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# Clinical Correlation of Transcription Factor SOX3 in Cancer: Unveiling Its Role in Tumorigenesis

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Posted Date: 18 April 2024

doi: 10.20944/preprints202404.1199.v1

Keywords: SOX3; Cancer; Apoptosis; EMT; Invasion; Migration; Cell cycle; Proliferation; Prognosis; Therapeutics



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# Clinical Correlation of Transcription Factor SOX3 in Cancer: Unveiling Its Role in Tumorigenesis

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Abstract Members of the SOX (SRY-related HMG-box) family of transcription factors are crucial for embryonic development and cell fate determination. This review investigates the role of SOX3 in cancer, as aberrations in SOX3 expression have been implicated in several cancers, including osteosarcoma, breast, esophageal, endometrial, ovarian, gastric, hepatocellular carcinomas, glioblastoma, and leukemia. These dysregulations modulate key cancer outcomes such as apoptosis, epithelial-mesenchymal transition (EMT), invasion, migration, cell cycle, and proliferation, contributing to cancer development. SOX3 exhibits varied expression patterns correlated with clinicopathological parameters in diverse tumor types. This review aims to elucidate the nuanced role of SOX3 in tumorigenesis, correlating its expression with clinical and pathological characteristics in cancer patients and cellular models. By providing a comprehensive exploration of SOX3 involvement in cancer, this review underscores the multifaceted role of SOX3 across distinct tumor types. The complexity uncovered in SOX3 function emphasizes the need for further research to unravel its full potential in cancer therapeutics.

Keywords: SOX3; Cancer; Apoptosis; EMT; Invasion; Migration; Cell cycle; and proliferation

# 1. Introduction

Transcription factors (TFs) are pivotal in shaping cellular identities, directing cell differentiation, and orchestrating complex temporal-spatial gene expression profiles during embryonic development [1–3]. The discovery of *SRY* (sex-determining region Y), the founder of the SOX(SRY-related HMG box) protein family of TFs, marked a significant milestone in understanding TF biology in the context of sex determination [4]. The high-mobility group (HMG) box domain within SRY is highly conserved and shared with all 20 SOX protein members and has since gained prominence as a versatile DNA-binding motif [5–7].

SOX proteins are classified into nine groups based on phylogenetic analysis, each characterized by distinct functions and target genes [8,9]. Among these groups, SOXB1 genes (SOX1, 2, and 3) play a pivotal role in sustaining stem cell proliferation and maintaining multipotent characteristics. Conversely, other groups, such as SOXB2 (SOX14 and 21), SOXD (SOX5, 6, and 13), and SOXE (SOX8, 9, and 10), function as inhibitors of proliferation, promoting lineage-specific cell identities. In contrast, the SOXC group (SOX4, 11, and 12) drives proliferation and terminal differentiation across diverse lineages [58]. These proteins harbor an HMG-box DNA binding domain that facilitates nuclear localization but also alters DNA architecture by inducing a bend that leads to the recruitment of

additional proteins such as p53, Nanog, OCT4, and Wnt/ $\beta$ -catenin required to modulate cellular behavior and fate [10–14].

In recent years, aberrant expression and function of SOX proteins have emerged as a significant contributor to multiple cancer types. These TFs influence cell differentiation, proliferation, migration, invasion, and metastasis in several tumor types [15–20]. The pleiotropic nature of SOX proteins is underscored by their ability to regulate different gene sets in diverse cellular contexts and tissues [9,21]. Adding to the complexity of cancer research is the variability observed in tumor types, genetic mutations, tumor locations, stages, patient characteristics, treatment responses, and drug resistance. Within a single tumor, there is often considerable regional cellular heterogeneity, meaning that different tumor regions have distinct genetic profiles and behavioral characteristics, complicating treatment efficacy [22–25]. This complexity arises from the intricate interplay of genetic mutations and molecular interactions within the tumor microenvironment [26].

While there has been considerable progress in understanding the roles of SOX proteins in cancer, identifying specific SOX factors as tumor suppressors or carcinogenic modulators remains challenging. Significantly, SOX proteins within a single family group often exhibit functional overlap or redundancy in the discreet cellular environment. SOX2, a member of the SOX B1 group, has garnered significant attention as a prognostic, diagnostic, and therapeutic target in various cancer types [27–32]. However, limited attention has been devoted to another SOX B1 group member, SOX3, leaving its role in cancer relatively unexplored.

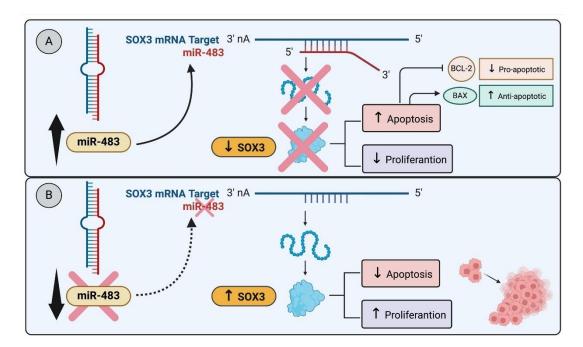
SOX3 expression in non-cancerous and cancerous tissues is widespread [33–36]. This review will discuss the involvement of SOX3 in multiple human neoplasms and present the SOX3 clinical correlation in concert with tumor behavior. Our objective is to shed light on the role of SOX3 in tumorigenesis by examining gene and protein expression patterns in clinical specimens and *in vitro* and *in vivo* models across various tumor types. Our review seeks to uncover the multifaceted role of SOX3 in cancer progression, potentially opening new avenues for understanding and targeting this TF in cancer therapeutics.

#### 2. SOX3 Involvement and Regulation of Cancer Hallmarks:

#### 2.1. SOX3 and Cell Death by Apoptosis

Apoptosis is an active, ATP-dependent form of cell death initiated through the activation of proteolytic cascades involving caspases. This process leads to both molecular and morphological alterations within cells, serving as a regulated process to eliminate cells with DNA damage, thus preventing the accumulation of mutations that could potentially lead to cancer [37]. The significance of apoptosis lies in its ability to safeguard the integrity of the cellular environment by orchestrating the removal of compromised cells. A recurrent theme is the potential involvement of SOX3 as either a promoter or inhibitor of apoptosis. This role appears to be contingent upon the specific type of cancer cells under consideration.

For example, in breast cancer cell lines MCF-7 and T-47D, both originating from invasive ductal carcinoma Luminal A molecular subtype and characterized by differentiated epithelial cells, a study revealed that miR-483, targeting SOX3, induced apoptosis leading to a reduction in cell proliferation (Figure 1A). Notably, miR-483 is down-regulated in both breast cancer tissues and Luminal A breast cancer cell lines [36] (Figure 1B). These findings strongly suggest that SOX3 may play a regulatory role as a blocker of apoptosis, specifically within the context of Luminal A breast cancer.



**Figure 1. (A)** The miR-483 targets SOX3, induces apoptosis, and reduces cell proliferation. **(B)** The down-regulation of miR-483 in breast cancer tissues and Luminal A cancer cell lines decreases apoptosis and induces cell proliferation [36]. **(A)** The miR-483 transfection into glioma cell lines directly targets SOX3 and downregulates SOX3, enhancing apoptosis in glioma cells [38].

In contrast, a recent study in breast cancer, utilizing the MDA-MB-231 cell line from invasive ductal carcinoma of the Triple Negative (TN) molecular subtype and characterized by undifferentiated epithelial cells with mesenchymal morphology, revealed no SOX3 expression. The transfection and expression of SOX3 into these cell lines resulted in the expression of pro-apoptotic markers leading to apoptosis, as detected by Annexin V/PI flow cytometry [39]. These disparate results highlight the complex and context-dependent role of SOX3 in the regulation of apoptosis across different molecular subtypes of breast cancer.

In a study conducted by Guo et al.[33], it was observed that the expression of SOX3 was significantly reduced in osteosarcoma (OS) cell lines. This decrease in SOX3 levels contributed to a higher concentration of OS cells in the G1 phase of interphase and triggered cell apoptosis. This phenomenon was further supported by the observation of lowered Bcl-2 levels, an anti-apoptotic marker, with an increase in the expression of the pro-apoptotic gene Bax [33]. Similarly, research by Shujing et al. showed that miR-483 directly targets and downregulates SOX3, which enhances apoptosis in glioma cells [38]. These works illustrate a shared pathway in different types of cancer cells where SOX3 suppression leads to increased apoptosis [38] (Figure 1A).

Yan et al.[35], investigated SOX3 expression in ovarian carcinoma tissues and SOX3 basal expression in six different ovarian cancer cell lines. SOX3 expression and localization in human ovarian cancer were detected mainly in cell nuclei, while normal ovarian tissue samples showed no SOX3 expression. To assess the effect of SOX3 overexpression and silencing in SK-OV-3 (human ovarian cancer cell line with epithelial-like morphology) and SK-OV-3-ip1 (more metastatic) apoptosis compared with their control cells were analyzed with Annexin V/Pi flow cytometry. Results revealed a higher percentage of apoptotic cells in SK-OV-3 and SK-OV-3-ip cell lines silenced for SOX3 [35], and a downregulation of apoptosis when SOX3 is overexpressed in these cell lines.

Comprehending the intricacies of apoptosis is essential for establishing precise anti-cancer strategies and discovering innovative therapeutic approaches focused on reinstating apoptotic regulation in cancerous cells. New therapeutic approaches aimed at modulating both the intrinsic and extrinsic pathways of apoptosis, either individually or in combination, hold promise for treating cancer. Oligonucleotides and small molecules designed to mimic the interaction between BH3 proapoptotic members and BCL-2 anti-apoptotic members within the BCL-2 family offer a means of

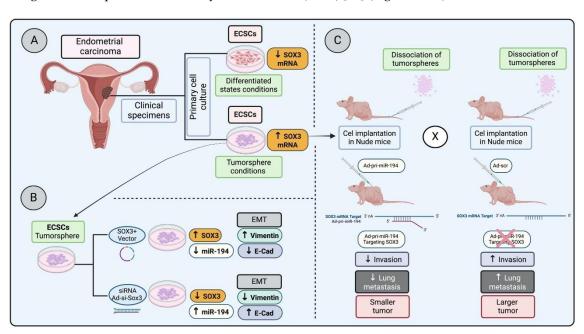
influencing mitochondrial membrane permeability and the release of cytochrome c. Additionally, the targeting of cell death receptors (DR) involved in activating the extrinsic apoptosis pathway through external signaling presents a potential mechanistic approach. Agonists capable of binding to DR and initiating cell death signaling show potential in this context [40].

# 2.2. SOX3 and Epithelial-Mesenchymal Transition (EMT)

Epithelial–mesenchymal transition (EMT) is a pivotal cellular process with far-reaching implications in oncology. EMT is a reversible program that transforms epithelial cells into mesenchymal cells, involving the loss of adherents junctions and the downregulation of cytokeratins and E-cadherin (epithelial-specific markers), and an increase of mesenchymal markers, such as fibronectin, N-cadherin, and vimentin [41]. In the context of cancer, EMT plays a crucial role in malignant progression by inducing traits such as tumor-initiating properties, motility, dissemination ability, and resistance to chemotherapy. Orchestrated by EMT-inducing transcription factors (EMT-TFs), such as SNAIL, SLUG, TWIST, and ZEB1/ZEB2, this epigenetic process operates independently of DNA sequence [42]. EMT in carcinoma cells depends on signals from the tumor-associated reactive stroma induced by EMT-TFs, shaping the tumor microenvironment. The detection of EMT-associated protein markers serves as a prognosis indicator of high-grade malignancy in various cancers, including prostate, lung, liver, pancreatic, and breast cancers [43–46].

Qiu et al.[47] identified *SOX3* as a metastasis-associated gene in OS, highlighting its mechanistic connection with the TFs SNAIL1 and MET. *SOX3* was overexpressed in 42 cases of human OS tissues in comparison with non-tumor samples. In addition, MG63 transfected with SOX3 exhibited elevated expression of MET markers, such as N-cadherin and Vimentin, and lower expression of epithelial markers, such as E-cadherin and Keratin 1, while the SOX3 silencing in U2OS cells increases epithelial markers and decreases mesenchymal markers, suggesting SOX3 involvement in EMT in OS cells [47].

In a study examining endometrial carcinoma stem cells (ECSCs) under both tumorsphere conditions and differentiated states—achieved by removing basic fibroblast growth factor (bFGF)—a significant decrease in SOX3 mRNA expression was observed in differentiated conditions compared to their undifferentiated tumorsphere counterparts (Figure 2A). This research took a further step by injecting dissociated undifferentiated cells from tumorspheres into nude mice, followed by the administration of Ad-pri-miR-194 targeting SOX3 mRNA (Figure 2C). These results highlight that silencing SOX3 led to reduced invasion and lung metastasis, pointing to SOX3 as a potential marker for ECSCs and suggesting its involvement in invasion, metastasis, and possibly in the regulation of epithelial-mesenchymal transition (EMT) [48] (Figure 2B,C).



