. Also, glyphosate decreases the levels of reduced glutathione (GSH) and increases the amounts of thiobarbituric acid-reactive species (TBARS) and protein carbonyls. In fact, simultaneous stimulating effects on the activities of glutathione peroxidase, glutathione reductase, glutathione S-transferase, γ -glutamyltransferase, catalase, superoxide dismutase, and glucose-6-phosphate dehydrogenase potentiate the consequences of downregulated GSH levels, further decrease GSH and increase the amounts of TBARS and protein carbonyls [55].

Phthalates, also known as phthalic acid diesters, are man-made chemicals that are used in several consumer and industrial goods [56]. They were found in many consumer products such as toys, pharmaceuticals, cosmetic products, building and construction materials, scent retainers, some medications, personal care products, etc. [57]. Male partners of infertile couples, who have been exposed to phthalate drugs, have poor sperm quality as compared to unexposed men [57,58]. In particular, exposure to phthalates decreases semen volume, total sperm count, and sperm concentration, and causes diverse morphological abnormalities of the sperm head [59,60], in addition to reducing sperm motility [60,61]. At the molecular level, phthalates, especially mono-(2-ethylhexyl) phthalate (MEHP), an active metabolite of di-2-ethylhexyl phthalate (DEHP), cause activation of peroxisome proliferator-activated receptor (PPAR) α and γ pathways [62]. The activation of both pathways stimulates PPAR:RXR (retinoid X receptor) heterodimers that compete for DNA binding sites required for gene transcription, impeding the synthesis of aromatase enzyme involved in sexual development [63]. In addition, MEHP decreases the production of steroidogenic proteins including steroidogenic acute regulatory (StAR), and cytochrome P450 side-chain cleavage (P450scc) proteins [31]. At high levels, it also inhibits the activity of 3β -hydroxysteroid dehydrogenases (3β -HSD) and 17β -hydroxysteroid dehydrogenases (17β -HSD) specific to Leydig cell function by increasing oxidative stress in Leydig cells; this inhibitory action leads to a decrease in testosterone synthesis [64].

Heavy metals, such as lead, cadmium, arsenic, mercury, barium, uranium and others, were also reported to reduce the semen and sperm quality in men. Data were published showing a close association between the presence of cadmium and barium in blood, and lead, cadmium, barium and uranium in seminal plasma, on the one hand, and increased risk for reduced sperm count, viability and motility, and abnormal sperm morphology, on the other hand [65]. An increase in reactive oxygen species (ROS) generation, leading to excessive lipis peroxidation and sperm DNA damage, is the main mechanism by which heavy metals affect male infertility [64], although they also act as endocrine disrupters which affect semen quality by altering the production of androgens and their action in the testis [31].

2.4. Heat

Excessive heat exposures in the workplace, or those caused by elective lifestyle habits, are known factors affecting spermatogenesis. The question of whether global warming also can play a role is to be evaluated with interest in future studies. In fact, any factor that causes a rise in scrotal temperature (normally 2–4°C lower than the core body temperature) can be blamed [67–69]. Even moderate scrotal temperature rise (1–1.5°C) was shown to impair sperm production (oligozoospermia, azoospermia) and to cause sperm morphological abnormalities (teratozoospermia) [70]. Alteration of spermatogenesis in men with a varicocele is actually related to scrotal temperature increase [68].

Exposure to heat can be self-imposed or nonelective, professional or environmental, transient or durable. Self-imposed exposures are related to elective lifestyle habits. As a matter of example, a study has revealed that the frequent use of tight underwear in men can increase scrotal temperature leading to decreased sperm concentration, total sperm count, motility, and male infertility [71]. Frequent exposure to wet heat (hot tubs, Jacuzzi, etc) also affects sperm count and motility and can disturb fertility in males [72,73]. Unlike self-imposed heat exposure, there are also conditions of nonelective exposures, mainly professional and environmental ones. Professional exposure to radiant heat, as seen in cases of people working in furnaces, bakeries, welding or ceramic factories, and in those working for long hours in kitchens, laundries, or dry cleaning, was shown to affect one or more components of semen quality in males [69,70,74]. Chronic environmental exposure to heat has been increasing over the past decades due to climate change. Inverse relationship between ambient temperature and sperm quality has been documented. Sperm concentration, motility and total amount per ejaculate are significantly lower in summer and higher in winter [75]. Another study, also including sperm motility evaluation, reported similar trends and suggested the winter and spring semen patterns to be compatible with increased fecundability, which may be a plausible explanation of the peak number of deliveries during the fall [76]. The consequences of self-imposed heat exposure are usually reversible and disappear when the causative lifestyle habit is corrected. This was demonstrated in some, but not all, patients who responded favourably to cessation of voluntary heat exposure (hot baths, Jacuzzi) [72,73]. On the other hand, it is evident that nonelective exposures, both professional ones and those related to non-modifiable environmental factors, are more difficult to control.

There appear to be several mechanisms mediating the effect of heat on spermatozoa. It was observed that higher scrotal temperatures result in a rise in testicular metabolism without the corresponding surge in blood supply, leading to local tissue hypoxia and oxidative stress [77]. Owing to the high levels of polyunsaturated fatty acids in the sperm plasma membrane [78], exposure to oxidative stress causes increased production of reactive oxygen species leading to sperm DNA fragmentation [79]. Independently, excessive heat exposure decreases sperm motility by downregulating mitochondrial activity via activation of glycogen synthase kinase-3 α and inhibition of mitochondrial protein import, thus reducing ATP levels [80]. Finally, even a transient rise in scrotal temperature decreases the level of the anti-apoptotic *B-cell lymphoma 2 (Bcl-2)* gene expression, which facilitates sperm apoptotic pathways [81].

2.5. Cigarette Smoking

Tobacco smoke is a toxic and carcinogenic mixture of more than 5,000 chemicals [82] some of which, including nicotine and its metabolites, carbon monoxide, benzopyrene, and cadmium, may have harmful effects on male germ cells [6]. A meta-analysis comprising 5865 men shows that cigarette smoking is associated with reduced sperm count and motility as well as overall deterioration of semen quality, this trend being more pronounced in moderate and heavy smokers than in occasional ones [83]. In addition to all conventional semen parameters, sperm chromatin condensation and sperm viability are also affected in smokers, proportionally to the number of cigarettes smoked per day and the duration of smoking [84,85]. Evaluation of the mechanisms underlying the effects of smoking on spermatozoa is complicated because of the presence of multiple potentially harmful chemicals in cigarette smoke. Among them, polycyclic aromatic hydrocarbons

(PAHs) display a preeminent role in accelerating germ cell death across all stages of spermatogenesis. This effect is mediated by the cytoplasmic transcription factor aryl hydrocarbon receptor (AHR) whose interaction with cigarette smoke components adversely affects the expression of genes associated with antioxidant mechanisms, cell proliferation and apoptosis, and cell cycle progression [86]. Besides, cigarette smoke-mediated crosstalk between AHR and nuclear factor erythroid 2-related factor 2 (NRF2), a transcription factor that is implicated in activating the expression of genes involved in the response to different cellular insults, and that between AHR-NRF2 and mitogen-activated protein kinase (MAPK) pathways induces cell death of spermatocytes [86]. Animal studies (rat) suggest that, like traditional smoking, chemicals derived from electronic cigarette vapours also have adverse effects on male fertility and semen parameters [87].

2.6. Alcohol Intake

Chronic heavy drinking, especially when associated with cigarette smoking, is linked to lower testosterone levels leading to impaired semen quality, while moderate alcohol consumption may not have significant adverse effects [88]. Daily use of alcohol by men of reproductive age was shown to affect semen volume and sperm morphology [89], to cause hormone imbalance [90], and to lengthen time to pregnancy [91]. The mechanisms of the damaging impact of alcohol on fertility are not yet fully discovered. Alcohol drinking was reported to cause a hormonal shift towards higher free estradiol/free testosterone ratio [92] and to augment the level of reactive oxygen species and DNA damage in male germ cells [93].

2.7. Psychological Stress

Stress can result from both physical and psychological conditions. Physical stress is dealt with in section 2.8. of this article. Prolonged exposure to psychological stress can reduce testosterone levels, impair sperm production, and decrease libido, in addition to promoting the acquisition of unhealthy behaviors, such as poor diet, lack of exercise, smoking, and excessive alcohol consumption [94]. Elevated levels of stress hormones, particularly cortisol, were shown to interfere with the secretion of gonadotropin-releasing hormone (GnRH) from the hypothalamus, thus decreasing the release from the pituitary gland of luteinizing hormone (LH) and follicle-stimulating hormone (FSH), both essential for stimulating the testis to produce testosterone needed for spermatogenesis [95,96]. High cortisol levels can also make germ cells more vulnerable to oxidative stress, causing sperm DNA damage and reduction of sperm motility and viability [97].

2.8. Inadequate Physical Activity

Research suggests that both insufficient and excessive physical activity can cause male fertility issues, although the connection with male fertility is complex and multifaceted [98]. Resistance exercise training was reported to improve markers of male fertility and reproductive capacity in infertile patients [99], while excessive exercise, such as intense training for marathons or Olympic events, can potentially lower testosterone levels, reduce sperm quality, and lead to infertility [100]. At appropriate levels, physical exercise attenuates inflammation, as reflected by reduced seminal pro-inflammatory cytokines (IL-1 β , IL-6, IL-8 and TNF- α), decreases the levels of oxidative stress markers (malondialdehyde and 8-isoprostane) and enhances the production of antioxidants (superoxide dismutase and catalase) that protect sperm DNA integrity [99]. However, the available studies have limitations, and more research is needed to fully understand the specific effects of different regimens of exercise and their impact on fertility status [98].

2.9. Use of Mobile Telephones and Portable Computers

Mobile telephones and other similar electronic devices may affect male fertility both through a thermal effect and a nonthermal effect. The thermal effect is related to the fact that mobile phones are often carried in trouser pockets near external male reproductive organs [101], and portable computers

are often placed on the user's lap (laptop computers). The consequences of testis exposure to heat are reviewed in section 2.4. of this article. The nonthermal effects of portable electronic devices are due to low-level radio-frequency electromagnetic fields (RF-EMF) [6]. A meta-analysis of ten studies, including 1492 samples, has concluded that exposure to RF-EMF negatively affects parameters of human sperm quality, mainly sperm motility and viability, while effects on sperm concentration were more equivocal [102]. Another meta-analysis, including eighteen studies with 3947 men and 186 rats, indicated that RF-EMF had detrimental effects on sperm motility and viability in human in-vitro studies and on sperm concentration and motility in animal studies, while no relationship with semen parameters was found in human in-vivo studies [103]. It was also reported that ex vivo exposure of human spermatozoa to a wireless internet-connected laptop decreased motility and induced DNA fragmentation by a nonthermal effect [104,105]. It was suggested that sperm exposure to electromagnetic fields may cause oxidation of phospholipids and produce high seminal reactive oxygen species (ROS) levels [6,104], but further research is still needed.

3. Individual Susceptibility to Negative Lifestyle and Environmental Factors

Susceptibility to negative lifestyle and environmental factors is highly individual and mainly conditioned by genetic predisposition, sperm epigenetics, systemic disease, and local affection of genitourinary system (Table 2).

