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Posted Date: 14 August 2024

doi: [10.20944/preprints202408.1008.v1](https://doi.org/10.20944/preprints202408.1008.v1)

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Article

Yield Stability of Drought Tolerant Sorghum [*Sorghum bicolor* (L.) Moench] Genotypes in Southern and West Hararghe, Ethiopia

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Abstract: A multi-environment evaluation of sorghum genotypes was conducted across six environments in the 2021 main growing season in a randomized complete block design with three replications. The objectives of the study were to estimate the magnitude of genotypes by environment interaction (GEI) and grain yield stability of drought-tolerant sorghum genotypes across different environments. Data were subjected to analysis of variance, Additive Main Effects and Multiplicative Interaction (AMMI), and GGE biplot analysis. Combined analysis of variance revealed significant variations among genotypes, environments, and GEI for yield and yield-related traits, indicating that these factors significantly affected grain yield. The maximum mean grain yield value of genotypes due to the mean effect of the environment was obtained from G1 (5119.93 kg ha⁻¹), followed by G14 (4834.57 kg ha⁻¹), and G18 (4801.20 ha⁻¹), while the least mean grain yield was obtained from G3 (3314.50 kg ha⁻¹). The multiplicative variance of the treatment sum of squares due to GEI was partitioned into four principal component axes (PCA). Sum squares of the first and second interaction principal component axis (IPCA) explained 71.07% and 17.50% of the GEI variation, respectively. The IPCA1&2 mean squares were highly significant ($P \leq 0.01$), indicating the adequacy of the AMMI model with the first two IPCAs for cross-validation of grain yield variation. The magnitude of the GEI sum squares was 3.9 times that of the genotype sum squares for grain yield, indicating the presence of substantial differences in genotypic responses across environments. The results of cultivar superiority measure (Pi), yield stability index (YSI), AMMI stability value (ASV), regression coefficient (bi), and deviation from regression ($S^2 di$) depicted that genotypes G18, G22, G31, and 32 were the most stable genotypes for grain yield and biomass yield, respectively. AMMI2 biplot showed Jinka, Alduba, and Kako were the most discriminating environments as indicated by the long distance from the origin; whereas testing locations Meioso and Gato with short vector length indicated that these locations had less discriminating power on the genotypes' performance. The study has provided *precious* information on the yield stability status of the sorghum *genotypes* and the *best* environments for future improvement programs in Ethiopia.

Keywords: AMM1 biplot; ASV; Yield stability index; cultivar superiority measure

1. Introduction

Sorghum [*Sorghum bicolor* (L.) Monech] is the fifth most important cereal crop in the world, after wheat, rice, maize, and barley[1]. It is a C4 plant that has high photosynthetic efficiency and originated in the Ethiopian region [2]. Multi-environment trials are used to select the best-performing genotype for different locations and environmental conditions. Plant breeders need to identify drought-tolerant genotypes with stable yield performance in various environments. Developing countries require stable cultivars with high yields.

In Ethiopia, several studies have been conducted to investigate the interaction between genotype and environment when it comes to crops like sorghum [3–7], maize [8,9], wheat [10], teff [11], and finger millet [12]. However, there is limited information available on the impact of genotype,

environment, and GEI on sorghum yield using drought-tolerant genotypes in Ethiopia. To address this gap, a study was carried out to determine the extent of the effect of genotypes by environment interaction and to assess the performance and stability of drought-tolerant sorghum genotypes that show promise for adaptation to different conditions and cultivation under farmers' conditions in Ethiopia.

2. Materials and Methods

2.1. Description of the Study Area

The experiment was conducted at six locations during the 2021 main cropping season. These locations represent the main lowland sorghum-growing areas of the country (Figure 1).

