

Article

Not peer-reviewed version

Comparative Analysis of Comprehensive Genomic Profile in Thymomas and Recurrent Thymomas Reveals Potentially Actionable Mutations for Target Therapies

[Filippo Lococo](#) * , [Elisa De Paolis](#) , Jessica Evangelista , [Andrea Dell'Amore](#) , Diana Giannarelli , [Marco Chiappetta](#) , Annalisa Campanella , [Carolina SASSOROSSI](#) , Alessandra Cancellieri , [Fiorella Calabrese](#) , Emanuele Vita , [Angelo Minucci](#) , [Emilio Bria](#) , [Angelo Castello](#) , [Andrea Urbani](#) , [Federico Rea](#) , [Stefano Margaritora](#) , [Giovanni Scambia](#)

Posted Date: 17 July 2024

doi: 10.20944/preprints2024071356.v1

Keywords: NGS; recurrent thymoma; surgery; CGP; gene profile; clinical trials



Preprints.org is a free multidiscipline platform providing preprint service that is dedicated to making early versions of research outputs permanently available and citable. Preprints posted at Preprints.org appear in Web of Science, Crossref, Google Scholar, Scilit, Europe PMC.

Copyright: This is an open access article distributed under the Creative Commons Attribution License which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Disclaimer/Publisher's Note: The statements, opinions, and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions, or products referred to in the content.

Article

Comparative Analysis of Comprehensive Genomic Profile in Thymomas and Recurrent Thymomas Reveals Potentially Actionable Mutations for Target Therapies

Filippo Lococo ^{1,2,*}, Elisa De Paolis ^{3,4,†}, Jessica Evangelista ^{1,2,‡}, Andrea Dell'Amore ⁵, Diana Giannarelli ⁶, Marco Chiappetta ², Annalisa Campanella ², Carolina Sassorossi ², Alessandra Cancellieri ⁷, Fiorella Calabrese ⁸, Emanuele Vita ⁹, Angelo Minucci ⁸, Emilio Bria ^{9,10}, Angelo Castello ¹¹, Andrea Urbani ³, Federico Rea ⁵, Stefano Margaritora ^{1,2,‡} and Giovanni Scambia ^{12,‡}

¹ Università Cattolica del Sacro Cuore, 00168 Rome, Italy; jessica.evangelista@policlinicogemelli.it (J.E.); stefano.margaritora@policlinicogemelli.it (S.M.)

² Thoracic Surgery, Fondazione Policlinico Universitario A. Gemelli IRCCS, 00168 Rome, Italy; marco.chiappetta@policlinicogemelli.it (M.C.); annalisa.campanella@guest.policlinicogemelli.it (A.C.); carolina.sassorossi@guest.policlinicogemelli.it (C.S.)

³ Clinical Chemistry, Biochemistry and Molecular Biology Operations (UOC), Fondazione Policlinico Universitario A. Gemelli IRCCS, Rome, Italy; elisa.depaolis@policlinicogemelli.it (E.D.); andrea.urbani@policlinicogemelli.it (A.U.);

⁴ Departmental Unit of Molecular and Genomic Diagnostics, Genomics Core Facility, Gemelli Science and Technology Park (G-STeP), Fondazione Policlinico Universitario A. Gemelli IRCCS, Rome, Italy

⁵ Thoracic Surgery Unit, Department of Cardiac, Thoracic, Vascular Sciences and Public Health, University of Padova, Padova, Italy; andrea.dellamore@aopd.veneto.it (A.D.); federico.rea@unipd.it (F.R.)

⁶ Epidemiology and Biostatistics Facility, Gemelli Science and Technology Park (G-STeP), Fondazione Policlinico Universitario A. Gemelli IRCCS, 00168 Rome, Italy; diana.giannarelli@policlinicogemelli.it

⁷ Unit of Pathology, Fondazione Policlinico Gemelli IRCCS, Rome, Italy; alessandra.cancellieri@policlinicogemelli.it

⁸ Pathology Unit, Department of Cardiac, Thoracic, Vascular Sciences and Public Health, University of Padova, Padova, Italy; fiorella.calabrese@unipd.it (F.C.); angelo.minucci@policlinicogemelli.it (A.M.)

⁹ UOSD Oncologia Toraco-Polmonare, Comprehensive Cancer Center, Fondazione Policlinico Universitario Ago-stino Gemelli IRCCS, Rome, Italy; Medical Oncology, Department of Traslational Medicine and Surgery, Università Cattolica del Sacro Cuore, Rome, Italy; emanuele.vita@guest.policlinicogemelli.it (E.V.); emilio.bria@policlinicogemelli.it (E.B.);

¹⁰ UOC Oncologia Medica, Ospedale Isola Tiberina – Gemelli Isola, Roma, Italy

¹¹ Nuclear Medicine Department, Fondazione IRCCS Ca' Granda Ospedale Maggiore Policlinico, 20122 Milan, Italy; angelo.castello@policlinico.mi.it

¹² Division of Gynecologic Oncology, Fondazione Policlinico Universitario A. Gemelli-IRCCS, Rome, Italy; giovanni.scambia@policlinicogemelli.it

* Correspondence: filippo.lococo@policlinicogemelli.it; Tel.: +0039-06-356353

† Authors equally contribute to the paper.

‡ Authors equally contribute to the paper.

Abstract: Objective: Molecular profiles of thymomas and recurrent thymomas are far to be defined. Herein, we report analysis of comprehensive genetic profile (CGP) in a highly-selected cohort of recurrent thymomas.

Methods: Among a cohort of 426 thymomas, the tissue was available in 23 recurrent tumors for matching biomolecular results obtained from primary and relapses samples. A control-group composed by non-recurrent thymoma patients was selected by propensity-score match analysis. CGP was performed using the NGS TruSightOncology assay to evaluate TMB, MSI, and molecular alterations in 523 genes. Genomic results analysis was done according to clinical practice guidelines adopting the Tier classification system. Statistical association among different molecular pathways and clinical characteristics were assessed through the chi-square test.

Results: CGP does not change when comparing initial tumor with tumor relapse. Significantly higher



frequency of cell-cycle control genes alterations (100.0% vs 57.1%, $p=0.022$) are detected in patients with early-recurrence (<32 months) compared to late-recurrent cases. Conversely, alterations of DNA-repair genes were more frequently observed in early-stage Masaoka tumors ($p=0.019$). CGP were similar in recurrent thymomas and non-recurrent thymomas. Finally, based on NGS-results, an off-label treatment or clinical trial could be potentially proposed in >50% of cases (oncogenic Tier-IIC variants). **Conclusions:** CGP do not substantially differ between initial tumor vs tumor recurrence and recurrent thymomas vs non-recurrent thymomas. Cell-cycle control genes alterations are associated with an early recurrence after thymectomy. Multiple target therapies are potentially available by performing a comprehensive CGP, suggesting that a precision medicine approach on these patients could be further explored.

Keywords: NGS; recurrent thymoma; surgery; CGP; gene profile; clinical trials

1. Introduction

Thymomas are relatively rare tumors of epithelial thymic cells representing approximately 0.2–1.5% of all malignancies [1]. From a pathological point of view, the World Health Organization distinguish them into different types (so-called A, AB, B1, B2, and B3) based upon the relative proportion of the non-tumoral lymphocytic component, and the resemblance to normal thymic architecture [2].