3.1. Genetic Predisposition

Genome alterations can cause by themselves different pathological phenotypes. However, the penetrance of a number of genome alterations is incomplete and the development of the corresponding phenotype depends on other (epigenetic and environmental) factors. In other words, genetic and epigenetic predisposition can help explain why certain individuals exposed to the same load of a lifestyle or environmental harmful factor exhibit symptoms or signs of disease, while others do not [106]. For instance, men who are homozygous null for glutathione S-transferase mu 1 (*GSTM1*) gene have lower capacity to detoxify reactive metabolites of polycyclic aromatic hydrocarbons (PAHs) and are consequently more susceptible to the effects of air pollution on sperm chromatin [107]. Research shows that men with *GSTM1*-null and glutathione S-transferase theta 1 (*GSTT1*)-present genotypes are susceptible to infertility, particularly that resulting from 4-*n*-octylphenol exposure [106]. Moreover, polymorphism in several DNA repair genes, such as X-ray repair cross-complementing protein 1 (*XRCC1*), xeroderma pigmentosum group D 6 (*XPB6*) and 23 (*XPB23*), and cytochrome P450 1A1 (*CYP1A1*), was observed to be associated with high or medium DNA damage after exposure to air pollution [107]. Thus, understanding the underlying genetic mechanisms related to the pathophysiology of male infertility, and the impact of environmental exposures and lifestyle factors on gene expression might aid clinicians in developing individualized treatment strategies [108].

Up to 10% of cases of male subfertility and infertility were shown to have a genetic background [109]. The karyotype can identify numerical chromosome abnormalities, such as Klinefelter's syndrome (XXY), and other male infertility-associated anomalies, such as reciprocal and Robertsonian translocations [110]. Additional molecular analyses are required, even routinely, to identify variants of genes in both the Y chromosome and the autosomes of infertile men. A recent article reviews the known genes in which mutations or gene expression changes are associated with different infertility phenotypes including asthenozoospermia, multiple morphological anomalies of the sperm flagellum, nonobstructive azoospermia, obstructive azoospermia, oligozoospermia, and teratozoospermia [111]. According to the phenotype observed in each individual patient, the screening for deletions in these genes is highly recommended. Sperm mitochondrial DNA (mtDNA) is also prone to mutations and deletions most of which are correlated with elevated oxidative stress and sperm immotility [112–114]. The content of mtDNA in spermatozoa, measured as the ratio between the amount of mtDNA and nuclear DNA, may be used as an indicator for predicting the implication of sperm mitochondria in male infertility [115].

Table 2. Selected intrinsic susceptibility conditions and their interactions with external harmful factors. See main text for spelling out the abbreviations used.

Conditions	Examples	Interactions	References
Genetic	GSTM1 mutation	Decreased air pollutants	[106,107]
	<i>XRCC1</i> , <i>XRCC1</i> , <i>CYP1A1</i>	detoxification	[107]
	polymorphism	High DNA damage by air pollutants	[112–114]
	Mutations and deletions of mtDNA	Susceptibility to oxidative stress	
Epigenetic	<i>H19</i> hypomethylation	Susceptibility to effects of smoking	[121–123]
Systemic disease and medication	Insulin resistance and diabetes	Potential of factors causing oxidative stress and inflammation	[125–128]
	Infectious diseases	Potential of factors causing oxidative stress and inflammation	[136–147]
	Chemotherapeutic and antiepileptic drugs, paracetamol, aspirin, lansoprazole, marijuana	Potential of factors affecting spermatogenesis and sperm motility	[159–164]
Local affections	Semen microbiome	Can exert both potentiating and protective action	[165–174]
	Varicocele, orchitis and prostatitis	Potential of factors causing oxidative stress and inflammation	[175]

3.2. Epigenetic Factors

Various epigenetic factors regulate genes in male germ cells, and alterations of sperm epigenetics, potentially influenced by environmental exposures, can contribute to male infertility [110,116] and congenital disorders in progeny [110,116–120]. Epigenetic marks in mature spermatozoa include post-translational histone modifications, protamines, small non-coding RNA, and DNA methylation, and can be considered as a network that aims to establish and maintain genes' expression status [110,116,118]. A list of imprinted genes whose methylation status (hypomethylation or hypermethylation) is associated with different pathological sperm conditions has been published recently [110]. Among the imprinted genes, *H19* has been highlighted as a potential biomarker for developmental defects in human spermatozoa [121]. Actually, 100% methylation of the *H19*

imprinting control area was shown in normal individuals, while *H19* was hypomethylated in diabetic infertile males [122]. Furthermore, a significant positive correlation was observed between the degree of *H19* hypomethylation and decrease in sperm count and motility, and the risk was potentiated by smoking [123]. Other environmental factors, such as endocrine disrupting chemicals (EDCs), and lifestyle elements also have the potential to affect germline epigenetic markers, with sperm cells being the ultimate recipients of these changes and potential carriers of induced epimutations across generations through a mechanism known as paternal transgenerational epigenetic inheritance [124].

3.3. Systemic Disease and Medication

Interactions between systemic disease and lifestyle factors that influence semen quality have been mainly studied in the context of insulin resistance and diabetes, autoimmunity, systemic infectious diseases, and diseases requiring chronic medication.

Insulin resistance (IR) is a condition in which insulin circulating in blood is unable to properly stimulate glucose uptake and/or use by insulin-sensitive organs and tissues, leading to a compensatory increase in production of insulin in the pancreas and glucose in the liver [125]. IR is known to potentiate the adverse effects of many lifestyle and environmental factors on male fertility [126]. Indexes calculated on the basis of fasting insulin and glucose levels, namely homeostatic model assessment of insulin resistance (HOMA) index and Matsuda index, are the best and most extensively validated indicators for IR diagnosis [127]. Both isolated and accompanied by the action of external factors, IR reduces the production of testosterone, semen volume and the percentage of progressive spermatozoa [128]. Inversely, many lifestyle and environmental factors, such as obesity, smoking, lack of sleep or excessive physical activity, influence both IR and male fertility [16,129–131].

Mechanisms of effects on male fertility (marginally mentioned in section 2.1. of this article) are similar for IR and diabetes and have been reviewed recently [132,133]. Briefly, hyperglycemia reduces the sensitivity of pituitary gland to stimulation by hypothalamic gonadotropin-releasing hormone (GnRH), which reduces the secretion of FSH and LH by the pituitary gland and, consequently, disturbs spermatogenesis and testosterone production by the testis [133]. Also, the state of high blood glucose can shift the balance between oxidation and antioxidant defense in the body in favor of oxidation (oxidative stress) [134], resulting in inflammatory infiltration of neutrophils, increased protease secretion, and the production of a large amount of reactive oxygen species (ROS) and reactive nitrogen species (RNS) [133]. The hyperproduction of ROS and RNS causes oxidative damage in nuclear and mitochondrial DNA and activates the inflammatory signaling cascade in endothelial cells, releasing a large number of pro-inflammatory cytokines into blood [135,136]. Diabetes also affects epigenetic modifications in spermatogenesis [133] and may dysregulate imprinted genes in male germ cells (see Section 3.2. of this paper). In addition to reducing semen quality, diabetes can also cause erectile dysfunction [137,138].

Autoimmunity is another known systemic condition affecting spermatozoa. Antisperm antibodies, found in roughly 6% of infertile men [139], can significantly diminish sperm motility through agglutinating spermatozoa at specific regions (head to head, head to midpiece, head to tail, or non-specific binding), due to the binding of immunoglobulins to sperm surfaces [140]. This autoimmune condition can be idiopathic but is mostly found in homosexual men and patients with varicocele, testicular trauma, mumps, orchitis, congenital absence of the vas deferens, and spinal cord injury, as well as in those who have undergone vasectomy [141].

As to systemic infectious diseases, several bacterial and viral infections are associated with semen quality issues. Among sexually transmitted pathogens that commonly impair parameters of sperm quality, human papilloma virus (HPV) [142,143], herpes simplex virus (HSV) [144], *Ureaplasma urealyticum* [145], *Chlamydia trachomatis* [146], coronavirus disease 2019 (COVID-19) [147,148], Zika virus [149] and adeno-associated virus (AAV) [150] have been pointed out. It has been suggested that microorganisms usually affect the normal function of the male reproductive system through immune responses [151]. In many cases, the numbers of immune cells, and especially mast cells, are increased, and the proliferation of mast cells in the testis interstitium and in walls and

lumens of seminiferous tubules, along with collagen fibers deposition, hinders sperm production [152]. Bacterial infection also increases CD3 helper lymphocytes, B cells and natural killer (NK) cells (CD56), leading to increased level of antisperm antibodies and NK cell-mediated sperm damage [153].

Chronic medication, used to control some systemic diseases, can also be associated with male infertility [154]. In particular, there is sufficient evidence indicating a deleterious effect of chemotherapeutic drugs on spermatogenesis in cancer patients, aggravating the effects caused by the disease itself [155–158]. Some commonly used medications, namely psychotropic drugs (imipramine hydrochloride, desmethylimipramine, chlorpromazine, trifluoperazine, and nortriptyline hydrochloride) [159], antiepileptic drugs (phenytoin, carbamazepine, and valproate) [160], acetaminophen (paracetamol) [161], aspirin, [162], and lansoprazole [163], have adverse affects on sperm quality. In addition, regular consumption of recreational drugs, such as marijuana, was shown to affect both spermatogenesis and sperm motility [164].

3.4. Local Affections of Genitourinary System

Imbalance of semen microbiome, varicocele, orchitis and prostatitis are the most important local factors that may affect male fertility.

Recent research on the semen microbiome, utilizing sequencing technologies, has consistently shown that semen has its own microbiome, which is far more complex than previously thought, encompassing a diverse range of bacteria with both beneficial and detrimental effects on sperm quality and reproductive outcomes [165–167]. Despite its unique nature, semen microbiota shows a high interindividual variability resulting from interplay between environmental factors, personal hygiene habits, and age [168]. There are a number of studies addressing semen microbiota in relation with male fertility and reproductive health. They have been reviewed and critically analyzed recently, and it is shown that, despite much incongruence between data from individual reports, there are also points of agreement [169]. The main bacterial genus/species linked to different pathological conditions include *Neisseria* (semen hyperviscosity and oligoasthenoteratozoospermia) [165], *Klebsiella pneumoniae* (reduced sperm motility and increased apoptosis, semen hyperviscosity [165], *Prevotella* (oligoasthenozoospermia) [170], *Corynebacterium* (asthenoteratozoospermia) [171], *Mycoplasma* and *Ureaplasma* (azoospermia) [172], *Escherichia coli* (sperm acrosome and DNA damage) [173], *Pseudomonas* (oligoasthenoteratozoospermia) [165], and *Ralstonia* and *Stenotrophomonas* (asthenozoospermia) [171]. Interestingly, data obtained by a high-throughput sequencing method targeting V3 and V4 regions of *16S rRNA* gene have shown that, in addition to seminal plasma, the human testis also harbours potential bacterial signature, though in a low-biomass, and could contribute to the seminal microbiome composition [174].

Varicocele orchitis and prostatitis are other local affections of genitourinary tract that cause male infertility. A large number of studies have shown that they cause sperm dysfunction, mainly through oxidative stress, and are associated with overproduction of pro-inflammatory cytokines and abnormal spermatogenesis [175]. In addition, varicocele causes a rise in scrotal temperature with its negative effect on spermatogenesis (see section 2.4. of this article).

4. Personalized Management

Since the effects of lifestyle and environmental factors on male fertility (see section 2 of this article) are highly dependent on a particular combination of susceptibility factors (see section 3 of this article), which is unique to each individual, the management of the resulting pathological condition also needs to be strictly individualized.

4.1. Prevention

Among the preventive measures aimed at the reduction of risk for human health damage in general, and reproductive health in particular, collective ones and individual ones can be

distinguished. The collective measures are carried out by governments and use habitual tools of centralized regulation, such as promotion of social health-care programs, ban on certain products and industrial procedures, and selective imposition of others. At present, this kind of measures are mainly focused on the implement and increase of health taxes on tobacco, alcohol, and sugar-sweetened beverages, control of hypertension, medical checkups for prevention and early detection of cancer, reduction of both indoor and ambient air pollution, and increasing consumption of healthy and decreasing that of unhealthy food by incentivizing companies and empowering consumers [176].

