

Article

Not peer-reviewed version

---

# Efficient Bioprocess for Mixed Pet Waste Depolymerization Using Crude Cutinase

---

[Virender Kumar](#) , [Reinhard Wimmer](#) , [Cristiano Varrone](#) \*

Posted Date: 28 February 2025

doi: [10.20944/preprints202502.2241.v1](https://doi.org/10.20944/preprints202502.2241.v1)

Keywords: Mixed PET waste; Bioprocess; Depolymerization; Cutinase



Preprints.org is a free multidisciplinary platform providing preprint service that is dedicated to making early versions of research outputs permanently available and citable. Preprints posted at Preprints.org appear in Web of Science, Crossref, Google Scholar, Scilit, Europe PMC.

Copyright: This open access article is published under a Creative Commons CC BY 4.0 license, which permit the free download, distribution, and reuse, provided that the author and preprint are cited in any reuse.

Disclaimer/Publisher's Note: The statements, opinions, and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions, or products referred to in the content.

Article

# Efficient Bioprocess for Mixed PET Waste Depolymerization Using Crude Cutinase

Virender Kumar, Reinhard Wimmer, Cristiano Varrone \*

Department of Chemistry and Bioscience, Aalborg University, Fredrik Bajers Vej 7H, 9220 Aalborg, Denmark

\* Correspondence: cva@bio.aau.dk

**Abstract:** In recent years, several plastic-degrading enzymes with efficient depolymerization ability for PET have been reported. Here, we report a bioprocess for mixed PET waste depolymerization using crude extracellularly expressed enzymes in *E. coli*. The enzymes namely FastPETase, LCC, and LCCICCG were screened to depolymerize amorphous PET powder and films of different sizes and crystallinity. FastPETase, LCC, and LCCICCG achieved approximately 25, 34, and 70% depolymerization, respectively, when applied to 13 g L-1 of PET film, powder, or mixed waste in optimized enzyme conditions without any pH control. The yield of terephthalic acid in the hydrolytic process was maximum for LCCICCG followed by LCC and FastPETase. Finally, extracellular LCCICCG producing *E. coli* cells were cultivated using minimal medium supplemented with 0.1 % ammonium chloride and 1% glycerol as nitrogen and carbon sources in a bioreactor with final protein content and specific activity of  $119 \pm 5$  mg L-1 and  $1232 \pm 18$  U mg-1 respectively. Nearly complete depolymerization of 13 g L-1 PET and 23.8 g L-1 post-consumer PET was achieved in 50 h using crude LCCICCG supernatant, without enzyme purification, at 62°C. A bioprocess was thus developed to depolymerize 100 g L-1 mixed PET trays and bottle waste (MW1 and MW2), reaching 78 % and 50% yield at 62°C with a crude enzyme loading of 2.32 mg g-1 PET in 60 h. The results demonstrate an easy PET depolymerization strategy that could be exploited in large-scale facilities for efficient plastic waste treatment.

**Keywords:** mixed PET waste; bioprocess; depolymerization

---

## 1. Introduction

Polyethylene terephthalate (PET) is one of the most important plastics in volume and accounts for 18% of global plastic production. The huge consumption and usage generate a huge amount of discarded PET plastics and its subsequent mismanagement severely damages the ecological environment [4,5]. Globally, only 9% of plastic waste is recycled, leaving the rest accumulating in landfills or natural environments, where it persists for centuries. Approximately 19-23 million tons of plastic waste leaks into aquatic ecosystems, polluting lakes, rivers, and seas, and this figure could triple by 2040 without intervention (UNEP). Ellen Mc Arthur Foundation report (breaking the plastic wave) predicted that there will be more plastic waste than fish biomass in the Sea by 2050. A recent study employed machine learning to foresee that annual mismanaged plastic waste will nearly double to 121 million metric tons by 2050. Annual greenhouse gas emissions from plastic waste are also projected to grow by 37% to 3.35 billion tons of CO<sub>2</sub> [6]. This underscores the urgent need for sustainable waste management solutions. To overcome this problem, PET plastics recycling (thermo-chemical, mechanical, and biological) has been envisaged as an efficient and effective approach. It extends PET's life cycle, reduces non-renewable resource consumption, decreases plastic pollution, and protects the ecological environment [5,6]. However, PET thermo-mechanical recycling approaches have some drawbacks related to waste sourcing and their decreased mechanical properties during the extrusion process [7]. The mixed plastic waste further complicates these

processes due to its complex nature (types, composition, color, and additives). It is difficult to recycle it due to different polymer types and contamination. The recycling process for mixed plastics is energy-intensive, less efficient, and results in downcycled materials with limited utility due to cross-contamination. Biological depolymerization mediated by enzymes has emerged as an efficient and sustainable alternative for plastic treatment. Plastic waste recycling negates harmful environmental consequences by utilizing less energy-intensive processes, and fewer chemicals, and recovering valuable components from plastic waste. The enzymatic hydrolysis of polyethylene terephthalate (PET) offers a sustainable recycling pathway. It aligns with global efforts to reduce plastic pollution and foster a more sustainable materials economy [1,8–10]. This approach helps close the loop in a circular economy and offers a more environmentally friendly alternative to chemical recycling methods, which often involve harsher conditions and generate more waste [11]. PET monomers are linked by ester bonds, which can be enzymatically hydrolyzed, producing terephthalic acid (TPA) and ethylene glycol (EG) [2]. After the discovery of PET hydrolyzing cutinase two decades ago, several hydrolases have been identified and improved for PET hydrolysis [1,7,12,13]. Recently, two mutants of thermostable *IsPETase* [15,16] from the bacterium *Ideonella sakaiensis*, namely FastPETase [17] and HotPETase [9] were reported for improved PET depolymerization. In another study, a leaf-branch compost cutinase (LCC) [18] was engineered to get LCC<sup>ICCG</sup> mutant [19]. The enzyme paved the way for the first industrial PET recycling plant unit in France run by Carbios. Recently, Arnal et al. [2] compared four reported PET hydrolases (LCC<sup>ICCG</sup>, FAST-PETase, HotPETase, and PES-H1<sup>L92F/Q94Y</sup>) and demonstrated their ability to perform PET hydrolysis in industrial settings. They also discussed critical parameters to upscale an enzyme process for industrial deployment indicating that the crystallinity of PET, surface area, reaction temperature, the efficiency of the enzyme vs recrystallization, and substrate loading are among the most critical factors. It is now well known that PET hydrolases prefer amorphous regions of PET and no PET hydrolases have been reported to act efficiently on highly crystalline forms of the polymer, typically found in consumer products [2,20,21]. It is necessary to perform a pretreatment to convert PET to its amorphous state to meet the high enzymatic conversion essential to meet techno-economic goals. The thermostability, expression yields, and enzyme activity are also crucial factors in reaching industrial scales. The pretreatment of postconsumer waste and crystallinity of PET powder and enzyme production platform negatively affect the overall process. The extent of PET breakdown, substrate loading, enzyme price (production and purification cost), and enzyme loading are all key cost drivers [22]. The enzyme purification process requires high energy, chemicals, and water consumption with an overall negative impact on the environment. Therefore, it would be worthwhile to assess the potential of crude enzyme on PET hydrolysis focusing on high yield enzyme production in the extracellular supernatant. In this study, we have compared the depolymerization efficiency of three PET hydrolases (FastPETase, LCC, and LCC<sup>ICCG</sup>) on different PET substrate types such as powder or films, amorphous or crystalline, pure or mixed waste, and optimized key variables in small-scale. We demonstrated the extracellular expression of LCC<sup>ICCG</sup> in *E. coli* and the utilization of crude supernatant to catalyze efficient depolymerization of pure PET powder, films, post-consumer trays, and mixed trays and bottle waste (originated from Spain). A bioprocess has been developed for the efficient depolymerization of mixed PET waste in a 1L bioreactor and subsequent recovery and purification of TPA from the reaction mixture. It is a sustainable, low-cost PET hydrolytic process that completely hydrolyzes mixed PET waste into its monomers at the reactor scale.

## 2. Materials and Methods

### 2.1. Materials, Plasmids, and Enzyme

All the chemicals used in the study were of analytical grade. The p-Nitrophenol butyrate (pNPB), TPA, and MHET were purchased from Sigma-Aldrich, USA. All the plasmids and gene constructs were synthesized by Gene Universal, USA. The sequences for FastPETase, LCC, and LCC<sup>ICCG</sup> [17–20] were retrieved from the PDB database, codon-optimized, cloned into pET26b (+) and

pET28a (+) vector from Gene Universal, USA. PET films (0.25 mm amorphous and crystalline) were purchased from Goodfellow Cambridge Limited. PET powders of different sizes (0.05, 0.1, 0.25, 0.75; Table 1) were kindly provided by Dr. Alessandro Pellis, University of Genova, Italy. Mixed plastic wastes were received from different sources/origins and assigned the acronyms MW1-2 (Table 1) and cryo-milled PET powder (0.08 mm) as part of the UPLIFT project from AIMPLAS, Valencia, Spain. These wastes were composed of only PET (MW1 and MW2) and were micronized to obtain the sample size of 1 mm in diameter by ultra-centrifugal grinder (RetschTM 171 ZM200, Spain) or cryo-milled to obtain different sizes (Table 1).

**Table 1.** Size and crystallinity of different PET materials used in the study.

Sample Code	Sample name	Size or thickness (mm)	X <sub>c</sub> (%)
P80A	PET powder cryo-milled	0.08	9.3
P750A	PET powder	0.75	16.6
P750C	PET powder	0.75	37.7
P250C	PET powder	0.25	42.6
P100C	PET powder	0.1	39.1
P750C	PET powder	0.05	37.7
F250A	PET film (Amorphous)	0.25	0.6
F250C	PET film (Crystalline Film) Goodfellow	0.25	35.1
MW1	PET trays (real waste, origin Spain) cryo-milled at < 500 microns	<0.5	15.9
MW2	PET Bottle (real waste, origin: Spain) cryo-milled at > 500 microns	<0.5	28

\*P (powder), A (amorphous).

