

Review

Not peer-reviewed version

---

# Advancements in Neurosurgical Intraoperative Histology With Particular Emphasis on Stimulated Raman Scattering (SRS) Microscopy

---

[Ali A. Mohamed](#) \* , Emma Sargent , Cooper Williams , Zev Karve , Karthik Nair , [Brandon Lucke-wold](#) \*

Posted Date: 4 April 2024

doi: [10.20944/preprints202404.0354.v1](https://doi.org/10.20944/preprints202404.0354.v1)

Keywords: deep neural networks; raman histology; digital histopathological assessment, frozen sectioning, cytologic preparations



Preprints.org is a free multidiscipline platform providing preprint service that is dedicated to making early versions of research outputs permanently available and citable. Preprints posted at Preprints.org appear in Web of Science, Crossref, Google Scholar, Scilit, Europe PMC.

Copyright: This is an open access article distributed under the Creative Commons Attribution License which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Disclaimer/Publisher's Note: The statements, opinions, and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions, or products referred to in the content.

Review

# Advancements in Neurosurgical Intraoperative Histology with Particular Emphasis on Stimulated Raman Scattering (SRS) Microscopy

Ali A. Mohamed <sup>1,2</sup>, Emma Sargent <sup>1</sup>, Cooper Williams <sup>1</sup>, Zev Karve <sup>1</sup>, Karthik Nair <sup>1</sup> and Brandon Lucke-wold <sup>3,\*</sup>

<sup>1</sup> Charles E. Schmidt College of Medicine, Florida Atlantic University; amohamed2020@health.fau.edu, esargent2022@health.fau.edu, cooperwillia2023@health.fau.edu, zkarve2023@health.fau.edu, knair2021@health.fau.edu

<sup>2</sup> College of Engineering and Computer Science, Florida Atlantic University; amohamed2020@health.fau.edu

<sup>3</sup> Department of Neurosurgery, University of Florida; brandon.lucke-wold@neurosurgery.ufl.edu

\* Correspondence: brandon.lucke-wold@neurosurgery.ufl.edu

**Abstract:** Despite their relatively low incidence globally, CNS tumors remain amongst the most lethal cancers, with only a few other malignancies surpassing them in 5-year mortality rates. Treatment decisions for brain tumors heavily rely on histopathological analysis, particularly intraoperatively, to guide surgical interventions and optimize patient outcomes. Frozen sectioning has emerged as a vital intraoperative technique, allowing for highly accurate, rapid analysis of tissue samples, although it poses challenges regarding interpretive errors and tissue distortion. Raman histology, based on Raman spectroscopy, has shown great promise in providing label-free, molecular information for accurate intraoperative diagnosis, aiding in tumor resection and identification of neurodegenerative disease. Techniques including Stimulated Raman Scattering (SRS), Coherent Anti-Stokes Raman Scattering (CARS), Surface-Enhanced Raman Scattering (SERS), and Tip-Enhanced Raman Scattering (TERS) have profoundly enhanced the speed and resolution of Raman imaging. Similarly, Confocal Laser Endomicroscopy (CLE) allows for real-time imaging and rapid intraoperative histologic evaluation of specimens. While CLE is primarily utilized in gastrointestinal procedures, its application in neurosurgery is promising, particularly in the context of gliomas and meningiomas. This review focuses on discussing the immense progress in intraoperative histology within neurosurgery and provides insight into the impact of these advancements on enhancing patient outcomes..

**Keywords:** deep neural networks; raman histology; digital histopathological assessment; frozen sectioning; cytologic preparations

## 1. Introduction

Cancers of the brain and central nervous system (CNS) are comprised of a varied group of pathology stemming from brain and spinal cord tissue, with the World Health Organization (WHO) classifying over 150 differing brain tumor subtypes [1]. Intracranial and spinal cord malignancies have remained steadfast amongst ranking within the most fatal cancers, estimating that only esophageal, lung, hepatic and pancreatic tumors stand ahead of CNS tumor pathology in 5-year mortality rates [2]. While CNS cancers are considered rare when contrasted to other classifications of malignancy (with statistics showing that CNS cancer comprised 1.7% of global cancer cases as compared to 12.2% for cancers of the lung), their place amongst the most fatal cancers has been stout for several years [3,4]. While the 5-year survival rates of brain and CNS cancer are enduringly morbid, mild improvements in the frequency of intracranial malignancy cases have been noted in recent

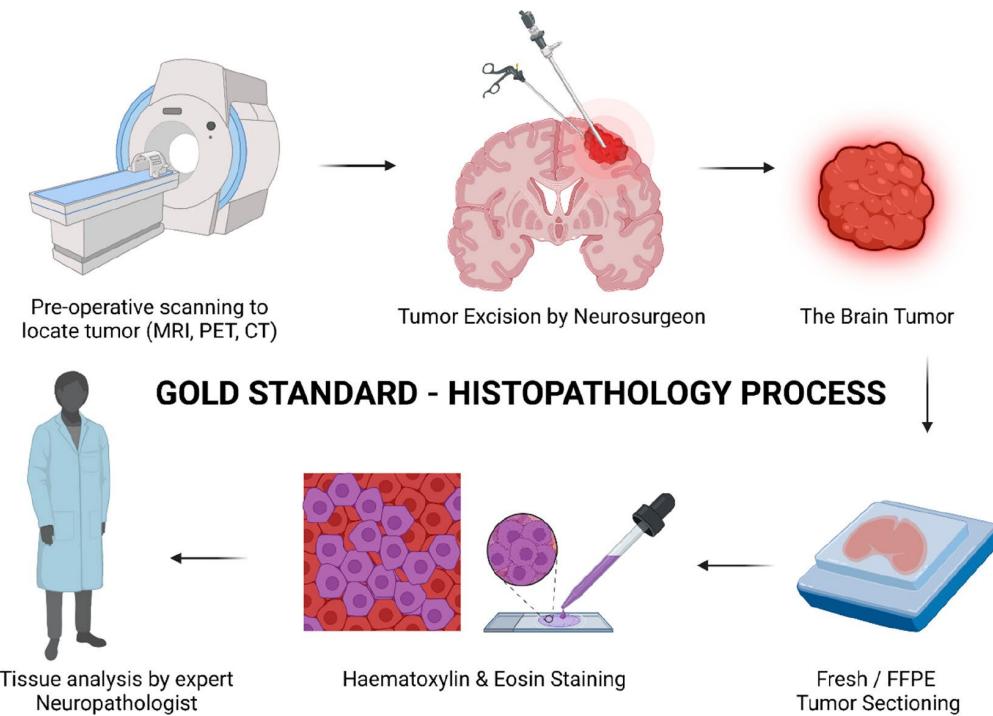
decades, as exemplified by the 0.8% decrease in incidence noted between 2008 and 2017 within North America [5].