However, Silva et al.[49] demonstrates *in vitro* induction of SOX3 expression results in a decreased expression of the mesenchymal marker N-cadherin (NCAD) and TFs SNAIL, ZEB1, and ZEB2, which play crucial roles in EMT. This aligns with earlier studies that identified elevated levels of SOX3 as key to inhibiting EMT, as seen by the reduced expression of SNAIL in the MCF-7 breast cancer cell line [50]. Moreover, the study observed that MDA-MB-231 cells overexpressing transiently transfected SOX3 exhibited changes in EMT-related TFs and upregulation in E-cadherin (ECAD) gene expression, further substantiating the role of SOX3 in blocking EMT [49].

Understanding and characterizing EMT programs are important in clinical oncology, as they contribute to the elevated resistance of mesenchymal carcinoma cells to various treatment regimens, including chemotherapy and immunotherapy.

# 2.3. SOX3 and Cell Invasion and Migration

Epithelial-mesenchymal transition (EMT) is followed by invasion and migration of cancer cells, ultimately leading to metastasis. Notably, an increase in SOX3 expression has been associated with gastric cancer characterized by lymph node metastasis, primary tumor invasion, and high TNM tumor graduation system [51]. In this study, comprehensive investigations, which included gastric cancer cell lines, a zebrafish *in vivo* model, as well as clinical samples of patients with gastric cancer, were employed. *In vitro* experiments demonstrated that silencing *SOX3* reduced the expression of Matrix metalloproteinase-9 (MMP-9). ChIP-PCR confirmed direct transcriptional regulation of the *MMP-9* promoter by SOX3, establishing a pivotal role in the SOX3 transcriptional regulation of MMP-9, a protein crucial for cell invasion and migration processes [51].

In osteosarcoma (OS), Guo et al.[33] conducted a SOX3 knockdown in an OS cell line using a Transwell assay, suppressing migration and invasion of OS cells with reduced SOX3 levels. However, this effect was not observed in control cells with basal levels of SOX3 expression [33]. In a parallel study involving ovarian cancer cells, clinical samples, and *in vitro* approaches, Yan et al.[35], reported that silencing SOX3 in SK-OV-3 cells decreases its ability to migrate and metastasize [35].

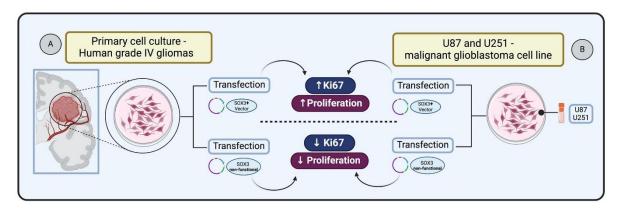
Malignant glioblastoma (GBM) is an aggressive cancer characterized by its invasive behavior. In an effort to investigate the SOX3 influence on GBM behavior, Vicentic et al. [34] induced SOX3 overexpression in GBM cell lines. Following the overexpression in U87 and U251 GBM cells, they utilized both Transwell migration and Matrigel assays to assess the cells' behaviors. The results demonstrated that cells increased SOX3 expression lead to enhanced migration and invasion capabilities *in vitro* [34]. Building on this, Pan et al. [52] discovered that reducing SOX3 expression elevates the migration of the U251 glioblastoma cells, as supported by a wound-healing assay, while not influencing the cellular invasive capabilities at 48 hours [52]. Furthermore, research by Shujing et al. [38], which targeted SOX3 with its repressor miR-483, found that downregulating SOX3 suppresses both cell migration and invasion, providing insights into the complex role of SOX3 in GBM behavior.

In esophageal squamous carcinoma samples from 118 patients, both gene and protein expression were evaluated using RT-qPCR coupled with immunohistochemistry, showing no significant correlation in clinical evaluations of primary tumor invasion with SOX3 [53]. Similarly, employing comparable approaches, Feng et al.[54] showed no correlation between SOX3 expression and clinicopathological factors such as tumor emboli and microvascular invasion in hepatocellular carcinoma [54]. Collectively, these observations suggest that SOX3 plays a role in invasion and migration depending on tumor subtype and spatial distribution.

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Cancer is associated with deregulated cell cycle controls. Defects in checkpoints and cyclin-dependent kinase (CDK) activity can drive unrestrained proliferation, increase genomic instability, and contribute to cancer progression and treatment resistance [55]. SOX3 appears to have a role in cell cycle progression. Knockdown experiments with SOX3 resulted in a G1 arrest in OS cells, accompanied by a decrease in the proportion of osteosarcoma MG63 and U2OS cells in the S and G2/M phases [33]. To understand the mechanism of cell cycle alteration in SOX3 knockdown cells, western blot for Cdc25A, cyclin D1, and PCNA protein quantification indicates a decrease in the expression of these three proteins relative to the control cells [33].

SOX3 is known to be involved in central nervous system (CNS) development during embryogenesis. Experiments demonstrating SOX3 gain of function, through cDNAs encoding HMG box of chick Sox3 in expression constructs, followed by electroporated into the neural tube of Hamburger-Hamilton (HH) stage 10 chick embryos, have revealed its capacity to sustain cells as self-renewing progenitors [56]. Holmberg et al.[57] investigated whether SOX3 exhibits this regulatory capacity in glioma cells. They transfected primary cultures derived from human grade IV gliomas with vectors expressing either full-length SOX3 or a dominant negative version of SOX3 (SOX3EnR-Myc). Glioma cells expressing SOX3 showed the presence of cell cycle marker Ki67 within 24 hours following transfection. In contrast, glioma cells transfected with the non-functional SOX3 were prompted to exit the cell cycle, resulting in a reduction of cells positive for Ki67 after 24 hours. These results indicate that SOX3 can maintain glioma cells in an undifferentiated and proliferating state, while the active repression of SOX3 target genes causes glioma cells to exit the cell cycle [57] (Figure 3A).



**Figure 3. (A)** Primary glioma cells transfected and overexpressing SOX3 increases Ki67 expression and cell proliferation status [57]. **(B)** Glioblastoma cells transfected and overexpressing SOX3 increase Ki67 and enhance cellular proliferation [34].

In the context of malignant glioblastoma (GBM), Vicentic et al.[34] identified the upregulation of SOX3 as a key factor in enhancing proliferation. The study involved overexpressing SOX3 in U87 and U251 cell lines transiently transfected with pcDNA3.1/SOX3 construct, leading to a significant increase in proliferation in both cell lines. This effect was confirmed by anti-phosphohistone H3 (pH3) immunostaining, indicating an elevated number of dividing cells and increased expression of the Ki67 marker, which is indicative of enhanced cellular proliferation [34] (Figure 3B).

After conducting functional *in vitro* experiments involving both the overexpression and silencing of SOX3 in ovarian cancer cell lines (SK-OV-3), Yan et al.[35] assessed the influence of SOX3 on cell proliferation using a CCK8 assay, enabling the quantification of live cells, along with assays to assess colony formation. In ovarian cancer cells, upregulation of SOX3 was found to elevate cell proliferation rate, while inhibition reduced cell proliferation. Moreover, the overexpression of SOX3 induced an increased formation of colonies [35].

In contrast, when applying a similar overexpression and silencing approach to SOX3 while utilizing the CCK-8 assay to count cells, Shen et al.[51] observed that SOX3 exerted little effect on cell

proliferation in gastric cancer cell lines. This discrepancy, again, highlights the context-dependent nature of SOX3 influence on cell proliferation, suggesting that its impact varies across different cancer types.

SOX3, part of the SOXB1 family, is emerging as a critical regulator in the cell cycle and proliferation, interacting with various TFs and regulatory proteins. These interactions likely play roles in crucial phases such as the G1 to S phase transition or cell cycle checkpoint regulation, thus affecting the cell cycle's timing and progression [59]. Interestingly, SOX3 role is multifaceted; Turchi et al. <sup>60</sup> highlighted its anti-proliferative influence in Plasmacytoid Dendritic Cells (PDC) from primary glioblastoma tumors, presenting a contrast to its previously noted contribution to cancer cell proliferation and tumor progression in different cancers, including gliomas [34,51].

This duality in SOX3 function emphasizes the complexity of gene regulation within cancer biology, where SOX3 interactions are pivotal and exceedingly context-dependent. Despite the established significance of interactions with other SOX family members, such as SOX2 and SOX4, with cyclins/CDKs and key signaling pathways like Notch and Wnt, specific documentation regarding SOX3 involvement in these pathways remains scarce [59]. This lack of detailed insight underscores a significant gap in our understanding of the molecular mechanisms through which SOX3 influences the cell cycle and cancer progression, highlighting the need for further research to unravel the intricacies of SOX3 regulatory roles and interactions within the cell cycle and beyond.

# 3. SOX3 Investigation and Clinical Correlation in Different Types of Cancer:

SOX3 investigation in different types of cancer: osteosarcoma (OS), breast cancer (BC), gastric cancer (GC), endometrial cancer (EC), esophageal cancer, hepatocellular carcinoma (HCC), lung, ovarian, and Acute myeloid leukemia (AML), and its clinical relation with SOX3 expression is summarized and described in Table 1.

#### 3.1. SOX3 in Osteosarcoma

Osteosarcoma (OS) is a rare primary malignant sarcoma in bone, exhibiting osteoid production alongside malignant mesenchymal cells. It ranks as the third most common cancer in adolescence, with an annual incidence of 5.6 cases per million among children under 15. Osteosarcoma typically arises sporadically, with chromosomal abnormalities identified in about 70% of tumor specimens, often involving mutations in tumor-suppressor genes or DNA helicases [60].

In a study involving osteosarcoma patients (n=70), higher gene (RT-qPCR) and protein expression of SOX3 were observed when compared to benign bone lesions [47]. Additionally, *in vitro* studies using osteosarcoma cell lines indicated that silencing SOX3 expression in osteosarcoma leads to reduced aggressiveness, including proliferation and invasion. Further, there is a positive correlation between SOX3 and genes involved in the epithelial-mesenchymal transition (EMT), suggesting a potential regulatory role. This identifies SOX3 as a potential therapeutic target for metastasis in osteosarcoma [33,47] (Table 1).

#### 3.2. SOX3 in Ovarian Cancer

Ovarian cancers (OC) rank as the second leading cause of gynecological cancer-related deaths. Despite advances in treatment, survival rates for stage III and IV Epithelial Ovarian cancer (EOC) remain at 40% and 20%, respectively. OC treatments involve surgery and platinum-based chemotherapy. However, recurrence is frequent, and current screening methods, including clinical examination and assessment of tumor markers, offer limited benefit in overall survival. Risk factors for OC include age, nulliparity, endometriosis, obesity, and smoking. In addition, hereditary factors, such as BRCA mutations, play a significant role. While molecular subtyping through immunohistochemistry aids diagnosis and prognosis, the quest for new diagnostic and prognostic markers persists [61].

Positive nuclear accumulation of SOX3 in immunohistochemistry staining was found in human ovarian cancer tissue samples, contrasting with the negative staining observed in control ovarian

tissue [35]. This variability in SOX3 expression among human ovarian cancer cell lines highlights its differential behavior depending on the cell type, with metastatic cell lines showing increased SOX3 expression, potentially tying SOX3 to malignant transformations within ovarian tumors. Intriguingly, studies examining the response to chemotherapy drugs like cisplatin found that SOX3 expression was lower in tissues that were more resistant to the drug, suggesting a complex involvement of SOX3 in the progression of ovarian cancer [35] (Table 1).

Expanding upon these insights, Matsumoto et al. [62] explored the impact of anaplastic lymphoma kinase (ALK) overexpression in ovarian cancer [62]. Their research indicates that ALK's overexpression significantly impacts the biological behavior of ovarian high-grade serous carcinoma (HGSC) and is transcriptionally regulated by the SOXB1 subgroup, which includes SOX3. In HGSC cell lines overexpressing SOX3, ALK expression increased, contributing to the aggressive phenotypic characteristics of HGSC [62]. This body of research not only underscores the pivotal role of SOX3 in ovarian cancer but also connects it with other key molecular players like ALK, offering a better understanding of the molecular underpinnings of ovarian cancer and highlighting potential targets for therapeutic intervention.