## 2.2. Expression and Purification of Enzymes

FastPETase, LCC, and LCC<sup>ICCG</sup> were expressed in the electrocompetent *Escherichia coli* strain BL21 (DE3) using pET28a(+) (FastPETase) and pET26b(+) vector (LCC and LCC<sup>ICCG</sup>). The sequence of LCC and LCC<sup>ICCG</sup> contained N-terminal PelB peptide for periplasmic expression. A single colony of *E. coli* BL21 (DE3) cells containing the gene FastPETase or LCC or LCC<sup>ICCG</sup> was transferred in 5 mL LB medium or M9 medium (Table S1) supplemented with 50 µg mL<sup>-1</sup> kanamycin and incubated at 37°C for 5 h. One mL of starter culture was transferred to a 50 mL LB<sub>Kan</sub> medium or M9 medium and incubated at 37°C and 190 rpm till the OD<sub>600</sub> reached 0.6-0.8. The protein production was induced using 0.5 mM (FastPETase) or 1 mM (LCC and LCC<sup>ICCG</sup>) IPTG and further incubated at 20°C (FastPETase) and 30°C (LCC and LCC<sup>ICCG</sup>) for 24 h respectively. The cells were harvested by centrifugation at 8000 g for 20 min at 4°C. For FastPETase, the cell pellet was resuspended (10% wet weight) in lysis or sample loading buffer (50 mM potassium phosphate buffer, 300 mM NaCl, 5 mM Imidazole; pH 7.5). The cells were lysed using sonication (45% amplitude; 9 sec on, 10 sec off; 15 min, and 2 cycles). The lysate was centrifuged at 12000 g for 30 min and pellet was discarded and the supernatant was used for further purification. For LCC and LCC<sup>ICCG</sup>, the supernatant was retained, and the pH was adjusted to 7.5. The supernatant was equilibrated with imidazole (5 mM) and 300 mM NaCl to mimic loading buffer conditions. The HisTrap FF column was pre-equilibrated (5 column volume) with 50 mM potassium phosphate buffer, 300 mM NaCl, 5 mM Imidazole; pH 7.5 (AKTA explorer system, GE Healthcare). The 0.22 µm filtered cell-free supernatant was loaded onto the column 2 mL min<sup>-1</sup>). After loading, the column was washed with wash buffer (50 mM potassium phosphate buffer, 300 mM NaCl, 20 mM Imidazole; pH 7.5) with 4 column volumes (2 mL min<sup>-1</sup>) to remove unbound proteins. The protein was eluted (1 mL min<sup>-1</sup>) with elution buffer (50 mM potassium phosphate buffer, 300 mM NaCl, 200 mM Imidazole; pH 7.5). The eluted fractions were checked on

the SDS PAGE. The purified fractions were pooled, dialyzed, and concentrated using 3 kDa Amicon filters (4000 g) against 50 mM potassium phosphate buffer (pH 7.5). The purified enzyme was used for the depolymerization of PET powder and film.

### 2.3. Estimation of Esterase Activity and Protein Concentration

Esterase activity was measured at 50°C (FastPETase) and 65°C (LCC and LCC<sup>ICCG</sup>) using p-NPB as a substrate as previously reported [23,24]. Solution A was composed of 86 µL of 50 mM p-nitrophenol butyrate (p-NPB) and 1000 µL of 2-butanol and solution B was prepared using 40 µL of Solution A and 1 mL of 50 mM potassium phosphate buffer (pH 8.0). A final assay mixture made up of 200 µL of solution B and 20 µL of enzyme solution (diluted if required). The change in absorbance was recorded at 405 nm due to the hydrolytic release of p-nitrophenol (p-NP) was measured using a microplate reader (Biotek, USA). One unit is defined as the amount of enzyme required to hydrolyze 1 µmol of substrate per minute under the given assay conditions. The standard of p-NP was prepared (10 µM-100 µM). The protein concentration was determined using Bradford assay [23,25]. Briefly, 4 µL of the protein solution was added into the wells of a 96-well micro-titer plate and 196 µL of Bradford solution and incubated for 5 min. The potassium phosphate buffer (50 mM, pH 8.0) was used as blank, and the absorption was measured after 5 min at 595 nm. The protein concentration was calculated using BSA as a standard (1-100 µg mL<sup>-1</sup>).

### 2.4. Evaluating Enzyme Performance on a Small Scale on Different Powder and Films

For all the experiments, 13.0±0.5 mg) of PET amorphous film (size 0.5×1 cm, thickness 0.25 mm GoodFellow) or powder (Table 1) were incubated with 0.5 mg g<sup>-1</sup> PET of purified enzyme (FastPETase, LCC, and LCC LCC<sup>ICCG</sup>) in 1.0 mL buffer (50 mM phosphate buffer, pH 8.0) in separate reactions. The Eppendorf tubes were incubated in a thermo-shaker (constant vertical shaking) at an optimum temperature of 50°C [13], 65°C and 72°C [19] for FastPETase and LCC, and LCC<sup>ICCG</sup> respectively. The blank (buffer) and control (film/powder and buffer without enzymes) were also incubated. The reaction was monitored for 96 h and the released monomers were quantified using HPLC-DAD analysis. The monomer concentration is expressed and presented as molar yield (Supplementary file. S1.1).

### 2.5. Evaluation of Different Reaction Variables for PET Hydrolysis

Different 50 mM buffer systems, such as KH<sub>2</sub>PO<sub>4</sub>-K<sub>2</sub>HPO<sub>4</sub> (pH 6.5-7.5), KH<sub>2</sub>PO<sub>4</sub>-NaOH (pH 8.0-9.0), glycine-NaOH (pH 9.0-10.0), NaHCO<sub>3</sub>-Na<sub>2</sub>CO<sub>3</sub> (pH 10.0-11.0), were tested for the depolymerization of PET film. Different enzyme concentrations (mg g<sup>-1</sup>PET) for FastPETase, LCC, and LCC<sup>ICCG</sup> (0.1, 0.25, 0.5, 1, and 2 mg g<sup>-1</sup> PET) were used to optimize the effect of enzyme loading on PET depolymerization. 13.0 mg of PET powder (0.08 mm cryo-milled amorphous) and film (0.2 mm amorphous) were incubated in 1 mL KH<sub>2</sub>PO<sub>4</sub>-NaOH buffer (50 mM, pH 8.0) for 96 h in different enzyme concentrations. The incubation temperature (30, 40, 50, 65, 72, and 80°C), and reaction time (2, 4, 8, 16, 24, 30, 36, 48, 60, 72, and 96 h) were also studied.

### 2.6. Evaluating Enzyme Performance in 10 mL (Without pH Control) and 1L Bioreactor (pH Control)

The depolymerization of PET film (0.25 mm amorphous), PET powder (0.08 mm amorphous), PET tray waste, and PET bottle waste from Spain was performed in 10 mL falcon tubes in the optimized conditions without any pH control and subsequently in a 1L bioreactor (Biostat A, Sartorius) under controlled conditions. 13 g L<sup>-1</sup> PET film was added to 1000 mL of 50 mM phosphate-NaOH buffer (pH 8.0) and 0.5 mg g<sup>-1</sup>PET (FastPETse) and 1.0 mg g<sup>-1</sup> PET (LCC and LCC<sup>ICCG</sup>) in separate reactions. The reaction was performed at 50, 65, and 72°C for FastPETase, LCC, and LCC<sup>ICCG</sup> respectively. The initial pH of the reaction was adjusted to 8.0 using 1M NaOH. The samples were collected at different time intervals and monomers were quantified using HPLC analysis. The

reaction was stopped after 96 h, residual PET films were recovered from the reaction, and weight loss (Supplementary file, S1.2) was measured.

### 2.7. Production of LCC<sup>ICCG</sup> in the Bioreactor and PET Hydrolysis

For seed preparation, *E. coli* containing LCC<sup>ICCG</sup> was streaked on LB agar plates (50 mg L<sup>-1</sup> Kanamycin) and incubated at 37°C overnight. One single colony was inoculated to 5 mL M9 medium (Table S3) in a 50 mL falcon for overnight growth at 37°C and 180 rpm. 1 mL of this seed culture was transferred to a 250-mL flask containing 50 mL fresh M9 medium to grow for another 12 h until an OD<sub>600</sub> of 1.2–1.8 was reached. This culture was used to inoculate the 2 L batch bioreactor (Biostat A, Sartorius) at 2% (v/v). The exponentially growing cells from the seed culture were transferred into the bioreactor to initiate the fermentation (*t* = 0 h). The dissolved oxygen level of the growth experiments was set at 20% of air saturation by cascade controls of agitation speed. A two-stage temperature control was implemented i.e., 37°C for the growth phase during 0–4 h and 30°C for enzyme production in the remainder of the run. The pH was maintained at 7.2 throughout the run by feeding 1 M sodium hydroxide solution or 1M HCl solution. When the O.D<sub>600</sub> reached 0.6–0.8, the temperature was lowered to 30°C and the protein production was induced with 1 mM IPTG and kept for growth till 24 h. The cell debris was removed by centrifugation at 10000 g for 15 min. The pH of the supernatant (crude LCC<sup>ICCG</sup>) was adjusted to 8.0 (using 1M NaOH) in the same reactor, and the reaction temperature was adjusted to 62°C (the maximum temperature that could be reached in the bioreactor) and at a fixed rpm speed of 600 using a Rushton impeller to provide efficient mixing. 13 g L<sup>-1</sup> PET material (film or powder) was added to the supernatant and monitored for 96 h. The pH of the reaction was maintained at 8.0 with 1M NaOH. The monomers and oligomers released were measured using HPLC at different intervals, and quantitative NMR was used to calculate the mass balance.

### 2.8. Depolymerization of Mixed PET Waste Using LCC<sup>ICCG</sup> in 1L

LCC<sup>ICCG</sup> was produced using *E. coli* BL21 cells as described in the above section in a 2L bioreactor as described earlier. After growth, the cell debris was removed and the supernatant was concentrated using a filtration device (10 kDa, Merck Millipore) to 1L. Different PET materials i.e. PET tray (commercial; 24 g L<sup>-1</sup>), PET tray waste (100 g L<sup>-1</sup>), and PET bottle waste (100 g L<sup>-1</sup>) were depolymerized in separate experiments using approximately 0.8 mg g<sup>-1</sup> PET of ICCG in a 1L bioreactor (Biostat A-Sartorius) under cascade pH control (pH 8.0 using 1M NaOH) and 62°C for 96 h.. The temperature regulation was performed in the water-jacketed bioreactor, and a Rushton impeller was used to maintain constant agitation at 600 rpm. The reaction was initiated by the addition of PET substrate (mix tray or bottle waste) into the reactor containing crude LCC<sup>ICCG</sup> at 62°C, 600 rpm, and cascade pH control at 8.0. The samples of 5 mL from the PET hydrolysis were collected at different time intervals from the sampling port. After the completion of the reaction, the residual PET and protein were removed by centrifugation. TPA was recovered from the reaction mixture using a modified protocol [26]. The purification of TPA from PET hydrolysate was performed by increasing the pH to over 9.0 to ensure the solubilization of all the monomers followed by centrifugation at 10000 rpm to remove any debris of denatured proteins. The pH of the supernatant was decreased (to 2.0) stepwise to monitor the precipitation of TPA. The precipitates were washed with cold acidified water, lyophilized, and analyzed by <sup>1</sup>H-NMR for purity.