The choice of treatment regimen against brain tumors most often anchors on information received from histopathologic analysis of cancerous tissue, an approach that can even be applied intra-operatively to guide for necessity of retrieving additional tissue for biopsy or improvement of margins with regards to tumor resection and overall morbidity and mortality outcomes [6]. Examples of such growing technique modalities include Raman spectroscopy, frozen sectioning, and confocal laser endomicroscopy [7].

Despite being regarded by some as one of the oldest medical specialties in human history (with some historians arguing for the evidence of neurosurgical techniques performed dating back to 10,000 BC), neurosurgery still stands as one of the most dynamic fields within medicine today [8]. Given the wide variety in etiology, clinical presentation, and pathohistological features touted amongst the variety of potential intracranial and CNS tumors, neurosurgery continues to discover new avenues for greater treatment interventions and more efficient intraoperative analyses. Given that the current prevalence of said pathology has continued to remain primarily stagnant (with only marginal reductions in case prevalence since 2008), it is imperative that current clinicians remain vigilant in aiming to appreciate the medley of possible treatment approaches that are practically applicable in the world of neurosurgical techniques. This review specifically aims to discuss advancements in intraoperative histology in the realm of neurosurgery, highlighting the role of these innovations in improving patient outcomes [3,4].

## 2. Frozen Sectioning and Cytologic Preparations

Intraoperative analysis of tissue samples is crucial to allow for a surgeon to determine established treatment protocols and margins in the neurosurgical setting. Proper analysis of tissue requires hardening of the matrix to allow for sectioning. This led to the introduction of frozen sectioning into the field of intraoperative pathology (Figure 1). The first instance of this technique dates to 1905 when Dr. Louis B. Wilson simply used cold winter air to help freeze his samples [9]. Dr. Wilson then progressed to using the CO<sub>2</sub> microtome and staining samples with methylene blue before many other advancements were made to the current state of frozen cytology [10]. The most current form of frozen sectioning includes the utilization of the cryostat set between -20 to -30 degrees Celsius and frozen aerosol sprays to help set the sample [11]. A study by Novis and Zarbo found that 90% of frozen sectioning block turnaround times were within 20 minutes from the time the pathologist received the sample [12]. This fast turnaround time has allowed for the expansion of intraoperative diagnosis of lesions, rather than waiting for the traditional pathology report after the operation has been completed.



**Figure 1.** Schematic representation of the long and tedious histopathology process. The patients need to undergo initial scanning to identify the tumor. This is followed by a sample section which further undergoes multiple processing steps taking longer time and needs to be confirmed by a neuropathologist which increases the chances of error. An error or partial removal of tumor leads to invasive surgery, again affecting the health and quality of life of patient. (Created with Biorender.com). Murugappan S, Tofail SAM, Thorat ND. Raman Spectroscopy: A Tool for Molecular Fingerprinting of Brain Cancer. *ACS Omega*. 2023;8(31). doi:10.1021/acsomega.3c01848. Copyright © 2023 The Authors. *CS Omega* published by American Chemical Society [13].

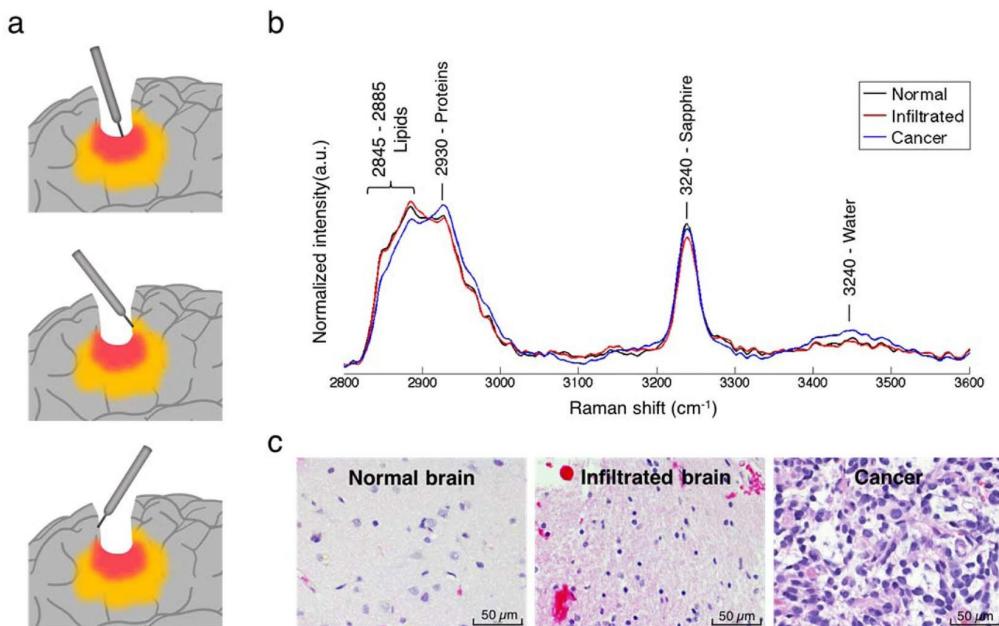
Frozen sectioning has been proven to be a fast intraoperative process, yet many individuals have questioned the accuracy of such procedures. A study performed by Khoddami et al. (2015) found an accuracy rate of 99.5% for the frozen sectioned samples compared to the gold standard of formalin-fixed and paraffin-embedded samples [14]. The same article reported a sensitivity and specificity of 91.4% and 99.7%, respectively. The high level of accuracy associated with frozen sectioning was repeated in a study performed by Kang et al. (2019) which displayed an intraoperative accuracy of 98.8% compared to the permanent histological diagnosis [15]. Another study demonstrated a high correlation of 95.6% between the intraoperative diagnosis and the final diagnosis [16]. These results have demonstrated a high level of accuracy of intraoperative frozen sectioning, but several limitations must be considered while utilizing the technique.

