

Review

Not peer-reviewed version

# m6A Methylation in Regulation of Antiviral Innate Immunity

---

Ivan Karandashov , [Anastasiya Kostyusheva](#) , [Artyom Kachanov](#) , [Maria Dukich](#) , Natalia Ponomareva , [Sergey Brezgin](#) , [Alexander Lukashev](#) , Vadim S Pokrovsky , [Vladimir Chulanov](#) , [Dmitry Kostyushev](#) \*

Posted Date: 14 March 2024

doi: [10.20944/preprints202403.0853.v1](https://doi.org/10.20944/preprints202403.0853.v1)

Keywords: TLR; RLR; herpesviruses; hepatitis viruses; coronaviruses; retroviruses; flaviviruses; adenoviruses



Preprints.org is a free multidiscipline platform providing preprint service that is dedicated to making early versions of research outputs permanently available and citable. Preprints posted at Preprints.org appear in Web of Science, Crossref, Google Scholar, Scilit, Europe PMC.

Copyright: This is an open access article distributed under the Creative Commons Attribution License which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Disclaimer/Publisher's Note: The statements, opinions, and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions, or products referred to in the content.

Review

# m6A Methylation in Regulation of Antiviral Innate Immunity

Ivan Karandashov <sup>1</sup>, Anastasiya Kostyusheva <sup>1</sup>, Artyom Kachanov <sup>1</sup>, Maria Dukich <sup>1,2</sup>, Natalia Ponomareva <sup>1,3,4</sup>, Sergey Brezgin <sup>1,5</sup>, Alexander Lukashev <sup>1</sup>, Vadim S. Pokrovsky <sup>6,7,8</sup>, Vladimir Chulanov <sup>9</sup> and Dmitry Kostyushev <sup>1,5,10,\*</sup>

<sup>1</sup> Laboratory of Genetic Technologies, Martsinovsky Institute of Medical Parasitology, Tropical and Vector-Borne Diseases, First Moscow State Medical University (Sechenov University), Moscow, Russia

<sup>2</sup> Faculty of Virology, Lomonosov Moscow State University, Moscow, Russia

<sup>3</sup> Division of Biotechnology, Sirius University of Science and Technology, Sochi, Russia

<sup>4</sup> Department of Pharmaceutical and Toxicological Chemistry, Sechenov First Moscow State Medical University, Moscow, Russia

<sup>5</sup> Division of Biotechnology, Sirius University of Science and Technology, Sochi, Russia

<sup>6</sup> Shemyakin-Ovchinnikov Institute of Bioorganic Chemistry, Russian Academy of Sciences, Moscow, Russia

<sup>7</sup> Blokhin National Medical Research Center of Oncology, Moscow, Russia

<sup>8</sup> People's Friendship University, Moscow, Russia

<sup>9</sup> Department of Infectious Diseases, First Moscow State Medical University (Sechenov University), Moscow, Russia

<sup>10</sup> Faculty of Bioengineering and Biotechnologies, Lomonosov Moscow State University, Moscow, Russia

\* Correspondence: dkostushev@gmail.com

**Abstract:** The epitranscriptomic modification m6A is a prevalent RNA modification that plays a crucial role in the regulation of various aspects of RNA metabolism. It has been found to be involved in a wide range of physiological processes and disease states. Of particular interest is the role of m6A machinery and modifications in viral infections, serving as an evolutionary marker for distinguishing between self and non-self entities. In this review article, we present a comprehensive overview of the epitranscriptomic modification m6A and its implications in the interplay between viruses and their host, focusing on immune responses and viral replication. We outline future research directions that highlight the role of m6A in viral nucleic acid recognition, initiation of antiviral immune responses, and modulation of antiviral signaling pathways. Additionally, we discuss the potential of m6A as a prognostic biomarker and a target for therapeutic interventions in viral infections.

**Keywords:** TLR; RLR; herpesviruses; hepatitis viruses; coronaviruses; retroviruses; flaviviruses; adenoviruses

## 1. Introduction

In 1974, Desrosiers, Friderici, and Rottman described a new modification of RNA, N6-methyladenosine (m6A), in Novikov's hepatoma cells [1]. m6A is a post-transcriptional RNA modification involving the addition of an extra methyl group to nitrogen 6 of adenosine. Currently, more than 150 types of RNA modifications have been identified [2], of which m6A is the most prevalent [3].

The m6A modification has been observed in almost all types of RNA, including messenger RNA (mRNA) [4], ribosomal RNA (rRNA) [5], and microRNA [6]. m6A typically occurs at the consensus motif RRm6ACH ([G/A/U][G/A]m6AC[U/A/C]). Ke *et al.* demonstrated that 93% of m6A modifications in partially spliced chromatin-associated RNA were found within exonic regions although intronic sequences are three-fold more abundant [7]. In addition, Dominissi *et al.* showed that 87% of m6A were located in exons longer than 400 nucleotides. Most m6A markers (37%) were found in coding sequences; 28% were localized in the 400-nucleotide window centered on stop



codons; 20% were located in the 3' untranslated region (3'-UTR); and 12% in transcription start sites (TSS). The relative enrichment of m6A bases was highest in stop codon region and TSS [8].

The m6A modification plays a role in almost all biochemical processes related to RNA metabolism, influencing RNA stability [9] and regulating nuclear export of mRNA [10], splicing [11], and translation [12].

Viral RNA can also undergo methylation [13], and m6A influences various processes that regulate the viral cycle [14]. Thanks to m6A markers, cells can recognize its RNA as “self” and protect it from the innate immune system, whereas genetic material of viruses can frequently be recognized foreign because it does not bear m6A-modified nucleotides [15]. However, viruses have learned to evade cellular immunity by utilizing m6A modifications [16].

In this manuscript, we provide a focused review of the role of m6A in regulating innate immunity and shaping antiviral defenses. We consolidate recent results demonstrating the role of m6A methylation in viral immune evasion and antiviral immune signaling.

## 2. Regulation of m6A Modification

The m6A modification is dynamically reversible by three groups of enzymes: methyltransferases (writers), demethylases (erasers), and proteins that recognize m6A-modified RNA (readers). The reversibility of m6A methylation is controlled by writers and erasers, while reader proteins recognize the modified adenosines and regulate the associated biological functions.

So far, four genes encoding methyltransferases have been identified in the human genome - methyltransferase-like (METTL) 3 (METTL3), METTL5, METTL16, and zinc finger CCHC-type containing 4 (ZCCHC4). In addition to these methyltransferases, other proteins participating in m6A catalysis perform collaborative functions, including METTL14, Wilms tumor 1-associating protein (WTAP), Vir-like m6A methyltransferase associated (VIRMA), Cbl proto-oncogene like 1 (HAKAI), zinc finger CCCH-type containing 13 (ZC3H13), and RNA-binding motif 15/15B (RBM15/15B). In most cases, m6A methylation is catalyzed by the methyltransferase complex (MTC) consisting of METTL3, METTL14, WTAP, RBM15/15B, ZC3H13, VIRMA, and HAKAI, which is typically localized in the nucleus, except for in certain cancer cell lines [17,18]. METTL3 directly adds m6A modifications onto RNA, and METTL14 stabilizes the conformation of METTL3's catalytic center. METTL14 is also crucial for substrate recognition [19]. In addition, Liu *et al.* have also identified that both METTL14 and METTL3 individually exhibit methyltransferase activity; however, the activity of the METTL3-METTL14 complex is markedly higher [20]. The remaining proteins of this complex perform various specific functions: WTAP targets the METTL3-METTL14 complex to nuclear speckles [17,21], and RBM15/15B binds the m6A complex and interacts with specific U-rich sites in XIST RNA. Data show that knockdown of both RBM15 and RBM15B results in impaired XIST-mediated gene silencing, suggesting that RBM15 and RBM15B are necessary for MTC recruitment to XIST RNA. Additionally, WTAP is essential for facilitating the interaction between RBM15/RBM15B and the methylation complex [22]. ZC3H13 regulates MTC's localization to the nucleus, as knocking down ZC3H13 results in cytoplasmic localization of METTL3, METTL14, VIRMA, WTAP, and HAKAI, the latter of which bridges WTAP to the mRNA-binding factor NITO [23]. VIRMA also plays a crucial role in preferential 3'-UTR m6A modification, but has no impact on long exon preferential methylation [24]. Studies on *Drosophila melanogaster* have shown that HAKAI is crucial for MTC stabilization, as its knockout results in decreased methylation activity. We were unable to find studies regarding the role of HAKAI in mammals, but HAKAI is considered to be conserved between humans and *Drosophila* [25].

Methylation of 18S and 28S rRNA, as well as small nuclear RNA (snRNA) U6, is carried out by the heterodimeric methyltransferase complex METTL5 and the stabilizing coactivator protein TRMT112 [26]. m6A modification of 26S rRNA is regulated by methyltransferase ZCCHC4. ZCCHC4 is localized in nucleoli, where it interacts with RNA-binding proteins involved in ribosome biogenesis and RNA metabolism [27]. The methyltransferase METTL16 is distributed to both the nucleus and the cytoplasm [28,29], and interacts with snRNA U6, rRNA, and pre-mRNA [30,31]. Notably, METTL16 can regulate the expression of MAT2A, which is directly responsible for the synthesis of S-adenosylmethionine [32].

Protein erasers act as demethylases, removing m6A modifications from RNA. Currently, only two demethylases from the FeII/α-KG-dependent dioxygenase AlkB family associated with m6A methylation have been described. The first demethylase, identified in 2011, is the fat mass and obesity-associated protein (FTO) [33], which regulates processing and alternative splicing in adipocytes through m6A demethylation [34]. Mathiyalagan *et al.* discovered that FTO-dependent demethylation of m6A regulates intracellular  $\text{Ca}^{2+}$  dynamics and sarcomeres in cardiomyocytes [35]. FTO's activity as an m6A methyltransferase is interrelated with lipid accumulation control in muscle, as this enzyme plays a regulatory role in activating AMP kinase (AMPK) [36]. Wu *et al.* demonstrated that suppressing FTO significantly reduces the expression of CCNA2 and CDK2 genes, which play a key role in cell cycle regulation. This leads to delayed transitioning of cells exposed to insulin into the G2 phase of the cell cycle. Moreover, the level of m6A methylation of CCNA2 and CDK2 mRNA is substantially increased upon FTO suppression [37].

Another demethylase is the related alpha-ketoglutarate-dependent dioxygenase alkB homolog 5 (ALKBH5), described in 2013 [10]. ALKBH5 primarily interacts with m6A in RNA and is most highly expressed in testes and lungs. ALKBH5 is believed to regulate the assembly of mRNA processing factors [10]. Notably, m6A sites are the only substrates for ALKBH5, whereas FTO can erase other modifications like 2-O-dimethyladenosine, N1-methyladenosine, 3-methylthymine, and 3-methyluracil [38,39].

Reader proteins are the major factors that execute biological functions in m6A-modified RNA by recognizing and binding to methylated transcripts. Three major groups of reader proteins regulate m6A-mediated functions. The first group is proteins containing the conservative YT521-B domain (YTH) [40], which consists of approximately 150 amino acids distributed across three  $\alpha$ -helices and six  $\beta$ -sheets [41]. In humans, five proteins with the YTH domain have been identified: three paralogs of YTH domain family 1–3 (YTHDF1, YTHDF2, and YTHDF3) and distinct proteins of YTH domain containing 1–2 (YTHDC1 and YTHDC2). All of these proteins have structural differences. In particular, YTHDC1 contains the YTH domain surrounded by charged regions containing glutamic acid, arginine, or proline residues. YTHDC2 is the most complex protein in this group, containing not only the YTH domain but also several helicase domains and two ankyrin repeats, possessing ATPase and 3'→5'-helicase activity [42]. The YTHDF family of proteins, together with the YTH domain, also consist of disordered regions enriched in proline, glutamine, and asparagine [41].

YTHDF1–3 and YTHDC2 mainly localize in the cytoplasm, while YTHDC1 is predominantly distributed in the nucleus. YTHDF1 enhances the translation of m6A-modified RNAs by a yet unclarified mechanism; YTHDF1 binds RNA near the stop codon and interacts with translation initiation factor complex 3, which is part of the translation initiation complex [12]. YTHDF2's functions are primarily associated with mRNA degradation in the cytosol. It can activate the CCR4-NOT deadenylase complex, involved in mRNA deadenylation and degradation [43], and the P/MRP ribonuclease complex, which initiates endoribonucleolytic cleavage of mRNA. YTHDF3 is the least studied of this group. Li *et al.* noted that YTHDF3 regulates translation by interacting with YTHDF1 and with the 40S and 60S ribosome subunits [44]. However, it has also been determined that YTHDF3, along with YTHDF2, can participate in degrading RNA [45]. YTHDC1 actively participates in the regulation of transcription, splicing, and RNA export from the nucleus [46], while YTHDC2 contributes to enhancing translation efficiency and mRNA degradation through its helicase activity [47].