### 2.9. Advanced Analysis

#### 2.9.1. High-Performance Liquid Chromatography (HPLC)

HPLC analysis was performed for all the PET hydrolysis reactions to quantify the monomers (TPA, MHET released. 500  $\mu$ L samples were taken, centrifuged at 20,000 rpm for 10 min, and the supernatant was diluted 1:1 with methanol. TPA, MHET, and BHET standards were prepared in 50% methanol (v/v). The samples were analyzed using a Dionex Ultimate 3000 system, fitted with a diode

array detector. A C18 column (Phenomenex Luna 5  $\mu$ m, 250 mm x 4.6 mm) at 30 °C was used for the separation of the products of the reaction, with 0.1% formic acid in water (A) and methanol (B) as the mobile phase. The solvent gradient was as follows: 10 % B (0-5 min), increased linearly to 50% (5-17 min), and 100 % B (17-20 min). The flow rate was 1 mL min<sup>-1</sup> and the injection volume was 2  $\mu$ L, with detection at 241 nm and a total run time of 20 min.

#### 2.9.2. Nuclear Magnetic Resonance (NMR)

Samples were prepared by mixing 500  $\mu$ L of supernatant with 25  $\mu$ L (200 mM) of 3-(Trimethylsilyl) -2,2,3,3-tetradeuteropropionic acid sodium salt (TSP-D<sub>4</sub>) in D<sub>2</sub>O. <sup>1</sup>H-NMR spectra were recorded on a BRUKER AVIII-600 MHz NMR spectrometer, equipped with a cryogenic CPP-TCI probe. Spectra were recorded at 298.1 K with a standard 1D pulse sequence with an acquisition time of 2.73 s (64k complex data points, spectral width of 20 ppm). The relaxation delay was set to 25.5 s. During the last 5 s of the relaxation delay, a weak continuous-wave irradiation of  $\gamma B_1/2\pi = 70$  Hz was applied for water suppression. Resonance assignment in product mixtures was done by comparing chemical shifts with spectra of pure standards. [11]

#### 2.9.3. Differential scanning Calorimetry (DSC) Analysis of PET Materials

The thermal behavior of the PET film and powder was determined by using DSC. The DSC experiment was carried out using NETZSCH Proteus® for Differential Scanning Calorimetry (DSC). In the first scan, each sample was heated to 300°C at a heating rate of 15°C/min. The sample was then cooled at the same rate before reheating the samples to 300°C. The test samples, weighing about 8.0±2.0 mg, were placed in a T-zero thermionic aluminum pan for analysis and all tests were carried out under a nitrogen atmosphere. Percentage of crystallinity (X<sub>c</sub>) was calculated based on the equation (Eq. 1) below:

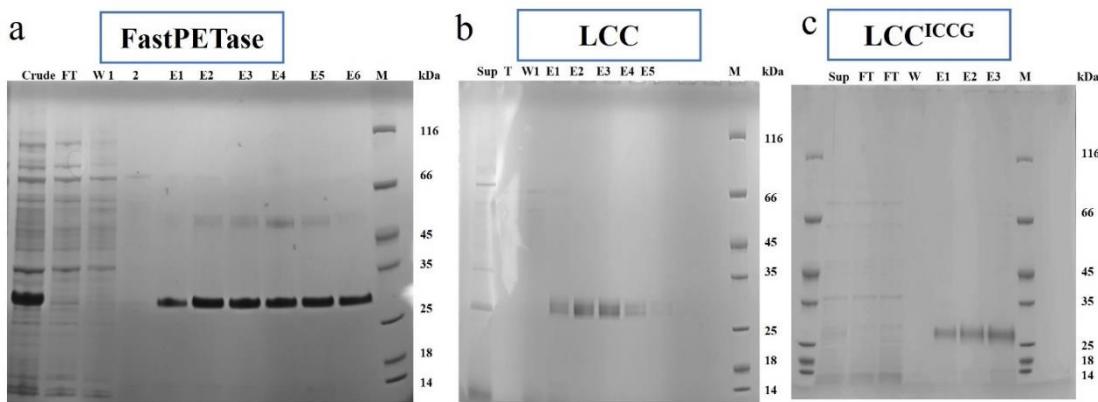
$$X_c = \frac{\Delta H_m - \Delta H_c}{wt * \Delta H_f} \times 100 \quad (1)$$

$\Delta H_m$  is the enthalpy of melting that can be determined by integrating the endothermic melting peak,  $\Delta H_c$  is the enthalpy of cold crystallization and determined by integrating the exothermic cold crystallization peak, wt is the weight fraction of polyester in the plastic, and  $\Delta H_f$  100% is the enthalpy of melting for a fully crystalline polymer and taken from literature as 140.1 J/g.

### 3. Results and Discussion

#### 3.1. Expression, Production, and Purification of FastPETase, and LCC, LCC<sup>ICCG</sup>

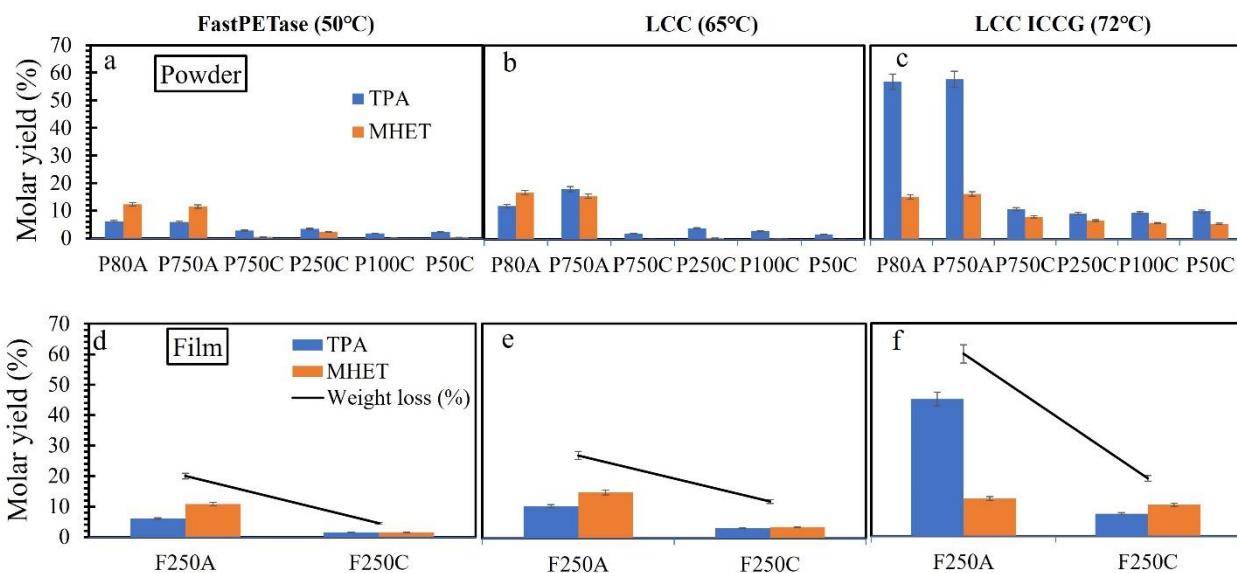
FastPETase, LCC, and LCC<sup>ICCG</sup> were expressed in the *Escherichia coli* strain BL21 (DE3). FastPETase was obtained from the cytoplasmic fraction (Figure 1A) after the cell lysis whereas LCC and LCC<sup>ICCG</sup> were obtained in extracellular supernatant (Figure 1b and c). LCC and LCC<sup>ICCG</sup> contain N-terminal pelB leader peptide that guides protein into the periplasmic space. The pelB peptide does not correspond to the active transport of the enzymes to extracellular space. However, the enzymes were recovered from the culture supernatant. It could be attributed to the phospholipase activity of these enzymes especially cutinases [28,29] which makes the cell membrane permeable and the cause leaking of the enzyme out of the cell. There was no cell cytotoxicity observed during the growth phase and enzyme production. After expression and purification, a final yield of 26, 35, and 32 mg L<sup>-1</sup> of culture was obtained (Table S1).



**Figure 1.** SDS PAGE analysis of (a) FastPETase, (b) LCC, and (c) LCC<sup>ICCG</sup>. Lanes contain different samples and are named as follows: cell lysate (crude), Extracellular Supernatant (sup), Flowthrough (FT) Wash (W), Eluates (E), and protein marker (M). The proteins appear as a single band (FastPETase 30.1 kDa) with actual molecular weight of 27.9 kDa, (LCC 29.9 kDa) with actual molecular weight of 29.0 kDa, and (LCC<sup>ICCG</sup> 27.3 kDa) with actual molecular weight of 28.6 kDa. The gels were prepared twice with the gel shown representative of the set.

### 3.2. Enzymatic Depolymerization of Different Powders and Films in Small-Scale

FastPETase, LCC, and LCC<sup>ICCG</sup> are among the best polyester hydrolases reported so far and are known to depolymerize aliphatic and aromatic polyesters including PET [2,17–19]. The purified enzymes were used for PET depolymerization experiments on a small scale (1 mL) without any pH control. Different PET powders and films have been used in this study and all the materials were characterized using DSC analysis (Tables 1 and S2, Figure S10). Preliminary tests were made, using the purified enzyme (0.5 mg g<sup>-1</sup>PET), 13 g L<sup>-1</sup> of PET film or powder, and 50 mM bicin NaOH buffer at the respective optimum temperature of the enzyme i.e. 50, 65, and 72°C for FastPETase, LCC and LCC<sup>ICCG</sup> in 1.0 mL reactions. After 96 h, the weight loss was measured for films and powders and the residual supernatant was analyzed by HPLC. FastPETase had lower molar yields (%) i.e., 6.2, 12.3 (P80A), and 6.0 and 10.8 % (P750A) of TPA and MHET respectively relative to the total PET compared to the other two enzymes (Figure 2a). LCC treated P80A and P750A reactions molar yield of 12 and 18.1% TPA and 16.7 and 15.5 % MHET respectively (Figure 2b). LCC<sup>ICCG</sup> exhibited maximum yield of TPA (57 and 58 mol%) and MHET (15.3 and 16 mol%) during depolymerization of cryo-milled PET powders (P80A, P750A) (Figure 2c). The depolymerization of PET 0.2 mm Goodfellow PET by FastPETase resulted in 20% weight loss (Figure 2d) which was lower than LCC<sup>ICCG</sup> (60%) and LCC (27%). LCC<sup>ICCG</sup> treated PET film (F250A) also had maximum weight loss (60%) and yielded 45.1% TPA and 12.3 % of MHET (Figure 2f).



