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Article

Cytotoxic Potential of the Monoterpene Isoespintanol against Human Tumor Cell Lines

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Abstract: Cancer is a disease that encompasses multiple and different malignant conditions and is among the leading causes of death in the world. Therefore, the search for new pharmacotherapeutic options and potential candidates that can be used as treatments or adjuvants to control this disease is urgent. Natural products, especially those obtained from plants, have played an important role as a source of specialized metabolites with recognized pharmacological properties against cancer, therefore, they are an excellent alternative to be used. The objective of this research was to evaluate the action of the monoterpene isoespintanol (ISO) against the human tumor cell lines MDA-MB-231, A549, DU145, A2780, A2780-cis and the non-tumor line MRC-5. Experiments with MTT and fluorescence with propidium iodide (PI), DAPI and green plasma revealed the cytotoxicity of ISO against these cells; furthermore, morphological and chromogenic studies revealed the action of ISO on cell morphology and the inhibitory capacity on reproductive viability to form colonies in MDA-MB-231 cells. Likewise, 3D experiments validated the damage in these cells caused by this monoterpene. These results serve as a basis for progress in studies of the mechanisms of action of these compounds and the development of derivatives or synthetic analogues with a better antitumor profile.

Keywords: antitumor activity; isoespintanol; monoterpene; cell lines

1. Introduction

Non-communicable diseases (NCDs), also known as chronic diseases, are medical conditions that are associated with long duration and slow progress; currently, they are responsible for the majority of deaths globally. Cancer is one of the four main NCDs with the highest number of deaths [1,2]. In 2020, almost 10 million deaths were attributed to this disease [3,4]. It is a illness that affects any part of the body, presenting typical characteristics such as abnormal cells, with growth and potential to spread to different parts of the body. According to estimates by the World Health Organization (WHO) in 2015, cancer is the first or second cause of death before the age of 70 in 91 out of 172 countries and ranks third or fourth in 22 other countries. The incidence of this disease has increased rapidly in recent years [5,6]. The most common cancers in 2020, in terms of new cases, included: breast (2.26 million cases); lung (2.21 million cases); colorectal (1.93 million cases); prostate (1.41 million cases); skin other than melanoma (1.20 million cases); and gastric (1.09 million cases) [4].

Despite the existence of various methods to treat cancer, there are limitations inherent to each method and also to access to the affected part. Chemotherapy drugs have been used for years, but serious side effects on patients have limited their use [7]. Furthermore, resistance to chemotherapy is a major obstacle to the effective treatment of this disease [8]. In this context, the search and development of novel, chemo-preventive and anti-tumor compounds that act against tumor cells with minimal effect on healthy cells is urgent. Various investigations with natural or synthetic compounds have documented their ability to inhibit the growth of tumor cells both *in vitro* and *in*

vivo; many of these compounds have antioxidant, antiproliferative and proapoptotic effects in several types of cancer [9–12].

Monoterpenes are a group of natural products that have been widely studied for their therapeutic potential against various pathologies [13]. They have been the research subject in the discovery of antineoplastic drugs, since they have a potential antitumor effect and low toxicity. Furthermore, the structural diversity of these compounds provides greater plasticity when interacting with these cells [7,14–16].

Isoespintanol (ISO) [2-isopropyl-3,6-dimethoxy-5-methylphenol] is a mono-terpene first obtained from the aerial parts of *Eupatorium saltense* (Asteraceae) [17], and its synthesis was previously reported [18]. Additionally, it has been extracted from *Oxandra xylopioides* (Annonaceae). Chemical and biological activities such as antioxidant [19], anti-inflammatory [20], antispasmodic [21], vasodilator [22] and cryoprotective of canine semen, has been reported for this compound [23], as well as insecticidal [24] and antifungal activities against *Colletotrichum* [25]. Furthermore, its activity against human pathogenic bacteria and yeasts has also been documented [26–29]. The cytotoxicity of this compound has been studied in the line of immortalized African green monkey kidney epithelial cells (VERO) [28], human peripheral blood lymphocytes [30] and murine macrophages (RAW 264.7) [31]. However, there are no reports on its potential against tumor cells, then we hypothesize that ISO has a cytotoxic effect against them based on the wide range of described activities. The purpose of this research was to evaluate the antitumor activity of ISO against lung (A549), breast (MDA-MB-231), prostate (DU145) and ovarian (A2780 and A2780-cis) tumor cell lines, contributing to the search for new compounds of natural origin that can serve as adjuvants in the treatment of cancer.

2. Results

2.1. MTT Assay

The cytotoxic activity of ISO was evaluated against different human tumor cell lines (MDA-MB-231, A549, DU145, A2780 and A2780-cis) and the non-tumor line (MRC-5) using the MTT assay. From the data obtained, the IC₅₀ values (inhibitory concentration of 50% of cell viability) of the ISO were calculated for all the cell lines under study (Table 1). The ISO showed cytotoxicity against all the cell lines evaluated.

Table 1. IC₅₀ values of ISO on MDA-MB-231, A549, DU145, A2780, A2780-cis and MRC5 monolayers.

| Cell lines | IC ₅₀ ISO (μM) |
|------------|---------------------------|
| MDA-MB-231 | 52.39 ± 3.20 |
| A549 | 59.70 ± 2.74 |
| DU145 | 47.84 ± 3.52 |
| A2780 Cis | 60.35 ± 8.4 |
| A2780 | 42.15 ± 1.39 |
| MRC5 | 39.95 ± 3.76 |

2.2. Cell Morphology Assay

The assay to evaluate the effect of ISO on the morphology of the MDA-MB-231 and MRC-5 cell lines revealed morphological changes in both lines. In cells treated at concentrations of 0.5IC₅₀ (26.2 μM and 19.97 μM, for MDA-MB-231 and MRC-5 respectively), no significant changes were observed, while at concentrations of IC₅₀ (52.4 and 39.95 μM respectively) and 2IC₅₀ (104.8 and 79.9 μM, respectively) after 24 h of treatment, important morphological changes were observed such as nuclear and cytoplasmic condensation, decrease in cell confluency and cell fragmentation surrounded by membranes (smaller cells); these changes were observed in individual cells, which is characteristic of death by apoptosis [32]. The tests with propidium iodide (PI) showed cells stained red, evidencing

cell death, which grew as the IC_{50} values increased, so we could propose that ISO induces cell death by apoptosis in both cell types (Figures 1 and 2).