The second group of readers includes several heterogeneous nuclear ribonucleoproteins (HNRNP): HNRNPC, HNRNPG, and HNRNPA2B1. HNRNPC selectively binds to unstructured RNA regions during pre-mRNA processing and separates transcripts into mRNA and uridine-rich snRNA [48]. HNRNPG binds to arginine-glycine-glycine (RGG) regions and regulates alternative splicing of pre-mRNA by interacting with the phosphorylated C-terminal domain of RNA polymerase II [49]. HNRNPA2B1 accelerates the processing of primary microRNAs by interacting with the microprocessor complex subunit DGCR8 [50].

The third group of readers consists of three highly conserved insulin-like growth factor 2 mRNA-binding proteins (IGF2BP): IGF2BP1, IGF2BP2, and IGF2BP3. At the N-terminus, IGF2BP

proteins have two RNA recognition motifs (RRM); four hnRNP-K homology (KH) domains are located at the C-terminus [51]. KH domains are mainly responsible for protein-RNA binding. RRM domains regulate the stability of IGF2BP-RNA complexes [52]. The primary function of IGF2BP proteins is maintaining stability of target RNAs [53]. It is suggested that virtually any cellular protein can act as reader for m6A, enabling highly tunable and diverse control of RNA metabolism [54].

### 3. Mechanisms of the Antiviral Innate Immune Response

Organisms are constantly exposed to exogenous antigens, including pathogenic microorganisms like bacteria and viruses. To eliminate such pathogens and maintain the host's health, two interrelated immune systems have evolved: innate and acquired immunity. The immune system is tightly regulated to avoid the risk of non-specific recognition of self-antigens, which can not only impair the ability to eliminate pathogens but also negatively impact the host. Therefore, cells possess various regulatory mechanisms for both innate immune perception and the transmission of signals that initiate antimicrobial response reactions.

Recognition of nucleic acids is one strategy by which cells can detect infectious agents. In recent years, tremendous progress has been made in understanding how cells can activate the immune response at the molecular level. Binding of intracellular nucleic acids to a range of specialized sensors activates downstream signaling cascades, leading to the production of type I interferons (IFN) and pro-inflammatory cytokines, triggering corresponding systemic immune responses.

After cells are infected by a virus, the innate immune response is triggered by molecules called pathogen recognition receptors (PRRs), molecules that recognize specific molecular structures of pathogens and play a crucial role in innate immunity by activating the immune response upon contact with pathogens. Currently, five subfamilies of PRRs have been described: membrane Toll-like receptors (TLRs), RIG-I-like receptors (RLRs), Nod-like receptors (NLRs), AIM2-like receptors (ALRs), and C-type lectin receptors (CLRs) [55]. In most cases, membrane PRRs are actively expressed in immune cells, such as macrophages or dendritic cells (DCs). Intracellular receptors actively participate in processes related to the activation of the immune response, including apoptosis, phagocytosis, and the regulation of gene activity.

PRRs can interact with pathogen-associated molecular patterns (PAMPs) and damage-associated molecular patterns (DAMPs). In addition to intracellular receptors, viral genetic material in the cytoplasm can be identified by several factors, the most common of which are cyclic GMP-AMP synthase (cGAS), interferon-inducible protein 16 (IFI16), and absent in melanoma 2 (AIM2).

PAMPs are molecular patterns that consist of conserved microbial and viral components (nucleic acids, proteins, and carbohydrates) [56], and DAMPs are molecules released during cellular stress or tissue damage. These molecules act as endogenous danger signals, triggering inflammatory reactions and activating the innate immune system during non-infectious inflammatory processes [57].

Three main classes of PRRs can interact with viral genetic material: RLR [58], TLR [59], and a set of cytosolic sensors [60–62]. RLRs interact with viral RNA, which is transcribed from RNA and DNA viruses. TLRs can directly interact with various substrates, such as double-stranded RNA (dsRNA; TLR-3), single-stranded RNA (ssRNA; TLR-7/TLR-8), and CpG DNA (TLR-9) in endolysosomes upon viral entry.

After pathogen recognition, a cascade of protein reactions is initiated, leading to the production of various protective molecules in the organism, including IFNs, pro-inflammatory cytokines, and chemokines [63].

#### 3.1. Recognition of Viruses by RLP

Three proteins with DExD/H-box helicase activity belong to the RLP subfamily: RIG-I, melanoma differentiation association gene 5 (MDA5), and laboratory of genetics and physiology 2 (LGP2). RLRs include a central DExD/H-box helicase/ATPase domain and a C-terminal regulatory domain that binds to RNA and zinc ligands. RIG-I and MDA5 have two tandem caspase recruitment domains (CARDs) at their N-terminus [64,65] RIG-I recognizes relatively short ssRNA and dsRNA (up to 1 kb) with triphosphate or diphosphate fragments at the 5'-end [66]. In contrast, MDA5 detects

longer dsRNA (over 1 kb) formed during viral replication [67]. Further investigation is warranted to elucidate the function of LGP2, though LGP2 can interact with viral dsDNA and ssRNA, modulating the functions of RIG-I and MDA5. Conflicting findings suggest that LGP2 may exert a dual role in regulating RIG-I and MDA5 activity, either negatively or positively influencing MDA5 [61]. LGP2 has been observed to enhance the action of MDA5 [68], but can inhibit the action of the TRAF ubiquitin ligase, thereby negatively regulating the innate immune response [69].

RLRs are expressed in virtually all tissues. Upon binding to a virus, RIG-I and MDA5 undergo a conformational change, exposing the CARD domain [70,71], which binds to the CARD domain at the N-terminus of mitochondrial antiviral signaling (MAVS). MAVS is anchored to the outer membrane of the mitochondria through the C-terminal transmembrane domain, and forms functional aggregates upon activation [72]. This complex includes TNF receptor-associated factors (TRAF) and kinases TBK1 and I $\kappa$ B- $\epsilon$  (IKKe). The complex also consists of subunits IKK $\alpha$ , IKK $\beta$ , and IKK $\gamma$  of the tripartite-activated protein kinase (TAK1), which subsequently stimulates the activity of interferon regulatory factor 3 (IRF3) and/or IRF7, as well as nuclear factor kappa B (NF- $\kappa$ B) [73]. This cascade stimulates the production and release of IFNs, cytokines, and IFN-stimulated genes (ISGs). In addition to mitochondria, MAVS is found in peroxisomes, where it activates the expression of IFN III genes [74].

### 3.2. Recognition of Viral Nucleic Acids by TLRs

TLRs are located on the cell surface or inside the cell in organelles such as the endoplasmic reticulum (ER), endosomes, lysosomes, or endolysosomes. TLR synthesis occurs in the ER. Subsequently, TLRs exit the ER with the help of the protein UNC93B1 and move into endosomes through the plasma membrane or directly through the Golgi complex [59,75]. Each TLR molecule has an N-terminal domain with leucine-rich repeats (LRR), a transmembrane domain, and a Toll/IL-1 receptor (TIR) cytoplasmic domain at the C-terminus. The LRR domain recognizes PAMPs, and the TIR domain activates downstream signaling pathways. Viral nucleic acids are typically recognized by intracellular TLR3, TLR7, TLR8, and TLR9. TLR3 recognizes viral dsRNA, TLR7 and TLR8 recognize ssRNA, and TLR9 recognizes the oligodeoxynucleotide CpG [76].

The cellular antiviral response mediated by TLR is based on the recruitment of adapters containing the TIR domain (MyD88, TRIF, TIRAP/MAL, or TRAM). The TLR signaling pathway can be broadly divided into two pathways: MyD88-dependent and TRIF-dependent [77]. In the MyD88-dependent pathway, either the IKK complex or the MAPK pathway can be activated. The IKK complex releases NF- $\kappa$ B, which translocates to the nucleus and activates the expression of pro-inflammatory cytokine genes. The MAP kinase cascade is responsible for the formation of the transcription factor AP-1, which is also directed at cytokine genes. The TRIF-dependent pathway interacts with TRAF6 and TRAF3. In turn, TRAF6 activates the NF- $\kappa$ B signaling cascade and, consequently, the production of cytokines. TRAF3 induces the expression of IFN-I genes.

### 3.3. Recognition of Viruses Using Cytosolic Sensors

Viral DNA present in the cytosol of eukaryotic cells is also a PAMP. Cytoplasmic viral DNA can be recognized by a set of sensors like cGAS, IFI16, and AIM2. Additionally, several other proteins like DDX41 and RNA polymerase III have been noted to initiate innate immune responses in cells [78]. cGAS binds to the sugar-phosphate backbone of dsDNA without sequence specificity, allowing it to recognize a wide variety of DNA types. Due to this property, cGAS can detect numerous DNA species. The C-terminal domain of cGAS includes a nucleotidyltransferase. Upon binding to viral DNA, cGAS catalyzes the production of cGAMP from ATP and GTP. cGAMP, in turn, acts as a secondary messenger and can activate stimulator of IFN genes (STING) [79,80].

Another sensor of viral DNA in the cytosol is IFI16, a member of the PYHIN protein family. IFI16 is predominantly localized in the nucleus but can shuttle between the nucleus and the cytoplasm. IFI16 contains a pyrin domain at the N-terminus and two HIN200 domains at the C-terminus. It interacts with viral DNA through the HIN200 domain, after which IFI16 interacts with cGAS, initiating cGAMP production [81].

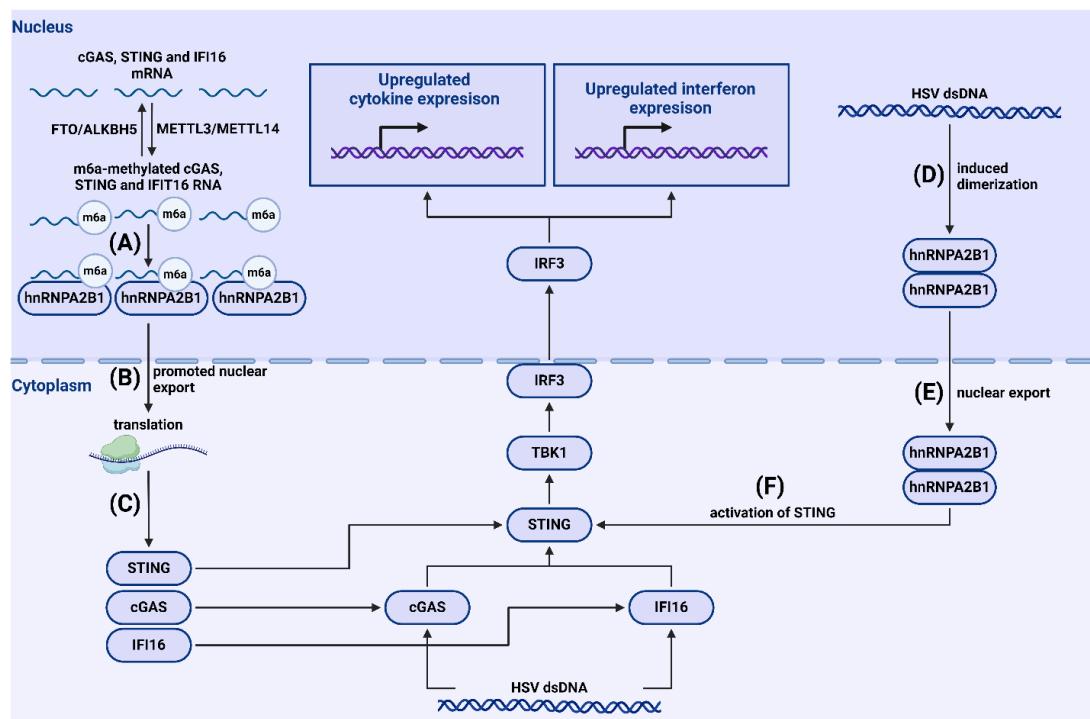
Also belonging to the PYHIN family is the AIM2 protein, which also recognizes viral DNA. Like IFI16, AIM2 consists of two domains: a pyrin domain at the N-terminus and an HIN200 domain at the C-terminus. The HIN200 domain is responsible for DNA binding, while the pyrin domain interacts with the pyrin domain of the adapter molecule ASC. AIM2 initiates caspase 1-dependent activation of inflammation, leading to the production of interleukin-1 $\beta$  (IL-1 $\beta$ ) and IL-18 [82].

Other sensors of viral DNA and RNA have been described in the cytosol, including DDX41 [83], RNA polymerase III [84], DNA-PK [85], OAS [86], and many other factors. However, there are still numerous questions regarding the role of these factors and intracellular immune signaling pathways, as well as the ways in which viruses evade detection. All these questions require further detailed description and study.