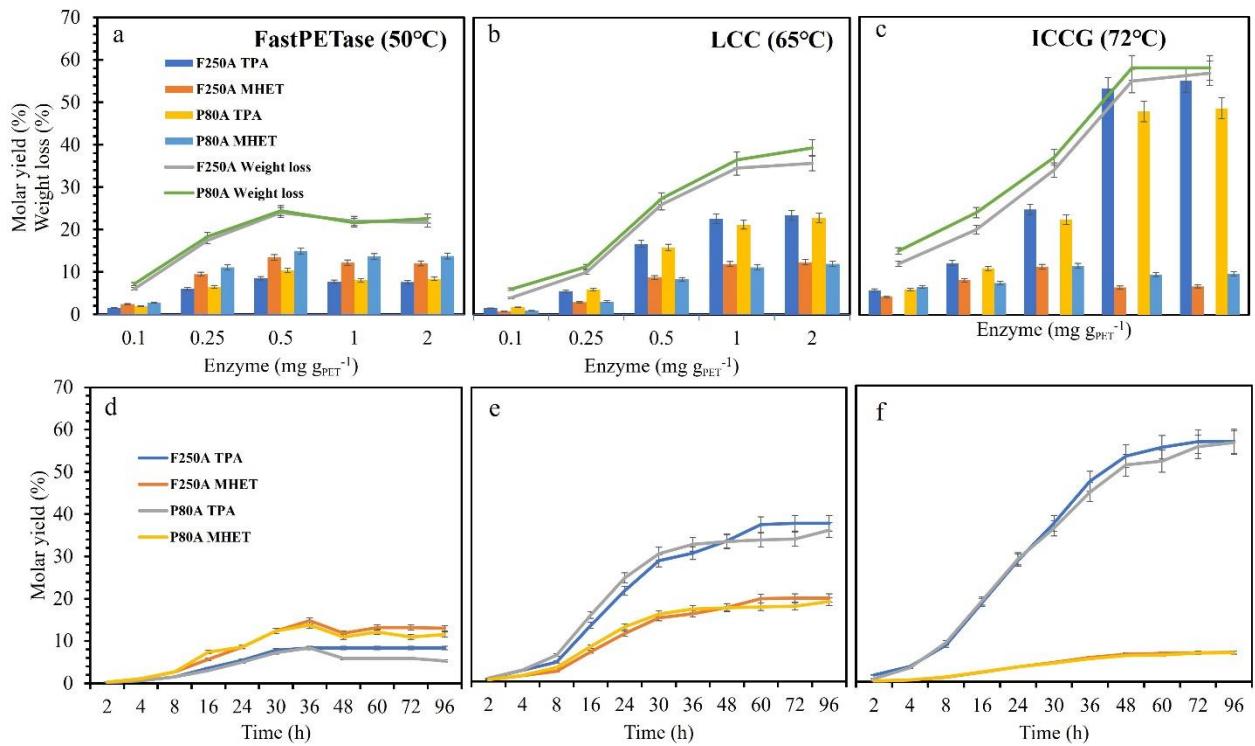
**Figure 2.** Depolymerization of different PET powders and films (amorphous and crystalline) by (a and d) FastPETase, (b and e) LCC, and (c and f) LCC<sup>ICCG</sup>. The reaction mixture was composed of 50 mM Bicin-NaOH buffer (pH 8.0), 13 g L<sup>-1</sup> PET powder or film, and 0.5 mg g<sup>-1</sup> PET enzyme. The reaction was incubated for 96 h at the respective optimum temperature of the enzymes i.e. 50°C (FastPETase), 65°C (LCC), and LCC<sup>ICCG</sup> (72°C) without any pH control.

The effect of increasing crystallinity of powder and films on the depolymerization and monomer release by these enzymes was evident where the enzyme efficiency was severely reduced both for powders and films (Figure 2a-f). LCC<sup>ICCG</sup> was the most active enzyme on crystalline PET (Figure 2f) but had 3 times less depolymerization than on amorphous PET. The crystallinity, chain mobility, molecular size, surface topography, and hydrophobicity of PET greatly influence enzyme performance [11,29–32]. A crystallinity exceeding 20% has been proposed to significantly impede the enzymatic depolymerization process as evidenced in quenched PET powders (27.6% crystallinity) [2,12]. Additionally, the crystalline PET powder of different sizes did not show any improvement in terms of total depolymerization by the enzymes (Figure 2a–c). Brizendine et al. [33] also demonstrated that particle size reduction of PET only affects the initial rate of hydrolysis but not the overall conversion.

### 2.3. Optimization of Different Parameters for Enzymatic PET Hydrolysis

After the preliminary tests, P80A and F250A were used in subsequent experiments. Enzyme loading for efficient conversion is an important parameter of enzymatic processes such as PET hydrolysis. PET weight loss and monomer release reached a maximum (24.4% weight loss, molar yield of 10.3 and 15% of TPA and MHET) when 0.5 mg g<sup>-1</sup> PET FastPETase was used at 50°C in potassium phosphate-NaOH buffer (pH 8.0) after 96 h (Figure 3a). Interestingly, the PET hydrolysis did not improve when the concentration of FastPETase was increased to 1 and 2 mg g<sup>-1</sup> PET. A recent study showed the low thermostability of FastPETase during PET hydrolysis at different reactor scales compared to other variants [2]. LCC on the other hand showed increased depolymerization of both powder and film (Figure 3b) with an increase in enzyme loading, reaching a maximum weight loss of 32 % at a concentration of 1 mg g<sup>-1</sup> PET. The performance of ICCG at 1 mg g<sup>-1</sup> PET enzyme loading was superior where the depolymerization was approximately 58% and TPA and MHET molar yield of 55% and 7 % respectively. LCC<sup>ICCG</sup> has been reported to convert 200 g L<sup>-1</sup> amorphized PET to 90% over 10 h at 3 mg g<sup>-1</sup> PET enzyme loading [19]. Similar results were obtained by Brizendine et al. [33] when depolymerization was performed at 65°C and 100 g L<sup>-1</sup> loading of PET film (1x1 cm) but in 48 h. The amount of TPA compared to MHET also kept increasing (Figure 3c) with increased enzyme loading. A possible explanation could be that the enzyme performs endo cleavage first on the

polymer, so at lower enzyme concentrations, more oligomers are released than monomers or MHET is primarily released but then hydrolyzed further.



**Figure 3.** Depolymerization (Weight loss and monomer release) of PET powder (P80A) and film (F250A) by (a) FastPETase, (b) LCC, and (c) LCC<sup>ICCG</sup>. The reaction mixture contained 50 mM KH<sub>2</sub>PO<sub>4</sub>-NaOH buffer (pH 8.0), 13 g L<sup>-1</sup> PET powder, or film at different enzyme loading (0.1, 0.25, 0.5, 1.0, and 2.0 mg g<sup>-1</sup> PET). It was incubated for 96 h at the respective optimum temperature of the enzymes i.e., 50°C (FastPETase), 65°C (LCC), and LCC<sup>ICCG</sup> (72°C) without any pH control. Depolymerization (monomer release) of PET powder (P80A) and film (F250A) by (d) FastPETase (0.5 mg g<sup>-1</sup> PET), (e) LCC (1 mg g<sup>-1</sup> PET), and (f) LCC<sup>ICCG</sup> (1 mg g<sup>-1</sup> PET) at different time intervals (2, 4, 8, 16, 24, 30, 36, 48, 60, 72, and 96 h) in 50 mM KH<sub>2</sub>PO<sub>4</sub>-NaOH buffer (pH 8.0), 13 g L<sup>-1</sup> PET powder, or film.

The effect of using different buffers and pH for PET hydrolysis was investigated. FastPETase (50°C), LCC (65°C), and LCC<sup>ICCG</sup> (72°C) treated PET exhibited weight loss in all buffer solutions with a pH ranging from 6.5 to 11.0 (Figure S2a-c). The weight loss was maximum in 50 mM potassium phosphate NaOH buffer (at pH 8.0) for all three enzymes and efficient depolymerization in the pH range of 7.0-9.0. Also PET powder depolymerization was most efficient in 50 mM KH<sub>2</sub>PO<sub>4</sub>-NaOH buffer (pH 8.0 and 9.0) for FastPETase (22%), LCC (31%) and LCC<sup>ICCG</sup> (58%). There was a rapid decline in depolymerization efficiency as the pH dropped below 7.0 due to the release of acidic hydrolysis products (TPA and MHET) which decreased the reaction mixture's pH.

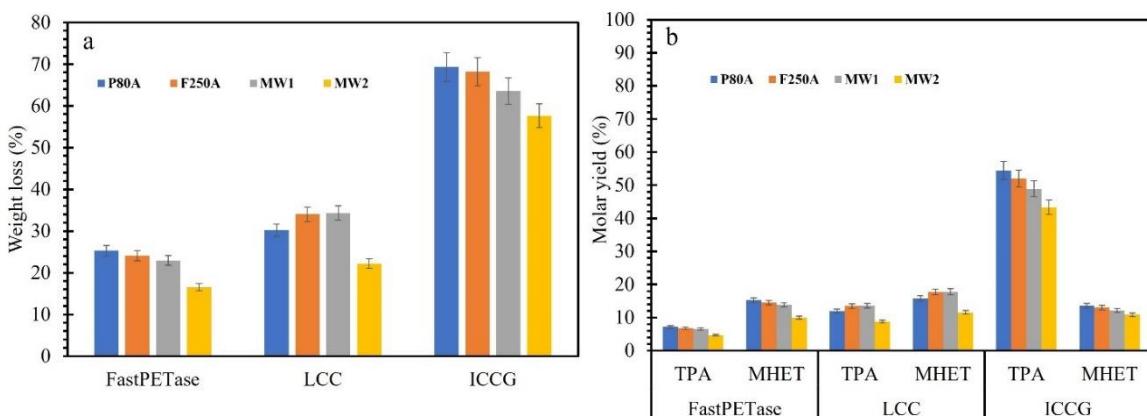
Interestingly, LCC<sup>ICCG</sup> catalyzed PET hydrolysis produced more TPA than MHET compared to LCC and FastPETase in the buffer conditions tested. Here, the lower molarity (50 mM) was chosen to mimic the industrial reaction environments where hydrolysis is performed in water rather than specialized buffered solutions to reduce operational costs [2].