The efficacy of individual preventive measures depends on the willingness and objective capacity of persons exposed to potentially harmful factors to change their habits so as to reduce the load. Dietary habits should be modified to reduce popular Western-style diet, low in vitamins and high in processed products [177]. On the other hand, dietary patterns based on foods rich in antioxidant and anti-inflammatory compounds, such as vitamin C, vitamin E, vitamin D, folate, β -carotene, selenium, zinc, cryptoxanthin and lycopene, and low in saturated fatty acids and trans-fatty acids appear to help prevent male infertility [177]. Voluntary exposures, such as smoking, alcohol, inadequate exercise, heat, or incorrect positioning of electronic devices (see Section 2 of this article) should be modified. If they cannot be modified easily, such as cases of professional exposures, or if the exposed person is reticent to adopt the lifestyle change recommended, they should be at least taken into consideration. This will make it possible to search for objective markers of their impact in each individual. Since most of the effects of lifestyle and environmental factors converge towards oxidative stress [13,14], essays for the evaluation of oxidative stress markers and their local effects in semen [178–181] can be considered. A recent study suggested platelet mitochondrial function and endogenous coenzyme Q10 levels as markers of mitochondrial health in infertile men [182]. Data obtained by using such markers can serve as a basis for patient-tailored preventive antioxidant medications and long-term evaluation of their effects.

4.2. Intervention

The consequences of exposure to many lifestyle and environmental factors for male fertility can be reversible. Thus, spontaneous fertility improvement can be observed in many symptomatic men who quit such exposure. For instance, a recent study showed that smoking cessation had a positive effect on sperm concentration, semen volume, and total sperm count [183]. In other cases, treatment of the health issues predisposing to the effects of external lifestyle and environmental factors, if they can be identified, can also restore fertility or at least improve the existing fertility problem. This was shown to be the case for the use of metformin, a drug employed in both diabetes and insulin resistance without the presence of diabetes, which improved sperm count, morphology, and motility in men suffering from these pathologies [184]. Similarly, beneficial effects of oral antibiotics and anti-inflammatory agents on semen parameters, sperm chromatin structure and sperm DNA integrity in men with infertility diagnosis were reported [185], and oral treatment with high doses of antioxidant vitamins C and E improved both sperm DNA integrity [186] and assisted reproduction outcomes [187]. Other medical therapies, including gonadotropins, selective estrogen receptor modulators, and aromatase inhibitors, can also be considered [188]. It is important to note that the population to be treated by these noninvasive measures has to be adequately selected as in inadequately selected patients such therapies might have deleterious effects.

If neither lifestyle modification nor the above noninvasive treatments manage to improve male fertility, the recourse to assisted reproduction (AR) is needed. Depending on the degree of sperm damage, the use of different AR techniques, including intrauterine insemination (IA), conventional in-vitro fertilization (IVF), and IVF assisted by intracytoplasmic sperm injection (ICSI) or intracytoplasmic morphologically-selected sperm injection (IMSI) can be envisaged. IA and conventional IVF may be used in cases of moderate impairment of sperm count (oligozoospermia) and motility (asthenozoospermia) and of the presence of antisperm isoantibodies in the female partner, while ICSI or IMSI are required for severe oligozoospermia and asthenozoospermia as well as for high percentage of morphologically abnormal spermatozoa (teratozoospermia). ICSI,

developed and first successfully used in 1992 [189], can overcome even the most severe issues of sperm count, motility and morphology by selecting healthy-appearing spermatozoa and injecting them individually into each oocyte. However, high degrees of sperm DNA fragmentation seriously decrease the chance of livebirth after ICSI by affecting embryo viability and implantation potential without being detectable during the first 3-5 days after fertilization [190]. An empirical study [191] has shown that livebirths in cases of severe sperm DNA fragmentation can be improved by using IMSI, a modification of ICSI introduced in 2001 [192]. Methods for evaluation of sperm DNA fragmentation are not compatible with sperm survival, so that the use of high optical magnifications to select spermatozoa in the context of IMSI cannot detect DNA damage directly [193]. To understand how IMSI can improve AR outcomes as compared to ICSI, it is needed to have a look at the principal DNA-protective mechanism inherent in spermatozoa. In healthy spermatozoa DNA is shielded against oxidative damage by its close association with protamins in highly condensed sperm chromatin structure. This protection is ineffective in regions in which sperm chromatin condensation is defective, and this condition is reflected by the presence of intranuclear vacuoles that can be observed by electron microscopy (Figure 2). Even large intranuclear vacuoles are mostly undetectable with optical magnifications used in conventional ICSI but do appear at the magnification used in

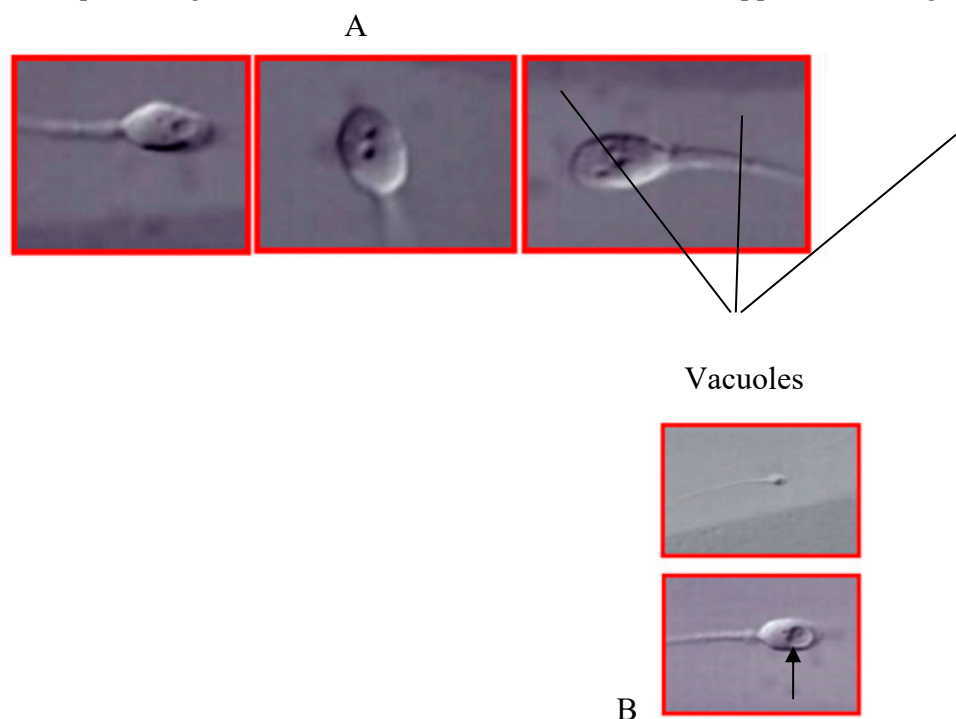


Figure 2. (A) Scanning electron micrographs of human spermatozoa with different-sized intranuclear vacuoles. (B) A large intranuclear vacuole observed in a living human spermatozoon at magnifications used in ICSI (upper image, barely seen) and IMSI (lower image, arrow) settings. Adapted from Tesarik [116].

IMSI (Figure 2) so that injection of spermatozoa with such vacuoles into oocytes can be avoided [191,193]. If the nocive action of a given lifestyle or environmental factor leads to an irreversible blockage of germ cell development at the round spermatid stage, round spermatid injection (ROSI) into oocytes may represent the treatment of last chance [194]. Pregnancies and births were achieved with the use of both freshly recovered spermatids [194,195] and those resulting from in vitro development of primary spermatocytes into spermatids [196]. Besides IVF, ICSI, IMSI and ROSI, varicocele is another invasive intervention to be considered; it was reported to improve semen quality and fertility in men with varicocele [175], though there is no common agreement on this effect.

5. Conclusions

Under contemporary lifestyle and environmental conditions, male fertility can be affected by a number of potentially harmful factors related to unhealthy habits or involuntary professional or nonprofessional exposures. The degree of the resulting fertility impairment is highly individual and is given by an interplay between the external factors and inherent susceptibility to them. If a man is exposed to a known external factor, his susceptibility should be evaluated by appropriate diagnostic methods. In many cases, the damage caused to the reproductive system is reversible and can be repaired spontaneously when the action of the factor(s) in question ceases. If the exposure cessation is difficult to achieve (obesity, professional exposures, stress) or accept (smoking, alcohol), or if the causative agent comes from sources independent of individual human will (air pollution, harmful chemicals present in food, beverages, cosmetics, etc.), symptomatic treatment, mainly based on individualized oral intake of antioxidants, can be envisaged. In the most severe cases, the recourse to assisted reproduction techniques is possible.

Methodological Considerations: The choice of the studies included in this review article was based exclusively on their novelty and scientific impact. Both meta-analyses and original research studies were valued equally according to these criteria. Eventual conflicting results were mentioned and discussed.

Funding: No funding was received for this study.

Conflict of Interest: The author declares no conflict of interest.

References

1. Preventing noncommunicable diseases (NCDs) by reducing environmental risk factors. Geneva: World Health Organization; 2017 (WHO/FWC/EPE/17.1). Licence: CC BY-NC-SA 3.0 IGO.
2. Pakholok, O. (2013). The Idea of Healthy Lifestyle and Its Transformation Into Health-Oriented Lifestyle in Contemporary Society. *Sage Open*. 2013, 3(3). doi: 10.1177/2158244013500281.
3. Kwon, H.-a.; Kim, S. Variation in the characteristics of everyday life and meaning of urban housing due to the transition of social structure: Focusing on articles published in lifestyle magazines. *Sustainability*. 2017, 9, 1298. doi: 10.3390/su9081298.
4. Farhud, D.D. Impact of lifestyle on health. *Iran J Public Health*. 2015, 44(11):1442-1444.
5. Tabish S.A. Lifestyle diseases: Consequences, characteristics, causes and control. *J Cardiol Curr Res*. 2017, 9(3): 00326. doi: 10.15406/jccr.2017.09.00326.
6. Balawender, K.; Orkisz, S. The impact of selected modifiable lifestyle factors on male fertility in the modern world. *Cent European J Urol*. 2020,73(4):563-568. doi: 10.5173/cej.2020.1975.
7. Ilacqua, A.; Izzo, G.; Emerenziani, G.P.; Baldari, C.; Aversa, A. Lifestyle and fertility: the influence of stress and quality of life on male fertility. *Reprod Biol Endocrinol*. 2018, 16:115. doi: 10.1186/s12958-018-0436-9
8. Carlsen, E.; Giwercman, A.; Keiding, N.; Skakkebaek, N.E. Evidence for decreasing quality of semen during past 50 years. *BMJ*. 1992, 305(6854):609-613. doi: 10.1136/bmj.305.6854.609.
9. Splingart, C.; Frapsauce, C.; Veau, S.; Barthélémy, C.; Royère, D.; Guérif, F. Semen variation in a population of fertile donors: Evaluation in a French centre over a 34-year period. *Int J Androl*. 2012, 35(3):467-474. doi: 10.1111/j.1365-2605.2011.01229.x.
10. Sengupta, P.; Dutta, S.; Krajewska-Kulak, E. The Disappearing sperms: Analysis of Reports Published Between 1980 and 2015. *Am J Mens Health*. 2017, 11(4):1279-1304. doi: 10.1177/1557988316643383.
11. Wang, T.; Wang, Q.; Fan, Z.; Xu, R.; Deng, X.; Li, Y.; Liang, S.; Lv, Z.; Huang, S.; Duan, Y.G.; Zhang, X.; Liu, Y. Association between central obesity and semen quality: A cross-sectional study in 4513 Chinese sperm donation volunteers. *Andrology*. 2024, 12(2):316-326. doi: 10.1111/andr.13471.
12. Bisht, S.; Faiq, M.; Tolahunase, M.; Dada, R. Oxidative stress and male infertility. *Nat Rev Urol*. 2017, 14(8):470-485. doi: 10.1038/nrurol.2017.69.
13. Alahmar, AT. Role of oxidative stress in male infertility: An updated review. *J Hum Reprod Sci*. 2019, 12(1):4-18. doi: 10.4103/jhrs.JHRS_150_18.

