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Article

Larvicidal Activity of Hemp Extracts and Cannabidiol against the Yellow Fever Mosquito *Aedes aegypti*

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Simple Summary: The present study examined whether extracts of hemp leaves were toxic to *Aedes aegypti* larvae and determined which compound(s) were responsible for the toxicity. We found that larvae, from both insecticide-resistant and -susceptible strains were killed by hemp leaf extract within 48 h of exposure. Furthermore, we found that an abundant cannabinoid (cannabidiol) within the extract was the primary active compound. This study suggests that hemp extracts and cannabidiol are potentially valuable sources for developing biopesticides to control mosquitoes.

Abstract: To mitigate pyrethroid resistance in mosquito vectors of emerging and re-emerging human pathogens, there is an urgent need to discover insecticides with novel modes of action. Natural alternatives, such as extracts derived from plants, may serve as substitutes for traditional synthetic insecticides if they prove to be sustainable, cost-effective, and safe for non-target organisms. Hemp (*Cannabis sativa*) is a sustainable plant known to produce various secondary metabolites with insecticidal properties, including terpenoids and flavonoids. The goal of this study was to assess the larvicidal activity of hemp leaf extract on 1st instar mosquito larvae from both pyrethroid-susceptible (PS) and pyrethroid-resistant (PR) strains of *Aedes aegypti*. Another goal was to identify which components of the extracts were responsible for any observed larvicidal activity. We found that methanol extract of hemp leaves induced similar concentration-dependent larvicidal activity against PS (LC₅₀: 4.4 ppm) and PR (LC₅₀: 4.3 ppm) strains within 48 hours. Partitioning of the dried leaf extract between methanol and revealed that full larvicidal activity was restricted to the methanol fraction. Analysis of this fraction by gas chromatography-mass spectrometry and nuclear magnetic resonance showed it to be dominated by cannabidiol (CBD). Larvicidal assays using authentic CBD confirmed this compound was primarily responsible for the toxicity of hemp leaf extract against both strains. We conclude that hemp leaf extracts and CBD have the potential to serve as viable sources for the development of novel mosquito larvicides.

Keywords: hemp; extracts; cannabidiol (CBD); mosquito; larvicide; insecticide resistance

1. Introduction

Mosquitoes are considered the most dangerous animals on Earth, because they are vectors of numerous pathogens that cause deadly and debilitating diseases in humans and domestic animals, including malaria, West Nile virus, and heartworm [1–4]. The yellow fever mosquito *Aedes aegypti* is a vector of several arboviruses of medical importance, including chikungunya, dengue, yellow fever, and Zika. To limit transmission of mosquito-borne arboviruses, humans have relied heavily on insecticides to control the larval and/or adult life stages of mosquitoes. Synthetic insecticides, such as pyrethroids, are critical chemical tools used to control *Ae. aegypti*; however, the widespread use of insecticides with limited modes of action has led to resistance [5–7]. Additionally, the overuse of these insecticides has led to detrimental impacts on both human and environmental health [8]. Because of

this, the discovery of alternatives to synthetic pesticides, like biopesticides, is crucial to improving mosquito management.

Plant secondary metabolites constitute a diverse library of chemicals that mediate a wide array of interactions, including biotic defense, competition, and mutualism [9]. As such, they provide a rich resource for the discovery of compounds for pharmaceuticals, antibiotics, and biopesticides. [9–12].

The hemp plant, *Cannabis sativa*, is a rapidly emerging high value specialty crop that can be cultivated for the production of a wide array of products [13]. For example, hemp seeds, which have numerous health benefits, are used to produce a diversity of goods including milk, dietary supplements, animal feed, and oil [14–18]. Furthermore, hemp bark and stems are used for industrial production of textiles, paper and even concrete [17,19,20]. Hemp leaves and inflorescences are also used in many products (e.g., essential oils) because of their high content of secondary metabolites [21]. Both leaves and inflorescences produce more than 1,000 different secondary metabolites including highly volatile and aromatic terpenes and phenols, as well as semi-volatile phytocannabinoids [21,22]. Phytocannabinoids are specialized metabolites that were first discovered in *C. sativa*, but have also been found in *Helichrysum umbraculigerum* Less [23,24], *Amorpha fruticosa* L. (Fabaceae), *Glycyrrhiza foetida* and several *Rhododendron* species [24].

