

# Analysis of the Setomimycin Biosynthetic Gene Cluster from *Streptomyces nojiriensis* JCM3382 and Evaluation of Its $\alpha$ -Glucosidase inhibitory activity using Molecular Docking and Molecular Dynamics Simulations

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**Table S1. Putative *S.nojiriensis* JCM 3382 BGCs\*.**

Regions	Type	From	To	Most similar known cluster		Similarity
Region 1	Terpene	40,991	61,998	Avermitilol	Terpene	100%
Region 2	Lanthipeptide-class-iv PKS-like Butyrolactone	119,303	167,470	Streptocollin	RiPP:Lanthipeptide	100%
Region 3	Thiopeptide LAP	168,443	197,954	Lactazole	RiPP:Thiopeptide	55%
Region 4	NRPS-like NRPS	432,526	479,497	Antipain	NRP	100%
Region 5	T2PKS	1,066,919	1,139,413	Spore pigment	Polyketide	58%
Region 6	NRPS/NRPS-like	1,561,900	1,631,094	Tambromycin	NRP	92%
Region 7	NRPS/T1PKS	1,796,196	1,861,151	Coelichelin	NRP	72%
Region 8	Butyrolactone	1,961,621	1,972,592	Neocarzinostatin	Iterative type I + Eneidyne type I polyketide	4%
Region 9	NI-Siderophore	3,622,602	3,652,383	Desferrioxamin B/E	Other	100%
Region 10	T2PKS	4,276,901	4,349,362	Setomimycin	Polyketide	100%
Region 11	RiPP-like	4,584,818	4,595,105			
Region 12	NRPS-like/T1PKS/NRPS	5,057,273	5,117,610	Bleomycin A2/B2	NRP + Polyketide + Saccharide	7%
Region 13	CDPS	6,208,634	6,229,392	BD-12	NRP	17%
Region 14	NI-Siderophore	6,607,990	6,640,997	Kinamycin	Polyketide	13%
Region 15	Hydrogen-cyanide	6,812,909	6,825,953	Aborycin	RiPP	21%
Region 16	RiPP-like	6,891,612	6,902,943			
Region 17	Terpene	7,011,883	7,034,099	Toxoflavin Fervenulin	Other	14%
Region 18	Lanthipeptide-class-iv	7,095,057	7,117,990			
Region 19	Terpene	7,404,584	7,431,728	Hopene	Terpene	61%

Region 20	T1PKS/HgIE-KS	7,504,501	7,555,916	Hexacosalactone A	Other	9%
Region 21	NRPS/NRPS-like	7,604,900	7,720,913	Leupeptin Pr/Ac	NRP	100%
Region 22	T1PKS	7,832,794	8,024,104	Linearmycin A-C	Type I polyketide	100%
Region 23	Lanthipeptide-class-iv	8,122,015	8,144,813			
Region 24	NRPS-like	8,186,135	8,229,356	Lipstatin	NRP	42%
Region 25	Terpene	8,288,028	8,309,149	Bombyxamycin A/ B	Polyketide	3%
Region 26	Terpene	8,385,926	8,407,281	2-Methylisoborneol	Terpene	100%
Region 27	Terpene	8,477,920	8,498,825	Monensin	Polyketide	5%
Region 28	Melanin	8,514,987	8,542,302	Istamycin	Saccharide	4%
Region 29	NI-Siderophore	8,627,590	8,658,781	Gausemycin A/B	NRP + Saccharide	4%
Region 30	T3PKS	8,688,189	8,729,250	Alkylresorcinol	Polyketide	100%
Region 31	CDPS/NAPAA	8,765,178	8,817,045	$\epsilon$ -Poly-L-lysine	NRP	100%

\*Predicted biosynthetic gene clusters of *S.nojiriensis* JCM 3382 (antiSMASH version 7.1.0).