14. Takalani, N.B.; Monageng, E.M.; Mohlala, K.; Monsees, T.K.; Henkel, R.; Opuwari, C.S. (2023). Role of oxidative stress in male infertility. *Reproduction and Fertility*, 4(3):e230024. Retrieved Feb 16, 2025, from <https://doi.org/10.1530/RAF-23-0024>.
15. Evans, E.P.P.; Scholten, J.T.M.; Mzyk, A.; Reyes-San-Martin, C.; Llumbet, A.E.; Hamoh, T.; Arts, E.G.J.M.; Schirhagl, R.; Cantineau, A.E.P. Male subfertility and oxidative stress. *Redox Biol.* 2021, 46:102071. doi: 10.1016/j.redox.2021.102071.
16. Mancini, A.; Oliva, A.; Vergani, E.; Festa, R.; Silvestrini, A. The Dual Role of Oxidants in Male (In)fertility: Every ROSe Has a Thorn. *Int J Mol Sci.* 2023, 24(5):4994. doi: 10.3390/ijms24054994.
17. Venigalla, G.; Ila, V.; Dornbush, J.; Bernstein, A.; Loloi, J.; Pozzi, E.; Miller, D.; Ramasamy, R. Male obesity: Associated effects on fertility and the outcomes of offspring. *Andrology.* 2023, 13(1):64-71. doi: 10.1111/andr.13552.
18. Barbagallo, F.; Condorelli, R.A.; Mongioi, L.M.; Cannarella, R.; Cimino, L.; Magagnini, M.C.; Crafa, A.; La Vignera, S.; Calogero A.E. Molecular mechanisms underlying the relationship between obesity and male infertility. *Metabolites.* 2021, 11(12):840. doi: 10.3390/metabo11120840.
19. Chavarro, J.E.; Toth, T.L.; Wright, D.L.; Meeker, J.D.; Hauser, R. Body mass index in relation to semen quality, sperm DNA integrity, and serum reproductive hormone levels among men attending an infertility clinic. *Fertil Steril.* 2010, 93(7):2222–2231. doi: 10.1016/j.fertnstert.2009.01.100.
20. Suganami, T.; Nishida, J.; Ogawa, Y. A paracrine loop between adipocytes and macrophages aggravates inflammatory changes: role of free fatty acids and tumor necrosis factor alpha. *Arterioscler Thromb Vasc Biol.* 2005, 25(10):2062–2068. doi: 10.1161/01.ATV.0000183883.72263.13.
21. Dupont, J.; Pollet-Villard, X.; Reverchon, M.; Mellouk, N.; Levy, R. Adipokines in human reproduction. *Horm Mol Biol Clin Investig.* 2015, 24(1):11–24. doi: 10.1515/hmbci-2015-0034. PMID: 26574894.
22. Bellastella, G.; Menafrà, D.; Puliani, G.; Colao, A.; Savastano, S.; Obesity programs of nutrition, Education, Research and Assessment (OPERA) Group. How much does obesity affect the male reproductive function? *Int J Obes Suppl.* 2019, 9:50–64. doi: 10.1038/s41367-019-0008-2.
23. Agbaje, I.M.; Rogers, D.A.; McVicar, C.M.; McClure, N.; Atkinson, A.B.; Mallidis, C.; Lewis, S.E. Insulin dependant diabetes mellitus: Implications for male reproductive function. *Hum Reprod.* 2007, 22(7):1871–1877. doi: 10.1093/humrep/dem077.
24. Liu, Y.; Ding, Z. Obesity, a serious etiologic factor for male subfertility in modern society. *Reproduction.* 2017, 154(4):R123–R131. doi: 10.1530/REP-17-0161.
25. Li, L.; Law, C.; Lo Conte, R.; Power, C. Intergenerational influences on childhood body mass index: the effect of parental body mass index trajectories. *Am J Clin Nutr.* 2009, 89(2):551–557. doi: 10.3945/ajcn.2008.26759.
26. Huang, Q.; Ma, C.; Chen, L.; Luo, D.; Chen, R.; Liang, F. Mechanistic Insights into the Interaction Between Transcription Factors and Epigenetic Modifications and the Contribution to the Development of Obesity. *Front Endocrinol (Lausanne).* 2018, 9:370. doi: 10.3389/fendo.2018.00370.
27. Keyhan, S.; Burke, E.; Schrott, R.; Huang, Z.; Grenier, C.; Price, T.; Raburn, D.; Corcoran, D.L.; Soubry, A.; Hoyo, C.; Murphy, S.K.. Male obesity impacts DNA methylation reprogramming in sperm. *Clin Epigenet.* 2021, 13(1):17. doi: 10.1186/s13148-020-00997-0.
28. Soubry, A.; Guo, L.; Huang, Z.; Hoyo, C.; Romanus, S.; Price, T.; Murphy, S.K. Obesity-related DNA methylation at imprinted genes in human sperm: Results from the TIEGER study. *Clin Epigenetics.* 2016, 8:51. doi: 10.1186/s13148-016-0217-2.
29. El Hajj, N.; Zechner, U.; Schneider, E.; Tresch, A.; Gromoll, J.; Hahn, T.; Schorsch, M.; Haaf, T. Methylation status of imprinted genes and repetitive elements in sperm DNA from infertile males. *Sex Dev.* 2011, 5(2):60–69. doi: 10.1159/000323806.
30. Tunc, O.; Bakos, H.W.; Tremellen, K. Impact of body mass index on seminal oxidative stress. *Andrologia.* 2011, 43(2):121–128. doi: 10.1111/j.1439-0272.2009.01032.x.
31. Kumar, N.; Singh, A.K. Impact of environmental factors on human semen quality and male fertility: a narrative review. *Environ Sci Eur.* 2022, 34:6. doi: 10.1186/s12302-021-00585-w.

32. Deng, Z.; Chen, F.; Zhang, M.; Lan, L.; Qiao, Z.; Cui, Y.; An, J.; Wang, N.; Fan, Z.; Zhao, X.; Li, X. Association between air pollution and sperm quality: A systematic review and meta-analysis. *Environ Pollut.* 2016, *208(Pt B)*:663-669. doi: 10.1016/j.envpol.2015.10.044.
33. Giudice, L.C. Environmental toxicants: hidden players on the reproductive stage. *Fertil Steril.* 2016, *106(4)*:791-794. doi: 10.1016/j.fertnstert.2016.08.019.
34. Chiang, C.; Mahalingam, S.; Flaws, J.A. Environmental Contaminants Affecting Fertility and Somatic Health. *Semin Reprod Med.* 2017, *35(3)*:241-249. doi: 10.1055/s-0037-1603569.
35. Zhang, J.; Cai, Z.; Ma, C.; Xiong, J.; Li, H. Impacts of Outdoor Air Pollution on Human Semen Quality: A Meta-Analysis and Systematic Review. *Biomed Res Int.* 2020, *2020*:7528901. doi: 10.1155/2020/7528901.
36. Jurewicz, J.; Dziewirska, E.; Radwan, M.; Hanke, W. Air pollution from natural and anthropic sources and male fertility. *Reprod Biol Endocrinol.* 2018, *16*:109. doi: 10.1186/s12958-018-0430-2.
37. Calogero, A.E.; La Vignera, S.; Condorelli, R.A.; Perdichizzi, A.; Valenti, D.; Asero, P.; Carbone, U.; Boggia, B.; De Rosa, N.; Lombardi, G.; D'Agata, R.; Vicari, L.O.; Vicari, E.; De Rosa, M. Environmental car exhaust pollution damages human sperm chromatin and DNA. *J Endocrinol Invest.* 2011, *34(6)*:e139-43. doi: 10.1007/BF03346722.
38. Takeda, K.; Tsukue, N.; Yoshida, S. Endocrine-disrupting activity of chemicals in diesel exhaust and diesel exhaust particles. *Environ Sci.* 2004, *11(1)*:33-45.
39. Carré, J.; Gatimel, N.; Moreau, J.; Parinaud, J.; Léandri, R. Does air pollution play a role in infertility?: a systematic review. *Environ Health.* 2017, *16(1)*:82. doi: 10.1186/s12940-017-0291-8.
40. Gaspari, L.; Chang, S.S.; Santella, R.M.; Garte, S.; Pedotti, P.; Taioli, E. Polycyclic aromatic hydrocarbon-DNA adducts in human sperm as a marker of DNA damage and infertility. *Mutat Res.* 2003, *535(2)*:155-160. doi: 10.1016/s1383-5718(02)00297-8.
41. Mocarelli, P.; Gerthoux, P.M.; Patterson, D.G. Jr.; Milani, S.; Limonta, G.; Bertona, M.; Signorini, S.; Tramacere, P.; Colombo, L.; Crespi, C.; Brambilla, P.; Sarto, C.; Carreri, V.; Sampson, E.J.; Turner, W.E.; Needham, L.L. Dioxin exposure, from infancy through puberty, produces endocrine disruption and affects human semen quality. *Environ Health Perspect.* 2008, *116(1)*:70-77. doi: 10.1289/ehp.10399.
42. Schultz, R.; Suominen, J.; Värre, T.; Hakovirta, H.; Parvinen, M.; Toppari, J.; Pelto-Huikko, M. Expression of aryl hydrocarbon receptor and aryl hydrocarbon receptor nuclear translocator messenger ribonucleic acids and proteins in rat and human testis. *Endocrinology.* 2003, *144(3)*:767-776. doi: 10.1210/en.2002-220642. PMID: 12586752.
43. Manfo, F.P.; Jubendradass, R.; Nantia, E.A.; Moundipa, P.F.; Mathur, P.P. Adverse effects of bisphenol A on male reproductive function. *Rev Environ Contam Toxicol.* 2014, *228*:57-82. doi: 10.1007/978-3-319-01619-1_3.
44. Cariati, F.; D'Uonno, N.; Borrillo, F.; Iervolino, S.; Galdiero, G.; Tomaiuolo, R. "Bisphenol a: an emerging threat to male fertility". *Reprod Biol Endocrinol.* 2019, *17(1)*:6. doi: 10.1186/s12958-018-0447-6.
45. Rahman, M.S.; Kwon, W.S.; Lee, J.S.; Yoon, S.J.; Ryu, B.Y.; Pang, M.G. Bisphenol-A affects male fertility via fertility-related proteins in spermatozoa. *Sci Rep.* 2015, Mar 5:9169. doi: 10.1038/srep09169.
46. Murata, M.; Kang, J.H. Bisphenol A (BPA) and cell signaling pathways. *Biotechnol Adv.* 2018, *36(1)*:311-327. doi: 10.1016/j.biotechadv.2017.12.002.
47. Barbonetti, A.; Castellini, C.; Di Giammarco, N.; Santilli, G.; Francavilla, S.; Francavilla, F. In vitro exposure of human spermatozoa to bisphenol A induces pro-oxidative/apoptotic mitochondrial dysfunction. *Reprod Toxicol.* 2016, *66*:61-67. doi: 10.1016/j.reprotox.2016.09.014.
48. Bretveld, R.; Brouwers, M.; Ebisch, I.; Roeleveld, N. Influence of pesticides on male fertility. *Scand J Work Environ Health.* 2007, *33(1)*:13-28. doi: 10.5271/sjweh.1060.
49. Turusov, V.; Rakitsky, V.; Tomatis, L. Dichlorodiphenyltrichloroethane (DDT): ubiquity, persistence, and risks. *Environ Health Perspect.* 2002, *110(2)*:125-128. doi: 10.1289/ehp.02110125.
50. Bhatia, R.; Shiau, R.; Petreas, M.; Weintraub, J.M.; Farhang, L.; Eskenazi, B. Organochlorine pesticides and male genital anomalies in the child health and development studies. *Environ Health Perspect.* 2005, *113(2)*:220-224. doi: 10.1289/ehp.7382.