The temperature of incubation was also optimized for these enzymes (Figure S3a-c). All these enzymes exhibited maximum depolymerization at their respective reported optimum temperature i.e. 50°C, 65°C, and 72°C for FastPETase, LCC, and LCC<sup>ICCG</sup> respectively [17-19] (Figure S3). As expected, the thermostability analysis suggested that FastPETase is least stable in all tested temperatures (50, 60, and 72°C) after 36h followed by LCC and LCC<sup>ICCG</sup> (Figure S4). Furthermore, PET hydrolysis efficiency was monitored over time. It can be seen from Figure 3d that FastPETase

achieved maximum depolymerization and molar yield (13.75% MHET and 8 % TPA) in 33 h. The depolymerization process slowed down after 36 h and no significant changes were observed after longer incubation times (Figure 2c). This can be due to poor thermostability and short half-life of FastPETase, changes in pH or a combination thereof [2]. LCC showed an improved depolymerization performance (Figure 3e). The released monomer (for both powder and film) increased with prolonged incubation time and molar yield of 26% and 24% of TPA and 14.5% and 13% of MHET (P80A and F250A) in 60 h. LCC<sup>ICCG</sup> also exhibited maximum depolymerization after 60 h (Figure 3f) but with a higher molar yield of TPA (54%) and less MHET (7%). The TPA yield marginally increased to 56.5% after 72 hours and remained constant after that. Brizendine et al. [33] observed differences between the large-scale (bioreactor conditions) and small-scale reactions for MHET concentrations where large-scale reactions generally built up less MHET than small-scale ones. LCC<sup>ICCG</sup> will preferentially release TPA due to the inhibitory effect of MHET. The reactions can be complemented by adding a carboxylesterase or MHETase to further improve the depolymerization efficiency [34,35].

#### 2.4. PET Depolymerization in 10 mL

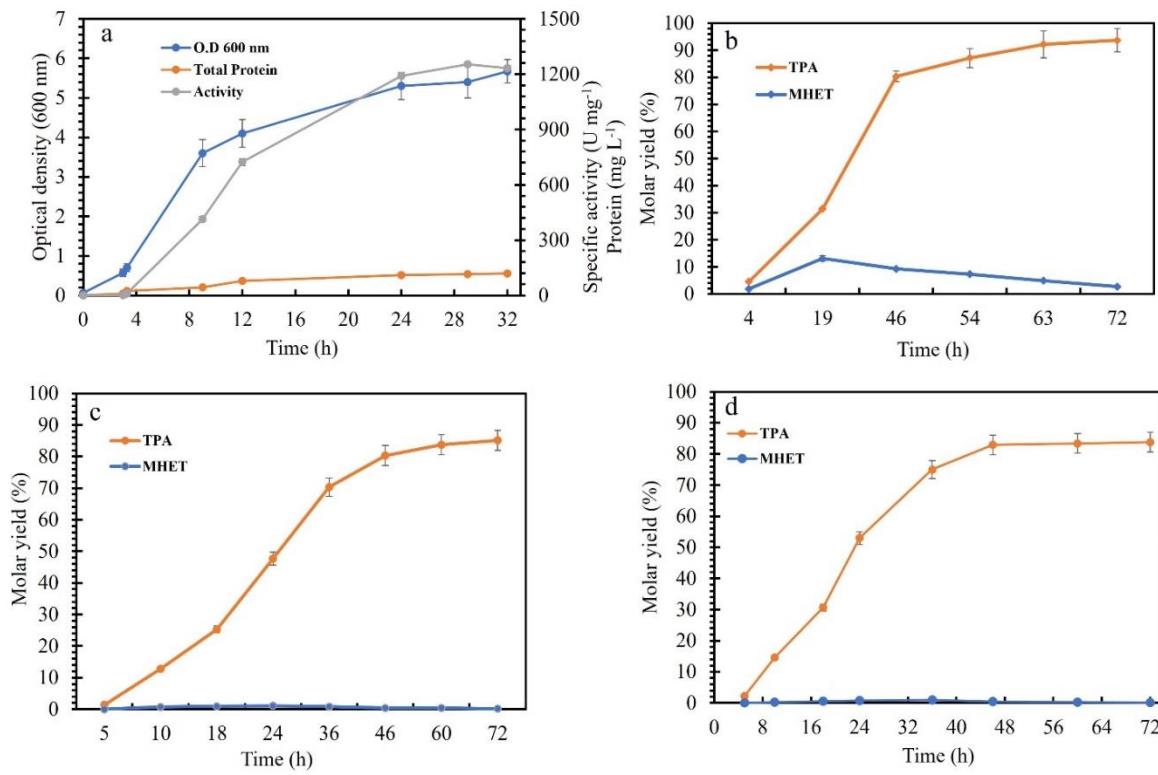
The previous experiments were performed on a 1 mL scale (50 mM phosphate-NaOH, pH 8.0) without a constant pH control, which may have a significant effect on the overall depolymerization of PET [2]. First scale-up (10 mL) was performed in the reaction conditions optimized earlier with P80A, F250A, and mixed PET waste MW1, and MW2 without any pH control. The overall depolymerization improved compared to the 1 mL scale. The depolymerization of P80A, F250A, and mixed PET waste MW1 (Xc16%), and MW2 (Xc 28%) by FastPETase led to a weight loss of 25, 24, 23, and 15% respectively (Figure 4a). LCC catalyzed depolymerization of PET reached 34% weight loss for F250A and MW1, but only 22% weight loss for MW2. ICCG outperforms the other two in terms of weight loss and monomer molar yield for all the substrates tested (Figure 4a). A weight loss of 69, 68, 63, and 56% was recorded for the depolymerization of P80A, F250A, MW1, and MW2 respectively. The release of TPA was maximum for LCC<sup>ICCG</sup> followed by LCC and FastPETase (Figure 2b). The interesting result is the depolymerization of MW2 (Xc=28%) by ICCG where 57% weight loss (molar yield of 43% TPA and 11% MHET) was achieved compared to LCC (22% weight loss, 8.9% and 11.6% molar yield of TPA and MHET). There is a difference in the observed weight loss of powders and the mass loss obtained from molar yield due to the loss of powder particles during washing, processing, and analysis. Thomsen et al. [31] demonstrated that LCC<sup>ICCG</sup> performs relatively better than PETase at higher crystallinity (15-25%), however, the depolymerization efficiency was decreased as the crystallinity was increased.



**Figure 4.** (a) Depolymerization (Weight loss) of PET powder (P80A) and film (F250A), mixed PET waste MW1 and MW2 by FastPETase, LCC, and LCC<sup>ICCG</sup>. The reaction was performed in a 10 mL volume containing 50 mM KH<sub>2</sub>PO<sub>4</sub>-NaOH buffer (pH 8.0), 13 g L<sup>-1</sup> PET powder, or film or MW 1 mg g<sup>-1</sup> enzyme for 48 h at the respective optimum temperature of the enzymes i.e., 50°C (FastPETase), 65°C (LCC), and LCC<sup>ICCG</sup> (72°C) without any pH control. (b) The released monomers were measured by HPLC after the completion of the reaction and expressed as molar yield (%).

