

Communication

Not peer-reviewed version

New Isomeric Dipyridothiazine Dimers – Synthesis, Structural Characterization and Cytotoxicity Screening

[Emilia Martula](#) , [Beata Morak-Młodawska](#) * , [Małgorzata Jeleń](#) , [Paulina Strzyga-Łach](#) , [Marta Struga](#) ,
[Katarzyna Żurawska](#) , Anna Kasprzycka

Posted Date: 6 May 2024

doi: 10.20944/preprints202405.0291.v1

Keywords: phenothiazines; diazaphenothiazines; 2D NMR; structural analysis; Way2Drug; cytotoxicity



Preprints.org is a free multidiscipline platform providing preprint service that is dedicated to making early versions of research outputs permanently available and citable. Preprints posted at Preprints.org appear in Web of Science, Crossref, Google Scholar, Scilit, Europe PMC.

Copyright: This is an open access article distributed under the Creative Commons Attribution License which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Disclaimer/Publisher's Note: The statements, opinions, and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions, or products referred to in the content.

Communication

New Isomeric Dipyrithiothiazine Dimers – Synthesis, Structural Characterization and Cytotoxicity Screening

Emilia Martula ¹, Beata Morak-Młodawska ^{2,*}, Małgorzata Jeleń ², Paulina Strzyga-Łach ³, Marta Struga ³, Katarzyna Żurawska ⁴ and Anna Kasprzycka ^{4,5}

¹ Doctoral School of The Medical University of Silesia, Poland; d201074@365.sum.edu.pl

² Department of Organic Chemistry, Faculty of Pharmaceutical Sciences, The Medical University of Silesia, Jagiellońska 4, 41-200 Sosnowiec, Poland; manowak@sum.edu.pl

³ Chair and Department of Biochemistry, Medical University of Warsaw, 02-097 Warsaw, Poland; paulina.strzyga-lach@wum.edu.pl (P.S.-Ł); marta.struga@wum.edu.pl (M.S.)

⁴ Biotechnology Centre, The Silesian University of Technology, Krzywoustego Street 8, 44-100 Gliwice, Poland; katarzyna.hopko@polsl.pl (K.Ż.); anna.kasprzycka@sum.edu.pl (A.K.)

⁵ Department of Organic Chemistry, Bioorganic Chemistry and Biotechnology, Faculty of Chemistry, Silesian University of Technology, Krzywoustego Street 4, 44-100 Gliwice, Poland

* Correspondence: bmlodawska@sum.edu.pl; Tel.: +48-32-364-16-04

Abstract: New dimers of selected dipyrithiothiazines with an *m*-xylene linker have been designed, synthesized and characterized. Their structure was identified using ¹H and ¹³C NMR as well as 2D NMR techniques (COSY, ROESY, HSQC and HMBC experiments) and HR MS spectrometry. The compounds were subjected to preliminary in silico biological profile tests using the Way2Drug server and analysis of the probable cytotoxic effect on various cancer cell lines. Preliminary in vitro cytotoxicity tests were performed against normal cell lines (HaCaT) and five cancer cell lines (SW480, SW620, MDA-MB-231, A-549, LN-229) showing moderate activity. An analysis of the structure-activity relationship was performed.

Keywords: phenothiazines; diazaphenothiazines; 2D NMR; structural analysis; Way2Drug; cytotoxicity

1. Introduction

Phenothiazines are tricyclic heterocyclic compounds in which two benzene rings have been joined by nitrogen and sulfur atoms to form a central 1,4-thiazine ring. There are completely synthetic compounds that have no precursor of their own in the animate natural world [1,2]. Although phenothiazine was discovered more than 140 years ago by German researcher August Bernthsen to this day the molecule is still very popular in the world of medical, pharmaceutical as well as chemical research [3–5].

In the 1950s, phenothiazines containing dialkylaminoalkyl substituents in their structure triggered revolutionary changes in psychiatry by showing phenomenal antipsychotic activity to treat schizophrenia or manic states. These drugs are still used in medical treatment to this day [1]. Scientific research in the area of phenothiazines is constantly being carried out, revealing many further interesting pharmacological activities, including: anticancer [6–10], antimicrobial [11,12], antifungal [13], antiviral [14,15], antioxidant [16] or reversal of multidrug resistance activities [17]. These topics are rich resulting in numerous review papers published each year around the world [18–22]. In 2006, English researcher Mitchell named the phenothiazine “parent molecule”, which fully reflects the application nature of this chemical compound [23].

The structure of phenothiazines is also modified at various levels, which leads to the creation of new, previously undescribed derivatives with interesting biological activities. [24]. Such a group of

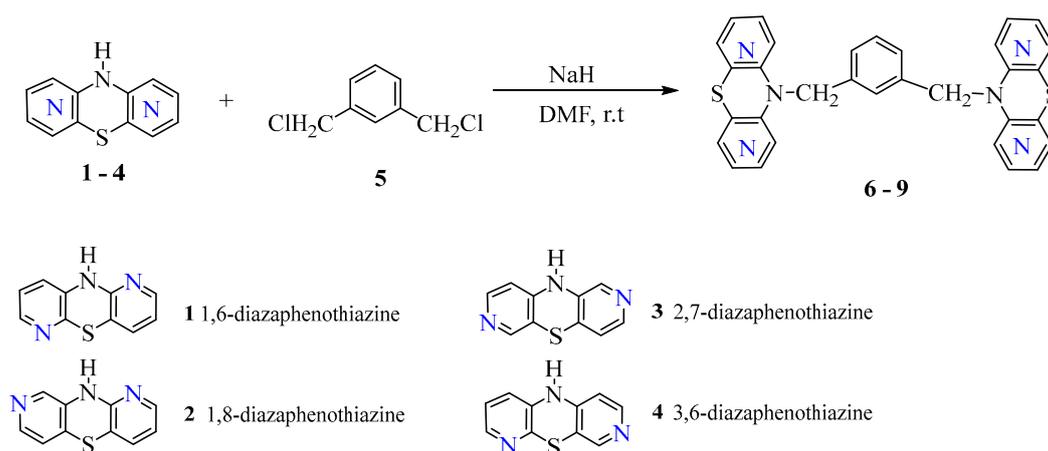
modified phenothiazines includes dipyridthiazines having two pyridine rings in their structure instead of benzene rings [21]. Dipyridthiazines show valuable anticancer activities involving induction of apoptosis via mitochondrial activation of it as well as partial DNA intercalation [25–27]. Dipyridthiazines also showed significant immunosuppressive and anti-inflammatory activity by inhibiting TNF alpha and interleukin 6 levels [28].