In addition to medicinal uses, phytocannabinoids possess insecticidal and/or antifeedant activity against herbivorous insects. For example, as hemp plants produce more phytocannabinoids, lepidopteran larvae (*Trichoplusia ni* and *Manduca sexta*) have shown to feed less on plant tissue [25,26]. Additionally, larval survival and growth of *T. ni* is reduced when cannabidiolic acid (CBDA) and cannabigerolic acid (CBGA) were incorporated in larval diets [25]. Moreover, cannabidiol (CBD) oil (3%) impairs development and lowers survival rates in stored product insect pests (*Plodia interpunctella*, *Oryzaephilus surinamensis*, and *Tribolium confusum*) [27].

Hemp also produces over 100 terpenes, more than 40 noncannabinoid phenolics, and more than 30 flavonoids [22,28]. Notably, the terpenes linalool, eucalyptol, p-cymene, and thymol have shown larvicidal and/or adulticidal activity to various mosquitoes species [29–31]. Thus, the hemp plant produces a variety of secondary metabolites with potential for mosquito control.

Consistent with this notion, hemp-based essential oils and extracts are larvicidal and adulticidal against multiple mosquito species [32–37]. However, cross resistance of pyrethroid-resistant mosquito strains to hemp extracts and the specific compound(s) responsible for mosquitocidal activities within the extracts are unknown. To address these gaps of knowledge, the objectives of this study were to 1) compare the larvicidal potency of hemp leaf extracts against pyrethroid-susceptible (PS) and pyrethroid-resistant (PR) strains of *Ae. aegypti* and 2) identify the principal active larvicidal ingredients in the extracts that elicit mortality.

2. Materials and Methods

2.1. Hemp Plants

The hemp material used to obtain the raw extract was grown under controlled greenhouse conditions (24°C 14:10 L:D cycle). The cuttings for new plants used for the experiment were obtained from previously grown hemp mother plants (variety Tango Kush). In brief, fresh shoots were cut using a sanitized razor blade, and the cuttings were placed in a 500 ppm DIP 'N' Grow Hormonal Rooting Concentrate (Clackamas, OR, USA) for 30 sec. The cuttings were then transferred to a 34 ct double strip OASIS wedge (Grow It Depot, Long Branch NJ, USA) filled with Pro-Mix BX soilless medium (Quebec, Canada) and placed in a mist chamber at controlled conditions. When root growth was adequate, hemp plants were transplanted into 38 L plastic pots and placed in a greenhouse room maintained at 25°C, 16:8 (L:D) by an Argus Control System (Conviron Company, British Columbia, Canada).

2.2. Hemp Extract

Leaves were removed from the hemp plants 7 months after planting and air-dried at 25°C for 7 days. Dried vegetative material was pulverized using a coffee grinder; 150 g of powdered material

was extracted in 4 L methanol for 3 weeks at 20°C, shaking the solution for 2-3 min daily. The incubated solution was then filtered through a membrane filter paper on top of a Porcelain Buchner funnel (Fisherbrand, Waltham MA, USA) attached to a Little Giant pressure vacuum pump (Gelman Instrument Company, Ann Arbor, MI, USA). The solution was placed in a Hei-Vap Rotory Evaporator (Heidolph NA, Wood Dale, IL, USA) at 30°C and 65 rpm to remove the methanol. The resulting crude residue (dried extract) was kept at 4°C and aliquots were resuspended in 100% acetone immediately prior to use in bioassays.

2.3. Methanol and Hexane Partitioning

We partitioned 50 mg of dried extract between 20 mL hexane and 20 mL methanol using a 60 mL separatory funnel (Chemglass Life Sciences LLC, Vineland, NJ, USA), with the resulting partitions collected into separate 40 mL glass vials (Thermo Scientific, Rockwood, TN, USA). The solutions were then evaporated to dryness under nitrogen at room temperature, using a Reacti-Vap (Thermo Scientific, Waltham, MA, USA) connected to a Nitrox UHPLCMS12 nitrogen generator (Domnick Hunter, Gateshead, UK). The residues were resuspended in acetone to reach the desired concentration for mosquito bioassays or gas chromatography/nuclear magnetic resonance analysis.