51. Tavares, R.S.; Amaral, S.; Paiva, C.; Baptista, M.; Ramalho-Santos, J. In vitro exposure to the organochlorine p,p'-DDE affects functional human sperm parameters. *Chemosphere*. 2015, *120*:443-446. doi: 10.1016/j.chemosphere.2014.08.075.
52. Pant, N.; Shukla, M.; Upadhyay, A.D.; Chaturvedi, P.K.; Saxena, D.K.; Gupta, Y.K. Association between environmental exposure to p, p'-DDE and lindane and semen quality. *Environ Sci Pollut Res Int*. 2014, *21(18)*:11009-16. doi: 10.1007/s11356-014-2965-x.
53. Longnecker, M.P. Invited Commentary: Why DDT matters now. *Am J Epidemiol*. 2005, *162(8)*:726-8. doi: 10.1093/aje/kwi277.
54. Anifandis, G.; Amiridis, G.; Dafopoulos, K.; Daponte, A.; Dovolou, E.; Gavriil, E.; Gorgogietas, V.; Kachpani, E.; Mamuris, Z.; Messini, C.I.; Vassiou, K.; Psarra, A.G. The In Vitro Impact of the Herbicide Roundup on Human Sperm Motility and Sperm Mitochondria. *Toxics*. 2017, *6(1)*:2. doi: 10.3390/toxics6010002.
55. de Liz Oliveira Cavalli, V.L.; Cattani, D.; Heinz Rieg, C.E.; Pierozan, P.; Zanatta, L.; Benedetti Parisotto, E.; Wilhelm Filho, D.; Mena Barreto Silva, F.R.; Pessoa-Pureur, R.; Zamoner, A. Roundup disrupts male reproductive functions by triggering calcium-mediated cell death in rat testis and Sertoli cells. *Free Radic Biol Med*. 2013, *65*:335-346. doi: 10.1016/j.freeradbiomed.2013.06.043.
56. Radke, E.G.; Braun, J.M.; Meeker, J.D.; Cooper, G.S. Phthalate exposure and male reproductive outcomes: A systematic review of the human epidemiological evidence. *Environ Int*. 2018, *121(Pt 1)*:764-793. doi: 10.1016/j.envint.2018.07.029.
57. Broe, A.; Pottgård, A.; Hallas, J.; Ahern, T.P.; Fedder, J.; Damkier, P. Association between use of phthalate-containing medication and semen quality among men in couples referred for assisted reproduction. *Hum Reprod*. 2018, *33(3)*:503-511. doi: 10.1093/humrep/dey009.
58. Cai, H.; Zheng, W.; Zheng, P.; Wang, S.; Tan, H.; He, G.; Qu, W. Human urinary/seminal phthalates or their metabolite levels and semen quality: A meta-analysis. *Environ Res*. 2015, *142*:486-494. doi: 10.1016/j.envres.2015.07.008.
59. Caporossi, L.; Alteri, A.; Campo, G.; Paci, E.; Tranfo, G.; Capanna, S.; Papaleo, E.; Pigni, D.; Viganò, P.; Papaleo, B. Cross Sectional Study on Exposure to BPA and Phthalates and Semen Parameters in Men Attending a Fertility Center. *Int J Environ Res Public Health*. 2020, *17(2)*:489. doi: 10.3390/ijerph17020489.
60. Bloom, M.S.; Whitcomb, B.W.; Chen, Z.; Ye, A.; Kannan, K.; Buck Louis, GM. Associations between urinary phthalate concentrations and semen quality parameters in a general population. *Hum Reprod*. 2015, *30(11)*:2645-2657. doi: 10.1093/humrep/dev219.
61. Thurston, S.W.; Mendiola, J.; Bellamy, A.R.; Levine, H.; Wang, C.; Sparks, A.; Redmon, J.B.; Drobnis, E.Z.; Swan, S.H. Phthalate exposure and semen quality in fertile US men. *Andrology*. 2016, *4(4)*:632-638. doi: 10.1111/andr.12124.
62. Lovekamp-Swan, T.; Jetten, A.M.; Davis, B.J. Dual activation of PPARalpha and PPARgamma by mono-(2-ethylhexyl) phthalate in rat ovarian granulosa cells. *Mol Cell Endocrinol*. 2003, *201(1-2)*:133-141. doi: 10.1016/s0303-7207(02)00423-9.
63. Rehman, S.; Usman, Z.; Rehman, S.; Aldraihem, M.; Rehman, N.; Rehman, I.; Ahmad, G. Endocrine disrupting chemicals and impact on male reproductive health. *Transl Androl Urol*. 2018, *7(3)*:490-503. doi: 10.21037/tau.2018.05.17.
64. Zhao, Y.; Ao, H.; Chen, L.; Sottas, C.M.; Ge, R.S.; Li, L.; Zhang, Y. Mono-(2-ethylhexyl) phthalate affects the steroidogenesis in rat Leydig cells through provoking ROS perturbation. *Toxicol In Vitro*. 2012, *26(6)*:950-955. doi: 10.1016/j.tiv.2012.04.003.
65. Sukhn, C.; Awwad, J.; Ghantous, A.; Zaatari, G. Associations of semen quality with non-essential heavy metals in blood and seminal fluid: data from the Environment and Male Infertility (EMI) study in Lebanon. *J Assist Reprod Genet*. 2018, *35(9)*:1691-1701. doi: 10.1007/s10815-018-1236-z.
66. Jamal, M.; Ghaffari, M.A.; Hoseinzadeh, P.; Hashemitabar, M.; Zeinali, M. Human Sperm Quality and Metal Toxicants: Protective Effects of some Flavonoids on Male Reproductive Function. *Int J Fertil Steril*. 2016, *10(2)*:215-223. doi: 10.22074/ijfs.2016.4912.
67. Ivell, R. Lifestyle impact and the biology of the human scrotum. *Reprod Biol Endocrinol*. 2007, *5*:15. doi: 10.1186/1477-7827-5-15.

68. Garolla, A.; Torino, M.; Miola, P.; Caretta, N.; Pizzol, D.; Menegazzo, M.; Bertoldo, A.; Foresta, C. Twenty-four-hour monitoring of scrotal temperature in obese men and men with a varicocele as a mirror of spermatogenic function. *Hum Reprod.* 2015, *30*(5):1006-1013. doi: 10.1093/humrep/dev057.
69. Al-Otaibi, S.T. Male infertility among bakers associated with exposure to high environmental temperature at the workplace. *J Taibah Univ Med Sci.* 2018, *13*(2):103-107. doi: 10.1016/j.jtumed.2017.12.003.
70. Hamerezaee, M.; Dehghan, S.F.; Golbabaee, F.; Fathi, A.; Barzegar, L.; Heidarnejad, N. Assessment of semen quality among workers exposed to heat stress: A cross-sectional study in a steel industry. *Saf Health Work.* 2018, *9*(2):232-235. doi: 10.1016/j.shaw.2017.07.003.
71. Mínguez-Alarcón, L.; Gaskins, A.J.; Chiu, Y.H.; Messerlian, C.; Williams, P.L.; Ford, J.B.; Souter, I.; Hauser, R.; Chavarro, J.E. Type of underwear worn and markers of testicular function among men attending a fertility center. *Hum Reprod.* 2018, *33*(9):1749-1756. doi: 10.1093/humrep/dey259.
72. Hassun Filho, P.A. Re: Wet heat exposure: a potentially reversible cause of low semen quality in infertile men. *Int Braz J Urol.* 2007, *33*(2):269-270. doi: 10.1590/s1677-55382007000200023.
73. Shefi, S.; Tarapore, P.E.; Walsh, T.J.; Croughan, M.; Turek, P.J. Wet heat exposure: a potentially reversible cause of low semen quality in infertile men. *Int Braz J Urol.* 2007, *33*(1):50-56; discussion 56-57. doi: 10.1590/s1677-55382007000100008.
74. Barazani, Y.; Katz, B.F.; Nagler, H.M.; Stember, D.S. Lifestyle, environment, and male reproductive health. *Urol Clin North Am.* 2014, *41*(1):55-66. doi: 10.1016/j.ucl.2013.08.017.
75. Mao, H.; Feng, L.; Yang, W.X. Environmental factors contributed to circannual rhythm of semen quality. *Chronobiol Int.* 2017, *34*(3):411-425. doi: 10.1080/07420528.2017.
76. Levitas, E.; Lunenfeld, E.; Weisz, N.; Friger, M.; Har-Vardi, I. Seasonal variations of human sperm cells among 6455 semen samples: a plausible explanation of a seasonal birth pattern. *Am J Obstet Gynecol.* 2013, *208*(5):406.e1-6. doi: 10.1016/j.ajog.2013.02.010.
77. Reyes, J.G.; Farias, J.G.; Henríquez-Olavarrieta, S.; Madrid, E.; Parraga, M.; Zepeda, A.B.; Moreno, R.D. The hypoxic testicle: physiology and pathophysiology. *Oxid Med Cell Longev.* 2012, *2012*:929285. doi: 10.1155/2012/929285.
78. Vernet, P.; Aitken, R.J.; Drevet, J.R. Antioxidant strategies in the epididymis. *Mol Cell Endocrinol.* 2004, *216*(1-2):31-39. doi: 10.1016/j.mce.2003.10.069.
79. Mahfouz, R.; Sharma, R.; Thiyagarajan, A.; Kale, V.; Gupta, S.; Sabanegh, E.; Agarwal, A. Semen characteristics and sperm DNA fragmentation in infertile men with low and high levels of seminal reactive oxygen species. *Fertil Steril.* 2010, *94*(6):2141-2146. doi: 10.1016/j.fertnstert.2009.12.030.
80. Gong, Y.; Guo, H.; Zhang, Z.; Zhou, H.; Zhao, R.; He, B. Heat Stress Reduces Sperm Motility via Activation of Glycogen Synthase Kinase-3 α and Inhibition of Mitochondrial Protein Import. *Front Physiol.* 2017, *8*:718. doi: 10.3389/fphys.2017.00718.
81. Rao, M.; Xia, W.; Yang, J.; Hu, L.X.; Hu, S.F.; Lei, H.; Wu, Y.Q.; Zhu, C.H. Transient scrotal hyperthermia affects human sperm DNA integrity, sperm apoptosis, and sperm protein expression. *Andrology.* 2016, *4*(6):1054-1063. doi: 10.1111/andr.12228.
82. Talhout, R.; Schulz, T.; Florek, E.; van Benthem, J.; Wester, P.; Opperhuizen, A. Hazardous compounds in tobacco smoke. *Int J Environ Res Public Health.* 2011, *8*(2):613-628. doi: 10.3390/ijerph8020613.
83. Sharma, R.; Harlev, A.; Agarwal, A.; Esteves, S.C. Cigarette smoking and semen quality: A new meta-analysis examining the effect of the 2010 World Health Organization Laboratory Methods for the Examination of Human Semen. *Eur Urol.* 2016, *70*(4):635-645. doi: 10.1016/j.eururo.2016.04.010.
84. Mostafa, R.M.; Nasrallah, Y.S.; Hassan, M.M.; Farrag, A.F.; Majzoub, A.; Agarwal, A. The effect of cigarette smoking on human seminal parameters, sperm chromatin structure and condensation. *Andrologia.* 2018, *50*(3). doi: 10.1111/and.12910.
85. Osadchuk, L.; Kleshchev, M.; Osadchuk, A. Effects of cigarette smoking on semen quality, reproductive hormone levels, metabolic profile, zinc and sperm DNA fragmentation in men: results from a population-based study. *Front Endocrinol (Lausanne).* 2023, *14*:1255304. doi: 10.3389/fendo.2023.1255304.
86. Esakky, P.; Moley, K.H. Paternal smoking and germ cell death: A mechanistic link to the effects of cigarette smoke on spermatogenesis and possible long-term sequelae in offspring. *Mol Cell Endocrinol.* 2016, *435*:85-93. doi: 10.1016/j.mce.2016.07.015.