It is an important distinction to make that the high level of accuracy of frozen sectioning does not make it a viable replacement for paraffin-embedded tissue techniques. One reason that it cannot be used as a replacement is the possibility of interpretative errors in frozen sections due to the freezing process distorting the architecture [17]. This interpretative error occurs commonly in differentiating cells that are spindle shaped. This includes differentiating between cerebellopontine angle meningiomas and schwannomas [18]. Also, frozen sectioning provides the possibility that oligodendrogloma is mischaracterized as a high-grade astrocytoma. The perinuclear halo traditionally seen with paraffin-embedded oligodendroglia-based cells is lost in the freezing process [19]. The loss of a perinuclear halo along with freezing artifacts causes oligodendroglia to be angulated similar to astrocyte cells. Errors with frozen sectioning extend beyond these issues listed, but it is a valuable tool that can still provide accurate information. Proper anatomical knowledge, good surgical technique, and expert pathological interpretation of neural tissue can help reduce errors in differentiation between cell lines and structures [20].

Overall, frozen sectioning is a valuable intraoperative tool to help provide guidance for the surgeon on margins, diagnosis, and possible complications. Limitations exist in the usage of frozen sectioning due to the possibility of artifact and tissue distortion created during the freezing process. However, close cooperation between the surgeon and the pathologist can help to limit errors. The pathologist should have the opportunity to look at radiologic images before the procedure to be able to compare histological findings to what is seen in imaging modalities. The combination of radiologic images and histology has been shown to improve accuracy and avoid interpretative errors with frozen sectioning [11]. Continuing, proper selection of tumor for the procedure improves accuracy. Masses firmer in consistency, such as fibroblastic meningioma, showed better yields from frozen cytology than more friable masses [21]. Lastly, usage of intraoperative frozen cytology along with other diagnostic techniques such as crush smear has been shown to improve diagnostic accuracy [22]. In conclusion, proper usage of frozen cytology has been shown to be a valuable tool in the belt of neurosurgeons to provide greater guidance for intraoperative decision-making.

### 3. Raman histology

In recent years, the development of various intraoperative neurohistological analyses has emerged based on advancements in the application of Raman spectroscopy (Figure 2). Raman spectroscopy was first developed in 2008 and is based on the Raman scattering phenomenon, which posits that spontaneous inelastic scattering of light will occur based on the molecular characteristics of the sample [23–25]. Therefore, Raman spectroscopy allows for the rapid, non-destructive acquisition of label-free information directly from the sample's chemical characteristics [23,24]. As most neuropathologies involve alterations at the molecular level, Raman technology has been increasingly efficacious in rapidly diagnosing and treating ischemic or traumatic brain injuries, neurodegenerative diseases, and brain tumors [23]. Several techniques for enhancing Raman spectroscopy have been developed, as spontaneous Raman spectroscopy alone is not strong enough to provide high-resolution images. These methods include but are not limited to stimulated Raman histology (SRH), coherent anti-Stokes Raman scattering (CARS), surface-enhanced Raman scattering (SERS), and tip-enhanced Raman scattering (TERS) [26]. For instance, compared to conventional methods, Stimulated Raman Histology (SRH) microscopy allows for rapid and accurate detection of the extent of brain tumor infiltration through high-resolution imaging [27]. A review of existing literature indicates that as Raman histological techniques continue to develop and improve, they will become an essential element in the neurosurgeon's armamentarium for accurate intraoperative diagnosis and decision-making [28].



**Figure 2.** (a) Schematic depiction of in vivo RS measurements taken in the surgical cavity during glioma resection using the handheld contact probe in dense cancer (red), infiltrated brain (yellow) and surrounding normal brain. (b) In vivo high wavenumber Raman spectra of dense cancer, infiltrated brain and normal brain, averaged over all samples. (c) Representative H&E-stained micrographs for each tissue type. Desroches, J., Jermyn, M., Pinto, M. et al. A new method using Raman spectroscopy for in vivo targeted brain cancer tissue biopsy. *Sci Rep* 8, 1792 (2018). <https://doi.org/10.1038/s41598-018-20233-3>. Copyright© 2018 The Authors. *Scientific Reports* (*Sci Rep*) published by Springer Nature Limited [29].

### 3.1. Indications

Raman technology has very promising indications in oncological neurosurgery, as it allows for the detection of microscopic cancer cell infiltration and therefore more accurate resection of brain tumors and diagnosis of tumor subtypes [30]. Perhaps the primary advantage of Raman spectroscopy is its ability to rapidly produce histological images, allowing the neurosurgeon to achieve empirically clear tumor margins and enhancing the prognosis of neuro-oncologic patients [30]. Raman-based imaging methods can also be used in the detection of neurodegenerative diseases such as Alzheimer Disease (AD). Raman spectroscopy has been shown to greatly improve the accuracy of diagnosing such disorders by detecting the presence of neurofibrillary tangles, amyloid- $\beta$  plaques, and tau protein in tissues [31]. A study by Ryzhikova et al. suggests that RS analysis of CSF may allow for early detection of Alzheimer's, as they were able to diagnose AD with 84% specificity and sensitivity [31,32]. Another study by Lochoki et al. successfully used RA to detect amyloid-beta peptides in tissue samples from AD patients [31,33].

Raman-based imaging techniques may also be effective in analyzing ischemic metabolic changes in stroke and TBI patients. For example, the release of cytochrome C from damaged neurons can be measured by RS to elucidate the extent of ischemic cell death [23].

### 3.2. Techniques

#### 3.2.1. Scattered Raman Spectroscopy

Scattered Raman spectroscopy has promising clinical applications in the detection of healthy versus necrotic or diseased brain tissue as well as the detection of biomarkers that may indicate the stage of the pathology [34]. Raman imaging uses monochromatic light that interacts with the vibrational modes of a sample, causing inelastic scattering of photons. The Raman phenomenon can be analyzed through Stokes or anti-Stokes scattering, where the scattered beams have a lower or higher frequency than that of the incident beam, respectively [34,35]. The resultant peaks represent the Raman spectrum and the energy shift is measured as the "Raman shift", which indicates the various concentrations of specific molecules based on the strength of each spectroscopic peak. This allows each protein, lipid, and DNA molecule to be distinguished through its unique vibrational fingerprint [34]. However, due to the relatively weak signal given by spontaneous Raman spectroscopy, various techniques have been developed to amplify the signal, including Stimulated Raman spectroscopy (SRS), coherent anti-Stokes Raman scattering (CARS) microscopy, surface-enhanced Raman scattering (SERS) microscopy, and tip-enhanced Raman scattering (TERS) microscopy [26,34]. Furthermore, the invention of the femtosecond laser has allowed for the development of other SRS techniques with minimal collateral tissue damage, as its pulse duration is a minuscule  $10^{-15}$  seconds [36].