2.4. Gas Chromatography-Mass Spectrometry (GC-MS) and Nuclear Magnetic Resonance (NMR)

GC-MS analyses were performed using an Agilent Technologies 7890A GC equipped with a 7683B auto-sampler and interfaced to a 5975C inert mass selective detector (Agilent Technologies, Santa Clara, CA, USA). A 1- μ l sample was injected splitless at 280 °C, with a constant He flow of 1.1 ml/min. The GC column was HP-5MS, 30 m x 250 μ m diam x 0.25 μ m film thickness. The oven was programmed with an initial temperature of 35 °C for 1 min, increased 7 °C/min to 100 °C, then 25 °C/min to 280 °C held for 10 min., with the MS transfer line at 280 °C. The MS was operated in scan mode using m/z 19-450 with source at 230°C and quadrupole at 150 °C. GC peaks were identified by searching their mass spectra against the NIST/EPA/NIH Mass Spectral Library and comparing relative retention times against published values. To quantitate CBD, a calibration curve was prepared from a CBD isolate (99%, Extract Labs, Boulder, CO) dissolved in acetone, and injected into the GC-MS (19.4 - 525 ng, linear R^2 = 0.98).

Hemp extracts and methanol/hexane partitions were also analyzed via proton NMR. They were first dissolved in 0.6 mL of deuterated chloroform before analysis with a Bruker AVANCE III 400 MHz NMR (Bruker, Billerica, MA, USA). Resonances were analyzed and compared with previously published data [38].

2.5. *Aedes Aegypti* Colony

Larvae of *Ae. aegypti* from Liverpool (strain LVP-IB12, MRA-735, contributed by David W. Severson) and Puerto Rico (strain Puerto Rico, NR-48830, contributed by G.G. Clark & J.J. Becnel) strains were reared from eggs using established methods [39,40]. The original eggs were provided by the Centers for Disease Control and Prevention for distribution by BEI Resources, NIAID, NIH. Larvae of the Puerto Rico strain used in the present study were over 30-times resistant to the pyrethroid cypermethrin compared to the Liverpool strain (Supplemental Figure S1). From here, we refer to the Liverpool strain as pyrethroid-susceptible (PS) and the Puerto Rico strain as pyrethroid-resistant (PR). Larvae from both strains were nourished with fish food (Tropical Tablets, Tetramin, Blacksburg, VA, USA). Adult mosquitoes were provided 10% sucrose solution ad libitum. For additional egg production, adult females were fed defibrinated rabbit blood (Hemostat Laboratories, Dixon, CA, USA) using a membrane feeder (Hemotek, Blackburn, UK). All mosquitoes were reared in environmentally controlled chambers held at 28°C and 80% RH, with a 12:12 L:D cycle.

2.6. Larval Bioassay

Larvicidal activities of hemp extracts, methanol/hexane fractions, and CBD isolates against 1st instar larvae were determined using an established bioassay [40]. In brief, to each well of a 24-well

Falcon® Multiwell plate (Becton Dickinson Labware, Franklin Lakes, NJ, USA) the following was added: 985 μ L of deionized H₂O (diH₂O), six 1st-instar *Ae. aegypti*, 5 μ L of food solution (13 mg/mL of finely ground fish food flakes; Tetramin, Blacksburg, VA, USA), and 10 μ L of a treatment (hemp extract, methanol fraction, hexane fraction, CBD isolate, or suitable solvent control). The hemp extract and the methanol and hexane fractions were serially diluted with acetone to 100, 33, 11, 3.6, and 1.2 ppm. CBD isolate was serially diluted with acetone to 20, 6.0, 2.2, 0.7, and 0.24 ppm. The CBD isolate concentrations were selected to match the corresponding CBD concentrations in the hemp extracts as determined by GC-MS. The plates were held under standard rearing conditions, and larval mortality was assessed at 48 h. Larvae were considered dead if they did not move after gently touching their abdomen with a fine needle or pipette tip. All mortality values were corrected for solvent control mortality using Abbott's formula [41]. If solvent control mortality exceeded 20% then the trial was excluded.

