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## Article

# Histochemical Localization and Cytotoxic Potential of Alkaloids in *Phaedranassa lehmannii*

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**Abstract:** Plants of the subfamily Amaryllidoideae are a source of unique and bioactive alkaloids called Amaryllidaceae alkaloids. The study of their anticancer potential has intensified in recent years. This work aims to locate and characterize the profile of cytotoxic alkaloids biosynthesized and stored in different tissues of *Phaedranassa lehmannii* Regel using different histochemical methods and chromatographic analysis. The histochemical analysis in the bulbs revealed the presence of alkaloids at the basal edge of scale-like leaves and bud apical zone. The GC-MS analysis indicated that the bulbs biosynthesize alkaloids distributed in crinine- (9.80 µg/g DW), galanthamine- (8.04 µg/g DW), lycorine- (7.38 µg/g DW) and narciclasine-type (3.75 µg/g DW). The root biosynthesizes alkaloids distributed mostly in lycorine- (225.29 µg/g DW) and galanthamine-type (0.723 µg/g DW) alkaloids. The total alkaloids biosynthesized by the root (324.93 µg/g DW) exceeded eleven times the abundance of the alkaloids identified in the bulbs (28.97 µg/g DW). In addition, the total alkaloids fractions exhibited a cytotoxic effect dose-dependent in the evaluated concentrations with IC<sub>50</sub> values of 11.76 ± 0.99 µg/mL and 2.59 ± 0.56 µg/mL against human lung (A549) cancer cells and 8.00 ± 1.35 µg/mL and 18.74 ± 1.99 µg/mL against gastric (AGS) cancer cells. The present study provided evidence to locate and characterize the alkaloids of *P. lehmannii* grown under nursery conditions as a species producing potential antiproliferative alkaloids.

**Keywords:** *Phaedranassa lehmannii*; amaryllidaceae alkaloids; cytotoxicity; histochemical localization

## 1. Introduction

The family Amaryllidaceae is a monophyletic group, represented by approximately 1650 species distributed in 73 genera. Three clades are recognized, represented in the subfamilies Agapanthoideae, Allioideae, and Amaryllidoideae. The subfamily Amaryllidoideae is made up of monocotyledonous, bulbous, and flowering species [1]. The subfamily Amaryllidoideae has increased its level of importance due to the presence of unique isoquinoline alkaloids biogenetically derived from tyrosine and phenylalanine, and known as Amaryllidaceae alkaloids [2,3]. These alkaloids are characterized by their biological effects such as neuroprotective, enzyme inhibitors of cholinesterase and butyrylcholinesterase, anti-inflammatory, and antiviral, among others [2]. Galanthamine has demonstrated a superior response in clinical terms in comparison with other Amaryllidaceae alkaloids [4], and is approved for the treatment of Alzheimer's disease [5]. On the other hand, Amaryllidaceae alkaloids have been isolated and tested also as anti-proliferative agents for different types of cancer [6,7]. The biological potential shows that Amaryllidaceae alkaloids could be used in the pharmacological field of cancer [1,8].

In Colombia, 18 genera and 48 species of the Amaryllidoideae subfamily have been reported, of which 11 genera and 27 species are native with 9 endemic species [9]. *Phaedranassa*, one of these genera, correspond to perennial herbs, up to 60 cm tall, with bulbs 2-6 cm in diameter, globose, and with cream to brown tunic. The species of the genus are distributed in tropical and premontane forests of the southern Cordillera Occidental and Nudo de los Pastos, reaching the center of Ecuador [9]. Although phytochemical analysis of *Phaedranassa* have shown three different Amaryllidaceae alkaloid types (lycorine, crinine/haemanthamine, and galanthamine), the alkaloids identified vary according to the species [10,11]. *Phaedranassa cinerea*, *Phaedranassa cuencana*, and *Phaedranassa ventricosa* are characterized by producing lycorine- and galanthamine-type alkaloids [7,10]. In *Phaedranassa brevifolia*, *Phaedranassa dubia*, *Phaedranassa glauciflora* and *Phaedranassa tunguraguae*, the presence of crinine/haemanthamine-, lycorine- and galanthamine-type alkaloids is notable [10-12]. In addition, alkaloids such as crinine-, lycorine- and narcyclasine-type, have been reported in *Phaedranassa lehmannii* [13,14]. Other less frequent alkaloid types are homolycorine and montanine, which were found in fewer species [10,12]. In this way, the production of crinine- and lycorine-type alkaloids in *Phaedranassa* species is highlighted.