87. Vivarelli, F.; Canistro, D.; Cirillo, S.; Cardenia, V.; Rodriguez-Estrada, M.T.; Paolini, M. Impairment of testicular function in electronic cigarette (e-cig, e-cigs) exposed rats under low-voltage and nicotine-free conditions. *Life Sci.* 2019, 228:53-65. doi: 10.1016/j.lfs.2019.04.059.
88. Sansone, A.; Di Dato, C.; de Angelis, C.; Menafra, D.; Pozza, C.; Pivonello, R.; Isidori, A.; Gianfrilli, D. Smoke, alcohol and drug addiction and male fertility. *Reprod Biol Endocrinol.* 2018, 16(1):3. doi: 10.1186/s12958-018-0320-7.
89. Li, Y.; Lin, H.; Li, Y.; Cao, J. Association between socio-psycho-behavioral factors and male semen quality: systematic review and meta-analyses. *Fertil Steril.* 2011, 95(1):116-123. doi: 10.1016/j.fertnstert.2010.06.031.
90. Condorelli, R.A.; Calogero, A.E.; Vicari, E.; La Vignera, S. Chronic consumption of alcohol and sperm parameters: our experience and the main evidences. *Andrologia.* 2015, 47(4):368-379. doi: 10.1111/and.12284.
91. Hassan, M.A.; Killick, S.R. Negative lifestyle is associated with a significant reduction in fecundity. *Fertil Steril.* 2004, 81(2):384-92. doi: 10.1016/j.fertnstert.2003.06.027.
92. Hansen, M.L.; Thulstrup, A.M.; Bonde, J.P.; Olsen, J.; Håkonsen, L.B.; Ramlau-Hansen, C.H. Does last week's alcohol intake affect semen quality or reproductive hormones? A cross-sectional study among healthy young Danish men. *Reprod Toxicol.* 2012, 34(3):457-462. doi: 10.1016/j.reprotox.2012.06.004.
93. Aboulmaouahib, S.; Madkour, A.; Kaarouch, I.; Sefrioui, O.; Saadani, B.; Copin, H.; Benkhalifa, M.; Louanjli, N.; Cadi, R. Impact of alcohol and cigarette smoking consumption in male fertility potential: Looks at lipid peroxidation, enzymatic antioxidant activities and sperm DNA damage. *Andrologia.* 2018, 50(3). doi: 10.1111/and.12926.
94. Szkodziak, F.; Krzyżanowski, J.; Szkodziak, P. Psychological aspects of infertility. A systematic review. *J Int Med Res.* 2020, 48(6):300060520932403. doi: 10.1177/0300060520932403.
95. Kirby, E.D.; Geraghty, A.C.; Ubuka, T.; Bentley, G.E.; Kaufer, D. Stress increases putative gonadotropin inhibitory hormone and decreases luteinizing hormone in male rats. *Proc Natl Acad Sci U S A.* 2009, 106(27):11324-1139. doi: 10.1073/pnas.0901176106.
96. Odetayo, A.F.; Akhigbe, R.E.; Basse, G.E.; Hamed, M.A.; Olayaki, L.A. Impact of stress on male fertility: role of gonadotropin inhibitory hormone. *Front Endocrinol (Lausanne).* 2024, 14:1329564. doi: 10.3389/fendo.2023.1329564.
97. Nargund, V.H. Effects of psychological stress on male fertility. *Nat Rev Urol.* 2015, 12(7):373-382. doi: 10.1038/nrurol.2015.112.
98. Belladelli, F.; Basran, S.; Eisenberg, M.L. Male Fertility and Physical Exercise. *World J Mens Health.* 2023, 41(3):482-488. doi: 10.5534/wjmh.220199.
99. Hajizadeh Maleki, B.; Tartibian, B. Resistance exercise modulates male factor infertility through anti-inflammatory and antioxidative mechanisms in infertile men: A RCT. *Life Sci.* 2018, 203:150-160. doi: 10.1016/j.lfs.2018.04.039.
100. Minas, A.; Fernandes, A.C.C.; Maciel Júnior, V.L.; Adami, L.; Intasqui, P.; Bertolla, R.P. Influence of physical activity on male fertility. *Andrologia.* 2022, 54(7):e14433. doi: 10.1111/and.14433.
101. Agarwal, A.; Singh, A.; Hamada, A.; Kesari, K. Cell phones and male infertility: a review of recent innovations in technology and consequences. *Int Braz J Urol.* 2011, 37(4):432-454. doi: 10.1590/s1677-55382011000400002.
102. Adams, J.A.; Galloway, T.S.; Mondal, D.; Esteves, S.C.; Mathews, F. Effect of mobile telephones on sperm quality: a systematic review and meta-analysis. *Environ Int.* 2014, 70:106-112. doi: 10.1016/j.envint.2014.04.015.
103. Liu, K.; Li, Y.; Zhang, G.; Liu, J.; Cao, J.; Ao, L.; Zhang, S. Association between mobile phone use and semen quality: a systemic review and meta-analysis. *Andrology.* 2014, 2(4):491-501. doi: 10.1111/j.2047-2927.2014.00205.x.
104. Delli Muti, N.; Salvio, G.; Ciarloni, A.; Perrone, M.; Tossetta, G.; Lazzarini, R.; Bracci, M.; Balercia, G. Can extremely low frequency magnetic field affect human sperm parameters and male fertility? *Tissue Cell.* 2023, 82:102045. doi: 10.1016/j.tice.2023.102045.
105. Avendaño, C.; Mata, A.; Sanchez Sarmiento, C.A.; Doncel, G.F. Use of laptop computers connected to internet through Wi-Fi decreases human sperm motility and increases sperm DNA fragmentation. *Fertil Steril.* 2012, 97(1):39-45.e2. doi: 10.1016/j.fertnstert.2011.10.012.

106. Hu, W.; Chen, M.; Wu, W.; Lu, J.; Zhao, D.; Pan, F.; Lu, C.; Xia, Y.; Hu, L.; Chen, D.; Sha, J.; Wang, X. Gene-gene and gene-environment interactions on risk of male infertility: Focus on the metabolites. *Environ Int.* 2016, *91*:188-195. doi: 10.1016/j.envint.2016.02.025.
107. Rubes, J.; Selevan, S.G.; Sram, R.J.; Evenson, D.P.; Perreault, S.D. GSTM1 genotype influences the susceptibility of men to sperm DNA damage associated with exposure to air pollution. *Mutat Res.* 2007, *625(1-2)*:20-28. doi: 10.1016/j.mrfmmm.2007.05.012.
108. Gunes, S.; Esteves, S.C. Role of genetics and epigenetics in male infertility. *Andrologia.* 2021, *53(1)*:e13586. doi: 10.1111/and.13586.
109. Laan, M.; Kasak, L.; Punab, M. Translational aspects of novel findings in genetics of male infertility-status quo 2021. *Br Med Bull.* 2021, *140(1)*:5-22. doi: 10.1093/bmb/ldab025.
110. Montjean, D.; Beaumont, M.; Natiq, A.; Louanjli, N.; Hazout, A.; Miron, P.; Liehr, T.; Cabry, R.; Ratbi, I.; Benkhalifa, M. Genome and epigenome disorders and male infertility: Feedback from 15 years of clinical and research experience. *Genes (Basel).* 2024, *15(3)*:377. doi: 10.3390/genes15030377.
111. Pereira, R.; Sousa, M. Morphological and molecular bases of male infertility: A closer look at sperm flagellum. *Genes.* 2023, *14(2)*: 383. doi: 10.3390/genes14020383.
112. Ruiz-Pesini, E.; Lapeña, A.C.; Díez-Sánchez, C.; Pérez-Martos, A.; Montoya, J.; Alvarez, E.; Díaz, M.; Urriés, A.; Montoro, L.; López-Pérez, M.J., Enríquez, J.A. Human mtDNA haplogroups associated with high or reduced spermatozoa motility. *Am J Hum Genet.* 2000, *67(3)*:682-696. doi: 10.1086/303040.
113. Selvi Rani, D.; Vanniarajan, A.; Gupta, N.J.; Chakravarty, B.; Singh, L.; Thangaraj, K. A novel missense mutation C11994T in the mitochondrial ND4 gene as a cause of low sperm motility in the Indian subcontinent. *Fertil Steril.* 2006 Dec;*86(6)*:1783-5. doi: 10.1016/j.fertnstert.2006.04.044.
114. Baklouti-Gargouri, S.; Ghorbel, M.; Ben Mahmoud, A.; Mkaouar-Rebai, E.; Cherif, M.; Chakroun, N.; Sellami, A.; Fakhfakh, F.; Ammar-Keskes, L. Identification of a novel m.9588G > a missense mutation in the mitochondrial COIII gene in asthenozoospermic Tunisian infertile men. *J Assist Reprod Genet.* 2014 May;*31(5)*:595-600. doi: 10.1007/s10815-014-0187-2.
115. Kao, S.H.; Chao, H.T.; Liu, H.W.; Liao, T.L.; Wei, Y.H. Sperm mitochondrial DNA depletion in men with asthenospermia. *Fertil Steril.* 2004, *82(1)*:66-73. doi: 10.1016/j.fertnstert.2003.11.056.
116. Tesarik, J. Acquired sperm DNA modifications: Causes, consequences, and potential solutions. *EMJ.* 2019, *4(3)*: 83-95.
117. Jenkins, T.G.; Aston, K.I.; James, E.R.; Carrell, D.T. Sperm epigenetics in the study of male fertility, offspring health, and potential clinical applications. *Syst Biol Reprod Med.* 2017, *63(2)*: 69-76, DOI: 10.1080/19396368.2016.1274791.
118. Santi, D.; De Vincentis, S.; Magnani, E.; Spaggiari, G. Impairment of sperm DNA methylation in male infertility: a meta-analytic study. *Andrology.* 2017, *5(4)*:695-703. doi: 10.1111/andr.12379.
119. Pollard, C.A.; Jenkins, T.G. Epigenetic mechanisms within the sperm epigenome and their diagnostic potential. *Best Pract Res Clin Endocrinol Metab.* 2020, *34(6)*:101481. doi: 10.1016/j.beem.2020.101481.
120. Hosseini, M.; Khalafiyani, A.; Zare, M.; Karimzadeh, H.; Bahrami, B.; Hammami, B.; Kazemi, M. Sperm epigenetics and male infertility: unraveling the molecular puzzle. *Hum Genomics.* 2024, *18(1)*:57. doi: 10.1186/s40246-024-00626-4.
121. Li, X.P.; Hao, C.L.; Wang, Q.; Yi, X.M.; Jiang, Z.S. H19 gene methylation status is associated with male infertility. *Exp Ther Med.* 2016, *12(1)*:451-456. doi: 10.3892/etm.2016.3314.
122. Poplinski, A.; Tüttelmann, F.; Kanber, D.; Horsthemke, B.; Gromoll, J. Idiopathic male infertility is strongly associated with aberrant methylation of MEST and IGF2/H19 ICR1. *Int J Androl.* 2010, *33(4)*:642-649. doi: 10.1111/j.1365-2605.2009.01000.x.
123. Dong, H.; Wang, Y.; Zou, Z.; Chen, L.; Shen, C.; Xu, S.; Zhang, J.; Zhao, F.; Ge, S.; Gao, Q.; Hu, H.; Song, M.; Wang, W. Abnormal methylation of imprinted genes and cigarette smoking: Assessment of their association with the risk of male infertility. *Reprod Sci.* 2017, *24(1)*:114-123. doi: 10.1177/1933719116650755.
124. Cescon, M.; Chianese, R.; Tavares, R.S. Environmental impact on male (in)fertility via epigenetic route. *J Clin Med.* 2020, *9(8)*:2520. doi: 10.3390/jcm9082520.
125. Santolero, D.; Titchenell, P.M. Resolving the paradox of hepatic insulin resistance. *Cell Mol Gastroenterol Hepatol.* 2019, *7(2)*:447-456. doi: 10.1016/j.jcmgh.2018.10.016.