Dimeric derivatives of dipyridthiazines in which two dipyridthiazine units were connected by selected organic lines, such as *o*-xylene, *p*-xylene and 2,6-dimethylpyridine (lutidine), have recently shown highly promising anticancer activities in relation to colon (SW480) and breast (MCF 7) cancer lines. Additionally, these compounds were characterized by relatively low cytotoxicity in relation to normal myoblast cell (L6) lines [29]. This work is a continuation of this research. The synthesis of further new dimeric systems of dipyridthiazines (1,6-, 1,8-, 2,7- and 3,6-diazaphenothiazines **1-4**) has been described, in which two dipyridthiazine units have been linked by an *m*-xylene linker. The structure of new dimers was clearly confirmed by nuclear magnetic resonance spectroscopy (^1H , ^{13}C NMR), two-dimensional COSY, ROESY, HSQC, HMBC spectra and high-resolution mass spectrometry HR MS. In silico analyzes of probable molecular targets were performed using the Way2Drug server. The new derivatives were tested for anticancer activity in relation to cancer cell lines: human primary colon cancer (SW480), human metastatic colon cancer (SW620), human breast adenocarcinoma (MDA-MB-231), human lung carcinoma (A-549), human glioblastoma (LN-229), and to human immortal keratinocyte cell line from adult human skin (HaCaT).

2. Results & Discussion

2.1. Chemistry Part

The preparation of new dipyridthiazine dimers was carried out using previously synthesized, selected dipyridthiazines (10*H*-1,6-, 1,8-, 2,7- and 3,6-diazaphenothiazines **1-4**), for which the promising anticancer activities was determined [29]. In the next step, selected dipyridthiazines were subjected to alkylation reactions with α,α' -dichloro-*m*-xylene. The reactions were carried out in anhydrous DMF in the presence of the strong base NaH at room temperature, which led to the formation of the final dimers **6-9** (Scheme 1). The crude solid reaction products were purified using column chromatography. All new compounds were obtained in good yields (79-86%).



Scheme 1. Synthesis of isomeric dipyridthiazine dimers **6-9**.

It is worth noting that the obtained dimers are specific structural analogues of dimers with the 2,6-dimethylpyridine system, which were described in a previous publication [29].

2.2. Structural Identification Using Spectroscopic Methods

Identification of the structure of new organic compounds is a fundamental problem both in organic chemistry and in the drug design process [30,31]. Due to the fact that chemical reactions may be accompanied by rearrangements or parallel or subsequent reactions, proving the structure of the obtained product is essential. For this purpose, advanced NMR spectroscopic techniques and high-resolution HR MS mass spectrometry are used. The identification of the molecule's structure was carried out on the basis of ^1H , ^{13}C NMR spectra, two-dimensional experiments: 2D NMR correlations: COSY (COrrelation Spectroscopy), ROESY (Rotating frame nuclear Overhauser Effect Spectroscopy), HSQC (Heteronuclear Single Quantum Correlation), HMBC (Heteronuclear Multiple Bond Correlation) and HR MS mass spectrometry.

Structural analysis was performed for dimer **8**, which contains two 2,7-diazaphenothiazine units connected by an *m*-xylene liner. The ^1H NMR spectrum shows ten signals in the form of singlet (s), doublet (d) and degraded triplet (t) with different intensities, which correspond to proton signals in the molecule. In order to assign individual signals to appropriate protons, two-dimensional COSY and ROESY correlation spectra were performed. In the ROESY spectrum, spatial interactions of the CH_2 group with neighboring protons of both the *m*-xylene system and 2,7-diazaphenothiazine were observed (Figure 1.). The COSY spectrum shows the coupling of the CH_2 group at 5.06 ppm with a singlet at 6.94 ppm, which was assigned to the H_a proton of the *m*-xylene system, and with a doublet at 7.25 ppm with twice the intensity, which was assigned to H_b protons. In the ROESY spectrum, there is a visible correlation of the CH_2 group with a singlet at 7.7 ppm, which was assigned to the H_i proton of the 2,7-diazaphenothiazine system and with a doublet at 6.35 ppm, which corrects to the H_j signal. The ROESY spectrum also shows the interaction of the H_i proton with a doublet at 6.97 ppm, which is assigned to the H_k proton, which is coupled to the H_l proton at 8.10 ppm. In the COSY spectrum, it can be observed that the H_m proton is coupled to the H_n proton at 7.96 ppm. In this way, all signals were assigned to the protons present in compound **8**. The results of the analysis are summarized in Table 1. Then, the individual carbon signals in the ^{13}C NMR spectrum were assigned, first using the two-dimensional HSQC spectrum showing the carbons directly adjacent to the protons. Then, using the HMBC spectrum, quaternary carbons were assigned. The results are summarized in Table 1. The molecular mass and purity of compound were confirmed by high-resolution mass spectrometry (HR MS).