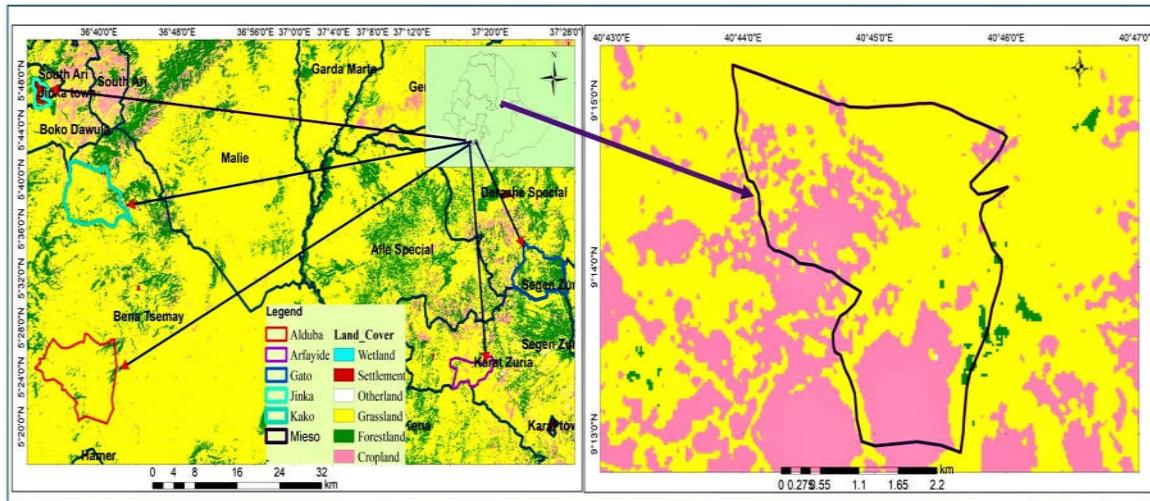


Figure 1. Study area map. Different colors designate land cover of study locations.

2.2. Trial Materials

Thirty-four genotypes were selected based on their drought performance from the moisture stress trial conducted at Weito (Appendix Tables A1 and A2), and two standard check varieties were included. A list of genotypes used for genotype-by-environment experiments is provided in (Table 2).

Table 2. List of genotypes used for genotypes by environment experiments.

Serial no.	Genotypes	Code	Source
1	27907	G1	EBI
2	69313	G2	EBI
3	69330	G3	EBI
4	70084	G4	EBI
5	70154	G5	EBI
6	70373	G6	EBI
7	71018	G7	EBI
8	74654	G8	EBI
9	74669	G9	EBI
10	74679	G10	EBI
11	74680	G11	EBI
12	74684	G12	EBI
13	74686	G13	EBI
14	74691	G14	EBI
15	74693	G15	EBI

16	74704	G16	EBI
17	74705	G17	EBI
18	200617	G18	EBI
19	201453	G21	EBI
20	204602	G22	EBI
21	204619	G23	EBI
22	204629	G24	EBI
23	204631	G25	EBI
24	204633	G26	EBI
25	204634	G27	EBI
26	206285	G28	EBI
27	206286	G29	EBI
28	213008	G30	EBI
29	213017	G31	EBI
30	213019	G32	EBI
31	213026	G33	EBI
32	214010	G34	EBI
33	214109	G35	EBI
34	216906	G36	EBI
35	Dekeba(Standard check)	G19	MARC
36	Melkam(Standard check)	G20	MARC

EBI =Ethiopian Biodiversity Institute (EBI), MARC=Melkassa Agricultural Research Center.

2.2.1. Experimental Design and Procedures

The experiment used randomized complete block design with three replications at all locations with plot size of 3.75mX5m. Seeds were drilled into 5-meter-long paired rows spaced 0.75 meters apart. Weeds were removed from the plots 2-3 weeks after sowing. The seed rate was 15 kg/ha, and plots received NPS fertilizer at planting (19 kg/ha N, 38 kg/ha P2O5, and 7 kg/ha S), with an additional 23 kg/ha of nitrogen in the form of urea applied at 45 days after planting. Data was collected from the four middle rows.

2.2.2. Data Collection

Data was collected on sorghum based on descriptors developed by the International Board for Plant Genetic Resources [13]. At maturity, yield components were recorded, including panicle length, panicle weight, panicle yield, and thousand kernel weight. Data was collected on a plot basis and plant basis, including measurements of straw weight, grain yield, total biomass, harvest index, and panicle weight.

2.2.3. Data Analyses

Various statistical software packages were used to analyze the data. SAS software 9.0 was used for combined analyses of variance and mean comparison with the LSD test [14]. Additionally, GEA-R Version 4.1 was used for several analyses, including AMMI analysis and GGE biplot stability analysis [15].

Analysis of Variance for Individual location and Combined Data over the Location

The data from different locations were analyzed using a mixed linear model through the analysis of variance based on Gomez and Gomez [16]. Bartlett's test was used to check the homogeneity of error variances before combining the analysis [17]. The combined analysis of variance was conducted using SAS software 9.0[14] to determine the differences between genotypes across and among environments and their interaction. In the combined analysis, genotypes were considered as fixed

while locations were considered a random variable. The following model was used for ANOVA of data of individual location:

$$Y_{ij} = \mu + G_i + B_j + e_{ij}$$

where; Y_{ij} = observed value of genotype i in block j , μ = Grand mean of the experiment, G_i = the effect of genotype i , B_j = the effect of block j , e_{ij} = the error of genotype i in block j .