126. Zańko, A.; Martynowicz, I.; Citko, A.; Konopka, P.; Paszko, A.; Pawłowski, M.; Szczerbiński, Ł.; Siewko, K.; Krętowski, A.J.; Kuczyński, W.; Milewski, R. The influence of lifestyle on male fertility in the context of insulin resistance-identification of factors that influence semen quality. *J Clin Med*. 2024, *13*(10):2797. doi: 10.3390/jcm13102797.
127. Sasaki, N.; Ueno, Y.; Higashi, Y. Indicators of insulin resistance in clinical practice. *Hypertens Res*. 2024, *47*:978–980. doi: 10.1038/s41440-023-01566-7.
128. Zańko, A.; Siewko, K.; Krętowski, A.J.; Milewski, R. Lifestyle, insulin resistance and semen quality as co-dependent factors of male infertility. *Int J Environ Res Public Health*. 2022, *20*(1):732. doi: 10.3390/ijerph20010732.
129. Asare-Anane, H.; Bannison, S.B.; Ofori, E.K.; Ateko, R.O.; Bawah, A.T.; Amanquah, S.D.; Oppong, S.Y.; Gandau, B.B.; Ziem, J.B. Tobacco smoking is associated with decreased semen quality. *Reprod Health*. 2016, *13*(1):90. doi: 10.1186/s12978-016-0207-z.
130. Du, C.Q.; Yang, Y.Y.; Chen, J.; Feng, L.; Lin, W.Q. Association between sleep quality and semen parameters and reproductive hormones: A cross-sectional study in Zhejiang, China. *Nat Sci Sleep*. 2020, *12*:11-18. doi: 10.2147/NSS.S235136.
131. Józków, P.; Rossato, M. The Impact of Intense Exercise on Semen Quality. *Am J Mens Health*. 2017, *11*(3):654-662. doi: 10.1177/1557988316669045.
132. Maresch, C.C.; Stute, D.C.; Fleming, T.; Lin, J.; Hammes, H.P.; Linn, T. Hyperglycemia induces spermatogenic disruption via major pathways of diabetes pathogenesis. *Sci Rep*. 2019, *9*(1):13074. doi: 10.1038/s41598-019-49600-4.
133. Huang, R.; Chen, J.; Guo, B.; Jiang, C.; Sun, W. Diabetes-induced male infertility: potential mechanisms and treatment options. *Mol Med*. 2024, *30*(1):11. doi: 10.1186/s10020-023-00771-x.
134. Yaribeygi, H.; Sathyapalan, T.; Atkin, S.L.; Sahebkar, A. Molecular mechanisms linking oxidative stress and diabetes mellitus. *Oxid Med Cell Longev*. 2020, *2020*:8609213. doi: 10.1155/2020/8609213.
135. Esposito, K.; Nappo, F.; Marfella, R.; Giugliano, G.; Giugliano, F.; Ciotola, M.; Quagliaro, L.; Ceriello, A.; Giugliano, D. Inflammatory cytokine concentrations are acutely increased by hyperglycemia in humans: role of oxidative stress. *Circulation*. 2002, *106*(16):2067-2072. doi: 10.1161/01.cir.0000034509.14906.ae.
136. Donath, M.Y.; Shoelson, S.E. Type 2 diabetes as an inflammatory disease. *Nat Rev Immunol*. 2011, *11*(2):98-107. doi: 10.1038/nri2925.
137. Corona, G.; Giorda, C.B.; Cucinotta, D.; Guida, P.; Nada, E.; Gruppo di studio SUBITO-DE. Sexual dysfunction at the onset of type 2 diabetes: the interplay of depression, hormonal and cardiovascular factors. *J Sex Med*. 2014, *11*(8):2065-2073. doi: 10.1111/jsm.12601.
138. Kouidrat, Y.; Pizzol, D.; Cosco, T.; Thompson, T.; Carnaghi, M.; Bertoldo, A.; Solmi, M.; Stubbs, B.; Veronese, N. High prevalence of erectile dysfunction in diabetes: a systematic review and meta-analysis of 145 studies. *Diabet Med*. 2017, *34*(9):1185-1192. doi: 10.1111/dme.13403.
139. McLachlan, R.I. Basis, diagnosis and treatment of immunological infertility in men. *J Reprod Immunol*. 2002, *57*(1-2):35-45. doi: 10.1016/s0165-0378(02)00014-1.
140. Cui, D.; Han, G.; Shang, Y.; Liu, C.; Xia, L.; Li, L.; Yi, S. Antisperm antibodies in infertile men and their effect on semen parameters: a systematic review and meta-analysis. *Clin Chim Acta*. 2015, *444*:29-36. doi: 10.1016/j.cca.2015.01.033.
141. Shibahara, H.; Shiraiishi, Y.; Suzuki, M. Diagnosis and treatment of immunologically infertile males with antisperm antibodies. *Reprod Med Biol*. 2005, *4*(2):133-141. doi: 10.1111/j.1447-0578.2005.00102.x.
142. Lai, Y.M.; Lee, J.F.; Huang, H.Y.; Soong, Y.K.; Yang, F.P.; Pao, C.C. The effect of human papillomavirus infection on sperm cell motility. *Fertil Steril*. 1997, *67*(6):1152-1155. doi: 10.1016/s0015-0282(97)81454-9.
143. Nasser, S.; Monavari, S.H.; Keyvani, H.; Nikkhoo, B.; Vahabpour Roudsari, R.; Khazeni, M. The prevalence of Human Papilloma Virus (HPV) infection in the oligospermic and azospermic men. *Med J Islam Repub Iran*. 2015, *29*:272.
144. Kurscheidt, F.A.; Damke, E.; Bento, J.C.; Balani, V.A.; Takeda, K.I.; Piva, S.; Piva, J.P.; Irie, M.M.T.; Gimenes, F.; Consolaro, M.E.L. Effects of Herpes Simplex Virus Infections on Seminal Parameters in Male Partners of Infertile Couples. *Urology*. 2018, *113*:52-58. doi: 10.1016/j.urology.2017.11.050.

145. Yang, T.; Zou, Y.; Zhou, W.; Ruan, Z.; Kong, Y.; Zhou, Y.; Zhang, J.; Xie, X. Clonal diversity of Ureaplasma species and its relationship with oligozoospermia and semen quality in Chinese infertile males. *Eur J Clin Microbiol Infect Dis*. 2018, *37*(10):1957-1963. doi: 10.1007/s10096-018-3331-6.
146. López-Hurtado, M.; Flores-Salazar, V.R.; Gutierrez-Trujillo, R.; Guerra-Infante, F.M. Prevalence, concordance and reproductive sequelae after Chlamydia trachomatis infection in Mexican infertile couples. *Andrologia*. 2020, *52*(10):e13772. doi: 10.1111/and.13772.
147. Li, H.; Xiao, X.; Zhang, J.; Zafar, M.I.; Wu, C.; Long, Y.; Lu, W.; Pan, F.; Meng, T.; Zhao, K.; Zhou, L.; Shen, S.; Liu, L.; Liu, Q.; Xiong, C. Impaired spermatogenesis in COVID-19 patients. *EClinicalMedicine*. 2020, *28*:100604. doi: 10.1016/j.eclinm.2020.100604.
148. Apaydin, T.; Sahin, B.; Dashdamirova, S.; Dincer Yazan, C.; Elbasan, O.; Ilgin, C.; Bilgin, H.; Cam, H.K.; Bahramzada, G.; Kucuk, A.; Haklar, G.; Ilikso Gozu, H. The association of free testosterone levels with coronavirus disease 2019. *Andrology*. 2022, *10*(6):1038-1046. doi: 10.1111/andr.13152.
149. Joguet, G.; Mansuy, J.M.; Matusali, G.; Hamdi, S.; Walschaerts, M.; Pavili, L.; Guyomard, S.; Prisant, N.; Lamarre, P.; Dejuq-Rainsford, N.; Pasquier, C.; Bujan, L. Effect of acute Zika virus infection on sperm and virus clearance in body fluids: a prospective observational study. *Lancet Infect Dis*. 2017, *17*(11):1200-1208. doi: 10.1016/S1473-3099(17)30444-9.
150. Erles, K.; Rohde, V.; Thaele, M.; Roth, S.; Edler, L.; Schlehofer, J.R. DNA of adeno-associated virus (AAV) in testicular tissue and in abnormal semen samples. *Hum Reprod*. 2001, *16*(11):2333-2337. doi: 10.1093/humrep/16.11.2333.
151. Chen, J.; Chen, J.; Fang, Y.; Shen, Q.; Zhao, K.; Liu, C.; Zhang, H. Microbiology and immune mechanisms associated with male infertility. *Front Immunol*. 2023, *14*:1139450. doi: 10.3389/fimmu.2023.1139450.
152. Hussein, M.R.; Abou-Deif, E.S.; Bedaiwy, M.A.; Said, T.M.; Mustafa, M.G.; Nada, E.; Ezat, A.; Agarwal, A. Phenotypic characterization of the immune and mast cell infiltrates in the human testis shows normal and abnormal spermatogenesis. *Fertil Steril*. 2005, *83*(5):1447-1453. doi: 10.1016/j.fertnstert.2004.11.062.
153. Seshadri, S.; Flanagan, B.; Vince, G.; Lewis-Jones, D.J. Detection of subpopulations of leucocytes in different subgroups of semen sample qualities. *Andrologia*. 2012, *44 Suppl 1*:354-361. doi: 10.1111/j.1439-0272.2011.01189.x.
154. Dcunha, R.; Hussein, R.S.; Ananda, H.; Kumari, S.; Adiga, S.K.; Kannan, N.; Zhao, Y.; Kalthur, G. Current insights and latest updates in sperm motility and associated applications in assisted reproduction. *Reprod Sci*. 2022, *29*(1):7-25. doi: 10.1007/s43032-020-00408-y.
155. Hendry, W.F.; Stedronska, J.; Jones, C.R.; Blackmore, C.A.; Barrett, A.; Peckham, M.J. Semen analysis in testicular cancer and Hodgkin's disease: pre- and post-treatment findings and implications for cryopreservation. *Br J Urol*. 1983, *55*(6):769-773. doi: 10.1111/j.1464-410x.1983.tb03423.x.
156. Williams, D.H. 4th; Karpman, E.; Sander, J.C.; Spiess, P.E.; Pisters, L.L.; Lipshultz, L.I. Pretreatment semen parameters in men with cancer. *J Urol*. 2009, *181*(2):736-740. doi: 10.1016/j.juro.2008.10.023.
157. Dias, T.R.; Agarwal, A.; Pushparaj, P.N.; Ahmad, G.; Sharma, R. Reduced semen quality in patients with testicular cancer seminoma is associated with alterations in the expression of sperm proteins. *Asian J Androl*. 2020, *22*(1):88-93. doi: 10.4103/aja.aja_17_19.
158. Drechsel, K.C.E.; Broer, S.L.; van Breda, H.M.K.; Stoutjesdijk, F.S.; van Dulmen-den Broeder, E.; Beishuizen, A.; Wallace, W.H.; Körholz, D.; Mauz-Körholz, C.; Hasenclever, D.; Cepelova, M.; Uyttebroeck, A.; Ronceray, L.; Twisk, J.W.R.; Kaspers, G.J.L.; Veening, M.A. Semen analysis and reproductive hormones in boys with classical Hodgkin lymphoma treated according to the EuroNet-PHL-C2 protocol. *Hum Reprod*. 2024, *39*(11):2411-2422. doi: 10.1093/humrep/deae204.
159. Levin, R.M.; Amsterdam, J.D.; Winokur, A.; Wein, A.J. Effects of psychotropic drugs on human sperm motility. *Fertil Steril*. 1981, *36*(4):503-506.
160. Chen, S.S.; Shen, M.R.; Chen, T.J.; Lai, S.L. Effects of antiepileptic drugs on sperm motility of normal controls and epileptic patients with long-term therapy. *Epilepsia*. 1992, *33*(1):149-153. doi: 10.1111/j.1528-1157.1992.tb02298.x.
161. Banihani, S.A. Effect of paracetamol on semen quality. *Andrologia*. 2018, *50*(1). doi: 10.1111/and.12874.