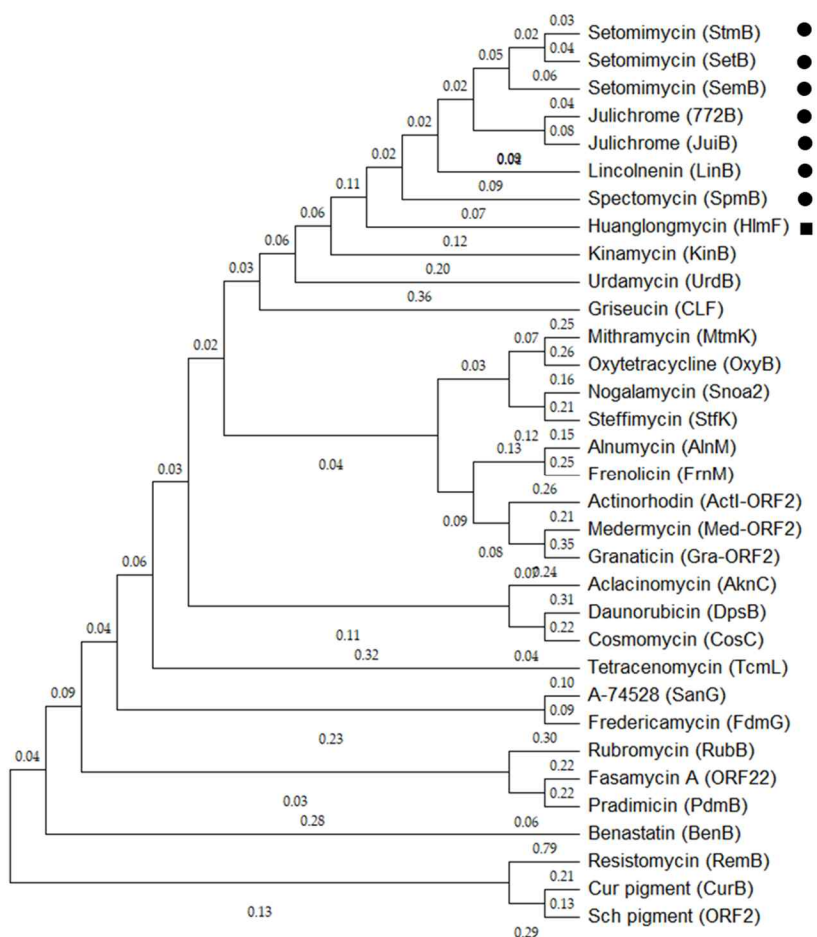
**Table S2. Genetic organization of the setomimycin BGCs\*.**

<i>S.nojiriensis</i> JCM3382			<i>S.aurantiacus</i> JA4570 (GenBank/Identities/Positives/Gaps)	<i>S.justiciae</i> RA-WS2 (GenBank/Identities/Positives/Gaps)
QTI46191.1	StmR (199aa)	TetR family transcriptional regulator	EPH44807.1/90%/92%/0%	MCW8383185.1/87%/93%/0%
QTI46192.1	StmP (206aa)	Hypothetical protein		
QTI46193.1	StmJ (504aa)	Drug resistance transporter	EPH44808.1/92%/95%/0%	MCW8383184.1/87%/93%/0%
QTI46194.1	StmK (217aa)	Type II polyketide cyclase (TcmI)	EPH44809.1/75%/86%/0%	MCW8383183.1/69%/83%/0%
				MCW8383182.1/58%/66%/6% (StmP-like)
QTI46195.1	StmL (142aa)	NTP-2 family protein	EPH44810.1/76%/86%/0%	MCW8383181.1/79%/89%/0%
QTI46196.1	StmG (268aa)	Sensory transduction protein(regX3)	EPH44811.1/87%/92%/0%	MCW8383180.1/79%/87%/0%
QTI46197.1	StmI (406aa)	Cytochrome P450	EPH44812.1/97%/99%/0%	MCW8383179.1/92%/95%/0%
QTI46198.1	StmM (64aa)	[3Fe-4S]-like ferredoxin	EPH44813.1/89%/95%/0%	MCW8383178.1/77%/90%/0%
QTI46199.1	StmN (152aa)	NTP-2 family protein	EPH44814.1/86%/91%/0%	MCW8383177.1/76%/83%/0%
QTI46200.1	StmH (216aa)	Type II polyketide cyclase (TcmI)	EPH44815.1/85%/90%/0%	MCW8383176.1/66%/75%/9%
QTI46201.1	StmA (423aa)	$\beta$ -ketoacyl synthase (KS $\alpha$ )	EPH44816.1/94%/97%/0%	MCW8383175.1/94%/97%/0%
QTI46202.1	StmB (397aa)	Chain length factors (KS $\beta$ )	EPH44817.1/93%/96%/0%	MCW8383174.1/90%/94%/0%
QTI46203.1	StmC (88aa)	Acyl carrier protein	EPH44818.1/86%/92%/0%	MCW8383173.1/86%/90%/3%
QTI46204.1	StmD (261aa)	C9 Ketoreductase	EPH44819.1/98%/99%/0%	MCW8383172.1/92%/95%/0%
QTI46205.1	StmE (317aa)	Aromatase/cyclase	EPH44820.1/88%/93%/1%	MCW8383171.1/79%/87%/0%
QTI46206.1	StmF (299aa)	$\alpha/\beta$ Fold hydrolase	EPH44821.1/85%/90%/1%	MCW8383170.1/79%/85%/0%
QTI46207.1	StmO (63aa)	Hypothetical protein	EPH44822.1/57%/62%/0%	
QTI46208.1	ORF1 (292aa)	4'-phosphopantetheinyl transferase superfamily protein		
QTI46209.1	ORF2 (199aa)	Phosphoribosylanthranilate isomerase		