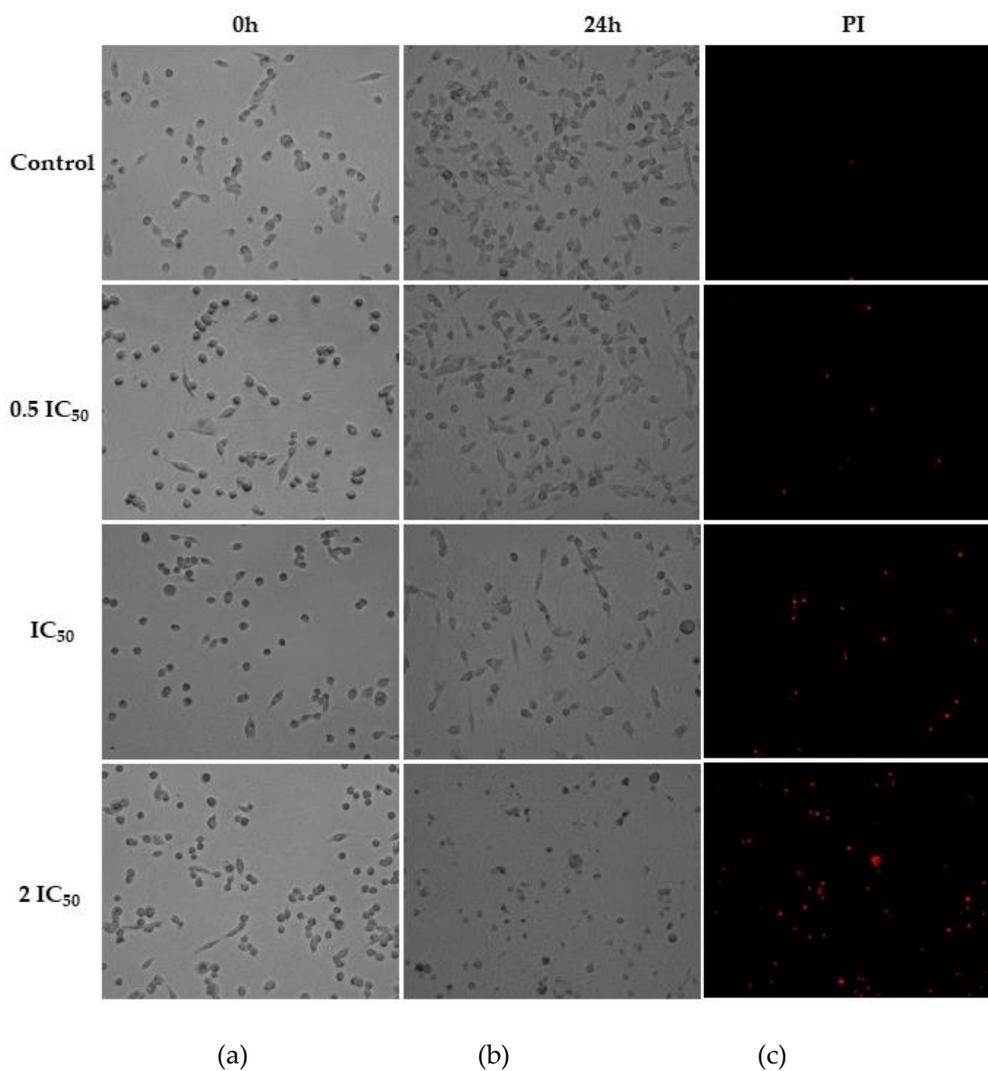


Figure 1. Bright field microscopy (a and b) and fluorescence with propidium iodide (c) of morphological study in breast tumor cells (MDA-MB-231) after treatment with ISO ($0.5IC_{50}$, IC_{50} and $2IC_{50}$) at 0 and 24 hours. Significant morphological changes are observed with the IC_{50} and $2IC_{50}$ treatments at 24 hours. The cells were photographed with the CELENA® S Digital Imaging System (Logos biosystems).

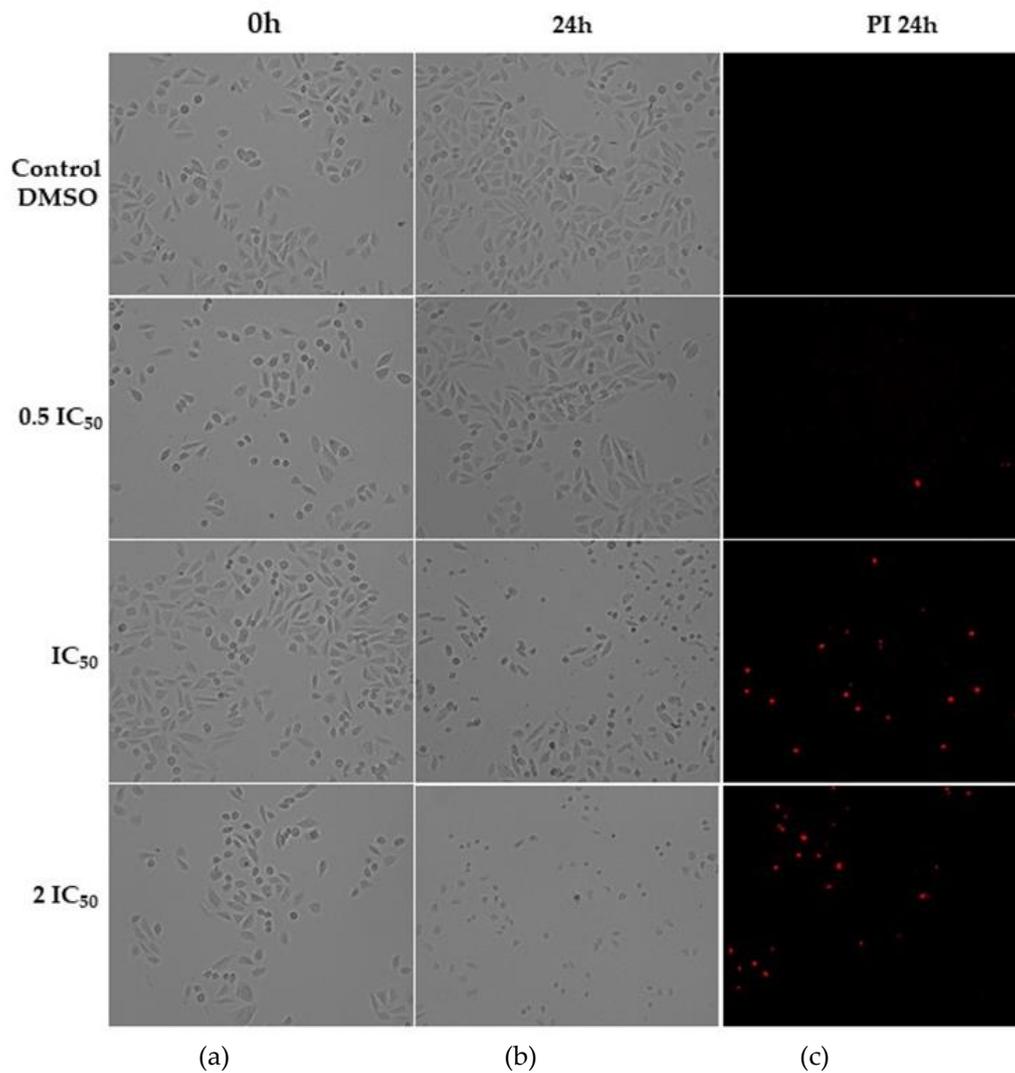


Figure 2. Bright field microscopy (a and b) and fluorescence with propidium iodide (c) of morphological study in non-tumor lung cells (MRC-5) after treatment with ISO (0.5IC₅₀, IC₅₀ and 2IC₅₀) at 0 and 24 h. Morphological changes are observed with IC₅₀ and 2IC₅₀ treatments at 24 hours. The cells were photographed with the CELENA® S Digital Imaging System (Logos biosystems).

The morphological changes of the MDA-MB-231 cells treated with ISO were also evidenced with fluorescence studies using green plasma and DAPI, as seen in Figure 3. The increase in the ISO concentration (IC₅₀ and 2IC₅₀) caused a greater effect on the morphology of these cells.