Combined analysis of variance over locations was carried out using the following statistical model:

$$Y_{ijk} = \mu + G_i + E_j + G_E_{ij} + B_k(j) + e_{ijk}$$

where; Y_{ijk} = observed value of genotype i in block k of environment (location) j , μ = Grand mean of the experiment, G_i = the effect of genotype i , G_E_{ij} = the interaction effect of genotype i with environment j , $B_k(j)$ = the effect of block k in location (environment) j , e_{ijk} = the error effect of genotype i in block k of environment j . Mean separation was done using Duncan multiple range test to discriminate the genotypes and identify superior ones based on yield.

Stability Analysis

Analysis of variance only detects genotype by environment interaction effects. To determine the stability of a genotype's performance across different environments, breeders need additional information. Stability depends on the genetic structure of the cultivar's population and the genotype of individual plants. So, the significance of genotype by environment interaction was further analyzed using stability parameters. Means of genotypes for grain yield across locations were analyzed using SAS [18]. AMMI model, biplot technique, and AMMI stability value analysis were computed per standard procedures.

AMMI Analysis

The Additive Main effect and Multiplicative Interaction (AMMI) model analysis was performed for grain yield and biomass. The AMMI model equation is given as:

$$y_{ij} = \mu + G_i + E_j + (\sum K_n V_{ni} S_{nj}) + Q_{ij} + e_{ij}$$

where, y_{ij} = is the observed yield of genotype i in environment j

μ = is the grand mean, G_i = the additive effect of the i^{th} genotype (genotype means minus the grand mean), E_j = is the additive effect of the j^{th} environment (environment mean deviation), K_n = is the eigenvalues of the PCA axis n , V_{ni} and S_{nj} = are scores for the genotype i and environment j for the PCA axis n , Q_{ij} = is the residual for the first n multiplicative components, e_{ij} = is the error

AMMI Stability Value (ASV)

The AMMI stability value as described by Purchase [19] was calculated as follows:

$$\text{ASV} = \sqrt{\left[\frac{\text{IPCAA 1 Sum of squares}}{\text{IPCAA 2 Sum of squares}} (\text{IPCAA 1 scores}) \right]^2 + [\text{IPCAA 2 scores}]^2}$$

Where;

ASV = AMMI stability value, IPCAA1 = interaction principal component analysis 1, IPCAA2 = interaction principal component analysis 2, SSIPCAA1 = sum of square of the interaction principal component one, SSIPCAA2 = sum of square of the interaction principal component two.

3. Results and Discussion

3.1. Combined Analysis of Variance

In the study, significant differences were found in the environment, genotype, and genotype by environment interactions for all traits studied (Table 1). Our findings are in agreement with an earlier study on bread wheat [20]. The AMMI analysis revealed that a large portion of the variation was attributed to environmental effects (69.29%) followed by genotype by environment interactions (24.49%) and genotype effects (6.22%). This suggests that the performance of sorghum genotypes

varied across different locations, highlighting the need for further evaluation of genotypes with wider adaptability and testing them in diverse environments. The diversity in environments indicated by the large sum of squares emphasizes the importance of testing sorghum genotypes at multiple locations to account for variations in climatic and soil conditions. These findings are consistent with earlier studies [21–23] on sorghum and support the idea that testing genotypes across various environments is crucial for understanding their performance and adaptation.

3.2. Mean Performance of Genotypes

Grain Yield

The analysis of variance results for each environment showed significant differences ($P \leq 0.01$) in grain yield among sorghum genotypes tested at multiple locations (Table 1). This aligns with previous studies on sorghum that also reported significant variations in grain yield among different varieties [24]. The tested genotypes exhibited varying performances for grain yield across the different environments, with the mean grain yield ranging from 1148.1 kg/ha for genotype G34 at Arfayide to 9137 kg/ha for genotype G36 at Jinka, with an overall environmental mean yield of 4250.12 kg/ha (Table 2). The average environmental grain yield varied from 1721.61 kg/ha at Arfayide to 6182.31 kg/ha at Jinka, while the average genotype grain yield across environments ranged from 3314.50 kg/ha for genotype G3 to 5119.93 kg/ha for G1 (Table 2). These findings indicate that it is crucial to evaluate sorghum genotypes in various environments to comprehend their adaptability and performance variations. Additionally, the study demonstrated that the GEI sum square magnitudes were around 3.9 times greater than those of the genotypes sum squares for grain yield. This suggests significant differences in genotypic responses across different environments, resulting in observed variations in genotypic responses across environments. These findings align with the research by Amare *et al.* [24], which also highlighted a threefold difference in the magnitude of the GEI sum of squares compared to wheat genotypes, indicating substantial variations in genotypic responses across environments. The larger sum of squares of GEI relative to genotypes underscores the significant differences in genotypic responses across environments, emphasizing the considerable variance in genotypic responses across different conditions. Consequently, GEI poses challenges to the selection process by complicating the assessment of genotypes' yield performance and weakening the correlation between genotypic and phenotypic values [25]. The GEI observed in this study follows a cross-over pattern, leading to shifts in genotype rankings for specific environments, making it challenging to interpret grain yield based solely on genotype and environment means. These findings are consistent with the findings of Tekle *et al.* [26] regarding mung bean.