Despite these tumors presented usually with an indolent behavior, the natural history is often unpredictable with recurrences reported to occur in 10–30% of patients even after 10 to 20 years [3,4] after radical resection (R0).

Tumor recurrences are generally located in the thorax (mostly in the pleural cavity) and are usually treated by loco-regional approach (combined or not with systemic treatment) with surgery staying as the gold standard approach when technically feasible [4].

Indeed, several studies [5–7] and meta-analysis [8] reported improved early- and long-term outcomes after surgery in recurrent thymoma patients, whereas few Authors support chemotherapy only (usually platinum-based protocols) in this setting [9,10].

Unfortunately, the clinical history of these neoplasms is very insidious: in fact, even after re-do surgery further recurrences of disease are very frequently reported [4,7]; similarly, in recurrent cases who underwent 1st line treatment, a disease progression is quite common and further lines of therapy are not standardized and become generally much less effective. Consequently, there is an urgent need for novel treatments for recurrent and platinum-resistant thymomas.

In the last decade, the wide implementation of high throughput technologies and Comprehensive Genomic Profiling (CGP) in solid tumors have allowed the identification of a broad spectrum of molecular aberrations and altered signaling pathways in TETs, leading to the definition of distinct molecular profiles in TETs. Several attempts to identify somatic mutations that characterize TETs have been made in recent years. Target-specific drugs for TETs have not been developed because the genomic aberrations in TETs are poorly understood [11].

Several studies have generally explored thymomas and thymic carcinoma in the same dataset [11,12] clearly demonstrating different biological aspects in terms of tumor mutational burden (TMB), microsatellite instability (MSI) status and molecular pathways. In particular, TMB has reported to be much higher in thymic carcinoma [11–13], this stays as a predictive factor of immune check-point inhibitors (ICI) efficacy. A recent meta-analysis [14] suggest that ICI could be a therapeutic option for selected patients with thymic carcinoma that are not amenable to curative radical treatment after first-line chemotherapy. On the other hand, no impressive changes in therapeutic paradigm of unresectable/recurrent thymomas progressed to platinum-based chemotherapy have been achieved so far.

In this framework, we reviewed a large cohort of surgically resected thymomas, performing a CGP on the surgical specimen of both primary and recurrent thymomas. A control group of non-recurrent thymomas was also selected (propensity-score match analysis) and their gene profiles also analyzed. The final aims of the present study were:

- To compare the CGP of recurrent thymoma patients vs non-recurrent thymoma patients;
- To explore the CGP of both primary and recurrent thymomas and identify associations with clinic-pathological variables;
- To evaluate actionable mutations detected in thymomas as target for new therapeutic approaches.

2. Materials and Methods

2.1. Study Design and Selection of Cases

This bicentric, observational, retrospective, cohort study was reviewed and approved by the Fondazione Policlinico Universitario “Agostino Gemelli” IRCCS, ethics committee and partner institutional review boards (study identification: 3027). Data on patients treated for thymoma recurrence from January 1, 2003, to January 1, 2023, in two high-volume centers were collected and retrospectively reviewed. The two centers were selected because they had high-volume, long-term experience and similar management in thymoma and recurrent thymoma patients. This study was conducted based on an overall surgical cohort of 426 TETs. Thymic carcinomas and neuroendocrine thymic tumors were excluded from the analysis, because of their biological and molecular differences from thymomas. After excluding cases with missing data, we selected patients who experienced a recurrence during follow-up (see Consort Diagram, Figure 1). A control-group was made by propensity score match analysis being composed by thymoma patients who did not develop any recurrence during a minimum of 5 yrs after initial surgery. A total of 43 patients experienced a thymoma recurrence after thymectomy (Overall Relapse Rate: 13.65%). Tissue was available for the analysis in 23 of them, these composing the Recurrent Thymomas Group (Rec_Thy). In these patients both primary and relapses tumor was analyzed and compared. Finally, 23 thymoma patients who have not reported any recurrence at least 5 years after thymectomy was identified from the entire surgical cohort of cases thanks to a propensity score match analysis (as reported below). Tissue was available in 14 cases, these composing the control group of Non-Recurrent Thymoma (NoRec_Thy).

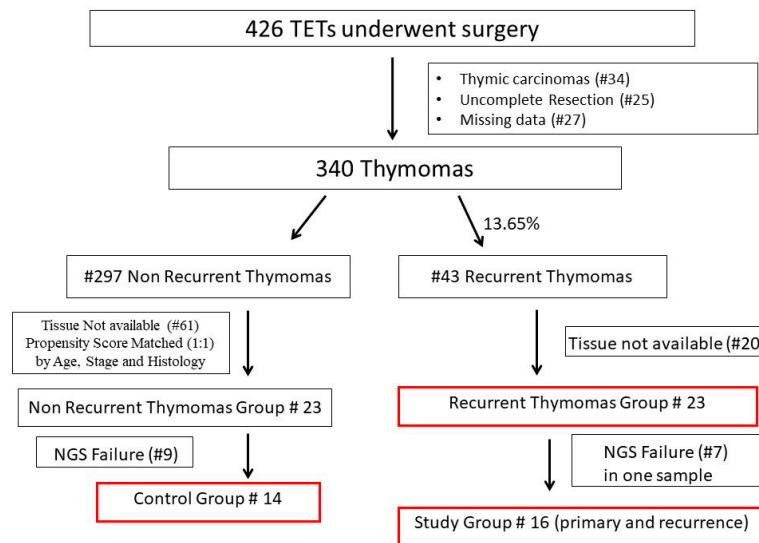


Figure 1. Consort Diagram of the Study Population.

2.2. Pathological Review

Thymomas were classified using the Masaoka-Koga staging classification [15], the eighth edition of the TNM staging system for thymic neoplasm [16] and the WHO classification system for TETs [17]. They were also organized into 2 groups based on a previously reported prediction model of recurrence [18]. On the basis of this model, patients with T1-T2 thymomas or T3 type A-AB-B1

thymomas had a significantly lower incidence of recurrence ("low-risk group") than those with T2-T3 type B2-B3 thymomas ("high-risk group"). All patients underwent surgery for both primary and recurrent thymoma with curative intent. We excluded patients with only radiological suspect of thymoma recurrence or with pathological confirmation achieved by small biopsy. Dedicated pathologist in each involved center (AC,FC) reviewed all specimens and re-evaluated histology according to the WHO-classification system. A centralized revision at the promoting center was performed in cases with doubts.

2.3. Comprehensive Genomic Profiling and Bioinformatics Analysis

Eosin-stained histology tissue slides were examined by dedicated pathologists to identify areas of at least 20% of tumor cells content. DNA was extracted using the AllPrep® DNA/RNA FFPE commercial kit (QI-AGEN®), according to manufacturer's procedures. Nucleic acids quality was assessed by using the Illumina Infinium FFPE QC kit (Illumina®) on the CFX Connect Real-Time PCR Detection System instrument (Bio-Rad®). The DNA quantitation was performed using the Qubit HS dsDNA fluorimetric assays (Life Technologies®) and only samples with a quantitation greater than 40 ng were analysed.