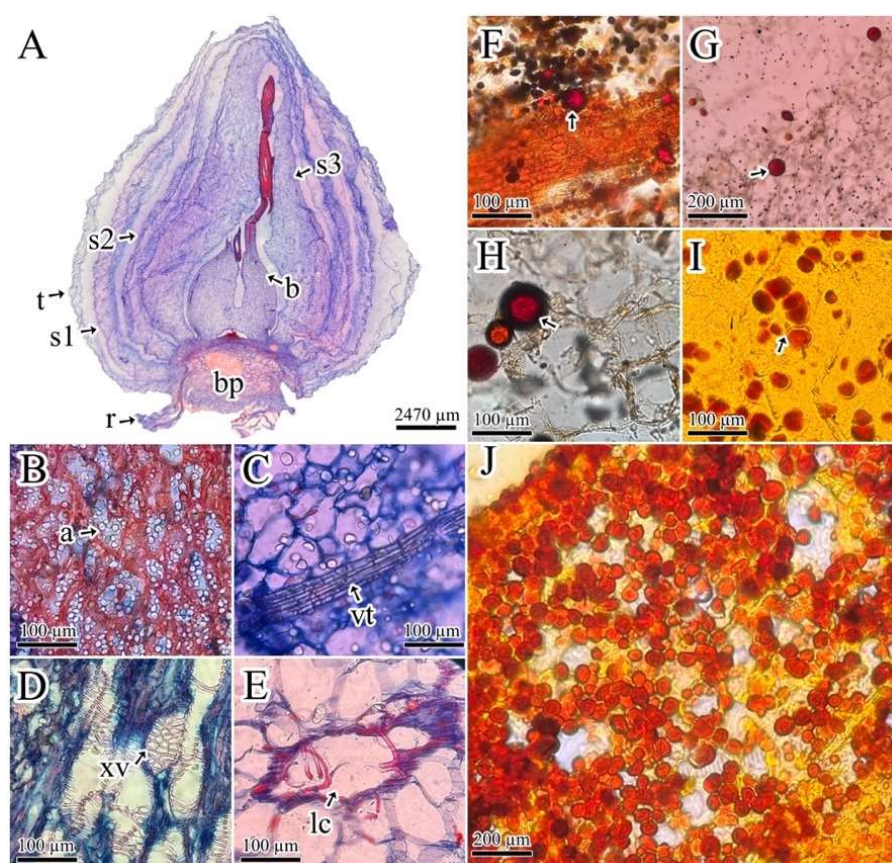
In the search for new molecules for the treatment and prevention of cancer, the vegetative organs of many Amaryllidoideae species have been analyzed as a source of numerous cytotoxic Amaryllidaceae alkaloids, particularly crinine- and lycorine-type alkaloids [15,16]. It has been suggested the correlation between the cytotoxic effect of Amaryllidaceae species and crinine- and lycorine-type alkaloids [7]. Simultaneously, the location, the composition of the chemical profile, and the distribution of alkaloids in producing organs are considered important productive features of the subfamily Amaryllidoideae, due to the quantity of compounds generally in low concentrations [2]. Histochemistry identification may yield valuable data on the producing organs and cell-type localization of the Amaryllidaceae alkaloids [17]. For this, different histochemical methods have been developed, which allow the characterization of plant structures where different compounds are distributed and accumulate, as well as the precise determination of tissues involved in the secretion of alkaloids [18]. However, studies of spatial distribution, localization, and accumulation of Amaryllidaceae alkaloids in different plant tissues (bulb, leaf, and root) are limited, especially in cytotoxic Amaryllidaceae alkaloids. In most cases, and for industrial purposes, histochemical studies have been performed on the main sources of galanthamine, and alternative sources of galanthamine for further optimization of compound production [17,19]. Therefore, this work aims to locate and characterize the alkaloids synthesized and stored by different histochemical methods and chromatographic analysis of plants grown under nursery conditions of *P. lehmannii*, a species with interesting pharmacological properties and a source of cytotoxic alkaloids [7].

