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Reactive Oxygen Species Mechanisms That Regulate Protein-Protein Interactions in Cancer

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Abstract: Reactive oxygen species (ROS) are produced during cellular metabolism and in response to environmental stress. While low levels of ROS play essential physiological roles, excess ROS can damage cellular components, leading to cell death or transformation. ROS can also regulate protein interactions in cancer cells, thereby affecting processes such as cell growth, migration, and angiogenesis. Dysregulated interactions occur via various mechanisms, including amino acid modifications, conformational changes, and alterations in complex stability. Understanding ROS-mediated changes in protein interactions is crucial for targeted cancer therapies. In this review, we examine the role that ROS mechanisms in regulating pathways through protein-protein interactions.

Keywords: ROS; PPI; cancer; degradation; phosphorylation; kinases; phosphatase

1. Introduction

The regulation of protein-protein interactions serves as the fundamental framework governing most cellular processes [1,2]. However, these interactions are not static; they are subject to the dynamic influence of reactive oxygen species (ROS), which are key players in redox biology (Figure 1) [1,3]. ROS, generated during normal cellular metabolism and in response to environmental stressors, have a multifaceted impact on the delicate balance of these interactions [4,5]. In this context, we explore the multifaceted role of ROS in protein-protein interactions, from the modulation of protein conformation to indirect effects on kinase and phosphatase activity, stabilization of protein complexes through disulfide bond formation, and regulation of protein degradation. The nuanced interplay between ROS and protein-protein interactions provides a window into the web of cellular signaling, offering insights into various physiological processes, disease mechanisms, and potential therapeutic interventions. This exploration draws upon recent research findings that have delved into the latest advancements in redox biology, shedding light on the remarkable versatility of ROS in shaping the intricacies of cellular communication [6–9].

Protein-protein interactions are the backbone of almost all essential cellular processes [10,11]. These interactions dictate the behavior of signaling networks [12], the coordination of cellular responses [13,14], and the fine-tuning of regulatory mechanisms [15,16]. However, protein-protein interactions are far from static. One of the most influential orchestrators of this biological symphony is reactive oxygen species (ROS) [17,18]. These chemically reactive molecules, arising as byproducts of cellular metabolism and environmental influences, can signal by inducing altered interactions: For example, they modulate tetradecanoyl-phorbol-13-acetate-induced interactions of the Raf kinase inhibitory protein causing its dissociation from heat shock protein 60/mitogen-activated protein kinase (ERK and JNK) complex [19,20].

The impact of ROS on protein-protein interactions is multifaceted. ROS can alter protein conformation [21,22], indirectly affecting the activity of crucial enzymes such as kinases [23] and phosphatases [24], stabilizing protein complexes through disulfide bond formation, and even regulating protein degradation pathways [25,26]. This complex interplay between ROS and protein-

protein interactions serves as a critical nexus in the broader landscape of redox biology, offering insights into fundamental physiological processes, underlies mechanisms of disease, and has the potential for innovative therapeutic interventions [27,28]. We draw from the latest advancements in the field of redox biology, showcasing the most recent research findings and shedding light on the remarkable adaptability of ROS in sculpting the intricacies of cellular communication. With every revelation, we move one step closer to unraveling the mysteries of how ROS, those double-edged molecular swords, shape the dynamics of protein-protein interactions and, in turn, the fate of cells and organisms. In this review, we examine the pathways and implicated mechanisms in different tumor types affected by ROS. In the next paragraphs, we examine several important pathways and the mechanisms involved in modulating their PPIs in different types of tumors.

2. Pathways

2.1. The Ras-PI3K-Akt Pathway in Pancreatic Cancer

The Ras-PI3K-Akt pathway controls cellular growth and proliferation in several different cancer types, including pancreatic cancer [29,30]. ROS can disrupt this pathway by oxidizing specific cysteine residues in Ras and PI3K, which prevents their interaction and activation of downstream signaling. This leads to the inhibition of Akt signaling and subsequent cell death. Treatment with the ROS-inducing agent menadione causes a decrease in Akt phosphorylation and subsequent cell death in pancreatic cancer cells, which is mediated by the disruption of the Ras-PI3K-Akt pathway [31,32]. ROS-induced oxidation of a cysteine residue C118 in Ras prevents its interaction with PI3K and inhibits downstream Akt signaling [33]. Thus, ROS can target specific cysteine residues in Ras and PI3K proteins, which leads to the disruption of their protein-protein interactions and subsequent inhibition of the normal PI3K-Akt signaling pathway. Similarly, in PI3K, cysteine residues in the p110 α subunit, such as C862, are susceptible to ROS-induced oxidation. Oxidation of these cysteine residues disrupts the interaction of PI3K with Ras and other signaling molecules, leading to inhibition of the PI3K-Akt pathway [34].

In addition to targeting cysteine residues in Ras and PI3K, ROS can also affect the activity of upstream signaling molecules such as receptor tyrosine kinases (RTKs) [35,36]. For example, ROS can oxidize specific cysteine residues in the extracellular domain of EGFR (Figure 2), leading to activation of its tyrosine kinase activity and subsequent downstream signaling. This can lead to activation of the Ras-PI3K-Akt pathway and subsequent cancer growth [37]. Introduction of the ROS-inducing agent menadione causes a reduction in Akt phosphorylation and subsequent cell death in pancreatic cancer cells. This effect is primarily mediated by disruption of the Ras-PI3K-Akt pathway, emphasizing the key role of ROS in inhibiting this signaling cascade [38]. In Ras, cysteine residue 118 (C118) has been identified as the prime target for ROS-induced oxidation. The oxidation of C118 prevents Ras from interacting with PI3K, which in turn hampers downstream Akt signaling. Similarly, in PI3K, cysteine residues located within the p110 α subunit such as C862 are susceptible to ROS-induced oxidation. Oxidation of these cysteine residues disrupts PI3K's interaction with Ras and other vital signaling molecules, resulting in the inhibition of the PI3K-Akt pathway [39,40].

Previous studies have highlighted the significance of ROS-induced oxidation in targeting specific cysteine residues within the Ras and PI3K proteins, effectively disrupting their protein-protein interactions and consequently suppressing the PI3K-Akt signaling pathway. ROS does not limit the effects on Ras and PI3K alone. They can also influence the activity of upstream signaling molecules, including receptor tyrosine kinases (RTKs). For example, ROS oxidize specific cysteine residues in the extracellular domain of epidermal growth factor receptor (Figure 2) (EGFR). This oxidation event leads to the activation of EGFR tyrosine kinase activity and subsequent initiation of downstream signaling events. This in turn activates the Ras-PI3K-Akt pathway and promotes cancer growth (Figure 2) [41]. These findings underscore the multifaceted role of ROS in influencing the Ras-PI3K-Akt pathway in pancreatic cancer, primarily through oxidation of critical cysteine residues within key proteins. Such studies contribute significantly to our understanding of the molecular mechanisms involved in cancer development and provide potential avenues for targeted cancer

therapy. Overall, ROS-induced oxidative modifications of cysteine residues on key signal transducers can disrupt protein-protein interactions critical for the activation of the Ras-PI3K-Akt pathway, which leads to the inhibition of downstream signaling and subsequent cell death [42].

2.2. Bcl-2 Family Proteins in Breast Cancer

The Bcl-2 family of proteins plays a critical role in regulating apoptosis and the survival of cancer cells [43,44]. In breast cancer cells, ROS can disrupt protein-protein interactions between Bcl-2 and pro-apoptotic members of the Bcl-2 family, such as Bax and Bak [45]. This disruption leads to the activation of apoptosis and subsequent cell death. Treatment with the ROS-inducing agent paraquat causes an increase in the Bax/Bcl-2 ratio and subsequent apoptosis in breast cancer cells, which was mediated by the disruption of the Bcl-2/Bax interaction [46]. One example is the interaction between Bax and Bak, which are two pro-apoptotic members of the Bcl-2 family. ROS induces the formation of disulfide bonds between cysteine residues in Bax and Bak, enhancing their interaction and promoting the formation of apoptotic pores in the mitochondrial membrane. This ultimately leads to cytochrome c release and the activation of the caspase cascade, resulting in apoptosis [47]. The reversible formation of disulfide bonds between interacting proteins is a target of ROS, and in the future, the identification of more instances may provide avenues for the targeted protection of such interactions. Disulfide bond formation, which has the potential to mediate extensive yet fully reversible structural and functional changes, rapidly adjusting the activity of the protein to prevailing oxidant levels, is a frequent target of ROS [44,48].

ROS can also affect the activity of anti-apoptotic members of the Bcl-2 family, such as Bcl-2 and Mcl-1. ROS can oxidize cysteine residues in Bcl-2 and Mcl-1, leading to changes in their conformation and activity [49,50]. These changes can alter the balance between pro- and anti-apoptotic Bcl-2 family members and contribute to the development and progression of cancer. Overall, ROS-induced oxidative modifications can affect the interactions between members of the Bcl-2 family, lead to changes in apoptotic signaling and cell survival. Dysregulation of the Bcl-2 family is a hallmark of many types of cancer, and understanding the role of ROS in regulating these interactions could provide insights into the development of novel cancer therapeutics. In the greater context, these ROS-induced oxidative modifications significantly affect interactions within the Bcl-2 family. Dysregulation of the Bcl-2 family is a hallmark of numerous cancer types. Delving into the role of ROS in this regulation presents an exciting avenue for the development of novel and targeted cancer therapeutics.