A pan-cancer CGP was performed using the TruSight Oncology 500 High-Throughput (TSO500HT) assay. TSO500HT allows identification of low-frequency somatic variants as Single Nucleotide Variants (SNVs), Insertions and Deletions (indels), splice variants, and Copy Number Alterations (CNVs, i.e. gain/amplification) in 523 genes related to cancer susceptibility and treatment, along with the major immunotherapy biomarkers (TMB, MSI). Genomic DNA was sheared and converted into libraries with addition of Unique Molecular Identifiers (UMIs). The NGS was performed on the NovaSeq6000 platform (Illumina). Output data evaluation was obtained using VELSERA Clinical Genomics Workspace tool and only genomic profiling characterized by a sequencing data with a median depth of coverage > 500X was considered in the final review of molecular results. A cut-off of 5% of Variant Allele Frequency (VAF%) was adopted. MSI was calculated from 130 loci.

Interpretation and final reporting of the detected variants were performed according to expertly curated genomic databases, clinical practice guidelines, FDA therapeutics indications, clinical trials availability, and medical interpretations. Tier classification system of the Association for Medical Pathology, the American Society of Clinical Oncology, and the College of American Pathologists was adopted [19]. Only molecular alterations predicted to be oncogenic/likely oncogenic were evaluated (Tier I-II), based on the annotations in mutational databases as COSMIC [20], OncoKb [21], ClinVar [22], or with pertinent literature evidences. Molecular alterations were considered as clinically relevant (Tier IIC) if targetable by drugs available in different clinical contexts or if represent enrollment criteria in a registered clinical trial for the specific clinical context. Variants of unknown significance (Tier III) were excluded. The ESMO Precision Medicine Working Group recommendations were considered for a follow-up germline target test according to annotations in germline mutational databases, types of alteration, and VAF% [23].

2.4. Statistical Analysis

As a first step, a propensity score approach was used to select the control group among patients without recurrence; with this method we identified a subgroup of patients without recurrence within 5 years after surgery to undergo CGP. The propensity was based on the nearest neighbour method with a caliper of 1.5 standard deviations considering sex, age, presence of Myasthenia Gravis, Masaoka-Koga staging and histology.

As second step, comparing CGP on primary tumors and recurrence on the same patient a paired approach was implemented: kappa statistics was used to measure concordance in the presence of alterations and the McNemar test was calculated to assess marginal homogeneity.

As an overall approach, data were summarized using absolute counts and percentages for categorical items and median and range when referring to quantitative variables. Association among

different pathways and clinical and demographical characteristics were assessed through the chi-square test. IBM-SPSS v.28.0 and R v.4.1.2 softwares were used for analysis.

3. Results

3.1. Clinical and Pathological Characteristics

The main patient's characteristics and pathological features of Rec_Thy Group and NoRec_Thy are summarized in Table 1. In details, Rec_Thy patients were relative young (median age = 51 yrs) and presented a recurrence several months after thymectomy (median disease-free interval -DFI- of 32 months). They had mostly Masaoka Stage II-III Type-B thymoma and were often treated with neoadjuvant therapy before surgery. These variables were balanced in the control group (NoRec_Thy) with similar distribution of age, Masaoka Stage and histology (see Table 1). According to the classification reported above, thymomas were classified at high-risk in 75% of Rec_Thy and 64.3% of NoRec_Thy.

Table 1. The main patient's characteristics and pathological features of Rec_Thy Group and NoRec_Thy.

	Rec_Thy (n = 23 pts)	No Rec_Thy (n=14 pts)
GENDER		
M	13 (56.5%)	10 (71.4%)
F	10 (43.5%)	4 (28.6%)
AGE (median, range)	51y (27y-83y)	59y (16y-82y)
MG	8 (34.8%)	7 (50.0%)
MASAOKA*		
II	5 (21.8%)	5 (35.7%)
III	12 (52.2%)	7 (52.2%)
IV	3 (13.0%)	2 (14.3%)
NEOADJUVANT TREATMENT	10/20 (50.0%)	6/14 (42.9%)
HISTOLOGY WHO		
AB	0 (0%)	1 (7.1%)
B1	9 (39.2%)	2 (14.3%)
B2	7 (30.4%)	9 (64.3%)
B3	7 (30.4%)	2 (14.3%)
^RISK CLASS*		
Low-Risk	5 (25.0%)	5 (35.7%)
High-Risk	15 (75.0%)	9 (64.3%)
STAGE*		
II	4 (20.0%)	6 (42.8%)
III	11 (55.0%)	5 (35.7%)
IV	5 (25.0%)	3 (21.5%)
DFI (median, range)**	32m (6m-132m)	/
ADJUVANT TREATMENT	7 (30.0%)	7 (50.0%)

*only on primary tumor; ** DFI= Time between thymectomy and relapse (months); ^ according to criteria reported in [18].

3.2. Overall Genomic Results (Entire Cohort)

Globally, most patients showed reportable oncogenic/likely oncogenic molecular alterations, observed in 81% of cases, with a low rate of oncogenic mutations/case (min: 0 - max: 6) and with several genes appeared only once in the cohort (Figure 2).

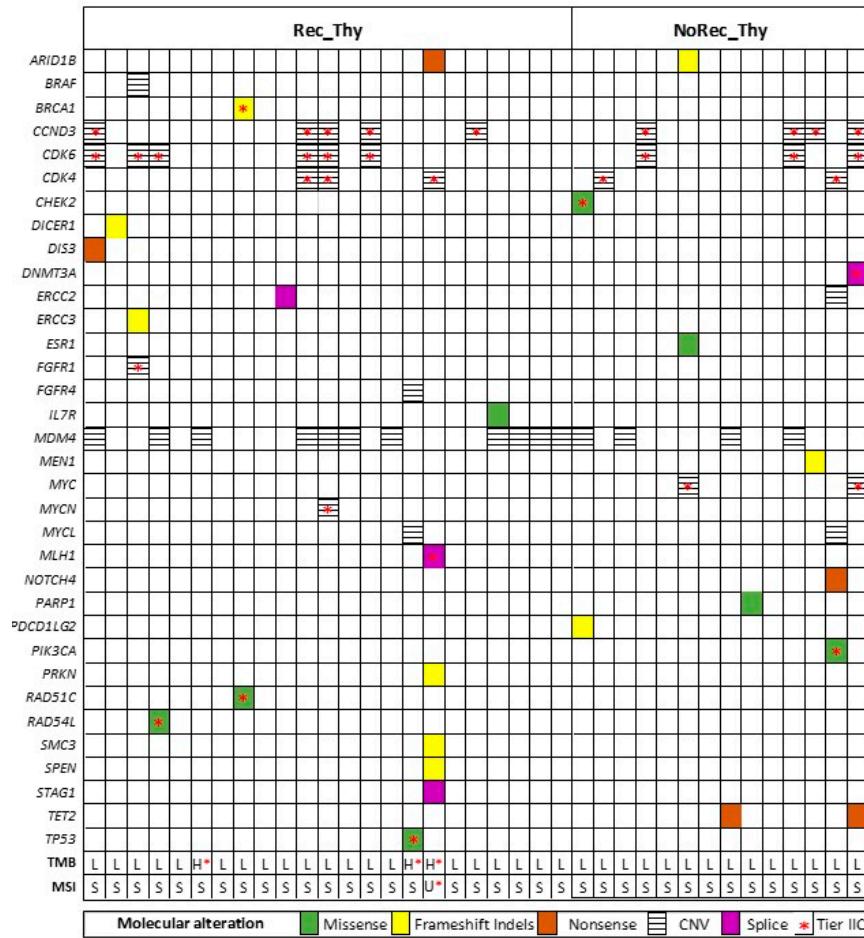


Figure 2. CGP profile of Thymomas. The figure shows the molecular alterations and the genomic signatures identified in the cohorts of Rec_Thy and NoRec_Thy.