The highest mean grain yields across the environment were recorded for the genotypes (G1 and G14), with a mean yield of (5119.93 and 4834.57 kg ha⁻¹), respectively. Whereas, the lowest mean grain yield was recorded from the genotype (G3) with an average mean yield of 3314.50 kg ha⁻¹ (Table 2), indicating that the tested genotypes had inconsistent performance across the tested environments. In this study, most of the tested genotypes gave relatively good grain yield performance, and it could be suggested that there is an opportunity to get high-yielding sorghum genotypes for future variety development. The large variation due to the environments in our study also confirmed the high diversity of weather conditions during growing seasons and the locations had different soil types, temperature, and rainfall as well as altitude, directly affecting the performances of the genotypes. Thus, the selection and development of sorghum varieties in the future should follow environment-specific approaches. This result is in agreement with the work of Yitayeh *et al.* [22] and Abiy [27] on the early maturing sorghum genotype, who reported that the performance of sorghum genotypes was different from location to location, similar to that of Tegegn *et al.* [28] in finger millet. Ranking based on the genotype-focused scaling assumed that stability and mean yield were equally important [29].

The best candidate genotypes were expected to have a high mean seed yield with stable performance across all test locations. However, such genotypes are very rare to find in practice.

Therefore, high-yielding and relatively stable genotypes can be considered as a reference for genotype evaluation [30].

Table 1. Analysis of variance of morphological traits between genotypes (G), environment (E) and GEI interaction for 36 sorghum genotypes across six locations.

Trait		DF		MD		GFP		PHT		
Source	of	df	MS	Percent	MS	Percent	MS	Percent	MS	Percent
variation										
Environment(E)		5	3026.16**	28.64	17357.93**	56.66	8170.96**	44.99	120104.56*	28.38
Genotype (G)		35	150.20**	9.95	150.84**	3.45	77.04**	3.01	13494.25**	22.32
GxE interactions		17	71.95**	23.84	61.46**	7.02	72.61**	13.97	2023.64**	16.74
Rep(E)		5	226.2**	5.14	3167.80**	24.82	1337.36**	17.64	9735.66**	5.52
Error		12	40.77		32.42	29.39	8.06	44.27	20.45	44.27
		42								27.05
		0								
Trait		PL		PY		PW		SW		
Source	of	df	MS	Percent	MS	Percent	MS	Percent	MS	Percent
variation										
Environment(E)		5	3631.75**	34.65	418611.32*	58.85	366168.77*	40.70	419507928	47.34
Genotype (G)		35	137.87**	9.21	2025.96*	3.18	7848.58**	6.11	15762275**	12.45
GxE interactions		17	85.18**	28.44	3042.56**	14.97	5260.07**	20.46	2353159**	9.29
Rep(E)		5	82.24**	1.88	9413.07**	3.18	19546.56**	5.21	70190040**	19.01
Error		12	32.21	25.82	1779.34	21.01	2946.62	27.51	1255266	11.90
		42								
		0								
Trait		BM		TKW		GY		HI		
Source	of	df	MS	Percent	MS	Percent	MS	Percent	MS	Percent
variation										
Environment(E)		5	597883554	43.59	1799.68**	43.90	298875131**	48.30	419507928	39.58
Genotype (G)		35	16989501*	8.67	74.47**	12.72	3897432**	4.14	15762275**	7.68
GxE interactions		17	4667812**	11.91	16.20**	13.83	3031950**	16.98	2353159**	21.12
Rep(E)		5	103932563	18.18	75.54**	4.42	12269443**	4.77	70190040**	3.24
Error		12	**	17.65	12.27	0.06	1877757	25.81	1255266	28.38
		42	2881993							
		0								

*, **, =Significant at 5% and 1% level of probability respectively, DF=days to flowering, DM=days to maturity, GFP=grain filling period PHT=plant height (cm), PL=panicle length (cm), PY=panicle yield (g), PW=panicle weight (g), SW=Straw weight (kg), BM= biomass (kg), TKW= thousand- kernel weight (g) and GY=grain yield (kg).