A key player in the activation of cellular immunity by the aforementioned agents is STING, an ER membrane protein that consists of a cytosolic N-terminal domain, four transmembrane helices forming the transmembrane domain, and a cytosolic ligand-binding domain (LBD) to which the C-terminal domain is attached. In cells, STING exists as a V-shaped dimer [87]. The LBD binds to cGAMP [88], and the C-terminal tail contains the PXPLRXD motif and is necessary for the activation of TANK-binding kinase 1 (TBK1) [89]. After activation, TBK1 phosphorylates IRF3, as mentioned earlier. Consequently, IRF3 translocates to the nucleus and induces the synthesis of anti-inflammatory cytokines and type I IFNs [90].

### 3.4. Recognition of Viruses in the Nucleus

Usually, PRRs are localized on the plasma membrane, in endosomes, or in the cytoplasm. However, some nuclear proteins can also serve as viral sensors. For example, the aforementioned IFI16, cGAS, and hnRNPA2B1 recognize herpes simplex virus 1 (HSV1) in the nucleus [91–93]. In uninfected cells, hnRNPA2B1 is methylated. Upon HSV1 infection, hnRNPA2B1 forms a complex with viral DNA, dimerizes, and is demethylated by JMJD6. This results in its cytoplasmic translocation followed by activation of TBK1, enhanced phosphorylation of IRF3, and activation of the immune response mediated by IFN I signaling (Figure 1) [94]. Additionally, Carpenter *et al.* demonstrated that hnRNPA2B1 factor bind HSV1 DNA in the nucleus, amplifying IFN $\beta$  antiviral signaling (Figure 1) [93].



**Figure 1. The role of hnRNPA2B1 in regulation of nuclear foreign DNA sensing.** (A) Nuclear hnRNPA2B1 binds to m6a-methylated cGAS, STING and IFI16 mRNA and (B) promotes their nuclear export. (C) Enhanced nuclear export of mRNA increases protein synthesis from mRNA of cGAS, STING and IFI16, resulting in enhanced cytosolic foreign DNA sensing and antiviral signaling. (D) Homodimerization of hnRNPA2B1 in HSV infection. (E) Homodimerized hnRNPA2B1 exits the nucleus. (F) Activation of TBK1-IRF3 pathway via STING by hnRNPA2B1 in the cytoplasm.

Gentili *et al.* found that nuclear cGAS enhances innate immune responses. Associated with centromeres and DNA repetitive sequences, nuclear cGAS can synthesize cGAMP and stimulate innate immune activity in primary DCs [95]. Nuclear cGAS can also bind to RNA viruses. In the nucleus, cGAS interacts with PRMT5 and facilitates symmetric demethylation of histone H3 arginine 2 at IFN-I promoter element, thus promoting interaction of activated IRF3 with this promoter and enhancing production of IFN I and CXCL10 [96].

Similar to its role as a cytoplasmic viral sensor, IFI16 functions as a viral sensor in the nucleus. Upon binding to viral DNA, IFI16 activates the nuclear protein DNA-PK, which, in turn, phosphorylates IFI16 at T149 [97]. This modification determines IFI16's subcellular localization and protein nuclear export, and promotes synthesis of type I IFNs. IFI16 undergoes a conformational change to adopt a filamentous structure during the course of infection, leading to its localization to viral replication sites [98]. After exiting to the cytoplasm, IFI16 can bind and activate STING [99].

As shown by Diner *et al.*, IFIX, another protein of the PYHIN family, also acts as a viral sensor in the nucleus. The authors noted that upon IFIX overexpression, HSV1 titers decreased almost three-fold, while IFIX knockdown significantly increased viral titers, suggesting that IFIX is an antiviral factor [145]. Crow and Cristea discovered that HSV-1 has acquired mechanisms to block IFIX function via proteasome-dependent degradation of the pyrin domain. Using immunoprecipitation mass-spectrometry (IP-MS), the authors demonstrated that IFIX interacts with components of the ubiquitin-proteasome system and transcriptional regulators during HSV-1 infection [100].

A nuclear sensor for viral dsRNA is scaffold attachment factor A (SAFA). SAFA interacts with dsRNA, oligomerizes, and activates DNA topoisomerase 1 and SWI/SNF-related matrix-associated actin-dependent regulator of chromatin subfamily A member 5, which, in turn, regulate the synthesis of enhancers for antiviral genes, including IFN- $\beta$ 1 [101].

Other note-worthy regulators are the non-POU domain-containing octamer binding (NONO) and hexamethylene bis-acetamide-inducible protein 1 (HEXIM1). NONO has been found to recognize the conserved capsid region of human immunodeficiency virus (HIV) and binds to cGAS [102]. HEXIM1 binds to the long non-coding RNA (lncRNA) NEAT1 and forms a complex that activates the cGAS-STING pathway [103].

#### 4. m6A Methylation of Antiviral Response Factors

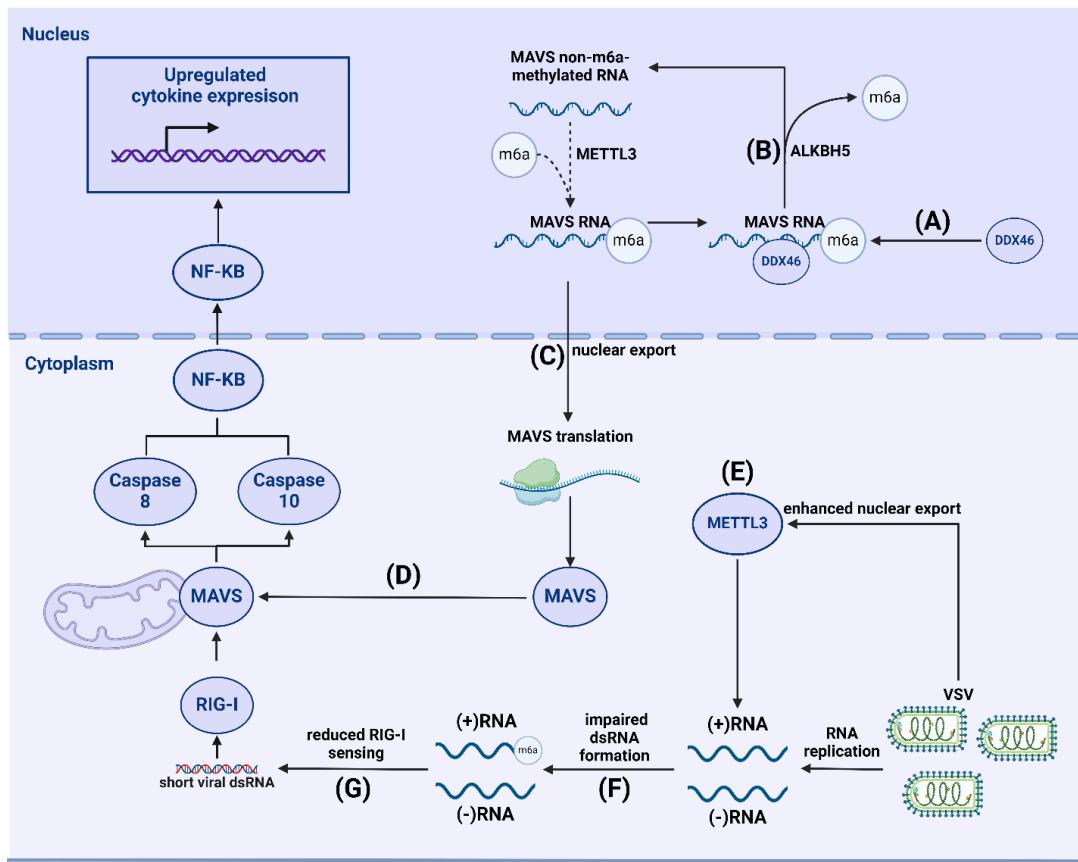
One important function of m6A modification is the activation and regulation of intracellular immunity. An early study showed that depleting METTL14 induces synthesis of type I IFNs, while depleting the demethylase ALKBH5 reduces type I IFN expression [104]. Further, m6A was found to be responsible for complex regulation of distinct antiviral signaling programs.

##### 4.1. m6A-Dependent Regulation of the RLP Signaling Pathway

###### 4.1.1. m6A and Stimulation of Immunity

m6A modification plays a crucial role in the RLP-dependent antiviral response pathway. m6A methylation is necessary for the export of MAVS, TRAF3, and TRAF6 mRNA factors from the nucleus. Zheng *et al.* found that during viral infection, the eraser protein ALKBH5 is activated, preventing the export of these transcripts from the nucleus and thus inhibiting IFN synthesis (Figure 2) [105]. Hesser and Walsh demonstrated that the expression of YTHDF readers was suppressed to varying degrees during infection with cowpox virus (VacV) and HSV1. The authors noted that decreased levels of YTHDF1, YTHDF2, and, to a lesser extent YTHDF3, led to increased expression

of ISGs and activation of antiviral response mechanisms [106], indicating the crucial role of readers in shaping the innate immune response. Knocking out METTL14 in macrophages infected with Sendai virus or encephalomyocarditis virus, Qin *et al.* found that demethylation of MAVS, which regulates the RLR-induced signaling pathway, enhanced expression, preserved protein stability, and consequently amplified IFN synthesis [107]. The authors also observed a gradual increase in the expression of eraser proteins FTO and ALKBH5 during infection with these viruses. Further research is needed to investigate the possibility of demethylation by eraser proteins and the role of demethylated MAVS.



**Figure 2. Interplay between VSV infection, m6a methylation and RIG-I-dependent RNA sensing.** (A) VSV causes DDX46 binding to m6a-methylated MAVS mRNA. (B) DDX46 recruits ALKBH5 and facilitates MAVS mRNA demethylation, (C) preventing its nuclear export. (D) Reduced nuclear export of MAVS mRNA leads to impaired RIG-I-mediated foreign RNA sensing. (E) VSV infection causes METTL3 translocation to the cytoplasm, where it methylates VSV (+) RNA. (F) M6a-methylation of VSV (+) RNA prevents dsRNA formation. (G) ssRNA is less prone to RIG-I sensing.

#### 4.1.2. Viral Evasion of RLP-Mediated Immune Responses

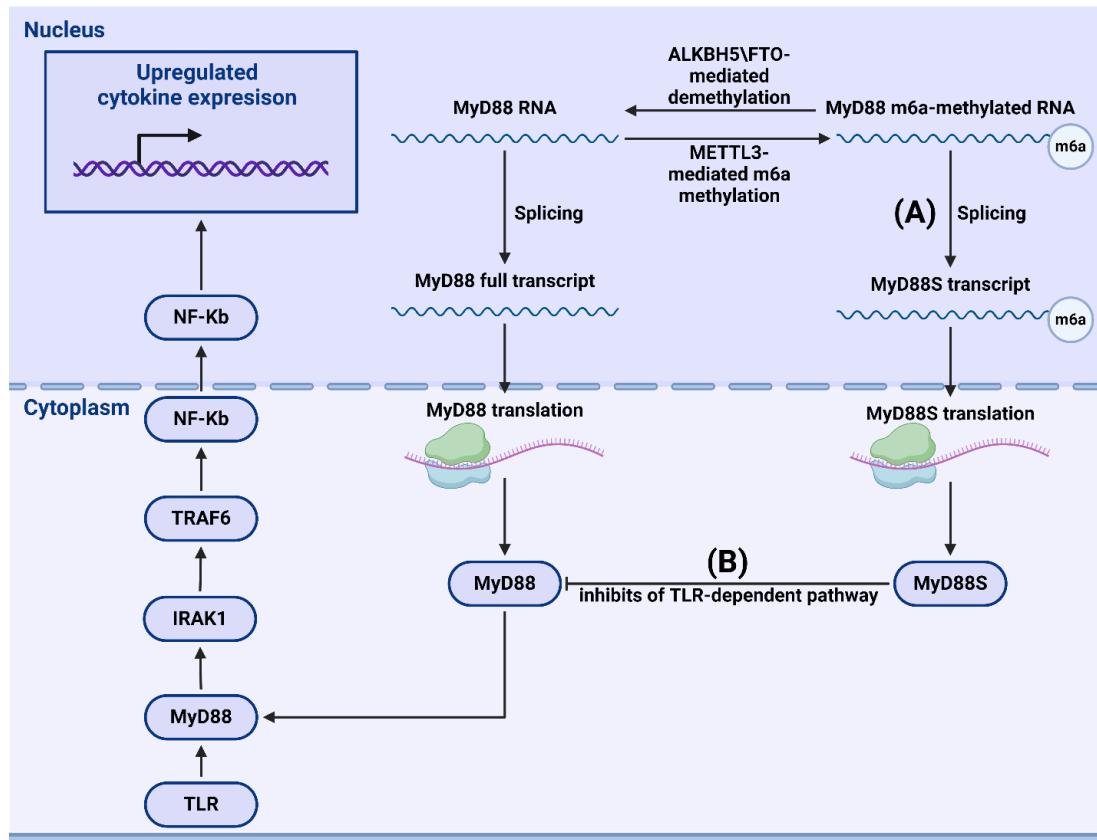
Viruses have developed strategies to evade RLP-mediated immunity. According to Li *et al.*, the 3'-UTR of the genome of severe acute respiratory syndrome coronavirus clade 2 (SARS-CoV-2) is enriched with m6A-modified nucleotides. SARS-CoV-2 utilizes the host's METTL3 methyltransferase for innate immune response evasion. Depleting METTL3 reduced m6A methylation in the SARS-CoV-2 genome and resulted in decreased viral RNA levels. It also led to reduced m6A methylation and expression of host proviral genes, including but not limited to NRP1, TRIM4, and SMAD4. In contrast, expression of innate immune response effector genes was upregulated in METTL3- and METTL14-depleted cells. This upregulation was due to the increased binding of m6A-unmethylated viral RNA to RIG-I, which was further confirmed by introducing mutations into SARS-CoV-2 m6A sites [108].