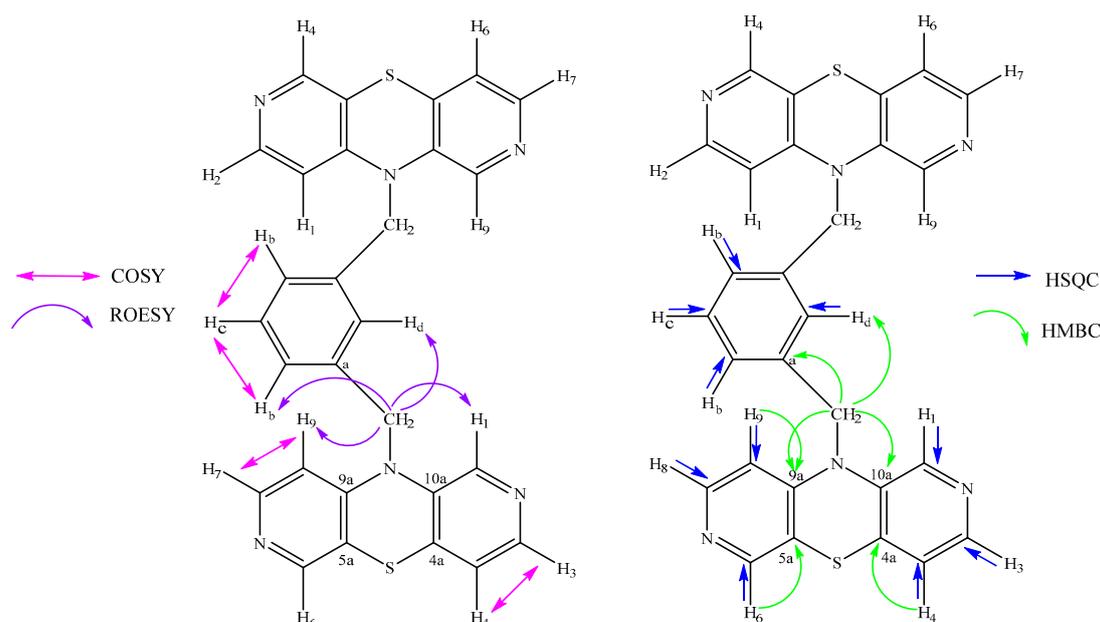


Figure 1. Assignment of ^1H and ^{13}C NMR and interactions of ^1H - ^1H (COSY and ROESY) and ^1H - ^{13}C (HSQC and HMBC) in compound **8** with particular emphasis on the interactions of the CH_2 group.

Table 1. Tabulated list of ^1H - ^1H and ^1H - ^{13}C correlations of a compound 8.

^1H NMR (ppm)	ROESY	COSY	^{13}C NMR (ppm)	HSQC	HMBC
			C _{CH2} 51.21	5.06-51.21	
			C ₉ 109.49	H ₉ 6.35-109.53	
			C _{9a} 117.19	H _d 6.94-121.74	H _c 7.45-C _a 134.46
CH ₂ 5.06			C _d 121.74	H ₄ 6.97-123.39	CH ₂ 5.06-C _{5a} 151.47
H ₉ 6.35	5.06-7.70		C ₄ 123.39	H _b 7.25-126.06	CH ₂ 5.06- C _{4a} 137.22
H _d 6.94	5.06-6.35	5.06-6.94	C _b 126.06	H _c 7.45-130.79	H ₃ 8.10- C _{10a} 133.35
H ₄ 6.97	5.06-7.25	5.06-7.25	C _c 130.79	H _i 7.70-135.56	H ₈ 7.98- C _{9a} 117.19
H _b 7.25	5.06-6.94	7.25-7.45	C _{10a} 133.35	H ₈ 7.98-147.14	
H _c 7.45	6.35-8.01	6.35-7.96	C _a 134.46	H ₆ 8.01-143.44	
H _i 7.70	7.70-6.97	6.97-8.10	C ₁ 135.56	H ₃ 8.10-145.05	
H ₈ 7.98	6.97-8.10		C _{4a} 137.22		
H ₆ 8.01			C ₆ 143.44		
H ₃ 8.10			C ₃ 145.05		
			C ₈ 147.14		
			C _{5a} 151.47		

2.3. In Silico Target Prediction

The new dimers **6-9** were analyzed in silico using the Way2Drug server, which includes the PASS (*Prediction of Activity Spectra for Substances*) application, which allows predicting the potential biological activities of new molecules and probability of cytotoxicity toward cancer cell lines [32,33]. The obtained results are summarized in Tables 2 and 3.

The results indicated that dimers **6-9** have a high probability of biological activity aimed at the anticancer potential associated with mechanism of stimulation of histone deacetylase, inhibition of glycosylphosphatidylinositol phospholipase D, which may directly lead to activation of the mitochondrial apoptosis pathway. Probability of action in neurodegenerative diseases, antipsychotic and antiallergic activity as well as cytochrome 450 inhibition has been demonstrated.

Table 2. Probability (%) of activities spectrum of dimers (6-9) using PASS program.

No.	Probability of Activity Spectrum				
6	(28%) Histone deacetylase stimulant	(77%) Glycosylphosphatidylinositol phospholipase D inhibitor	(47%) Transcription factor inhibitor	(33%) Alzheimer's disease treatment	(46%) Antiallergic
7	(29%) Mitochondrial processing peptidase inhibitor	(83%) Glycosylphosphatidylinositol phospholipase D inhibitor	(18%) Antipsychotic	(30%) Cytochrome P450 inhibitor	(10%) MAP3K8 inhibitor
8	(75%) Neurodegenerative diseases treatment	(84%) Glycosylphosphatidylinositol phospholipase D inhibitor	(65%) Histone deacetylase stimulant	(42%) Alzheimer's disease treatment	(41%) Cytochrome P450 inhibitor
9	(76%) Histone deacetylase stimulant	(66%) Antiallergic	(60%) Neurodegenerative diseases treatment	(58%) Alzheimer's disease treatment	(59%) Antiasthmatic

More importantly, in addition to the probable mechanisms of biological activity, the PASS program indicated the likelihood of cytotoxic effects on breast, kidney, colon, pancreatic and ovarian cancer cell lines, as well as on non-small cell lung cancer.

Table 3. The probable cytotoxic effect (%) on various cancer cell lines of compound 6-9.