Recurrent defective pathways were identified. Molecular alterations accounted on genes involved in cell cycle resulted as recurrent in this study, with amplifications in CCND3 (16%), CDK4/6 (27%), and MDM4 (32%) genes as the most involved.

Alterations in DNA damage repair (DDR) pathways including Homologous Recombination (HR), Nucleotide Excision Repair (NER), and Mismatch Repair (MMR) were identified. In particular, Loss-of-Function (LoF) SNVs in DDR were identified in 22% of patients, without any recurrently mutated gene. Among HR, we identified LoF mutations in BRCA1, RAD51C, RAD54L, and CHEK2. One patient resulted as carrier of MLH1 LoF oncogenic mutation, with a predicted impairment of MMR system.

Other dysregulated pathways included RTK family signalling (with FGFR1/4 amplifications in 5% of patients) and PI3K/AKT/mTOR activation (ESR1 and PIK3CA genes in 5% of patients).

Amplifications in MYC oncprotein family (MYC, MYCL, MYCN genes) was identified in 10% of patients. In addition, alterations in epigenetic regulatory genes as TET2 and DNMT3A were rarely identified in the cohort (8% of patients). TP53 oncogenic variant was identified in one case of our series (3%).

TMB status resulted as low across all samples, together with MSI stable status (i.e. MMR-proficient). In only one patient, we observed a high TMB, probably related to MLH1 mutation that could lead to the accumulation of somatic frameshift and SNVs. For this patient we were not able to

calculate the MSI status (failure to cover the 130 MSI sites). According to ESMO guidelines, follow up-germline testing was not recommended for the enrolled patients.

3.3. CGP Differences in Recurrent Thymoma vs Non Recurrent Thymoma

From the comparative evaluation of the two study groups of Rec_Thy and NoRec_Thy, no overall significant differences emerged in the molecular analysis (Table 2). Oncogenic alterations was reported in 83% of recurrent thymomas vs 78% of non-recurrent thymomas ($p=0.76$). The rate of clinically relevant alterations (Tier-IIC) is similar in the two groups, with 43% of recurrent thymomas vs 57% of non-recurrent thymomas. Looking into the distribution and types of oncogenic alterations, the same percentage of cases with dysregulation of the two main pathways of cell cycle (74% in recurrent vs 64% in non-recurrent) and DDR (22% in recurrent vs 21% in non-recurrent) was identified. Even if accounted in a limited number of cases, dysregulations in epigenetic regulatory genes and PI3K/AKT pathway genes were identified only in the non-recurrent group of thymomas (14%). On the contrary, alterations in RTK-RAS family signalling cascade were detected only in recurrent thymomas (FGFR1/4, BRAF) (13%). No differences in MSI and TMB status were identified in the two groups.

Table 2. Distribution of genetic alterations between Recurrent thymomas and Non-Recurrent Thymomas.

GROUP	All Patients (#37)	Rec_Thy (#23)	NoRec_Thy (#14)	p-value
Pathway cell cycle	26 (70%)	17 (73.9%)	9 (64.3%)	$p=0.53$
Pathway DNA repair	8 (22%)	5 (21.7%)	3 (21.4%)	$p=0.98$
At least 1 alteration	30 (81%)	19 (82.6%)	11 (78.6%)	$p=0.76$
Clinically relevant alteration	18 (49%)	10 (43%)	8 (57%)	$p=0.83$

3.4. CGP Differences in Primary vs Recurrent Thymoma and Inter-Relationship with Clinic-Pathological Variables

No significant differences in CGP emerged from the comparative evaluation of matched primary and recurrence tissue biopsies. As reported in Table 3, similar frequencies of samples with at least one oncogenic/likely oncogenic alteration were observed when comparing primary thymomas and recurrent thymomas (Kappa statistics -0.049 $p=0.84$; McNemar $p=0.73$).

In details, genes belonging to cell cycle pathway were similarly altered in both primary (37%) and their recurrences (50%) (Kappa statistics -0.09 $p=0.30$, McNemar $p=0.69$). Comparable results were obtained evaluating the distribution of DNA damage repair alterations, occurring at 19% of primary tumor and 12% at matched recurrences (Kappa statistics 0.29 $p=0.23$, McNemar $p=0.99$). TMB was low in both primary thymomas and their recurrences, with no remarkable modification between samples (data not shown).

Table 3. Distribution of genetic alterations between Primary Thymomas and Recurrent Thymomas.

GROUP	Primary_Thy	Recurrent_Thy	p-value
Pathway cell cycle	6 (37.5 %)	9 (56.2%)	$p=0.30$
Pathway DNA repair	2 (12.5%)	3 (18.7%)	$p=0.23$
At least 1 alteration	9 (56.2%)	11 (68.7%)	$p=0.84$

On the contrary, when evaluating the distribution (see Table 4) of at least one genomic alteration in Rec_Thy with early-recurrence (DFI<32 months) we found a higher proportion of samples with at least 1 mutation compared to Rec_Thy with DFI>32 months others (100% vs 71.4%, $p=0.082$).

More interestingly, more cell-cycle control genes alterations were observed in early-recurrence Rec_Thy compared with others (100.0% vs 57.1%, $p=0.022$) while a similar distribution of alteration of gene of DNA-repair (25% vs 25%, $p=0.99$) was found.

Finally, by exploring the associations between other clinical variables and gene mutations, we observed a significantly higher frequency of genetic alteration in DNA-repair pathways in early

Masaoka-Stage tumors (see Table 4) while similar gene profile distribution was found according to age, presence of M.G., histology and classes of risk.

Table 4. Inter-relationship between clinic-pathological variables and gene mutations in Recurrent Thymoma.