2.3. E-cadherin and β -catenin in Colorectal Cancer

ROS can also regulate the activity of the Wnt/β-catenin/E-cadherin pathway, which plays a critical role in cell proliferation, differentiation, and apoptosis [51,52]. E-cadherin is a critical cell adhesion molecule that plays a key role in maintaining epithelial tissue integrity. In colorectal cancer, elevated ROS levels can disrupt the protein-protein interactions between E-cadherin and β-catenin, leading to the activation of Wnt/β-catenin signaling and subsequent tumor growth [53]. Treatment with the ROS-inducing agent H2O2 causes a decrease in E-cadherin/β-catenin interaction and subsequent activation of Wnt/β-catenin signaling in colorectal cancer cells [54]. ROS can induce the oxidation of cysteine residues in E-cadherin, which leads to disruption of its interactions with β catenin and subsequent destabilization of adherens junctions [55]. This results in increased cell motility and invasion, which can contribute to the metastasis of cancer cells. Additionally, ROS can oxidize the cysteine residues in β -catenin, which affects its activity and stability. Oxidation of β catenin leads to its phosphorylation and subsequent degradation, which can result in decreased levels of β-catenin and subsequent changes in downstream signaling pathways [56]. ROS can induce the oxidation of specific cysteine residues in key components of this pathway, such as Dishevelled and Axin, which can lead to changes in downstream signaling and subsequent cancer growth [57,58]. Overall, ROS-induced oxidative modifications can affect the interactions between E-cadherin and βcatenin, leading to changes in cell adhesion and motility, which can contribute to cancer cell metastasis [59].

In colorectal cancer, elevated ROS levels have been observed to disrupt protein-protein interactions between E-cadherin and β -catenin. This disruption has far-reaching consequences, leading to the activation of the Wnt/ β -catenin signaling pathway, which, in turn, fuels tumor growth [60,61]. Notably, the introduction of the ROS-inducing agent H2O2 has been shown to decrease the interaction between E-cadherin and β -catenin, subsequently activating Wnt/ β -catenin signaling in colorectal cancer cells [62]. This study exemplifies the critical role of ROS in modulating these interactions within cancer contexts. The destabilization of the E-cadherin/ β -catenin interaction enhances cell motility and invasiveness, contributing to the metastatic potential of cancer cells [63]. Oxidation of β -catenin can lead to phosphorylation and degradation, resulting in reduced β -catenin levels and consequential alterations in downstream signaling pathways [64]. In conclusion, the impact of ROS-induced oxidative modifications on the interactions between E-cadherin and β -catenin has profound implications for cell adhesion, motility, metastasis, and the development of colorectal cancer as well as other types of cancer.

2.4. HIF-1 α and p300

The transcription factor HIF-1 α plays a critical role in regulating cellular responses to hypoxia in cancer cells. In glioblastoma cells, elevated ROS levels can disrupt the protein-protein interactions between HIF-1 α and the transcriptional co-activator p300, which prevents the activation of HIF-1 α target genes and subsequent tumor growth [65-67]. The ROS-inducing agent menadione causes a decrease in HIF-1 α target gene expression and subsequent cell death in glioblastoma cells, which was mediated by the disruption of the HIF- 1α /p300 interaction. Another study showed that ROS-induced oxidation of specific cysteine residues in p300 disrupted its interaction with HIF-1 α and inhibited its transcriptional activity [68,69]. ROS can affect the activity of HIF-1α by inducing the oxidation of specific cysteine residues in its oxygen-dependent degradation domain (ODD), which stabilizes the protein and enhances its transcriptional activity. ROS can affect the activity of p300 by inducing the oxidation of specific cysteine residues in its histone acetyltransferase (HAT) domain, which affects its ability to acetylate histones and activate transcription. Additionally, ROS can also affect the interactions between p300 and other transcription factors, such as p53 and NF-κB, which can alter downstream signaling pathways and subsequent cancer growth. p300 is a multifunctional coactivator protein that interacts with a wide range of transcription factors to activate gene expression. One of the key transcription factors that interacts with p300 is p53, which is a tumor suppressor protein that plays a critical role in preventing cancer by inducing cell cycle arrest, DNA repair, and apoptosis in response to cellular stress. ROS can modulate the activity of p53 by inducing the oxidation of specific cysteine residues within its DNA-binding domain, which alters its ability to bind to DNA and activate transcription. The interaction between p300 and p53 is critical for the regulation of p53-mediated transcriptional activation. P300 acts as a coactivator of p53 by acetylating histones at the promoter regions of p53 target genes, which leads to increased expression of these genes. ROS-induced oxidation of p300 can affect its ability to acetylate histones and activate transcription, which in turn affects the ability of p53 to induce its target genes [70,71].

The transcription factor HIF- 1α , a key regulator of cellular responses to hypoxia [72], finds its activity modulated by ROS-induced disruption in its protein-protein interactions with the transcriptional co-activator p300. This disturbance ultimately prevents the activation of HIF- 1α target genes, thereby inhibiting tumor growth. A study employing the ROS-inducing agent menadione revealed a decrease in HIF- 1α target gene expression, leading to glioblastoma cell death, all mediated by the disruption of the HIF- 1α /p300 interaction. Another investigation uncovered how ROS-induced oxidation of specific cysteine residues in p300 disrupted its interaction with HIF- 1α , impeding its transcriptional activity. Additionally, p300, a versatile coactivator protein, forms interactions not only with HIF- 1α but also with other transcription factors such as p53 and NF- κ B. ROS-induced oxidation of p300, particularly within its histone acetyltransferase (HAT) domain, impacts its ability to acetylate histones and activate transcription. This interference extends to the interactions between p300 and other transcription factors, ultimately resulting in alterations to downstream signaling pathways and influencing cancer growth.

P53, a critical tumor suppressor protein, plays a fundamental role in preventing cancer by inducing cell cycle arrest, DNA repair, and apoptosis in response to cellular stress such as DNA damage. ROS, however, can modulate p53's activity by oxidizing specific cysteine residues within its DNA-binding domain. This oxidative event alters p53's DNA binding capacity and its ability to activate transcription. Concurrently, ROS-induced oxidation of p300 can also affect its capacity to acetylate histones and activate transcription [73,74]. The interplay between p300 and p53 holds particular significance in the regulation of p53-mediated transcriptional activation. P300 acts as a coactivator for p53 by acetylating histones at the promoter regions of p53 target genes, resulting in increased gene expression. The impact of ROS-induced oxidation of p300 extends to this critical interaction, affecting p300's ability to acetylate histones and, consequently, influencing p53's capacity to induce its target genes. Dysregulation of the p300/p53 pathway has been implicated in the development and progression of various cancer types. Delving into the role of ROS in regulating these interactions provides valuable insights into the mechanisms governing cancer growth and opens new avenues for potential therapeutic targets in the battle against cancer.

2.5. Effects of ROS on Protein-Protein Interactions in Pathways of Brain and Lung Cancers

Reactive oxygen species (ROS) are critical regulators of protein-protein interactions (PPIs) within the signaling pathways of brain and lung cancers. These interactions significantly influence cancer progression, metastasis, and response to therapy. In brain and lung cancers, ROS can modify proteins through redox reactions, which particularly affect cysteine residues. These modifications can alter the structure and function of proteins, impacting PPIs that are essential for signaling pathways involved in cancer progression. For example, ROS can modify the KEAP1 protein, leading to the stabilization of NRF2, which is crucial for cellular defense mechanisms against oxidative stress in lung cancer [75,76]. ROS influence several signaling pathways by modulating PPIs. In brain cancers, as in other types of tumors, ROS can affect interactions within the PI3K/Akt pathway, which is vital for cell survival and proliferation. Alterations in PPIs within this pathway can lead to enhanced cancer cell growth and resistance to apoptosis. Similarly, in lung cancer, ROS can modify PPIs involved in the epithelial-mesenchymal transition (EMT), a process that facilitates metastasis. By altering interactions between proteins such as E-cadherin and vimentin, ROS promote the invasive capabilities of cancer cells [77]. Moreover, ROS can impact PPIs in pathways related to angiogenesis and invasion in brain cancers. For instance, ROS can modulate interactions between vascular endothelial growth factor (VEGF) and its receptors, promoting angiogenesis and contributing to the aggressive nature of these tumors [78]. ROS act as signaling molecules that can either promote or inhibit cancer progression depending on their concentration. At moderate levels, ROS activate signaling cascades involving mitogen-activated protein kinase (MAPK), extracellular signalregulated kinase (ERK), and nuclear factor kappa-light-chain-enhancer of activated B cells (NF-κB), which promote cancer cell survival and proliferation. At higher concentrations, ROS can induce apoptosis through intrinsic and extrinsic pathways, involving mitochondrial damage and death receptor activation, respectively [76,79].

2.6. Interaction between Reactive Oxygen Species (ROS) and Lipids: Implications for Protein-Protein Interactions

Reactive oxygen species (ROS) are pivotal in the oxidation of lipids, a process known as lipid peroxidation, which significantly influences protein-protein interactions (PPIs) in cells. This process is especially relevant in the context of cancer, where lipid peroxidation products can alter signaling pathways and contribute to oncogenic processes. Lipid peroxidation is initiated when ROS, such as hydroxyl radicals, attack polyunsaturated fatty acids (PUFAs) in cell membranes. This reaction results in the formation of lipid radicals, which propagate chain reactions leading to the production of lipid peroxides. These lipid peroxides can decompose into reactive aldehydes, such as 4-hydroxynonenal (4-HNE), malondialdehyde (MDA), and acrolein, which are capable of forming adducts with proteins, nucleic acids, and other cellular components [80,81]. Among the lipid

peroxidation products, 4-HNE is particularly significant due to its high reactivity. 4-HNE can form covalent adducts with cysteine, histidine, and lysine residues on proteins, thereby altering their structure and function. This modification can disrupt PPIs and affect signaling pathways involved in cell proliferation, apoptosis, and stress responses. For example, 4-HNE can modify proteins involved in the NF-κB signaling pathway, leading to altered transcriptional activity and promoting oncogenic signaling [82].