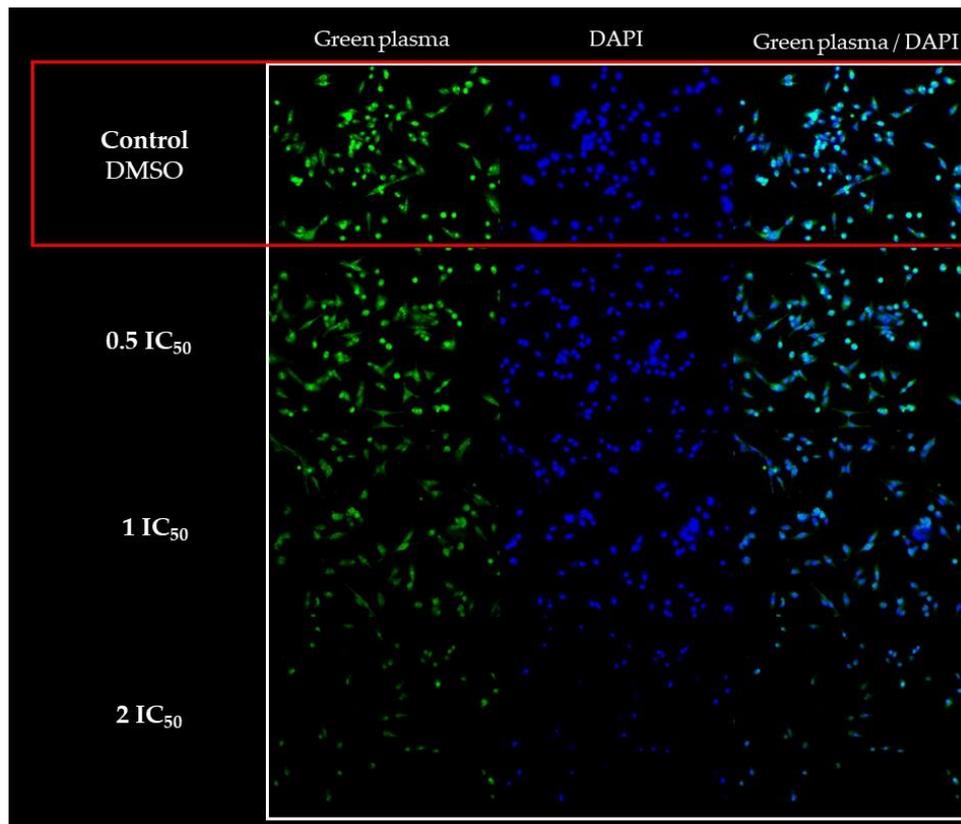


Figure 3. Morphological changes in breast tumor cells (MDA-MB-231) after treatment with ISO ($0.5IC_{50}$, IC_{50} and $2IC_{50}$) for 24 h. Double fluorescent staining (green plasma/DAPI). The cells were photographed with the CELENA® S Digital Imaging System (Logos biosystems).

2.3. Clonogenicity Assay

MDA-MB-231 breast tumor cells and the non-tumor lung cell line MRC-5 were treated with ISO ($0.5IC_{50}$, IC_{50} , $2IC_{50}$) for 48 h, to evaluate their ability to survive, reproduce and form colonies after the treatment with this monoterpene. The cells were evaluated after a period of 10 days. The number of colonies of MDA-MB-231 cells treated with ISO decreased significantly. In contrast, the number of colonies in MRC-5 cells treated with the ISO only decreased significantly at the concentration of $2IC_{50}$ (Figure 4). The antiproliferative effect of ISO was higher in MDA-MB-231 cells compared to MRC-5 cells, demonstrating a dose-dependent effect.

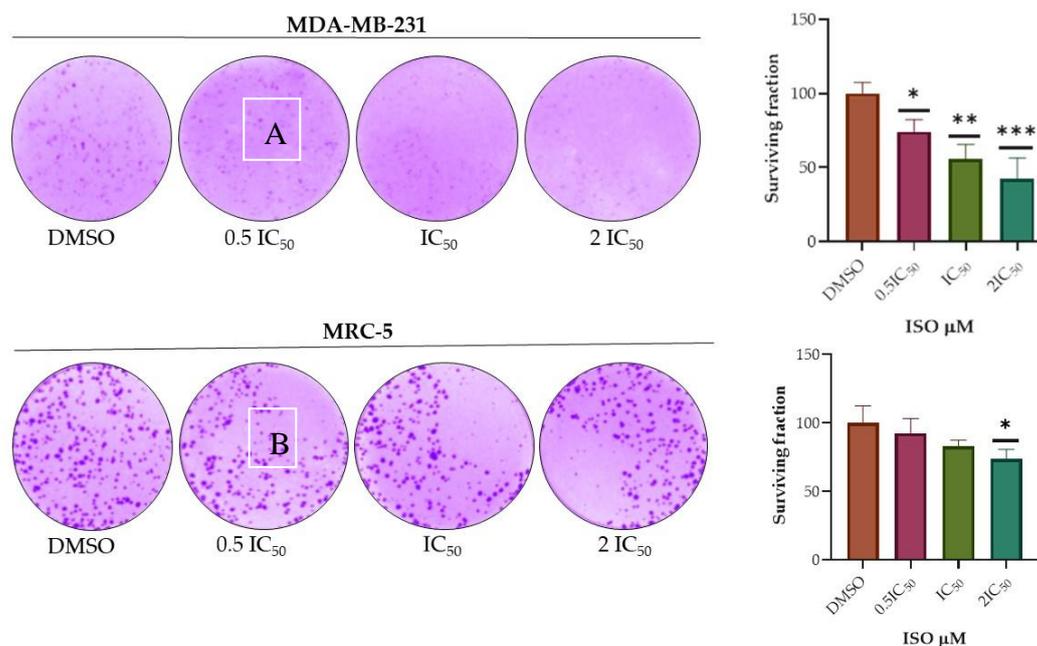


Figure 4. Colony formation by the breast tumor cell line MDA-MB-231 and the non-tumor lung cell line MRC-5 after plating and treatment with ISO ($0.5IC_{50}$, IC_{50} , $2IC_{50}$) after 10 days of incubation. The decrease in the number of colonies in MDA-MB-231 cells is significantly greater compared to MRC-5 cells.

2.4.3. D Assay

The 3D cell culture model offers advantages that are not possible in the traditional 2D model, bringing it closer to the *in vivo* model. In 3D culture, cells are organized in a three-dimensional way that favors cell-cell and cell-matrix interactions. The 3D tumor cell environment features a differentiated proliferation rate, oxygen gradient, nutrient gradient, acidic pH, the presence of apoptotic and necrotic cell death, regulation of gene expression and drug resistance. While in the 2D model the cells are forced to adapt to a rigid surface, the distribution of nutrients and oxygen occurs uniformly, there is less cell-cell interaction and dead cells are removed [32-36].

The experiment tested the ISO at five different concentrations: 0.5, 1, 2, 4, and 8 times the IC_{50} value determined in the 2D experiment against the MDA-MB-231 cell line. The spheroid cultured in the standard medium served as the control. The spheroid control maintained its cell density and spherical shape throughout the analyzed periods. In contrast, the spheroid exposed to the ISO exhibited cell detachment after 8 days. Notably, $2IC_{50}$, $4IC_{50}$, and $8IC_{50}$ concentrations highlighted the expected trend, with spheroids displaying greater resistance than monolayer cells. The most significant impact was observed at higher concentrations. After 10 days, the spheroid dissolved, accompanied by a noticeable decrease in cell density at $4IC_{50}$ and $8IC_{50}$ concentrations, indicating the compound's efficacy, as shown in Figure 5. This observation was further validated by images using DAPI and PI markers, confirming the compound's action. Only live cells were observed in the control group after 10 days, with DAPI marking the cells. However, treatment with the compound at various concentrations resulted in cell death, as indicated by a PI marker (Figure 6).