#### 3.2.2. Stimulated Raman Scattering (SRS) Microscopy

Stimulated Raman scattering (SRS) microscopy utilizes two lasers, the pump beam, and the Stokes beam, to amplify the Raman signal given by specific bonds in the sample and generate image contrast [37]. Inverted microscopes with two lenses are usually used for SRS; the first lens projects beams onto the sample and the second funnels the photons into the detector [25]. The Stokes beam is typically a fixed wavelength while the Pump beam is a tunable wavelength and both pulse at a rate

of 80 MHz. The Stokes and Pump beams must overlap for SRS to occur; this is achieved through the microscope's dichroic mirror and optical delay line [38]. The Stokes beam is then reduced to a fixed frequency of 1-20 MHz using an optic modulator and optic filters to elucidate the SRS signal [25,38]. A large area photodiode detector and optical filters are then used to retrieve the unmodulated beam from the overlapping Stokes and Pump beams, and the signal is finally converted to a histological image through specialized computer software [38].

### 3.2.3. Coherent Anti-Stokes Raman Scattering (CARS)

Coherent anti-Stokes Raman scattering (CARS) microscopy is one method that was developed to overcome the imaging speed limitations of spontaneous Raman scattering [39]. As in SRS, the Pump and Stokes beams concurrently excite the sample such that the frequency difference between the beams corresponds to the vibrational frequency of a specific chemical bond in the sample. However, in addition to stimulated Raman gain ( $\omega_s$ ) and loss ( $\omega_p$ ), CARS produces two much stronger signals: coherent anti-Stokes Raman scattering ( $(\omega_p - \omega_s) + \omega_p$ ) and coherent Stokes Raman scattering ( $\omega_p - (\omega_p - \omega_s)$ ) frequencies [39]. For this reason, CARS is significantly more sensitive than spontaneous Raman spectroscopy and allows for extremely rapid imaging [40]. Furthermore, the nonlinear nature of CARS allows for 3-D sectioning of thick tissues, and tissue damage is minimized as CARS occurs in the ground electric state [40].

### 3.2.4. Surface-Enhanced Raman Scattering (SERS) Microscopy

Another method of enhancing the sensitivity of spontaneous Raman scattering is surface-enhanced Raman scattering (SERS) microscopy [38]. The SERS method was discovered serendipitously when it was observed that a roughened silver electrode significantly increased the Raman signal of an adsorbed pyridine sample [41]. SERS utilizes a metal probe to absorb molecules and concentrate electromagnetic energy through surface plasmons, therefore significantly enhancing the scattering of photons [42,43]. The probe must be composed of a transition metal, a noble metal, or a semiconductor to allow for molecules to be rapidly analyzed on the surface of the metal [43]. The use of such metal probes enhances the incident light and therefore increases Raman scattering, providing a faster and clearer image [38].

### 3.2.5. Tip-Enhanced Raman Scattering (TERS) Microscopy

Tip-enhanced Raman spectroscopy (TERS) operates based on the chemical sensitivity of the SERS method, with the added benefit of high spatial resolution at the sub-nanometer level via scanning probe microscopy (SPM) [44]. Like SRS, CARS, and SERS, TERS is rapid, accurate, and non-destructive. The TERS method involves a sharp metal SPM tip with an enhanced electromagnetic field at the apex due to the resonance of surface plasmons, thereby significantly augmenting the Raman signal of molecules in its vicinity [44]. This increased signal strength allows TERS to overcome the diffraction limit of SRS and SERS and convey improved sub-diffraction limit spatial resolution [45].

## 3.3. Outcomes

Several *ex vivo* studies have found Raman spectroscopy to be highly specific and highly sensitive; Zhang et al. reported the detection of brain tumors by Raman spectroscopy (RS) to be 97% sensitive and 98.5% specific [46,47]. Another *ex vivo* study by Aguiar et al. resulted in 97.4% sensitivity and 100% specificity in distinguishing meningiomas, medulloblastomas, and glioblastomas from health tissue [46,48]. These highly promising *ex vivo* results have laid the foundation for *in vivo* use of Raman spectroscopy. In a study of 13 glioma patients, RS was used to detect invasive cancer cells that would not be detected with T1 or T2 MRI. It was found that RS was able to intraoperatively detect residual grade 2-4 gliomas with 93% sensitivity and 91% specificity [49].

In 2017, Orringer et al. described the first application of intraoperative stimulated Raman scattering (SRS) microscopy and developed the stimulated Raman histology (SRH) interpretation technique, which utilizes SRS images to produce simulated hematoxylin and eosin (H&E) stains [24]. They found that in a sample of 30 patients the intraoperative use of SRH has over 92% accuracy when compared to conventional histological methods in diagnosing brain tumors, and 90% accuracy in elucidating tumor subtypes. In another study on the efficacy of SRH, they created a residual network to predict the presence of tumor cells, non-tumor cells, and low-quality images. Compared to the neuropathologist's analysis of the samples, the SRH-based residual network predicted the presence of tumor cells with 90.2% accuracy in under four minutes [24].

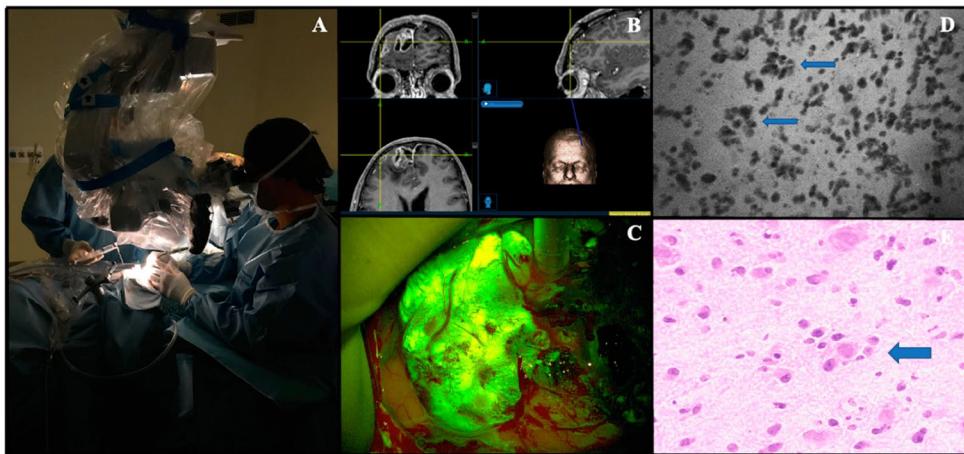
The use of Coherent anti-Stokes Raman scattering (CARS) has also illustrated promising results by significantly decreasing the time required for image collection. In a landmark study by Evans et al., the speed of generating CARS images was found to be 30 images per second, compared to a 30-minute CARS image collection time in 1999 [50]. This reduction in image generation time greatly enhances intraoperative decision-making as well as patient prognoses.