No.	Probability of Cytotoxicity towards Cancer Cell Lines				
			Non-small cell		
6	Breast cancer	Melanoma	lung carcinoma	Colon carcinoma	Pancreatic carcinoma
	MDA-MB-468 (62%)	UACC-257(51%)	NCI-H322M (49%)	HCT-116 (42%)	YAPC (43%)
7	Breast cancer	Melanoma	Renal carcinoma	Colon carcinoma	Pancreatic carcinoma
	MDA-MB-468 (59%)	UACC-257 (55%)	786 (45%)	HCT-116 (39%)	YAPC (44%)
8	Breast cancer	Melanoma	Renal carcinoma	Colon carcinoma	Ovarian adenocarcinoma
	MDA-MB-468 (39%)	UACC-257 (54%)	786-0 (49%)	HCT-116 (55%)	OVCAR-4 (43%)
9	Breast cancer	Melanoma	Renal carcinoma	Colon carcinoma	Ovarian adenocarcinoma
	MDA-MB-468 (44%)	UACC-257 (49%)	786-0 (44%)	HCT-116 (54%)	OVCAR-4 (37%)

2.4. Biological Part – Analysis of Cytotoxicity towards Cancer and Normal Cells

Taking into account the results of in silico simulations and the previously promising anticancer activities and low cytotoxicity of the obtained dimers with an *o*-, *p*-xylene and 2,6-dimethylpyridine (lutidine) linker [29], the new derivatives 6-9 were tested for anticancer activity. The cytotoxicity and antiproliferative activity of new dimers was determined in vitro in relation to the following five cancer cell lines: human primary colon cancer (SW480), human metastatic colon cancer (SW620), human breast adenocarcinoma (MDA-MB-231), human lung carcinoma (A-549), human glioblastoma (LN-229), and to the normal line: human immortal keratinocyte cell line from adult human skin (HaCaT) using the MTT assay. The study used doxorubicin and cisplatin as reference drugs. Data are expressed as mean SD, IC₅₀ (μM) – the half-maximal growth inhibitory concentration after cultured the cells for 72 h with the individual compound and the SI (Selectivity Index), which was calculated using formula: SI = IC₅₀ for normal cell line/IC₅₀ cancer cell line.

The results obtained are summarized in the Table 4. The results showed that the compounds 6-9 were not toxic to the normal keratinocyte HaCaT cell line in the range of tested concentration. The analysis of anticancer activity showed that the tested derivatives were mostly inactive (IC₅₀> 100 μM). The derivative 6 showed poor activity in relation to SW480, MDA-MB-231 and A-549 cell lines (IC₅₀> 81-92 μM) and the derivative 7 in relation to MDA-MB-231 the cell line (IC₅₀> 67 μM).

Table 4. Cytotoxic activity (IC₅₀, μM) of studied compounds estimated by the MTT assay.

No.	Cancer Cells								Normal Cells		
	SW480		SW620		MDA-MB-231		A-549		LN-229		HaCaT
	IC ₅₀	SI	IC ₅₀								
6	92.6 ± 5.31	1.08	>100	1	83.2 ± 3.38	1.20	81.5 ± 5.71	1.22	>100	1	>100
7	>100	0.93	>100	0.93	67.3 ± 4.13	1.38	>100	0.93	>100	0.93	92.7 ± 5.00
8	>100	1	>100	1	>100	1	>100	1	>100	1	>100
9	>100	1	>100	1	96.5 ± 1.37	1.03	>100	1	>100	1	>100
Doxorubicin	0.7 ± 0.1	0.4	0.3 ± 0.1	1.0	1.6 ± 0.23	0.19	0.2 ± 0.09	1.5	1.1 ± 0.12	0.27	0.3 ± 0.1
Cisplatin	10.4 ± 0.9	0.6	6.7 ± 1.1	0.9	7.8 ± 0.98	0.81	3.2 ± 1.24	5.08	2.6 ± 0.15	2.42	6.3 ± 0.7

The obtained cytotoxicity results were surprising when analyzed in relation to the previously obtained results of anticancer activity of structural analogues containing the 2,6-dimethylpyridine

(lutidine) system Figure 2 [29]. These compounds showed activity in the range of IC_{50} = 0.1-20 μ M towards MCF and IC_{50} = 4.5-25 μ M SW480 cell lines. This led us to consider the reason for the deactivation of the obtained derivatives 6-9.

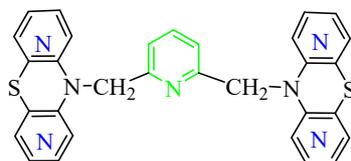


Figure 2. Structure of the previously described isomeric anti-cancer active dimers with the 2,6-dimethylpyridine (lutidine) system [29].

For this purpose, measurements of the electron densities of the selected dimer **8** containing 2,7-diazaphenothiazine in its structure and its analogue with the lutidine system were performed using the Gaussian program [34]. A visualization of the obtained results is presented in Figures 3 and 4 using Avogadro program [35]. These measurements showed that the presence of the lutidine system causes a differentiation of electron charges on thiazine nitrogen atoms in 1,4-thiazine rings directly connected to the CH_2 moiety (Figure 4). Additionally, the lutidine system contains a basic nitrogen atom with a lone electron pair, which causes the entire molecule to be enriched with electrons. This situation is not observed in dimer **8** with *m*-xylene linker (Figure 3).

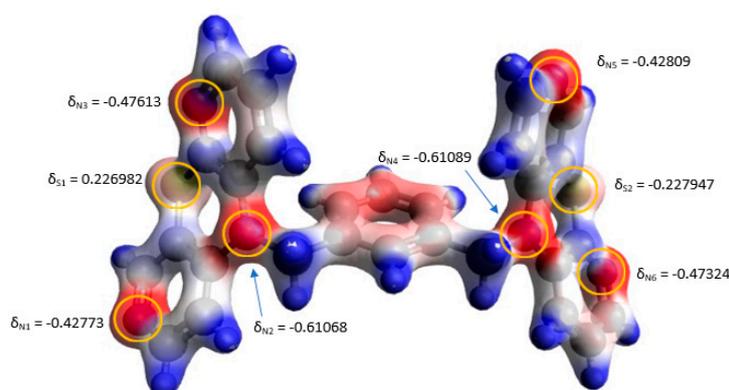


Figure 3. Visualization of the electron density of the compound **8** (electron density was calculated in the Gaussian program, visualized in the Avogadro program). Electron density is represented by a color gradient, with red representing higher electron density and dark blue representing lower electron density.