\*Genetic organization, putative functions, and amino acid homology of the setomimycin BGCs from *S.nojiriensis* JCM 3382, *S.aurantiacus* JA4570, and *S.justiciae* RA-WS2.

**Table S3. Comparison of amino acid homology between the N- and C-termini of StmH and StmK.**

	StmH-N (Identity/Similarity)	StmH-C (Identity/Similarity)	StmK-N (Identity/Similarity)	StmK-C (Identity/Similarity)
StmH-N	-	-	-	-
StmH-C	39.68%/52.38%	-	-	-
StmK-N	33.86%/48.82%	37.90%/59.68%	-	-
StmK-C	27.73%/46.22%	34.75%/55.93%	33.33%/51.22%	-



**Figure S1.** Phylogenetic analysis of a cluster of 42 known KS $\beta$  genes in Type II polyketide BGCs. The alignment was created by use of ClustalX and edited manually. The phylogenetic tree was constructed using MEGA11 with the method of maximum likelihood. It indicates that the putative KS $\beta$  gene StmB is located in the KS $\beta$  group for biaryl nonaketide (●) and huanglongmycin (■) biosynthesis. Sequences included StmB (QTI46202.1), SetB (EPH44817.1), SemB (MCW8383174.1), LinB (ANS69118.1), SpmB (MCI3903331.1), JuiB (QNL10614.1), 772B (EPJ36720.1), and HlmF (AXL88815.1).

<b>ActIII</b>	MATQDSEVALV	<b>TGATSGIG</b>	LEIARRLGKEGLRVFVCARGEELRRTLKELREAGVEADGRTCDVRSVPEIEALVAAVVERYGPVDLV	<b>NNAG</b>	<b>PGGGAT</b>	AELADELWLD	VVETNLTGVFR	120
<b>HedA</b>	--MSRPQTAFV	<b>TGVSSGIG</b>	LAVARTLAARGIAVYGCARDAKNVSAAVDGLRAAGHDVDGSSCDVTSTDEVHAAVAAVERFGPIGILV	<b>NSAG</b>	<b>NCGGET</b>	ADLDDALWAD	VLDTNLTGVFR	118
<b>StmD</b>	MTQEQRVALV	<b>TGATSGIG</b>	LAVTRLLAGQGHRVFIGSRTEDKVLTTVKQLQEEQLDVDGTTCDVRSVPEIEALVAAVVERYGPVDLV	<b>NNAG</b>	<b>PGGGAT</b>	AELADELWLD	VVETNLTGVFR	120
<b>BexD</b>	MTDQEGRTAVV	<b>TGATSGIG</b>	LAVSKLLARRGLRVFLGARTAENVATTVKELRDAGFEADGAACDVRSGEDVTAfvRAAVDRYGPVGVLV	<b>NNAG</b>	<b>PGGGVT</b>	AEIADELWLD	VVIETNLTsvFR	120