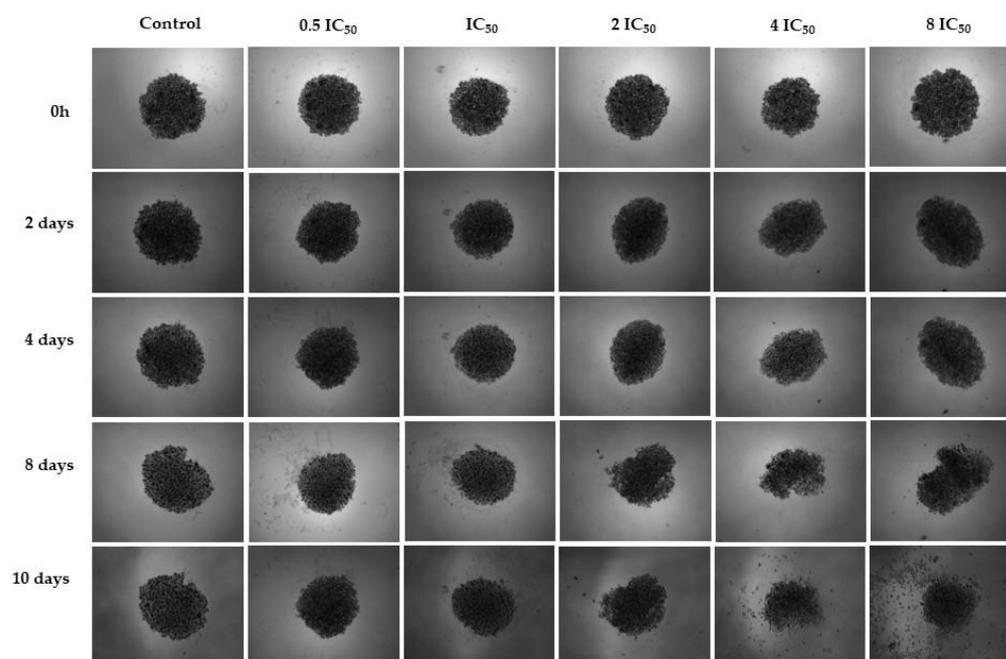


Figure 5. Bright field microscopy of MDA-MB-231 cell spheroids treated with ISO ($2IC_{50}$, $4IC_{50}$ and $8IC_{50}$) and without treatment. Cell detachment was evident after 8 days of treatment with ISO.

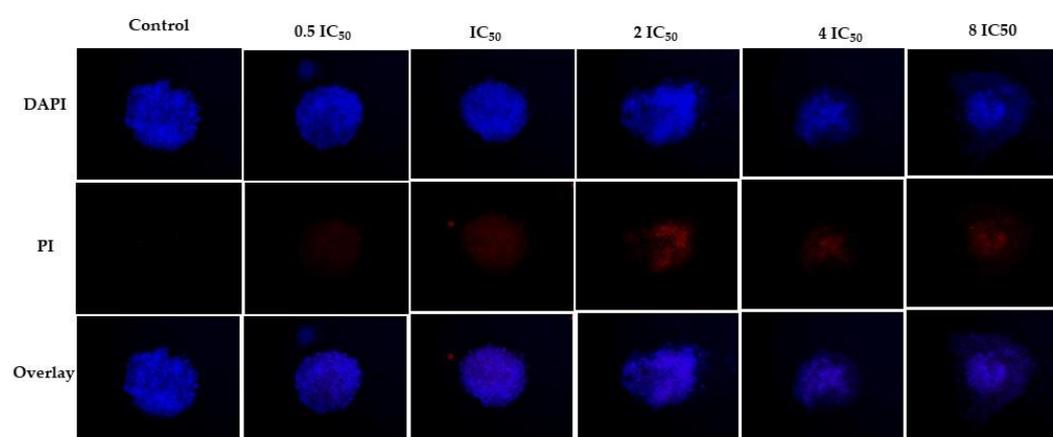


Figure 6. Fluorescence microscopy of MDA-MB-231 cell spheroids, labeled with DAPI and PI, treated with ISO ($2IC_{50}$, $4IC_{50}$ and $8IC_{50}$) and without treatment. Cell detachment was evident after 8 days of treatment with ISO. PI staining shows cell death at all concentrations evaluated.

3. Discussion

The great cellular heterogeneity in the tumor microenvironment and the astonishing ability of cancer cells to mutate, and thus evade almost any therapeutic intervention, supports the lethality of the disease and has proven to be immune to many decades of efforts to overcome it [37]. Cancer drugs account for approximately a quarter of total new drug discovery and approvals each year. Although some of the treatments used help some patients, the great challenges faced in cancer make other therapeutic strategies necessary to reduce the burden of the disease [38]. Therefore, the search and development of new compounds with anti-tumor potential, that are effective and safe, as well as the development of new treatment strategies with better patient tolerance, is urgent today. Since ancient times, natural products have made an important contribution to pharmacotherapy, especially in cancer and infectious diseases. In this scenario, plants play a primary role as a source of specialized

metabolites with recognized medicinal properties [39,40]. Due to their wide chemical diversity, these metabolites can be used directly as bioactive compounds, as drug prototypes, or used as pharmacological tools for different targets [41].

In this research, we demonstrate that the monoterpene ISO extracted from *O. xylopioides* inhibits the development of the tumor cell lines MDA-MB-231, A549, DU145, A2780-cis, A2780 and the non-tumor line MRC-5, with IC_{50} values of 52.39 μ M, 59.70 μ M, 47.84 μ M, 60.35 μ M, 42.15 μ M and 39.95 μ M, respectively. Monoterpenes have been well documented for their anti-tumor activity and chemopreventive activity against various types of cancer. Among the mechanisms of action that may explain the antitumor activity of these compounds is the induction of phase II carcinogen-metabolizing enzymes, resulting in carcinogenic detoxification. The tumor suppressive chemopreventive activity of these compounds may be due to the induction of apoptosis and/or the inhibition of post-translational isoprenylation of cell growth regulatory proteins and transforming growth factor 1. Therefore, monoterpenes seem to act through multiple mechanisms in cancer chemoprevention and chemotherapy [42].

ISO caused changes in the morphology of MDA-MB-231 cells (Figures 1–3), such as compaction of the cell nucleus and cytoplasm, and cell fragments surrounded by membrane, which is indicative of cell death by apoptosis. These results are consistent with studies reported with citral, which can eliminate drug-resistant breast cancer cells (MDA-MB-231) in a spheroid model, by inducing apoptosis [14] and inhibit the growth of MCF-7 breast cells, with an arrest of the cell cycle in the G2/M phase and induction of apoptosis; besides, a decrease in the synthesis of prostaglandin E2 was observed, which supports a possible chemopreventive effect of this compound [40].

Other monoterpenes, such as limonene, have been documented with chemopreventive activity, stimulating the detoxification of carcinogenic compounds and limiting tumor growth and angiogenesis in various cancer models. The anticancer activity of limonene was related to the inhibition of tumor initiation, growth and angiogenesis and the induction of cancer cell apoptosis [43,44]. The activity of geraniol has also been documented against cancer of the prostate, intestine, liver, kidney and skin, due to the induction of apoptosis and the increase in the expression of proapoptotic proteins; furthermore, it has been indicated that the synergy of this compound with other drugs can further increase the range of chemotherapeutic agents [45]. The monoterpenoids nerol, geranial and neral have been reported with antineoplastic activity in animal and cellular models in various types of cancer; these compounds have been found to activate multiple antitumor responses, such as apoptosis, autophagic cell death, cytostasis, and necrosis. These multitarget mechanisms may provide superior therapeutic effects, reducing the adaptive resistance of tumor cells [46].

We also evaluated the ability of MDA-MB-231 and MRC-5 cells to survive and reproduce to form colonies after ISO treatment. The results show the capacity of ISO to inhibit colony formation in both cell types at concentrations of IC_{50} and $2IC_{50}$, with the effect observed on the MDA-MB-231 tumor cell line being significantly greater, compared to the effect observed on the non-tumor cell line MRC-5. These results are consistent with those documented with other monoterpenes, for which antiproliferative capacity has been demonstrated. Thymoquinone has shown its effectiveness in inhibiting different stages of cancer, such as proliferation, migration and invasion. In addition, it acts as an anticancer agent against different human cancers, by inducing apoptosis and regulating levels of pro- and anti-apoptotic genes, and inhibition of metastasis [47]. Furthermore, monoterpenes such as iridoid glycosides also exert inhibitory effects on numerous cancers; these compounds inhibit cancer growth by inducing cell cycle arrest or by regulating signaling pathways related to apoptosis. In addition, they suppress the expression and activity of matrix metalloproteinases (MMP), which reduces the migration and invasiveness of cancer cells. The antiangiogenic mechanism of iridoid glycosides was found to be closely related to the transcriptional regulation of proangiogenic factors, i.e., vascular endothelial growth factors (VEGF) and cluster of differentiation 31 (CD31). In this way, these compounds can alleviate or prevent the rapid progression of cancer and metastasis [48].