2.7. Statistical Analysis

Data analysis and plotting were performed using GraphPad Prism (version 6.07) software (GraphPad, San Diego, CA, USA). Median lethal concentrations (LC₅₀) were determined by plotting percent mortalities against log transformations of the concentration tested. A non-linear regression ('log(agonist) vs. normalized response' function) was used to best fit the data and calculate LC₅₀. Statistical comparison of LC₅₀ values was performed through sum-of-squares F-tests ($\alpha = 0.05$).

3. Results

3.1. Hemp Leaf Extract Toxicity against 1st Instar Larvae

Application of hemp leaf extracts to the rearing water of 1st instar *Ae. aegypti* caused concentration-dependent mortality in both PS and PR strains within 48 h that reached 100%. The LC₅₀ value of leaf extracts in the PS strain (4.4 ppm; 95% CI = 4.0 - 4.8 ppm) was not different ($p = 0.85$; $F = 0.03$) than that of the PR strain (4.3 ppm; 95% CI = 2.37-7.70 ppm).

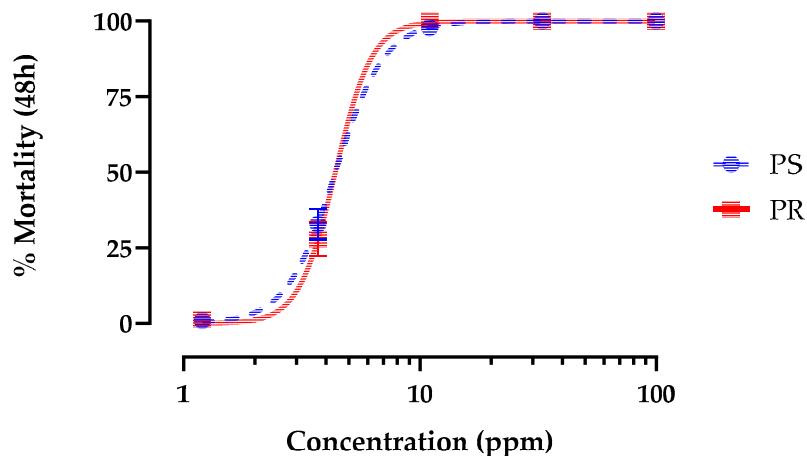


Figure 1. Concentration-response curves for 48 h larvicidal activity of hemp leaf extract against 1st instar pyrethroid-susceptible (blue) and pyrethroid-resistant (red) *Ae. aegypti*. Values plotted are means \pm standard errors of the mean (SEM) based on 28 replicates of 6 larvae per concentration (1.2, 3.7, 11, 33, 100 ppm).

3.2. Methanol and Hexane Partition Toxicity against 1st Instar Larvae

Dried hemp leaf extract was partitioned between methanol and hexane to separate polar and nonpolar constituents, respectively. These partitions were then tested in parallel with the original unpartitioned hemp leaf extract for larvicidal activity against the PS strain. Both the methanol

partition and unfractionated leaf extract produced concentration-dependent mortality within 48 h that reached 100% (Figure 2). However, the LC₅₀ of the methanol partition (4.3 ppm; 95% CI = 3.8-4.9 ppm) was ~2.5-times less potent ($p < 0.001$, $F = 88.1$,) than that of the unfractionated leaf extract (1.7 ppm; 95% CI = 1.4-2.0 ppm). In contrast, the hexane fraction did not elicit concentration-dependent mortality, reaching only 25% mortality at the maximal concentration tested (Figure 2).