Qiu *et al.* found that METTL3 is a strong internal inhibitor of the innate immune response in vascular stomatitis virus (VSV)-infected cells, suppressing innate immunity signal transduction and inhibiting IRF synthesis. VSV infection robustly enhances translocation of METTL3 but not METTL14 into the cytoplasm. Using PAR-CLIP-seq and miCLIP-seq, the group identified METTL3 binding sites on positive-sense RNAs of VSV, unlike negative-sense genomic RNA. m6A methylation of viral RNA impeded dsRNA formation, making it less prone to sensing by RIG-I and MDA5 and subsequently reducing IFNB-1 production (Figure 2)[109].

#### 4.2. m6A-Dependent Regulation of the TLR Signaling Pathway

##### 4.2.1. m6A and Stimulation of Cellular Immunity through the TLR Pathway

m6A methylation can also influence the TLR-mediated signaling pathway. In their study of dental pulp inflammation, Feng *et al.* showed that knocking down methyltransferase METTL3 regulated alternative splicing of the TLR-mediated antiviral immune pathway participant MyD88, resulting in the production of MyD88S splice variant, which is known to inhibit inflammatory cytokine production. Depletion of METTL3 reduced the accumulation of inflammatory cytokines and suppressed the activation of NF- $\kappa$ B and MAPK signaling pathways (Figure 3) [110]. Similarly, METTL3 overexpression in a mouse model of inflammatory bowel diseases resulted in increased immune response via the NF- $\kappa$ B pathway [111]. Using m6A-modified in vitro RNA oligonucleotides, Karikó *et al.* demonstrated that m6A modification reduced the ability of RNA to induce cytokine secretion mediated by TLR3, TLR7, and TLR8 [112], indicating the crucial role of this methylation in pathogen recognition and differentiation between pathogen and host nucleic acids. Tong *et al.* used CRISPR tools to demonstrate that macrophages with METTL3 deficiency exhibit reduced activation associated with TLR signaling. Decreased methylation of IRAKM factor led to increased level of IRAKM, which inhibited signal transduction in TLR-dependent macrophage activation [113]. Additionally, m6A modification of CD40, CD80, and TIRAP transcripts promoted DC activation and DC-based immune response activation of TLR [114]. Notably, experiments by Geng *et al.* on brown croaker fish (*Miichthys miiuy*) showed that, in cells infected with *Siniperca chuatsi* rhabdovirus and *Vibrio anguillarum* bacteria, METTL3 inhibited the innate immune response via methylation of TRIF and MYD88, mRNA factors in the TLR pathway that are further degraded through YTHDF2- and YTHDF3-dependent mechanisms. In contrast, YTHDF1 promoted the translation of MYD88 RNA [115].



**Figure 3. Impact of m6a methylation on TLR-dependent antigen sensing.** (A) M6a methylation of MyD88 mRNA promotes alternative splicing and MyD88S transcript formation. (B) MyD88S protein prevents TLR-dependent signal transduction.

#### 4.2.2. m6A-Dependent Regulation of Cytosolic and Nuclear Sensors

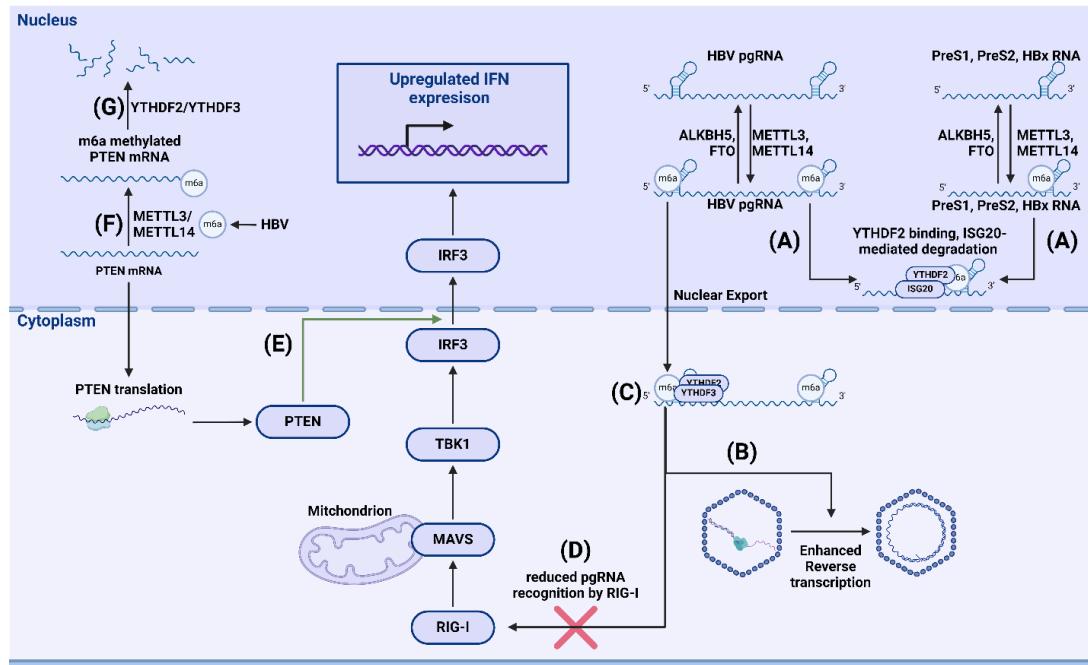
m6A methylation plays a crucial role in recognizing viruses in the nucleus. As previously mentioned, one nuclear sensor is the reader protein hnRNPA2B1. Wang *et al.* showed that, after binding to HSV-1 DNA in the nucleus, hnRNPA2B1 is demethylated at arginine-226 by the arginine demethylase JMJD6, after which it exits to the cytoplasm and activates the TBK1-IRF3 pathway in a STING-dependent manner. In vitro knockdown of hnRNPA2B1 or in vivo knockout of this factor in mice reduced the expression of IFN-I induced by DNA virus HSV-1; no such reduction was seen in RNA viruses. hnRNPA2B1 also promotes m6A modification and translocation to the cytoplasm of cGAS, STING, and IFI16 mRNA, enhancing their expression and ensuring a robust immune response (110). Balzarolo *et al.* identified that m6A modification augments recognition of cytosolic dsDNA through the cGAS-STING pathway. m6A methylation of 5'-GATC-3' motifs increases the immunogenicity of synthetic dsDNA in mouse macrophages and DC [116]. Song *et al.* showed that m6A can regulate lncRNAs associated with the cGAS-STING pathway, stimulating several key components of the cGAS-STING signaling pathway, including STAT1, DNA sensors cGAS and hnRNPA2B1, and the IFN receptor IFNAR1 [117].

#### 4.3. m6A-Dependent Regulation of Type I Interferon Synthesis

McFadden *et al.* conducted an extensive study and demonstrated that the MTC complex methylates a series of IFN-stimulated genes. Depletion of the METTL3/METTL14 complex resulted in decreased expression of ISGs IFITM1 and MX1 while the expression of ISG15 and EIF2AK2 remained unchanged. m6A modification in the 3'-UTR of IFITM1 enhances its translation through interaction with YTHDF1. The authors also noted that knockdown and overexpression of METTL3/14 proportionally altered the expression of type I IFN, but did not alter the expression of MX1, ISG15,

or EIF2AK2 [14]. Ge *et al.* observed that, in cells with WTAP knockdown, the expression of phosphorylated and total IRF3, which affects IFN synthesis, was reduced, and its dimerization was disrupted. In addition to IRF3, WTAP degradation led to decreased levels of IFNAR1, another key factor in IFN-I signaling. The authors also noted that in peripheral blood mononuclear cells (PBMC) infected with vesicular stomatitis virus with enhanced GFP, HSV1, or intracellular HSV-60 (a synthetic analog of HSV1 DNA), WTAP was degraded via the ubiquitination-proteasome pathway due to activation of IFN-I signaling. WTAP degradation resulted in reduced m6A methylation of IRF3 and IFNAR1 mRNA, resulting in reduced IRF3 and IFNAR1 protein levels and blocking of IFN-1 mediated innate response. The conclusion was that WTAP-IRF3/IFNAR1 signaling may serve as negative feedback pathway of INF-I signaling [118].

Imam *et al.* found that m6A-methylated hepatitis B virus (HBV) RNA is degraded by ISG20 via YTHDF2 m6A reader recruitment. They found that YTHDF2 protein was crucial for recognizing m6A-methylated viral RNA. After recognition, YTHDF2 recruited ISG20 to facilitate viral RNA degradation [119]. Kim *et al.* also reported that HBV expression induces m6A modification of PTEN mRNA, thus reducing PTEN RNA stability mainly through interaction with YTHDF2 and YTHDF3. PTEN increases nuclear import of IRF-3 and promotes subsequent IFN synthesis; thus, decreased PTEN expression due to the lower stability of m6A-methylated mRNA hinders IFN production. Data from patients with hepatocellular carcinoma (HCC), HBV-positive patients with HCC, and patients negative for both HBV and HCC also suggest that HBV-induced m6A-methylation of PTEN might be responsible for HCC development via disruption of the PI3K pathway (Figure 4) [120].



**Figure 4. Impact of m6a methylation on RLR-sensing of HBV.** (1) M6a methylation of 3' epsilon stem loop results in HBV RNA binding with YTHDF2 and subsequent ISG20-mediated degradation. (2) M6a methylation of 5' epsilon stem loop enhances reverse transcription through currently unknown mechanism. (3) 5' epsilon stem loop methylation causes binding of YTHDF2 and YTHDF3 to pgRNA. (4) YTHDF2/3 bound to pgRNA reduce pgRNA sensing by RIG-I. (5) PTEN enhances nuclear import of IRF3, leading to upregulated IFN production. (6) HBV infection increases m6a modification of PTEN mRNA. (7) Methylated PTEN mRNA is recognized by YTHDF2/YTHDF3 and subsequently degraded.

Similarly, Zhang *et al.* showed that knocking down the reader protein YTHDF3 reduced the replication of VSV, HSV1, and encephalomyocarditis virus in infected cells. Subsequently, the authors used RNA sequencing to demonstrate that YTHDF3 knockout cells exhibited higher expression of various ISGs, including IFIT1, CXCL10, MX1, and OAS1a. However, it was noted that

YTHDF3 did not bind to ISGs or signaling factors of the JAK-STAT pathway and did not affect the stability of ISG mRNA. According to eCLIP-seq data, even basal levels of YTHDF3 could inhibit IFN synthesis by promoting FOXO3 protein translation. FOXO3 is known to negatively regulate ISG expression. YTHDF3 binds FOXO3 mRNA independently of METTL3-mediated m6A modifications and promotes its translation through interaction with poly-A binding protein and eukaryotic translation initiation factor 4 gamma 2 [121].

## 5. The Role of the m6A Modifications in the Genetic Material of Viruses

In addition to its involvement in cellular processes, the m6A modification has been shown to impact the regulation of the viral life cycle. The presence of m6A markers in viruses has been recognized for several decades, but its functional significance has been poorly understood just until recently. Initially identified in nuclear viral DNA, the role of m6A was later described for RNA viruses. Upon infection, writer and eraser proteins can translocate from the nucleus to the cytoplasm, where they regulate methylation of viral nucleic acids. Further investigation of m6A methylation in viruses is crucial for understanding how it influences the viral epitranscriptome and may provide insights for developing novel prognostic biomarkers and antiviral therapies.