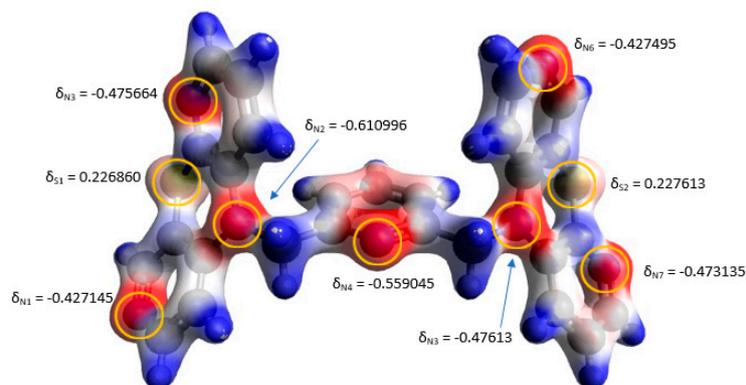


Figure 4. Visualization of the electron density of the structural analogue of compound **8** with a pyridine ring as a linker (electron density was calculated in the Gaussian program, visualized in the

Avogadro program). Electron density is represented by a color gradient, with red representing higher electron density and dark blue representing lower electron density.

Additionally, bond length measurements were made, with particular attention paid to the connecting linkers. In the dimer containing lutidine, there are CH₂-C_{lutidine} bonds with a length of 1.516 Å and a characteristic C_{lutidine}-N_{lutidine} bond with a length of 1.362 Å. These bonds are essentially shorter than the analogous bonds found in dimer **8** (CH₂-C_{m-xylene} a length of 1.521 Å, C_{m-xylene} - C_{m-xylene} bond with a length of 1.405 Å) (Supplementary materials). The performed structural analysis explains the weak anticancer activity of the obtained dimers with the *m*-xylene system, while also showing that subtle differences in bond lengths and electron density have a significant impact on the biological activity of the new compounds.

3. Materials and Methods

3.1. Chemistry

The NMR spectra were recorded on Bruker Avance spectrometers (¹H at 600 MHz, ¹³C at 150 MHz) in CDCl₃ and DMSO-d₆. Two-dimensional COSY, ROESY, HSQC and HMBC spectra of compound **8** were recorded on a Bruker Avance spectrometer at 600 MHz. The HR MS spectra (ESI – Electro Spray Ionisation) was run on a Bruker Impact II. The thin layer chromatography was performed on silica gel 60 F₂₅₄ (Merck 1.05735) with CHCl₃-EtOH (10:1 v/v) and on Al₂O₃ 60 F₂₅₄ neutral (type E) (Merck 1.05581) with CHCl₃-EtOH (10:1 v/v) as eluents. Melting points were determined in open capillary tubes on a Boetius melting point apparatus.

The parent starting dipyridthiazines **1-4** (**Scheme 1**) were prepared according to reference [29].

3.1.1. General Procedure for Synthesis of Compounds (6-9)

To a solution of selected diazaphenothiazine (**1-4**) (0.201 mg, 1 mmol) in dry DMF (10 mL), NaH (0.028 g, 1.2 mmol, 60% NaH in mineral oil was washed out with hexane) was added. The reaction mixture was stirred at room temperature for 1 h and then linker α,α' -dichloro-*m*-xylene (0.5 mmol) was added and the stirring was continued for 24 h. The mixture was poured into water (15 ml), extracted with CHCl₃ (3 × 10 ml) and dried using Na₂SO₄. The obtained product was purified by column chromatography (aluminum oxide, CHCl₃-Ethanol 10:1) to give:

1,3-bis((10*H*-dipyrido [2,3-*b*:2',3'-*e*][1,4]thiazin-10-yl)methyl)benzene (**6**)

(214 mg, 85%) mp = 268-270 °C

¹H NMR (CDCl₃) δ : 5.16 (s, 2H, CH₂), 6.70 (dd, 1H), 6.78 (m, 1H), 6.84 (m, 1H), 6.93 (s, 1H), 7.18 (d, 1H), 7.31 (t, 1H), 7.37 (m, 1H), 7.68 (m, 1H), 7.82 (d, 1H). ¹³C NMR: 47.58, 79.65, 115.14, 119.10, 121.95, 122.56, 124.59, 125.45, 129.28, 135.11, 137.54, 142.85, 143.17, 145.41, 151.89, 162.77. HR MS (EI) *m/z* for: [C₂₈H₂₁N₆S₂ + H] calc. 505.1264, Found: 505.1283 (100%).

1,3-bis((10*H*-dipyrido[3,2-*b*:3',4'-*e*][1,4]thiazin-10-yl)methyl)benzene (**7**)

(209 mg, 83%) mp = 262-264 °C

¹H NMR (CDCl₃) δ : 5.42 (s, 2H, CH₂), 6.90 (m, 1H), 7.17 (m, 2H), 7.20 (m, 1H), 7.21 (m, 1H), 7.29 (s, 2H), 8.05 (m, 1H), 8.14 (d, 1H), 8.19 (s, 1H). ¹³C NMR: 48.02, 111.95, 120.20, 121.59, 125.52, 126.67, 128.23, 130.03, 135.05, 140.91, 143.84, 147.14, 151.13, 152.90. HR MS (EI) *m/z* for: [C₂₈H₂₁N₆S₂ + H] calc. 505.1264, Found: 505.1283 (100%).