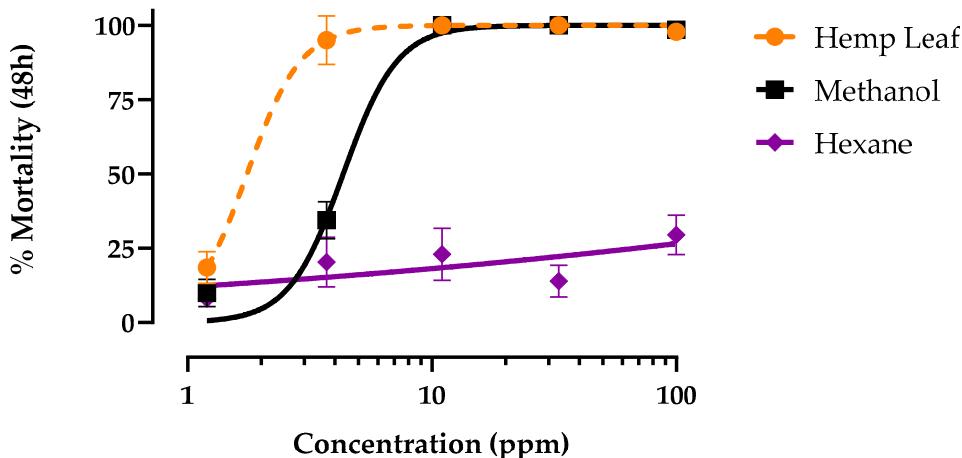


Figure 2. Concentration-response curves for 48 h larvicidal (1st instar) activity of methanol (black) and hexane (purple) fractions of hemp leaf extract (orange) against pyrethroid-susceptible *Ae. aegypti*. Values are means \pm SEM based on 12 replicates of 6 larvae per concentration (1.2, 3.7, 11, 33 and 100 ppm).

3.3. GC-MS and NMR Analysis

GC-MS identified CBD as the most abundant compound in the original dried leaf extract and both partitions; however, the methanol partition contained ~4x more CBD (80% of CBD found in dried leaf extract) compared to the hexane partition (20%) (Figure 3B). Additionally, the analysis indicated the presence of other compounds like α - and β -caryophellene and bisabolol in both the unpartitioned extract and hexane partition, but not the methanol partition (Supplemental Table S1). The ¹H NMR spectra of the leaf extracts measured in deuterated chloroform were superposable with the ¹H NMR spectrum reported by Barthlott, et al [38]. Since CBD was observed as the major compound, quick ¹³C NMR was collected to confirm its presence in the extract (Supplemental Figure S2). Moreover, signals arising from proton (¹H) of the active methanol fraction showed CBD as the major compound and a trace of THC analogs (Supplemental Figure S3). Resonances arising from the protons of bisabolol were also observed (δ 5.36, brs; δ 5.12, brt; and singlet methyls at δ ~1.6 ppm; Supplemental Figure S4); they were similar to those reported by Cerneau and co-workers [42].