### 5.1. Retroviruses

HIV-1, a representative of the *Retroviridae* family, is a positive ssRNA virus that replicates in the nucleus and is susceptible to m6A methylation. Tirumuru *et al.* demonstrated that METTL3 and METTL14 knockdown resulted in inhibited HIV-1 Gag protein expression. Conversely, knocking down ALKBH5 and FTO resulted in higher HIV protein expression; notably, FTO knockdown caused a significant increase in HIV protein expression. m6A modifications in HIV-1 genome can interact with cellular YTHDF1–3 reader proteins. Partially knocking down YTHDF1 or YTHDF3 in Jurkat cells resulted in three- to four-fold increase in HIV-1 infection, while only a slight increase was observed after YTHDF2 knockdown. Overexpressing each of the YTHDF1–3 proteins significantly reduced the level of HIV-1 reverse transcriptase products by four- to ten-fold. This downregulation of reverse transcription was attributed to mRNA degradation, mediated by YTHDF1–3 [122]. In contrast, in the study by Tsai *et al.*, a three-fold increase was observed in the overall expression of HIV-1 viral RNA in cells with YTHDF2 protein overexpression 24 hours after infection. The authors noted that YTHDF2 and another reader, YTHDC1, bind to m6A sites in HIV-1 transcripts. Knockdown of YTHDC1 increased viral Gag expression, while overexpressing YTHDC1 reduced viral RNA expression. Deep sequencing of splice forms demonstrated that YTHDC1 can regulate alternative splicing of HIV-1 transcripts [123]. Chen *et al.* assessed HIV content in cells with different levels of expression of FTO and ALKBH5 eraser proteins, finding that decreasing methylation levels enhanced binding to RIG-I [124].

### 5.2. Orthomyxoviruses

Influenza A virus (IAV) belongs to the *Orthomyxoviridae* family, characterized by negative-sense segmented RNA genomes that replicate in the host cell nucleus. Courtney *et al.* investigated how m6A affects the expression and replication of IAV genes. The authors demonstrated that METTL3 knockout and chemical inhibition of methylation using 3-deazaneplanocin A in A549 cells reduced IAV replication levels by suppressing viral mRNA and protein expression. The study also showed that increased regulation of the reader protein YTHDF2 significantly enhanced IAV replication and viral particle production. Additionally, the locations of m6A residues on viral mRNA were mapped using RNA-protein crosslinking and immunoprecipitation. These mapping data were confirmed by generating mutant forms of IAV, in which eight prominent m6A sites on the plus-strand mRNA/cRNA of hemagglutinin (HA) or nine m6A sites on the minus-strand vRNA of HA were mutated. In these IAV mutants, selectively lower levels of mRNA and HA protein were expressed, while the expression of other IAV mRNA and proteins remained unaffected [125]. Additionally, Zhu *et al.* identified the involvement of another reader protein, YTHDC1, which was found to be

upregulated in IAV-infected cells. Using mass spectrometry and co-immunoprecipitation experiments, the authors showed that YTHDC1 interacts with the non-structural protein NS1 of IAV, thereby promoting viral replication. Mutations in NS1 (NS1 R38A and K41A) disrupted its ability to inhibit NS segment splicing and hindered its interaction with YTHDC1. Knockdown of YTHDC1 enhanced NS segment splicing, while restoring YTHDC1 reversed this effect. The study concluded that the interaction between NS1 and YTHDC1 plays a critical role in regulating NS mRNA splicing during IAV infection [11].

### 5.3. Flaviviruses

Flaviviruses are a group of viruses characterized by positive-sense ssRNA genomes that replicate in the cytoplasm. This viral family includes pathogens such as yellow fever virus, West Nile virus, dengue virus, tick-borne encephalitis virus, Zika virus (ZIKV), and hepatitis C virus (HCV). Lichinchi *et al.* observed that ZIKV can undergo m6A methylation mediated by host METTL3 and METTL14 as well as by ALKBH5 and FTO. m6A methylation was shown to negatively regulate ZIKV production. Knockdown of YTHDF1-3 readers increased viral RNA levels, while overexpressing YTHDF1-3 decreased levels of extracellular viral RNA [126]. Gokhale *et al.* conducted a comprehensive study mapping m6A modification sites in HCV, dengue virus, yellow fever virus, ZIKV, and West Nile virus. In addition to mapping, the authors investigated the role of m6A methylation in HCV and noted that depleting METTL3 and METTL14 increased the rate of HCV infection, promoting the formation of infectious viral particles without affecting viral RNA replication. Depleting the m6A demethylase FTO had the opposite effect. Additionally, YTHDF readers reduced the production of HCV particles and localized to viral assembly sites. All three YTHDF proteins bound to HCV RNA at the same sites, and their depletion increased the production of HCV particles [127]. The role of the YTHDC2 reader was established in the study by Kim and Siddiqui. Using cells with a knockout of the YTHDC2 helicase domain (YTHDC2-E332Q), they showed that the expression levels of HCV core protein were higher in cells transfected with wild-type YTHDC2 but significantly lower in cells transfected with YTHDC2-E332Q. Additionally, in the same study, the authors noted that suppressing METTL3 and METTL14 increased the stability of HCV RNA [128]. In experiments with HBV and HCV, Kim *et al.* showed that METTL3/METTL14 depletion led to increased viral RNA recognition by RIG-I with subsequent increases in IFN1 production. Furthermore, RNA binding by YTHDF2 and YTHDF3 was found to be the reason for decreased RIG-I sensing [129]. The mechanism behind the evasion of the RIG-I-mediated antiviral response through m6A methylation and mimicry of cellular RNAs has been identified in representatives of the *Pneumoviridae*, *Paramyxoviridae*, and *Rhabdoviridae* viral families [130,131].

Gokhale *et al.* found negative regulation of HCV viral particle production by METTL3/METTL14 methyltransferases and YTHDF1, YTHDF2, and YTHDF3 readers. Additionally, FTO demethylase positively regulated HCV particle production, while ALKBH5 levels did not affect viral particle production [127]. Further, m6A methylation of HCV RNA was shown to downregulate viral particle production and reduce the efficiency of RIG-I-mediated viral RNA sensing.

### 5.4. Coronaviruses

SARS-CoV-2, a  $\beta$ -coronavirus with positive-sense ssRNA, contains eight m6A modifications in its genome, as shown via m6A-seq and miCLIP analysis conducted by Liu *et al.* The authors also demonstrated that m6A RNA methylation negatively regulates the life cycle of SARS-CoV-2. In Huh7 cells infected with SARS-CoV-2, abundant METTL14 and ALKBH5 relocated to the cytoplasm, where coronavirus genomic RNA replication occurs. Virus replication and the percentage of SARS-CoV-2-positive cells significantly increased after METTL3 and METTL14 knockdown. Knocking down YTHDF2, but not YTHDF1 or YTHDF3, promoted viral infection and replication. SARS-CoV-2 can influence host cell m6A methylation, as the overall intensity of m6A significantly increased in Huh7 cells infected with SARS-CoV-2 compared to uninfected Huh7 cells [132]. Burgess *et al.* demonstrated that METTL3, YTHDF1, and YTHDF3 are necessary for the replication of human  $\beta$ -coronaviruses. The catalytic function of METTL3 plays a crucial role in the efficient synthesis of viral RNA within

the first 24 hours post infection, leading to subsequent accumulation of viral proteins [133]. Liu *et al.* conducted a systematic analysis of m6A in different strains of SARS-CoV-2 causing mild or severe forms of COVID-19, and concluded that the presence of more m6A modifications in the N region of SARS-CoV-2 correlated with weaker pathogenicity. The authors described several methylation sites that may be associated with viral pathogenicity, such as site 74 located in the transcription regulatory sequence of the 5'-UTR of SARS-CoV-2, and a similar site, 29 707, in the 3'-UTR [134].

### 5.5. *Hepadnaviruses*

HBV is a hepatotropic DNA virus from *Hepadnaviridae* family, characterized by a complex life cycle involving a stage of reverse transcription. Imam *et al.* used m6A-seq analysis to identify m6A methylation sites within a conserved motif located in the epsilon stem loop at the 3' end of all HBV mRNA, as well as at the 5' and 3' ends of pregenomic RNA (pgRNA). Their results showed that m6A methylation in the 5'-epsilon stem loop of pgRNA was necessary for pgRNA reverse transcription, while m6A methylation in the 3'-epsilon stem loop of pgRNA destabilized all HBV transcripts [135]. Moreover, m6A RNA modification of HBV is necessary for efficient replication of the virus in hepatocytes. As shown by Murata *et al.*, m6A modifications primarily occur in the coding region of HBx, and mutating m6A sites decreased HBV and HBs RNA levels. The authors also assessed the impact of the m6A methylation inhibitor cycloleucine on HBV. Cycloleucine reduced HBV RNA levels in cells and HBs levels in a dose-dependent manner, while 3-[4,5-dimethylthiazol-2-yl]-5-[3-carboxymethoxyphenyl]-2-[4-sulfophenyl]-2H-tetrazolium (MTS) analysis showed that cell viability was unaffected. METTL3 knockdown also reduced HBV and HBs RNA levels, indicating the crucial role of m6A in the HBV life cycle [136]. Kim *et al.* found that m6A methylation of HBV transcripts regulates their intracellular distribution. Transcripts of HBV in cells transfected with m6A mutant plasmids predominantly accumulated in the nucleus compared to wild-type cells. The authors also noted that knocking out YTHDC1 and fragile X mental retardation protein (FMRP) affected the nuclear export of m6A-modified HBV transcripts. MeRIP analysis showed that most m6A-modified viral RNAs located to the cytoplasm, but when cells are depleted of YTHDC1 or FMRP, m6A-methylated viral RNAs accumulated in the nucleus [137].

Studies by Imam *et al.* demonstrated that depleting METTL3 and METTL14 resulted in increased expression of HBV HBs and HBc proteins. Similarly, depleting FTO and ALKBH5 demethylases decreased expression of said proteins, suggesting that m6A methylation downregulates HBV protein expression. Notably, depleting YTHDF2 and YTHDF3 also resulted in increased expression of HBs and HBc proteins. Further investigations uncovered more than two-fold increase in pgRNA stability in cells depleted of either YTHDF2 or METTL3/METTL14; this was ruled to be due to reduced stability of m6A-modified HBV RNA. This effect might be due to YTHDF2-mediated RNA degradation. The role of m6A methylation in the interaction between HBV and cells is further discussed by Kostyusheva *et al.* [138].

Zhang *et al.* conducted a bioinformatics analysis of m6A regulators associated with immune infiltration in HBV-related HCC. The authors demonstrated that m6A influences oncogenesis, tumor microenvironment, and patient prognosis in HBV-related HCC [139].

### 5.6. *Adenoviruses*

Adenoviruses (AdV), dsDNA viruses, undergo replication in the nucleus utilizing cellular RNA polymerase II. It was demonstrated that methyltransferases METTL3, METTL14, and WTAP, and the reader protein YTHDC1, translocate to sites of viral RNA biosynthesis in A549 cells infected with AdV5 within 18 hours of infection, suggesting that m6A modification may play a role in regulating late viral RNA splicing efficiency. Knockout of METTL3 or WTAP did not affect the splicing efficiency of the E1A gene, expressed during early infection stages, but significantly reduced splicing efficiency of the fiber gene, expressed later. Similar changes were observed in cells lacking YTHDC1, albeit to a lesser extent [140]. Hajikhezri *et al.* noted that the absence of FXR1 reduced the accumulation of viral capsid protein during human AdV-5 infection in cell culture. The authors found that the FXR1 protein accumulates at a late stage of human AdV-5 infection and forms distinct subcellular condensates.

CLIP-qPCR revealed that the endogenous FXR1 protein binds to m6A-modified viral capsid mRNA [141].

### 5.7. Herpesviruses

Transcripts of the HSV-1 genome can contain 12 sites of m6A methylation. Feng *et al.* noted that the expression of methyltransferases METTL3 and METTL14 and readers YTHDF1–3 increased in the early stages of HSV-1 infection and decreased in the later stages; the expression of demethylases FTO and ALKBH5 was reduced [142]. Hesser and Walsh also observed that the infection time of HSV1 suppresses YTHDF proteins [106]. Jansens *et al.* found that suppressing m6A-methylated HSV transcripts depended on the YTHDF reader protein family and correlated with the localization of these proteins to enlarged P-bodies [143]. Wang *et al.* identified that METTL3 is aberrantly expressed in mouse corneal endothelial cells and human umbilical vein endothelial cells infected with HSV1. HSV-1 infection may lead to increased levels of m6A in endothelial cells, but this does not always correlate with METTL3 levels [144]. Xu *et al.* demonstrated that the level of m6A RNA modification changed after infection of oral epithelial cells with HSV-1, likely regulated by changes in the expression of demethylases FTO and ALKBH5 [145].

## 6. Conclusions and Future Directions

Despite RNA methylation being described over forty years ago, the expanding significance of epitranscriptomic markers is still to be understood. m6A methylation is the most common and well-studied of these markers, but its role in virology was practically unstudied until recently. Now, viral epitranscriptomics has actively started to develop. Increasing attention is given to the role of m6A in regulating the reverse transcription and translation of viral genetic material, as well as the mechanisms of viral evasion of the host organism's immune system. m6A modifications can determine the strength and duration of the activation of the innate immune response. Studying the impact of m6A methylation on innate antiviral immunity, viral life cycles, and the ability of viruses to mask and evade sensor cells holds numerous perspectives and practical applications.