1,3-bis((10*H*-dipyrido[3,4-*b*:3',4'-*e*][1,4]thiazin-10-yl)methyl)benzene (**8**)

(216 mg, 86%) mp = 256-257 °C

¹H NMR (CDCl₃) δ : 5.06 (s, 2H, CH₂), 6.35 (d, 1H, H₉), 6.94 (s, 1H, H_{d-m-xylene}), 6.97 (d, 1H, H₄), 7.25 (d, 1H, H_{b-m-xylene}), 7.45 (t, 1H, H_{c-m-xylene}), 7.70 (s, 1H, H_i), 7.99 (d, 1H, H₈), 8.01 (s, 1H, H₆), 8.10 (d, 1H, H₃). ¹³C NMR: 51.08 C_{CH₂}, 109.41 C₉, 117.19 C_{9a}, 121.56 C_d, 123.48 C₄, 126.06 C_b, 130.79 C_c, 133.35 C_{10a}, 134.46 C_a, 135.56 C₁, 137.22 C_{4a}, 143.44 C₆, 145.70 C₃, 147.14 C₈, 151.47 C_{5a}. HR MS (EI) *m/z* for: [C₂₈H₂₁N₆S₂ + H] calc. 505.1264 Found: 505.1229 (100%).

1,3-bis((10*H*-dipyrido[2,3-*b*:4',3'-*e*][1,4]thiazin-5-yl)methyl)benzene (**9**)

(199 mg, 79%), mp = 288-290 °C

^1H NMR (CDCl_3) δ : 5.04 (s, 2H, CH_2), 6.44 (d, 1H), 6.81 (m, 1H), 6.88 (m, 1H), 6.97 (s, 1H), 7.27 (m, 1H), 7.44 (t, 1H), 7.90 (d, 1H), 7.95 (d, 1H), 8.02 (s, 1H). ^{13}C NMR: 50.88, 79.56, 110.32, 123.30, 123.52, 124.36, 126.67, 130.56, 136.07, 142.94, 145.93. HR MS (EI) m/z for: $[\text{C}_{28}\text{H}_{21}\text{N}_6\text{S}_2 + \text{H}]$ calc. 505.1264, Found: 505.1299 (100%).

3.2. *In Silico* Target Prediction

The prediction of biological targets and cytotoxicity towards cancer cell lines was performed using the Way2Drug server [32,33]. This server analyzes the biological activity of an organic drug based on molecular recognition against its database.

3.3. Biological Evaluation

3.3.1. Cell Line and Culture

Human primary colon cancer (SW480) and metastatic colon cancer (SW620), human breast adenocarcinoma (MDA-MB-231), human lung carcinoma (A-549), human glioblastoma (LN-229), and human immortal keratinocyte (HaCaT) cell lines were obtained from the American Type Culture Collection (ATCC) in Rockville, USA. These cell lines were cultured in the recommended medium according to ATCC protocols: SW480 and SW620 in Minimum Essential Medium (MEM), and MDA-MB-231, A-549, LN-229, and HaCaT cells in Dulbecco's Modified Eagle Medium (DMEM). The culture medium was supplemented with 10% fetal bovine serum (FBS), penicillin (100 U/mL), and streptomycin (100 $\mu\text{g/mL}$). Cells were maintained in a humidified incubator at 37°C with 5% CO_2 . Passaging was performed when cells reached 80%–90% confluence using 0.25% trypsin (Gibco Life Technologies, Gibco, Grand Island, NY, USA) treatment before being utilized for experiments.

3.3.2. MTT Cell Viability Assay

Cell viability was determined through enzymatic conversion of 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide salt (MTT) to insoluble formazan crystals by mitochondrial dehydrogenases present in viable cells. Cells were seeded into 96-well plates at a density of 1×10^4 cells per well and allowed to adhere for 24 hours at 37°C in a CO_2 incubator. Following the 24-hour incubation period, the culture medium was replaced with fresh medium, and cells were exposed to varying concentrations of the compounds (ranging from 10 to 100 μM), then further incubated for 72 hours at 37°C in a CO_2 incubator. Untreated cells served as controls. Subsequently, MTT solution (0.5 mg/mL in serum-free medium) was added, and samples were incubated for 4 hours at 37°C in a CO_2 incubator. Following this, the medium was removed, and formazan crystals were solubilized by adding an isopropanol-DMSO mixture (1:1). The absorbance of the dissolved crystals was measured at a wavelength of 570 nm using a UVM 340 reader (ASYS Hitech GmbH, Austria). IC_{50} values were calculated using GraphPad Prism 8 software (GraphPad Software).

3.4. Quantum Mechanical Calculations

Calculation of the electron density and bond lengths were carried out with B3LYP functional and 6-31+G(d,p) basis set, using Gaussian16 software [34] and visualized using the Avogadro program [35].

3.5. The Statistical Analysis

The statistical analysis was performed using GraphPad Prism 9 software (Graph Pad Software, San Diego, CA, USA). The results were reported as mean \pm SD from a minimum of three independent experiments. Statistical significance of differences between values was assessed using analysis of variance with Dunnett's multiple comparison post hoc test, and significance was defined as $p < 0.05$.

4. Conclusions

An effective synthesis of new dimers of selected dipyrithiazines with an *m*-xylene linker was presented. The structure of new derivatives has been identified using ¹H and ¹³C NMR and advanced techniques 2D NMR (COSY, ROESY, HSQC and HMBC experiments). The compounds were subjected to preliminary in silico screening using the Way2Drug server to determine molecular targets and the probability of cytotoxicity towards various types of cancer cells. In the next step cytotoxicity studies have been performed against normal cell lines (HaCaT) and five cancer cell lines (SW480, SW620, MDA-MB-231, A-549, LN-229). The compounds showed weak anticancer activity and low cytotoxicity compared to previously described structural analogues with the 2,6-dimethylpyridine (lutidine) system [29]. The obtained results of cytotoxicity of new dimers are in opposition to the results of in silico tests, which confirms the validity of performing experimental tests. To explain the low activity of the obtained derivatives, quantum mechanical calculations regarding electron densities and bond lengths were performed to explain structural differences, which directly affect the conformations of the molecule, as well as interactions with molecular targets.

Supplementary Materials: The following supporting information can be downloaded at the website of this paper posted on Preprints.org. The following supporting information of spectra analysis and in silico analysis are downloaded.