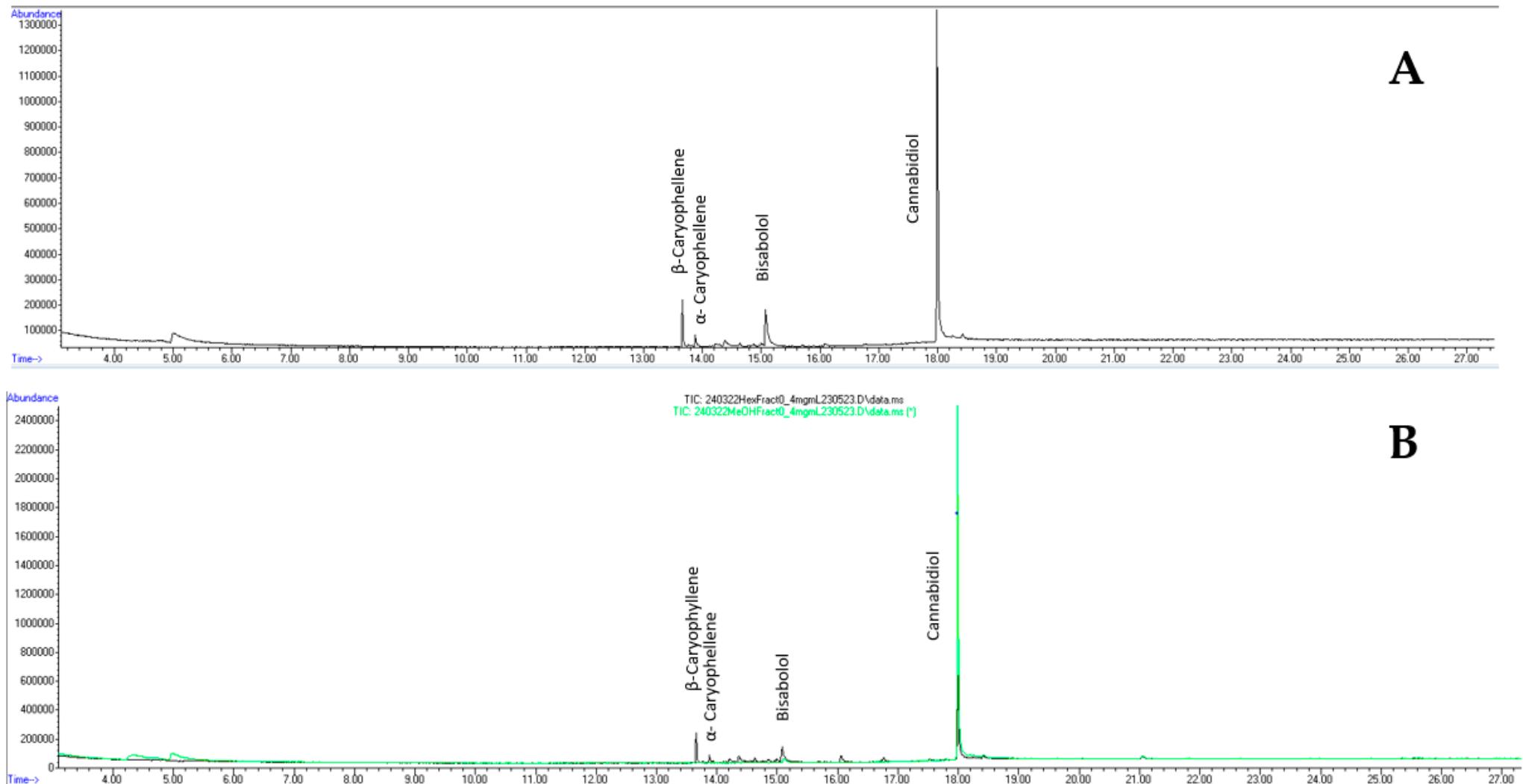


Figure 3. Gas chromatogram of dried hemp leaf extract (A), methanol fraction (green; B) and hexane fraction (black; B).

3.4. CBD Toxicity against 1st Instar Larvae

Authentic CBD produced concentration-dependent mortality in PS 1st instar larvae within 48 h that reached 100% and was indistinguishable from hemp leaf extract when standardized for CBD concentration (Figure 5). The LC₅₀ of CBD (0.59 ppm, 95% C.I.=0.52-0.67 ppm) was similar ($F = 0.95$, $p > 0.32$), to hemp leaf extract (0.53 ppm, 95% C.I.=0.46-0.62 ppm) in PS 1st instar *Ae. aegypti* (Figure 5). Moreover, like the hemp leaf extract, CBD elicited similar ($F = 0.26$, $p > 0.6$) larvicidal potency against PR (LC₅₀ = 0.83 ppm, 95% C.I.=0.69-0.99 ppm) and PS (LC₅₀ = 0.88 ppm, 95% C.I. = 0.75-1.04 ppm) strains (Figure 6).