**TGxxxGxG**
**NNAG PGGxxT**

<b>ActIII</b>	VTKQVLKAGGMLERGTGRIVNI A	<b>S</b>	STGGKQGVHAAP	<b>Y</b>	<b>S</b>	<b>A</b>	<b>S</b>	<b>K</b>	HGVVGF	KALGLELARTGITVNAVCPGFVETPMAASVREHYSDIWEVSTEEAFDRITARVPIGRYVQPSEVAEMVAYLI	240
<b>HedA</b>	VTREVLRAGGMREAGWGRIVNIA	<b>S</b>	STGGKQGVMAAP	<b>Y</b>	<b>T</b>	<b>A</b>	<b>S</b>	<b>K</b>	HGVVGF	TKSVGFELAKTGITVNAVCPGYVETPMAERVREGYARHWGVTEQEVHERFNAKIPLGRYSTPEEVAGLVGYLV	238
<b>StmD</b>	LTREVLTTGGLRDKSWGRVINI A	<b>S</b>	STAGKQGVVLGAP	<b>Y</b>	<b>S</b>	<b>A</b>	<b>S</b>	<b>K</b>	HGVVGF	KALGNELAPTGITVNAVCPGYVETPMAERVQRNYSRLSGAPESAILEKFQAKIPLGRYSTPEEVAGLVGYLA	240
<b>BexD</b>	VTREVLTTGGMVAAGHGRIVNI A	<b>S</b>	STAGKQGVVLGAP	<b>Y</b>	<b>S</b>	<b>A</b>	<b>S</b>	<b>K</b>	HGVVGF	KALGNELAPAGITVNAVCPGYVETPMAERVQRGYAAAYGTSEDEVLKKFQAKIPLGRYSTPDEVAGLVGYLV	240

**YxxxK**

<b>ActIII</b>	GPGAAAVTAQALNVCGLGNY	261
<b>HedA</b>	TDAAASITAQALNVCGLGNY	259
<b>StmD</b>	SDTAASITSQALNVCGLGNF	261
<b>BexD</b>	TDTAGSITAQALNVCGLGNF	261

**Figure S2.** Sequence alignment among various type II PKS KRs. Sequences included hedamycin (HedA), actinorhodin (ActIII), BE-7585A (BexD), and setomimycin (StmD) KRs. Multiple-sequence alignment was performed using Clustal Omega. The SDR proteins motif (TGxxxGxG, PGxxxT, NNAG and YxxxK) were shown in red.

**StfQ** MPTLGKDTLQYTATVAAPAPLVFDLVAGLENWPQF **H**GPSVHAEP LGRRDGAEFQHWVVIDRTRVTRWRAR **W**RFDQSALRIG**Y**TYDPAEPAAPAQHGWEWFRRLSDASTEVRVE **Y**EL—L 118  
**StmE** MPSE—LRTSEHEIKIAGPAENVYRLLAQAEKWPQL **F**PPNIYVDHVERGEREERIRI—WATAGDTVKHWTS**H**TLDPDKLRIG**F**RQEVSTHPVAAMGGSWQVEALPSGETLVRLS **H**DFRAA 118  
**BexL** ----MREAHDILVDAPADEVYRLVAEVANWPRI **F**PPTVFDHVERGEGTERIRI—WATANGEPKNWTSR**R**ELDPAARRIS**F**HQEVSTPPVAEMSGTWIVEPVSAATAKVRL **H**AYRAV 114  
  
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**StfQ** GDEGDA—AL----ADRELRELVCVT—DAAERHEERRDLVDFEDPLFVAGAVDDAYTYLYEADKWPERIPHVARLVMEERVP NIQ**F**DMDTTTPDGSHTTRSVRVCLPGDKIVYKQIQ 232  
**StmE** DD—DSLTLWLEEVDRNSRSELSLAKAESRDAADELTCSEFEDTVQIRGAAQDVYDFIYEANLWADRLPHVITVRLDEPTPGVQT **L**EMDTLSKDGSSHTTKSHRVGLAPHRIAYKQTT 236  
**BexL** GDDPGGLAWIDRAVDTNSRSELAALKHVEL—VTNPELTFSTDTVRIDAPAKDVYDFVDQAALWAERLPHVSSVDLREPSPLQV **L**RMDTRAKDGSVHTTESVRVCFPHHRIVYKQTT 232  
  
.\* .. : . \* : \* . : . : . : \* . \* \* : : . . \* . \* : : \* \* : \* : \* \* \* : : \* \* \* \* \* \* \* \* : . : \* . \* \* \*  
  
**StfQ** LPKLLTGHTGHWKFTPTREGFV—LGARHTATIKP—SALPILGEGTTVL DARKYLRRVLSANSMG **N**LRLAKAFAEERAGV---- 309  
**StmE** LPAL**I**TLHTGHWTFTPNDD—GVAASSQHTFTVNTDNIKVLGADATVQDAIAYVRTALSTNSKA **I**LGIAKEYAEGLAGAGRS 317  
**BexL** LPAL**M**TLHTGRWDFAEPPGGTTASSEHTVVLNTANIKVLGAGAGVAEAREFVRTALSTNSRA **I**LGFAKDHAEARP----- 309  
  