The formation of 4-HNE-protein adducts can have profound effects on PPIs. These modifications can either inhibit or enhance protein interactions, depending on the specific proteins and pathways involved. For instance, in cancer cells, 4-HNE has been shown to modify key signaling proteins, thereby influencing pathways such as MAPK and PI3K/Akt, which are crucial for cell survival and proliferation. The role of lipid peroxidation in modulating PPIs and signaling pathways highlights its potential as a therapeutic target. By understanding the specific modifications induced by lipid peroxidation products like 4-HNE, novel therapeutic strategies can be developed to inhibit these interactions and restore normal signaling. Antioxidants and inhibitors of lipid peroxidation are being explored as potential therapies to mitigate the effects of ROS-induced lipid damage in cancer. The formation of reactive aldehydes such as 4-HNE and their ability to form adducts with proteins underscore the complexity of ROS interactions in cancer biology. Understanding these mechanisms offers valuable insights for developing targeted therapies aimed at disrupting these pathological interactions.

4-Hydroxynonenal (4-HNE) is a highly reactive aldehyde formed during lipid peroxidation, and it plays a significant role in altering protein-protein interactions (PPIs) through its ability to form covalent adducts with proteins. These modifications can profoundly affect cellular signaling pathways and contribute to oncogenic processes.

2.7. Mechanisms of 4-HNE Interaction with Proteins

4-HNE primarily interacts with proteins by forming covalent adducts with nucleophilic amino acid residues such as cysteine, histidine, lysine, and arginine. This interaction can occur through Michael addition or the formation of Schiff bases, leading to changes in protein structure and function [83,84]. For instance, 4-HNE can modify the catalytic sites of enzymes or bind to residues outside the active site, thereby altering enzyme activity and protein interactions [85]. The formation of 4-HNE-protein adducts can disrupt PPIs by altering the structural conformation of proteins. This disruption can lead to changes in signaling pathways that regulate cell proliferation, apoptosis, and stress responses. For example, 4-HNE can modify proteins involved in the NF-κB signaling pathway, leading to altered transcriptional activity and promoting inflammation and oncogenic signaling. In addition to its role in signaling pathways, 4-HNE can influence the dynamics of PPIs by modifying proteins such as tubulins, which are essential for microtubule formation. 4-HNE adduction to tubulin can impair its polymerization, leading to microtubule disorganization and affecting cellular processes such as mitosis and intracellular transport.

2.8. Biological Implications and Oncogenic Potential of 4-HNE

The ability of 4-HNE to form adducts with proteins makes it a potent modulator of cellular processes. These modifications can lead to the activation of stress-response pathways and contribute to the development and progression of cancer. For instance, 4-HNE has been shown to modulate ligand-independent signaling by membrane receptors such as EGFR, acting as a sensor of external stress and initiating signaling cascades that affect cell growth and survival [86]. Given the significant impact of 4-HNE on PPIs and signaling pathways, targeting 4-HNE-protein interactions presents a potential therapeutic strategy. Understanding the specific proteins and pathways affected by 4-HNE can lead to the development of interventions aimed at mitigating its oncogenic effects. Antioxidants and compounds that can detoxify 4-HNE or prevent its formation are being explored as potential therapies to counteract the deleterious effects of lipid peroxidation products. In summary, 4-HNE plays a crucial role in altering protein-protein interactions, thereby influencing cellular signaling and contributing to oncogenic processes. Its ability to modify proteins and disrupt PPIs highlights the

complexity of its role in cancer biology and underscores the potential for therapeutic interventions targeting 4-HNE-mediated modifications.

2.9. Therapeutic Implications

Understanding the specific PPIs affected by ROS in brain and lung cancers provides opportunities for therapeutic interventions. Targeting these interactions can disrupt critical signaling pathways, potentially inhibiting cancer progression. For example, therapies that modulate ROS levels can either increase ROS to induce cancer cell death or decrease ROS to prevent tumor progression, depending on the cancer type and stage. Antioxidants may be used to reduce ROS levels, thereby inhibiting pro-tumorigenic signaling pathways. However, caution is necessary as antioxidants can also protect cancer cells from ROS-induced apoptosis. Alternatively, therapies that specifically target ROS-sensitive PPIs could provide a more effective approach, especially when combined with other treatments that manipulate ROS levels. In summary, ROS significantly impact protein-protein interactions in pathways critical to brain and lung cancers. By understanding these interactions, researchers can develop targeted therapies that exploit the dual nature of ROS, offering potential advancements in cancer treatment [77,79].

3. Mechanisms

3.1. Direct ROS-Mediated Mechanisms

Reactive oxygen species (ROS) can modulate protein complex interactions in cancer cells through multiple mechanisms [28]. ROS can affect protein conformation: they oxidize amino acid residues in proteins, leading to changes in protein structure and function. These changes can alter the conformation of proteins that are part of a protein complex and as a consequence alter the interactions between these proteins [35,87]. The first mechanism involves changes in protein conformation: For example, ROS-mediated oxidation of the transcription factor NF-κB can lead to a conformational change that disrupts its interaction with its inhibitor, IκB, thereby activating NF-κB target genes [88]. In its inactive state, NF-κB is held in check by its inhibitor, IκB, which prevents NF-κB from translocating to the cell nucleus where it would initiate gene transcription. However, the introduction of ROS can lead to the oxidation of specific amino acid residues within NF-κB. While cysteine is the primary target for oxidation in NF-κB and its regulatory proteins, it's worth noting that ROS can potentially oxidize other amino acids, such as arginine, lysine, proline, threonine, tryptophan, phenylalanine, and histidine. Also, ROS can directly oxidize IkB at cysteine sites, inducing the nuclear translocation of NF-kB. This oxidative event initiates a conformational change in the transcription factor, disrupting its interaction with IkB. On the other hand, the IKK complex, which regulates NFκB activation, is also a target for ROS-mediated oxidation. Specifically: H₂O₂ can inactivate IKK by oxidizing cysteine 179 of IKKβ, leading to S-glutathionylation and reduced NF-κB signaling. Conversely, H₂O₂ can activate IKK in some cell types by forming disulfide bonds between Cys54 and Cys347 of the IKKγ/NEMO subunit

This disruption frees NF-κB from its inhibitory partner, allowing it to translocate into the nucleus and activate its target genes, which are involved in a plethora of cellular responses, making NF-κB a central regulator of processes like inflammation and immune reactions [89]. Reactive oxygen species (ROS), such as superoxide, hydrogen peroxide, and hydroxyl radicals, have the remarkable ability to influence the conformation and activity of proteins within cells [90], with several proteins acting as redox sensors. This dynamic interplay between ROS and protein conformation hinges on the oxidative modification of specific amino acid residues within these proteins, ultimately culminating in structural changes that can profoundly impact their functions [91]. The NF-κΒ/ΙκΒ interaction exemplifies this interplay between ROS and protein conformation, underscoring the role of ROS as signaling molecules that can alter vital cellular responses.

Another notable example is the FOXO transcription factor family, important regulators of the cellular stress response and cellular antioxidant defense [92]. This family of proteins is regulated by ROS as well as other stressful stimuli that elicit the formation of ROS, by modulating FoxO activity

at multiple levels, including posttranslational modifications of FOXOs (such as phosphorylation and acetylation) [93], interaction with coregulators [94], alterations in FoxO subcellular localization [95], protein synthesis and stability. ROS-induced post-translational modifications play a significant role, including phosphorylation by activated c-Jun N-terminal kinase (JNK), which promotes FOXO nuclear localization and transcriptional activity. Oxidative stress also leads to acetylation of FOXO proteins via activation of p300/CREB-binding protein, and can induce monoubiquitination, affecting FOXO activity. FOXO proteins engage in redox-sensitive interactions with co-regulators and antioxidant peroxiredoxins. Increased intracellular ROS facilitates FOXO localization to the nucleus, where it is transcriptionally active. ROS can also influence FOXO protein stability and there is evidence for redox-sensitive regulation of FOXO gene expression. While the search results don't specify exact amino acids modified in FOXO proteins by ROS, cysteine residues are often targets for oxidation due to their reactive thiol groups. The specific amino acids modified and the exact mechanisms can vary depending on the particular FOXO protein (FOXO1, FOXO3a, FOXO4, or FOXO6) and the cellular context. These diverse mechanisms collectively contribute to the complex regulation of FOXO transcription factors by ROS, influencing their activity and function in cellular processes [96].

3.2. Indirect ROS-Mediated Mechanisms

Indirect mechanisms involve ROS affecting protein-protein interactions through the modulation of protein kinases and phosphatases. Changes in protein phosphorylation can affect protein conformation and alter protein-protein interactions. For example, ROS-mediated activation of the protein kinase JNK can lead to the phosphorylation of the transcription factor c-Jun, altering its conformation and enhancing its interaction with other proteins such as ATF-6, p38, and ERKs. The phosphorylation of c-Jun by JNK creates a sophisticated regulatory mechanism without inducing large-scale conformational changes. As an intrinsically disordered protein, c-Jun's regulation occurs through a precise temporal pattern of phosphorylation at four specific residues within its transactivation domain. This sequential modification generates three distinct functional states of c-Jun, each with unique interaction profiles and cellular roles. The unphosphorylated state recruits the MBD3 repressor, while the doubly-phosphorylated state (at serines 63 and 73) favors binding to the TCF4 co-activator. The fully phosphorylated state, involving all four sites, attenuates JNK signaling by disfavoring TCF4 binding [97]. These phosphorylation-dependent states modulate c-Jun's ability to form complexes with various partners, including components of the AP-1 transcription factor. Additionally, phosphorylation enhances c-Jun's stability by reducing its susceptibility to proteasomal degradation. This intricate system allows c-Jun to encode multiple functional states in response to JNK activation, enabling a nuanced and complex signaling response from a single kinase input [98,99].