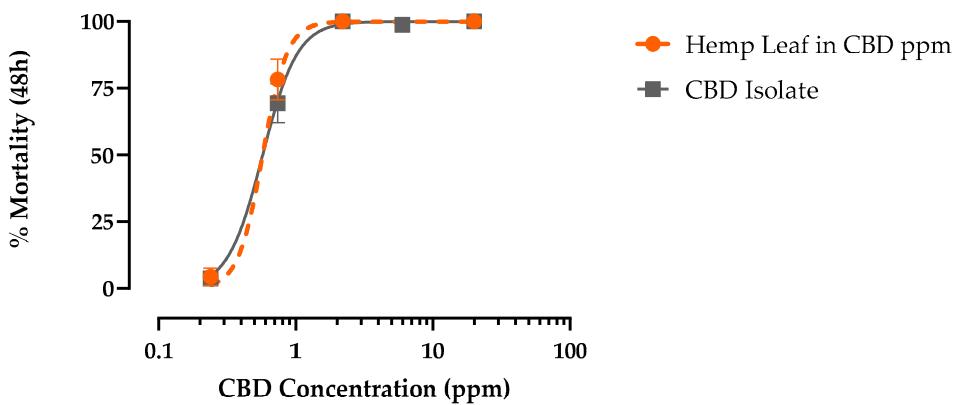


Figure 5. Concentration-response curves for 48 h larvicidal (1st instar) activity of CBD (grey) against pyrethroid-susceptible *Ae. aegypti*. Values plotted are means \pm SEM based on 12 replicates (n) of 6 larvae per concentration (0.2, 0.7, 2.2, 6.0, and 20 ppm).

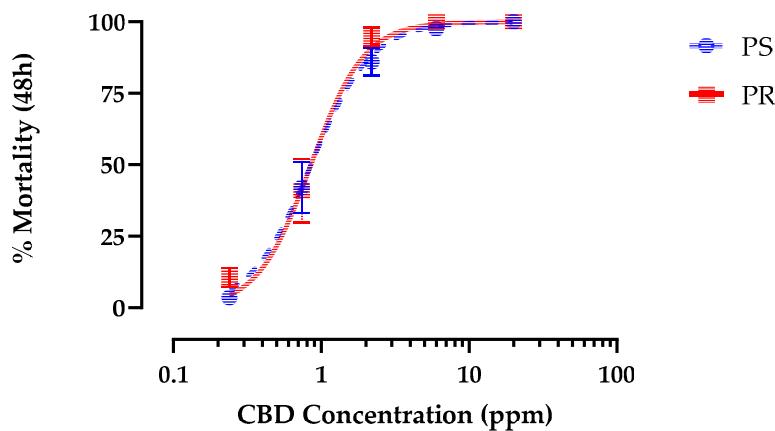


Figure 6. Concentration-response curves for 48 h larvicidal activity of CBD against pyrethroid-susceptible (blue) and pyrethroid-resistant (red) strains *Ae. aegypti*. Values are means \pm SEM based on 16 replicates of 6 larvae per concentration (0.2, 0.7, 2.2, 6.0, and 20 ppm).

4. Discussion

We demonstrated that hemp leaf extracts elicit concentration-dependent larvicidal activity against *Ae. aegypti*. These results are consistent with previous studies that have found concentration-dependent larvicidal activity of hemp extracts against other mosquitoes, including *Culex quinquefasciatus*, *Anopheles stephensi*, *An. gambiae*, *Ae. albopictus*, and *Ae. aegypti* [31–35,43]. In addition, we demonstrated that hemp leaf extracts exhibited similar toxic potency against a PR strain of *Ae. aegypti* that has both target-site (*kdr*) and metabolic resistance to pyrethroids [5,7,44,45]. This finding suggests that hemp leaf extracts have potential to bypass pyrethroid-resistance in mosquito larvae.

We also identified CBD as the primary active ingredient within the hemp extract responsible for its larvicidal activity. CBD was by far the most abundant compound in our hemp leaf extracts and methanol partitions as detected by GC-MS and proton NMR. Terpenes, which have previously been speculated as the primary larvicidal compounds in hemp extracts [36,46–50] were of nominal abundance. Moreover, the hexane fraction of our dried hemp extract, in which terpenes are expected to partition, was minimally larvicidal. Importantly, the larvicidal potency of CBD isolate matched that of the hemp leaf extract in both the PS and PR strains of *Ae. aegypti*. Moreover, the larvicidal potency of the methanol partition of the leaf extract was slightly weaker than that of the unpartitioned dried leaf extract, but far superior to that of the hexane partition of the leaf extract, consistent with the relative abundances of CBD in these samples.