Other terpenes and their derivatives have demonstrated significant anti-cancer potential. The sesquiterpenes β -caryophyllene (BCP) and β -caryophyllene oxide (BCPO) have important anticancer

activities, affecting the growth and proliferation of numerous cancer cells, properties that have been related to their chemical structure [49]. Thus, BCPO contains exocyclic methylene and epoxide functional groups, which is why it binds covalently to proteins and DNA bases through sulfhydryl and amino groups. For this reason, BCPO reveals a high potential to be a signaling modulator in cancerous tumor cells. The anticancer activities of both sesquiterpenes can be exerted by suppressing cell growth and inducing apoptosis [50]. 2-cyano-3,12-dioxooleana-1,9(11)-dien-28-oic acid (CDDO), and its C28 modified derivative methyl-ester (CDDO-Me, also known as bardoxolone methyl), are two synthetic derivatives of oleanolic acid (natural triterpene). These molecules have been widely investigated for their great capacity to exert antiproliferative, antiangiogenic and antimetastatic activities, and to induce apoptosis and differentiation in cancer cells. At doses greater than 100 nM, CDDO and CDDO-Me are capable of modulating the differentiation of a variety of cell types, both tumor cell lines and primary culture cells, while at micromolar doses these compounds exert an anticancer effect of multiple ways; by inducing extrinsic or intrinsic apoptotic pathways, or autophagic cell death, by inhibiting telomerase activity, altering mitochondrial functions through inhibition of the Lon protease and blocking the deubiquitylating enzyme USP7 [51]. We also validated the effectiveness of ISO against MDA-MB-231 cells through 3D experiments, using images that used DAPI and PI markers, being the first report of this type of assay in the ratification of cell damage by this compound.

The results of this research highlight the effect of ISO against the human tumor cell lines MDA-MB-231, A549, DU145, A2780, A2780-cis and the non-tumor line MRC-5; suggesting as possible mechanisms of action on MDA-MB-231 cells the induction of apoptosis and inhibition of cell proliferation. Additional studies are necessary to delve into the mechanisms of antitumor action of this monoterpene as a possible adjuvant in the treatment of this disease.

4. Materials and Methods

4.1. Obtaining and Identification of Isoespinanol

ISO was obtained as a crystalline amorphous solid from the petroleum benzene extract of *O. xylopioides* leaves, and its structural identification was performed by GC-MS, ¹H-NMR, ¹³C-NMR, DEPT, COSY ¹H-¹H, HMQC, and HMBC. Information related to obtaining and identifying the ISO was reported in our previous study [29].

4.2. Cell Lines and Reagents

The cell lines, MDA-MB-231 (triple negative human breast adenocarcinoma of mesenchymal phenotype, ATCC No. HTB-26); A549 (alveolar basal cell epithelial adenocarcinoma of the human lung, ATCC No. CCL-185); DU145 (prostate tumor, ATCC No. HTB-81); A2780-cis; (cisplatin-resistant human ovarian tumor, ECACC No. 93112517); A2780 (human ovarian tumor, ECACC No. 93112519) and MRC-5 (human non-lung tumor, ATCC No. CCL-171) were obtained from the American Type Culture Collection (ATCC) (Rockville, Maryland, United States). Dimethylsulfoxide (DMSO), trypan blue and (3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide) [MTT], were obtained from Sigma-Aldrich®, St. Louis, Missouri, United States; crystal violet (CV) was obtained from Synth, Diadema, Sao Paulo, Brazil; propidium iodide was obtained from BD Biosciences, San Jose, California, United States; trypsin and fetal bovine serum (FBS) were obtained from Vitrocell, Campinas, Sao Paulo, Brazil; phosphate-buffered saline (PBS) was prepared with sodium chloride, potassium chloride, disodium phosphate and monopotassium phosphate obtained from Synth, Diadema, Sao Paulo, Brazil; Dulbecco's Modified Eagle culture medium (DMEM) was prepared with DMEM (D5523), gentamicin and amphotericin B obtained from Sigma-Aldrich®, St. Louis, Missouri, United States, penicillin obtained from Vitrocell, Campinas, Sao Paulo, Brazil, and sodium bicarbonate obtained from Neon, Suzano, Sao Paulo, Brazil; RPMI medium was prepared with RPMI-1640 (R6504), gentamicin and amphotericin B obtained from Sigma-Aldrich®, St. Louis, Missouri, United States, and penicillin obtained from Vitrocell, Campinas, Sao Paulo, Brazil; green plasma,

CellMask™ and DAPI (4',6-Diamidino-2-phenylindole dilactate) were obtained from Invitrogen by Thermo Fisher Scientific, Eugene, Oregon, United States.

4.3. Cell Cultures

The A2780-cis and A2780 cell lines, was cultured in RPMI 1640 medium supplemented with 10% fetal bovine serum (FBS) and 1% penicillin/streptomycin and the MDA-MB-231, A549, DU145 and MRC-5 lines were cultured in DMEM medium and were maintained in a humidified atmosphere with 5% CO₂ at 37 °C. Medium changes were performed every 2 to 3 days for maintenance. Subcultures were carried out twice a week until reaching approximately 80% confluency (using an inverted microscope, Nikon Eclipse TS 100) for the assays.

4.4. MTT Assay

The cytotoxic activity of ISO against human tumor cell lines was evaluated by the colorimetric MTT assay, which is based on the ability of dehydrogenase enzymes of metabolically viable cells to reduce tetrazolium rings and form formazan crystals; consequently, the number of viable cells is directly proportional to the level of formazan produced [52,53]. To do this, a density of 1.5×10^4 cells/well in 150 μ L of RPMI 1640 medium (for A2780 and A2780-cis) and DMEM (for MDA-MB-231, A549, DU145 and MRC-5) were seeded in 96-well microplates (Kasvi), allowing their adhesion and proliferation during 24 h of incubation at 37 °C with 5% CO₂. After this time, 0.75 μ L of the ISO stock solution dissolved in DMSO at different concentrations (4.88 to 625 μ M) was added to the final reaction wells, the plates were incubated for 48 h at 37 °C. After incubation, 50 μ L of MTT (1 mg/mL) was added to each well. The plates were incubated with 5% CO₂ at 37 °C for 4 h. Subsequently, the MTT was discarded and the boxes were allowed to dry at room temperature, then the formazan crystals deposited at the bottom of each well were dissolved in 100 μ L of isopropanol. Absorbance was determined at an optical density (OD) of 540 nm using an Epoch 2 microplate reader (Biotek). Cells with DMSO were used as controls. The experiments were performed three times in triplicate. The results of the tests were expressed by dose-response curves using 8 concentrations of ISO previously described. IC₅₀ values (50% inhibitory concentration of the cell population) were calculated from the fit ($R^2 > 0.95$) of the Hill slope curve of the experimental data using nonlinear regression analysis in GraphPad Prism version 8.0 software.