Table 2. Mean grain yield (kg ha^{-1}) of 36 sorghum genotypes across six different test locations.

Genotype	Jinka	Kako	Alduba	Arfayide	Gato	Mieso	Mean
G1	7694 ^{abcd}	5852 ^{abc}	8440 ^{ab}	1816.0 ^{abcde}	3669.6 ^a	3248 ^{abcd}	5119.93
G2	7749 ^{abcd}	3639 ^c	5096 ^{bcd^{efg}}	1957.2 ^{abcde}	1455.8 ^{bc}	3311 ^{abcd}	3868.00
G3	5176 ^{abcd}	3555 ^c	4992 ^{d^{c^{efg}}}	1331.6 ^{bcde}	1269.4 ^c	3563 ^{abcd}	3314.50
G4	4957 ^{bcd}	6080 ^{abc}	4851 ^{defg}	1034.9 ^e	4092.1 ^{ab}	4859 ^{abc}	4312.33
G5	4134 ^d	5354 ^{abc}	4277 ^{efg}	1333.3 ^{bcde}	3581.1 ^a	4596 ^{abc}	3879.23
G6	5406 ^{abcd}	5347 ^{abc}	2677 ^g	1837.3 ^{abcde}	3490.6 ^a	2737 ^{bcd}	3582.48
G7	7798 ^{abcd}	6844 ^a	5511 ^{bcd^{efg}}	1503.7 ^{bcde}	3864.1 ^{ab}	2878 ^{bcd}	4733.13
G8	5596 ^{abcd}	5665 ^{abc}	4533 ^{defg}	1639.2 ^{bcde}	3694.4 ^{ab}	5063 ^{abc}	4365.10
G9	6058 ^{abcd}	5230 ^{abc}	7816 ^{abcd}	1612.5 ^{bcde}	3658.1 ^a	3211 ^{abcd}	4597.60
G10	3879 ^d	4167 ^{bc}	5872 ^{abcde^{f^g}}	1960.0 ^{abcde}	2447.2 ^{abc}	3893 ^{abcd}	3703.03
G11	8869 ^{ab}	5301 ^{abc}	6528 ^{abcde^f}	1573.3 ^{bcde}	3173.0 ^a	3074 ^{abcd}	4753.05
G12	4353 ^{cd}	4876 ^{abc}	8336 ^{abc}	2010.6 ^{abcd}	3675.1 ^a	4941 ^{abc}	4698.62
G13	6425 ^{abcd}	6093 ^{abc}	5126 ^{bcd^{efg}}	1277.0 ^{cde}	3887.6 ^{ab}	2737 ^{bcd}	4257.60
G14	8333 ^{abc}	5267 ^{abc}	6654 ^{abcde^f}	1917.5 ^{abcde}	3539.9 ^a	3296 ^{abcd}	4834.57
G15	4467 ^{cd}	5923 ^{abc}	4109 ^{efg}	1475.6 ^{bcde}	3916.1 ^{ab}	5678 ^a	4261.45
G16	7674 ^{abcd}	4350 ^{bc}	4543 ^{defg}	1787.7 ^{abcde}	3113.2 ^a	4485 ^{abcd}	4325.48
G17	8209 ^{abc}	4350 ^{bc}	5333 ^{bcd^{efg}}	1360.5 ^{bcde}	3665.4 ^a	4015 ^{abcd}	4488.82
G18	7658 ^{abcd}	5722 ^{abc}	6383 ^{abcde^f}	1158.5 ^{ed}	3415.7 ^a	4470 ^{abcd}	4801.20
G19	4556 ^{cd}	4706 ^{abc}	5291 ^{bcd^{efg}}	2234.1 ^{ab}	2585.8 ^{abc}	3141 ^{abcd}	3752.32
G20	4636 ^{cd}	4706 ^{abc}	4365 ^{efg}	1410.4 ^{bcde}	3416.4 ^a	4563 ^{abc}	3849.47
G21	5484 ^{abcd}	4765 ^{abc}	4230 ^{efg}	2020.7 ^{abcd}	3199.3 ^a	3300 ^{abcd}	3833.17
G22	6019 ^{abcd}	7274 ^a	6519 ^{abcde^f}	1583.5 ^{bcde}	3488.7 ^a	3530 ^{abcd}	4735.70
G23	7649 ^{abcd}	5101 ^{abc}	4285 ^{efg}	2013.8 ^{abcd}	3161.9 ^a	3093 ^{abcd}	4217.28
G24	4952 ^{bcd}	4303 ^{bc}	7467 ^{abcde}	1867.7 ^{abcde}	2579.5 ^{abc}	5270 ^{ab}	4406.53
G25	6015 ^{abcd}	4134 ^{bc}	5956 ^{abcde^f}	2136.3 ^{abc}	2985.3 ^{ab}	1778 ^d	3834.10
G26	5548 ^{abcd}	3534 ^c	5600 ^{bcd^{efg}}	1226.7 ^{cde}	2610.7 ^{abc}	3584 ^{abcd}	3683.90
G27	5168 ^{abcd}	4862 ^{abc}	6198 ^{abcde^f}	1459.3 ^{bcde}	4045.2 ^{ab}	4078 ^{abcd}	4301.75
G28	5342 ^{abcd}	5559 ^{abc}	5353 ^{bcd^{efg}}	2151.1 ^{abc}	3149.0 ^a	4007 ^{abcd}	4260.18
G29	4067 ^d	4683 ^{abc}	5084 ^{bcd^{efg}}	1828.5 ^{abcde}	3008.4 ^{ab}	2567 ^{bcd}	3539.65
G30	7212 ^{abcd}	3957 ^c	4312 ^{efg}	2638.8 ^a	2722.7 ^{ab}	2723 ^{bcd}	3927.58
G31	7339 ^{abcd}	5362 ^{abc}	6257 ^{abcde^f}	1860.7 ^{abcde}	3676.4 ^a	4211 ^{abcd}	4784.35
G32	6064 ^{abcd}	5849 ^{abc}	5481 ^{bcd^{efg}}	2088.1 ^{abcd}	3451.3 ^a	4080 ^{abcd}	4502.23
G33	5627 ^{abcd}	4548 ^{abc}	9022 ^a	1837.0 ^{abcde}	3316.1 ^a	4381 ^{abcd}	4788.52
G34	7669 ^{abcd}	5647 ^{abc}	5931 ^{abcde^f}	1148.1 ^{de}	3758.4 ^{ab}	4252 ^{abcd}	4734.25
G35	5644 ^{abcd}	4934 ^{abc}	5719 ^{abcde^f}	1983.8 ^{abcd}	3626.5 ^a	2489 ^{cd}	4066.05
G36	9137 ^a	3554 ^c	3531 ^{fg}	1902.9 ^{abcde}	3353.8 ^a	4022 ^{abcd}	4250.12
Mean	6182.31	5030.36	5602.17	1721.61	3270.66	3753.44	