## 2. Results

### 2.1. Histochemical Analysis

*P. lehmannii* bulbs develop a thin tunic and three scale-like leaves that protect the bud (Figure 1A). Scales leaves are made up of thin-walled polyhedric parenchyma cells. The scaly leaf closest to the bud develops lysogenic cavities in the apical zone (Figure 1E). In addition, the bulbs present a basal plate formed by vascular tissue (xylem vessels) and parenchyma flattened cells, which have amyloplasts storage (Figure 1B and Figure 1D), and a central bud provided by a base formed by parenchyma-like storage cells, while the bud apical zone has developing vascular tissue (Figure 1C). Dragendorff's and Wagner's reagents with reddish-brown staining, and Lugol's reagent brown, showed alkaloid storage at the basal edge of scale-like leaves and the bud apical zone with the absence of alkaloids on the basal plate (Figure 1F-J).





**Figure 1.** *P. lehmannii* bulbs anatomy. A. Anatomic details of bulbs tissues; B. Amyloplast storage in the basal plate; C. Bud developing vascular tissue; D. Xylem vessels; E. Lysogenic cavities; F-J. Detection of alkaloids in the bud and scale leaves; F-G. Wagner's reagent; H. Lugol's reagent; I-J. Dragendorff's reagent. Arrow shows alkaloid storage: a = amyloplast, b = bud, bp = basal plate, lc = lysogenic cavities, r = root, s1 = scale leaf 1, s2 = scale leaf 2, s3 = scale leaf 3, t = tunic, vt = vascular tissue, xv = xylem vessels.

## 2.2. Chromatographic Analysis of Alkaloids from *P. lehmannii*

After histochemical and phytochemical detection with different staining reagents, the alkaloids present in the bulb and root of *P. lehmannii* were analyzed by GC-MS. In the total alkaloid fraction of bulbs, eight alkaloids were detected with a predominant abundance of crinine- (9.80  $\mu\text{g/g}$  DW), followed by galanthamine- (8.04  $\mu\text{g/g}$  DW) and lycorine-type (7.38  $\mu\text{g/g}$  DW) alkaloids (Figure 2., Table S1). The alkaloids 8-O-demethylmaritidine (8.77  $\mu\text{g/g}$  DW) and lycorine (6.79  $\mu\text{g/g}$  DW) were the most abundant. Interestingly, the root has biosynthesis of nine alkaloids with high diversity and abundance. Lycorine-type alkaloids (225.29  $\mu\text{g/g}$  DW) predominate in the root, with a high prevalence of anhydrolycorine (133.7  $\mu\text{g/g}$  DW) and lycorine (73.79  $\mu\text{g/g}$  DW), being the most abundant compound in the total alkaloid fraction, followed by galanthamine-type (72.35  $\mu\text{g/g}$  DW) and narciclasine-type (13.74  $\mu\text{g/g}$  DW) alkaloids. Additionally, the results showed that the alkaloids biosynthesized by the root (324.93  $\mu\text{g/g}$  DW) exceed eleven times the abundance of the alkaloids identified in the bulbs (28.97  $\mu\text{g/g}$  DW).