#### 3.3. SOX3 in Breast Cancer

Breast cancer (BC) stands out as the most prevalent and challenging malignancy affecting women, representing one of the most widespread cancers globally. Its complexity is evident in diverse tumor types characterized by distinct morphology, behavior, and clinical implications. The great heterogeneity in BC poses challenges in understanding and treating the disease. Categorically, BC is divided into three major molecular subtypes based on the positive or negative expression of estrogen or progesterone receptors and human epidermal growth factor 2 (ERBB2 (or Human Epidermal Growth Factor Receptor 2 (HER2)): hormone receptor-positive/ERBB2 negative (70% of patients), ERBB2 positive (15%-20%), and triple-negative (tumors lacking all three standard molecular markers; 15%). Breast cancer treatment is intricately linked to the cancer molecular subtype and stage. Early diagnosis significantly enhances survival rates, with a noteworthy 90% chance of survival within 5 years [63,64].

Various scientific investigations have reported changes in SOX genes and/or protein expression in human breast cancer, suggesting that SOX genes contribute significantly to key aspects of breast cancer genesis and progression. In contrast to studies with SOX2, showing it to be associated with aggressive BC and an indicator of poor prognosis [29,65,66], there are limited reports on the role of SOX3 in BC (Table 1).

Mehta et al.[67] extensively analyzed the transcript profile of SOX family gene expression in breast cancer subtypes [67]. Samples were organized by PAM50 molecular subtype, and patterns of SOX gene expression were determined for 1,052 human breast tumors and 94 adjacent standard samples from the TCGA dataset. The analysis revealed altered expression of several SOX genes relative to adjacent normal breast tissue and within the context of the PAM50 molecular subtypes. Notably, SOX3 was excluded from the analysis due to missing or insufficient data (expression values present in >80% of samples) [67] (Table 1).

In a study with invasive ductal carcinoma (IDC) cell lines MCF-7 and MDA-MB-231, Silva et al.[39], reported transcript expression of SOX3 and cytoplasmic localization of SOX3 in MCF-7 cells characterized as an epithelial-like cell and Luminal A molecular subtype. In contrast, no SOX3 mRNA or protein was detected in MDA-MB-231 cells, classified as mesenchymal-like and a triple-negative molecular subtype [39]. Following transfection of MDA-MB-231 cells with a SOX3 expression vector, cells upregulated pro-apoptotic genes and increased the apoptotic rate, supported by Annexin V/PI flow cytometry, indicating SOX3 involvement in apoptosis regulation [39]. The study further investigated SOX3 immunohistochemistry localization and quantification in 27 IDC patient samples, along with its correlation with Pro-caspase-3 immunoreactivity. Interestingly, positive cases for Pro-caspase-3 were negative for SOX3, and the weak staining pattern, with significant SOX3 cytoplasmic localization and a low score (average of 25%), was associated with the

cell's aggressive behavior, indicating downregulation of SOX3 with resistance to apoptosis phenotype [39] (Table 1).

In summary, the action of SOX3 in BC, whether as an oncogenic or tumor suppressor, appears to be linked and dependent on the breast cancer histological, molecular, and grade subtype.

# 3.4. SOX3 in Esophageal Cancer

Esophageal cancer, originating in the esophageal epithelium, poses a significant challenge due to low cure rates, especially with late diagnosis [68]. This malignancy is often linked to specific genetic alterations, including mutations in the TP53 tumor suppressor gene, alterations in the CDKN2A gene, and amplifications of the ERBB2 (HER2) oncogene, among others. In esophageal squamous cell carcinoma (ESCC), notable elevations in SOX3 expression were observed compared to non-neoplastic samples [68] (Table 1).

Clinicopathologic correlation studies indicated that increased SOX3 expression in ESCC is significantly associated with regional lymph node metastasis (RLNM) and advanced TNM staging. This suggests that SOX3 holds promise as a valuable biomarker for prognostic prediction in esophageal cancer and a potential therapeutic target for ESCC [53].

#### 3.5. SOX3 in Gastric Cancer (GC)

Gastric Cancer (GC) is a complex and unresolved clinical challenge, marked by heterogeneity and particularly high mortality rates in advanced and metastatic stages. This condition remains a significant health concern, with poor overall survival statistics, especially prevalent in Asian and South American countries. Given the variable outcomes associated with the different disease subtypes, the urgency for improved treatment and early detection strategies is evident. Ongoing research explores various emerging therapies and targets in the quest for more effective management strategies [69].

In gastric cancer (GC), there is an observed elevation in serum SOX3 expression compared to healthy individuals. This increased SOX3 expression in GC is intricately linked to differentiation, lymph node metastasis, and tumor invasion. The correlation with metastasis is proposed to be influenced by SOX3 positive modulation of matrix metalloproteinase-9 (MMP-9), a key player in cancer cell migration [15,51] (Table 1). Consequently, SOX3 emerges as a promising prognostic factor in GC patients, showing potential oncogenic properties and positioning itself as a candidate for targeted intervention aimed at suppressing cancer progression.

#### 3.6. SOX3 in Glioma and Glioblastoma (GBM)

Glioblastoma (GBM) is recognized as the most aggressive brain tumor due to its rapid and infiltrating growth progression. GBM diagnosis depends on histopathological examination. The molecular subtype of GBM is crucial for both diagnostic accuracy and treatment [70].

In a study led by Vicentic et al.[34], clinical tissue samples and *in vitro* models were employed to investigate the role of SOX3 in GBM. Immunohistochemical analysis of clinical samples confirmed the presence of SOX3 in the nucleus of all analyzed tumor samples, revealing elevated SOX3 expression in GBM samples compared to non-tumoral brain tissues. Similarly, Yuan et al.[71] and Shujing et al.[38] examined glioma tumor tissues and adjacent normal brain tissues, and SOX3 gene expression was upregulated in glioma tissue clinical specimens compared to that in adjacent normal tissues [38,71] (Table 1).

In a series of in vitro studies, the role of SOX3 in glioma and glioblastoma was investigated, revealing its varied impact on cancer cell behavior [34,38]. Shujing et al.[38] noted that glioma cell lines (LN18 and LN229) exhibited an elevation in SOX3 transcripts when compared to normal brain cell lines (HEB), indicating a potential link between SOX3 expression and glioma pathogenesis. Building on this observation, Vicentic et al.[34] explored SOX3 expression across a broader spectrum of glioblastoma cell lines (U87, U373, U251, A172, and T98). They found that SOX3 expression varied significantly among these lines, and importantly, cells transfected to overexpress SOX3 showed

enhanced proliferation, viability, migration, and invasion capabilities [34]. Further extending the investigation into SOX3's role, Jason et al. [72] used RNA sequencing (RNA-seq) to identify several genes, including SOX3, that were differentially expressed in glioblastoma, correlating with increased tumor invasiveness, malignancy, and a poor prognosis for patients [72] (Table 1).

In a comprehensive exploration of SOX3 in glioblastoma, Pan et al.[52] through ONCOMINE and CCLE bioinformatic databases found SOX3 overexpression in glioblastoma tissues compared to normal tissues [52]. This was complemented by a prognostic analysis using LinkedOmics and GEPIA databases, which presented a positive correlation between higher SOX3 levels and improved overall survival rates in GBM patients, suggesting SOX3 potential as a prognostic biomarker [52]. However, this finding was met with controversy, as Shujing et al. [38] identified a connection between SOX3 upregulation and poorer patient outcomes in glioma, highlighting the complex role of SOX3 in glioblastoma and glioma pathology (Table 1).

Further investigations into the functional role of SOX3 through *in vitro* studies using the U251 glioblastoma cell line demonstrated that downregulating SOX3 positively impacted the woundhealing rate, indicating its influence on cell migration [52]. Vicentic et al. [34] furthered this line of inquiry, showing that SOX3 overexpression not only enhanced migration but also increased viability, proliferation, and invasion of glioblastoma cells, while reducing autophagy [34]. This effect was particularly pronounced in glioblastoma stem cells and oncospheres, emphasizing SOX3 significant role in tumor aggression and stem cell properties [34].

Expanding our understanding of the regulatory mechanisms of SOX3 was a series of *in silico* experiments and functional assays, revealing that miR-483 and miR-483-3p target SOX3, impacting its expression and thereby affecting tumor cell behavior [38]. This interaction was supported by the findings of Yuan et al. [71], who confirmed the inhibitory effect of miR-483-3p on SOX3 through Dual-Luciferase reporter assays, linking SOX3 upregulation to increased cell proliferation and anti-apoptotic activity [71] (Table 1). Additionally, the study by Turchi et al. [73] shed light on the RNA-binding protein CELF2's role as an epigenetic regulator that indirectly represses SOX3, promoting a proliferative tumor cell phenotype and correlating with more aggressive tumor behavior [73]. This relationship between CELF2 and SOX3 underscores the intricate regulatory networks influencing glioblastoma cell dynamics [73].

Additionally, the expression of SOX3 was associated with the presence of the cell cycle marker Ki67, reinforcing its pivotal role in maintaining glioma cells in a proliferative state and promoting malignant behavior in GBM cells [34,57] (Table 1).

Lastly, research by Scuderi et al. [74] introduced a potential therapeutic angle by demonstrating that the inhibitor BX795 significantly reduced SOX3 expression in various glioblastoma cell lines, pointing to the targeting of SOX3 pathways as a promising approach to improve disease outcomes [74] (Table 1). Together, these findings paint a complex but enlightening picture of SOX3s multifaceted role in glioblastoma and glioma, offering valuable insights into its potential as a biomarker and therapeutic target in the fight against these diseases.

# 3.7. SOX3 in Hepatocellular Carcinoma (HCC)

Hepatocellular carcinoma (HCC) stands out as the predominant primary liver malignancy. Risk factors for HCC include chronic liver disease and cirrhosis, with viral hepatitis and excessive alcohol intake ranking as the foremost contributors [75].

A study by Feng et al.[54] delved into SOX3 mRNA expression and protein immunolocalization in HCC tissues compared to non-tumor counterparts. The findings revealed a significant upregulation of SOX3 mRNA and protein expression in HCC tissues compared to adjacent non-tumor regions. Elevated SOX3 expression was associated with advanced tumor progression and worse prognosis in HCC patients. The correlation between SOX3 expression and clinicopathological features further indicated that high SOX3 expression was linked to lower tumor capsule formation, poorer tumor differentiation grades, and worse TNM classification [54] (Table 1).

The immunolocalization of SOX3 by IHC demonstrated that SOX3 was predominantly localized in the nucleus of tumor cells [54], aligning with its expected role as a TF. These findings underscore the significance of SOX3 as a potential prognostic indicator in HCC and inform treatment strategies.

#### 3.8. SOX3 in Endometrial Carcinoma (EC)

Endometrial carcinoma (EC) is the predominant cancer within the uterine corpus, constituting over 83% of cases. The SOX gene family, particularly SOX2, plays a significant role in carcinogenesis, maintaining cancer stem cell (CSC) pluripotency and regulating cell differentiation, proliferation, and survival. Expression of SOX3 has been identified in EC tissues, correlating with multipotency observed in endometrial tumorspheres cultivated in stem cell medium. These tumorspheres serve as a cancer stem cell model indicative of SOX3 as an ECSC marker. A positive correlation between SOX3 and miR-194 expression with undifferentiated ECSCs was noted, and SOX3 overexpression sustained pluripotency in EC tumorspheres. Elevated SOX3 expression appeared to enhance the epithelial-mesenchymal transition (EMT) process in ECSCs, suggesting its potential to impact clinical outcomes in EC patients [48] (Table 1).

# 3.9. Acute Myeloid Leucemia (AML)

Acute myeloid leukemia (AML) is a complex and heterogeneous disease characterized by rapid cellular proliferation, an aggressive clinical course, variable prognosis, and generally high mortality. A study conducted by Tosica et al. 2018, examined SOX3 gene expression in clinical samples from AML patients and its correlation with clinicopathological aspects. The analysis revealed higher SOX3 expression in 22% of the analyzed AML patients, with a corresponding complete remission rate of 55%. Furthermore, patients with high SOX3 expression exhibited a lower Disease-free survival (DFS) than those with low expression, although the difference lacked statistical significance. Overall survival (OS) mirrored the DFS findings, indicating that patients displaying high SOX3 expression had OS of 3 months, not significantly shorter than the 7 months observed in patients with low SOX3 expression [76] (Table 1).