## 2.5. Depolymerization of Different PET Substrates Using Crude LCC<sup>ICCG</sup> in a Bioreactor (1L)

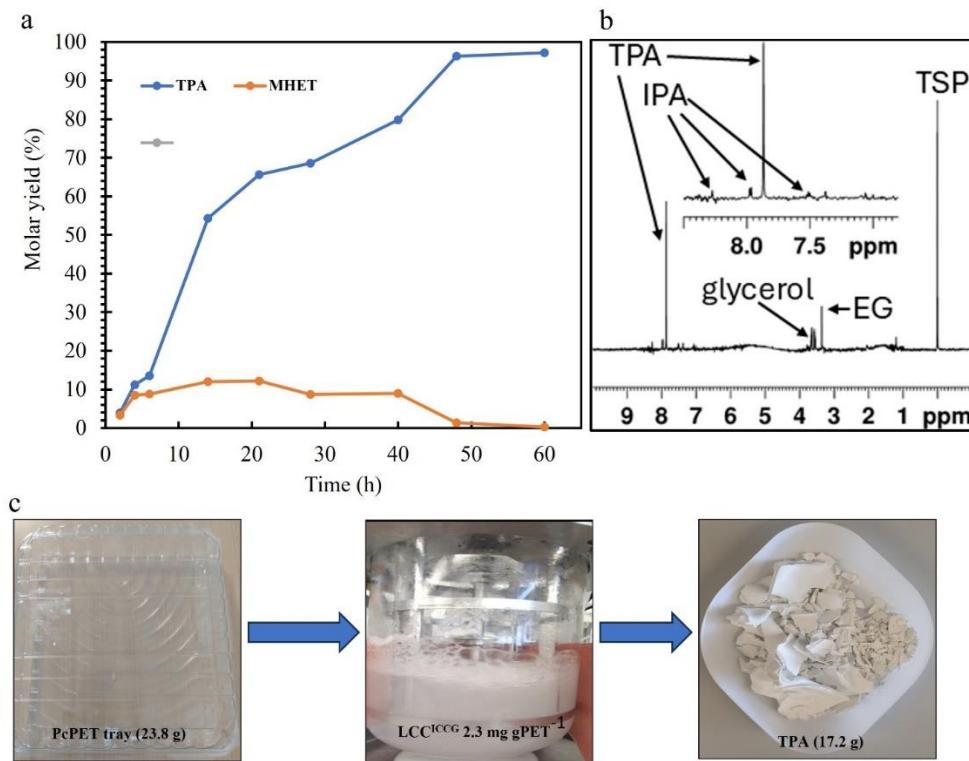
In the PET hydrolytic process, pH is a crucial factor with a huge impact on the overall depolymerization. We performed depolymerization of different PET substrates (PET film, Powder) in the 1L reactor in the optimized conditions i.e., 13 g L<sup>-1</sup> substrate loading (PET film or powder). The *E. coli* BL21 cells containing LCC<sup>ICCG</sup> were grown in an M9 medium (0.1% ammonium chloride and 0.5% glycerol, 20% pO<sub>2</sub>, pH 7.2). After induction with IPTG, the cells were grown for 32 h (Figure 5a). The final OD<sub>600</sub> of 5.7, total protein content in the supernatant of 119 mg L<sup>-1</sup>, and pNPB activity of 1232 U mg<sup>-1</sup> was reached in the bioreactor. Similarly, a fed-batch fermentation (1L) using LB medium or autoinduction medium with glucose feeding and IPTG induction led to the production of 12 g L<sup>-1</sup> total protein (10-15% of the target protein LCC) [36] which is higher than the current study. However, it is to be noted that in the current study, the production of the enzyme was performed in a minimal medium and 0.2 % glycerol in a batch reactor. 30% of the total protein produced extracellularly accounts for LCC<sup>ICCG</sup>. The enzyme cost during the PET hydrolysis is a key cost driver and a balanced production process with lower costs and higher enzyme yields will be an advantage [22]. After removing cell debris, one-half of the supernatant (1L) was used for purification and the other half for the bioprocess using the crude enzyme. With the purified enzyme-catalyzed reaction with film F250A, MHET concentration reached a maximum molar yield of 13.1 % after 19 h and decreased after that, whereas TPA formation reached the maximum of 94% molar yield in 72 h (Figure 5b) and was also verified by quantitative NMR (Figure S5). However, almost 90% of TPA was released in 25 h showing the efficiency of this enzyme. The molar yield of TPA (Figure 5c and S6) and powder using the crude supernatant reached 90% in approximately 46 h. However, MHET release was very low compared to TPA. NMR spectrum revealed the presence of isophthalic acid and some unknown aromatic compounds (Figure S5 and Figure S6). Another interesting difference is the lag phase during the initial hours (5-7 h) for PET film (using a purified enzyme and crude supernatant) which was not evident for powder. Brizendine et al. [33] also observed a lag phase 1x1cm films which seemed to be dependent on the surface area and enzyme concentration. It can be explained by the fact that the enzyme performs an endo cleavage on PET which would not release monomers during the initial phase [31,33]. They also reported that PET particle size does not have an appreciable effect on total depolymerization [33,37,38].



**Figure 5.** (a) Growth profile, total protein content in the supernatant, and specific activity of LCC<sup>ICCG</sup> producing *E. coli* BL21 cells in a 2L fermenter after induction with IPTG at 2.2 hours (O.D<sub>600</sub> 0.6-0.8). (b) Depolymerization profile (monomer release) of PET film (F250A) using the purified LCC<sup>ICCG</sup>, (c) depolymerization of PET film (F250A), and (d) PET powder (P80A) using crude supernatant of LCC<sup>ICCG</sup>. The reaction was performed in a 1L volume containing 50 mM KH<sub>2</sub>PO<sub>4</sub>-NaOH buffer (pH 8.0), 13 g L<sup>-1</sup> PET powder, or film, and 1 mg g<sup>-1</sup> PET enzyme at 62°C with pH control (1M NaOH). The monomer release is measured by HPLC.

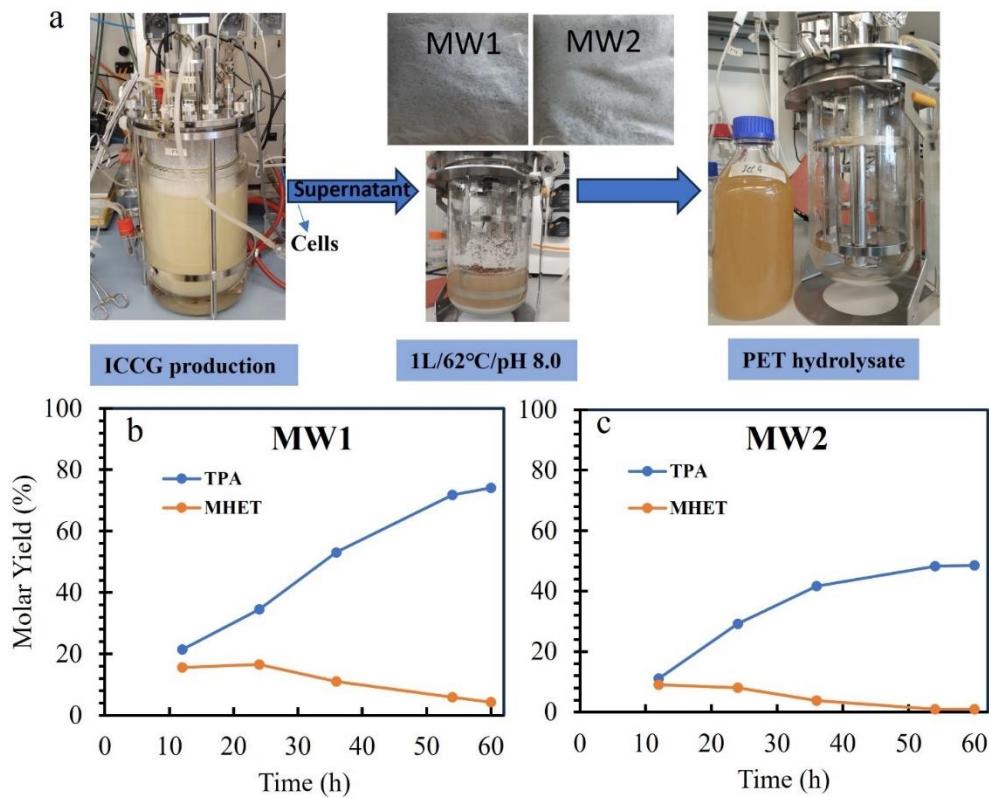
## 2.6. Mixed PET Waste Depolymerization

The depolymerization efficiency using crude enzyme was further tested on post-consumer PET tray from supermarket (used for packaging). The PET tray weighing 23.8 g was cut into pieces (Figure 6) and mixed with ICCG crude supernatant (2.3 mg g<sup>-1</sup> PET) and incubated at 62°C. A 94% depolymerization (based on residual films) of PET was achieved in 60 h with a TPA molar yield of 97% as measured by HPLC (Figure 6a) and 17.2 g TPA (90% molar yield) was recovered (Figure 6b, c). Tournier et al. [19] reported that LCC<sup>ICCG</sup> (3 mg g<sup>-1</sup> PET) can depolymerize 90% of 200 g L<sup>-1</sup> PET (amorphized and micronized) in 10 h at 72°C and a similar conversion was reached at 100 g L<sup>-1</sup> PET (1x1 cm) and 65°C but in 48 h [33].



**Figure 6.** (a) Depolymerization profile (mol% of TPA and MHET relative to PET) of pcPET tray (1x1 cm) using the LCC<sup>ICCG</sup>. The reaction was performed in a 1L volume containing 50 mM KH<sub>2</sub>PO<sub>4</sub>-NaOH buffer (pH 8.0), 2.3 mg g<sup>-1</sup> PET of crude LCC<sup>ICCG</sup>, 23.8 g L<sup>-1</sup> PET tray at 62°C with pH control (1M NaOH). The monomers released were measured using HPLC. (b) NMR spectrum of TPA recovered after 60 h. Apart from TPA, the sample contains IPA which is part of commercial PET, EG from the hydrolysis of PET, glycerol from the culture medium, and TSP-d4 as a chemical shift standard. (c) Post-consumer PET tray used for the enzymatic reaction and TPA recovered after the hydrolysis.

In the present study, 94% conversion was reached in 60 h at 2.4 g L<sup>-1</sup> PET loading with the crude supernatant but at a slightly lower temperature i.e., 62°C. Furthermore, we tested the crude supernatant on MW1 (mixed PET waste tray from Spain) and MW2 (mixed PET waste bottles from Spain) at 100 g L<sup>-1</sup> PET loadings and 2.5 mg g<sup>-1</sup> PET of crude enzyme (Figure 7a and S7). It is interesting to notice that the depolymerization of MW1 ( $X_c=16\%$ ) was nearly 78% in 60 h (Figure 7b) compared to MW2 ( $X_c=28\%$ ) where 50% depolymerization was recorded. The effect of increasing crystallinity was evident in the overall depolymerization as previously reported [30,32].



**Figure 7.** (a) Schematic representation of  $\text{LCC}^{\text{ICCG}}$  production using recombinant *E. coli* BL21 cells in 2L reactor. The supernatant was concentrated 2 times using ultrafiltration (10 kDa). The mixed plastic wastes MW1 and MW2 were added to the supernatant in separate reactions (100 g  $\text{L}^{-1}$  PET). The reaction was performed in 1L volume containing approximately 2.5 mg  $\text{g}^{-1}$  PET of crude  $\text{LCC}^{\text{ICCG}}$  at 62°C with pH control (1M NaOH). HPLC measurements of TPA and MHET during depolymerization of mixed PET waste (b) MW1, and (c) MW2.

Currently,  $\text{LCC}^{\text{ICCG}}$  is among the best enzymes that depolymerize crystalline PET ( $X_c < 20\%$ ) as efficiently as amorphous PET. However, the depolymerization activity decreases sharply after the crystallinity is increased as observed in this work and reported by others [31,39]. The NMR analysis of the MW1 and MW2 hydrolysate revealed the presence of isophthalic acid, an unknown aromatic compound, and oligomers of MHET and BHET other than the TPA, MHET and EG (Figure S8 and S9). TPA tends not to be very soluble in the reaction mixture and will certainly affect the mixing and mass transfer. However, it has been reported that the addition of NaOH neutralizes the acidic products released during PET depolymerization. The base addition also leads to the formation of soluble disodium terephthalate (13% w/w) between 25 and 70 °C [2,40]. The current process runs at 100 g/L substrate loading (10%), so the process is well below these limits. If the process is run at 20-30% solid loadings, the problem of solid mixing and mass transfer will arise due to insoluble TPA. However, it can be negated using higher concentrations of NaOH (to form TPA salt) and ensuring that TPA salt concentrations do not go beyond the solubility limits. Recently, Chen et al. [41] expressed and secreted FastPETase in *Pichia pastoris* and used crude supernatant to depolymerize 90% of 5 g  $\text{L}^{-1}$  PET (0.5%) in 18 h in a 10 L reactor. Similarly, LCC variants were produced in *E. coli*, and crude supernatant was applied to degrade 10 g  $\text{L}^{-1}$  PET with 97.5% depolymerization [42]. Table S4 summarizes recent studies on PET wastes at the bioreactor scale. However, in the current work, 90% depolymerization of 100 g  $\text{L}^{-1}$  PET (10%) mixed PET tray waste was achieved in a 1L reactor. The use of enzymes in a bioprocess such as this presents a sustainable PET utilization strategy without the need for enzyme purification. However, microbial strains with high expression and secretion ability would be highly beneficial for such processes to be successful.