Author Contributions: Conceptualization, B.M.-M. and E.M.; methodology, E.M., P.S., M. S., K.H, A.K; software, E.M., B.M.-M. and M.J., P.S., M. S., K.H, A.K.; validation, E.M.; formal analysis, E.M., B.M.-M., M.J., M.J., P.S., M. S., K.H, A.K; investigation, E.M., B.M.-M.; resources, E.M., B.M.-M., M.J., P.S., M. S., K. H, A.K; data curation, E.M., B.M.-M.; writing—original draft preparation, E.M. and B.M.-M.; writing—review and editing, B.M.-M., E.M., M. J., visualization, B.M.-M.; supervision, E.M. and B.M.-M.; project administration, B.M.-M.; funding acquisition, E.M., B.M.-M. All authors have read and agreed to the published version

Funding: Emilia Martula was supported by a research subsidy from the Medical University of Silesia in Katowice (grant BNW-2-015/N/3/F); APC was funded by the Metropolis of Upper Silesia and Zagłębie Basin as part of the implementation of the project entitled “Support for scientific activity of doctoral students and employees at the doctoral level of the Silesian Medical University in Katowice”, which was implemented within the framework of the Metropolitan Science Support Fund program for 2022–2024 (grants agreement no. RW/27/20243, PCTT/621/2024).

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Data is contained within the article and Supplementary Materials.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. López-Muñoz F, Alamo C, Cuenca E, Shen W.W, Clervoy P, Rubio G. History of the Discovery and Clinical Introduction of Chlorpromazine. *Annals of Clinical Psychiatry* **2005**, *17*, 113–135, doi:10.1080/10401230591002002.
2. Jaszczyszyn A, Gašiorowski K, Świątek P, Malinka W, Cieślik-Boczula K, Petrus J, Czarnik-Matusewicz B. Chemical structure of phenothiazines and their biological activity. *Pharmacological Reports* **2012**, *64*, 16–23, doi:10.1016/s1734-1140(12)70726-0.
3. Otręba M, Stojko J, Rzepecka-Stojko A. Phenothiazine derivatives and their impact on the necroptosis and necrosis processes. *A review Toxicology* **2023**, *492*, 1-9, doi:10.1016/j.tox.2023.153528.
4. Kumar A, Vigato C, Boschi D, Lolli M.L, Kumar D. Phenothiazines as anti-cancer agents: SAR overview and synthetic strategies. *European Journal of Medicinal Chemistry* **2023**, *254*, 115337, doi:10.1016/j.ejmech.2023.115337.
5. Swoboda D, Nycz J.E, Karaush-Karmazin N, Minaev B, Książek M, Kusz J, Podsiadły R. Synthesis and Spectroscopic Characterization of Selected Phenothiazines and Phenazines Rationalized Based on DFT Calculation. *Molecules* **2022**, *27*, 7519, 1-26, doi: 10.3390/molecules27217519.
6. Li J.J. *Heterocyclic Chemistry in Drug Discovery*. Wiley: Hoboken **2013**, *52*, 1–16, doi:10.1002/anie.201308187.
7. Otręba M, Stojko J, Rzepecka-Stojko A. The role of phenothiazine derivatives in autophagy regulation: A systematic review. *Journal of Applied Toxicology* **2023**, *43*, 474–489, doi:10.1002/jat.4397.