Furthermore, Surface-enhanced Raman scattering (SERS) microscopy has been shown to significantly improve prognosis in post-resection glioma. Han et al. used a SERS scanner intraoperatively until all Raman signals were completely absent in the tissue, and longitudinal MRI analysis indicated that this SERS-guided resection greatly reduced glioma recurrence rate in rat models. SERS has also been used to distinguish acidic margins of gliomas, as extracellular acidosis is a marker for cancer cell infiltration [51]. Jin et al. used a SERS navigation system to locate these acidic margins in animal models as well as in excised tissue from glioma patients and found that the level of acidity correlates with the level of cancer cell proliferation and density. The post-operative survival of animal models was greatly enhanced compared to conventional methods, indicating a promising prognosis for glioma patients [51].

Finally, tip-enhanced Raman scattering (TERS) microscopy achieves both the enhancement of SERS and the high spatial resolution of SPM, thereby overcoming the diffraction limit for more accurate images [52]. As TERS is a relatively new technique, the literature shows limited *in vivo* analysis of SERS in neurosurgery.

#### 4. Confocal Laser Endomicroscopy

Confocal laser endoscopy has grown in popularity for intraoperative, real-time imaging (Figure 3). The use of real-time imaging is especially useful in tumor resections as surgeons can effectively evaluate histoarchitecture and the need for resection of specific pieces of tissue. Using intraoperative imaging reduces the time required for tissue collection and standard biopsy procedures involving in-person pathologists. Instead, surgeons can employ the CLE in the OR and send the live camera feed to a pathologist remotely or in person for analysis. With a pathologist on alert, analysis of histoarchitecture can be done instantly. As such, the surgeon can more effectively execute the resection [53]. CLE has been widely implemented in gastroenterology because of its remarkable accuracy in detecting neoplasia of the colon [54]. However, the technology has yet to reach widespread practice in neurosurgery. The gold standard remains frozen sectioning.



**Figure 3.** Confocal Laser Microscopy *in vivo* Convivo case (Besta Neurological Institute, Milan, Italy). (A). the confocal probe is dressed with its appropriate sterile sheath and used directly upon cerebral surface. (B). Preoperative magnetic resonance with contrast administration images loaded on the neuronavigation system (Stealth S8-Medtronic) of a right frontal parasagittal anaplastic oligodendrogloma, IDH mutant (WHO grade III). (C). Intraoperative view of fluorescein-guided removal of the tumor under YELLOW560 filter activation on Pentero microscope (Carl Zeiss Meditec). (D). Convivo *in vivo* image taken at the center of the tumor showing tumor cells along with typical perineural satellitosis (small arrows), that can be easily found on relative histopathological image as well (H-E, big arrow, (E)). Restelli F, Pollo B, Vetrano IG, Cabras S, Broggi M, Schiariti M, Falco J, de Laurentis C, Raccuia G, Ferroli P, et al. Confocal Laser Microscopy in Neurosurgery: State of the Art of Actual Clinical Applications. *Journal of Clinical Medicine*. 2021; 10(9):2035. <https://doi.org/10.3390/jcm10092035>. Copyright© 2021 The Authors. *Journal of Clinical Medicine* published by MDPI, Basel, Switzerland [55].

#### 4.1. Indications

Because CLE allows surgeons an insight into the histological and cytologic features of tissues, most indications are neoplastic in nature. It allows surgeons to determine if a tissue is dysplastic, neoplastic, or healthy. More specifically, CLE is used to investigate, monitor, and resect tissue samples from meningiomas, gliomas, and pituitary adenomas.

#### 4.2. Techniques

CLE devices are classified as either probe-based or endoscope-based. Either way, the laser is directed until it is in direct contact with the tissue of interest. The laser works by sending light via a straight line into the tissue. Light rays then reflect off the tissue back towards the laser. Reflections are redirected through the same pinhole as the light source (thus the name confocal). The sensor is placed proximally to the pinhole, excluding all the light that is scattered or refocused out of the imaging plane. By doing so, the laser is able to increase the spatial resolution of the image [56,57]. An additional measure to increase resolution has been the advent of a photoactive contrast enhancer. Contrast is commonly fluorescein sodium [58,59].

#### 4.3. Outcomes

Focusing on image quality, brightness, contrast, and resolution are parameters to examine. Brightness and contrast have been shown to be higher for *in vivo* imaging that uses CLE relative to *ex vivo* images that use traditional biopsy methods [58,60]. Another non-clinical measure critical to CLE is the reduction of time spent on tissue analysis. In 2016, Martirosyan et al. conducted one of the first preclinical tests for feasibility. The study showed a decrease in time as the system was only in use for an average of 15.7 minutes; a marked improvement from the typical 20+ minutes required for frozen histology [61].

Feasibility of CLE was further confirmed by determining comparable, if not better, accuracy with decreased time spent in analysis [62–64]. First, CLE was compared to frozen histology in ex vivo studies. In doing so, concordance was found to be 80% for determining correct diagnosis and 93.3% for categorizing patterns at the tumor core [65].

Only a handful of studies have been performed to show the use of CLE in vivo. In the first, intraoperative CLE was shown to correctly diagnose neoplasia while reducing operation time. The device was reported as easy to use due to its similarity to other microsurgical instruments [66].

A later study showed in a 30-patient sample that CLE could be used with nearly identical accuracy as frozen sectioning and permanent histology. The diagnostic accuracy, sensitivity, and specificity reported for CLE vs. frozen section were 94%, 94%, and 100%; for CLE vs. permanent histology 92%, 90%, and 94%, respectively [67].