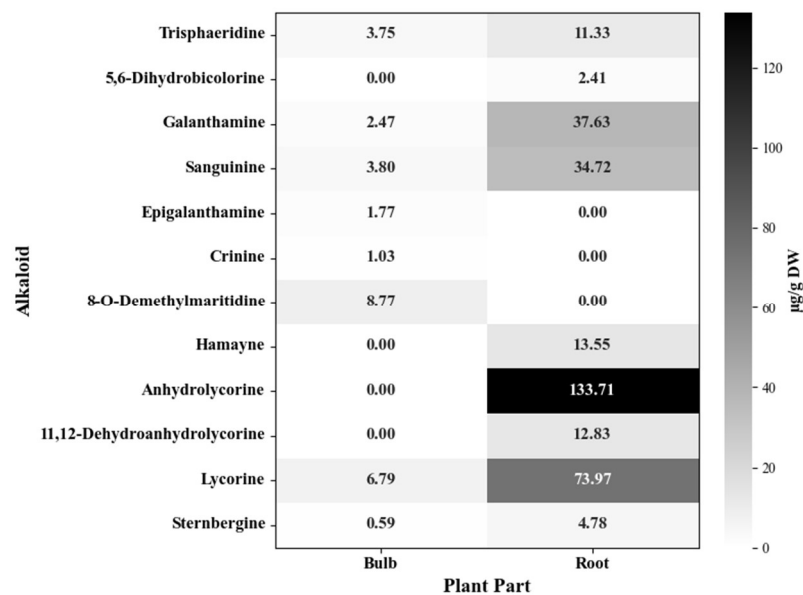
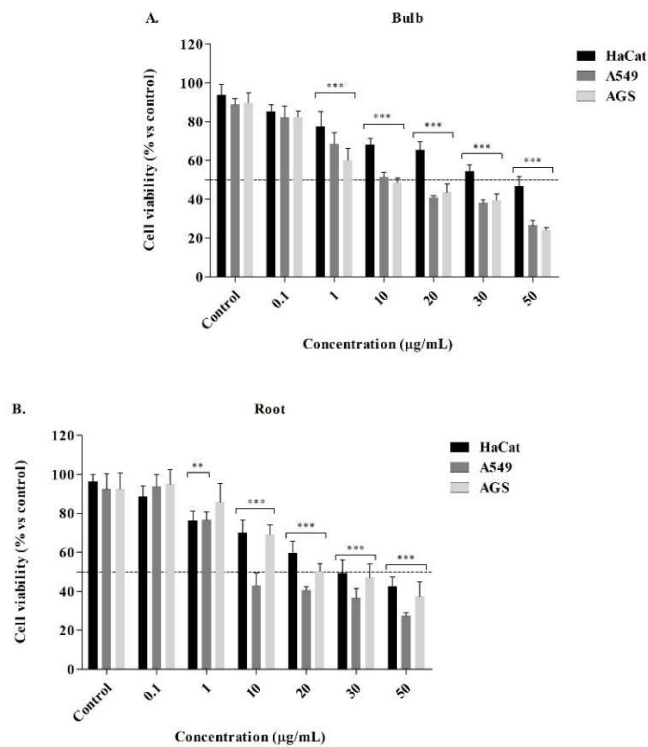


Figure 2. Alkaloids identified in bulb and root from *P. lehmannii*.

2.3. Cytotoxic Activity from *P. lehmannii* alkaloids

The alkaloids fraction of bulbs and roots showed a cytotoxic effect in the evaluated concentrations with IC<sub>50</sub> values of 11.76 ± 0.99 µg/mL and 2.59 ± 0.56 µg/mL against human lung (A549) cancer cells, and 8.00 ± 1.35 µg/mL and 18.74 ± 1.99 µg/mL against gastric (AGS) cancer cells for 48 h exposure, determined by the reduction of 3-(4,5-dimethylthiazol-2) 2,5- diphenyltetrazolium bromide (MTT) to formazan. The alkaloids showed a concentration-dependently decreased in cell viability with statistically significant differences for concentrations greater than 1 µg/mL (Figure 3A and 3B). Lycorine and doxorubicin were tested as standards (positive control) with IC<sub>50</sub> values of 4.97 ± 0.89 µg/mL and 5.52 ± 0.21 µg/mL, respectively (Table 1). Therefore, the values showed that alkaloids biosynthesized in the root generate a high cytotoxic effect on lung cancer cells, compared to lycorine. Alkaloid fractions were also evaluated on human keratinocytes (HaCat, non-cancer control cells) to evaluate the degree of selectivity of the alkaloids by determination of the selectivity index (SI, ratio of IC<sub>50</sub>). The SI values higher than 1.0 indicate that compounds or extracts have anticancer specificity. If SI values are much greater than 1.0, the samples are highly selective [20]. Based on this, the SI data shown in Table 1 indicate that alkaloids of bulbs, root, and lycorine standard exhibit a high degree of cytotoxic selectivity against lung and gastric cancer cells with values between 2.11 and 15.29.



**Figure 3.** Cell viability of alkaloid fractions of the bulb (A) and root (B) in gastric AGS and lung A549 cancer cells. Human keratinocytes HaCat were used as control cells. The results correspond to the mean  $\pm$  SEM (n=3). \*\*\*P $\leq$ 0.001 indicates statistically significant differences between control (DMSO 0.1%) and treatments according to Dunnett's Multiple Comparison Test.