Investigating how m6A methylation can influence the expression of genes involved in the innate immune response may provide a comprehensive answer to how m6A modifications regulate the transcription and translation of key genes associated with immunity.

Understanding the role of m6A modifications in the recognition of viral RNA by PRRs and the subsequent activation or inhibition of antiviral pathways warrants further investigation. Some factors recognizing viral DNA may be present not only in the cytosol but also in the nucleus [81]. This raises additional questions: how can proteins associated with innate immunity differentiate between host and viral DNA, and what role does m6A play in these processes?

Studying the functions of proteins linked to m6A methylation is also crucial. Since m6A is a dynamic regulation, understanding the role of the proteins regulating these modifications can provide an overall insight into the regulation of m6A modification during immune reactions and, consequently, the dynamism of immune responses.

Recently, the role of lncRNAs in the organization of intracellular immunity was actively investigated [146]. Exploring the influence of m6A methylation on lncRNAs associated with innate immunity will allow understanding how m6A modifications in lncRNAs can affect their stability, localization, and interactions with other cellular components.

m6A is not the only modification to play a role in the organization of intercellular immunity [112]. Researching the interaction between m6A methylation and other epigenetic modifications in the context of innate immunity, such as how m6A interacts with DNA methylation, histone modifications, and other RNA modifications during the formation of immune responses, will establish additional mechanisms for regulating antiviral immune reactions.

An important direction is exploring the potential impact on m6A regulators as a therapeutic strategy for activating innate immune responses. Understanding regulatory mechanisms could open opportunities for developing interventions aimed at enhancing or suppressing immune reactions in various viral diseases. Studying how disrupting regulation of m6A modifications is linked to

impairments in signaling pathways of antiviral immunity will help identify new diagnostic markers and therapeutic targets.

Investigating the involvement of m6A methylation in the innate immune response holds great prospects for further research into host-pathogen interactions and immune regulation. Studying m6A methylation's role may have broad implications for the development of therapeutic agents related to modulating immune responses and treating viral diseases.

**Funding:** This research was funded by RSF grant № 22-75-10032. Section 2 was supported by the Ministry of Science and Higher Education of Russia no. 075-01551-23-00 (FSSF-2023-0006) (Vadim S. Pokrovsky).

## References

1. R. Desrosiers, K. Friderici, F. Rottman, Identification of methylated nucleosides in messenger RNA from Novikoff hepatoma cells., *Proc. Natl. Acad. Sci. U. S. A.* 71 (1974) 3971–3975. <https://doi.org/10.1073/pnas.71.10.3971>.
2. P. Boccalotto, M.A. Machnicka, E. Purta, P. Piatkowski, B. Baginski, T.K. Wirecki, V. de Crécy-Lagard, R. Ross, P.A. Limbach, A. Kotter, M. Helm, J.M. Bujnicki, MODOMICS: a database of RNA modification pathways. 2017 update., *Nucleic Acids Res.* 46 (2018) D303–D307. <https://doi.org/10.1093/nar/gkx1030>.
3. K. Beemon, J. Keith, Localization of N6-methyladenosine in the Rous sarcoma virus genome., *J. Mol. Biol.* 113 (1977) 165–179. [https://doi.org/10.1016/0022-2836\(77\)90047-x](https://doi.org/10.1016/0022-2836(77)90047-x).
4. S. Zaccara, R.J. Ries, S.R. Jaffrey, Reading, writing and erasing mRNA methylation., *Nat. Rev. Mol. Cell Biol.* 20 (2019) 608–624. <https://doi.org/10.1038/s41580-019-0168-5>.
5. B.E. Maden, Identification of the locations of the methyl groups in 18 S ribosomal RNA from *Xenopus laevis* and man., *J. Mol. Biol.* 189 (1986) 681–699. [https://doi.org/10.1016/0022-2836\(86\)90498-5](https://doi.org/10.1016/0022-2836(86)90498-5).
6. T. Berulava, S. Rahmann, K. Rademacher, L. Klein-Hitpass, B. Horsthemke, N6-adenosine methylation in MiRNAs, *PLoS One.* 10 (2015) e0118438. <https://doi.org/10.1371/journal.pone.0118438>.
7. S. Ke, A. Pandya-Jones, Y. Saito, J.J. Fak, C.B. Vågbø, S. Geula, J.H. Hanna, D.L. Black, J.E.J. Darnell, R.B. Darnell, m(6)A mRNA modifications are deposited in nascent pre-mRNA and are not required for splicing but do specify cytoplasmic turnover., *Genes Dev.* 31 (2017) 990–1006. <https://doi.org/10.1101/gad.301036.117>.
8. D. Dominissini, S. Moshitch-Moshkovitz, S. Schwartz, M. Salmon-Divon, L. Ungar, S. Osenberg, K. Cesarkas, J. Jacob-Hirsch, N. Amariglio, M. Kupiec, R. Sorek, G. Rechavi, Topology of the human and mouse m6A RNA methylomes revealed by m6A-seq., *Nature.* 485 (2012) 201–206. <https://doi.org/10.1038/nature11112>.
9. X. Wang, Z. Lu, A. Gomez, G.C. Hon, Y. Yue, D. Han, Y. Fu, M. Parisien, Q. Dai, G. Jia, B. Ren, T. Pan, C. He, N6-methyladenosine-dependent regulation of messenger RNA stability., *Nature.* 505 (2014) 117–120. <https://doi.org/10.1038/nature12730>.
10. G. Zheng, J.A. Dahl, Y. Niu, P. Fedorcsak, C.-M. Huang, C.J. Li, C.B. Vågbø, Y. Shi, W.-L. Wang, S.-H. Song, Z. Lu, R.P.G. Bosmans, Q. Dai, Y.-J. Hao, X. Yang, W.-M. Zhao, W.-M. Tong, X.-J. Wang, F. Bogdan, K. Furu, Y. Fu, G. Jia, X. Zhao, J. Liu, H.E. Krokan, A. Klungland, Y.-G. Yang, C. He, ALKBH5 is a mammalian RNA demethylase that impacts RNA metabolism and mouse fertility., *Mol. Cell.* 49 (2013) 18–29. <https://doi.org/10.1016/j.molcel.2012.10.015>.
11. Y. Zhu, R. Wang, J. Zou, S. Tian, L. Yu, Y. Zhou, Y. Ran, M. Jin, H. Chen, H. Zhou, N6-methyladenosine reader protein YTHDC1 regulates influenza A virus NS segment splicing and replication., *PLoS Pathog.* 19 (2023) e1011305. <https://doi.org/10.1371/journal.ppat.1011305>.
12. X. Wang, B.S. Zhao, I.A. Roundtree, Z. Lu, D. Han, H. Ma, X. Weng, K. Chen, H. Shi, C. He, N(6)-methyladenosine Modulates Messenger RNA Translation Efficiency., *Cell.* 161 (2015) 1388–1399. <https://doi.org/10.1016/j.cell.2015.05.014>.
13. S. Lavi, A.J. Shatkin, Methylated simian virus 40-specific RNA from nuclei and cytoplasm of infected BSC-1 cells., *Proc. Natl. Acad. Sci. U. S. A.* 72 (1975) 2012–2016. <https://doi.org/10.1073/pnas.72.6.2012>.
14. M.J. McFadden, A.B.R. McIntyre, H. Mourelatos, N.S. Abell, N.S. Gokhale, H. Ipas, B. Xhemalçe, C.E. Mason, S.M. Horner, Post-transcriptional regulation of antiviral gene expression by N6-methyladenosine., *Cell Rep.* 34 (2021) 108798. <https://doi.org/10.1016/j.celrep.2021.108798>.
15. M.J. McFadden, S.M. Horner, N(6)-Methyladenosine Regulates Host Responses to Viral Infection., *Trends Biochem. Sci.* 46 (2021) 366–377. <https://doi.org/10.1016/j.tibs.2020.11.008>.
16. M. Lu, Z. Zhang, M. Xue, B.S. Zhao, O. Harder, A. Li, X. Liang, T.Z. Gao, Y. Xu, J. Zhou, Z. Feng, S. Niewiesk, M.E. Peeples, C. He, J. Li, N(6)-methyladenosine modification enables viral RNA to escape recognition by RNA sensor RIG-I., *Nat. Microbiol.* 5 (2020) 584–598. <https://doi.org/10.1038/s41564-019-0653-9>.

17. E. Schöller, F. Weichmann, T. Treiber, S. Ringle, N. Treiber, A. Flatley, R. Feederle, A. Bruckmann, G. Meister, Interactions, localization, and phosphorylation of the m(6)A generating METTL3-METTL14-WTAP complex., *RNA*. 24 (2018) 499–512. <https://doi.org/10.1261/rna.064063.117>.
18. J. Choe, S. Lin, W. Zhang, Q. Liu, L. Wang, J. Ramirez-Moya, P. Du, W. Kim, S. Tang, P. Sliz, P. Santisteban, R.E. George, W.G. Richards, K.-K. Wong, N. Locker, F.J. Slack, R.I. Gregory, mRNA circularization by METTL3-eIF3h enhances translation and promotes oncogenesis., *Nature*. 561 (2018) 556–560. <https://doi.org/10.1038/s41586-018-0538-8>.
19. P. Wang, K.A. Doxtader, Y. Nam, Structural Basis for Cooperative Function of Mettl3 and Mettl14 Methyltransferases., *Mol. Cell*. 63 (2016) 306–317. <https://doi.org/10.1016/j.molcel.2016.05.041>.
20. J. Liu, Y. Yue, D. Han, X. Wang, Y. Fu, L. Zhang, G. Jia, M. Yu, Z. Lu, X. Deng, Q. Dai, W. Chen, C. He, A METTL3-METTL14 complex mediates mammalian nuclear RNA N6-adenosine methylation., *Nat. Chem. Biol.* 10 (2014) 93–95. <https://doi.org/10.1038/nchembio.1432>.
21. X.-L. Ping, B.-F. Sun, L. Wang, W. Xiao, X. Yang, W.-J. Wang, S. Adhikari, Y. Shi, Y. Lv, Y.-S. Chen, X. Zhao, A. Li, Y. Yang, U. Dahal, X.-M. Lou, X. Liu, J. Huang, W.-P. Yuan, X.-F. Zhu, T. Cheng, Y.-L. Zhao, X. Wang, J.M. Rendtlew Danielsen, F. Liu, Y.-G. Yang, Mammalian WTAP is a regulatory subunit of the RNA N6-methyladenosine methyltransferase., *Cell Res.* 24 (2014) 177–189. <https://doi.org/10.1038/cr.2014.3>.
22. D.P. Patil, C.-K. Chen, B.F. Pickering, A. Chow, C. Jackson, M. Guttman, S.R. Jaffrey, m(6)A RNA methylation promotes XIST-mediated transcriptional repression., *Nature*. 537 (2016) 369–373. <https://doi.org/10.1038/nature19342>.
23. J. Wen, R. Lv, H. Ma, H. Shen, C. He, J. Wang, F. Jiao, H. Liu, P. Yang, L. Tan, F. Lan, Y.G. Shi, C. He, Y. Shi, J. Diao, Zc3h13 Regulates Nuclear RNA m(6)A Methylation and Mouse Embryonic Stem Cell Self-Renewal., *Mol. Cell*. 69 (2018) 1028–1038.e6. <https://doi.org/10.1016/j.molcel.2018.02.015>.
24. Y. Yue, J. Liu, X. Cui, J. Cao, G. Luo, Z. Zhang, T. Cheng, M. Gao, X. Shu, H. Ma, F. Wang, X. Wang, B. Shen, Y. Wang, X. Feng, C. He, J. Liu, VIRMA mediates preferential m(6)A mRNA methylation in 3'UTR and near stop codon and associates with alternative polyadenylation., *Cell Discov.* 4 (2018) 10. <https://doi.org/10.1038/s41421-018-0019-0>.
25. P. Bawankar, T. Lence, C. Paolantoni, I.U. Haussmann, M. Kazlauskiene, D. Jacob, J.B. Heidelberger, F.M. Richter, M.P. Nallasivan, V. Morin, N. Kreim, P. Beli, M. Helm, M. Jinek, M. Soller, J.-Y. Roignant, Hakai is required for stabilization of core components of the m(6)A mRNA methylation machinery., *Nat. Commun.* 12 (2021) 3778. <https://doi.org/10.1038/s41467-021-23892-5>.
26. E.M. Turkalj, C. Vissers, The emerging importance of METTL5-mediated ribosomal RNA methylation., *Exp. Mol. Med.* 54 (2022) 1617–1625. <https://doi.org/10.1038/s12276-022-00869-y>.
27. R. Pinto, C.B. Vågbø, M.E. Jakobsson, Y. Kim, M.P. Baltissen, M.-F. O'Donohue, U.H. Guzmán, J.M. Małecki, J. Wu, F. Kirpekar, J. V Olsen, P.-E. Gleizes, M. Vermeulen, S.A. Leidel, G. Slupphaug, P.Ø. Falnes, The human methyltransferase ZCCHC4 catalyses N6-methyladenosine modification of 28S ribosomal RNA., *Nucleic Acids Res.* 48 (2020) 830–846. <https://doi.org/10.1093/nar/gkz1147>.
28. J.A. Brown, C.G. Kinzig, S.J. DeGregorio, J.A. Steitz, Methyltransferase-like protein 16 binds the 3'-terminal triple helix of MALAT1 long noncoding RNA., *Proc. Natl. Acad. Sci. U. S. A.* 113 (2016) 14013–14018. <https://doi.org/10.1073/pnas.1614759113>.
29. D.J. Nance, E.R. Satterwhite, B. Bhaskar, S. Misra, K.R. Carraway, K.D. Mansfield, Characterization of METTL16 as a cytoplasmic RNA binding protein., *PLoS One*. 15 (2020) e0227647. <https://doi.org/10.1371/journal.pone.0227647>.
30. A.S. Warda, J. Kretschmer, P. Hackert, C. Lenz, H. Urlaub, C. Höbartner, K.E. Sloan, M.T. Bohnsack, Human METTL16 is a N(6)-methyladenosine (m(6)A) methyltransferase that targets pre-mRNAs and various non-coding RNAs., *EMBO Rep.* 18 (2017) 2004–2014. <https://doi.org/10.15252/embr.201744940>.
31. R. Su, L. Dong, Y. Li, M. Gao, P.C. He, W. Liu, J. Wei, Z. Zhao, L. Gao, L. Han, X. Deng, C. Li, E. Prince, B. Tan, Y. Qing, X. Qin, C. Shen, M. Xue, K. Zhou, Z. Chen, J. Xue, W. Li, H. Qin, X. Wu, M. Sun, Y. Nam, C.-W. Chen, W. Huang, D. Horne, S.T. Rosen, C. He, J. Chen, METTL16 exerts an m(6)A-independent function to facilitate translation and tumorigenesis., *Nat. Cell Biol.* 24 (2022) 205–216. <https://doi.org/10.1038/s41556-021-00835-2>.
32. K.E. Pendleton, B. Chen, K. Liu, O. V Hunter, Y. Xie, B.P. Tu, N.K. Conrad, The U6 snRNA m(6)A Methyltransferase METTL16 Regulates SAM Synthetase Intron Retention., *Cell*. 169 (2017) 824–835.e14. <https://doi.org/10.1016/j.cell.2017.05.003>.
33. G. Jia, Y. Fu, X. Zhao, Q. Dai, G. Zheng, Y. Yang, C. Yi, T. Lindahl, T. Pan, Y.-G. Yang, C. He, N6-methyladenosine in nuclear RNA is a major substrate of the obesity-associated FTO., *Nat. Chem. Biol.* 7 (2011) 885–887. <https://doi.org/10.1038/nchembio.687>.
34. X. Zhao, Y. Yang, B.-F. Sun, Y. Shi, X. Yang, W. Xiao, Y.-J. Hao, X.-L. Ping, Y.-S. Chen, W.-J. Wang, K.-X. Jin, X. Wang, C.-M. Huang, Y. Fu, X.-M. Ge, S.-H. Song, H.S. Jeong, H. Yanagisawa, Y. Niu, G.-F. Jia, W. Wu, W.-M. Tong, A. Okamoto, C. He, J.M. Rendtlew Danielsen, X.-J. Wang, Y.-G. Yang, FTO-dependent demethylation of N6-methyladenosine regulates mRNA splicing and is required for adipogenesis., *Cell Res.* 24 (2014) 1403–1419. <https://doi.org/10.1038/cr.2014.151>.