162. Stutz, G.; Zamudio, J.; Santillán, M.E.; Vincenti, L.; de Cuneo, M.F.; Ruiz R.D. The effect of alcohol, tobacco, and aspirin consumption on seminal quality among healthy young men. *Arch Environ Health*. 2004, 59(11):548-552. doi: 10.1080/00039890409603432.
163. Banihani, S.A.; Khasawneh, F.H. Effect of lansoprazole on human sperm motility, sperm viability, seminal nitric oxide production, and seminal calcium chelation. *Res Pharm Sci*. 2018, 13(5):460-468. doi: 10.4103/1735-5362.236839.
164. du Plessis, S.S.; Agarwal, A.; Syriac, A. Marijuana, phytocannabinoids, the endocannabinoid system, and male fertility. *J Assist Reprod Genet*. 2015, 32(11):1575-88. doi: 10.1007/s10815-015-0553-8.
165. Monteiro, C.; Marques, P.I.; Cavadas, B.; Damião, I.; Almeida, V.; Barros, N.; Barros, A.; Carvalho, F.; Gomes, S.; Seixas, S. Characterization of microbiota in male infertility cases uncovers differences in seminal hyperviscosity and oligoasthenoteratozoospermia possibly correlated with increased prevalence of infectious bacteria. *Am J Reprod Immunol*. 2018, 79(6):e12838. doi: 10.1111/aji.12838.
166. Altmäe, S.; Franasiak, J.M.; Mändar, R. The seminal microbiome in health and disease. *Nat Rev Urol*. 2019, 16(12):703-721. doi: 10.1038/s41585-019-0250-y.
167. Farahani, L.; Tharakan, T.; Yap, T.; Ramsay, J.W.; Jayasena, C.N.; Minhas, S. The semen microbiome and its impact on sperm function and male fertility: A systematic review and meta-analysis. *Andrology*. 2021, 9(1):115-144. doi: 10.1111/andr.12886.
168. Ma, Z.S.; Li, L. Semen Microbiome Biogeography: An Analysis Based on a Chinese Population Study. *Front Microbiol*. 2019, 9:3333. doi: 10.3389/fmicb.2018.03333.
169. Chatzokou, D.; Tsarna, E.; Davouti, E.; Siristatidis, C.S.; Christopoulou, S.; Spanakis, N.; Tsakris, A.; Christopoulos, P. Semen Microbiome, Male Infertility, and Reproductive Health. *Int. J. Mol. Sci*. 2025, 26:1446. <https://doi.org/10.3390/ijms26041446>.
170. Baud, D.; Pattaroni, C.; Vulliemoz, N.; Castella, V.; Marsland, B.J.; Stojanov, M. Sperm Microbiota and Its Impact on Semen Parameters. *Front Microbiol*. 2019, 10:234. doi: 10.3389/fmicb.2019.00234.
171. Yang, H.; Zhang, J.; Xue, Z.; Zhao, C.; Lei, L.; Wen, Y.; Dong, Y.; Yang, J.; Zhang, L. Potential Pathogenic Bacteria in Seminal Microbiota of Patients with Different Types of Dysspermatism. *Sci Rep*. 2020, 10:6876. doi: 10.1038/s41598-020-63787-x.
172. Okwelogu, S.I.; Ikechebelu, J.I.; Agbakoba, N.R.; Anukam, K.C. Microbiome Compositions From Infertile Couples Seeking *In Vitro* Fertilization, Using 16S rRNA Gene Sequencing Methods: Any Correlation to Clinical Outcomes? *Front Cell Infect Microbiol*. 2021, 11:709372. doi: 10.3389/fcimb.2021.709372.
173. Folliero, V.; Santonastaso, M.; Dell'Annunziata, F.; De Franciscis, P.; Boccia, G.; Colacurci, N.; De Filippis, A.; Galdiero, M.; Franci, G. Impact of *Escherichia coli* Outer Membrane Vesicles on Sperm Function. *Pathogens*. 2022, 11(7):782. doi: 10.3390/pathogens11070782.
174. Molina, N.M.; Plaza-Díaz, J.; Vilchez-Vargas, R.; Sola-Leyva, A.; Vargas, E.; Mendoza-Tesarik, R.; Galán-Lázaro, M.; Mendoza-Ladrón de Guevara, N.; Tesarik, J.; Altmäe, S. Assessing the testicular sperm microbiome: a low-biomass site with abundant contamination. *Reprod Biomed Online*. 2021, 43(3):523-531. doi: 10.1016/j.rbmo.2021.06.021.
175. Jensen, C.F.S.; Østergren, P.; Dupree, J.M.; Ohl, D.A.; Sønksen, J.; Fode, M. Varicocele and male infertility. *Nat Rev Urol*. 2017, 14(9):523-533. doi: 10.1038/nrurol.2017.98.
176. Alleyne, G.; Coll-Seck, A.M.; Frieden, T.R.; Tufton, C. Fourth time a charm?—How to make the UN High-Level Meeting on Noncommunicable Diseases effective. *JAMA*. Published online February 21, 2025. doi:10.1001/jama.2025.1431.
177. Salas-Huetos, A.; Bulló, M.; Salas-Salvadó, J. Dietary patterns, foods and nutrients in male fertility parameters and fecundability: a systematic review of observational studies. *Hum Reprod Update*. 2017, 23(4):371-389. doi: 10.1093/humupd/dmx006.
178. Agarwal, A.; Bui, A.D. Oxidation-reduction potential as a new marker for oxidative stress: Correlation to male infertility. *Investig Clin Urol*. 2017 Nov;58(6):385-399. doi: 10.4111/icu.2017.58.6.385. *Investig Clin Urol*. 2017, 58(6):385-399. doi: 10.4111/icu.2017.58.6.385.
179. Agarwal, A.; Qiu, E.; Sharma, R. Laboratory assessment of oxidative stress in semen. *Arab Journal of Urology*. 2018, 16(1):77-86. doi: 10.1016/j.aju.2017.11.008.

180. Katerji, M.; Filippova, M.; Duerksen-Hughes, P. Approaches and methods to measure oxidative stress in clinical samples: Research applications in the cancer field. *Oxid Med Cell Longev.* 2019, 2019:1279250. doi: 10.1155/2019/1279250.
181. Agarwal, A.; Baskaran, S.; Parekh, N.; Cho, C.L.; Henkel, R.; Vij, S.; Arafa, M.; Panner Selvam, M.K.; Shah, R. Male infertility. *Lancet.* 2021, 397(10271):319-333. doi: 10.1016/S0140-6736(20)32667-2.
182. Sumbalová, Z.; Rausová, Z.; Kucharská, J.; Šranko, P.; Harbulák, P.; Svitok, P.; López-Lluch, G.; Gvozdjaková, A. Platelet Mitochondrial Function and Endogenous Coenzyme Q₁₀ Levels Could Be Used as Markers of Mitochondrial Health in Infertile Men: A Pilot Study. *Int J Mol Sci.* 2024, 26(1):268. doi: 10.3390/ijms26010268.
183. Kulaksiz, D.; Toprak, T.; Tokat, E.; Yilmaz, M.; Ramazanoglu, M.A.; Garayev, A.; Sulukaya, M.; Degirmentepe, R.B.; Allahverdiyev, E.; Gul, M.; Verit, A. Sperm concentration and semen volume increase after smoking cessation in infertile men. *Int J Impot Res.* 2022, 34(6):614-619. doi: 10.1038/s41443-022-00605-0.
184. Morgante, G.; Tosti, C.; Orvieto, R.; Musacchio, M.C.; Piomboni, P.; De Leo, V. Metformin improves semen characteristics of oligo-terato-asthenozoospermic men with metabolic syndrome. *Fertil Steril.* 2011, 95(6):2150-2152. doi: 10.1016/j.fertnstert.2010.12.009.
185. Adel Domínguez, M.A.; Cardona Maya, W.D.; Mora Topete, A. Sperm DNA fragmentation: focusing treatment on seminal transport fluid beyond sperm production. *Arch Ital Urol Androl.* 2025, 30:13128. doi: 10.4081/aiua.2025.13128.
186. Greco, E.; Iacobelli, M.; Rienzi, L.; Ubaldi, F.; Ferrero, S.; Tesarik, J. Reduction of the incidence of sperm DNA fragmentation by oral antioxidant treatment. *J Androl.* 2005, 26(3):349-353. doi: 10.2164/jandrol.04146.
187. Greco, E.; Romano, S.; Iacobelli, M.; Ferrero, S.; Baroni, E.; Minasi, M.G.; Ubaldi, F.; Rienzi, L.; Tesarik, J. ICSI in cases of sperm DNA damage: beneficial effect of oral antioxidant treatment. *Hum Reprod.* 2005, 20(9):2590-2594. doi: 10.1093/humrep/dei091.
188. Sharma, A.; Minhas, S.; Dhillo, W.S.; Jayasena, C.N. Male infertility due to testicular disorders. *J Clin Endocrinol Metab.* 2021, 106(2):e442-e459. doi: 10.1210/clinem/dgaa781.
189. Palermo, G.; Joris, H.; Devroey, P.; Van Steirteghem, A.C. Pregnancies after intracytoplasmic injection of single spermatozoon into an oocyte. *Lancet.* 1992, 340(8810):17-18. doi: 10.1016/0140-6736(92)92425-f.
190. Tesarik, J.; Greco, E.; Mendoza, C. Late, but not early, paternal effect on human embryo development is related to sperm DNA fragmentation. *Hum Reprod.* 2004, 19(3):611-615. doi: 10.1093/humrep/deh127
191. Hazout, A.; Dumont-Hassan, M.; Junca, A.M.; Cohen Bacrie, P.; Tesarik, J. High-magnification ICSI overcomes paternal effect resistant to conventional ICSI. *Reprod Biomed Online.* 2006, 12(1):19-25. doi: 10.1016/s1472-6483(10)60975-3.
192. Bartoov, B.; Berkovitz, A.; Eltes, F. Selection of spermatozoa with normal nuclei to improve the pregnancy rate with intracytoplasmic sperm injection. *N Engl J Med.* 2001, 345:1067-1068. doi: 10.1056/NEJM200110043451416.
193. Tesarik, J.; Mendoza-Tesarik, R.; Mendoza, C. Sperm nuclear DNA damage: update on the mechanism, diagnosis and treatment. *Reprod Biomed Online.* 2006, 12(6):715-721. doi: 10.1016/s1472-6483(10)61083-8.
194. Tesarik, J.; Mendoza, C.; Testart, J. Viable embryos from injection of round spermatids into oocytes. *N Engl J Med.* 1995, 333(8):525. doi: 10.1056/NEJM199508243330819.
195. Tanaka, A.; Suzuki, K.; Nagayoshi, M.; Tanaka, A.; Takemoto, Y.; Watanabe, S.; Takeda, S.; Irahara, M.; Kuji, N.; Yamagata, Z.; Yanagimachi, R. Ninety babies born after round spermatid injection into oocytes: survey of their development from fertilization to 2 years of age. *Fertil Steril.* 2018, 110(3):443-451. doi: 10.1016/j.fertnstert.2018.04.033.
196. Tesarik, J.; Bahceci, M.; Ozcan, C.; Greco, E.; Mendoza, C. Restoration of fertility by in-vitro spermatogenesis. *Lancet.* 1999, 353(9152):555-556. doi: 10.1016/S0140-6736(98)04784-9.

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.