Kinases and phosphatases play a pivotal role in protein phosphorylation, a post-translational modification, significantly influencing protein conformation and, consequently, protein-protein interactions [100,101]. This reversible modification serves as a molecular switch regulating various cellular processes. The alteration in the phosphorylation state of a protein can induce changes in its conformation, thereby affecting its interactions with other proteins. One illustrative example of ROSmediated modulation through kinases involves the protein kinase JNK. ROS can stimulate JNK activity, leading to the phosphorylation of specific target proteins. In particular, JNK activation can result in the phosphorylation of the transcription factor c-Jun. This phosphorylation event causes a conformational change in c-Jun, rendering it more receptive to interactions with other proteins. In this case, c-Jun's altered conformation enhances its affinity for another transcription factor, ATF-2 (Activating Transcription Factor 2). The strengthened interaction between c-Jun and ATF-2 can subsequently influence gene expression patterns, impacting cellular responses. In essence, ROS indirectly affect protein-protein interactions by triggering a cascade of events that ultimately lead to changes in protein phosphorylation. The ramifications of these modifications can extend beyond individual proteins and influence the complex network of interactions within the cell. the mechanism of c-Jun phosphorylation and its effects on protein interactions are more nuanced than a simple conformational change. JNK phosphorylates c-Jun at four specific residues within its transactivation domain (TAD) in a precise temporal pattern. Serine 63 and serine 73 are phosphorylated more rapidly than threonine 91 and threonine 93. This sequential phosphorylation creates three distinct functional states of c-Jun, each with unique interaction profiles. The unphosphorylated c-Jun recruits the MBD3 repressor, maintaining a transcriptionally repressed state. When serine 63 and serine 73 are phosphorylated, c-Jun transitions to an active state where the MBD3 repressor is displaced, allowing binding of the transcriptional activator TCF4. This doubly-phosphorylated state triggers c-Jun's transcriptional activity. Subsequently, the slower phosphorylation of threonine 91 and threonine 93 leads to a fully phosphorylated state that disfavors TCF4 binding, attenuating JNK signaling and returning c-Jun to a transcriptionally inactive state. These phosphorylation-dependent states modulate c-Jun's ability to form complexes with various partners, including components of the AP-1 transcription factor such as JunB, JunD, c-Fos, and ATF. Additionally, phosphorylation enhances c-Jun's stability by reducing its susceptibility to proteasomal degradation, leading to its accumulation. This intricate system allows c-Jun to encode multiple functional states in response to JNK activation, enabling a nuanced and complex signaling response from a single kinase input. It's important to note that c-Jun is an intrinsically disordered protein, and the phosphorylation events do not induce largescale conformational changes in the traditional sense. Instead, they create local changes in charge and structure that significantly alter c-Jun's functional interactions and cellular behavior. This mechanism provides a sophisticated regulatory system that fine-tunes c-Jun's activity in response to ROSmediated JNK activation, allowing for precise control of cellular responses to oxidative stress.

3.3. ROS-Mediated Protein Complex Stability

Another mechanism involves protein complex stability: ROS can affect protein complex stability by oxidizing cysteine residues in proteins, forming disulfide bonds that stabilize them. For instance, the formation of disulfide bonds between subunits of the transcription factor AP-1 can enhance its stability and promote the activation of target genes. Reactive oxygen species (ROS) play a crucial role in regulating the stability and activity of protein complexes, particularly transcription factors like AP-1, through the oxidation of cysteine residues. This mechanism is exemplified in the redox-sensitive regulation of the AP-1 complex, a dimeric transcription factor composed of proteins from the Jun, Fos, and ATF families. The c-Jun subunit of AP-1 contains three cysteine residues (Cys-269, Cys-320, and Cys-325) in its DNA-binding domain that are susceptible to oxidation. Under oxidative conditions, these cysteines can form intramolecular or intermolecular disulfide bonds, with Cys-269 capable of bonding with either Cys-320 or Cys-325. This oxidation-induced disulfide bond formation significantly enhances c-Jun's DNA-binding activity by increasing its affinity for target DNA sequences. Moreover, oxidation promotes the formation of c-Jun homodimers or heterodimers with other AP-1 components like c-Fos, further stabilizing the AP-1 complex. The c-Fos subunit also contains a redox-sensitive region in its DNA-binding domain, where cysteine oxidation enhances its DNA-binding activity and ability to form stable dimers with c-Jun. Importantly, this process is reversible, allowing for dynamic regulation of AP-1 activity in response to changing cellular redox conditions. Reducing agents can break these disulfide bonds, potentially decreasing AP-1 activity. This redox regulation mechanism provides cells with a rapid and reversible means to respond to oxidative stress, enabling quick activation of AP-1-dependent genes involved in various cellular processes, including stress response, proliferation, and apoptosis. However, it's crucial to note that while oxidation can enhance AP-1 activity, excessive oxidation may lead to protein dysfunction, underscoring the importance of maintaining a delicate balance between oxidation and reduction for optimal AP-1 function and cellular homeostasis [102,103].

3.4. ROS-Mediated Protein Degradation

A third mechanism involves protein degradation: Reactive oxygen species (ROS) impact the regulation of protein degradation [104]. One key player in cellular protein degradation is the proteasome, responsible for breaking down intracellular proteins, some of which are components of essential protein complexes. ROS-mediated modulation of proteasome activity can lead to alterations

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in the stability and functionality of these complexes, presenting a critical aspect of redox biology [105]. The proteasome's primary role is to maintain cellular homeostasis by degrading proteins tagged for disposal. This process involves the recognition and subsequent degradation of proteins, including those participating in various protein complexes. ROS can affect proteasome activity through various mechanisms, including direct oxidative modifications. When the proteasome's activity is perturbed, changes in protein turnover can impact complex stability and function. Ongoing research in redox biology explores ROS-mediated proteasome modulation, encompassing specific mechanisms involved, downstream consequences, and implications for various cellular processes and disease states. Overall, ROS can modulate protein complex interactions in cancer cells through multiple mechanisms, including changes in protein conformation, protein-protein interactions, protein complex stability, and protein degradation. These changes can alter cellular signaling and gene expression, contributing to cancer development and progression.

4. Therapeutic Approaches Targeting ROS Levels and Protein-Protein Interactions in Cancer

4.1. Current Use of Antioxidant Medications and Supplements in Cancer Therapy

Antioxidants have long been considered as potential adjuncts in cancer therapy due to their ability to neutralize reactive oxygen species (ROS) and protect cells from oxidative stress. Common antioxidants include vitamin C, vitamin E, selenium, and glutathione, which are either administered as dietary supplements or as part of therapeutic regimens. The rationale behind using antioxidants in cancer therapy lies in their potential to reduce the oxidative damage caused by elevated ROS levels, which is a hallmark of many cancers. However, the use of antioxidants in cancer treatment is complex and controversial. On one hand, antioxidants might protect normal cells from the oxidative damage induced by chemotherapy and radiation therapy, reducing side effects. On the other hand, they could also protect cancer cells, potentially reducing the efficacy of ROS-generating treatments and promoting tumor survival. Some studies suggest that antioxidants might interfere with the prooxidant effects of certain chemotherapies, which rely on ROS to kill cancer cells, thereby leading to reduced treatment efficacy [106].

Moreover, recent research has shown that ROS also play a role in disrupting protein-protein interactions (PPIs) within cancer cells. By oxidizing specific amino acid residues, ROS can alter the structure and function of proteins, leading to the formation or disruption of protein complexes that are crucial for cancer cell survival and proliferation. Therefore, while antioxidants might protect normal tissues, they could also inadvertently stabilize harmful PPIs in cancer cells, further complicating their therapeutic use [107].

4.2. Variation of ROS Levels across Different Stages of Cancer

ROS levels are not static and can vary significantly across different stages of cancer development and progression. In early stages, cancer cells often exhibit moderate increases in ROS, which can drive mutations and promote oncogenic signaling pathways. As the tumor progresses, metabolic reprogramming and mitochondrial dysfunction lead to further increases in ROS levels, contributing to more aggressive phenotypes and increased resistance to therapy. In advanced stages, cancer cells often adapt by upregulating antioxidant defenses to survive in the high-ROS environment. This adaptation is critical for maintaining the balance between ROS-driven proliferation and avoiding ROS-induced cell death. The variation in ROS levels across cancer stages highlights the potential for stage-specific therapeutic strategies. For example, in early-stage cancers, enhancing ROS levels might promote cancer cell death, while in late-stage cancers, targeting the antioxidant systems that allow cancer cells to thrive in high-ROS conditions might be more effective. The fluctuation of ROS levels also affects PPIs. In early stages, ROS-induced modifications might disrupt critical tumor suppressor interactions, while in advanced stages, the stabilization of oncogenic PPIs by ROS could promote cancer progression. This dynamic suggests that targeting the ROS-PPI axis could be a promising therapeutic strategy, with different approaches needed depending on the stage of the cancer [108,109].

4.3. The Role of Genetic Modifiers in ROS Sensitivity and Development of ROS-Targeting Therapeutic Agents

Genetic factors play a crucial role in determining a cancer cell's sensitivity to ROS. Variations in genes involved in ROS production, detoxification, and repair of ROS-induced damage can significantly influence how cancer cells respond to oxidative stress. For instance, mutations in genes like TP53, which encodes the tumor suppressor protein p53, can impair the cell's ability to respond to ROS-induced DNA damage, leading to increased mutation rates and cancer progression [110]. The understanding of genetic modifiers has paved the way for the development of ROS-targeting therapeutic agents. These agents aim to exploit the differential ROS sensitivity between normal and cancer cells. For example, drugs that inhibit antioxidant defenses in cancer cells can selectively increase ROS to lethal levels, while sparing normal cells that have lower ROS levels. Additionally, targeting specific PPIs that are stabilized by ROS in cancer cells offers another avenue for therapy. By disrupting these PPIs, it may be possible to selectively kill cancer cells without harming normal tissue. These approaches highlight the potential of integrating genetic information with ROS-targeting strategies to develop personalized cancer therapies. By understanding the genetic landscape of a tumor, clinicians can better predict how it will respond to ROS modulation, thereby improving the efficacy of treatments [111–113].