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**Figure S3.** Sequence alignment of StmE and polyketide ARO/CYCs. StmE is extremely conserved with R70 in the monodomain ARO/CYCs TcmN, WhiE, and ZhuI, whereas StfQ is substituted with a bulky W72. StmE is also highly conserved with F35, F81, and H103, which are seen in BexL rather than StfQ

(a)

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TcmI      MAYRALMVL RMDPAD-AEHVAAFAEHD T-TELPLEIGVRR RVLFRFHDL VMHLIEADD-DIMERLYQARSHPLFQEVNERVGQYLTPYA-QDWEELKDSKAEVFYSWTAPDS----- 109
AbxD      -MATTLIVARLKPGDHRDQISRLFAES DT-TELPDLVGVQE RRLTFKDL YFHLVVRTDE-ALSKLTPQHDLPLFRSISEAMDEYVTPYE-GAWGSVEQASARQFYHWRGLGRVQP- 113
StmK-C    -TGTVVTITRMDQAV-IPAVTSIFREF ES-AESSRPTGIRRRQLFAFKDLCIHIQEYTGSGADIAQQVASDIWSGKVVRLPEL-APEQ-----PAANGPASRFYGWEAS----- 102
StmH-N    -MHSSVIVGRIKPGS-TEAVAGVFAGRDG-EPGSTRNLLRRQLFEYKGIYIHLQDHTQENL TEVVEHGS-----DEAFTPFLDAYDPVAGTAPSGQVATRFYEWHGQPV SDEHR 106
StmK-N    -MRTMVTVAKMDQNS-IPEVSKLFAEFD S-TEIPNMIGLRRRQLFIHDGIQIHL YDFAEKSDQAAL EKVKTDPRMGRLAIDMKAFVTPYEPETWNSPADSTAFCFHDWEGEPIENPGP 115
StmH-C    -MYSTVIVARLNPAD-IGTVAKLFGDFD TGTELPHTMGTRRRQLFAYNGLYFHIQDFESDNGGELIERAKSDPRFVQIQD LLYISVLDPETWRSPSDAMASRFYTWQVTA----- 110

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(b)

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StmK-C    TGTVVTITRMDQAVIPAVTSIFREFES-AESSRPTGIRRRQLFAFKDLCIHIQEYTG SNG 59
StmH-N    MHSSVIVGRIKPGSTEAVAGVFAGRDG-EPGSTRNLLRRQLFEYKGIYIHLQDHTQENL 58
StmH-C    MYSTVIVARLNPADIGTVAKLFGDFD TGTELPHTMGTRRRQLFAYNGLYFHIQDFESDNG 60
StmK-N    MRTMVTVAKMDQNSIPEVSKLFAEFD S-TEIPNMIGLRRRQLFIHDGIQIHL YDFAEKSD 59

      : * : :.      *: :* : * . ***** ... : *: :. ..
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**Figure S4.** Comparison of the amino acid sequences of TcmI, AbxD, StmH, and StmK proteins. (a) Alignment of the complete amino acid sequence of TcmI, AbxD, and the N-terminus and C-terminus of StmH and StmK. (b) Alignment of the N-terminus and C-terminus of StmH and StmK with each other. Among the three amino acids that play a role in controlling and catalyzing cyclizaion and aromatization in TcmI and AbxD, R40 and H51 are conserved in both the N-terminus and C-terminus of StmH/StmK, whereas Y49 is only conserved in the N-terminus and C-terminus of StmH, and, the C-terminus of StmK of D27 is substituted with Asp.

(a)

**StmF** MTTAPVAPPPAR----SIDQRHITLDADGIMLSARLAQPVHVPPRATIVALHGA**GMSAG**YFDGPAHPETSLLAVAAELGFTAVAI DRPGYGRSAGRLPYGQGVVAQSHTLAAALRRLVAQ  
**SetF** MTTAPVAPFTGRSADPRIDRRHLTLDADGVTL SARLARPAHVPPRATVVALHGA**GMSAG**YFDGPAHPETSLLTAAELGFTAVAVDRPGYGDSAGRLPHGQGVAAQADTLAAALRGLLAR  
**SemF** MTTAPPAPPPPAP---VVDRLITLDADGIPLSARLALPTHVPPRATVVALHGA**GMSAA**YFDGPAHPETSLLTAAELGFAAVAVDRPGYGASAGQLPYGQGVGDQAATLSAALRSLSAF  
**AlpS** -----MASRTKARAARPARVTLNCLAHAGAGVASYR-----DWASAVGDGVDVHALPLPGR-EARRREPRLTTRADLLADLLPGLLTARR