35. P. Mathiyalagan, M. Adamiak, J. Mayourian, Y. Sassi, Y. Liang, N. Agarwal, D. Jha, S. Zhang, E. Kohlbrenner, E. Chepurko, J. Chen, M.G. Trivieri, R. Singh, R. Bouchareb, K. Fish, K. Ishikawa, D. Lebeche, R.J. Hajjar, S. Sahoo, FTO-Dependent N(6)-Methyladenosine Regulates Cardiac Function During Remodeling and Repair., *Circulation.* 139 (2019) 518–532. <https://doi.org/10.1161/CIRCULATIONAHA.118.033794>.

36. W. Wu, J. Feng, D. Jiang, X. Zhou, Q. Jiang, M. Cai, X. Wang, T. Shan, Y. Wang, AMPK regulates lipid accumulation in skeletal muscle cells through FTO-dependent demethylation of N(6)-methyladenosine., *Sci. Rep.* 7 (2017) 41606. <https://doi.org/10.1038/srep41606>.

37. R. Wu, Y. Liu, Y. Yao, Y. Zhao, Z. Bi, Q. Jiang, Q. Liu, M. Cai, F. Wang, Y. Wang, X. Wang, FTO regulates adipogenesis by controlling cell cycle progression via m(6)A-YTHDF2 dependent mechanism., *Biochim. Biophys. Acta. Mol. Cell Biol. Lipids.* 1863 (2018) 1323–1330. <https://doi.org/10.1016/j.bbalip.2018.08.008>.

38. J. Wei, F. Liu, Z. Lu, Q. Fei, Y. Ai, P.C. He, H. Shi, X. Cui, R. Su, A. Klungland, G. Jia, J. Chen, C. He, Differential m(6)A, m(6)A(m), and m(1)A Demethylation Mediated by FTO in the Cell Nucleus and Cytoplasm., *Mol. Cell.* 71 (2018) 973–985.e5. <https://doi.org/10.1016/j.molcel.2018.08.011>.

39. G. Jia, C.-G. Yang, S. Yang, X. Jian, C. Yi, Z. Zhou, C. He, Oxidative demethylation of 3-methylthymine and 3-methyluracil in single-stranded DNA and RNA by mouse and human FTO., *FEBS Lett.* 582 (2008) 3313–3319. <https://doi.org/10.1016/j.febslet.2008.08.019>.

40. P. Stoilov, I. Rafalska, S. Stamm, YTH: a new domain in nuclear proteins., *Trends Biochem. Sci.* 27 (2002) 495–497. [https://doi.org/10.1016/s0968-0004\(02\)02189-8](https://doi.org/10.1016/s0968-0004(02)02189-8).

41. C. Xu, X. Wang, K. Liu, I.A. Roundtree, W. Tempel, Y. Li, Z. Lu, C. He, J. Min, Structural basis for selective binding of m 6 A RNA by the YTHDC1 YTH domain, *Nat. Chem. Biol.* 10 (2014) 927–929.

42. M.N. Wojtas, R.R. Pandey, M. Mendel, D. Homolka, R. Sachidanandam, R.S. Pillai, Regulation of m(6)A Transcripts by the 3'→5' RNA Helicase YTHDC2 Is Essential for a Successful Meiotic Program in the Mammalian Germline., *Mol. Cell.* 68 (2017) 374–387.e12. <https://doi.org/10.1016/j.molcel.2017.09.021>.

43. H. Du, Y. Zhao, J. He, Y. Zhang, H. Xi, M. Liu, J. Ma, L. Wu, YTHDF2 destabilizes m(6)A-containing RNA through direct recruitment of the CCR4-NOT deadenylase complex., *Nat. Commun.* 7 (2016) 12626. <https://doi.org/10.1038/ncomms12626>.

44. A. Li, Y.-S. Chen, X.-L. Ping, X. Yang, W. Xiao, Y. Yang, H.-Y. Sun, Q. Zhu, P. Baidya, X. Wang, D.P. Bhattarai, Y.-L. Zhao, B.-F. Sun, Y.-G. Yang, Cytoplasmic m(6)A reader YTHDF3 promotes mRNA translation., *Cell Res.* 27 (2017) 444–447. <https://doi.org/10.1038/cr.2017.10>.

45. H. Shi, X. Wang, Z. Lu, B.S. Zhao, H. Ma, P.J. Hsu, C. Liu, C. He, YTHDF3 facilitates translation and decay of N(6)-methyladenosine-modified RNA., *Cell Res.* 27 (2017) 315–328. <https://doi.org/10.1038/cr.2017.15>.

46. I.A. Roundtree, G.-Z. Luo, Z. Zhang, X. Wang, T. Zhou, Y. Cui, J. Sha, X. Huang, L. Guerrero, P. Xie, E. He, B. Shen, C. He, YTHDC1 mediates nuclear export of N(6)-methyladenosine methylated mRNAs., *Elife.* 6 (2017). <https://doi.org/10.7554/elife.31311>.

47. Y. Mao, L. Dong, X.-M. Liu, J. Guo, H. Ma, B. Shen, S.-B. Qian, m(6)A in mRNA coding regions promotes translation via the RNA helicase-containing YTHDC2., *Nat. Commun.* 10 (2019) 5332. <https://doi.org/10.1038/s41467-019-13317-9>.

48. A. McCloskey, I. Taniguchi, K. Shinmyozu, M. Ohno, hnRNP C tetramer measures RNA length to classify RNA polymerase II transcripts for export., *Science.* 335 (2012) 1643–1646. <https://doi.org/10.1126/science.1218469>.

49. N. Liu, K.I. Zhou, M. Parisien, Q. Dai, L. Diatchenko, T. Pan, N6-methyladenosine alters RNA structure to regulate binding of a low-complexity protein., *Nucleic Acids Res.* 45 (2017) 6051–6063. <https://doi.org/10.1093/nar/gkx141>.

50. C.R. Alarcón, H. Goodarzi, H. Lee, X. Liu, S. Tavazoie, S.F. Tavazoie, HNRNPA2B1 Is a Mediator of m(6)A-Dependent Nuclear RNA Processing Events., *Cell.* 162 (2015) 1299–1308. <https://doi.org/10.1016/j.cell.2015.08.011>.

51. J.L. Bell, K. Wächter, B. Mühleck, N. Pazaitis, M. Köhn, M. Lederer, S. Hüttelmaier, Insulin-like growth factor 2 mRNA-binding proteins (IGF2BPs): post-transcriptional drivers of cancer progression?, *Cell. Mol. Life Sci.* 70 (2013) 2657–2675. <https://doi.org/10.1007/s00018-012-1186-z>.

52. T. Schneider, L.-H. Hung, M. Aziz, A. Wilmen, S. Thaum, J. Wagner, R. Janowski, S. Müller, S. Schreiner, P. Friedhoff, S. Hüttelmaier, D. Niessing, M. Sattler, A. Schlundt, A. Bindereif, Combinatorial recognition of clustered RNA elements by the multidomain RNA-binding protein IMP3., *Nat. Commun.* 10 (2019) 2266. <https://doi.org/10.1038/s41467-019-09769-8>.

53. H. Huang, H. Weng, W. Sun, X. Qin, H. Shi, H. Wu, B.S. Zhao, A. Mesquita, C. Liu, C.L. Yuan, Y.-C. Hu, S. Hüttelmaier, J.R. Skibbe, R. Su, X. Deng, L. Dong, M. Sun, C. Li, S. Nachtergale, Y. Wang, C. Hu, K. Ferchen, K.D. Greis, X. Jiang, M. Wei, L. Qu, J.-L. Guan, C. He, J. Yang, J. Chen, Recognition of RNA N(6)-methyladenosine by IGF2BP proteins enhances mRNA stability and translation., *Nat. Cell Biol.* 20 (2018) 285–295. <https://doi.org/10.1038/s41556-018-0045-z>.

54. O. Manners, B. Baquero-Perez, A. Whitehouse, m6A: widespread regulatory control in virus replication, *Biochim. Biophys. Acta (BBA)-Gene Regul. Mech.* 1862 (2019) 370–381.

55. D. Walsh, J. McCarthy, C. O'Driscoll, S. Melgar, Pattern recognition receptors--molecular orchestrators of inflammation in inflammatory bowel disease., *Cytokine Growth Factor Rev.* 24 (2013) 91–104. <https://doi.org/10.1016/j.cytofr.2012.09.003>.

56. D. Eletto, F. Mentucci, A. Voli, A. Petrella, A. Porta, A. Tosco, *Helicobacter pylori Pathogen-Associated Molecular Patterns: Friends or Foes?*, *Int. J. Mol. Sci.* 23 (2022). <https://doi.org/10.3390/ijms23073531>.

57. J.S. Roh, D.H. Sohn, *Damage-Associated Molecular Patterns in Inflammatory Diseases.*, *Immune Netw.* 18 (2018) e27. <https://doi.org/10.4110/in.2018.18.e27>.