G= genotype, G1-G36 (Genotypes one up to thirty six), a=highest, b=medium, c=poor, d=poorest, e-g=bad mean grain yield, genotype having same letters are same in mean yield.

3.3. Stability Analyses

Additive Main Effects and Multiplicative Interaction Analysis

The Additive Main Effects and Multiplicative Interaction (AMMI) analysis of variance for grain yield (kg/ha) and biomass (kg/ha) of 36 sorghum genotypes tested across six environments is presented in Table 4. The analysis revealed that genotypes had a significant impact on grain yield, while the environment ($p \leq 0.001$) also had a significant effect on grain yield. This finding aligns with a study by Jifar *et al.* [31] on tef genotypes, where they observed significant effects ($P \leq 0.01$) attributed to the environment, genotype, and their interaction ($G \times E$) on seed yield and yield components.

In this research, environmental factors explained the majority of the variance in grain yield (69.29%), with genotype-environment interaction (24.49%) and genotype (6.22%) following. The

impact of the environment was the most significant, while that of genotype was the least. The result aligns with earlier studies on sorghum [32,33]. Thus, the primary source of variation in grain yield was attributed to environments, suggesting their diversity and potential subdivision into mega-environments. Significant discrepancies among environments predominantly accounted for the variability in grain yield. Similar outcomes have been noted in other studies on sorghum [22,34], where environments displayed a higher sum of squares compared to genotypes.

The interaction between genotype and environment had a significant impact on grain and biomass. Two principal components were found to be the most precise predictors for both grain and biomass. These components explained 73.86% of grain yield variability and 88.57% of the total GEI sum of squares in biomass. These findings have been observed by several researchers across various locations and years [35,36]. Similar findings have been documented for sorghum genotypes assessed across various locations and years [3,37,32]. Moreover, the mean square of IPCA1 was higher than that of IPCA2, IPCA3, and IPCA4 for grain and biomass, indicating variations in genotype performance due to GEI. This aligns with previous studies on sorghum [3,33,38].

Table 4. Analysis of variance for the AMMI model for grain and biomass yield.