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**StmF** YETGAGIFLM**GHSFG**SKPALQIAADGTVP E IIGLDLSGCGAEYLVPTAPGTRAGSWKLN--WGPLRLYPPGTFQSSIGVVAPAPERELADAAQWPTVFAALAERIRVPVRF TFAEHEAW  
**SetF** HETGAGIFLM**GHSFG**CKPALRIAADGTVPGLGLDL SGCGAEYLAAPGAPATRAGSWKLN--WGPLRLYPPGTFQSSIDVVAPAPVRELADAAHWPTVFPALAERVRVPVRF TFAEHEAW  
**SemF** CAPDAGIFLV**GHSFG**SKPALRMAADGRVPGLLGLDVSGCGDEYLVPTTPGTRAGSWKLN--WGPLRLYPPGTFQAGLAVVAPPERELADAARWPEEFAALAGRVRVPVRF TFAEHEAW  
**AlpS** GPYALY**GHSMG**ALVGYTLTRALADAGAPP--LFLAVGACPPHTTTALVDGADLPDEDLLPLDITIGSLPPGASASPGGLWRRTFLPVLRDDRLAKSLRNAALDPVTGGPVDV PVLVFA

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**StmF** WRRD-HQALTRLRSRLVAAPRVLIDHQPGAG**NISL**GWAA RAYHLRALGFAEECLHRRSPGTEQRA-  
**SetF** WRRD-HEALTRLRNRLAAAPRVLIDHQPDAG**NISL**GWAA RAYHLRALGFAEECLRRYAADGKVVHP  
**SemF** WRRD-APTLARLRNRLDAAPRVRIDHQPDAG**NISL**GWAA RSYHLRALSF AEECLRRRTGGEGRQP-  
**AlpS** GSD**D**PLAAPAALRHQQWTTDLIELHTVSGG**H**---FFASSAGLAQHVG RACRDHVNAQPAGGRR--

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(b)

**SpmF** MTGPQGGVALAPQSPSTAGAPTTRPPSAARRPAGPAPRPAPPVPLPPWTPSPGVRQVTLAAGDGTLSGLLALPRGERPRATVLALHGA **GTSAA**YFDGQAHPGTSLLT LAADLGYAVL  
**JuiF** -----MTVAQPAVAGGVRGISL-DADGITLSALLALPPQAPCRAVIVALHGA **GMSA**YFHGSAHPDTSFLGLATSLGFGVV  
**H1mB** -----MTKAPP-----KAPPTAPPTAPPTAPAAVSAKVGRITL-QAGGITVSALLARPLYAPPRATVVALHGA **GMSAA**YFHGQTRPETSLLT LAADLGF AAV  
**LinF** -----MTAA-----GSAAARPPATVRRITL-DADGTPLSALLALPEHTPPRATVVALHGA **GMSAA**YFHGTARPDT SFLALAADLGF AAV  
**SemF** -----MTTAPP-----APPP---PAPVVDRLITL-DADGIPLSARLALPTHVPPRATVVALHGA **GMSAA**YFDGPAHPETSLLT LAELGFAAV  
**StmF** -----MTTAPV-----APPPAR----SIDQRHITL-DADGIMLSARLAQPVHVPPRATIVALHGA **GMSAG**YFDGPAHPETSLLAVAAELGFTAV