4.5. Cell Morphology Assay

The effect of ISO on the morphology of the MDA-MB-231 cell line was evaluated. For the assay, 1500 μ L of a cell suspension was added to 12-well dishes (1×10^5 cells/well) and incubated for 24 h at 37 °C and 5% CO₂. Then, the cells were treated with 7.5 μ L of the ISO (0.5IC₅₀, IC₅₀, 2IC₅₀) and incubated for 48 h at 37 °C with 5% CO₂. Subsequently, the cells were observed and photographed at 0 h and 24 h. Then, the medium was removed from the cells and they were fixed with 1 mL of methanol for 10 min. Next, the methanol was removed and 500 μ L of PBS and 200 μ L of green plasma were added to the wells, the plates were incubated for 30 min. Subsequently, the green plasma was removed and 500 μ L of PBS and 200 μ L of DAPI were added, the plates were incubated again for 5 min. Staining with propidium iodide (PI) [100 μ L] for 10 min was also performed. Then, the cells were photographed with the help of the digital image capture system CELENA® S *Digital Imaging System* (Logos biosystems). The morphology of cells treated with ISO was compared with the morphology of cells treated with 0.5% DMSO, used as a negative control. The experiments were carried out in triplicate.

4.6. Clonogenicity Assay

To evaluate the reproductive viability of MDA-MB-231 cells after treatment with ISO, a clonogenicity test [49] was performed. The criterion used was the number of colonies. 1500 μ L of the cell suspension were seeded in 6-well plates (1000 cells/well) and incubated at 37 °C with 5% CO₂ for 24 h. Then, the cells were treated with 10 μ L of ISO (0.5IC₅₀, IC₅₀, 2IC₅₀), and incubated for 48 h.

Subsequently, the medium was discarded from the dishes, the cells were washed with 2 ml of PBS and 2 mL of DMEN culture medium supplemented with 10% FBS were added, the dishes were incubated (5% CO₂ at 37 °C) for 10 days. Finally, the supernatant was discarded and the cells were washed with PBS and fixed with a solution of methanol (2 ml) and acetic acid (3:1 (v/v)) for 5 min. Colonies were stained with crystal violet (0.05% crystal violet, 1% formaldehyde, 1X PBS and 1% methanol). Colony counting was performed with ImageJ image analysis software [54] and Prisma software version 8.0 was used. To determine the plating efficiency [PE] (which allows knowing the number of colonies formed based on the cells that were seeded) and the survival fraction [SF] (which allows knowing the number of colonies developed after applying the treatment with ISO) [55], to calculate the PE and the SF, the formulas were used [56]:

1. $PE = \frac{N_0 \text{ of colonies formed}}{N_0 \text{ of seeded cells}} \times 100\%$
2. $SF = \frac{N_0 \text{ of colonies formed after treatment}}{N_0 \text{ of seeded cells}} \times PE$

4.7. D Assay

For the 3D cell culture assay, the 96-well Bioprint Kit from Magnetic 3D Cell Culture Technology (m3D-Greiner) was used. First, 150 µL of a solution containing nanoparticles (NanoShuttle - PL) was added to a cell culture flask containing the MDA-MB-231 cell line. After 24 hours, the nanoparticle cells were plated in a 96-well repellent culture plate (3000 cells/well). The culture plate was placed under a magnetic drive to form spheroids. The plate was kept in an incubator (37°C 5% CO₂), and the formation and growth of the spheroids were monitored using a CELENA® S *Digital Imaging System* microscope (Logos Biosystems). After 4 days, different concentrations of the compound were added to the spheroids, and the experiment was monitored for 10 days, with 50% of the culture media being replaced every 48 hours during the treatment period. The fluorescent markers DAPI and PI were added on the last day of treatment.

4.8. Data Analysis

Results were analyzed using GraphPad Prism version 8.0 software and ImageJ image analysis software.

5. Conclusions

This research evaluated the activity of ISO against five human tumor cell lines and one non-tumor line. The results show that ISO has a cytotoxic effect against these cells lines studied, affecting cell morphology and interrupting their reproductive viability. Thus, ISO becomes a promising compound of great interest to continue investigating the mechanisms of action against tumor cells and thus contribute to the search for alternatives that help in the control of challenging diseases such as cancer.

Author Contributions: Conceptualization, O.I.C.M., F.V.R., and A.A.O.; methodology, O.I.C.M., F.V.R., D.B.F., K.Z., T.T., J.S.M., and A.A.O.; formal analysis, O.I.C.M., F.V.R., K.Z., T.T., J.S.M., D.B.F., and A.A.O.; investigation, O.I.C.M., F.V.R., D.B.F., K.Z., T.T., and A.A.O.; resources, O.I.C.M., F.V.R., A.A.O. and G.S.P.; writing—original draft preparation, O.I.C.M., F.V.R., and A.A.O.; writing—review and editing, O.I.C.M., G.S.P., F.V.R., K.Z., D.B.F., T.T., J.S.M., and A.A.O.; visualization, O.I.C.M., F.V.R., J.S.M., and A.A.O.; supervision, O.I.C.M., F.V.R., G.S.P., and A.A.O.; funding acquisition, O.I.C.M., F.V.R., A.A.O. and G.S.P. All authors have read and agreed to the published version of the manuscript.