## 5. Conclusions

The field of neurosurgery faces ongoing challenges in the diagnosis and treatment of CNS malignancies. Intraoperative histology has emerged as a critical tool in guiding surgical decision-making, with techniques such as frozen sectioning, Raman histology, and Confocal Laser Endomicroscopy (CLE) offering invaluable insights into tumor pathology. Frozen sectioning provides rapid intraoperative analysis in tumor resection, and while highly accurate, tissue distortion and interpretive errors remain challenges. Stimulated Raman histology (SRH) offers high-resolution imaging and has demonstrated efficacy in the diagnosis of brain tumors and their subtypes. Surface-enhanced Raman scattering (SERS) microscopy has shown promise in detecting tumor margins and reducing glioma recurrence rates, and tip-enhanced Raman scattering (TERS) microscopy conveys enhanced spatial resolution. Coherent anti-Stokes Raman scattering (CARS) microscopy allows for rapid imaging and improved intraoperative decision-making, and Confocal Laser Endomicroscopy (CLE) enables real-time visualization of histoarchitecture and therefore precise tumor resection. Incorporation of these intraoperative histological techniques into clinical practice holds great potential for improving patient outcomes in neurosurgical oncology. Continued investigation and refinement of these methods are essential to further advance their utility and address existing limitations. By harnessing the power of intraoperative histology, neurosurgeons can strive towards more precise and personalized treatments for patients with CNS malignancies.

**Author Contributions:** Conceptualization, A.M.; methodology, A.M. and B.L.; validation, A.M. and B.L.; investigation, A.M., E.S., C.W., Z.K., K.N.; writing—original draft preparation, A.M., E.S., C.W., Z.K., K.N., B.L.; writing—review and editing, A.M., E.S., C.W., Z.K., K.N., B.L.; visualization, A.M., B.L.; supervision, A.M., B.L.; project administration, A.M., B.L. All authors have read and agreed to the published version of the manuscript.

**Funding:** This research received no external funding.

**Institutional Review Board Statement:** Not applicable.

**Informed Consent Statement:** Not applicable.

**Data Availability Statement:** Not applicable.

**Conflicts of Interest:** The authors declare no conflict of interest.

## References

1. Louis DN, Perry A, Reifenberger G, et al.: The 2016 World Health Organization Classification of Tumors of the Central Nervous System: a summary. *Acta Neuropathol.* 2016, 131: 10.1007/s00401-016-1545-1
2. Mattiuzzi C, Lippi G: Current cancer epidemiology. *J Epidemiol Glob Health.* 2019, 9: 10.2991/jegh.k.191008.001
3. Santucci C, Carioli G, Bertuccio P, et al.: Progress in cancer mortality, incidence, and survival: A global overview. *European Journal of Cancer Prevention.* 2020, 29: 10.1097/CEJ.0000000000000594
4. Ilic I, Ilic M: International patterns and trends in the brain cancer incidence and mortality: An observational study based on the global burden of disease. *Heliyon.* 2023, 9: 10.1016/j.heliyon.2023.e18222

5. Cioffi G, Waite KA, Edelson JL, Kruchko C, Ostrom QT, Barnholtz-Sloan JS: Changes in survival over time for primary brain and other CNS tumors in the United States, 2004–2017. *J Neurooncol.* 2022, 160: 10.1007/s11060-022-04138-w
6. Mannen H: Historical review of development of neuro-histological techniques and three-dimensional reconstruction of individual neurons. In: *Clinical Neurology*. 2006.
7. Orringer DA, Pandian B, Niknafs YS, et al.: Rapid intraoperative histology of unprocessed surgical specimens via fibre-laser-based stimulated Raman scattering microscopy. *Nat Biomed Eng.* 2017, 1: 10.1038/s41551-016-0027
8. Nikova A, Birbilis T: The Basic Steps of Evolution of Brain Surgery. *Maedica (Bucur).* 2017, 12:
9. Gal AA: The centennial anniversary of the frozen section technique at the Mayo Clinic. *Arch Pathol Lab Med.* 2005, 129:1532–5. 10.5858/2005-129-1532-TCAOTF
10. Wheeler TM: ORIGIN AND DEVELOPMENT OF AMERICAN SURGICAL PATHOLOGY. *Trans Am Clin Climatol Assoc.* 2020, 131:326–34.
11. Jaafar H: Intra-operative frozen section consultation: concepts, applications and limitations. *Malays J Med Sci.* 2006, 13:4–12.
12. Novis DA, Zarbo RJ: Interinstitutional comparison of frozen section turnaround time. A College of American Pathologists Q-Probes study of 32868 frozen sections in 700 hospitals. *Arch Pathol Lab Med.* 1997, 121:559–67.
13. Murugappan S, Tofail SAM, Thorat ND: Raman Spectroscopy: A Tool for Molecular Fingerprinting of Brain Cancer. *ACS Omega.* 2023, 8: 10.1021/acsomega.3c01848
14. Khoddami M, Akbarzadeh A, Mordai A, Bidari-Zerehpoush F, Alipour H, Samadzadeh S, Alipour B: Diagnostic accuracy of frozen section of central nervous system lesions: a 10-year study. *Iran J Child Neurol.* 2015, 9:25–30.
15. Kang M, Chung DH, Kim NR, et al.: Intraoperative Frozen Cytology of Central Nervous System Neoplasms: An Ancillary Tool for Frozen Diagnosis. *J Pathol Transl Med.* 2019, 53:104–11. 10.4132/jptm.2018.11.10
16. Uematsu Y, Owai Y, Okita R, Tanaka Y, Itakura T: The usefulness and problem of intraoperative rapid diagnosis in surgical neuropathology. *Brain Tumor Pathol.* 2007, 24:47–52. 10.1007/s10014-007-0219-z
17. Chand P, Amit S, Gupta R, Agarwal A: Errors, limitations, and pitfalls in the diagnosis of central and peripheral nervous system lesions in intraoperative cytology and frozen sections. *J Cytol.* 2016, 33:93–7. 10.4103/0970-9371.182530
18. Plesec TP, Prayson RA: Frozen section discrepancy in the evaluation of central nervous system tumors. *Arch Pathol Lab Med.* 2007, 131:1532–40. 10.5858/2007-131-1532-FSDITE
19. Mitra S, Kumar M, Sharma V, Mukhopadhyay D: Squash preparation: A reliable diagnostic tool in the intraoperative diagnosis of central nervous system tumors. *J Cytol.* 2010, 27:81–5. 10.4103/0970-9371.71870
20. Gandon-Edwards RF, Donald PJ, Boggan JE: Intraoperative frozen section diagnosis in skull base surgery. *Skull Base Surg.* 1993, 3:159–63. 10.1055/s-2008-1060580
21. Savargaonkar P, Farmer PM: Utility of intra-operative consultations for the diagnosis of central nervous system lesions. *Ann Clin Lab Sci.* 2001, 31:133–9.
22. Rao S, Rajkumar A, Ehtesham MD, Duvuru P: Challenges in neurosurgical intraoperative consultation. *Neurol India.* 2009, 57:464–8. 10.4103/0028-3886.55598
23. Terrones O, Olazar-Intxausti J, Anso I, Lorizate M, Nieto-Garai JA, Contreras FX: Raman Spectroscopy as a Tool to Study the Pathophysiology of Brain Diseases. *Int J Mol Sci.* 2023, 24: 10.3390/ijms24032384
24. Orringer DA, Pandian B, Niknafs YS, et al.: Rapid intraoperative histology of unprocessed surgical specimens via fibre-laser-based stimulated Raman scattering microscopy. *Nat Biomed Eng.* 2017, 1:0027. 10.1038/s41551-016-0027
25. Freudiger CW, Min W, Saar BG, et al.: Label-Free Biomedical Imaging with High Sensitivity by Stimulated Raman Scattering Microscopy. *Science (1979).* 2008, 322:1857–61. 10.1126/science.1165758
26. Jones RR, Hooper DC, Zhang L, Wolverton D, Valev VK: Raman Techniques: Fundamentals and Frontiers. *Nanoscale Res Lett.* 2019, 14:231. 10.1186/s11671-019-3039-2
27. Wadiura LI, Kiesel B, Roetzer-Pejrimovsky T, et al.: Toward digital histopathological assessment in surgery for central nervous system tumors using stimulated Raman histology. *Neurosurg Focus.* 2022, 53:E12. 10.3171/2022.9.FOCUS22429
28. Hollon T, Lewis S, Freudiger CW, Sunney Xie X, Orringer DA: Improving the accuracy of brain tumor surgery via Raman-based technology. *Neurosurg Focus.* 2016, 40:E9. 10.3171/2015.12.FOCUS15557
29. Desroches J, Jermyn M, Pinto M, et al.: A new method using Raman spectroscopy for in vivo targeted brain cancer tissue biopsy. *Sci Rep.* 2018, 8: 10.1038/s41598-018-20233-3
30. Hollon T, Orringer DA: Label-free brain tumor imaging using Raman-based methods. *J Neurooncol.* 2021, 151:393–402. 10.1007/s11060-019-03380-z
31. Ranasinghe JC, Wang Z, Huang S: Raman Spectroscopy on Brain Disorders: Transition from Fundamental Research to Clinical Applications. *Biosensors (Basel).* 2022, 13:27. 10.3390/bios13010027