Source of variations	df	Grain yield			biomass		
		S.S.	Percent	Accumulated	MS	Percent	Accumulated
Genotype(G)	35	44594242*	6.22	6.22	1.99E+08	13.61	13.61
Environments(E)	5	496649374**	69.29	75.51	9.93E+08	67.78	81.39
Interactions(GxE)	175	175547472**	24.49	100.00	2.72E+08	18.61	100.00
PC1	39	75845622**	43.21	43.21	1.94E+08	71.07	71.07
PC2	37	53814007**	30.65	73.86	47685656	17.50	88.57
PC3	35	24616969**	14.02	87.88	25943186	9.52	98.10
PC4	33	15600190**	8.89	96.77	4233067	1.55	99.65
Residuals	31	5670685	3.23		953107	0.35	
Total	215	7.17E+08			1.46E+09		

*, **, significant at 5% & 1% probability level; ns, non-significant.

The Additive main effects and multiplicative interaction one (AMMI 1) biplot

AMMI 1 biplot shows genotype and environment interaction effects on yield [25]. The X-axis represents yield, while the Y-axis shows IPCA1 scores. It helps interpret interaction effects and evaluate adaptability (Figures 1 and 2). The study identified stable genotypes G21, G25, G13, G32, G35, G28, and G8, with G21 and G25 underperforming and G32, G8, G28, and G13 exhibiting high grain yield (Figure 1). For biomass, genotypes G20, G22, G5, G13, G30, G28, G35, G6, and G12 were stable but G22, G20, and G28 didn't perform well. Genotypes G11, G2, and G1 were generally adaptable to all environments, while G6, G15, and G29 exhibited specific adaptability. Yitayeh *et al.* [22] utilized this model to assess the yield stability of early maturing sorghum. Genotypes on the right of the perpendicular line midpoint had higher yields than those on the left. G13, G34, G14, G31, G18, G16, G7, G17, G11, and G1 had higher grain yields, while G26, G19, G20, G29, G5, and G3 had lower yields. G1, G11, G25, and G16 had higher biomass, while G19, G29, G12, G28, and G5 had lower yields (Figure 2).

In summary, Alduba, Jinka, and Kako were favorable testing locations while Arifyde, Gato, and Meioso were unfavorable. Genotypes G22 and G33 showed higher grain yields in Kako and Alduba. For biomass, Jinka and Kako were favorable while Arifyde, Gato, and Meioso were unfavorable. Locations far from the origin, such as Arifyde, Kako, and Jinka, played a crucial role in the genotype-environment interaction, affecting the stability of biomass performance. Genotypes G1 and 11 showed higher biomass at Jinka, indicating their adaptability to this location. Crossa *et al.* [25] also noted that Genotype and location combinations with IPCA1 scores of the same sign resulted in positive specific interaction effects, while combinations with opposite signs led to negative specific interactions.

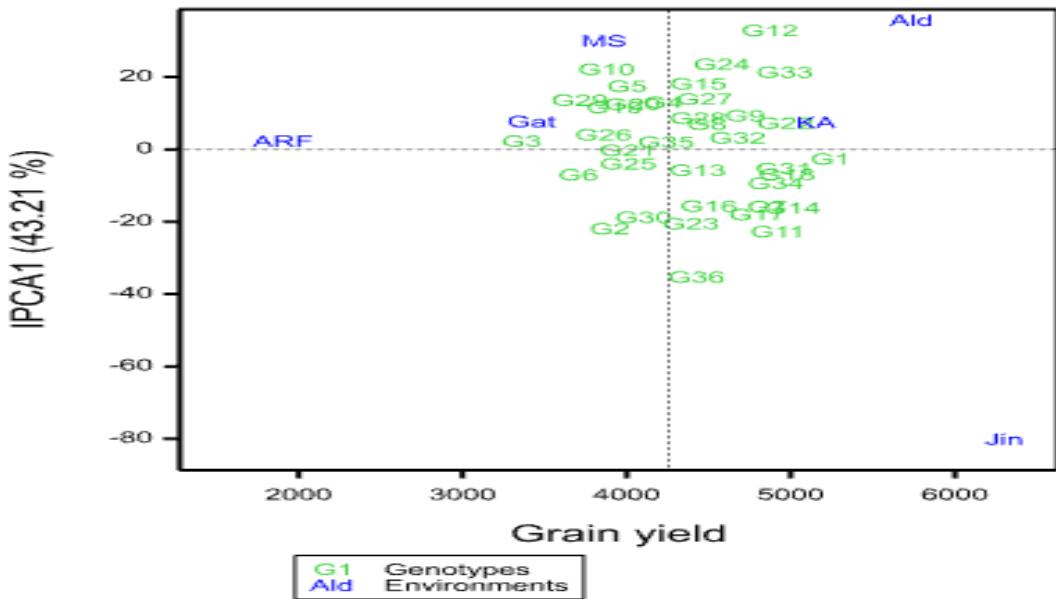


Figure 1. AMMI1 biplot of IPCA1 against grain yield for 36 sorghum genotypes tested across six locations (Jin:Jinka, KA:Kako, Ald:Alduba, ARF:Arifyde, Gat:Gato, MS:Meioso).