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References

1. Budreviciute, A.; Damiani, S.; Sabir, D. K.; Onder, K.; Schuller-Goetzburg, P.; Plakys, G.; Katileviciute, A.; Khoja, S.; Kodzius, R. Management and prevention strategies for non-communicable diseases (NCDs) and their risk factors. *Front. Public Health*. **2020**, *8*, 1–11. <https://doi.org/10.3389/fpubh.2020.574111>.
2. Siegel, R. L.; Miller, K. D.; Jemal, A. Cancer statistics, 2017. *CA. Cancer J. Clin.* **2017**, *67* (1), 7–30. <https://doi.org/10.3322/caac.21387>.
3. Ferlay, J.; Colombet, M.; Soerjomataram, I.; Parkin, D. M.; Piñeros, M.; Znaor, A.; Bray, F. Cancer statistics for the year 2020: An overview. *Int. J. Cancer* **2021**, *149* (4), 778–789. <https://doi.org/10.1002/ijc.33588>.
4. Sung, H.; Ferlay, J.; Siegel, R. L.; Laversanne, M.; Soerjomataram, I.; Jemal, A.; Bray, F. Global cancer statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA. Cancer J. Clin.* **2021**, *71* (3), 209–249. <https://doi.org/10.3322/caac.21660>.
5. Bray, F.; Ferlay, J.; Soerjomataram, I.; Siegel, R. L.; Torre, L. A.; Jemal, A. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA. Cancer J. Clin.* **2018**, *68* (6), 394–424. <https://doi.org/10.3322/caac.21492>.
6. Wang, J. J.; Lei, K. F.; Han, F. Tumor microenvironment: Recent advances in various cancer treatments. *Eur. Rev. Med. Pharmacol. Sci.* **2018**, *22* (12), 3855–3864. <https://doi.org/10.26355/eurrev-201806-15270>.
7. Silva, B. I. M.; Nascimento, E. A.; Silva, C. J.; Silva, T. G.; Aguiar, J. S. Anticancer activity of monoterpenes: A systematic review. *Mol. Biol. Rep.* **2021**, *48* (7), 5775–5785. <https://doi.org/10.1007/s11033-021-06578-5>.
8. Shrestha, S.; Song, Y. W.; Kim, H.; Lee, D. S.; Cho, S. K. Sageone, a diterpene from *Rosmarinus officinalis*, synergizes with cisplatin cytotoxicity in SNU-1 human gastric cancer cells. *Phytomedicine* **2016**, *23* (13), 1671–1679. <https://doi.org/10.1016/j.phymed.2016.09.008>.
9. Wang, H.; Shu, L.; Su, Z.; Fuentes, F.; Lee, J.-H.; Kong, A.-N. T. Plants against cancer: A review on natural phytochemicals in preventing and treating cancers and their druggability. *Anticancer. Agents Med. Chem.* **2012**, *12* (10), 1281–1305. <https://doi.org/10.2174/187152012803833026>.
10. Shamshoum, H.; Vlavcheski, F.; Tsiani, E. Anticancer effects of oleuropein. *BioFactors* **2017**, *43* (4), 517–528. <https://doi.org/10.1002/biof.1366>.
11. Hajdú, Z.; Hohmann, J.; Forgo, P.; Máthé, I.; Molnár, J.; Zupkó, I. Antiproliferative activity of *Artemisia asiatica* extract and its constituents on human tumor cell lines. *Planta Med.* **2014**, *80* (18), 1692–1697. <https://doi.org/10.1055/s-0034-1383146>.
12. Chen, W.; Liu, Y.; Li, M.; Mao, J.; Zhang, L.; Huang, R.; Jin, X.; Ye, L. Anti-tumor effect of α -pinene on human hepatoma cell lines through inducing G2/M cell cycle arrest. *J. Pharmacol. Sci.* **2015**, *127* (3), 332–338. <https://doi.org/10.1016/j.jphs.2015.01.008>.
13. Machado, T. Q.; Da Fonseca, A. C. C.; Duarte, A. B. S.; Robbs, B. K.; De Sousa, D. P. A Narrative review of the antitumor activity of monoterpenes from essential oils: An update. *Biomed Res. Int.* **2022**, *2022*. <https://doi.org/10.1155/2022/6317201>.
14. Nigjeh, S. E.; Yeap, S. K.; Nordin, N.; Kamalideghan, B.; Ky, H.; Rosli, R. Citral induced apoptosis in MDA-MB-231 spheroid cells. *BMC Complement. Altern. Med.* **2018**, *18* (1), 1–10. <https://doi.org/10.1186/s12906-018-2115-y>.
15. Zhang, Y.; Liu, J.; Chen, M.; Fan, Y.; Li, M.; Wang, A.; Liu, G.; Xu, Y.; Ren, X.; Xiao, Y. Isolation of a new monoterpenoid glycoside from anhua dark tea based on an NMR-guided method and its cytotoxic activity against MDA-MB-231 and SH-SY5Y cell lines. *Nat. Prod. Res.* **2020**, *36* (8), 2015–2020. <https://doi.org/10.1080/14786419.2020.1839465>.
16. Vandresen, F.; Falziroli, H.; Almeida Batista, S. A.; Da Silva-Giardini, A. P. B.; De Oliveira, D. N.; Catharino, R. R.; Ruiz, A. L. T. G.; De Carvalho, J. E.; Foglio, M. A.; Da Silva, C. C. Novel R-(+)-limonene-based thiosemicarbazones and their antitumor activity against human tumor cell lines. *Eur. J. Med. Chem.* **2014**, *79*, 110–116. <https://doi.org/10.1016/j.ejmech.2014.03.086>.
17. Morales, I.; De La Fuente, J.; Sosa, V. Componentes de *Eupatorium saltense*. *An Asoc Quim Argent* **1991**, *79* (3), 141–144.
18. Hocquemiller, R.; Cortes, D.; Arango, G. J.; Myint, S. H.; Cave, A. Isolement et synthèse de L'espintanol, nouveau monoterpène antiparasitaire. *J. Nat. Prod.* **1991**, *54*, 445–452. <https://doi.org/10.1021/np50074a015>.
19. Rojano, B.; Sáez, J.; Schinella, G.; Quijano, J.; Vélez, E.; Gil, A.; Notario, R. Experimental and theoretical determination of the antioxidant properties of isoespintanol (2-Isopropyl-3,6-Dimethoxy-5-Methylphenol). *J. Mol. Struct.* **2008**, *877*, 1–6. <https://doi.org/10.1016/j.molstruc.2007.07.010>.
20. Rojano, B.; Pérez, E.; Figadère, B.; Martin, M. T.; Recio, M. C.; Giner, R.; Ríos, J. L.; Schinella, G.; Sáez, J. Constituents of *Oxandra Cf. xylopioides* with anti-inflammatory activity. *J. Nat. Prod.* **2007**, *70* (5), 835–838. <https://doi.org/10.1021/np060333v>.

21. Gavilánez Buñay, T. C.; Colareda, G. A.; Ragone, M. I.; Bonilla, M.; Rojano, B. A.; Schinella, G. R.; Consolini, A. E. Intestinal, urinary and uterine antispasmodic effects of isoespintanol, metabolite from *Oxandra xylopioides* leaves. *Phytomedicine* **2018**, *51*, 20–28. <https://doi.org/10.1016/j.phymed.2018.06.001>.
22. Rinaldi, G. J.; Rojano, B.; Schinella, G.; Mosca, S. M. Participation of NO in the vasodilatory action of isoespintanol. *Vitae* **2019**, *26*, 78–83. <https://doi.org/10.17533/udea.vitae.v26n2a03>.
23. Usuga, A.; Tejera, I.; Gómez, J.; Restrepo, O.; Rojano, B.; Restrepo, G. Cryoprotective effects of ergothioneine and isoespintanol on canine semen. *Animals* **2021**, *11*, 2757. <https://doi.org/10.3390/ani11102757>.
24. Rojano, B. A.; Montoya, S.; Yépez, F.; Sáez, J. Evaluación de isoespintanol aislado de *Oxandra Cf. xylopioides* (Annonaceae) sobre *Spodoptera frugiperda* J.E. Smith (Lepidoptera: Noctuidae); 2007; *60*, 3691–3702.
25. Arango, N.; Vanegas, N.; Sáez, J.; García, C.; Rojano, B. Actividad antifúngica del isoespintanol sobre hongos del género *Colletotricum*. *Sci. Tech.* **2007**, *33*, 279–280. <https://doi.org/10.22517/23447214.6055>.
26. Contreras Martínez, O. I.; Angulo Ortiz, A.; Santafé Patiño, G. Antibacterial screening of isoespintanol, an aromatic monoterpene isolated from *Oxandra xylopioides* Diels. *Molecules* **2022**, *27* (22). <https://doi.org/10.3390/molecules27228004>.
27. Contreras Martínez, O. I.; Angulo Ortiz, A.; Santafé Patiño, G. Antifungal potential of isoespintanol extracted from *Oxandra xylopioides* Diels (Annonaceae) against intrahospital isolations of *Candida* spp. *Heliyon* **2022**, *8* (10). <https://doi.org/10.1016/j.heliyon.2022.e11110>.
28. Contreras Martínez, O. I.; Angulo Ortiz, A.; Santafé Patiño, G.; Peñata-Taborda, A.; Berrio Soto, R. Isoespintanol antifungal activity involves mitochondrial dysfunction, inhibition of biofilm formation, and damage to cell wall integrity in *Candida tropicalis*. *Int. J. Mol. Sci.* **2023**, *24* (12), 10187. <https://doi.org/10.3390/ijms241210187>.
29. Contreras Martínez, O. I.; Angulo Ortiz, A.; Santafé Patiño, G. Mechanism of antifungal action of monoterpene isoespintanol against clinical isolates of *Candida tropicalis*. *Molecules* **2022**, *27*, 5808. <https://doi.org/10.3390/molecules27185808>.
30. Marquez-Fernandez, M.; Munoz-Lasso, D.; Bautista Lopez, J.; Zapata, K.; Puertas Mejia, M.; Lopez-Alarcon, C.; Rojano, B. A. Effect of isoespintanol isolated from *Oxandra Cf. xylopioides* against DNA damage of human lymphocytes. *Pak. J. Pharm. Sci* **2018**, *31*, 1777–1782.
31. Zapata, K.; Arias, J.; Cortés, F.; Alarcon, C.; Durango, D.; Rojano, B. Oxidative stabilization of palm olein with isoespintanol (2-Isopropyl-3,6-Dimethoxy-5-Methylphenol) isolated from *Oxandra Cf. xylopioides*. *J. Med. Plants Res.* **2017**, *11*, 218–225. <https://doi.org/10.5897/JMPR2016.6270>
32. Jaganathan, H.; Gage, J.; Leonard, F.; Srinivasan, S.; Souza, G. R.; Dave, B.; Godin, B. Three-dimensional *in vitro* co-culture model of breast tumor using magnetic levitation. *Sci. Rep.* **2014**, *4*, 1–9. <https://doi.org/10.1038/srep06468>.
33. Brajša, K.; Trzun, M.; Zlatar, I.; Jelić, D. Three-dimensional cell cultures as a new tool in drug discovery. *Period. Biol.* **2016**, *118* (1), 59–65. <https://doi.org/10.18054/pb.2016.118.1.3940>.
34. Yamada, K. M.; Cukierman, E. Modeling tissue morphogenesis and cancer in 3D. *Cell* **2007**, *130* (4), 601–610. <https://doi.org/10.1016/j.cell.2007.08.006>.
35. Timm, D. M.; Chen, J.; Sing, D.; Gage, J. A.; Haisler, W. L.; Neeley, S. K.; Raphael, R. M.; Dehghani, M.; Rosenblatt, K. P.; Killian, T. C.; Tseng, H.; Souza, G. R. A High-throughput three-dimensional cell migration assay for toxicity screening with mobile device-based macroscopic image analysis. *Sci. Rep.* **2013**, *3*, 1–8. <https://doi.org/10.1038/srep03000>.
36. Huang, Z.; Yu, P.; Tang, J. Characterization of triple-negative breast cancer MDA-MB-231 cell spheroid model. *Onco. Targets. Ther.* **2020**, *13*, 5395–5405. <https://doi.org/10.2147/OTT.S249756>.
37. Liao, Z.; Tan, Z. W.; Zhu, P.; Tan, N. S. Cancer-associated fibroblasts in tumor microenvironment – accomplices in tumor malignancy. *Cell. Immunol.* **2018**, *343*, 0–1. <https://doi.org/10.1016/j.cellimm.2017.12.003>.
38. Mullard, A. Addressing cancer's grand challenges. *Nat. Rev. Drug Discov.* **2020**, *19* (12), 825–826. <https://doi.org/10.1038/d41573-020-00202-0>.
39. Aylate, A.; Agize, M.; Ekeru, D.; Kiros, A.; Ayledo, G.; Gendiche, K. *In-vitro* and *in-vivo* antibacterial activities of *Croton macrostachyus* methanol extract against *E. coli* and *S. aureus*. *Adv. Anim. Vet. Sci.* **2017**, *5*, 107–114. <https://doi.org/10.14737/journal.aavs/2017/5.3.107.114>.
40. Chaouki, W.; Leger, D. Y.; Liagre, B.; Beneytout, J. L.; Hmamouchi, M. Citral inhibits cell proliferation and induces apoptosis and cell cycle arrest in MCF-7 cells. *Fundam. Clin. Pharmacol.* **2009**, *23* (5), 549–556. <https://doi.org/10.1111/j.1472-8206.2009.00738.x>.
41. Avato, P. Editorial to the special issue – “Natural products and drug discovery”. *Molecules* **2020**, *25*, 1128. <https://doi.org/10.3390/molecules25051128>.
42. Crowell, P. L. Prevention and therapy of cancer by dietary monoterpenes. *J. Nutr.* **1999**, *129*, 775–778. <https://doi.org/10.1093/jn/129.3.775s>.
43. Araújo-Filho, H. G. de; dos Santos, J. F.; Carvalho, M. T. B.; Picot, L.; Fruitier-Arnaudin, I.; Groult, H.; Quintans-Júnior, L. J.; Quintans, J. S. S. Anticancer activity of limonene: A systematic review of target signaling pathways. *Phyther. Res.* **2021**, *35* (9), 4957–4970. <https://doi.org/10.1002/ptr.7125>.