Conclusions

The balance between the generation and elimination of reactive oxygen species (ROS) represents a critical determinant of cell fate and function. While ROS are vital participants in various physiological processes, their excessive accumulation can lead to detrimental damage to cellular components, ultimately resulting in cell death or malignant transformation. Importantly, ROSmediated alterations in protein interactions have emerged as central players in the progression of cancer, impacting key cellular processes such as proliferation, migration, and angiogenesis. These perturbed protein interactions occur through multiple mechanisms, including modifications of amino acid residues, such as cysteine, conformational changes, and disruption of complex stability. A deeper understanding of how ROS affect protein interactions provides valuable insights into the web of signaling pathways that regulate cancer progression. Recognizing the critical role of ROS in modulating these pathways and their underlying mechanisms is of paramount importance for the development of targeted cancer therapies. This review has endeavored to shed light on the multifaceted role of reactive oxygen species in shaping cellular pathways and interactions, with a particular focus on their implications in cancer. By further dissecting the interplay between ROS and protein interactions, will contribute to ongoing efforts to harness this knowledge for the development of innovative and precise therapeutic strategies in the fight against cancer.

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References

- Slater, O.; Miller, B.; Kontoyianni, M. Decoding Protein-protein Interactions: An Overview. Curr Top Med Chem 2020, 20, 855–882, doi:10.2174/1568026620666200226105312.
- Rabbani, G.; Baig, M.H.; Ahmad, K.; Choi, I. Protein-protein Interactions and their Role in Various Diseases and their Prediction Techniques. Curr Protein Pept Sci 2018, 19, 948–957, doi:10.2174/1389203718666170828122927.
- 3. Zhang, J.; Wang, X.; Vikash, V.; Ye, Q.; Wu, D.; Liu, Y.; Dong, W. ROS and ROS-Mediated Cellular Signaling. *Oxid Med Cell Longev* **2016**, *2016*, 4350965, doi:10.1155/2016/4350965.
- Zhou, R.; Tardivel, A.; Thorens, B.; Choi, I.; Tschopp, J. Thioredoxin-interacting protein links oxidative stress to inflammasome activation. *Nat Immunol* 2010, 11, 136–140, doi:10.1038/ni.1831.
- Wang, R.; Zhu, Y.; Lin, X.; Ren, C.; Zhao, J.; Wang, F.; Gao, X.; Xiao, R.; Zhao, L.; Chen, H.; et al. Influenza M2 protein regulates MAVS-mediated signaling pathway through interacting with MAVS and increasing ROS production. *Autophagy* 2019, 15, 1163–1181, doi:10.1080/15548627.2019.1580089.
- Zhang, B.; Pan, C.; Feng, C.; Yan, C.; Yu, Y.; Chen, Z.; Guo, C.; Wang, X. Role of mitochondrial reactive oxygen species in homeostasis regulation. *Redox Rep* 2022, 27, 45–52, doi:10.1080/13510002.2022.2046423.
- Sena, L.A.; Chandel, N.S. Physiological roles of mitochondrial reactive oxygen species. Mol Cell 2012, 48, 158–167, doi:10.1016/j.molcel.2012.09.025.
- Saikolappan, S.; Kumar, B.; Shishodia, G.; Koul, S.; Koul, H.K. Reactive oxygen species and cancer: A complex interaction. Cancer Lett 2019, 452, 132–143, doi:10.1016/j.canlet.2019.03.020.
- Jakubczyk, K.; Dec, K.; Kałduńska, J.; Kawczuga, D.; Kochman, J.; Janda, K. Reactive oxygen species sources, functions, oxidative damage. Pol Merkur Lekarski 2020, 48, 124–127.
- Sokolik, C.G.; Qassem, N.; Chill, J.H. The Disordered Cellular Multi-Tasker WIP and Its Protein-Protein Interactions: A Structural View. *Biomolecules* 2020, 10, 1084, doi:10.3390/biom10071084.
- 11. Malinverni, D.; Babu, M.M. Data-driven design of orthogonal protein-protein interactions. *Sci Signal* **2023**, *16*, eabm4484, doi:10.1126/scisignal.abm4484.
- 12. Calebiro, D.; Koszegi, Z.; Lanoiselée, Y.; Miljus, T.; O'Brien, S. G protein-coupled receptor-G protein interactions: a single-molecule perspective. *Physiol Rev* **2021**, *101*, 857–906, doi:10.1152/physrev.00021.2020.
- Cafarelli, T.M.; Desbuleux, A.; Wang, Y.; Choi, S.G.; De Ridder, D.; Vidal, M. Mapping, modeling, and characterization of protein-protein interactions on a proteomic scale. *Curr Opin Struct Biol* 2017, 44, 201– 210, doi:10.1016/j.sbi.2017.05.003.
- 14. Huttlin, E.L.; Bruckner, R.J.; Paulo, J.A.; Cannon, J.R.; Ting, L.; Baltier, K.; Colby, G.; Gebreab, F.; Gygi, M.P.; Parzen, H.; et al. Architecture of the human interactome defines protein communities and disease networks. *Nature* **2017**, *545*, 505–509, doi:10.1038/nature22366.
- Bachmann, V.A.; Bister, K.; Stefan, E. Interplay of PKA and Rac: fine-tuning of Rac localization and signaling. Small GTPases 2013, 4, 247–251, doi:10.4161/sgtp.27281.
- Yang, A.; Jude, K.M.; Lai, B.; Minot, M.; Kocyla, A.M.; Glassman, C.R.; Nishimiya, D.; Kim, Y.S.; Reddy, S.T.; Khan, A.A.; et al. Deploying synthetic coevolution and machine learning to engineer protein-protein interactions. *Science* 2023, 381, eadh1720, doi:10.1126/science.adh1720.
- 17. Alva, R.; Mirza, M.; Baiton, A.; Lazuran, L.; Samokysh, L.; Bobinski, A.; Cowan, C.; Jaimon, A.; Obioru, D.; Al Makhoul, T.; et al. Oxygen toxicity: cellular mechanisms in normobaric hyperoxia. *Cell Biol Toxicol* **2023**, 39, 111–143, doi:10.1007/s10565-022-09773-7.
- Abot, A.; Fried, S.; Cani, P.D.; Knauf, C. Reactive Oxygen Species/Reactive Nitrogen Species as Messengers in the Gut: Impact on Physiology and Metabolic Disorders. *Antioxid Redox Signal* 2022, 37, 394–415, doi:10.1089/ars.2021.0100.
- 19. Xu, T.; Gao, P.; Huang, Y.; Wu, M.; Yi, J.; Zhou, Z.; Zhao, X.; Jiang, T.; Liu, H.; Qin, T.; et al. Git1-PGK1 interaction achieves self-protection against spinal cord ischemia-reperfusion injury by modulating Keap1/Nrf2 signaling. *Redox Biol* **2023**, *62*, 102682, doi:10.1016/j.redox.2023.102682.
- Iachettini, S.; Ciccarone, F.; Maresca, C.; D' Angelo, C.; Petti, E.; Di Vito, S.; Ciriolo, M.R.; Zizza, P.; Biroccio, A. The telomeric protein TERF2/TRF2 impairs HMGB1-driven autophagy. *Autophagy* 2023, 19, 1479–1490, doi:10.1080/15548627.2022.2138687.
- Orru, R.; Pazmiño, D.E.T.; Fraaije, M.W.; Mattevi, A. Joint functions of protein residues and NADP(H) in oxygen activation by flavin-containing monooxygenase. *J Biol Chem* 2010, 285, 35021–35028, doi:10.1074/jbc.M110.161372.
- 22. Sumimoto, H. Structure, regulation and evolution of Nox-family NADPH oxidases that produce reactive oxygen species. FEBS J 2008, 275, 3249–3277, doi:10.1111/j.1742-4658.2008.06488.x.
- 23. Fialkow, L.; Wang, Y.; Downey, G.P. Reactive oxygen and nitrogen species as signaling molecules regulating neutrophil function. *Free Radic Biol Med* **2007**, 42, 153–164, doi:10.1016/j.freeradbiomed.2006.09.030.
- Wright, D.P.; Ulijasz, A.T. Regulation of transcription by eukaryotic-like serine-threonine kinases and phosphatases in Gram-positive bacterial pathogens. *Virulence* 2014, 5, 863–885, doi:10.4161/21505594.2014.983404.