**Table 1.** – SOX3 in different cancer types and its clinical correlation.

		Consistence of Community	Mathadalaav/	Main Results / Clinical
		Specimens/Sample	Methodology/	
Author	Tumor type	s	Technique	Correlation
				SOX3 gene expression was
				not different from healthy
				individuals. After the
				implementation of the "cut-
				off" value (3.60), it was
				detected 11 (22%) patients
				with high SOX3 expression.
				The complete remission rate
				of patients with high
		Clinical specimens:		expression of SOX3 was 55%.
		50 AML patients		In the survival analyses,
		with bone marrow		patients with increased
	Acute myeloid	and 12 healthy		expression of SOX3 showed
	leucemia	controls (bone		lower Disease-free survival
TOSIC et al., 2018 [76]	(AML)	marrow donors).	RT-qPCR	(DFS) compared to patients

	-	ı		
				with low expression of SOX3
				(4 vs. 14 months). Patients
				with SOX3 high expression
				had an overall survival (OS)
				of only 3 months, but it was
				not significantly shorter
				compared to the 7 months
				found in the patients with
				low SOX3 expression (Log-
				Rank = 3.434; p = 0.064).
				The miR-483 inhibitor
				upregulated the protein level
		Clinical specimens:		of SOX3. SOX3 expression
		62 patients.		was negatively correlated
		Cell lines: MCF-7,	Cell transfection; qRT-	with miR-483 expression in
		SKBR3, LCC2,	PCR; WB; Luciferase;	breast cancer tissues. The
		MDA-MB-453, T-	cell viability and	miR-483 could suppress
		47D, LCC9, and	proliferation (MTT);	breast cancer cell
		normal human	colony formation and	proliferation and promote
		breast cell line	apoptosis detection	cell apoptosis via targeting
CUI et al., 2019 [36]	Breast	MCF-10A	assay.	SOX3.
				The apoptotic rate was higher
				in cells transfected with
				pEF1-SOX3+ than in controls.
				MDA-MB-231 transfected
				with pEF1-SOX3+ showed
			Cell transfection with	upregulation of pro-
			SOX3 expression vector;	apoptotic CASP3, CASP8,
			Immunofluorescence;	CASP9, and BAX mRNA,
			cell viability and	contrasting with
			proliferation (MTT);	downregulation of BCL2
		Cell line: MDA-MB-	flow cytometry	anti-apoptotic mRNA,
SILVA et al., 2022 [39]	Breast	231	(apoptosis); RT-qPCR.	compared to controls.
				The nuclear expression of the
				SOX3 protein was detected in
				14% of the cases of ductal
				carcinoma, and the
		Clinical specimens:		expression of pro-Caspase-3
		İ		::: : F00/ FI HIG
		27 patients with		was positive in 50%. The IHC
		27 patients with breast invasive		negative nuclear expression

				13
				can be related to cells resistant to apoptosis.
SILVA et al., 2024 [49]	Breast	Cell lines: MDA-MB-231	Cell transfection with SOX3 expression vector; Viability test (MTT); RT-qPCR.	A downregulation in NCAD and an upregulation of ECAD expression, followed by SOX3 protein expression in the triple-negative breast cancer MDA-MB-231 cell line.
GONG et al., 2017 [48]	Endometrial carcinoma	Clinical specimens:  19 Endometrial carcinoma patients.  Samples (stage IB, n = 11; stage IC, n = 5; stage IIa, n = 3; age = 37–72 years).  Primary cell culture with the 19 EC, forming tumorspheres of in vitro experiments.  Implantation of tumorsphere cells into mice nude for in vivo experiments.	Constructs for overexpression and silencing SOX3; cell transfection; Flow cytometry, RT-qPCR, WB,	SOX3 contributes to endometrial cancer stem cell invasion and suggests that repression of SOX3 by microRNA-194 may have therapeutic potential to suppress endometrial carcinoma metastasis.
GONG et al., 2017 [48]	Esophageal	The vivo experiments.  Clinical specimens:	RT-qPCR; WB; Tissue microarrays;	The expression of SOX3 in esophageal squamous carcinoma (ESCC) was significantly higher than in non-neoplastic samples. SOX3 expression in ESCC significantly correlated with regional lymph node metastasis (RLNM) and advanced TNM. SOX3 may be a valuable biomarker for
LI et al., 2013 [53]	squamous	30 patients	Immunohistochemistry.	predicting prognosis and a

	_			
				potential therapeutic target
				for ESCC.
				SOX3 protein is involved in
				esophageal squamous
				carcinoma cell (ESCC)
				metastasis. SOX3 disruption
		Cell lines: ECA109,	Proliferation and	impaired ESCC cell
		SKGT-5, SKTG-4,	cytotoxicity assays	migration and invasion.
		TE-1, TE-3, TE-8;	(LDH); Oncomine;	Metastasis was significantly
		AND SV40-	migration and invasion;	inhibited when the SOX3
	Esophageal	immortalized non-	WB; MMPs activity; RT-	gene was disrupted by
CAI et al., 2016 [77]	squamous	tumorigenic	qPCR; xenograft model.	insertional mutagenesis.
C/11 Ct al., 2010[//]	squamous	tumongene	qr cre, xeriogram model.	SOX3 promotes tumor cell
				proliferation, migration, and
				invasion in vitro. SOX3
				promotes lymph node
				metastasis of the tumor <i>in</i>
				vivo. SOX3 could increase the
		Cell lines: TE-1, TE-		VEGF-C/D expression in
		10, TE-11, EC109,		ESCC cells both <i>in vivo</i> and <i>in</i>
		EC9706.		vitro. The high expression of
		Animal model:	WB; RT-qPCR; Invasion;	SOX3 upregulated the
		Tumorigenesis and	Scratch; MTT; Tube	expression of VEGF-C and
		axillary lymph		VEGF-D in ESCC and
	Esophageal	node metastasis in		
ZHENG et al., 2017 [78]	squamous	nude mice.	Immunohistochemistry.	metastasis.
ZHENG et al., 2017 [70]	squamous	nude niice.	minunonstochemistry.	
				Serum proteome profiling
				reveals differential
				expression of SOX3 protein,
				between pre- and post-
				operation for locally
				advanced gastric cancer.
				SOX3 is overexpressed in
		Clinian and an arise and	Dustain	gastric cancer tissues and is
		Clinical specimens:		associated with poor
		60 patients - 5 cases	TMT/iTRAQ labeling;	outcomes for gastric cancer.
		of early gastric		This study highlights the
		carcinoma and 55	liquid chromatography;	potentiality of the paired pre-
		locally advanced	WB; ELISA;	and post-operation serum
CHEN of al 2020 [51]	Costria	gastric carcinoma	Immunohistochemistry,	proteome signature for
SHEN et al., 2020 [51]	Gastric	cases.	Immunofluorescence.	detecting putative

			biomarkers for gastric
			carcinoma and reveals that
			SOX3 may serve as a
			candidate molecular marker
			for the prognosis and
			outcomes of gastric cancer
			patients.
		SOX3 mRNA silencing;	patients.
		Invasion assay;	
	Cell lines: AGS and	Xenograft model in	SOX3 promotes gastric
	MKN45 human		cancer cell invasion and
		, ,	migration through MMP9.
	gastric	Chromatin	
,	adenocarcinoma	immunoprecipitation.	
	Clinical		
	specimens:Glioblas		
	toma (GMB)		Identification of several
	obtained from		differentially expressed
	patients		genes, including SOX3,
	undergoing	DNA and RNA	associated with tumor
s	surgery was used to	Sequencing; Invasion;	invasiveness, malignancy,
C	obtain primary	Proliferation;	and unfavorable prognosis in
3	glioblastoma stem	Immunoblot Assay;	glioblastoma patients.
C	cells (GSCs).	Flow Cytometry;	
SON et al., 2019 [72] Glioblastoma		Immunohistochemistry.	
	Clinical specimens:		
1	13 samples for		SOX3 expression is higher in
i	immunohistochemi		glioblastoma samples than in
s	stry, 27 samples for		non-tumoral brain tissues.
I	RT-qPCR, Control		SOX3 protein expression in
r	non-tumoral brain		cell nuclei was observed in all
I	RNAs, and 23		analyzed tumor samples.
ICENTIC et al., 2019	samples for	Immunohistochemistry	anaryzeu tumor sampies.
4] Glioblastoma A	Ambion.	and RT-qPCR	
			Exogenous overexpression of
			SOX3 enhances proliferation,
	Cell lines: U87,	Transfection and	viability, migration, and
τ	U373, U251, A172	luciferase assay; WB;	invasion of glioblastoma
a	03/3, 0231, 111/2		·
	and T98; GNS166	MTT;	cells. The upregulation of
a		•	cells. The upregulation of SOX3 was accompanied by
	and T98; GNS166	,	

				and autophagy suppression
				in glioblastoma cells. SOX3
				expression was elevated in
				patient-derived glioblastoma
				stem cells as well as
				oncospheres derived from
				glioblastoma cell lines
				compared to their
				differentiated counterparts.
				Oncomine indicated the
				CCLE database showing
				SOX3 overexpressed in
				Glioblastoma with a fold
				change (FC) of 1.184
				compared to normal tissue.
				LinkedOmics and GEPIA
				databases showed that
				increased SOX3 improved
				overall survival (Logrank p =
				0.0432). The survival rate of
				high SOX3 patients is much
				higher than low SOX3
				patients (HR = 0.825), and
				SOX3 may serve as a
				prognostic biomarker set for
				GBM patients.
				Downregulation of SOX3
				increased the wound-healing
				rate in U251 cells at 48 h,
				suggesting SOX3 as an
			Bioinformatic; Cell	antioncogenic.
			transfection; Migration;	Downregulation of SOX3 has
			Transwell invasion	no significant effect on U251
PAN et al., 2021 [52]	Glioblastoma	Cell line: U251	assays; RT-qPCR.	cell invasion.
				Treatment of GBM with
				BX795 (inhibitor of TBK1-
				TANK-binding kinase)
				showed a significant
		Cell lines: U-		reduction in SOX3 gene and
	Glioblastoma	138MG, U-87 MG,	Cell viability; RT-qPCR;	protein expression in GBM
SCUDERI et al., 2021 [74]	Multiforme	U-138 and U-87.	WB and ELISA.	cells.

				1/
			Bioinformatics; Cell	
			transfection with SOX3	
			specific siRNAs and	The protein CELF2 acts as an
			CELF2-specific shRNA;	epigenetic regulator in
			Orthotopic Xenografts	glioma stem cells and can
			animal model; RNA	repress the SOX3 gene,
			sequencing; ChIP	promoting a proliferating
		Cell lines:	Sequencing (ChIP-Seq);	tumor cell phenotype. CELF2
		Plasmacytoid	Spheroid formation	was found to be a significant
		Dendritic Cells	assays;	point of tumor vulnerability
		(PDC) from	Immunofluorescence;	as its repression is sufficient
		primary	Immunohistochemistry;	to convert aggressive tumor
		glioblastoma tumor	Immunohistofluorescen	cells into cells without the
				ability to form tumors <i>in vivo</i> .
		(GB1, GB5, GB11)	ce; RNA	,
TUDGUL ( 1 2000 Fee)	Cl: 11 ·	and TG6 cells (T	Immunoprecipitation	
TURCHI et al., 2023 [73]	Glioblastoma	lymphoblast).	and PCR.	
		Clinical specimens:		
		24 human glioma		SOX3 maintains neural cells
	Glioma	samples.	RT-qPCR; WB;	as self-renewing progenitors,
	(Glioblastoma,	Cell line: Primary	Immunohistochemistry;	keeping cells on the cell cycle
	Astrocytoma,	cell culture.	Immunofluorescence	and in a proliferative state.
HOLMBERG et al., 2011	Oligoastrocyto	Animal model:	and in situ	1
[57]	ma)	Nude mice.	hybridization.	
				The expression level of SOX3
				in glioma was significantly
				higher compared with the
				normal tissues. SOX3
				upregulation was associated
				with patients predicted poor
				outcomes. SOX3 mRNA
		Clinical specimens:		expression was higher in
		40 patients' glioma		glioma cell lines (LN18 and
				LN229) than in normal cell
		1		lines (HEB). SOX3 is
		corresponding	D	downregulated by miR-483,
		adjacent tissue	Bioinformatic; Cells	inhibiting invasion,
		samples.	transfection with SOX3	migration and promoting
		Cell lines: Gliocyte	expression vector and	apoptosis of glioma cells,
		HEB and glioma	miR-483; Luciferase	suggesting that miR- 483 can
	l	cells LN18 and	assay; RT-qPCR; WB and	be a potential target for
SHUJING et al., 2020 [38]			transwell invasion assay.	1