	Pathway cell cycle	Pathway DNA repair	At least 1 alteration
Rec_Thy (n=23)	11/23 (47.8%)	3/23 (13.0%)	14/23 (60.9%)
Masaoka Stage		p=0.019	
II (n=5)	p=0.121 5/5 (100.0%)	3/5 (60.0%)	5/5 (100.0%)
III-IV (n=18)	12/18 (66.6%)	2/18 (11.1%)	14/18 (77.8%)
Age		p=0.635	
<51 (n=11)	p=0.896 5/11 (45.6%)	1/11 (9.1%)	8/11 (72.7%)
>51 (n=12)	6/12 (50.0%)	2/12 (16.7%)	6/12 (50.0%)
Miastenia Gravis		p=0.960	
Yes (n=8)	p=0.661 3/8 (37.5%)	1/8 (12.5%)	4/8 (50.0%)
No (n=15)	8/15 (53.3%)	2/15 (13.3%)	10/15 (66.7%)
RISK Class*		p=0.770	
Low (n=5)	p=0.121 4/5 (80.0%)	1/5 (20.0%)	5/5 (100.0%)
High (n=18)	9/18 (50.0%)	4/18 (22.2%)	11/18 (61.1%)
DFI		p=0.960	
<32 months (n=9)	p=0.022 9/9 (100.0%)	2/9 (22.2%)	9/9 (100.0%)
>32 months (n=14)	8/14 (57.1%)	3/14 (21.4%)	10/14 (71.4%)

Risk Classes as defined in [18].

3.5. Actionable Mutations for New Therapeutic Approaches

Overall, based on CGP profiling, off-label treatments approved in different disease entities or clinical trials potentially recruiting patients with mutated TETs has been identified. To note, no directly actionable genomic alterations (classifiable as Tier I) could be identified in our patients due to the lack of FDA/EMA approved molecular target therapy in thymoma clinical setting. Looking into the global actionability, approved treatments or clinical trials could be potentially recommended for 49% of analysed patients (18 out of 37). Supplementary Table 1 showed clinical trials potentially including thymomas in which the molecular characterization of tissue sample and the presence of a specific biomarker represent an enrollment criterion. Approved or experimental therapies mainly encompass Cyclin-Dependent Kinase (CDKs) inhibitors, PARP (Poly ADP-ribose) inhibitors, and Tyrosine Kinases (TKs) inhibitors.

4. Discussion

In this study, we took advantage of a robust series of recurrent TETs for which comprehensive GCP was conducted and compared with a control group of non-recurrent thymomas. Taken together, our results provide with a unique insight into molecular pathways activated in recurrent thymomas, paving the way for precision medicine approaches using targeted agents or experimental drugs in a large part of them. To our knowledge, our cohort is the largest reported so far, focusing on recurrent thymoma, this representing a specific subset of thymomas where the standard of care is still a matter of debate.

Despite recent evidences [8] promotes the role of surgical treatment for recurrent thymomas, the high rate of re-recurrences [3–5,24] suggests that surgery alone could fail to achieve a complete control of disease at this stage. On the other side, systemic treatment including immune check point inhibitors (ICI) [25,26] or somatostatin-receptor-targeting therapies (alone or with prednisone) [26] showed controversial results.

As a consequence of this, at today the strategy of care in recurrent thymomas remains an intriguing issue where exploring the role of molecular-targeted strategies after/prior to surgical resection.

In the present study, CGP data confirm a relatively low mutational burden, as emerged from literature [11–13,28–30]. Most studies highlight a limited number of molecular alterations, with no gene found to be mutated with a frequency exceeding 10% [11–13,28,29].

This may in part explain the paucity of effective molecular-target therapy. Literature data regarding pre-clinical and clinical evaluation of target drugs in TETs showed attractive results mainly in TC context [31].

Looking into the global actionability of our molecular findings, approved treatments or clinical trials could be potentially recommended for almost 49% of thymoma patients analysed herein. Similar data emerged from the EORTC-SPECTA/Arcagen study for rare tumours (53.8%) [28] and a lower percentage (27%) from the SPECTRALung platform [29].

The recommendations mainly encompassed CDKs inhibitors, PARPi, RTK inhibitors, and PI3K/mTOR inhibitors. Loss of cell cycle control emerged as a common occurrence in thymomas [29,31,32] and the most recurrent in our cohort (27%). Targeting D-type Cyclins in tumours expressing amplified CDK4/6 and CCND3 is widely investigated in solid and haematological malignancies (see Supplementary Table 1). A growing number of CDKs inhibitors are currently tested in clinical trials enrolling advanced/recurrent solid tumors as pan-CDKs inhibitors or more selective CDKs inhibitors.

Palbociclib, Ribociclib, and Abemaciclib are FDA-approved for hormone receptor-positive (HR+) breast cancer treatment. For patients with TETs, the utility of Palbociclib and Milciclib maleate CDKs inhibitor (PHA-848125AC) are under investigation in the phase II (NCT03219554 and NCT01301391 trials, respectively. Pre-clinical and phase-I supporting studies highlighted that in thymomas the negative expressions of p21 and p27 (natural inhibitors of CDKs) significantly correlates with poor prognosis for disease-free survival [33] and objective partial response type B3 and C thymic malignancies [34].

Interestingly in the present CGP analysis we found a significant higher alterations rate of Cyclin-Group genes in patients who experienced an early-recurrence compared with others (100.0% vs 57.1%, $p=0.022$), this suggesting a potential link between these genes and the biological aggressiveness in thymomas.

Moreover, CDK4/6 pathway hyper-activation are associated with worse prognosis in TC [35]. It is known that many other proteins interact with CDK4/6 and modulate the cell-cycle, as MDM2/MDM4 and TP53. TP53 mutation has been reported approximately in 3% of Thymomas as also identified in the present study [31,36]. MDM4 is significantly amplified (14% up to 43%) in several cancers types [37]. Here we identified MDM4 alterations in similar percentage (32%).

Additionally, DDR pathways alteration was reported in the 22% of patients. We not identified recurrent mutated targets in this subset. Defects in HRD pathway represent the molecular basis of synthetic lethality of PARP inhibition and FDA/EMA approved drugs are available in different settings (Olaparib, Talazoparib, Rucaparib). The role of DDR was largely unexplored in TETs. Few literature observations, mainly BRCA1/2 and ATM, are available regarding single case or families with sporadic/recurrent thymomas [38–40]. Among these, a patient with BRCA2-mutated thymoma showed a significant clinical benefit from treatment with Olaparib, with imaging showing overall stabilization of her disease [41].

Recommendations also encompassed TK-inhibitors. Experimental and clinical data regarding the potential role of VEGFR1/3 and FGFR1/4 driven angiogenesis dysregulation in TETs was also assumed [42]. The pan-RTK inhibitor Sunitinib is currently in NCCN guidelines for treatment of advanced TC and under investigation in a phase II clinical trial enrolling TC and thymomas patients [43]. Additionally, the NCT02307500 clinical trial evaluating the multikinase inhibitor Regorafenib is active for thymoma B2/3 patients in progression after chemotherapy.

Finally, alterations in PI3K/AKT/mTOR pathway are present in 5% of cases in our series, according to the TGCA PanCancer Atlas. Pre-clinical data suggested that subsets of thymomas activate the PI3K pathway through upregulation of a large microRNA cluster on chr19q13.42 with a marked reduction of cell viability [44]. In this context, the insulin-like growth factor-1 receptor (IGF1R) inhibitors cixutumumab and the mTOR inhibitor everolimus were investigated with a partial

response (NCT00965250 and NCT02049047, respectively) [45,46]. Everolimus is in NCCN guidelines for the treatment of thymomas and TC progressed after chemotherapy. Modest activity of the buparlisib, an oral pan-PI3K inhibitor, in relapsed or refractory thymomas, resulted from the NCT02220855 clinical trial [47]. These studies provide evidences to support further evaluation of PI3K/Akt pathway targeting in patients with advanced thymoma.