- Heo, A.J.; Kim, S.B.; Ji, C.H.; Han, D.; Lee, S.J.; Lee, S.H.; Lee, M.J.; Lee, J.S.; Ciechanover, A.; Kim, B.Y.; et al. The N-terminal cysteine is a dual sensor of oxygen and oxidative stress. *Proc Natl Acad Sci U S A* 2021, 118, e2107993118, doi:10.1073/pnas.2107993118.
- Lee, P.; Chandel, N.S.; Simon, M.C. Cellular adaptation to hypoxia through hypoxia inducible factors and beyond. Nat Rev Mol Cell Biol 2020, 21, 268–283, doi:10.1038/s41580-020-0227-y.
- Tang, D.; Kang, R.; Berghe, T.V.; Vandenabeele, P.; Kroemer, G. The molecular machinery of regulated cell death. Cell Res 2019, 29, 347–364, doi:10.1038/s41422-019-0164-5.
- 28. Ray, P.D.; Huang, B.-W.; Tsuji, Y. Reactive oxygen species (ROS) homeostasis and redox regulation in cellular signaling. *Cell Signal* **2012**, 24, 981–990, doi:10.1016/j.cellsig.2012.01.008.
- Yamaguchi, M.; Yosiike, K.; Watanabe, H.; Watanabe, M. The marine factor 3,5-dihydroxy-4-methoxybenzyl alcohol suppresses growth, migration and invasion and stimulates death of metastatic human prostate cancer cells: targeting diverse signaling processes. *Anticancer Drugs* 2022, 33, 424–436, doi:10.1097/CAD.000000000001306.
- Tsai, W.-B.; Long, Y.; Park, J.-R.; Chang, J.T.; Liu, H.; Rodriguez-Canales, J.; Savaraj, N.; Feun, L.G.; Davies, M.A.; Wistuba, I.I.; et al. Gas6/Axl is the sensor of arginine-auxotrophic response in targeted chemotherapy with arginine-depleting agents. Oncogene 2016, 35, 1632–1642, doi:10.1038/onc.2015.237.
- Petricciuolo, M.; Davidescu, M.; Fettucciari, K.; Gatticchi, L.; Brancorsini, S.; Roberti, R.; Corazzi, L.; Macchioni, L. The efficacy of the anticancer 3-bromopyruvate is potentiated by antimycin and menadione by unbalancing mitochondrial ROS production and disposal in U118 glioblastoma cells. *Heliyon* 2020, 6, e05741, doi:10.1016/j.heliyon.2020.e05741.
- Marchionatti, A.M.; Picotto, G.; Narvaez, C.J.; Welsh, J.; Tolosa de Talamoni, N.G. Antiproliferative action of menadione and 1,25(OH)2D3 on breast cancer cells. J Steroid Biochem Mol Biol 2009, 113, 227–232, doi:10.1016/j.jsbmb.2009.01.004.
- 33. Burgoyne, J.R.; Haeussler, D.J.; Kumar, V.; Ji, Y.; Pimental, D.R.; Zee, R.S.; Costello, C.E.; Lin, C.; McComb, M.E.; Cohen, R.A.; et al. Oxidation of HRas cysteine thiols by metabolic stress prevents palmitoylation in vivo and contributes to endothelial cell apoptosis. *FASEB J* 2012, 26, 832–841, doi:10.1096/fj.11-189415.
- Oeste, C.L.; Díez-Dacal, B.; Bray, F.; García de Lacoba, M.; de la Torre, B.G.; Andreu, D.; Ruiz-Sánchez, A.J.; Pérez-Inestrosa, E.; García-Domínguez, C.A.; Rojas, J.M.; et al. The C-terminus of H-Ras as a target for the covalent binding of reactive compounds modulating Ras-dependent pathways. PLoS One 2011, 6, e15866, doi:10.1371/journal.pone.0015866.
- 35. Hu, C.-T.; Mandal, J.P.; Wu, W.-S. Regulation on tumor metastasis by Raf kinase inhibitory protein: New insight with reactive oxygen species signaling. *Tzu Chi Med J* **2021**, *33*, 332–338, doi:10.4103/tcmj.tcmj_296_20.
- Wu, W.-S.; Wu, J.-R.; Hu, C.-T. Signal cross talks for sustained MAPK activation and cell migration: the potential role of reactive oxygen species. *Cancer Metastasis Rev* 2008, 27, 303–314, doi:10.1007/s10555-008-9112-4
- Ge, X.; Zhang, Y.; Huang, F.; Wu, Y.; Pang, J.; Li, X.; Fan, F.; Liu, H.; Li, S. EGFR tyrosine kinase inhibitor Almonertinib induces apoptosis and autophagy mediated by reactive oxygen species in non-small cell lung cancer cells. *Hum Exp Toxicol* 2021, 40, S49–S62, doi:10.1177/09603271211030554.
- 38. Kim, Y.J.; Shin, Y.K.; Sohn, D.S.; Lee, C.S. Menadione induces the formation of reactive oxygen species and depletion of GSH-mediated apoptosis and inhibits the FAK-mediated cell invasion. *Naunyn-Schmiedeberg's Arch Pharmacol* **2014**, *387*, 799–809, doi:10.1007/s00210-014-0997-x.
- 39. Messina, S.; De Simone, G.; Ascenzi, P. Cysteine-based regulation of redox-sensitive Ras small GTPases. *Redox Biology* **2019**, *26*, 101282, doi:10.1016/j.redox.2019.101282.
- 40. Mott, H.R.; Carpenter, J.W.; Campbell, S.L. Structural and Functional Analysis of a Mutant Ras Protein That Is Insensitive to Nitric Oxide Activation. *Biochemistry* **1997**, *36*, 3640–3644, doi:10.1021/bi962790o.
- Chen, C.-H.; Cheng, T.-H.; Lin, H.; Shih, N.-L.; Chen, Y.-L.; Chen, Y.-S.; Cheng, C.-F.; Lian, W.-S.; Meng, T.-C.; Chiu, W.-T.; et al. Reactive Oxygen Species Generation Is Involved in Epidermal Growth Factor Receptor Transactivation through the Transient Oxidization of Src Homology 2-Containing Tyrosine Phosphatase in Endothelin-1 Signaling Pathway in Rat Cardiac Fibroblasts. *Mol Pharmacol* 2006, 69, 1347–1355, doi:10.1124/mol.105.017558.
- 42. Truong, T.H.; Carroll, K.S. Redox regulation of epidermal growth factor receptor signaling through cysteine oxidation. *Biochemistry* **2012**, *51*, 9954–9965, doi:10.1021/bi301441e.
- 43. Strange, R.; Metcalfe, T.; Thackray, L.; Dang, M. Apoptosis in normal and neoplastic mammary gland development. *Microsc Res Tech* **2001**, *52*, 171–181, doi:10.1002/1097-0029(20010115)52:2<171::AID-JEMT1003>3.0.CO;2-T.
- 44. Lu, Q.L.; Abel, P.; Foster, C.S.; Lalani, E.N. bcl-2: role in epithelial differentiation and oncogenesis. *Hum Pathol* **1996**, 27, 102–110, doi:10.1016/s0046-8177(96)90362-7.
- 45. Pohl, S.Ö.-G.; Agostino, M.; Dharmarajan, A.; Pervaiz, S. Cross Talk Between Cellular Redox State and the Antiapoptotic Protein Bcl-2. *Antioxidants & Redox Signaling* **2018**, 29, 1215–1236, doi:10.1089/ars.2017.7414.