glloma tissue dinical specimens is upregulated compared to that in adjacent normal tissues. Bioinformatic tool TargetScan predicted SOX3 as the downstream target of mite-R83-3p Functional experiments of Dual-Luciferase reporter assay confirmed that milk 483-3p inhibited the activity of the SOX3-WT reporter. In vitro upregulated or SOX3 is pregulated in Final Dual Luciferase reporter assay confirmed that milk 483-3p inhibited the activity of the SOX3-WT reporter. In vitro upregulation of SOX3 expression or the inhibition of milk 483-3p expression or the inhibition of milk 483-3p expression peromotes the proliferation of U87 cells, which was blocked by UNCOMSC silencing. Anti-apoptotic protein Bel-2 (Cell line. HRB 5-Ethynyl-2- promotes the proliferation of U87 cells, which was blocked by UNCOMSC silencing. Anti-apoptotic protein Bel-2 (Cell line. HRB 5-Ethynyl-2- plasmids or milk 483-3p inhibitors. UNCOMSC silencing inhibit					
specimens is upregulated compared to that in adjacent normal tissues. Bioinformatic tool TargetScan predicted SOX3 as the downstream target of miR-483-ap Functional experiments of Dual-Luciferase reporter assay confirmed that milk 483-3p inhibited the activity of the SOX3-WT reporter. In vitro upregulation of SOX3 expression or the inhibition of miR-483-ap expression promotes the proliferation of of miR-483-ap expression promotes the proliferation of SOX3 expression was inhibited and the success of the south of the SOX3-WT reporter. In vitro upregulation of SOX3 expression was inhibited and to miR-483-ap expression promotes the proliferation of uR-483-ap expression promotes the proliferation of uR-483-ap expression of section with sox and adjacent normal brain tissues.  Cell line: HEB (Human normal glial cell) and Assay; Flow Cytometry; human glioma cell lines: U87, U251, LN229, and A172.  YUAN et al., 2022 [71] Glioma LN229, and A172.  Glioma LN239, and A172.  TNA390, Assay.  The mRNA expression of SOX3 is upregulated in the paticular carcinoma (HCC) itssues. The recurrence-free survival (RFS) rate of paticnts with high SOX3 ex					SOX3 gene expression in
compared to that in adjacent normal tissues. Bioinformatic tool TargetScan predicted SOX3 as the downstream target of miR-483-3p Functional experiments of Dual-Luciferase reporter assay confirmed that miR-483-3p inhibited the activity of the SOX3-WT reporter. In office our pregulation of SOX3 expression or the inhibition of miR-483-3p expression or miR-483-3p expressio					glioma tissue clinical
normal tissues. Bioinformatic tool TargetScan predicted SOX3 as the downstream target of miR-483-3p. Functional experiments of Dual-Luciferase reporter assay confirmed that miR-483-3p inhibited the activity of the SOX3-WT reporter. In vitro upregulation of SOX3 expression or the inhibition of miR-483-3p expression or the inhibition of miR-483-3p expression or more tissues and adjacent normal brain tissues  Cell line: HEB (Human normal dissues the proliferation of U87 cells, which was blocked by LINC00662 silencing the U87 cells, which was blocked by LINC00662 silencing promotes the proliferation of U87 cells, which was blocked by LINC00662 silencing promotes the proliferation of miR-483-3p expression was inhibited and reversed by co-transfection with SOX3 overexpression with SOX3 overexpression promotes the proliferation of U87 cells, which was blocked by LINC00662 silencing promotes the proliferation of U87 cells, which was blocked by LINC00662 silencing promotes the proliferation of U87 cells, which was blocked by LINC00662 silencing promotes the proliferation of miR-483-3p expression was inhibited and reversed by co-transfection with SOX3 overexpression with SOX3 overexpression was full apoptodic protein Bd-2 expression was inhibited and reversed by co-transfection with SOX3 overexpression was full apoptodic protein Bd-2 expression was full apoptodic protein Bd-2 expression was inhibited and reversed by co-transfection with SOX3 overexpression was full apoptodic protein Bd-2 expression was full apoptod					specimens is upregulated
Clinical specimens: 50 glioma tumor tissues and adjacent normal brain tissues Cell line: HEB (Human normal blines: U87, U251, Immunoprecipitation   LN229, and A172.  VUAN et al., 2022 [71] Glioma LN229, and A172.  To the service of mik-483-3p expression promotes the proliferation of of mik-483-3p expression promotes the proliferation of u87 cells, which was blocked by LINC00662 silencing Anti-apoptotic protein Bel-2 expression was inhibited and reversed by co-transfection with SOX3 overexpression plasmids or mik-483-3p expression was inhibited and reversed by co-transfection with SOX3 overexpression plasmids or mik-483-3p expression was inhibited and reversed by co-transfection with SOX3 overexpression plasmids or mik-483-3p expression was inhibited and reversed by co-transfection with SOX3 overexpression plasmids or mik-483-3p inhibitors. Lincom662 silencing Anti-apoptotic protein Bel-2 expression was inhibited and reverse d by co-transfection with SOX3 overexpression valuating the mik-483-3p inhibitors. Lincom662 silencing Anti-apoptotic protein Bel-2 expression was inhibited and reverse d by co-transfection with SOX3 overexpression was formal plasmids or mik-483-3p inhibitors. Lincom662 silencing Anti-apoptotic protein Bel-2 expression was inhibited and reverse d by co-transfection with SOX3 overexpression or mik-483-3p inhibitors. Lincom662 silencing Anti-apoptotic protein Bel-2 expression was inhibited and reverse d by co-transfection with SOX3 overexpression or mik-483-3p inhibitors. Lincom662 silencing Anti-apoptotic protein Bel-2 expression was inhibited and reverse d by co-transfection with SOX3 overexpression in mik-483-3p inhibitors. Lincom662 silencing Anti-apoptotic protein Bel-2 expression was inhibited and reverse d by co-transfection with SOX3 overexpression or mik-483-3p inhibitors. Lincom662 silencing Anti-apoptotic protein Bel-2 expression was inhibited and reverse d by co-transfection with SOX3 overexpression in mik-483-3p inhibitors. Lincom662 silencing Anti-apoptotic protein Bel-2 expressi					compared to that in adjacent
SOX3 as the downstream target of miR-483-3p. Functional experiments of Dual-Luciferase reporter assay confirmed that miR 483-3p inhibited the activity of the SOX3-WT reporter. In witro upregulation of SOX3 expression or the inhibition of miR-483-3p expression promotes the proliferation of U87 cells, which was blocked by LINC00662 silencing. Anti-apoptotic protein Bel-2 expression was inhibited and reversed by co-transfection with SOX3 overexpression was inhibited and reversed by co-transfection with SOX3 overexpression was inhibited and reversed by co-transfection with SOX3 overexpression was inhibited and reversed by co-transfection with SOX3 overexpression with SOX3 overexpression with SOX3 overexpression was inhibited and reversed by co-transfection with SOX3 overexpression with SOX3 overexpression with SOX3 overexpression was inhibited and reversed by co-transfection with SOX3 overexpression was inhibited and reversed by co-transfection with SOX3 overexpression was inhibited and reversed by co-transfection with SOX3 overexpression was inhibited and reversed by co-transfection with SOX3 overexpression was inhibited and reversed by co-transfection with SOX3 overexpression was inhibited and reversed by co-transfection with SOX3 overexpression was inhibited and reversed by co-transfection with SOX3 overexpression was inhibited and reversed by co-transfection with SOX3 overexpression was inhibited and reversed by co-transfection with SOX3 overexpression was inhibited and reversed by co-transfection with SOX3 overexpression was inhibited and reversed by LINC00662 silencing. Anti-apoptotic protein Bel-2 expression was inhibited and reversed by LINC00662 silencing. Anti-apoptotic protein Bel-2 expression over mix-483-3p inhibited and reversed by LINC00662 silencing. Anti-apoptotic protein Bel-2 expression was inhibited and reversed by LINC00662 silencing. Anti-apoptotic protein Bel-2 expression over transfer and protein protein Bel-2 expression over transfer and protein protein Bel-2 expression over transfer and pr					normal tissues. Bioinformatic
target of miR-483-3p. Functional experiments of Dual-Luciferase reporter assay confirmed that miR-483-3p inhibited the activity of the SOX3-WT reporter. In vitro upregulation of SOX3 expression or the inhibition of miR-483-3p expression promotes the proliferation of US7 cells, which was blocked by LINC00662 silencing. Anti-apoptotic protein Bcl-2 expression was inhibited and reversed by co-transfection WB;  Cell line: HEB 5-Ethynyl-2'- decayuridine (EdU) glial cell) and Assay; Flow Cytometry: human glioma cell transwell Assay; RNA lines: US7, U251, Immunoprecipitation  VUAN et al., 2022 [71] Glioma LIN229, and A172. (RIP) Assay.  The mRNA expression of SOX3 is upregulated in Hepatocellular carcinoma (HCC) tissues. The recurrence-free survival (RFS) rate of patients with high SOX3 expression was considerably lower than that of patients with basal SOX3 expression. SOX3 overexpression.					tool TargetScan predicted
Clinical specimens: 50 glioma tumor tissues and adjacent normal brain tissues Cell line: HEB (Human normal glial cell) and Assay; Cell line: HEB (Human glioma cell lines: US7, U251, LN229, and A172.  YUAN et al., 2022 [71] Glioma					SOX3 as the downstream
Clinical specimens: 50 glioma tumor tissues and adjacent normal brain tissues Cell line: HEB (Human normal glial cell) and human glioma cell lines: U87, U251, LN229, and A172.  YUAN et al., 2022 [71] Glioma				target of miR-483-3p.	
Clinical specimens: 50 glioma tumor tissues and adjacent normal brain tissues Cell line: HEB (Human normal glial cell) and human glioma cell lines: U87, U251, LN229, and A172.  Glioma  Clinical specimens: 50 glioma tumor tissues and adjacent normal brain tissues Cell Proliferation Assay; 5-Ethynyl-2'- deoxyuridine (EdU) glial cell) and human glioma cell lines: U87, U251, LN229, and A172.  Glioma  CRIP) Assay.  Transwell Assay; RNA lmmunoprecipitation (RIP) Assay.  The mRNA expression of SOX3 axis.  The mRNA expression of SOX3 is upregulated in Hepatocellular carcinoma (HCC) tissues. The recurrence-free survival (RFS) rate of patients with high SOX3 expression was considerably lower than that of patients with basal SOX3 expression. SOX3 overexpression was statistically correlated with					Functional experiments of
Clinical specimens: 50 glioma tumor tissues and adjacent normal brain tissues Cell line: HEB (Human normal glial cell) and human glioma cell lines: U87, U251, LN229, and A172.  TYUAN et al., 2022 [71] Glioma  Gliom					Dual-Luciferase reporter
Clinical specimens: 50 glioma tumor tissues and adjacent normal brain tissues Cell line: HEB (Human normal glial cell) and human glioma cell lines: U87, U251, LN229, and A172.  TYUAN et al., 2022 [71] Glioma  Gliom					assay confirmed that miR-
Clinical specimens: 50 glioma tumor tissues and adjacent normal brain tissues Cell line: HEB (Human normal glial cell) and human glioma cell lines: U87, U251, lines: U87, U25					-
Clinical specimens: 50 glioma tumor tissues and adjacent normal brain tissues Cell line: HEB (Human normal plial cell) and human glioma cell lines: U87, U251, lines: U87, U25					
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Clinical specimens: 50 glioma tumor tissues and adjacent normal brain tissues Cell Proliferation Assay; Cell line: HEB (Human normal glial cell) and human glioma cell human glioma cell LN229, and A172.  YUAN et al., 2022 [71] Glioma  Clinical specimens: 50 glioma tumor tissues and adjacent mormal brain tissues Cell Proliferation Assay; Cell Proliferation Assay; Cell Proliferation Assay; Sethynyl-2*- (Human normal dasay; Flow Cytometry; transwell Assay; RNA lines: U87, U251, Immunoprecipitation to SOX3 is upregulated in Hepatocellular carcinoma (HCC) tissues. The recurrence-free survival (RFS) rate of patients with high SOX3 expression was considerably lower than that of patients with basal SOX3 expression. SOX3 overexpression was statistically correlated with					
Clinical specimens: 50 glioma tumor tissues and adjacent normal brain tissues Cell line: HEB (Human normal glial cell) and human glioma cell lines: U87, U251, Immunoprecipitation (RIP) Assay.  Glioma  The mRNA expression of SOX3 is upregulated in Hepatocellular carcinoma (HCC) tissues. The recurrence-free survival (RFS) rate of patients with high SOX3 expression was statistically correlated with					
Clinical specimens: 50 glioma tumor tissues and adjacent normal brain tissues Cell line: HEB (Human normal glial cell) and human glioma cell lines: U87, U251, lines: U87, U251, LN229, and A172.  Glioma  Glioma  LN229, and A172.  U87 cells, which was blocked by LINC00662 silencing, Anti-apoptotic protein Bcl-2 expression was inhibited and reversed by co-transfection with SOX3 overexpression plasmids or miR-483-3p inhibitors. LINC00662 in					
Clinical specimens: 50 glioma tumor tissues and adjacent normal brain tissues Cell line: HEB (Human normal glial cell) and human glioma cell lines: U87, U251, Immunoprecipitation LN229, and A172.  Glioma  Clioma  Clioma  LN229, and A172.  Clioma  Clioma  Clioma  LN229, and A172.  Clioma  Cliom					
tissues and adjacent normal brain tissues  Cell line: HEB (Human normal glial cell) and lines: U87, U251, LN229, and A172.  YUAN et al., 2022 [71] Glioma  Glioma  Glioma  Glioma  Glioma  Cell New HEB (Human normal deoxyuridine (EdU) and lines: U87, U251, LN229, and A172.  Glioma  Glioma  LN229, and A172.  Glioma  Anti-apoptotic protein Bcl-2 expression was inhibited and reversed by co-transfection with SOX3 overexpression plasmids or miR-483-3p inhibitors. LINC00662 triggered tumor-promoting effects in gliomas via modulating the miR-483-3p/SOX3 axis.  The mRNA expression of SOX3 is upregulated in Hepatocellular carcinoma (HCC) tissues. The recurrence-free survival (RFS) rate of patients with high SOX3 expression was considerably lower than that of patients with basal SOX3 expression. SOX3 overexpression was statistically correlated with			Clinical specimens:		
tissues and adjacent normal brain tissues  Cell Proliferation Assay;  Cell line: HEB (Human normal glial cell) and human glioma cell lines: U87, U251, LN229, and A172.  Glioma  LN229, and A172.  Bioinformatic; RT-qPCR; WB; expression was inhibited and reversed by co-transfection with SOX3 overexpression with SOX3 overex			50 glioma tumor		
normal brain tissues Cell Proliferation Assay; Cell line: HEB 5-Ethynyl-2'- (Human normal glial cell) and human glioma cell lines: U87, U251, Immunoprecipitation LN229, and A172.  Glioma  LN229, and A172.  Glioma  Repart of patients with basal SOX3 expression was statistically correlated with sox3 overexpression was statistically correlated with sox3 overexpression was statistically correlated with sox3 overexpression was statistically correlated with			tissues and adjacent	Bioinformatic; RT-qPCR;	
tissues Cell line: HEB (Human normal glial cell) and human glioma cell lines: U87, U251, YUAN et al., 2022 [71] Glioma  LN229, and A172.  Cell Proliferation Assay; 5-Ethynyl-2'- deoxyuridine (EdU) Assay; Flow Cytometry; triggered tumor-promoting modulating the miR-483-			normal brain	WB;	_
Cell line: HEB (Human normal glial cell) and human glioma cell lines: U87, U251, LN229, and A172.  Glioma  Cell line: HEB (Human normal glial cell) and Assay; Flow Cytometry; triggered tumor-promoting modulating the miR-483-3p modulating the miR-483-3p modulating the miR-483-3p SOX3 axis.  The mRNA expression of SOX3 is upregulated in Hepatocellular carcinoma (HCC) tissues. The recurrence-free survival (RFS) rate of patients with high SOX3 expression was considerably lower than that of patients with basal SOX3 expression. SOX3 overexpression was statistically correlated with			tissues	Cell Proliferation Assay;	-
(Human normal glial cell) and Assay; Flow Cytometry; human glioma cell Transwell Assay; RNA lines: U87, U251, Immunoprecipitation LN229, and A172. (RIP) Assay.  The mRNA expression of SOX3 is upregulated in Hepatocellular carcinoma (HCC) tissues. The recurrence-free survival (RFS) rate of patients with high SOX3 expression was considerably lower than that of patients with basal SOX3 expression. SOX3 overexpression was statistically correlated with			Cell line: HEB	5-Ethynyl-2'-	_
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lines: U87, U251, Immunoprecipitation modulating the miR-483-3p/SOX3 axis.  The mRNA expression of SOX3 is upregulated in Hepatocellular carcinoma (HCC) tissues. The recurrence-free survival (RFS) rate of patients with high SOX3 expression was considerably lower than that of patients with basal SOX3 expression. SOX3 overexpression was statistically correlated with					
YUAN et al., 2022 [71]  Glioma  LN229, and A172. (RIP) Assay.  The mRNA expression of SOX3 is upregulated in Hepatocellular carcinoma (HCC) tissues. The recurrence-free survival (RFS) rate of patients with high SOX3 expression was considerably lower than that of patients with basal SOX3 expression. SOX3 overexpression was statistically correlated with			_	-	8
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of patients with basal SOX3 expression. SOX3 overexpression was statistically correlated with					high SOX3 expression was
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overexpression was statistically correlated with					of patients with basal SOX3
statistically correlated with					expression. SOX3
					overexpression was
Hepatocellular Clinical specimens: RT-qPCR; WB; less tumor capsule formation,					statistically correlated with
		Hepatocellular	Clinical specimens:	RT-qPCR; WB;	less tumor capsule formation,
FENG et al., 2017 [54] carcinoma 50 patients Immunohistochemistry. worse degrees of tumor	FENG et al., 2017 [54]	carcinoma	50 patients	Immunohistochemistry.	worse degrees of tumor