Our findings regarding the toxicity of CBD to mosquitoes align well with prior research suggesting CBD has toxic, antifeedant, and/or growth inhibiting properties against other insects. For example, larvae of three economically important lepidopteran pests, the tobacco hornworm (*Manduca sexta*), the corn earworm (*Helicoverpa zea*) and the fall army worm (*Spodoptera frugiperda*), all showed reduced size, weight loss, and decreased consumption rates when feeding on diets supplemented with CBD [26,51,52]; additionally, *M. sexta* larvae that consumed high doses of CBD experienced higher mortality [26]. Likewise, larvae of the cabbage looper, *T. ni*, consumed less leaf area on CBD-dominant *C. sativa* genotypes compared to the cannabinoid free genotypes, leading to a decrease in larval mass and increase of mortality [25]. Furthermore, in larvae of three common stored product insect pests, the meal moth (*P. interpunctella*), saw toothed grain beetle (*O. surinamensis*), and flour beetle (*T. confusum*), mortality significantly increased after exposure to grains that had been sprayed with high doses of CBD oil [27].

The specific mode of action of CBD toxicity against mosquitoes and other insects is unknown. Intriguingly, insects are one of the few animal groups that do not possess canonical cannabinoid receptors [59,60]. However, at least in mammalian systems, CBD is known to modulate a wide range of biochemical targets, including orthologs of known insecticide targets, such as sodium channels, potassium channels, calcium channels, transient receptor potential channels, G protein-coupled receptors, and acetylcholinesterase [23,53–56]. Thus, it is likely to affect multiple biochemical targets and tissues in insects. Notably, in ventral chain ganglia of *M. sexta* larvae, CBD treatment delayed the onset of electrophysiological responses to electrical stimuli, but the magnitude of the responses was enhanced [30]. Whether these neuromodulatory effects of CBD on the lepidopteran central nervous system contribute to its toxicity remains to be determined.

5. Conclusions

The present research has shown that hemp leaf extracts have larvicidal activity against PS and PR strains of *Ae. aegypti* and that CBD is the principal active ingredient in the extract. Altogether, our study provides additional evidence supporting the notion by others that hemp is a valuable resource for developing novel insecticides to control mosquitoes [33,43,57]. Notably, hemp extracts have been shown to be non-toxic to select non-target invertebrates [43,58] and may be beneficial to honeybee workers [59]. Hemp is also an emerging, readily cultivated crop in the U.S. [13,16,19,60] and its leaves are often discarded [33,57]. Thus, availability of raw material would not appear limited as it can be for other sources of biopesticides [46]. Further research is needed to elucidate the mode of toxic action in larvae and the potential toxic and repellent effects of hemp extracts and CBD on adult female mosquitoes. It is also necessary to examine whether CBD elicits synergistic effects when applied with other established insecticides, as has been found for other natural products [53,61].

Supplementary Materials: The following supporting information can be downloaded at the website of this paper posted on Preprints.org.

Author Contributions: Conceptualization, E.J.M.R., P.M.P., P.L.P., L.C., and L.R.; methodology, E.J.M.R., P.M.P., P.L.P., L.R., L.C., and N.A.; validation, E.J.M.R., P.M.P., P.L.P., and L.R.; formal analysis, E.J.M.R., P.M.P. and P.L.P.; investigation, E.J.M.R., P.M.P., P.L.P.; resources, E.J.M.R., P.L.P., L.C., L.R., and P.M.P.; data curation, E.J.M.R., and P.M.P.; writing—original draft preparation, E.J.M.R., P.L.P., and P.M.P.; writing—review and editing, E.J.M.R., P.L.P., L.R., L.C., N.A., and P.M.P.; visualization, E.J.M.R., P.M.P. and P.L.P.; supervision,

P.M.P.; project administration, P.M.P.; funding acquisition, E.J.M.R. and P.M.P. All authors have read and agreed to the published version of the manuscript.

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Data Availability Statement: Data is contained within the article or supplementary material.

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Conflicts of Interest: The authors declare no conflicts of interest.

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