Mutations in epigenetic regulatory genes as DNMT3A was reported in ~7% of thymomas in the AACR GENIE cohort (<https://genie.cbiportal.org/>) and in 10% of TETs in literature (together with TET genes alterations) [41]. From our data, 3 patients belonging from the No_Rec groups resulted as carrier of alterations in DNMT3A and TET2.

Refractory or recurrent TETs should not be integrated into clinical trials, mainly due to the rarity of the disease. Advantages in CGP adoption also relies in the possibility to access to large clinical trial designed to adopt the best target therapy according to genetic alterations (e.g. CUSTOM trial NCT01306045, NCT05667948, and NCT01385722).

Limitations, Points of Strength and Future Clinical Applications

This study presents some limitations both concerning the selection of cases and the methodology of the analysis. Firstly, thymoma patients have been selected in a relative long interval (>20 years) in a retrospective study. However, thymomas are almost rare tumors and recurrences are uncommon and usually occurring several years after surgery. Thus, a long observation time may be necessary to have acceptable number of cases and enough follow-up to perform survival analyses. Moreover, despite this is in our knowledge the study with the largest number of recurrent thymomas patients with GCP analysis, the sample size limited the generalization of our results that needed more cases to be confirmed. Concerning the methodology of analysis, FFPE specimens aged > 10 years from the present analysis, results in an almost high rate of extraction (low DNA sample quality) or sequencing (NGS metrics quality) failures (see Figure-1). To note, we adopted one of the largest CGP panels available to allow a wide molecular investigation. However, we cannot replicate some of the TETs molecular data previously available due to specific analytical characteristics of the sequencing solution adopted (e.g. lack of analytical validation for copy number loss, absence of CTF2I gene in the panel).

At the same time, we would like to enhance the points of strength of the present study. Firstly, the novelty of the topic analysed in the study that is an emerging and unexplored issue for future research and clinical applications. Indeed, since we showed as up to 50% of our recurrent thymoma patients presented with TIER II-C molecular alterations, these results opening some opportunities for innovative molecular-targeted strategies in this setting.

Moreover, considering that prospective studies on thymomas are clearly not feasible, the adoption of a comparative group of NoRec_Thy selected by a propensity-score match analysis from our real-world Institutional cohort of patients clearly stays as an added value.

Finally, the bioinformatics and analytical analysis of sequencing findings as here described, represent a state-of-art approach for clinical translational studies. The NGS-panel used contains a comprehensive pool of genes clearly associated to tumor biological characterization and with clinical relevance in terms of tar-get-therapy and trial enrolment. This feature maximized the interpretation of genomic results for translational purposes, allowing a proper integration with clinical data.

Concerning the clinical application of this study, while we have clearly showed that a CGP may be of high value for the management of (recurrent) thymomas, we need to consider the remarkable costs related to CGP analysis and the its overall clinical usefulness before suggesting to adopt CGP analysis on large scale. In this setting, the identification of the best candidates who will really benefit from CGP is a crucial point; while performing CGP-analysis in all thymoma patients is almost questionable considering that only 14% of them will experience a relapse [4,5], we may suggest to test only recurrent cases. Since gene profile does not change when comparing initial tumor with tumor relapse (as reported herein), CGP-analysis may be performed on the initial surgical tissue if the sample of the recurrent tumor is not available.

5. Conclusions

In the present analysis, we found that relevant molecular findings of recurrent TETs generally belong to cell cycle control pathway. CGP do not substantially differ between initial tumor vs tumor recurrence and recurrent thymomas vs non-recurrent thymomas. Cell-cycle control genes alterations are associated with an early recurrence after thymectomy. Multiple target therapies are potentially available by performing a comprehensive CGP, suggesting that a precision medicine approach on these patients could be further explored.

Supplementary Materials: The following supporting information can be downloaded at the website of this paper posted on Preprints.org. Supplementary Table 1: Molecular Targets detected in our analysis (classified as TIER IIC) and the relative clinical trials (ongoing or completed).

Author Contributions: Conceptualization, F.L.; methodology, F.L. and D.G.; software, D.G. and E.D.P.; validation, S.M., A.U., F.R., E.B. and A.Cas.; formal analysis, D.G.; investigation, A.D., M.C. and C.S.; resources, G.S. and A.M.; data curation, A.Cam., A. Can., F.C., E.D.P. and J.E.; writing—original draft preparation, E.V., F.L., E.D.P. and J.E.; writing—review and editing, S.M., A.M., A.Cas, E.B. and G.S.; visualization, J.E. and E.D.P.; supervision, G.S and S.M.; project administration, A.C., G.S. and F.L. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Institutional Review Board Statement: Approval was obtained from the ethics committee of Policlinico Universitario "A. Gemelli" IRCCS of Rome (protocol ID: 3027). The procedures used in this study adhere to the tenets of the Declaration of Helsinki.

Informed Consent Statement: Informed consent was obtained from all subjects involved in the study.

Data Availability Statement: Data are available from the Authors upon request.

Acknowledgments: We thank for their precious support Federica Pezzuto and the Thoracic Research Pathology Laboratory, Biomedical Campus Pietro D'Abano University of Padua.

Conflicts of Interest: The authors declare no conflicts of interest.

References

1. de Jong WK, Blaauwgeers JL, Schaapveld M, Timens W, Klinkenberg TJ, Groen HJ. Thymic epithelial tumours: a population-based study of the incidence, diagnostic procedures and therapy. *Eur J Canc* 2008. doi: 10.1016/j.ejca.2007.11.004
2. Marx A, Chan JK, Coindre JM, et al. The 2015 World Health Organization classification of tumors of the thymus: continuity and changes. *J Thorac Oncol*. 2015. <https://doi.org/10.1097/JTO.0000000000000654>.
3. Regnard JF, Magdeleinat P, Dromer C, Dulmet E, de Montpreville V, Levi JF, Levasseur P. Prognostic factors and long-term results after thymoma resection: a series of 307 patients. *J Thorac Cardiovasc Surg*. 1996. doi: 10.1016/S0022-5223(96)70265-9. PMID: 8751506.
4. Sandri A, Cusumano G, Lococo F, Alifano M, Granone P, Margaritora S, Cesario A, Oliaro A, Filosso P, Regnard JF, Ruffini E. Long-term results after treatment for recurrent thymoma: a multicenter analysis. *J Thorac Oncol*. 2014. doi: 10.1097/JTO.0000000000000370. PMID: 25393792.
5. Margaritora S., Cesario A., Cusumano G., Lococo F., Porziella V., Meacci E., Evoli A., Granone P. Single-centre 40-year results of redo operation for recurrent thymomas. *Eur. J. Cardiothorac. Surg.* 2011. doi: 10.1016/j.ejcts.2011.01.025
6. Chiappetta M, Lococo F, Zanfrini E, Moroni R, Aprile V, Guerrera F, Nachira D, Congedo MT, Ambrogi MC, Korasidis S, Lucchi M, Filosso PL, Ruffini E, Sperduto I, Meacci E, Margaritora S. The International Thymic Malignancy Interest Group Classification of Thymoma Recurrence: Survival Analysis and Perspectives. *J Thorac Oncol*. 2021. doi: 10.1016/j.jtho.2021.07.004. Epub 2021 Jul 10. PMID: 34256111.
7. Mizuno T, Okumura M, Asamura H, Yoshida K, Niwa H, Kondo K, Horio H, Matsumura A, Yokoi K. Surgical management of recurrent thymic epithelial tumors a retrospective analysis based on the Japanese nationwide database. *J. Thorac. Oncol.* 2015. doi: 10.1097/JTO.0000000000000378.
8. Chiappetta M, Grossi U, Sperduto I, Margaritora S, Marulli G, Fiorelli A, Sandri A, Mizuno T, Cusumano G, Hamaji M, Cesario A, Lococo F. Which Is the Best Treatment in Recurrent Thymoma? A Systematic Review and Meta-Analysis. *Cancers (Basel)*. 2021. doi: 10.3390/cancers13071559. PMID: 33805310; PMCID: PMC8036834.
9. Giaccone G, Wilmink H, Paul M.A., van der Valk P. Systemic treatment of malignant thymoma: A decade experience at a single institution. *Am. J. Clin. Oncol.* 2006. doi: 10.1097/01.coc.0000227481.36109.e7