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				differentiation, and worse
				TNM classification. Results
				suggested SOX3 plays an
				oncogenic role in HCC.
				SOX3 mRNA was not
				detected in serum from
				normal adult tissues. SOX3
				mRNA was detected in 2 out
				of 10 cell lines. SOX3 is not
				detectable in normal lung
				adult tissues. SOX3
				expression was detected in
				10% of adult lung cancer
				tissue. All patients with
		Clinical specimens:		antibodies against SOX3 or
		17 patients' serum		SOX21 had higher reactivity
		with lung cancer		against SOX1 and SOX2. The
		and 23 control		seroreactivity to SOX3 and
		patients.		SOX21 might be secondary to
		Cell line SK-LC-13;		the shared antigenic epitopes
		NCI-H69, 128, 146,		located within the highly
GURE et al., 2000		187, 209, 378, 889,	RT-qPCR; Northern Blot	conserved HMG box of SOX
[79]	Lung	740;	(NB).	proteins.
				SOX3 was overexpressed in
				most osteosarcoma tissues
				compared with that in bone
				cysts. SOX3 expression
				correlates with Snail1 and E-
				cadherin in human OS
		Clinical specimens:		tissues. The mechanistic link
		42 Osteosarcoma		among SOX3, Snail1, and
		tissues; non-tumor		EMT indicates SOX3 as a
		samples 42; and	RT-qPCR; WB and	potential therapeutic target
QIU et al., 2017 [47]	Osteosarcoma	bone cysts 28.	Immunohistochemistry.	for osteosarcoma metastasis.
			RT-qPCR; WB;	SOX3 promotes
			Luciferase assay;	osteosarcoma cell migration
			Chromatin	invasion and induces EMT
		Cell lines: U2OS,	immunoprecipitation;	upregulating Snail1
		SoSP-M, and MG-	Cell migration and	expression in osteosarcoma
QIU et al., 2017 [47]	Osteosarcoma	63	matrigel invasion; in vivo	cells.

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			lung metastasis model;	
			Immunohistochemistry.	
				Upregulation of SOX3
				mRNA and protein
				expression level in human
				osteosarcoma tissues. SOX3
		Clinical anadimona		
		Clinical specimens:		acts as an oncogene in
		70 patients with		osteosarcoma, and SOX3
		primary		inhibitors or downstream
		osteosarcoma and		effectors may be attractive
		20 patients with		targets for osteosarcoma
GUO et al., 2018 [33]	Osteosarcoma	bone cysts	RT-qPCR and WB	therapy.
				SOX3 knockdown in
				osteosarcoma cells inhibits
				the proliferation, induces G1
				phase arrest, induces
				apoptosis, suppresses the
				migration and invasion,
				suppresses tumor growth in a
				xenograft mouse model,
				decreases the EMT-
				promoting proteins (Twist,
				Snail, and MMP-9) and
				increased E-cadherin. SOX3
				acts as an oncogene in
			SOX3 mRNA silencing;	osteosarcoma, and SOX3
			WB; Cell proliferation;	inhibitors or downstream
		Cell lines: MG63	Cell cycle; Cell migration	effectors may be interesting
		and U2OS human	and invasion assays and	targets for osteosarcoma
GUO et al., 2018 [33]	Osteosarcoma	osteosarcoma cells	cell apoptosis analysis.	therapy.
		Clinical specimens:		SOX3 immunoreactivity in
		142 patients with		human ovarian tumor cells
		ovarian carcinoma,		was mainly localized to the
		28 patients with		nuclei. None of the normal
		borderline ovarian		ovarian tissue samples were
		cystadenoma, 33		positive for SOX3 expression,
		patients with		whereas SOX3-positive
		ovarian		epithelial cells were detected
YAN et al., 2016 [35]	Ovarian	cystadenoma, and	Immunohistochemistry.	in ovarian cystadenoma,
1111 Ct ally 2010 [00]	Varian	cyonactionia, and	y.	ii. Ovarian cystauchoma,

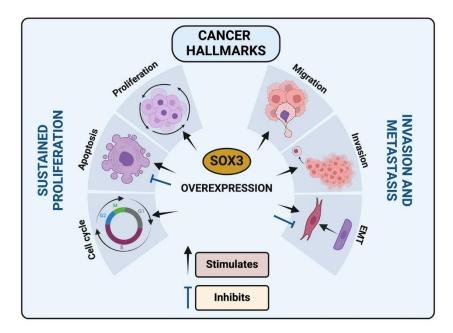
	-	i		,
		25 as normal		borderline ovarian tumors,
		controls.		and ovarian cancer epithelial
				tissues. SOX3 expression
				gradually increased from
				benign and borderline to
				malignant ovarian tumors.
				SOX3 may be involved in the
				malignant transformation of
				ovarian tumors and may be
				used as a supplementary
				indication in the diagnosis of
				epithelial ovarian cancer.
				SOX3 expression was
				different in each cell line.
				SOX3 promotes proliferation,
				migration, invasion
				and inhibits the adhesion of
				ovarian cancer cells. SOX3
				inhibits apoptosis of ovarian
				cancer cells. Overexpression
				of SOX3 leads to high
				phosphorylation of pro-
				metastatic proteins. SOX3
			Cell transfection; RT-	expression was relatively
			PCR; WB; Cell	
			immunofluorescence;	cell lines SKOV3-ip
		Cell lines: HO8910;	Cell proliferation;	
		HO8910-pm;	Colony formation; Cell	suggesting that SOX3 may
		SKOV3; SKOV3-ip;	migration and invasion;	play a key role in cell
		ES2; MCV-152; and		migration and tumor
YAN et al., 2016 [35]	Ovarian	Moody.	analysis.	metastasis.
	- 1,1		y	Overexpression of SOX2 or
		Clinical specimens:	Immunohistochemistry;	SOX3 enhanced both ALK
		135 cases of ovarian	In situ hybridization	and ELAVL3 promoter
		carcinomas.	fluorescence; Mutation	activities, suggesting the
		Cell lines: High-	Analysis of the ALK and	existence of ALK/Sox/HuC
		grade serous	TP53 Genes.	signaling loops. ALK
		carcinoma (HGSC)	Cells Transfection; RT-	overexpression was
		cell lines OVSAHO,	qPCR; WB; Flow	attributed to increased
	Ovarian	OVKATE, and	Cytometry; Spheroid	expression of
MATSUMOTO et al.,	Serous	OVCAR-3, and	assay; Cell counting	neuroendocrine markers,
2021 [62]	Carcinoma	ovarian clear cell	assay; Wound-Healing	including synaptophysin,

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	carcinoma	(CCC)	Assay	and	RNA	CD56, and B-cell lymphoma
	cell		sequenci	ng.		2, in HGSC tissues. These
	lines, OVISI	E, ES-2,				findings suggest that
	OVTOKO, I	KOC7C,				overexpression of full-length
	and TOV-21	G.				ALK may influence the
						biological behavior of HGSC
						through cooperation with
						ELAVL3 and Sox factors,
						leading to the establishment
						and maintenance of the
						aggressive phenotypic
						characteristics of HGSC.
						SOX3 expression increased in
						transfected cells with ALK-
						overexpressing vector but not
						in ALK-knockdown cells.
						Expression of SOX proteins
						was increased following ALK
						overexpression, suggesting
						the existence of a positive
						feedback loop between ALK
						and Sox factors. SOX3
						induces ALK (anaplastic
						lymphoma kinase)
						overexpression in ovarian
						serous carcinoma.

# 4. Conclusion and Future Perspective

This comprehensive review describes the multifaceted role of the SOX3 transcription factor within the cancer paradigm. The evidence presented underscores the complexity of SOX3 involvement in the modulation of critical cancer hallmarks, including apoptosis, EMT, invasion, migration, cell cycle regulation, and proliferation (Figure 4). It is apparent that SOX3 function is highly context-dependent, varying significantly across cancer types, and is influenced by the intricate interplay of genetic, epigenetic, and environmental factors. SOX3's dualistic nature as a potential tumor suppressor in certain contexts and a promoter of tumorigenesis in others presents both challenges and opportunities for therapeutic intervention. The correlation of SOX3 expression with clinical outcomes in various cancers emphasizes its potential as a prognostic biomarker and a molecular target for cancer therapy.