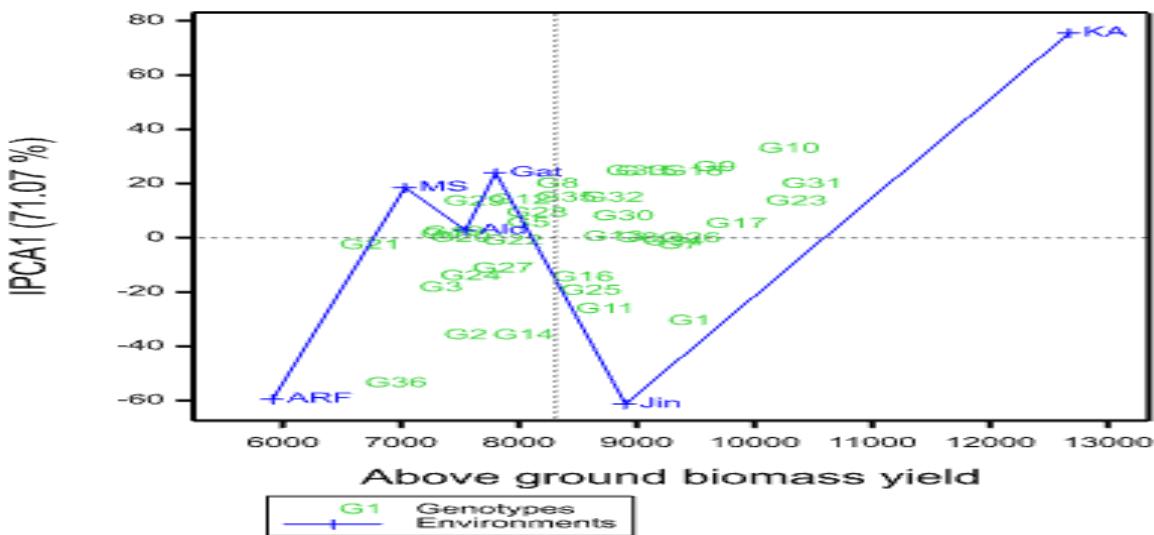


Figure 2. AMMI1 biplot of IPCA 1 against biomass for 36 sorghum genotypes tested across six locations.

3.4. AMMI Stability Value (ASV)

The Interaction Principal Component One (IPCA1) and Interaction Principal Component Two (IPCA2) scores in the AMMI model serve as stability indicators. The AMMI Stability Value (ASV) provides a balanced measure of stability [19]. Genotypes with lower ASV values are deemed stable, while those with higher ASV values are considered unstable. As per Appendix Table A1, G35 exhibited the highest stability with an ASV of 4.54, followed by G3 (6.60), G32 (7.51), G26 (8.29), G31 (9.17), G22 (9.57), and G18 (11.87) as the most unstable genotypes for grain yield. The stable genotypes (G31, G22, G32, and G18) showed mean grain yield above the grand mean, aligning with findings from Alemu *et al.* [39] and Yitayeh *et al.* [22] who utilized this stability parameter to assess sorghum genotypes' stability. Concerning biomass, genotype G26 (3.90) demonstrated the highest stability, followed by G4 (7.31), G13 (9.40), and G19 (10.90), while G36 (218.71), G2 (146.83), G14 (146.72), and G10 (133.48) were identified as the most unstable genotypes.

4. Conclusions

The combined analysis of variance showed significant variation among the sorghum genotypes, environments, and their interaction ($G \times E$). In the AMMI analysis, genotypes (G) and environments (E) contributed to 24.49% and 69.29% of the treatment sum of squares, respectively, whereas the $G \times E$ interaction accounted for 14.8%. The environment played a more significant role in the total sum of squares than genotypes. The two IPCAs validated the grain yield variation explained by genotype-environment interaction, explaining 73.86% of the interaction sum of squares. The AMMI1 and AMMI2 biplots identified the most stable genotypes for grain and biomass. For various stability measures, genotypes G18, G22, G31, and G32 were identified as the most stable genotypes for grain and biomass.

The study identified sorghum genotypes G18 and G31 as stable and high-yielding candidates for variety development programs in Ethiopia. These genotypes exhibit consistent performance across different environments and show promise for further enhancement in sorghum production.

Acknowledgements: This paper was supported by the Ministry of Education and Jinka University. We are grateful for their financial and logistical support.

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