10. Bott M, Wang H, Travis W, Riely G.J., Bains M, Downey R, Rusch V, Huang J. Management and outcomes of relapse after treatment for thymoma and thymic carcinoma. *Ann. Thorac. Surg.* 2011. doi: 10.1016/j.athoracsur.2011.07.078.
11. Shimada M, Taniguchi H, Yamaguchi H, Gyotoku H, Sasaki D, Kaku N, Senju C, Senju H, Imamura E, Takemoto S, Yamamoto K, Sakamoto N, Obase Y, Tsuchiya T, Fukuda M, Soda H, Ashizawa K, Fukuoka J, Nagayasu T, Yanagihara K, Mukae H. Genetic profile of thymic epithelial tumors in the Japanese population: an exploratory study examining potential therapeutic targets. *Transl Lung Cancer Res.* 2023. doi: 10.21037/tlcr-22-794. Epub 2023 Mar 23. PMID: 37197618; PMCID: PMC10183388.
12. Kurokawa K, Shukuya T, Greenstein RA, Kaplan BG, Wakelee H, Ross JS, Miura K, Furuta K, Kato S, Suh J, Sivakumar S, Sokol ES, Carbone DP, Takahashi K. Genomic characterization of thymic epithelial tumors in a real-world dataset. *ESMO Open.* 2023. doi: 10.1016/j.esmoop.2023.101627.
13. Girard N, Basse C, Schrock A, Ramkissoon S, Killian K, Ross JS. Comprehensive Genomic Profiling of 274 Thymic Epithelial Tumors Unveils Oncogenic Pathways and Predictive Biomarkers. *Oncologist.* 2022. doi: 10.1093/oncolo/oyac115.
14. Agrafiotis AC, Brandão M, Berghmans T, Durieux V, Jungels C. Immunotherapy and Targeted Therapies Efficacy in Thymic Epithelial Tumors: A Systematic Review. *Biomedicines.* 2023. doi: 10.3390/biomedicines11102722.
15. Koga K, Matsuno Y, Noguchi M, et al. A review of 79 thymomas: modification of staging system and reappraisal of conventional division into invasive and non-invasive thymoma. *Pathol Int.* 44 (1994), pp. 359-367
16. Brierley J, Gospodarowicz M.K., Wittekind C. TNM Classification of Malignant Tumours (8th ed.), John Wiley & Sons, Inc, Hoboken, NJ (2017)
17. Marx A, Ströbel P, Badve S.S., et al. ITMIG consensus statement on the use of the WHO histological classification of thymoma and thymic carcinoma: refined definitions, histological criteria, and reporting *J Thorac Oncol.* 9 (2014), pp. 596-611.
18. Liu H, Gu Z, Qiu B, Detterbeck FC, Roden AC, Ruffini E, Okumura M, Girard N, Xiang Y, Liu Y, Du Z, Hao Y, Fu J, Zhang P, Pang L, Chen K, Wang Y, Yu Z, Mao T, Fang W; AME Thoracic Surgery Cooperative Group. A Recurrence Predictive Model for Thymic Tumors and Its Implication for Postoperative Management: a Chinese Alliance for Research in Thymomas Database Study. *J Thorac Oncol.* 2020. doi: 10.1016/j.jtho.2019.10.018. Epub 2019 Nov 11. PMID: 31726106.
19. Li MM, Datto M, Duncavage EJ, Kulkarni S, Lindeman NI, Roy S, Tsimberidou AM, Vnencak-Jones CL, Wolff DJ, Younes A, Nikiforova MN. Standards and Guidelines for the Interpretation and Reporting of Sequence Variants in Cancer: A Joint Consensus Recommendation of the Association for Molecular Pathology, American Society of Clinical Oncology, and College of American Pathologists. *J Mol Diagn.* 2017. doi: 10.1016/j.jmoldx.2016.10.002. PMID: 27993330; PMCID: PMC5707196.
20. <https://cancer.sanger.ac.uk/signatures/>, last accessed 02/2024 (last accessed, 06/2024)
21. <https://www.oncokb.org/> (last accessed, 06/2024)
22. <https://www.ncbi.nlm.nih.gov/clinvar/> (last accessed, 06/2024)
23. Kuzbari Z, Bandlamudi C, Loveday C, Garrett A, Mehine M, George A, Hanson H, Snape K, Kulkarni A, Allen S, Jezdic S, Ferrandino R, Westphalen CB, Castro E, Rodon J, Mateo J, Burghel GJ, Berger MF, Mandelker D, Turnbull C. Germline-focused analysis of tumour-detected variants in 49,264 cancer patients: ESMO Precision Medicine Working Group recommendations. *Ann Oncol.* 2023. doi: 10.1016/j.annonc.2022.12.003. Epub 2022 Dec 16. PMID: 36529447.
24. Chiappetta M, Sassorossi C, Nachira D, Lococo F, Meacci E, Ruffini E, Guerrera F, Lyberis P, Aprile V, Lucchi M, Ambrogi MC, Bacchin D, Dell'Amore A, Marino C, Comacchio G, Roca G, Rea F, Margaritora S. Survival outcome after surgery in patients with thymoma distant recurrence. *J Thorac Oncol.* 2024. doi: 10.1016/j.jtho.2024.04.004.
25. Ak N, Aydiner A. Nivolumab treatment for metastatic thymic epithelial tumors. *J Oncol Pharm Pract.* 2021. doi: 10.1177/1078155220968150. Epub 2020 Oct 25. PMID: 33100181.
26. Song X, Fan J, Zhu L, Wang Z, He Y, Zhou C. The efficacy and safety of immunotherapy in thymic epithelial tumors: more effective, more risky: a systematic review. *J Thorac Dis.* 2021. doi: 10.21037/jtd-21-290.
27. Montella L, Ottaviano M, Morra R, Pietroluongo E, De Placido P, Tortora M, Sorrentino C, Facchini G, De Placido S, Giuliano M, Palmieri G. The Never-Ending History of Octreotide in Thymic Tumors: A Vintage or A Contemporary Drug? *Cancers (Basel).* 2022. doi: 10.3390/cancers14030774.
28. Morfouace M, Stevovic A, Vinches M, Golfinopoulos V, Jin DX, Holmes O, Erlich R, Fayette J, Croce S, Ray-Coquard I, Girard N, Blay JY. First results of the EORTC-SPECTA/Arcagen study exploring the genomics of rare cancers in collaboration with the European reference network EURACAN. *ESMO Open.* 2020. doi: 10.1136/esmoopen-2020-001075.
29. Morfouace M, Novello S, Stevovic A, Dooms C, Janžič U, Berghmans T, Dziadziuszko R, Gorlia T, Felip E, Paz-Ares L, Mazieres J, O'Brien M, Bironzo P, Vansteenkiste J, Lacroix L, Dingemans AC, Golfinopoulos V,