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SetF -----MTTAPV-----APFTGRSADPRIDRRHRTL-DADGVTLSARLARPAHVPPRATVVALHGA GMSAGYFDGPAHPETSLTLAAELGFTAV
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SpmF ALDRPGYGLSAARLPQGLPVAEQAATVRAALRGHAALHDTGAGVLLV AHSFGGKVALRMAADDRGDPSPVLLGLDVSGCGERYARPLAELTGPAGGVRWSGHWGPLRLYPPHTFRASAR
JuiF AVDRPGYGRSARQLPEGQGVADQATTLGAALRFLAARYDTGSGLFLL AHSFGGKAAALRIAADR----AVPELLGLDVSGCGAEYAVPLGDRE-PHGRQSRRLNWGPLRLYPPGTFRASGG
HlmB AVDRPGYGLSSAELPRGQTAAEQAAATLTAALGDLADRYDTGAGLFLL AHSFGGKPALRMAAEG----TLPALLGVDVSGCGDAYTAPPEPG-T-RDGGWRRSWGPLRLYPPGTFRSGAA
LinF ALDRPGYGLSARRLPEGQGVMEQATTIAAAVRDLTARHLPGAGVFL AHSFGGKPALGIAADG----LVPGLLGLAVSGCGDEYLVPTATT-G-RAAQWRLNWGPLRLYPPGTFRASGA
SemF AVDRPGYGASAGQLPYGQGVGDQAATLSAALRSLSAFCAPDAGIFLV GHSFGSKPALRMAADG----RVPGLLGLDVSGCGDEYLVPTTPG-T-RAGSWKLNWGPLRLYPPGTFAQGLA
StmF AIDRPGYGRSAGRLPYGQGVVAQSHTLAAALRRLVAQYETGAGIFLM GHSFGSKPALQIAADG----TVPEIIGLDLSGCGAEYLVPTAPG-T-RAGSWKLNWGPLRLYPPGTFAQSSIG
SetF AVDRPGYGDSAGRLPHGQGVAAQADTLAAALRGLLARHETGAGIFLM GHSFGCKPALRIAADG----TVPGLLGLDLGCGAEYLAAPGAPA-T-RAGSWKLNWGPLRLYPPGTFAQSSID
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SpmF MVSPVPPRELDAARWPGFAALAGRVVRVRFVTFEAEYALWCH DAPALARLRAGLRAAPRVSVDRQPGAG HNISLGWAARAYHLRALAFADDCLRR-----
JuiF VVAPVPSLELADAARWPGDFAGFAGRVVRVRFVTFEAEHEFWQH DARAVARLRSRLSGAPRVVTDHQPdag HNISLGWSARAYHLRVLGFVEECLRWRESRQRATPEEQEGT
HlmB VVARVPPGELADAARWPAEFGRLAGRIRVVRVRFVTFEAYETWWRH DPPALDRLRSLSAAPRVVIEHQPdag HNISLGWAARAYHLKALGFLEESMRHHGVGRESRT-----
LinF VVAPAPPRELADAGRWPETFRRIGGRIRVVRVRFVTFEAEHEHWRH DRRAVAQLRSRLHAAPRVLDHQPdag HNISLGWTARAYHLGVLGFVADCLRPDGDGKEHPA-----
SemF VVAPPPERELADAARWPEEFAALAGRVVRVRFVTFEAEHEAWWR DAPTLARLRNRLDAAPRVLDHQPdag HNISLGWAARSYHLRALSFEECLRRRTGGEGRQ-P-----
StmF VVAPAPERELADAAQWPTVFAALAERIRVVRVRFVTFEAEHEAWRR DHQALTRLRSRLVAAPRVLDHQPdag HNISLGWAARAYHLRALGFEECLHRRSPGTEQRA-----
SetF VVAPAPVRELADAAHWPTVFPALAERVRVVRVRFVTFEAEHEAWRR DHEALTRLRNRLAAAPRVLDHQPdag HNISLGWAARAYHLRALGFEECLRRYAADGKVVHP-----
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**Figure S5.** Sequence alignment of AlpS and biaryl polyketide type II TEs. AlpS and setomimycin type II TEs (a) and their respective biaryl polyketide TEs (b). The conserved GHSMG motifs are marked by azure colors, and the Ser-His-Asp triad is highlighted in red. Sequences included AlpS (AAR30163.1), SemF (MCW8383170.1), StmF (QTI46206.1), SetF (EPH44821.1), LinF (ANS69114.1), SpmF (MCI3903327.1), JuiF (QNL10618.1), and HlmB (AXL88811.1).