**Figure 4.** SOX3 overexpression in different cancer types can modulate cell cycle, apoptosis, proliferation, migration, invasion and epithelial-mesenchymal transition (EMT).

Future perspectives aim to elucidate the complex signaling pathways and interactions involving SOX3 to harness its full potential in the battle against cancer. Revealing the precise molecular mechanisms through which SOX3 influences cancer hallmarks is essential, including deciphering its interactions with other TFs, signaling pathways, and tumor microenvironment cells and components. As a perspective of SOX3 as a therapeutic target, the present review showed that this may include small molecule inhibitors, such as miRNAs and monoclonal antibodies, designed to either inhibit or enhance SOX3 function, tailored to the specific context of its role in various cancers [38,71,73,74]. Adding to that is essential the integration with Omics Data (genomics, transcriptomics, proteomics), which can help identify novel targets and pathways influenced by SOX3 [38,52,71,73]. Finally, to apply SOX3 in clinical correlation, it is necessary to expand the scope of clinical studies to explore the association between SOX3 expression levels, patient prognosis, and treatment responses across a broader range of cancer types. Such studies should aim to validate SOX3 as a biomarker for cancer diagnosis, prognosis, and prediction of therapeutic response. Investigating the potential of SOX3 as a therapeutic target involves developing and testing novel strategies to modulate its activity.

**Author Contributions:** H.L.D.P., A.P.G., A.U., D.Q. and J.C. designed, discussed the ideas and topics, conducted the literature review, wrote, revised and edit the manuscript. B.R.S., N.O.D., T.B.G.L., D.Y.O.T. conducted the literature rieview and revised the manuscript. E.F., F.H.S.S., B.M.L., B.A.C., E.G., D.C.C., E.T.P., S.M.Q. and A.E. wrote, edited and revised the manuscript. All authors read and approved the final manuscript.

**Funding statement:** Research in the HLDP lab is supported by Fundação de Amparo à Pesquisa do Estado de Minas Gerais (FAPEMIG - grant number APQ-01247-23) and CAPES Print Senior Visiting Professor program 05/2022. Research in the JSC lab is supported by the National Institute of General Medical Sciences of the National Institute of Health (R15CA267890-01).

Conflicts of Interest: The authors declare no conflict of interest.

# References

- 1. Lambert, S. A.; Jolma, A.; Campitelli, L. F.; Das, P. K.; Yin, Y.; Albu, M.; Chen, X.; Taipale, J.; Hughes, T. R.; Weirauch, M. T. The Human Transcription Factors. *Cell* **2018**, *172*, 650–665.
- 2. Sarkar, A.; Hochedlinger, K. The Sox Family of Transcription Factors: Versatile Regulators of Stem and Progenitor Cell Fate. *Cell Stem Cell* **2013**, 12, 15–30.
- 3. Kashimada, K.; Koopman, P. Sry: The master switch in mammalian sex determination. *Development* **2010**, 137, 3921–3930.

- 4. Berta, P.; Hawkins, J. B.; Sinclair, A. H.; Taylor, A.; Griffiths, B. L.; Goodfellow, P. N.; Fellous, M. Genetic evidence equating SRY and the testis-determining factor. *Nature* **1990**, *348*, 448–450.
- 5. Harley, V. R.; Lovell-badge, R.; Goodfellow, P. N. Definition of a consensus DNA binding site for SRY. *Nucleic Acids Res.* **1994**, 22, 1500–1501.
- 6. Harley, V. R.; Jackson, D. I.; Hextall, P. J.; Hawkins, J. R.; Berkovitz, G. D.; Sockanathan, S.; Lovell-Badge, R.; Goodfellow, P. N. DNA Binding Activity of Recombinant SRY from Normal Males and XY Females. *Science* (80-.). 1992, 255, 453–456.
- 7. Turner, M. E.; Ely, D.; Prokop, J.; Milsted, A. Sry, more than testis determination? *Am. J. Physiol. Regul. Integr. Comp. Physiol.* **2011**, 301.
- 8. Bowles, J.; Schepers, G.; Koopman, P. Phylogeny of the SOX family of developmental transcription factors based on sequence and structural indicators. *Dev. Biol.* **2000**, 227, 239–255.
- 9. Wegner, M. SURVEY AND SUMMARY From head to toes: the multiple facets of Sox proteins. *Nucleic Acids Res.* **1999**, *27*, 1409–1420.
- 10. Hou, L.; Srivastava, Y.; Jauch, R. Molecular basis for the genome engagement by Sox proteins. *Semin. Cell Dev. Biol.* **2017**, *63*, 2–12.
- 11. Hur, W.; Rhim, H.; Jung, C. K.; Kim, J. D.; Bae, S. H.; Jang, J. W.; Yang, J. M.; Oh, S. T.; Kim, D. G.; Wang, H. J.; et al. SOX4 overexpression regulates the p53-mediated apoptosis in hepatocellular carcinoma: Clinical implication and functional analysis in vitro. *Carcinogenesis* **2010**, *31*, 1298–1307.
- 12. Bernard, P.; Harley, V. R. Acquisition of SOX transcription factor specificity through protein-protein interaction, modulation of Wnt signalling and post-translational modification. *Int. J. Biochem. Cell Biol.* **2010**, 42, 400–410.
- 13. Swain, N.; Thakur, M.; Pathak, J.; Swain, B. SOX2, OCT4 and NANOG: The core embryonic stem cell pluripotency regulators in oral carcinogenesis. *J. Oral Maxillofac. Pathol.* **2020**, 24, 368.
- 14. Xu, Y. R.; Yang, W. X. SOX-mediated molecular crosstalk during the progression of tumorigenesis. *Semin. Cell Dev. Biol.* **2017**, *63*, 23–34.
- 15. Abadi, A. J.; Zarrabi, A.; Hashemi, F.; Zabolian, A.; Najafi, M.; Entezari, M.; Hushmandi, K.; Aref, A. R.; Khan, H.; Makvandi, P.; et al. The role of SOX family transcription factors in gastric cancer. *Int. J. Biol. Macromol.* **2021**, *180*, 608–624.
- 16. Jiang, J.; Wang, Y.; Sun, M.; Luo, X.; Zhang, Z.; Wang, Y.; Li, S.; Hu, D.; Zhang, J.; Wu, Z.; et al. SOX on tumors, a comfort or a constraint? *Cell Death Discov.* **2024**, *10*.
- 17. Castillo, S. D.; Sanchez-Cespedes, M. The SOX family of genes in cancer development: Biological relevance and opportunities for therapy. *Expert Opin. Ther. Targets* **2012**, *16*, 903–919.
- 18. Ashrafizadeh, M.; Taeb, S.; Hushmandi, K.; Orouei, S.; Shahinozzaman, M.; Zabolian, A.; Moghadam, E. R.; Raei, M.; Zarrabi, A.; Khan, H.; et al. Cancer and SOX proteins: New insight into their role in ovarian cancer progression/inhibition. *Pharmacol. Res.* **2020**, *161*, 105159.
- 19. Thu, K. L.; Becker-Santos, D. D.; Radulovich, N.; Pikor, L. A.; Lam, W. L.; Tsao, M. S. SOX15 and other SOX family members are important mediators of tumorigenesis in multiple cancer types. *Oncoscience* **2014**, *1*, 326–335
- 20. Tan, D. S.; Holzner, M.; Weng, M.; Srivastava, Y.; Jauch, R. SOX17 in cellular reprogramming and cancer. *Semin. Cancer Biol.* **2020**, *67*, 65–73.
- 21. Underwood, A.; Rasicci, D. T.; Hinds, D.; Mitchell, J. T.; Zieba, J. K.; Mills, J.; Arnold, N. E.; Cook, T. W.; Moustaqil, M.; Gambin, Y.; et al. Evolutionary Landscape of SOX Genes to Inform Genotype-to-Phenotype Relationships. *Genes (Basel)*. **2023**, *14*.
- 22. Qian, M.; Wang, D. C.; Chen, H.; Cheng, Y. Detection of single cell heterogeneity in cancer. *Semin. Cell Dev. Biol.* **2017**, *64*, 143–149.
- 23. Wu, F.; Fan, J.; He, Y.; Xiong, A.; Yu, J.; Li, Y.; Zhang, Y.; Zhao, W.; Zhou, F.; Li, W.; et al. Single-cell profiling of tumor heterogeneity and the microenvironment in advanced non-small cell lung cancer. *Nat. Commun.* **2021**, *12*, 1–11.
- 24. Nguyen, A.; Yoshida, M.; Goodarzi, H.; Tavazoie, S. F. Highly variable cancer subpopulations that exhibit enhanced transcriptome variability and metastatic fitness. *Nat. Commun.* **2016**, *7*, 1–13.
- 25. Ilan, Y.; Spigelman, Z. Establishing patient-tailored variability-based paradigms for anti-cancer therapy: Using the inherent trajectories which underlie cancer for overcoming drug resistance. *Cancer Treat. Res. Commun.* **2020**, *25*, 100240.
- 26. Yang, J.; Xu, J.; Wang, W.; Zhang, B.; Yu, X.; Shi, S. Epigenetic regulation in the tumor microenvironment: molecular mechanisms and therapeutic targets. *Signal Transduct. Target. Ther.* **2023**, 8.
- 27. Wuebben, E. L.; Rizzino, A. The dark side of SOX2: Cancer A comprehensive overview. *Oncotarget* **2017**, *8*, 44917–44943.
- 28. Mirzaei, S.; Paskeh, M. D. A.; Entezari, M.; Mirmazloomi, S. reza; Hassanpoor, A.; Aboutalebi, M.; Rezaei, S.; Hejazi, E. S.; Kakavand, A.; Heidari, H.; et al. SOX2 function in cancers: Association with growth, invasion, stemness and therapy response. *Biomed. Pharmacother.* **2022**, *156*, 113860.