**Table 1.** Cytotoxic potential of alkaloids *P. lehmannii*.

	IC <sub>50</sub> (µg/mL) $\pm$ SD <sup>1</sup>				
	A549 <sup>2</sup>	SI <sup>4</sup>	AGS <sup>2</sup>	SI <sup>4</sup>	HaCat <sup>3</sup>
Bulb	11.76 $\pm$ 0.99	4.39	8.00 $\pm$ 1.35	6.45	51.59 $\pm$ 7.98
Root	2.59 $\pm$ 0.56	15.29	18.74 $\pm$ 1.99	2.11	39.60 $\pm$ 7.86
Lycorine <sup>5</sup>	4.97 $\pm$ 0.89	7.92	4.07 $\pm$ 0.21	9.68	39.39 $\pm$ 0.94
Doxorubicin <sup>5</sup>	5.52 $\pm$ 0.21	0.81	5.84 $\pm$ 0.65	0.77	4.48 $\pm$ 0.45

<sup>1</sup> Results are the mean values standard deviations of three independent replications. <sup>2</sup> A549: lung cancer cell line and AGS: gastric cancer cell line. <sup>3</sup> HaCat: Human keratinocytes were used as control cells. <sup>4</sup> SI refers to the Selectivity Index (SI). <sup>5</sup> Positive controls.

3. Discussion

Alkaloids are molecules that plants use for chemical defense. Most plants produce alkaloids in one organ, then transport them through the xylem or phloem and store in organs such as seeds, flowers, leaves, or more specialized storage organs such as bulbs [21]. In addition, it is well known that the production of metabolites depends on conditions such as the state of development of the plant, and environmental conditions such as climate, location, and method of cultivation [22]. Specifically in the genus *Phaedranassa*, a chemical diversity in the production of Amaryllidaceae alkaloid has been demonstrated [10-14], and it could be suggested that the production is regulated by the processes of alkaloid biosynthesis in specific tissues or cells, depending largely on the level of differentiation and development, and then stored mainly in bulbs and roots, as occurs in other species of the subfamily [17]. However, few studies have examined the anatomy and histochemistry of *Phaedranassa* bulbs. In most cases, histochemical studies have been performed on the main sources of galanthamine for further optimization of the compound production [17,19].

In Amaryllidoideae species, a previous study to identify the alkaloids produced by *Rhodophiala bifida* highlights alkaloid concentrations in bulbs and roots [3]. While in *Narcissus pseudonarcissus* it has reported high alkaloid (galanthamine) storage in basal plate followed by bulb tissues [23]. Research on *Zephyranthes irwiniana* bulbs identified alkaloids in the chlorenchyma cells of the leaf and bulb margins and reported that data on the histolocalization of Amaryllidaceae alkaloids are scarce [24]. However, our results show that alkaloid storage in *P. lehmannii* occurs both in the roots and in the bulbs, in the latter, in the scale-like leaves, and in the apical zone of the buds, but not in the basal plate (Figure 1). This confirms the possible movement of alkaloids between tissues suggested by Amaryllidoideae species [25]. Nevertheless, further studies are required to characterize the transport of cytotoxic alkaloids and identify in which tissues their synthesis and storage occur. In this regard, it has been shown in *Hippeastrum papilio* that galantamine biosynthesis occurs in green tissues and is transferred to other plant organs, for example, bulbs and roots [17].

The results of the localization of alkaloids in plant tissues could provide a better understanding of the role and sites of biosynthesis and accumulation of these secondary metabolites. In addition, this contributes to the search for new sources and to the establishment of organogenesis processes aimed at the production of alkaloid-rich biomass, without endangering the biodiversity of wild populations [26,27]. It would be a strategy to increase the availability of cytotoxic alkaloids with interesting pharmacological activities such as lycorine, limited by nature in plants of the Amaryllidoideae subfamily [28,29].