44. Anandakumar, P.; Kamaraj, S.; Vanitha, M. K. D-limonene: A multifunctional compound with potent therapeutic effects. *J. Food Biochem.* **2021**, *45* (1), 1–10. <https://doi.org/10.1111/jfbc.13566>.
45. Mączka, W.; Wińska, K.; Grabarczyk, M. One hundred faces of geraniol. *Molecules.* **2020**, *25*, 3303. doi:10.3390/molecules25143303.
46. Silva, G. dos S. E.; Marques, J. N. de J.; Linhares, E. P. M.; Bonora, C. M.; Costa, É. T.; Saraiva, M. F. Review of anticancer activity of monoterpenoids: Geraniol, nerol, geranial and neral. *Chem. Biol. Interact.* **2022**, *362*. <https://doi.org/10.1016/j.cbi.2022.109994>.
47. Imran, M.; Rauf, A.; Khan, I. A.; Shahbaz, M.; Qaisrani, T. B.; Fatmawati, S.; Abu-Izneid, T.; Imran, A.; Rahman, K. U.; Gondal, T. A. Thymoquinone: A novel strategy to combat cancer: A review. *Biomed. Pharmacother.* **2018**, *106*, 390–402. <https://doi.org/10.1016/j.biopha.2018.06.159>.
48. Kim, C. W.; Choi, K. C. Potential roles of iridoid glycosides and their underlying mechanisms against diverse cancer growth and metastasis: Do they have an inhibitory effect on cancer progression? *Nutrients* **2021**, *13* (9). <https://doi.org/10.3390/nu13092974>.
49. Fidy, K.; Fiedorowicz, A.; Strzdała, L.; Szumny, A. β -caryophyllene and β -caryophyllene oxide—natural compounds of anticancer and analgesic properties. *Cancer Med.* **2016**, *5* (10), 3007–3017. <https://doi.org/10.1002/cam4.816>.
50. Park, K. R.; Nam, D.; Yun, H. M.; Lee, S. G.; Jang, H. J.; Sethi, G.; Cho, S. K.; Ahn, K. S. β -Caryophyllene oxide inhibits growth and induces apoptosis through the suppression of PI3K/AKT/MTOR/S6K1 pathways and ROS-mediated MAPKs activation. *Cancer Lett.* **2011**, *312* (2), 178–188. <https://doi.org/10.1016/j.canlet.2011.08.001>.
51. Borella, R.; Forti, L.; Gibellini, L.; De Gaetano, A.; De Biasi, S.; Nasi, M.; Cossarizza, A.; Pinti, M. Synthesis and anticancer activity of CDDO and CDDO-Me, two derivatives of natural triterpenoids. *Molecules* **2019**, *24* (22), 1–20. <https://doi.org/10.3390/molecules24224097>.
52. Mosmann, T. Rapid colorimetric assay for cellular growth and survival: Application to proliferation and cytotoxicity assays. *J. Immunol.* **1983**, *65*, 55–63.
53. Motlhatlego, K.; Ali, M.; Leonard, C.; Eloff, J.; McGaw, L. Inhibitory effect of newtonia extracts and myricetin-3-O-rhamnoside (myricitrin) on bacterial biofilm formation. *BMC Complement. Med. Ther.* **2020**, *20*, 358. <https://doi.org/10.1186/s12906-020-03139-4>.
54. Guzmán, C.; Bagga, M.; Kaur, A.; Westermarck, J.; Abankwa, D. Colony Area: An imageJ plugin to automatically quantify colony formation in clonogenic assays. *PLoS One* **2014**, *9* (3), 14–17. <https://doi.org/10.1371/journal.pone.0092444>.
55. Franken, N. A. P.; Rodermond, H. M.; Stap, J.; Haveman, J.; Van Bree, C. Clonogenic assay of cells *in vitro*. *Nat. Protoc.* **2006**, *1* (5), 2315–2319. <https://doi.org/10.1038/nprot.2006.339>.
56. Buch, K.; Peters, T.; Nawroth, T.; Sängler, M.; Schmidberger, H.; Langguth, P. Determination of cell survival after irradiation via clonogenic assay versus multiple MTT assay - a comparative study. *Radiat. Oncol.* **2012**, *7* (1), 1–6. <https://doi.org/10.1186/1748-717X-7-1>.

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