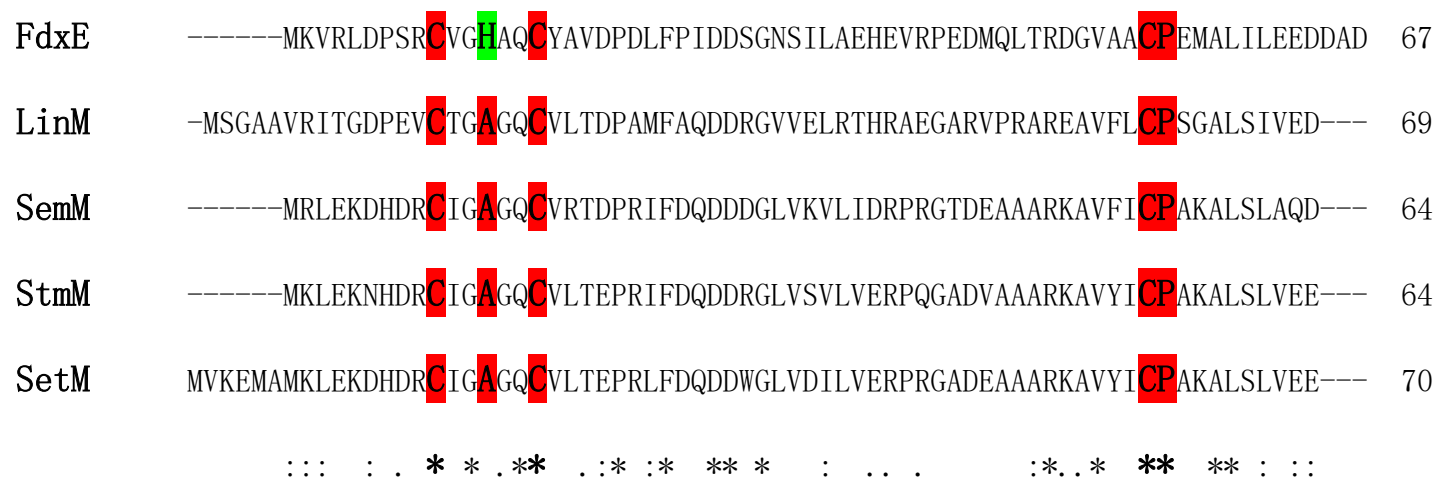


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SemI  LGQNLGRVELQVVFDLFRRLPDLRLAVPAEELNFKSDALVYGLYELPVTW  406
SetI  LGQNLGRLELQVVFDLFRRLPELRLAVPEEELSFKSDALVYGLYELPVTW  382
StmI  LGQNLGRLELQVVFDLFRRLPDLRLAVPEEELSFKSDALVYGLYELPVTW  406
SpmI  LGRNLARVELQTVFDLFRRLPGLRLAVGVEELPFKYDALVHGLRELPTW  407
LinI  LGQNLSRLELQIVFDLFRVPTLRLAVDEADLPFKYDALAFGLYELPVTW  407
      **: ** * *** *: * ** *: * ****.   * : * . . * *****

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**Figure S6.** Sequence alignment of P450 ezymes. Sequence alignment of actinomycetes P450s and StmI (a) and their respective biaryl polyketide P450s (b). Typical P450-fold, with helix L containing the signature sequence <sup>348</sup>FGHGXXHXCLG<sup>357</sup> containing the proximal axial thiolate ligand of the heme iron, <sup>355</sup>Cys, is conserved, and the highly conserved <sup>244</sup>A/GGxxT<sup>248</sup> motif and the <sup>283</sup>ExxR<sup>286</sup> motif in the K-helix, and the <sup>351</sup>GxxxC<sup>355</sup> motif in the heme binding ring is also conserved in StmI and its corresponding enzymes. All of them are marked in red. Amino acids conserved in biaryl polyketide P450 that are specific to the motif region are coloured green. Sequences included OxyA (GIF17054.1), OxyB (Q8RN04.1), AspB (WP\_030881046.1), 6KZS (WP\_003970936.1), JuiI (QNL10608.1), SemI (MCW8383179.1), SetI (EPH44812.1), StmI (QTI46197.1), SpmI (MCI3903336.1), and LinI (ANS69122.1).



**Figure S7.** Sequence alignment of FdxE and biaryl polyketide ferredoxins. The genes which encode [3Fe-4S] or [4Fe-4S] single cluster containing ferredoxins have unusual iron-sulfur cluster binding motif sequences, CXX?XXC(X)nCP, where '?' indicates a variable amino acid residue. Rather than a cysteine residue, which is highly conserved in [4Fe-4S] clusters, alanine or glycine residues at this position in [3Fe-4S] ferredoxin. The conserved CXXA/GXXC(X)nCP motif of [3Fe-4S] ferredoxin is highlighted in red. In FdxE, the Ala residue of the CXXA/GXXC(X)nCP motif is substituted by a His residue and is coloured green. Sequences included StmM (QTI46198.1), SetM (EPH44813.1), SemM (MCW8383178.1), LinM (ANS69121.1), and FdxE (O53937.1).