In the analysis of the identification of alkaloids in bulbs and roots from *P. lehmannii*, it was established that the alkaloids detected by CG-MS would be directly correlated with the positive staining observed for alkaloids accumulated in the central part of the root, and at the basal edge of scale-like leaves and in the bud apical zone, these two last of the bulbs. The fraction of total alkaloids of bulbs corresponded to a relative abundance of 28.97  $\mu\text{g/g DW}$  with a high prevalence of crinine-, galanthamine- and lycorine-type alkaloids, with an outstanding abundance of 8-O-demethylmaritidine, galanthamine, saguinine, and lycorine (Figure 2). Complementary, the production of alkaloids in the root (324.93  $\mu\text{g/g DW}$ ) was eleven times higher than that reported for bulbs. In addition, in the root predominant the lycorine-type alkaloids, mainly anhydrolycorine and lycorine, followed by galantamine-type alkaloids, with a predominance of galantamine. This shows that biosynthesis and accumulation of specific alkaloids can be limited to specific cell types, tissues, or organs [30,31].

This effect has been observed in Amaryllidoideae species such as *Lycoris radiata*, where leaves and roots contain significantly more alkaloids of interest than in scapes and bulbs [31]. In *Leucojum aestivum*, the biosynthesis and storage of galantamine is higher in bulb (0.0949 mg/g) than in root (0.0262 mg/g), while lycorine is higher in root (0.2328 mg/g) than bulb (0.1994 mg/g) [29]. Similarly, *Galanthus nivalis* produces twenty-two times more lycorine in root (0.1816 mg/g) than in bulb (0.0080 mg/g) [29,32]. Therefore, the results found for *P. lehmannii* are related to those reported for other plants of the subfamily Amaryllidoideae regarding the abundance of alkaloids such as lycorine and galanthamine. Interestingly, these two alkaloids are widely required by the pharmaceutical industry due to insufficient availability of natural resources and the significant increase in the demand for galanthamine for the treatment of neurodegenerative diseases [28] and biological interest of lycorine for the search of therapeutic alternatives in cancer [33,34].

Although the alkaloids identified in *P. lehmannii* have been reported in previous studies [7,8,13,14], differences in the concentration of alkaloids and the presence of others previously not reported, such as sternbergine, were evidenced. This is possibly due to a change in some biological, chemical, or environmental factor that influences the biosynthesis and accumulation of secondary metabolites [35]. Concerning other species of the genus, 70% of the alkaloids identified in *P. dubia* and *P. brevifolia* correspond to lycorine- and haemanthamine/crinine-type alkaloids [12]. In *P. cinerea*, *P. cuencana*, *P. dubia*, *P. glauciflora* and *P. tunguraguae* approximately 50% of the alkaloids correspond to the lycorine-type and 15% to the crinine/hemanthamine-type alkaloids [10]. Therefore, plants of the genus *Phaedrannassa* are chemically interesting for their alkaloid profiles, and pharmacologically of interest due to the cytotoxic potential that many of these alkaloids present [12].



The alkaloid fractions from *P. lehmannii* were evaluated in gastric (AGS) and lung (A549) cancer cells using lycorine as the reference alkaloid. The alkaloid fraction of the root showed a high cytotoxic effect on lung cancer cells with  $IC_{50}$  values of  $2.59 \pm 0.56 \mu\text{g/mL}$ , compared to the alkaloid fraction of the bulbs. The antiproliferative effect against lung cancer cells could be attributed to the high presence of lycorine-type alkaloids in the root ( $225.29 \mu\text{g/g DW}$ ), every time that lycorine induces high cell death with  $IC_{50}$  of  $4.07 \pm 0.21 \mu\text{g/mL}$ , with a high degree of selectivity. It has been previously reported that lycorine-type alkaloids and lycorine have cytotoxic efficacy against many cancers cell lines, including lung cancer [34,36-38]. Lycorine is a potent inducer of apoptosis, both in mitochondrial and death receptor-mediated apoptosis. This mode of action is related to the inhibition of migration and proliferation of cells with resistance to apoptosis [36]. Although multiple apoptotic mechanisms of lycorine are possible, depending on the type of cancer, in human lung cancer cells lycorine causes a G0/G1 phase arrest, induces an early stage of apoptosis and initiates mitochondrial dysfunction [38]. Therefore, lycorine has been suggested as an excellent candidate for combating cancers [34,36], and new lycorine derivatives have been synthesized to verify the antiproliferative effects against different cancer cell lines [39].