- 29. Rodrigues, R. C.; Almeida, C. P.; Oliveira, M. C. M.; Ferreira, E.; Ribeiro, T. S.; Borges, I. T.; Gomes, H. W.; Oliveira, C. A.; Puerto, H. L. Del; Martins, A. S. Overexpression of SOX2 is associated with poor prognosis in human breast cancer. *J. Clin. Images Med. Case Reports* **2021**, 2.
- 30. Lengerke, C.; Fehm, T.; Kurth, R.; Neubauer, H.; Scheble, V.; Müller, F.; Schneider, F.; Petersen, K.; Wallwiener, D.; Kanz, L.; et al. Expression of the embryonic stem cell marker SOX2 in early-stage breast carcinoma. *BMC Cancer* **2011**, *11*, 42.
- 31. Feng, X.; Lu, M. Expression of sex-determining region Y-box protein 2 in breast cancer and its clinical significance. *Saudi Med. J.* **2017**, *38*, 685–690.
- 32. Chen, Y.; Shi, L.; Zhang, L.; Li, R.; Liang, J.; Yu, W.; Sun, L.; Yang, X.; Wang, Y.; Zhang, Y.; et al. The molecular mechanism governing the oncogenic potential of SOX2 in breast cancer. *J. Biol. Chem.* **2008**, *283*, 17969–17978.
- 33. Guo, Y.; Yin, J.; Tang, M.; Yu, X. Downregulation of SOX3 leads to the inhibition of the proliferation, migration and invasion of osteosarcoma cells. *Int. J. Oncol.* **2018**, *52*, 1277–1284.
- 34. Vicentic, J. M.; Drakulic, D.; Garcia, I.; Vukovic, V.; Aldaz, P. SOX3 can promote the malignant behavior of glioblastoma cells. *Cell. Oncol.* **2018**, 42, 41–54.
- 35. Yan, Q.; Wang, F.; Miao, Y.; Wu, X. Sex-determining region Y-box3 (SOX3) functions as an oncogene in promoting epithelial ovarian cancer by targeting Src kinase. *Tumor Biol.* **2016**, *37*, 12263–12271.
- 36. Cui, K.; Zhang, H.; Wang, G. MiR-483 suppresses cell proliferation and promotes cell apoptosis by targeting SOX3 in breast cancer. *Eur. Rev. Med. Pharmacol. Sci.* **2019**, 23, 2069–2074.
- 37. Almeida, C. P.; Ferreira, M. C. F.; Silveira, C. O.; Campos, J. R.; Borges, I. T.; Baeta, P. G.; Silva, F. H. S.; Reis, F. M.; Del Puerto, H. L. Clinical correlation of apoptosis in human granulosa cells—A review. *Cell Biol. Int.* **2018**, 42, 1276–1281.
- 38. Lu, S.; Yu, Z.; Zhang, X.; Sui, L. MiR-483 targeted SOX3 to suppress glioma cell migration, invasion and promote cell apoptosis. *Onco. Targets. Ther.* **2020**, *13*, 2153–2161.
- 39. Silva, F. H. de S.; Underwood, A.; Almeida, C. P.; Ribeiro, T. S.; Souza-Fagundes, E. M.; Martins, A. S.; Eliezeck, M.; Guatimosim, S.; Andrade, L. O.; Rezende, L.; et al. Transcription factor SOX3 upregulated pro-apoptotic genes expression in human breast cancer. *Med. Oncol.* **2022**, *39*, 1–10.
- 40. Carneiro, B. A.; El-Deiry, W. S. Targeting apoptosis in cancer therapy. Nat. Rev. Clin. Oncol. 2020, 17, 395-417.
- 41. Ribatti, D.; Tamma, R.; Annese, T. Epithelial-Mesenchymal Transition in Cancer: A Historical Overview. *Transl. Oncol.* **2020**, *13*, 100773.
- 42. Dongre, A.; Weinberg, R. A. New insights into the mechanisms of epithelial–mesenchymal transition and implications for cancer. *Nat. Rev. Mol. Cell Biol.* **2019**, *20*, 69–84.
- 43. Hugo, H.; Ackland, M. L.; Blick, T.; Lawrence, M. G.; Clements, J. A.; Williams, E. D.; Thompson, E. W. Epithelial—mesenchymal and mesenchymal—epithelial transitions in carcinoma progression. *J. Cell. Physiol.* **2007**, 213, 374–383.
- 44. Lee, T. K.; Poon, R. T. P.; Yuen, A. P.; Ling, M. T.; Kwok, W. K.; Wang, X. H.; Wong, Y. C.; Guan, X.; Man, K.; Chau, K. L.; et al. Twist overexpression correlates with hepatocellular carcinoma metastasis through induction of epithelial-mesenchymal transition. *Clin. Cancer Res.* **2006**, *12*, 5369–5376.
- 45. Kase, S.; Sugio, K.; Yamazaki, K.; Okamoto, T.; Yano, T.; Sugimachi, K. Expression of E-cadherin and β-catenin in human non-small cell lung cancer and the clinical significance. *Clin. Cancer Res.* **2000**, *6*, 4789–4796.
- 46. Pirinen, R. T.; Hirvikoski, P.; Johansson, R. T.; Hollmén, S.; Kosma, V. M. Reduced expression of  $\alpha$ -catenin,  $\beta$ -catenin, and  $\gamma$ -catenin is associated with high cell proliferative activity and poor differentiation in non-small cell lung cancer. *J. Clin. Pathol.* **2001**, *54*, 391–395.
- 47. Qiu, M.; Chen, D.; Shen, C.; Shen, J.; Zhao, H.; He, Y. Sex-determining region Y-box protein 3 induces epithelial-mesenchymal transition in osteosarcoma cells via transcriptional activation of Snail1. *J. Exp. Clin. Cancer Res.* **2017**, *36*, 1–11.
- 48. Gong, B.; Yue, Y.; Wang, R.; Zhang, Y.; Jin, Q.; Zhou, X. Overexpression of microRNA-194 suppresses the epithelial-mesenchymal transition in targeting stem cell transcription factor Sox3 in endometrial carcinoma stem cells. *Tumor Biol.* **2017**, 39.
- 49. Silva, F. H. S.; Underwood, A.; Almeida, C. P.; Lima, B. M.; Veloso, Emerson Soares; Carvalho, Barbara Andrade; Ribeiro, T. S.; Cassali, Geovanni Dantas; Ferreira, E.; Del Puerto, H. L. Transcription Factor SOX3 Regulates Epithelial mesenchymal Transition in Human Breast Cancer Cell Line MDA-MB-231. *Ann. Breast Cancer* 2024, 7, 1–4.
- 50. Acloque, H.; Ocaña, O. H.; Matheu, A.; Rizzoti, K.; Wise, C.; Lovell-Badge, R.; Nieto, M. A. Reciprocal repression between Sox3 and Snail transcription factors defines embryonic territories at gastrulation. *Dev. Cell* **2011**, *21*, 546–558.
- 51. Shen, J.; Zhai, J.; Wu, X.; Xie, G.; Shen, L. Serum proteome profiling reveals SOX3 as a candidate prognostic marker for gastric cancer. *J. Cell. Mol. Med.* **2020**, 24, 6750–6761.
- 52. Pan, C.; Liang, L.; Wang, Z.; Zhang, B.; Li, Q.; Tian, Y.; Yu, Y.; Chen, Z.; Wang, X.; Liu, H. Expression and significance of SOX B1 genes in glioblastoma multiforme patients. *J. Cell. Mol. Med.* **2021**, *26*, 789–799.

- 53. Li, K.; Wang, R.; Jiang, Y.; Zou, Y.; Guo, W. Overexpression of Sox3 is Associated with Diminished Prognosis in Esophageal Squamous Cell Carcinoma. *Ann Surg Onco* **2013**, *20*, 459–466.
- 54. Feng, Y.; Xiao, F.; Yang, N.; Zhu, N.; Fu, Y.; Zhang, H.; Yang, G. Overexpression of Sox3 is associated with promoted tumor progression and poor prognosis in hepatocellular carcinoma. *Int J Exp Pathol* **2017**, *10*, 7873–7881.
- 55. Matthews, H. K.; Bertoli, C.; de Bruin, R. A. M. Cell cycle control in cancer. *Nat. Rev. Mol. Cell Biol.* **2022**, 23, 74–88.
- 56. Bylund, M.; Andersson, E.; Novitch, B. G.; Muhr, J. Vertebrate neurogenesis is counteracted by Sox1-3 activity. *Nat. Neurosci.* **2003**, *6*, 1162–1168.
- 57. Holmberg, J.; He, X.; Peredo, I.; Orrego, A.; Hesselager, G.; Ericsson, C.; Hovatta, O.; Oba-Shinjo, S. M.; Marie, S. K. N.; Nistér, M.; et al. Activation of neural and pluripotent stem cell signatures correlates with increased malignancy in human glioma. *PLoS One* **2011**, *6*, 1–10.
- 58. Chew, L. J.; Gallo, V. The Yin and Yang of Sox proteins: Activation and repression in development and disease. *J. Neurosci. Res.* **2009**, *87*, 3277–3287.
- 59. Muhr, J.; Hagey, D. W. The cell cycle and differentiation as integrated processes: Cyclins and CDKs reciprocally regulate Sox and Notch to balance stem cell maintenance. *BioEssays* **2021**, *43*, 1–12.
- 60. Misaghi, A.; Goldin, A.; Awad, M.; Kulidjian, A. A. Osteosarcoma: A comprehensive review. Sicot-J 2018, 4.
- 61. Sambasivan, S. Epithelial ovarian cancer: Review article. Cancer Treat. Res. Commun. 2022, 33, 100629.
- 62. Matsumoto, T.; Oda, Y.; Hasegawa, Y.; Hashimura, M.; Oguri, Y.; Inoue, H.; Yokoi, A.; Tochimoto, M.; Nakagawa, M.; Jiang, Z.; et al. Anaplastic Lymphoma Kinase Overexpression Is Associated with Aggressive Phenotypic Characteristics of Ovarian High-Grade Serous Carcinoma. *Am. J. Pathol.* **2021**, 191, 1837–1850.
- 63. Swaminathan, H.; Saravanamurali, K.; Yadav, S. A. Extensive review on breast cancer its etiology, progression, prognostic markers, and treatment. *Med. Oncol.* **2023**, *40*, 1–26.
- 64. Dai, X.; Xiang, L.; Li, T.; Bai, Z. Cancer hallmarks, biomarkers and breast cancer molecular subtypes. *J. Cancer* **2016**, *7*, 1281–1294.
- 65. Liu, P.; Tang, H.; Song, C.; Wang, J.; Chen, B.; Huang, X.; Pei, X.; Liu, L. SOX2 promotes cell proliferation and metastasis in triple negative breast cancer. *Front. Pharmacol.* **2018**, *9*, 1–8.
- 66. Rodriguez-Pinilla, S. M.; Sarrio, D.; Moreno-Bueno, G.; Rodriguez-Gil, Y.; Martinez, M. A.; Hernandez, L.; Hardisson, D.; Reis-Filho, J. S.; Palacios, J. Sox2: A possible driver of the basal-like phenotype in sporadic breast cancer. *Mod. Pathol.* **2007**, *20*, 474–481.
- 67. Mehta, G. A.; Khanna, P.; Gatza, M. L. Emerging Role of SOX Proteins in Breast Cancer Development and Maintenance. *J. Mammary Gland Biol. Neoplasia* **2019**, 24, 213–230.
- 68. Xu, J.; Cao, W.; Shao, A.; Yang, M.; Andoh, V.; Ge, Q.; Pan, H. W.; Chen, K. P. Metabolomics of Esophageal Squamous Cell Carcinoma Tissues: Potential Biomarkers for Diagnosis and Promising Targets for Therapy. *Biomed Res. Int.* **2022**, 2022.
- 69. Sexton, R. E.; Al Hallak, M. N.; Diab, M.; Azmi, A. S. Gastric cancer: a comprehensive review of current and future treatment strategies. *Cancer Metastasis Rev.* **2020**, *39*, 1179–1203.
- 70. Nørøxe, D. S.; Poulsen, H. S.; Lassen, U. Hallmarks of glioblastoma: A systematic review. *ESMO Open* **2016**, 1, 1–9.
- 71. Yuan, L.; Zhang, P.; Lu, Y.; Zhang, A.; Chen, X. LINC00662 Promotes Proliferation and Invasion and Inhibits Apoptosis of Glioma Cells Through miR-483-3p/SOX3 Axis. *Appl. Biochem. Biotechnol.* **2022**, 194, 2857–2871.
- 72. Sa, J. K.; Kim, S. H.; Lee, J. K.; Cho, H. J.; Shin, Y. J.; Shin, H.; Koo, H.; Kim, D.; Lee, M.; Kang, W.; et al. Identification of genomic and molecular traits that present therapeutic vulnerability to HGF-targeted therapy in glioblastoma. *Neuro. Oncol.* **2019**, *21*, 222–233.
- 73. Turchi, L.; Sakakini, N.; Saviane, G.; Polo, B.; Saurty-Seerunghen, M. S.; Gabut, M.; Gouillou, C. A.; Guerlais, V.; Pasquier, C.; Vignais, M. L.; et al. CELF2 Sustains a Proliferating/OLIG2+ Glioblastoma Cell Phenotype via the Epigenetic Repression of SOX3. *Cancers (Basel)*. **2023**, *15*, 1–20.
- 74. Scuderi, S. A.; Lanza, M.; Casili, G.; Esposito, F.; Colarossi, C.; Giuffrida, D.; Irene, P.; Cuzzocrea, S.; Esposito, E.; Campolo, M. TBK1 Inhibitor exerts antiproliferative effect on glioblastoma multiforme cells. *Oncol. Res.* **2021**, *28*, 779–790.
- 75. Balogh, J.; Victor, D.; Asham, E. H.; Burroughs, S. G.; Boktour, M.; Saharia, A.; Li, X.; Ghobrial, M.; Monsour, H. Hepatocellular carcinoma: a review. *J. Hepatocell. Carcinoma* **2016**, *Volume* 3, 41–53.
- 76. Tosic, N.; Petrovic, I.; Grujicic, N. K.; Davidovic, S.; Virijevic, M.; Vukovic, N. S.; Pavlovic, S.; Stevanovic, M. Prognostic significance of SOX2, SOX3, SOX11, SOX14 and SOX18 gene expression in adult de novo acute myeloid leukemia. *Leuk. Res.* **2018**, *67*, 32–38.
- 77. Cai, Q. Y.; Liang, G. Y.; Zheng, Y. F.; Tan, Q. Y.; Wang, R. W.; Li, K. Sox3 silencing inhibits metastasis and growth of esophageal squamous cell carcinoma cell via down-regulating GSK-3β. *Int. J. Clin. Exp. Pathol.* **2016**, *9*, 2939–2949.

- 78. Zheng, Y. F.; Li, K.; Cai, Q. Y.; Yang, L.; Tan, Q. Y.; Guo, W.; Wang, R. W. The effect of high Sox3 expression on lymphangiogenesis and lymph node metastasis in esophageal squamous cell carcinoma. *Am. J. Transl. Res.* **2017**, *9*, 2684–2693.
- 79. Güre, A. O.; Stockert, E.; Scanlan, M. J.; Keresztes, R. S.; Jäger, D.; Altorki, N. K.; Old, L. J.; Chen, Y. T. Serological identification of embryonic neural proteins as highly immunogenic tumor antigens in small cell lung cancer. *Proc. Natl. Acad. Sci. U. S. A.* **2000**, *97*, 4198–4203.

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