Besse B. Results of screening in early and advanced thoracic malignancies in the EORTC pan-European SPECTAlung platform. *Sci Rep.* 2022. doi: 10.1038/s41598-022-12056-0.

- 30. Jovanovic D, Markovic J, Ceriman V, Peric J, Pavlovic S, Soldatovic I. Correlation of genomic alterations and PD-L1 expression in thymoma. *J Thorac Dis.* 2020. doi: 10.21037/jtd-2019-thym-13.
- 31. Conforti F, Pala L, Giaccone G, De Pas T. Thymic epithelial tumors: From biology to treatment. *Cancer Treat Rev.* 2020. doi: 10.1016/j.ctrv.2020.102014.
- 32. Jardim DL, Millis SZ, Ross JS, Woo MS, Ali SM, Kurzrock R. Cyclin Pathway Genomic Alterations Across 190,247 Solid Tumors: Leveraging Large-Scale Data to Inform Therapeutic Directions. *Oncologist.* 2021. doi: 10.1634/theoncologist.2020-0509.
- 33. Baldi A, Ambrogi V, Mineo D, Mellone P, Campioni M, Citro G, Mineo TC. Analysis of cell cycle regulator proteins in encapsulated thymomas. *Clin Cancer Res.* 2005. doi: 10.1158/1078-0432.CCR-05-0070.
- 34. https://ascopubs.org/doi/10.1200/JCO.2018.36.15_suppl.8519
- 35. Aesif SW, Aubry MC, Yi ES, Kloft-Nelson SM, Jenkins SM, Spears GM, Greipp PT, Sukov WR, Roden AC. Loss of p16INK4A Expression and Homozygous CDKN2A Deletion Are Associated with Worse Outcome and Younger Age in Thymic Carcinomas. *J Thorac Oncol.* 2017. doi: 10.1016/j.jtho.2017.01.028.
- 36. <https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE1356> (last accessed 06/2024)
- 37. Markey MP. Regulation of MDM4. *Front Biosci (Landmark Ed).* 2011. doi: 10.2741/3780. PMID: 21196223.
- 38. Yi EJ, Park JH, Lee HW, Cho SY, Na II, Kang MC. BRCA1 gene mutation in thymic malignant melanoma. *Ann Thorac Surg.* 2013. doi: 10.1016/j.athoracsur.2012.12.017.
- 39. Nicodème F, Geffroy S, Conti M, Delobel B, Soenen V, Grardel N, Porte H, Copin MC, Laï JL, Andrieux J. Familial occurrence of thymoma and autoimmune diseases with the constitutional translocation t(14;20)(q24.1;p12.3). *Genes Chromosomes Cancer.* 2005. doi: 10.1002/gcc.20225.
- 40. Enkner F, Pichlhöfer B, Zaharie AT, Krunic M, Holper TM, Janik S, Moser B, Schlangen K, Neudert B, Walter K, Migschitz B, Müllauer L. Molecular Profiling of Thymoma and Thymic Carcinoma: Genetic Differences and Potential Novel Therapeutic Targets. *Pathol Oncol Res.* 2017. doi: 10.1007/s12253-016-0144-8
- 41. Principe DR, Kamath SD, Munshi HG, Mohindra NA. Metastatic Thymoma Harboring a Deleterious BRCA2 Mutation Derives Durable Clinical Benefit from Olaparib. *Oncologist.* 2020. doi: 10.1634/theoncologist.2019-0393.
- 42. Cimpean AM, Raica M, Encica S, Cornea R, Bocan V. Immunohistochemical expression of vascular endothelial growth factor A (VEGF), and its receptors (VEGFR1, 2) in normal and pathologic conditions of the human thymus. *Ann Anat.* 2008. doi: 10.1016/j.aanat.2007.05.003.
- 43. Thomas A, Rajan A, Berman A, Tomita Y, Brzezniak C, Lee MJ, Lee S, Ling A, Spittler AJ, Carter CA, Guha U, Wang Y, Szabo E, Meltzer P, Steinberg SM, Trepel JB, Loehrer PJ, Giaccone G. Sunitinib in patients with chemotherapy-refractory thymoma and thymic carcinoma: an open-label phase 2 trial. *Lancet Oncol.* 2015. doi: 10.1016/S1470-2045(14)71181-7.
- 44. Radovich M, Solzak JP, Hancock BA, Conces ML, Atale R, Porter RF, Zhu J, Glasscock J, Kesler KA, Badve SS, Schneider BP, Loehrer PJ. A large microRNA cluster on chromosome 19 is a transcriptional hallmark of WHO type A and AB thymomas. *Br J Cancer.* 2016. doi: 10.1038/bjc.2015.425.
- 45. Rajan A, Carter CA, Berman A, Cao L, Kelly RJ, Thomas A, Khozin S, Chavez AL, Bergagnini I, Scepura B, Szabo E, Lee MJ, Trepel JB, Browne SK, Rosen LB, Yu Y, Steinberg SM, Chen HX, Riely GJ, Giaccone G. Cixutumumab for patients with recurrent or refractory advanced thymic epithelial tumours: a multicentre, open-label, phase 2 trial. *Lancet Oncol.* 2014. doi: 10.1016/S1470-2045(13)70596-5.
- 46. Zucali PA, De Pas T, Palmieri G, Favaretto A, Chella A, Tiseo M, Caruso M, Simonelli M, Perrino M, De Vincenzo F, Toffalorio F, Damiano V, Pasello G, Garbella E, Ali M, Conforti F, Ottaviano M, Cioffi A, De Placido S, Giordano L, Bertossi M, Destro A, Di Tommaso L, Santoro A. Phase II Study of Everolimus in Patients With Thymoma and Thymic Carcinoma Previously Treated With Cisplatin-Based Chemotherapy. *J Clin Oncol.* 2018. doi: 10.1200/JCO.2017.74.4078.
- 47. Abu Zaid MI, Radovich M, Althouse S, Liu H, Spittler AJ, Solzak J, Badve S, Loehrer PJ Sr. A phase II study of buparlisib in relapsed or refractory thymomas. *Front Oncol.* 2022. doi: 10.3389/fonc.2022.891383

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.