32. Ryzhikova E, Ralbovsky NM, Sikirzhynska V, et al.: Raman spectroscopy and machine learning for biomedical applications: Alzheimer's disease diagnosis based on the analysis of cerebrospinal fluid. *Spectrochim Acta A Mol Biomol Spectrosc.* 2021, 248:119188. 10.1016/j.saa.2020.119188

33. Lochocki B, Morrema THJ, Ariese F, Hoozemans JJM, de Boer JF: The search for a unique Raman signature of amyloid-beta plaques in human brain tissue from Alzheimer's disease patients. *Analyst.* 2020, 145:1724–36. 10.1039/C9AN02087J

34. Ranasinghe JC, Wang Z, Huang S: Raman Spectroscopy on Brain Disorders: Transition from Fundamental Research to Clinical Applications. *Biosensors (Basel).* 2022, 13:27. 10.3390/bios13010027

35. Paraskevaidi M, Martin-Hirsch PL, Martin FL: Progress and Challenges in the Diagnosis of Dementia: A Critical Review. *ACS Chem Neurosci.* 2018, 9:446–61. 10.1021/acschemneuro.8b00007

36. Stern D: Corneal Ablation by Nanosecond, Picosecond, and Femtosecond Lasers at 532 and 625 nm. *Archives of Ophthalmology.* 1989, 107:587. 10.1001/archophth.1989.01070010601038

37. Orillac C, Hollon T, Orringer DA: Clinical Translation of Stimulated Raman Histology. *2022.* 225–36.10.1007/978-1-0716-1803-5\_12

38. Brzozowski K, Matuszyk E, Pieczara A, Firlej J, Nowakowska AM, Baranska M: Stimulated Raman scattering microscopy in chemistry and life science – Development, innovation, perspectives. *Biotechnol Adv.* 2022, 60:108003. 10.1016/j.biotechadv.2022.108003

39. Li S, Li Y, Yi R, Liu L, Qu J: Coherent Anti-Stokes Raman Scattering Microscopy and Its Applications. *Front Phys.* 2020, 8:. 10.3389/fphy.2020.598420

40. Evans CL, Xie XS: Coherent anti-Stokes Raman scattering microscopy: Chemical imaging for biology and medicine. *Annual Review of Analytical Chemistry.* 2008, 1:883–909. 10.1146/annurev.anchem.1.031207.112754

41. Aroca R: Surface-Enhanced Vibrational Spectroscopy. Wiley; 2006. 10.1002/9780470035641

42. Pérez-Jiménez AI, Lyu D, Lu Z, Liu G, Ren B: Surface-enhanced Raman spectroscopy: benefits, trade-offs and future developments. *Chem Sci.* 2020, 11:4563–77. 10.1039/D0SC00809E

43. Han XX, Zhao B, Ozaki Y: Surface-enhanced Raman scattering for protein detection. *Anal Bioanal Chem.* 2009, 394:1719–27. 10.1007/s00216-009-2702-3

44. Kumar N, Mignuzzi S, Su W, Roy D: Tip-enhanced Raman spectroscopy: principles and applications. *EPJ Tech Instrum.* 2015, 2:9. 10.1140/epjti/s40485-015-0019-5

45. Li Z, Persits N, Gray DJ, Ram RJ: Computational polarized Raman microscopy on sub-surface nanostructures with sub-diffraction-limit resolution. *Opt Express.* 2021, 29:38027. 10.1364/oe.443665