8. Lopes R.M, Souza A.C, Otręba M, Rzepecka-Stojko A, Tersariol I, Rodrigues T. Targeting autophagy by antipsychotic phenothiazines: potential drug repurposing for cancer therapy. *Biochemical Pharmacology* **2024**, *222*, 116075, doi:10.1016/j.bcp.2024.116075.
9. Xu F, Xi H, Liao M, Zhang Y, Ma H, Wu M, Xue Q, Sun H, Zhang Y, Xia Y. Repurposed antipsychotic chlorpromazine inhibits colorectal cancer and pulmonary metastasis by inducing G2/M cell cycle arrest, apoptosis, and autophagy. *Cancer Chemotherapy and Pharmacology* **2022**, *89*, 331–346, doi:10.1007/s00280-021-04386-z.
10. Li A, Chen X, Jing Z, Chen J. Trifluoperazine induces cellular apoptosis by inhibiting autophagy and targeting NUPR1 in multiple myeloma. *FEBS Open Bio* **2020**, *10*, 2097–2106, doi:10.1002/2211-5463.12960.
11. Posso M.C, Domingues F.C, Ferreira S, Silvestre S. Development of Phenothiazine Hybrids with Potential Medicinal Interest: A Review. *Molecules* **2022**, *27*, 276, doi:10.3390/molecules27010276.
12. Rineh A, Dolla N.K, Ball A.R, Magana M, Bremner J.B, Hamblin M.R, Tegos G.P, Kelso M.J. Attaching the NorA Efflux Pump Inhibitor INF55 to Methylene Blue Enhances Antimicrobial Photodynamic Inactivation of Methicillin-Resistant Staphylococcus aureus in Vitro and in Vivo. *ACS Infectious Diseases* **2017**, *3*, 756–766, doi:10.1021/acsinfecdis.7b00095.
13. Brilhante R.S.N, Gotay W.J.P, Pereira V.S, de Oliveira J.S, Pereira-Neto W.A, Castelo-Branco D.d.S.C.M, de Aguiar Cordeiro R, Sidrim J.J.C, Rocha M.F.G. Antifungal Activity of Promethazine and Chlorpromazine Against Planktonic Cells and Biofilms of Cryptococcus neoformans/Cryptococcus gattii Complex Species. *Medical Mycology* **2020**, *58*, 906–912, doi: 10.1093/mmy/myz140.
14. Piccini L.E, Castilla V, Damonte E.B. Inhibition of dengue virus infection by trifluoperazine. *Archives of Virology* **2022**, *167*, 2203–2212, doi:10.1007/s00705-022-05555-y.
15. Simanjuntak Y, Liang J.J, Lee Y.L, Lin Y.L Repurposing of prochlorperazine for use against dengue virus infection. *Journal of Infectious Diseases* **2015**, *211*, 394-404, doi:10. 1093/ infdis/ jiu377.
16. Egbujor M.C, Tucci P, Buttari B, Nwobodo D.C, Marini P, Saso L. Phenothiazines: Nrf2 activation and antioxidant effects. *Journal of Biochemical and Molecular Toxicology* **2024**, *38*, 23661, doi:10.1002/jbt.23661.
17. Takács D, Csonka Á, Horváth Á, Windt T, Gajdács M, Riedl Z, Hajós G, Amaral L, Molnár J, Spengler G. Reversal of ABCB1-related Multidrug Resistance of Colonic Adenocarcinoma Cells by Phenothiazines. *Anticancer Research* **2015**, *35*, 3245–3251.
18. Mosnaim A.D, Ranade V.V, Wolf M.E, Puente J, Valenzuela M.A. Phenothiazine molecule provides the basic chemical structure for various classes of pharmacotherapeutic agents. *American Journal of Therapeutics* **2006**, *13*, 261–273, doi:10.1097/01.mjt.0000212897.20458.63.
19. Ohlow M, Moosmann B. Phenothiazine: The seven lives of pharmacology's first lead structure. *Drug Discovery Today* **2011**, *16*, 119–131, doi:10.1016/j.drudis.2011.01.001.
20. Pluta K, Morak-Młodawska B, Jeleń M. Synthesis and properties of diaza-, triaza- and tetraazaphenothiazines. *Journal of Heterocyclic Chemistry* **2009**, *46*, 355-391, doi:10.1002/jhet.42.
21. Morak-Młodawska B, Pluta K, Jeleń M. Phenothiazines Modified with the Pyridine Ring as Promising Anticancer Agents. *Life* **2021**, *11*, 206, doi:10.3390/life11030206.
22. Kumar A, Vigato C, Boschi D, Lolli M.L, Kumar D. Phenothiazines as anti-cancer agents: SAR overview and synthetic strategies. *European Journal of Medicinal Chemistry* **2023**, *254*, 115337, doi:10.1016/j.ejmech.2023.115337.
23. S.C. Mitchell. Phenothiazine: the parent molecule. *Current Drug Targets* **2006**, *7*, 1181–1189, doi:10.2174/138945006778226552.
24. Gupta R.R, Kumar M. Synthesis, properties and reactions of phenothiazines. Phenothiazine and 1,4-Benzothiazines—Chemical and Biological Aspect. *Elsevier* **1988**, *108*, 1-161.
25. Zhang J, Chen M, Wenzhi Z, Okechukwu Nwabueze P, Morak-Młodawska B, Pluta K, Jeleń M, Md Akim A, Ang K.P, Ooi K.K. 10H-3,6-Diazaphenothiazines Induce G2/M Phase Cell Cycle Arrest, Caspase-dependent Apoptosis and Inhibits Cell Invasion of A2780 Ovarian Carcinoma Cells through Regulation on NF-κB and [BIRC6-XIAP] Complexes. *Drug Design, Development and Therapy* **2017**, *11*, 3045-3063, doi:10.2147/DDDT.S144415.
26. Skonieczna M, Kasprzycka A, Jeleń M, Morak-Młodawska B. Tri- and Pentacyclic Azaphenothiazine as Pro-Apoptotic Agents in Lung Carcinoma with a Protective Potential to Healthy Cell Lines. *Molecules* **2022**, *27*, 5255, doi:10.3390/molecules27165255.

27. Sochacka J, Pacholczyk M, Jeleń M, Morak-Młodawska B, Pluta K. Interaction of new tri-, tetra-, and pentacyclic azaphenothiazine derivatives with calf thymus DNA: Spectroscopic and molecular docking studies. *Spectrochimica Acta, Part A: Molecular and Biomolecular Spectroscopy* **2021**, 262, doi:10.1016/j.saa.2021.120105.
28. Morak-Młodawska B, Pluta K, Zimecki M, Jeleń M, Artym J, Kocięba M. Synthesis and selected immunological properties of 10-substituted 1,8-diazaphenothiazines. *Medicinal Chemistry Research* **2015**, 24, 1408-1418, doi:10.1007/s00044-014-1220-9.
29. Martula E, Morak-Młodawska B, Jeleń M, Okechukwu P, Balachandran A., Tehirunavukarasu P, Anamalay K, Ulaganathan V. Synthesis and structural characterization of novel dimers of dipyrithiazine as promising antiproliferative agents. *Molecules* **2023**, 28, 7662, doi:10.3390/molecules28227662.
30. Kiemle D, Webster F.X, Silverstein R.M, Bryce D.L. *Spectrometric Identification of Organic Compounds, 8th Edition*. John Wiley & Sons Inc, **2014**, ISBN: 978-0-470-61637-6.
31. Hermann Ch.K.F, Morrill T.C, Shriner R.L, Fuson R.C. *Systematic Identification of Organic Compounds, Ninth Edition*. Wiley, **2023**, ISBN: 9781119799665.
32. Way2Drug Understanding Chemical-Biological Interactions, Predictive Services. Available online: www.way2drug.com/passonline/index.php (accessed on 18 December 2023).
33. Druzhilovskiy D.S, Rudik A.V, Filimonov D.A, Glorizova T.A, Lagunin A.A, Dmitriev A.V, Pogodin P.V, Dubovskaya V.I, Ivanov S.M, Tarasova O.A, Bezhentsev V.M, Murtazaliev K.A, Semin M.I, Maiorov I.S, Gaur A.S, Sastry G.N, Poroikov V.V. Computational platform Way2Drug: from the prediction of biological activity to drug repurposing. *Russian Chemical Bulletin* **2017**, 66, 1832–1841, doi: 10.1007/s11172-017-1954-x.
34. Gaussian 16, Revision C.01, Gaussian, Inc., Wallingford CT, 2019.
35. Avogadro: an open-source molecular builder and visualization tool. Version 1.2.0. <http://avogadro.cc/> (accessed on 12 March 2024)

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.