#### 4. Materials and Methods

##### 4.1. Plant Material

The plant material of the plant grown under greenhouse conditions was obtained from the Plant Biotechnology Unit, Faculty of Engineering, Universidad Católica de Oriente. Rionegro, Colombia. One specimen of the species was deposited in the Herbarium of the University of Antioquia, Medellín, Colombia (voucher 5106). The material studied was collected with authorization from the Ministry of Environment with a genetic resource access contract #328. The selection criteria were material not in bloom and not more than 20 cm in length.

##### 4.2. Histochemical Analysis

Bulbs of *P. lehmannii* were fixated in formaldehyde, acetic acid, and ethanol (FAA) for 24 hours at  $6^{\circ}\text{C}$  [40]. After, each tissue was dehydrated in an ethanol series (50, 60, 70, 80, 90, 96 and 100%) and rinsed in HistoChoice® twice for 12 hours. Subsequently, the tissues were embedded in a solution 2:1 composed of HistoChoice® and Paraplast® paraffin for 2 hours and then in a 1:2 solution. Finally, the tissues were embedded in 100% Paraplast® paraffin for 12 hours for three times [41]. Sections were made in different cutting planes of 5 a  $7 \mu\text{m}$  thick, using a Leica RM2125 rotatory microtome. Staining was done with Safranin-Alcian Blue (Fasga) for structural differentiation of tissues [42]. Dragendorff and Wagner methods were used for the detection of alkaloids [43]. With Dragendorff reagents, the alkaloids were stained red or red-brown. In Wagner assay, the reagent gives colors similar to Dragendorff in the presence of alkaloids. Lugol's reagent, which is similar in chemical composition to Wagner reagent, produces a golden-brown reaction [43-45]. Sections were observed through an Olympus CX31 optical microscope.

##### 4.3. Extraction of Alkaloids

The treatment of plant material and extraction of alkaloids was carried out following protocols described previously [7,46]. The plants grown under greenhouse conditions were collected and washed with drinking water. After that, the bulbs were cut into pieces of 3 cm and dried to  $40^{\circ}\text{C}$  for 48 hours. The extract in methanol was made with dried and ground plant material, applying 15 min of ultrasonic baths (3 times), changing the solvent and evaporating the solvent at reduced pressure. The concentrated extract was dissolved in 10 mL 2%  $\text{H}_2\text{SO}_4$ , hexane ( $3 \times 10 \text{ mL}$ ) was used to remove neutral compounds and ammonium hydroxide was added to the aqueous fraction to adjust pH in the range of 9 to 10. Then, the alkaloids were extracted with chloroform ( $3 \times 50 \text{ mL}$ ). The organic solvent was evaporated using a rotary evaporator. To finish, for analysis in GC-MS 5mg of extract was dissolved in 500  $\mu\text{L}$  methanol.



#### 4.4. Chromatographic Analysis of Alkaloids

The analysis of alkaloids of *P. lehmannii* by GC-MS was performed according to the method described previously [47]. The alkaloid fraction was injected into an Agilent 7890 Gas Chromatograph equipped with a 5975C selective mass detector and electronic impact (EI) operating to 70 eV in splitless mode, programmed to acquire signals in scan mode between 40 and 400 daltons. In an HP-1 MS capillary column (30m x 0.25 mm x 0.25 $\mu$ m) were separated the alkaloids. It has a carrier gas flow (Helium) of 1 mL/min. The temperature ramp was as follows: 100-180°C at 15°C/minute, 180-300°C at 5°C/minute, and 10 minutes hot at 300°C. The temperature of the injector is kept at 250°C. Subsequently, 1  $\mu$ L of the sample was injected.

#### 4.5. Determination of the Alkaloid Profile

To identify alkaloids, the database "Amaryllidaceae Alkaloid Spectroteca, Agro Bio Institute (Bulgaria)" and data reported in scientific journals were used, to compare the fragments of the mass spectra of each molecule. The Kovats retention rates of the compounds were recorded with a standard calibration mixture of n-hydrocarbons (C7-C40). The percentage TIC (total ion current) was determined for each alkaloid. The codeine (50  $\mu$ g/mL) was used as an internal standard and with this, the abundance of each compound was calculated. The proportion of each compound in the extracts tested was expressed as a percentage of the total alkaloid content the area of GC-MS peaks depends on the concentration of the related compounds and the intensity of their mass spectral fragmentation.