#### 4. Conclusion

We evaluated the depolymerization efficiency of three efficient PET degrading enzymes namely FastPETase, LCC, and LCC<sup>ICCG</sup> on different powder and films in several conditions (enzyme loading, buffer, temperature, time) on a small scale and concluded that LCC<sup>ICCG</sup> is the best-performing enzyme on amorphous and crystalline PET, and mixed PET waste (tray and bottle waste). As reported earlier, the particle size does not affect the overall depolymerization process. However, if crystallinity of the substrate exceeds 25%, it becomes a bottleneck for efficient depolymerization. The LCC<sup>ICCG</sup> production was scaled to 2L and PET depolymerization of >90% was achieved in 50 h at 1.3% and 2.4% loading of amorphous PET and postconsumer PET respectively. Furthermore, 78% of 100 gL<sup>-1</sup> MW1 (X<sub>c</sub>=16%) and 50% of MW2 (X<sub>c</sub>=28%) were depolymerized in 60 h showing the effect of an increase in crystallinity on the depolymerization rate. In Summary, this work provides important foundation steps that can be extended to high-expression hosts to minimize the cost of enzyme purification and further improve the PET depolymerization steps.

**Author Contributions:** Virender Kumar: Conceptualization, Methodology, Investigation, Formal analysis, Writing an original draft. Reinhard Wimmer: Investigation, Formal analysis, Editing. Cristiano Varrone: Conceptualization, Methodology, Investigation, Formal Analysis, Review and Editing, Supervision, Funding Acquisition.

**Funding:** H2020 UPLIFT project (Grant agreement no. 953073).

**Data Availability Statement:** Data will be made available on request.

**Acknowledgments:** This work is funded by the European Union's Horizon 2020 Research and Innovation program under grant agreement No 953073, project UPLIFT (sUstainable PLastIcs for the Food and drink packaging indusTry). The authors also thank Michelle Hyldegård Mortensen for her assistance with the DSC analyzer. The authors are also thankful to Alessandro Pellis, University of Genova, Italy for providing the PET powder of different sizes and AIMPLAS, Valencia Spain for the supply of mixed PET waste. The NMR laboratory at Aalborg University is supported by the Obel Family, SparNord, and Carlsberg foundations.

**Conflicts of Interest:** The authors declare no competing financial interest.

## References

1. Tournier, V.; Duquesne, S.; Guillamot, F.; Cramail, H.; Taton, D.; Marty, A.; André, I. Enzymes' Power for Plastics Degradation. *Chemical Reviews* 2023, 123 (9). <https://doi.org/10.1021/acs.chemrev.2c00644>.
2. Arnal, G.; Anglade, J.; Gavalda, S.; Tournier, V.; Chabot, N.; Bornscheuer, U. T.; Weber, G.; Marty, A. Assessment of Four Engineered PET Degrading Enzymes Considering Large-Scale Industrial Applications. *ACS Catalysis* 2023, 13 (20), 13156–13166. <https://doi.org/10.1021/acscatal.3c02922>.
3. Zhang, S.; Li, M.; Zuo, Z.; Niu, Z. Recent Advances in Plastic Recycling and Upgrading under Mild Conditions. *Green Chemistry* 2023, 25 (18), 6949–6970. <https://doi.org/10.1039/D3GC01872E>.
4. Gibb, B. C. Plastics Are Forever. *Nature Chemistry* 2019, 11 (5), 394–395. <https://doi.org/10.1038/s41557-019-0260-7>.
5. Yang, W.; Wang, J.; Jiao, L.; Song, Y.; Li, C.; Hu, C. Easily Recoverable and Reusable P-Toluenesulfonic Acid for Faster Hydrolysis of Waste Polyethylene Terephthalate. *Green Chemistry* 2022, 24 (3), 1362–1372. <https://doi.org/10.1039/D1GC04567A>.
6. Raheem, A. B.; Noor, Z. Z.; Hassan, A.; Abd Hamid, M. K.; Samsudin, S. A.; Sabeen, A. H. Current Developments in Chemical Recycling of Post-Consumer Polyethylene Terephthalate Wastes for New Materials Production: A Review. *Journal of Cleaner Production* 2019, 225, 1052–1064. <https://doi.org/10.1016/j.jclepro.2019.04.019>.
7. Ragaert, K.; Delva, L.; Van Geem, K. Mechanical and Chemical Recycling of Solid Plastic Waste. *Waste Management* 2017, 69, 24–58. <https://doi.org/10.1016/j.wasman.2017.07.044>.
8. Zhang, S.; Hu, Q.; Zhang, Y.; Guo, H.; Wu, Y.; Sun, M.; Zhu, X.; Zhang, J.; Gong, S.; Liu, P.; Niu, Z. Depolymerization of Polyesters by a Binuclear Catalyst for Plastic Recycling. *Nature Sustainability* 2023, 6 (8), 965–973. <https://doi.org/10.1038/s41893-023-01118-4>.
9. Li, M.; Zhang, S. Tandem Chemical Depolymerization and Photoreforming of Waste PET Plastic to High-Value-Added Chemicals. *ACS catalysis* 2024, 14 (5), 2949–2958. <https://doi.org/10.1021/acscatal.3c05535>.
10. Niu, Z.; Zhang, S.; Xue, Y.; Wu, Y.; Zhang, Y.-X.; Tan, T. PET Recycling under Mild Conditions Via Substituent-Modulated Intramolecular Hydrolysis. *Chemical science* 2023, 14 (24), 6558–6563. <https://doi.org/10.1039/d3sc01161e>.
11. Jaime-Azuara, A.; Thomas Helmer Pedersen; Wimmer, R. Process Optimization by NMR-Assisted Investigation of Chemical Pathways during Depolymerization of PET in Subcritical Water. *Green Chemistry* 2023, 25 (7), 2711–2722. <https://doi.org/10.1039/d2gc04831k>.