- Blanco-Ayala, T.; Andérica-Romero, A.C.; Pedraza-Chaverri, J. New insights into antioxidant strategies against paraquat toxicity. Free Radical Research 2014, 48, 623–640, doi:10.3109/10715762.2014.899694.
- Xiao, D.; Zeng, Y.; Hahm, E.-R.; Kim, Y.-A.; Ramalingam, S.; Singh, S.V. Diallyl trisulfide selectively causes Bax- and Bak-mediated apoptosis in human lung cancer cells. *Environ Mol Mutagen* 2009, 50, 201–212, doi:10.1002/em.20431.
- Cremers, C.M.; Jakob, U. Oxidant Sensing by Reversible Disulfide Bond Formation*. *Journal of Biological Chemistry* 2013, 288, 26489–26496, doi:10.1074/jbc.R113.462929.
- Miles, J.A.; Hobor, F.; Trinh, C.H.; Taylor, J.; Tiede, C.; Rowell, P.R.; Jackson, B.R.; Nadat, F.A.; Ramsahye, P.; Kyle, H.F.; et al. Selective Affimers Recognise the BCL-2 Family Proteins BCL-xL and MCL-1 through Noncanonical Structural Motifs*. *Chembiochem* 2021, 22, 232–240, doi:10.1002/cbic.202000585.
- 50. Wolf, E.; Lento, C.; Pu, J.; Dickinson, B.C.; Wilson, D.J. Innate Conformational Dynamics Drive Binding Specificity in Anti-Apoptotic Proteins Mcl-1 and Bcl-2. *Biochemistry* **2023**, *62*, 1619–1630, doi:10.1021/acs.biochem.2c00709.
- 51. Anastas, J.N. Functional Crosstalk Between WNT Signaling and Tyrosine Kinase Signaling in Cancer. *Seminars in Oncology* **2015**, 42, 820–831, doi:10.1053/j.seminoncol.2015.09.020.
- 52. Krejci, P.; Aklian, A.; Kaucka, M.; Sevcikova, E.; Prochazkova, J.; Masek, J.K.; Mikolka, P.; Pospisilova, T.; Spoustova, T.; Weis, M.; et al. Receptor Tyrosine Kinases Activate Canonical WNT/β-Catenin Signaling via MAP Kinase/LRP6 Pathway and Direct β-Catenin Phosphorylation. *PLOS ONE* **2012**, *7*, e35826, doi:10.1371/journal.pone.0035826.
- Trejo-Solis, C.; Escamilla-Ramirez, A.; Jimenez-Farfan, D.; Castillo-Rodriguez, R.A.; Flores-Najera, A.; Cruz-Salgado, A. Crosstalk of the Wnt/β-Catenin Signaling Pathway in the Induction of Apoptosis on Cancer Cells. *Pharmaceuticals* 2021, 14, 871, doi:10.3390/ph14090871.
- 54. Catalano, T.; D'Amico, E.; Moscatello, C.; Di Marcantonio, M.C.; Ferrone, A.; Bologna, G.; Selvaggi, F.; Lanuti, P.; Cotellese, R.; Curia, M.C.; et al. Oxidative Distress Induces Wnt/β-Catenin Pathway Modulation in Colorectal Cancer Cells: Perspectives on APC Retained Functions. *Cancers (Basel)* **2021**, *13*, 6045, doi:10.3390/cancers13236045.
- Goitre, L.; Pergolizzi, B.; Ferro, E.; Trabalzini, L.; Retta, S.F. Molecular Crosstalk between Integrins and Cadherins: Do Reactive Oxygen Species Set the Talk? *Journal of Signal Transduction* 2011, 2012, e807682, doi:10.1155/2012/807682.
- Li, V.S.W.; Ng, S.S.; Boersema, P.J.; Low, T.Y.; Karthaus, W.R.; Gerlach, J.P.; Mohammed, S.; Heck, A.J.R.; Maurice, M.M.; Mahmoudi, T.; et al. Wnt Signaling through Inhibition of β-Catenin Degradation in an Intact Axin1 Complex. *Cell* 2012, 149, 1245–1256, doi:10.1016/j.cell.2012.05.002.
- 57. Miki, H.; Funato, Y. Regulation of intracellular signalling through cysteine oxidation by reactive oxygen species. *The Journal of Biochemistry* **2012**, *151*, 255–261, doi:10.1093/jb/mvs006.
- 58. Funato, Y.; Michiue, T.; Asashima, M.; Miki, H. The thioredoxin-related redox-regulating protein nucleoredoxin inhibits Wnt–β-catenin signalling through Dishevelled. *Nat Cell Biol* **2006**, *8*, 501–508, doi:10.1038/ncb1405.
- Cunnick, J.M.; Dorsey, J.F.; Standley, T.; Turkson, J.; Kraker, A.J.; Fry, D.W.; Jove, R.; Wu, J. Role of tyrosine kinase activity of epidermal growth factor receptor in the lysophosphatidic acid-stimulated mitogenactivated protein kinase pathway. *J Biol Chem* 1998, 273, 14468–14475, doi:10.1074/jbc.273.23.14468.
- Banskota, S.; Dahal, S.; Kwon, E.; Kim, D.Y.; Kim, J.-A. β-Catenin gene promoter hypermethylation by reactive oxygen species correlates with the migratory and invasive potentials of colon cancer cells. *Cell Oncol.* 2018, 41, 569–580, doi:10.1007/s13402-018-0391-7.
- 61. Sorolla, M.A.; Hidalgo, I.; Sorolla, A.; Montal, R.; Pallisé, O.; Salud, A.; Parisi, E. Microenvironmental Reactive Oxygen Species in Colorectal Cancer: Involved Processes and Therapeutic Opportunities. *Cancers* **2021**, *13*, 5037, doi:10.3390/cancers13205037.
- 62. Lin, S.; Li, Y.; Zamyatnin Jr., A.A.; Werner, J.; Bazhin, A.V. Reactive oxygen species and colorectal cancer. *Journal of Cellular Physiology* **2018**, 233, 5119–5132, doi:10.1002/jcp.26356.
- 63. Wong, S.H.M.; Fang, C.M.; Chuah, L.-H.; Leong, C.O.; Ngai, S.C. E-cadherin: Its dysregulation in carcinogenesis and clinical implications. *Critical Reviews in Oncology/Hematology* **2018**, *121*, 11–22, doi:10.1016/j.critrevonc.2017.11.010.
- 64. Daugherty, R.L.; Gottardi, C.J. Phospho-regulation of β-Catenin Adhesion and Signaling Functions. *Physiology* **2007**, *22*, 303–309, doi:10.1152/physiol.00020.2007.
- Galanis, A.; Pappa, A.; Giannakakis, A.; Lanitis, E.; Dangaj, D.; Sandaltzopoulos, R. Reactive oxygen species and HIF-1 signalling in cancer. *Cancer Letters* 2008, 266, 12–20, doi:10.1016/j.canlet.2008.02.028.
- 66. Hagen, T. Oxygen versus Reactive Oxygen in the Regulation of HIF-1<svg style="vertical-align:-0.216pt;width:14.1125px;" id="M1" height="10.725" version="1.1" viewBox="0 0 14.1125 10.725" width="14.1125" xmlns:xlink="http://www.w3.org/1999/xlink" xmlns="http://www.w3.org/2000/svg"> <g transform="matrix(.022,-0,0,-.022,.062,10.4)">path id="x1D736" d="M505 450h125l-163 -259q-1 -4 -1 -12q0 19 1 -23q9 -98 41 -98q36 0 58 76l24 -7q-34 -139 -121 -139q-32 0 -57.5 24t-31.5 55q-72 -77 -181 -77q-73 0 -114 44t-41 124q0 118 79 210q79 93 197 93q51 0 86 -27t50 -71zM369 281q0 141 -62 141q-43 0 -95 -84 q-52 -82 -52 -

- 67. Arany, Z.; Huang, L.E.; Eckner, R.; Bhattacharya, S.; Jiang, C.; Goldberg, M.A.; Bunn, H.F.; Livingston, D.M. An essential role for p300/CBP in the cellular response to hypoxia. *Proceedings of the National Academy of Sciences* **1996**, 93, 12969–12973, doi:10.1073/pnas.93.23.12969.
- Parandavar, E.; Yazdanparast, R. Differential impact of various reactive oxygen species (ROS) on HIF-1α/p53 direct interaction in SK-N-MC neuroblastoma cells. *Cell Biosci* 2017, 7, 1–10, doi:10.1186/s13578-017-0180-4.
- Kishore, C.; Sundaram, S.; Karunagaran, D. Vitamin K3 (menadione) suppresses epithelial-mesenchymaltransition and Wnt signaling pathway in human colorectal cancer cells. *Chemico-Biological Interactions* 2019, 309, 108725, doi:10.1016/j.cbi.2019.108725.
- Iyer, N.G.; Özdag, H.; Caldas, C. p300/CBP and cancer. Oncogene 2004, 23, 4225–4231, doi:10.1038/sj.onc.1207118.
- Meek, D.W. Regulation of the p53 response and its relationship to cancer1. Biochemical Journal 2015, 469, 325–346, doi:10.1042/BJ20150517.
- Dengler, V.L.; Galbraith, M.; Espinosa, J.M. Transcriptional regulation by hypoxia inducible factors. Crit Rev Biochem Mol Biol 2014, 49, 1–15, doi:10.3109/10409238.2013.838205.
- 73. Choi, K.-S.; Bae, M.-K.; Jeong, J.-W.; Moon, H.-E.; Kim, K.-W. Hypoxia-induced angiogenesis during carcinogenesis. *J Biochem Mol Biol* **2003**, *36*, 120–127, doi:10.5483/bmbrep.2003.36.1.120.
- 74. Vleugel, M.M.; Shvarts, D.; van der Wall, E.; van Diest, P.J. p300 and p53 levels determine activation of HIF-1 downstream targets in invasive breast cancer. *Hum Pathol* **2006**, *37*, 1085–1092, doi:10.1016/j.humpath.2006.03.015.
- Aggarwal, V.; Tuli, H.S.; Varol, A.; Thakral, F.; Yerer, M.B.; Sak, K.; Varol, M.; Jain, A.; Khan, Md.A.; Sethi,
 G. Role of Reactive Oxygen Species in Cancer Progression: Molecular Mechanisms and Recent Advancements. *Biomolecules* 2019, 9, 735, doi:10.3390/biom9110735.
- ArulJothi, K.N.; Kumaran, K.; Senthil, S.; Nidhu, A.B.; Munaff, N.; Janitri, V.B.; Kirubakaran, R.; Singh, S.K.; Gupt, G.; Dua, K.; et al. Implications of reactive oxygen species in lung cancer and exploiting it for therapeutic interventions. *Med Oncol* 2023, 40, 43, doi:10.1007/s12032-022-01900-y.
- 77. Hou, Y.; Wang, H.; Wu, J.; Guo, H.; Chen, X. Dissecting the pleiotropic roles of reactive oxygen species (ROS) in lung cancer: From carcinogenesis toward therapy. *Medicinal Research Reviews* **2024**, *44*, 1566–1595, doi:10.1002/med.22018.
- 78. Kong, J.; Fan, R.; Zhang, Y.; Jia, Z.; Zhang, J.; Pan, H.; Wang, Q. Oxidative stress in the brain–lung crosstalk: cellular and molecular perspectives. *Front Aging Neurosci* **2024**, *16*, 1389454, doi:10.3389/fnagi.2024.1389454.
- 79. Nakamura, H.; Takada, K. Reactive oxygen species in cancer: Current findings and future directions. *Cancer Sci* **2021**, *112*, 3945–3952, doi:10.1111/cas.15068.
- 80. Endale, H.T.; Tesfaye, W.; Mengstie, T.A. ROS induced lipid peroxidation and their role in ferroptosis. *Front Cell Dev Biol* **2023**, *11*, 1226044, doi:10.3389/fcell.2023.1226044.
- 81. Su, L.-J.; Zhang, J.-H.; Gomez, H.; Murugan, R.; Hong, X.; Xu, D.; Jiang, F.; Peng, Z.-Y. Reactive Oxygen Species-Induced Lipid Peroxidation in Apoptosis, Autophagy, and Ferroptosis. *Oxid Med Cell Longev* **2019**, 2019, 5080843, doi:10.1155/2019/5080843.
- 82. Verma, P.; Rishi, B.; George, N.G.; Kushwaha, N.; Dhandha, H.; Kaur, M.; Jain, A.; Jain, A.; Chaudhry, S.; Singh, A.; et al. Recent advances and future directions in etiopathogenesis and mechanisms of reactive oxygen species in cancer treatment. *Pathol. Oncol. Res.* **2023**, 29, 1611415, doi:10.3389/pore.2023.1611415.
- 83. Milkovic, L.; Zarkovic, N.; Marusic, Z.; Zarkovic, K.; Jaganjac, M. The 4-Hydroxynonenal–Protein Adducts and Their Biological Relevance: Are Some Proteins Preferred Targets? *Antioxidants (Basel)* **2023**, *12*, 856, doi:10.3390/antiox12040856.
- 84. Žarković, N.; Gęgotek, A.; Łuczaj, W.; Jaganjac, M.; Šunjić, S.B.; Žarković, K.; Skrzydlewska, E. Overview of the Lipid Peroxidation Measurements in Patients by the Enzyme-Linked Immunosorbent Assay Specific for the 4-Hydroxynonenal-Protein Adducts (4-HNE-ELISA). FBL 2024, 29, 153, doi:10.31083/j.fbl2904153.
- 85. Breitzig, M.; Bhimineni, C.; Lockey, R.; Kolliputi, N. 4-Hydroxy-2-nonenal: a critical target in oxidative stress? *Am J Physiol Cell Physiol* **2016**, *311*, C537–C543, doi:10.1152/ajpcell.00101.2016.
- Dwivedi, S.; Sharma, A.; Patrick, B.; Sharma, R.; Awasthi, Y.C. Role of 4-hydroxynonenal and its metabolites in signaling. *Redox Report* 2007, 12, 4–10, doi:10.1179/135100007X162211.
- Graf, P.C.F.; Jakob, U. Redox-regulated molecular chaperones. *Cell Mol Life Sci* 2002, 59, 1624–1631, doi:10.1007/pl00012489.
- 88. Lingappan, K. NF-κB in Oxidative Stress. Curr Opin Toxicol 2018, 7, 81–86, doi:10.1016/j.cotox.2017.11.002.
- Morgan, M.J.; Liu, Z. Crosstalk of reactive oxygen species and NF-κB signaling. Cell Res 2011, 21, 103–115, doi:10.1038/cr.2010.178.
- 90. Butterfield, D.A.; Hensley, K.; Cole, P.; Subramaniam, R.; Aksenov, M.; Aksenova, M.; Bummer, P.M.; Haley, B.E.; Carney, J.M. Oxidatively induced structural alteration of glutamine synthetase assessed by