#### 4.6. Cell Viability of Alkaloid Fraction

Cell culture: Cell viability of the total alkaloids fraction was evaluated in the gastric (AGS, CRL-1739<sup>TM</sup>) and lung (A549, CCL-185<sup>TM</sup>) cancer cell line. Human keratinocytes (HaCat, PCS-200-011<sup>TM</sup>) were used as a non-carcinogenic control cell line. The cells A549 and HaCat were grown cultured in Dulbecco Modified Eagle Medium-high glucose (DMEM) and AGS in Ham's F-12 nutrient medium (F12) supplemented with 10% heat-inactivated FBS, 100 U/mL penicillin and 100  $\mu$ g/mL streptomycin, 2 mM L-glutamine in a humidified atmosphere of 5% CO<sub>2</sub> and 95% air at 37°C. Cells were monitored on a Nikon Eclipse TS100 inverted phase contrast microscope. Cell viability experiments were performed when cells reached 75-80% confluence using 0.25% Trypsin 1 mM EDTA.

Cell viability by mitochondrial MTT reduction: Cell viability was determined using 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) assay. The total alkaloids fraction and lycorine standard were evaluated at concentrations of 50, 30, 20, 10, 1, 0.1  $\mu$ g/mL and doxorubicin at 10, 5, 2.50, 1.25, 0.62, 0.31  $\mu$ g/mL. All samples were prepared in dimethyl sulfoxide (DMSO). Cells were seeded in 96-well plates with a cell density of 2 x 10<sup>6</sup> cells/mL, incubated for 24 h at 37 °C with 5% CO<sub>2</sub>. After this time, cells were incubated with samples at 37 °C with 5% CO<sub>2</sub> for 48 h. Subsequently, 50  $\mu$ L of MTT (1.0 mg/mL buffered saline phosphate) was added. After, 4 h of incubation at 37 °C, the MTT culture medium was removed and replaced with 150  $\mu$ L DMSO to dissolve the formazan crystals. Plates were incubated in the dark with agitation for 2 h. Optical density was determined at 540 nm using a microplate reader. All experiments were conducted with three independent trials, each with six replicas.

#### 4.7. Statistical Analysis

The results are shown as the mean  $\pm$  SD. The statistical significance between the control group and the treatments with the alkaloid fraction was evaluated by a one-way analysis of variance (ANOVA) followed by the Dunnett multiple-comparison test, using the GraphPad Prism 5.0 data analysis system.

### 5. Conclusions

In summary, this study provides relevant information about the histochemical localization of alkaloids of Amaryllidaceae in the roots, and in the basal edge of scale-like leaves and bud apical

zone of *P. lehmannii*. Histochemical analysis of plants grown under nursery conditions of *P. lehmannii* showed that a possible site of alkaloid biosynthesis is the basal edge of scale-like leaves and the apical zone of buds. Where, lycorine-type alkaloids such as anhydrolycorine prevalence at root, and crinine-type alkaloids such as 8-O-demethylmaritidine in bulbs. Therefore, the roots of *P. lehmannii* represent a good source of lycorine. The bulbs constitute a significant part of the plant biomass and contain high levels of crinine-type alkaloids, not detected in the root, so they could also be considered as a valuable source of alkaloids. The cytotoxic potential against lung cancer cells of the root alkaloids was reported, mainly attributed to the presence of lycorine-type alkaloids. Consequently, the underground organs (bulb and root) of *P. lehmannii* are a source of alkaloids with cytotoxic potential, with interesting pharmacological properties in lung and gastric cancer.

**Supplementary Materials:** The following supporting information can be downloaded at: Preprints.org, **Table S1:** Amaryllidaceae alkaloids identified in *Phaedranassa lehmannii* by GC-MS.

**Author Contributions:** Conceptualization, E.O. and D.C.; methodology, L.M.T., H.L., J.M.R., I.Z.V. and M.D.; formal analysis, L.M.T. and I.Z.V.; investigation, L.M.T., H.L., J.M.R., I.Z.V. and M.D.; resources, E.O. and D.C.; writing—original draft preparation, E.O., D.C., L.T. and I.Z.V.; writing—review and editing, E.O., D.C. and L.M.T.; funding acquisition, E.O. and D.C. All authors have read and agreed to the published version of the manuscript.

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