46. Doran CE, Frank CB, McGrath S, Packer RA: Use of Handheld Raman Spectroscopy for Intraoperative Differentiation of Normal Brain Tissue From Intracranial Neoplasms in Dogs. *Front Vet Sci.* 2022, 8:. 10.3389/fvets.2021.819200

47. Zhang RR, Kuo JS: Detection of Human Brain Tumor Infiltration With Quantitative Stimulated Raman Scattering Microscopy. *Neurosurgery.* 2016, 78:N9–11. 10.1227/01.neu.0000481982.43612.7b

48. Aguiar RP, Silveira L, Falcão ET, Pacheco MTT, Zângaro RA, Pasqualucci CA: Discriminating Neoplastic and Normal Brain Tissues in Vitro Through Raman Spectroscopy: A Principal Components Analysis Classification Model. *Photomed Laser Surg.* 2013, 31:595–604. 10.1089/pho.2012.3460

49. Jermyn M, Desroches J, Mercier J, St-Arnaud K, Guiot M-C, Leblond F, Petrecca K: Raman spectroscopy detects distant invasive brain cancer cells centimeters beyond MRI capability in humans. *Biomed Opt Express.* 2016, 7:5129. 10.1364/BOE.7.005129

50. Evans CL, Potma EO, Puoris'haag M, Côté D, Lin CP, Xie XS: Chemical imaging of tissue in vivo with video-rate coherent anti-Stokes Raman scattering microscopy. *Proceedings of the National Academy of Sciences.* 2005, 102:16807–12. 10.1073/pnas.0508282102

51. Jin Z, Yue Q, Duan W, et al.: Intelligent SERS Navigation System Guiding Brain Tumor Surgery by Intraoperatively Delineating the Metabolic Acidosis. *Advanced Science.* 2022, 9:. 10.1002/adv.s.202104935

52. Zhang Z, Sheng S, Wang R, Sun M: Tip-Enhanced Raman Spectroscopy. *Anal Chem.* 2016, 88:9328–46. 10.1021/acs.analchem.6b02093

53. Abramov I, Park MT, Gooldy TC, et al.: Real-time intraoperative surgical telepathology using confocal laser endomicroscopy. *Neurosurg Focus.* 2022, 52:E9. 10.3171/2022.3.FOCUS2250

54. Shahid MW, Buchner AM, Coron E, et al.: Diagnostic accuracy of probe-based confocal laser endomicroscopy in detecting residual colorectal neoplasia after EMR: a prospective study. *Gastrointest Endosc.* 2012, 75:525–533.e1. 10.1016/j.gie.2011.08.024

55. Restelli F, Pollo B, Vetrano IG, et al.: Confocal laser microscopy in neurosurgery: State of the art of actual clinical applications. *J Clin Med.* 2021, 10:. 10.3390/jcm10092035

56. Aisenberg J: Gastrointestinal endoscopy nears "the molecular era". *Gastrointest Endosc.* 2008, 68:528–30. 10.1016/j.gie.2008.03.1075

57. Wang TD, Van Dam J: Optical biopsy: A new frontier in endoscopic detection and diagnosis. *Clinical Gastroenterology and Hepatology.* 2004, 2:744–53. 10.1016/S1542-3565(04)00345-3

58. Xu Y, Abramov I, Belykh E, Mignucci-Jiménez G, Park MT, Eschbacher JM, Preul MC: Characterization of ex vivo and in vivo intraoperative neurosurgical confocal laser endomicroscopy imaging. *Front Oncol.* 2022, 12: 10.3389/fonc.2022.979748
59. Restelli F, Mathis AM, Höhne J, Mazzapicchi E, Acerbi F, Pollo B, Quint K: Confocal laser imaging in neurosurgery: A comprehensive review of sodium fluorescein-based CONVIVO preclinical and clinical applications. *Front Oncol.* 2022, 12: 10.3389/fonc.2022.998384
60. Kiesslich R, Neurath MF: Chromoendoscopy and Other Novel Imaging Techniques. *Gastroenterol Clin North Am.* 2006, 35:605–19. 10.1016/j.gtc.2006.07.004
61. Martirosyan NL, Eschbacher JM, Kalani MYS, et al.: Prospective evaluation of the utility of intraoperative confocal laser endomicroscopy in patients with brain neoplasms using fluorescein sodium: experience with 74 cases. *Neurosurg Focus.* 2016, 40:E11. 10.3171/2016.1.FOCUS15559
62. Breuskin D, Szczygielski J, Urbschat S, Kim Y-J, Oertel J: Confocal Laser Endomicroscopy in Neurosurgery—An Alternative to Instantaneous Sections? *World Neurosurg.* 2017, 100:180–5. 10.1016/j.wneu.2016.12.128
63. Charalampaki P, Javed M, Daali S, Heiroth H-J, Igresa A, Weber F: Confocal Laser Endomicroscopy for Real-time Histomorphological Diagnosis. *Neurosurgery.* 2015, 62:171–6. 10.1227/NEU.0000000000000805
64. Pavlov V, Meyronet D, Meyer-Bisch V, et al.: Intraoperative Probe-Based Confocal Laser Endomicroscopy in Surgery and Stereotactic Biopsy of Low-Grade and High-Grade Gliomas. *Neurosurgery.* 2016, 79:604–12. 10.1227/NEU.0000000000001365
65. Acerbi F, Pollo B, De Laurentis C, et al.: Ex Vivo Fluorescein-Assisted Confocal Laser Endomicroscopy (CONVIVO® System) in Patients With Glioblastoma: Results From a Prospective Study. *Front Oncol.* 2020, 10: 10.3389/fonc.2020.606574
66. Höhne J, Schebesch K-M, Zoubaa S, Proescholdt M, Riemenschneider MJ, Schmidt NO: Intraoperative imaging of brain tumors with fluorescein: confocal laser endomicroscopy in neurosurgery. Clinical and user experience. *Neurosurg Focus.* 2021, 50:E19. 10.3171/2020.11.FOCUS20783
67. Abramov I, Park MT, Belykh E, et al.: Intraoperative confocal laser endomicroscopy: prospective in vivo feasibility study of a clinical-grade system for brain tumors. *J Neurosurg.* 2023, 138:587–97. 10.3171/2022.5.JNS2282

**Disclaimer/Publisher's Note:** The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.