- Juan, C.A.; Pérez de la Lastra, J.M.; Plou, F.J.; Pérez-Lebeña, E. The Chemistry of Reactive Oxygen Species (ROS) Revisited: Outlining Their Role in Biological Macromolecules (DNA, Lipids and Proteins) and Induced Pathologies. *Int J Mol Sci* 2021, 22, 4642, doi:10.3390/ijms22094642.
- 92. Klotz, L.-O.; Sánchez-Ramos, C.; Prieto-Arroyo, I.; Urbánek, P.; Steinbrenner, H.; Monsalve, M. Redox regulation of FoxO transcription factors. *Redox Biol* **2015**, *6*, 51–72, doi:10.1016/j.redox.2015.06.019.
- 93. Calnan, D.R.; Brunet, A. The FoxO code. Oncogene 2008, 27, 2276–2288, doi:10.1038/onc.2008.21.
- 94. Brunet, A.; Sweeney, L.B.; Sturgill, J.F.; Chua, K.F.; Greer, P.L.; Lin, Y.; Tran, H.; Ross, S.E.; Mostoslavsky, R.; Cohen, H.Y.; et al. Stress-dependent regulation of FOXO transcription factors by the SIRT1 deacetylase. *Science* **2004**, *303*, 2011–2015, doi:10.1126/science.1094637.
- Daitoku, H.; Hatta, M.; Matsuzaki, H.; Aratani, S.; Ohshima, T.; Miyagishi, M.; Nakajima, T.; Fukamizu, A. Silent information regulator 2 potentiates Foxo1-mediated transcription through its deacetylase activity. Proc Natl Acad Sci U S A 2004, 101, 10042–10047, doi:10.1073/pnas.0400593101.
- Klotz, L.-O.; Sánchez-Ramos, C.; Prieto-Arroyo, I.; Urbánek, P.; Steinbrenner, H.; Monsalve, M. Redox regulation of FoxO transcription factors. *Redox Biol* 2015, 6, 51–72, doi:10.1016/j.redox.2015.06.019.
- 97. Waudby, C.A.; Alvarez-Teijeiro, S.; Josue Ruiz, E.; Suppinger, S.; Pinotsis, N.; Brown, P.R.; Behrens, A.; Christodoulou, J.; Mylona, A. An intrinsic temporal order of c-JUN N-terminal phosphorylation regulates its activity by orchestrating co-factor recruitment. *Nat Commun* **2022**, *13*, 6133, doi:10.1038/s41467-022-33866-w.
- Kang, K.A.; Piao, M.J.; Madduma Hewage, S.R.K.; Ryu, Y.S.; Oh, M.C.; Kwon, T.K.; Chae, S.; Hyun, J.W. Fisetin induces apoptosis and endoplasmic reticulum stress in human non-small cell lung cancer through inhibition of the MAPK signaling pathway. *Tumour Biol* 2016, 37, 9615–9624, doi:10.1007/s13277-016-4864-x.
- 99. Xin, H.; Deng, Y.; Cao, J. Proviral insertion in murine lymphomas 2 promotes stomach cancer progression by regulating apoptosis via reactive oxygen species-triggered endoplasmic reticulum stress. *Biochem Biophys Res Commun* **2018**, *506*, 145–152, doi:10.1016/j.bbrc.2018.09.062.
- Klann, E.; Thiels, E. Modulation of protein kinases and protein phosphatases by reactive oxygen species: implications for hippocampal synaptic plasticity. *Prog Neuropsychopharmacol Biol Psychiatry* 1999, 23, 359–376, doi:10.1016/s0278-5846(99)00002-0.
- 101. Son, Y.; Kim, S.; Chung, H.-T.; Pae, H.-O. Reactive oxygen species in the activation of MAP kinases. *Methods Enzymol* **2013**, *528*, 27–48, doi:10.1016/B978-0-12-405881-1.00002-1.
- 102. Navarro-Antolín, J.; Hernández-Perera, O.; López-Ongil, S.; Rodríguez-Puyol, M.; Rodríguez-Puyol, D.; Lamas, S. CsA and FK506 up-regulate eNOS expression: role of reactive oxygen species and AP-1. *Kidney Int Suppl* **1998**, *68*, S20-24, doi:10.1046/j.1523-1755.1998.06807.x.
- 103. Yang, H.-L.; Lin, M.-W.; Korivi, M.; Wu, J.-J.; Liao, C.-H.; Chang, C.-T.; Liao, J.-W.; Hseu, Y.-C. Coenzyme Q0 regulates NFκB/AP-1 activation and enhances Nrf2 stabilization in attenuation of LPS-induced inflammation and redox imbalance: Evidence from in vitro and in vivo studies. *Biochim Biophys Acta* **2016**, 1859, 246–261, doi:10.1016/j.bbagrm.2015.11.001.
- 104. Han, Y.H.; Park, W.H. MG132 as a proteasome inhibitor induces cell growth inhibition and cell death in A549 lung cancer cells via influencing reactive oxygen species and GSH level. *Hum Exp Toxicol* **2010**, 29, 607–614, doi:10.1177/0960327109358733.
- Forrester, S.J.; Kikuchi, D.S.; Hernandes, M.S.; Xu, Q.; Griendling, K.K. Reactive Oxygen Species in Metabolic and Inflammatory Signaling. Circulation Research 2018, 122, 877–902, doi:10.1161/CIRCRESAHA.117.311401.
- Sayin, V.I.; Ibrahim, M.X.; Larsson, E.; Nilsson, J.A.; Lindahl, P.; Bergo, M.O. Antioxidants accelerate lung cancer progression in mice. Sci Transl Med 2014, 6, 221ra15, doi:10.1126/scitranslmed.3007653.
- Bansal, M.P. ROS, Redox Regulation and Signaling in Cancer Cells. In Redox Regulation and Therapeutic Approaches in Cancer; Bansal, M.P., Ed.; Springer Nature: Singapore, 2023; pp. 1–47 ISBN 978-981-9973-42-2.
- 108. Gorrini, C.; Harris, I.S.; Mak, T.W. Modulation of oxidative stress as an anticancer strategy. *Nat Rev Drug Discov* **2013**, *12*, 931–947, doi:10.1038/nrd4002.
- Liou, G.-Y.; Storz, P. Reactive oxygen species in cancer. Free Radic Res 2010, 44, 479–496, doi:10.3109/10715761003667554.
- 110. Vousden, K.H.; Prives, C. Blinded by the Light: The Growing Complexity of p53. *Cell* **2009**, 137, 413–431, doi:10.1016/j.cell.2009.04.037.
- 111. Chen, X.; Zhao, Y.; Luo, W.; Chen, S.; Lin, F.; Zhang, X.; Fan, S.; Shen, X.; Wang, Y.; Liang, G. Celastrol induces ROS-mediated apoptosis via directly targeting peroxiredoxin-2 in gastric cancer cells. *Theranostics* **2020**, *10*, 10290–10308, doi:10.7150/thno.46728.
- 112. Trachootham, D.; Alexandre, J.; Huang, P. Targeting cancer cells by ROS-mediated mechanisms: a radical therapeutic approach? *Nat Rev Drug Discov* **2009**, *8*, 579–591, doi:10.1038/nrd2803.

113. Wang, L.; Wang, C.; Tao, Z.; Zhao, L.; Zhu, Z.; Wu, W.; He, Y.; Chen, H.; Zheng, B.; Huang, X.; et al. Curcumin derivative WZ35 inhibits tumor cell growth via ROS-YAP-JNK signaling pathway in breast cancer. J Exp Clin Cancer Res 2019, 38, 460, doi:10.1186/s13046-019-1424-4.

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