12. Cui, Y.; Chen, Y.; Liu, X.; Dong, S.; Tian, Y.; Qiao, Y.; Mitra, R.; Han, J.; Li, C.; Han, X.; Liu, W.; Chen, Q.; Wei, W.; Wang, X.; Du, W.; Tang, S.; Xiang, H.; Liu, H.; Liang, Y.; Houk, K. N. Computational Redesign of a PETase for Plastic Biodegradation under Ambient Condition by the GRAPE Strategy. *ACS Catalysis* 2021, 11 (3), 1340–1350. <https://doi.org/10.1021/acscatal.0c05126>.
13. Bell, E. L.; Smithson, R.; Kilbride, S.; Foster, J.; Hardy, F. J.; Ramachandran, S.; Tedstone, A. A.; Haigh, S. J.; Garforth, A. A.; Day, P. J. R.; Levy, C.; Shaver, M. P.; Green, A. P. Directed Evolution of an Efficient and Thermostable PET Depolymerase. *Nature Catalysis* 2022, 5 (8), 673–681. <https://doi.org/10.1038/s41929-022-00821-3>.
14. Pfaff, L.; Gao, J.; Li, Z.; Jäckering, A.; Weber, G.; Mičan, J.; Chen, Y.; Dong, W.; Han, X.; Feiler, C.; Ao, Y.; Christoffel; Bednář, D.; Palm, G. J.; Lammers, M.; Jiří Damborský; Strodel, B.; Liu, W.; Bornscheuer, U. T.; Wei, R. Multiple Substrate Binding Mode-Guided Engineering of a Thermophilic PET Hydrolase. *ACS Catalysis* 2022, 12 (15), 9790–9800. <https://doi.org/10.1021/acscatal.2c02275>.
15. Han, X.; Liu, W.; Huang, J.-W.; Ma, J.; Zheng, Y.; Ko, T.-P.; Xu, L.; Cheng, Y.-S.; Chen, C.-C.; Guo, R.-T. Structural Insight into Catalytic Mechanism of PET Hydrolase. *Nature Communications* 2017, 8 (1). <https://doi.org/10.1038/s41467-017-02255-z>.
16. Yoshida, S.; Hiraga, K.; Takehana, T.; Taniguchi, I.; Yamaji, H.; Maeda, Y.; Toyohara, K.; Miyamoto, K.; Kimura, Y.; Oda, K. A Bacterium That Degrades and Assimilates Poly(Ethylene Terephthalate). *Science* 2016, 351 (6278), 1196–1199. <https://doi.org/10.1126/science.aad6359>.
17. Lu, H.; Diaz, D. J.; Czarnecki, N. J.; Zhu, C.; Kim, W.; Shroff, R.; Acosta, D. J.; Alexander, B. R.; Cole, H. O.; Zhang, Y.; Lynd, N. A.; Ellington, A. D.; Alper, H. S. Machine Learning-Aided Engineering of Hydrolases for PET Depolymerization. *Nature* 2022, 604 (7907), 662–667. <https://doi.org/10.1038/s41586-022-04599-z>.
18. Sulaiman, S.; Yamato, S.; Kanaya, E.; Kim, J.-J.; Koga, Y.; Takano, K.; Kanaya, S. Isolation of a Novel Cutinase Homolog with Polyethylene Terephthalate-Degrading Activity from Leaf-Branch Compost by Using a Metagenomic Approach. *Applied and Environmental Microbiology* 2011, 78 (5), 1556–1562. <https://doi.org/10.1128/aem.06725-11>.
19. Tournier, V.; Topham, C. M.; Gilles, A.; David, B.; Folgoas, C.; Moya-Leclair, E.; Kamionka, E.; Desrousseaux, M.-L.; Texier, H.; Gavalda, S.; Cot, M.; Guémard, E.; Dalibey, M.; Nomme, J.; Cioci, G.; Barbe, S.; Chateau, M.; André, I.; Duquesne, S.; Marty, A. An Engineered PET Depolymerase to Break down and Recycle Plastic Bottles. *Nature* 2020, 580 (7802), 216–219. <https://doi.org/10.1038/s41586-020-2149-4>.
20. Kawai, F.; Kawabata, T.; Oda, M. Current State and Perspectives Related to the Polyethylene Terephthalate Hydrolases Available for Biorecycling. *ACS Sustainable Chemistry & Engineering* 2020, 8 (24), 8894–8908. <https://doi.org/10.1021/acssuschemeng.0c01638>.
21. Erickson, E.; Gado, J. E.; Luisana Avilán; Bratti, F.; Brizendine, R. K.; Cox, P. A.; Gill, R.; Graham, R.; Kim, D.-J.; Gerhard König; Michener, W. K.; Saroj Poudel; Ramirez, K. J.; Shakespeare, T. J.; Zahn, M.; Boyd, E. S.; Payne, C. M.; DuBois, J. L.; Pickford, A.; Beckham, G. T. Sourcing Thermotolerant Poly(Ethylene Terephthalate) Hydrolase Scaffolds from Natural Diversity. *Nature Communications* 2022, 13 (1). <https://doi.org/10.1038/s41467-022-35237-x>.
22. Singh, A.; Rorrer, N. A.; Nicholson, S. R.; Erickson, E.; DesVeaux, J. S.; Avelino, A. F. T.; Lamers, P.; Bhatt, A.; Zhang, Y.; Avery, G.; Tao, L.; Pickford, A. R.; Carpenter, A. C.; McGeehan, J. E.; Beckham, G. T. Techno-Economic, Life-Cycle, and Socioeconomic Impact Analysis of Enzymatic Recycling of Poly(Ethylene Terephthalate). *Joule* 2021, 5 (9), 2479–2503. <https://doi.org/10.1016/j.joule.2021.06.015>.
23. Kumar, V.; Pellis, A.; Wimmer, R.; Vladimir Popok; de, J.; Varrone, C. Efficient Depolymerization of Poly(Ethylene 2,5-Furanoate) Using Polyester Hydrolases. *ACS Sustainable Chemistry & Engineering* 2024, 12 (26), 9658–9668. <https://doi.org/10.1021/acssuschemeng.4c00915>.
24. Ribitsch, D.; Acero, E. H.; Greimel, K.; Eiteljörg, I.; Trotscha, E.; Freddi, G.; Schwab, H.; Guebitz, G. M. Characterization of a New Cutinase From *Thermobifida Alba* for PET-Surface Hydrolysis. *Biocatalysis and Biotransformation* 2011, 30 (1), 2–9. <https://doi.org/10.3109/10242422.2012.644435>.
25. Bradford, M. M. A Rapid and Sensitive Method for the Quantitation of Microgram Quantities of Protein Utilizing the Principle of Protein-Dye Binding. *Analytical Biochemistry* 1976, 72 (1-2), 248–254. [https://doi.org/10.1016/0003-2697\(76\)90527-3](https://doi.org/10.1016/0003-2697(76)90527-3)
26. Li, R.; Greenchem; Ismail, M.; Abouhmad, A.; Warlin, N.; Pyo, S.-H.; Örn, O.; Al-Rudainy, B.; Tullberg, C.; Zhang, B.; Hatti-Kaul, R. Cite This: *Green Chem.* 2024, 26, 3863. <https://doi.org/10.1039/d3gc04728h>.
27. Su, L.; Woodard, R. W.; Chen, J.; Wu, J. Extracellular Location of *Thermobifida Fusca* Cutinase Expressed in *Escherichia Coli* BL21(DE3) without Mediation of a Signal Peptide. *Applied and Environmental Microbiology* 2013, 79 (14), 4192–4198. <https://doi.org/10.1128/aem.00239-13>.
28. Lizhu Aer; Qin, H.; Wo, P.; Feng, J.; Tang, L. Signal Peptide Independent Secretion of Bifunctional Dual-Hydrolase to Enhance the Bio-Depolymerization of Polyethylene Terephthalate. *Bioresource Technology* 2024, 391, 129884–129884. <https://doi.org/10.1016/j.biortech.2023.129884>.
29. Taniguchi, I.; Yoshida, S.; Hiraga, K.; Miyamoto, K.; Kimura, Y.; Oda, K. Biodegradation of PET: Current Status and Application Aspects. *ACS Catalysis* 2019, 9 (5), 4089–4105. <https://doi.org/10.1021/acscatal.8b05171>.

30. Tarazona, N. A.; Wei, R.; Brott, S.; Pfaff, L.; Bornscheuer, U. T.; Lendlein, A.; Rainhard Machatschek. Rapid Depolymerization of Poly(Ethylene Terephthalate) Thin Films by a Dual-Enzyme System and Its Impact on Material Properties. *Chem catalysis* 2022, 2 (12), 3573–3589. <https://doi.org/10.1016/j.chelet.2022.11.004>.
31. Thomsen, T. B.; Hunt, C. J.; Meyer, A. S. Influence of Substrate Crystallinity and Glass Transition Temperature on Enzymatic Degradation of Polyethylene Terephthalate (PET). *New Biotechnology* 2022, 69, 28–35. <https://doi.org/10.1016/j.nbt.2022.02.006>.
32. Pasula, R. R.; Lim, S.; Ghadessy, F. J.; Sana, B. The Influences of Substrates' Physical Properties on Enzymatic PET Hydrolysis: Implications for PET Hydrolase Engineering. *Engineering Biology* 2022, 6 (1), 17–22. <https://doi.org/10.1049/engb.2.12018>.
33. Brizendine, R. K.; Erickson, E.; Haugen, S. J.; Ramirez, K. J.; Miscall, J.; Salvachúa, D.; Pickford, A. R.; Sobkowicz, M. J.; McGeehan, J. E.; Beckham, G. T. Particle Size Reduction of Poly(Ethylene Terephthalate) Increases the Rate of Enzymatic Depolymerization but Does Not Increase the Overall Conversion Extent. *ACS Sustainable Chemistry & Engineering* 2022, 10 (28), 9131–9140. <https://doi.org/10.1021/acssuschemeng.2c01961>.
34. Barth, M.; Honak, A.; Oeser, T.; Wei, R.; Belisário-Ferrari, M. R.; Then, J.; Schmidt, J.; Zimmermann, W. A Dual Enzyme System Composed of a Polyester Hydrolase and a Carboxylesterase Enhances the Biocatalytic Degradation of Polyethylene Terephthalate Films. *Biotechnology Journal* 2016, 11 (8), 1082–1087. <https://doi.org/10.1002/biot.201600008>.
35. Knott, B. C.; Erickson, E.; Allen, M. D.; Gado, J. E.; Graham, R.; Kearns, F. L.; Pardo, I.; Topuzlu, E.; Anderson, J. J.; Austin, H. P.; Dominick, G.; Johnson, C. W.; Rorrer, N. A.; Szostkiewicz, C. J.; Copié, V.; Payne, C. M.; Woodcock, H. L.; Donohoe, B. S.; Beckham, G. T.; McGeehan, J. E. Characterization and Engineering of a Two-Enzyme System for Plastics Depolymerization. *Proceedings of the National Academy of Sciences of the United States of America* 2020, 117 (41), 25476–25485. <https://doi.org/10.1073/pnas.2006753117>.
36. Ya-Hue Valerie Soong; Abid, U.; Chang, A. C.; Ayafor, C.; Patel, A.; Qin, J.; Xu, J.; Lawton, C.; Wong, H.; Sobkowicz, M. J.; Xie, D. Enzyme Selection, Optimization, and Production toward Biodegradation of Post-Consumer Poly(Ethylene Terephthalate) at Scale. *Biotechnology Journal* 2023. <https://doi.org/10.1002/biot.202300119>.
37. de Castro, A. M.; Carniel, A.; Nicomedes Junior, J.; da Conceição Gomes, A.; Valoni, É. Screening of Commercial Enzymes for Poly(Ethylene Terephthalate) (PET) Hydrolysis and Synergy Studies on Different Substrate Sources. *Journal of Industrial Microbiology and Biotechnology* 2017, 44 (6), 835–844. <https://doi.org/10.1007/s10295-017-1942-z>.
38. Castro, A. M. de; Carniel, A.; Stahelin, D.; Chinelatto Junior, L. S.; Honorato, H. de A.; de Menezes, S. M. C. High-Fold Improvement of Assorted Post-Consumer Poly(Ethylene Terephthalate) (PET) Packages Hydrolysis Using *Humicola Insolens* Cutinase as a Single Biocatalyst. *Process Biochemistry* 2019, 81, 85–91. <https://doi.org/10.1016/j.procbio.2019.03.006>.
39. Oh, Y.-R.; Jang, Y.-A.; Song, J. K.; Eom, G. T. Secretory Production of an Engineered Cutinase in *Bacillus Subtilis* for Efficient Biocatalytic Depolymerization of Polyethylene Terephthalate. *Bioprocess and Biosystems Engineering* 2022, 45 (4), 711–720. <https://doi.org/10.1007/s00449-022-02690-3>.
40. Rezazadeh, A.; Thomsen, K.; Gavala, H. N.; Skiadas, I. V.; Fosbøl, P. L. Solubility and Freezing Points of Disodium Terephthalate in Water–Ethylene Glycol Mixtures. *Journal of Chemical & Engineering Data* 2021, 66 (5), 2143–2152. <https://doi.org/10.1021/acs.jcd.1c00052>.
41. Chen, C.-C.; Li, X.; Min, J.; Zeng, Z.; Ning, Z.; He, H.; Long, X.; Niu, D.; Peng, R.; Liu, X.; Yang, Y.; Huang, J.-W.; Guo, R.-T. Complete Decomposition of Poly(Ethylene Terephthalate) by Crude PET Hydrolytic Enzyme Produced in *Pichia Pastoris*. *Chemical Engineering Journal* 2023, 148418–148418. <https://doi.org/10.1016/j.cej.2023.148418>.
42. Fritzsche, S.; Holger Hübner; Oldiges, M.; Castiglione, K. Comparative Evaluation of the Extracellular Production of a Polyethylene Terephthalate Degrading Cutinase by *Corynebacterium Glutamicum* and Leaky *Escherichia Coli* in Batch and Fed-Batch Processes. *Microbial Cell Factories* 2024, 23 (1). <https://doi.org/10.1186/s12934-024-02547-2>.

**Disclaimer/Publisher's Note:** The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.