58. J. Rehwinkel, M.U. Gack, *RIG-I-like receptors: their regulation and roles in RNA sensing.*, *Nat. Rev. Immunol.* 20 (2020) 537–551. <https://doi.org/10.1038/s41577-020-0288-3>.

59. N.A. Lind, V.E. Rael, K. Pestal, B. Liu, G.M. Barton, *Regulation of the nucleic acid-sensing Toll-like receptors*, *Nat. Rev. Immunol.* 22 (2022) 224–235. <https://doi.org/10.1038/s41577-021-00577-0>.

60. A. Decout, J.D. Katz, S. Venkatraman, A. Ablasser, *The cGAS-STING pathway as a therapeutic target in inflammatory diseases.*, *Nat. Rev. Immunol.* 21 (2021) 548–569. <https://doi.org/10.1038/s41577-021-00524-z>.

61. M.J. McFadden, N.S. Gokhale, S.M. Horner, *Protect this house: cytosolic sensing of viruses.*, *Curr. Opin. Virol.* 22 (2017) 36–43. <https://doi.org/10.1016/j.coviro.2016.11.012>.

62. A. Dempsey, A.G. Bowie, *Innate immune recognition of DNA: A recent history.*, *Virology.* 479–480 (2015) 146–152. <https://doi.org/10.1016/j.virol.2015.03.013>.

63. O. Takeuchi, S. Akira, *Pattern recognition receptors and inflammation.*, *Cell.* 140 (2010) 805–820. <https://doi.org/10.1016/j.cell.2010.01.022>.

64. M. Yoneyama, M. Kikuchi, K. Matsumoto, T. Imaizumi, M. Miyagishi, K. Taira, E. Foy, Y.-M. Loo, M.J. Gale, S. Akira, S. Yonehara, A. Kato, T. Fujita, *Shared and unique functions of the DExD/H-box helicases RIG-I, MDA5, and LGP2 in antiviral innate immunity.*, *J. Immunol.* 175 (2005) 2851–2858. <https://doi.org/10.4049/jimmunol.175.5.2851>.

65. D. Luo, A. Kohlway, A.M. Pyle, *Duplex RNA activated ATPases (DRAs): platforms for RNA sensing, signaling and processing.*, *RNA Biol.* 10 (2013) 111–120. <https://doi.org/10.4161/rna.22706>.

66. D. Goubau, M. Schlee, S. Deddouche, A.J. Pruijssers, T. Zillinger, M. Goldeck, C. Schubert, A.G. Van der Veen, T. Fujimura, J. Rehwinkel, J.A. Iskarpatyoti, W. Barchet, J. Ludwig, T.S. Dermody, G. Hartmann, C. Reis e Sousa, *Antiviral immunity via RIG-I-mediated recognition of RNA bearing 5'-diphosphates.*, *Nature.* 514 (2014) 372–375. <https://doi.org/10.1038/nature13590>.

67. Q. Feng, S. V Hato, M.A. Langereis, J. Zoll, R. Virgen-Slane, A. Peisley, S. Hur, B.L. Semler, R.P. van Rij, F.J.M. van Kuppeveld, *MDA5 detects the double-stranded RNA replicative form in picornavirus-infected cells.*, *Cell Rep.* 2 (2012) 1187–1196. <https://doi.org/10.1016/j.celrep.2012.10.005>.

68. Z. Zhu, M. Zhang, L. Yuan, Y. Xu, H. Zhou, Z. Lian, P. Liu, X. Li, *LGP2 Promotes Type I Interferon Production To Inhibit PRRSV Infection via Enhancing MDA5-Mediated Signaling.*, *J. Virol.* 97 (2023) e0184322. <https://doi.org/10.1128/jvi.01843-22>.

69. J.-P. Parisien, J.J. Lenoir, R. Mandhana, K.R. Rodriguez, K. Qian, A.M. Bruns, C.M. Horvath, *RNA sensor LGP2 inhibits TRAF ubiquitin ligase to negatively regulate innate immune signaling.*, *EMBO Rep.* 19 (2018). <https://doi.org/10.15252/embr.201745176>.

70. D. Luo, A. Kohlway, A. Vela, A.M. Pyle, *Visualizing the determinants of viral RNA recognition by innate immune sensor RIG-I.*, *Structure.* 20 (2012) 1983–1988. <https://doi.org/10.1016/j.str.2012.08.029>.

71. F. Civril, M. Bennett, M. Moldt, T. Deimling, G. Witte, S. Schiesser, T. Carell, K.-P. Hopfner, *The RIG-I ATPase domain structure reveals insights into ATP-dependent antiviral signalling.*, *EMBO Rep.* 12 (2011) 1127–1134. <https://doi.org/10.1038/embor.2011.190>.

72. F. Hou, L. Sun, H. Zheng, B. Skaug, Q.-X. Jiang, Z.J. Chen, *MAVS forms functional prion-like aggregates to activate and propagate antiviral innate immune response.*, *Cell.* 146 (2011) 448–461. <https://doi.org/10.1016/j.cell.2011.06.041>.

73. R.B. Seth, L. Sun, C.-K. Ea, Z.J. Chen, *Identification and characterization of MAVS, a mitochondrial antiviral signaling protein that activates NF- $\kappa$ B and IRF 3.*, *Cell.* 122 (2005) 669–682. <https://doi.org/10.1016/j.cell.2005.08.012>.

74. E. Dixit, S. Boulant, Y. Zhang, A.S.Y. Lee, C. Odendall, B. Shum, N. Hacohen, Z.J. Chen, S.P. Whelan, M. Fransen, M.L. Nibert, G. Superti-Furga, J.C. Kagan, *Peroxisomes are signaling platforms for antiviral innate immunity.*, *Cell.* 141 (2010) 668–681. <https://doi.org/10.1016/j.cell.2010.04.018>.

75. K. Tabeta, K. Hoebe, E.M. Janssen, X. Du, P. Georgel, K. Crozat, S. Mudd, N. Mann, S. Sovath, J. Goode, L. Shamel, A.A. Herskovits, D.A. Portnoy, M. Cooke, L.M. Tarantino, T. Wiltshire, B.E. Steinberg, S. Grinstein, B. Beutler, *The Unc93b1 mutation 3d disrupts exogenous antigen presentation and signaling via Toll-like receptors 3, 7 and 9.*, *Nat. Immunol.* 7 (2006) 156–164. <https://doi.org/10.1038/ni1297>.

76. I. Botos, D.M. Segal, D.R. Davies, *The structural biology of Toll-like receptors.*, *Structure.* 19 (2011) 447–459. <https://doi.org/10.1016/j.str.2011.02.004>.

77. T. Kawai, S. Akira, *The role of pattern-recognition receptors in innate immunity: update on Toll-like receptors.*, *Nat. Immunol.* 11 (2010) 373–384. <https://doi.org/10.1038/ni.1863>.

78. H.-C. Lee, K. Chathuranga, J.-S. Lee, Intracellular sensing of viral genomes and viral evasion., *Exp. Mol. Med.* 51 (2019) 1–13. <https://doi.org/10.1038/s12276-019-0299-y>.

79. L. Sun, J. Wu, F. Du, X. Chen, Z.J. Chen, Cyclic GMP-AMP synthase is a cytosolic DNA sensor that activates the type I interferon pathway., *Science*. 339 (2013) 786–791. <https://doi.org/10.1126/science.1232458>.

80. A. Ablasser, M. Goldeck, T. Cavlar, T. Deimling, G. Witte, I. Röhl, K.-P. Hopfner, J. Ludwig, V. Hornung, cGAS produces a 2'-5'-linked cyclic dinucleotide second messenger that activates STING., *Nature*. 498 (2013) 380–384. <https://doi.org/10.1038/nature12306>.

81. V. Dell’Oste, D. Gatti, A.G. Giorgio, M. Gariglio, S. Landolfo, M. De Andrea, The interferon-inducible DNA-sensor protein IFI16: a key player in the antiviral response., *New Microbiol.* 38 (2015) 5–20.

82. V. Hornung, A. Ablasser, M. Charrel-Dennis, F. Bauernfeind, G. Horvath, D.R. Caffrey, E. Latz, K.A. Fitzgerald, AIM2 recognizes cytosolic dsDNA and forms a caspase-1-activating inflammasome with ASC., *Nature*. 458 (2009) 514–518. <https://doi.org/10.1038/nature07725>.

83. Z. Zhang, B. Yuan, M. Bao, N. Lu, T. Kim, Y.-J. Liu, The helicase DDX41 senses intracellular DNA mediated by the adaptor STING in dendritic cells., *Nat. Immunol.* 12 (2011) 959–965. <https://doi.org/10.1038/ni.2091>.

84. Y.-H. Chiu, J.B. Macmillan, Z.J. Chen, RNA polymerase III detects cytosolic DNA and induces type I interferons through the RIG-I pathway., *Cell*. 138 (2009) 576–591. <https://doi.org/10.1016/j.cell.2009.06.015>.

85. A. Dalet, E. Gatti, P. Pierre, Integration of PKR-dependent translation inhibition with innate immunity is required for a coordinated anti-viral response., *FEBS Lett.* 589 (2015) 1539–1545. <https://doi.org/10.1016/j.febslet.2015.05.006>.

86. V. Hornung, R. Hartmann, A. Ablasser, K.-P. Hopfner, OAS proteins and cGAS: unifying concepts in sensing and responding to cytosolic nucleic acids., *Nat. Rev. Immunol.* 14 (2014) 521–528. <https://doi.org/10.1038/nri3719>.

87. C. Shu, G. Yi, T. Watts, C.C. Kao, P. Li, Structure of STING bound to cyclic di-GMP reveals the mechanism of cyclic dinucleotide recognition by the immune system., *Nat. Struct. Mol. Biol.* 19 (2012) 722–724. <https://doi.org/10.1038/nsmb.2331>.

88. G. Shang, C. Zhang, Z.J. Chen, X.-C. Bai, X. Zhang, Cryo-EM structures of STING reveal its mechanism of activation by cyclic GMP-AMP., *Nature*. 567 (2019) 389–393. <https://doi.org/10.1038/s41586-019-0998-5>.

89. C. Zhang, G. Shang, X. Gui, X. Zhang, X.-C. Bai, Z.J. Chen, Structural basis of STING binding with and phosphorylation by TBK1., *Nature*. 567 (2019) 394–398. <https://doi.org/10.1038/s41586-019-1000-2>.

90. S. Liu, X. Cai, J. Wu, Q. Cong, X. Chen, T. Li, F. Du, J. Ren, Y.-T. Wu, N. V Grishin, Phosphorylation of innate immune adaptor proteins MAVS, STING, and TRIF induces IRF3 activation, *Science* (80-. ). 347 (2015).

91. A. Roy, A. Ghosh, B. Kumar, B. Chandran, IFI16, a nuclear innate immune DNA sensor, mediates epigenetic silencing of herpesvirus genomes by its association with H3K9 methyltransferases SUV39H1 and GLP., *eLife*. 8 (2019). <https://doi.org/10.7554/eLife.49500>.

92. M.H. Orzalli, N.M. Broekema, B.A. Diner, D.C. Hancks, N.C. Elde, I.M. Cristea, D.M. Knipe, cGAS-mediated stabilization of IFI16 promotes innate signaling during herpes simplex virus infection., *Proc. Natl. Acad. Sci. U. S. A.* 112 (2015) E1773–81. <https://doi.org/10.1073/pnas.1424637112>.

93. L. Wang, M. Wen, X. Cao, Nuclear hnRNPA2B1 initiates and amplifies the innate immune response to DNA viruses., *Science*. 365 (2019) eaav0758. <https://doi.org/10.1126/science.aav0758>.

94. S. Carpenter, D. Aiello, M.K. Atianand, E.P. Ricci, P. Gandhi, L.L. Hall, M. Byron, B. Monks, M. Henry-Bezy, J.B. Lawrence, L.A.J. O’Neill, M.J. Moore, D.R. Caffrey, K.A. Fitzgerald, A long noncoding RNA mediates both activation and repression of immune response genes., *Science*. 341 (2013) 789–792. <https://doi.org/10.1126/science.1240925>.

95. M. Gentili, X. Lahaye, F. Nadalin, G.P.F. Nader, E. Puig Lombardi, S. Herve, N.S. De Silva, D.C. Rookhuizen, E. Zueva, C. Goudot, M. Maurin, A. Bochnakian, S. Amigorena, M. Piel, D. Fachinetti, A. Londoño-Vallejo, N. Manel, The N-Terminal Domain of cGAS Determines Preferential Association with Centromeric DNA and Innate Immune Activation in the Nucleus., *Cell Rep.* 26 (2019) 2377–2393.e13. <https://doi.org/10.1016/j.celrep.2019.01.105>.

96. S. Cui, Q. Yu, L. Chu, Y. Cui, M. Ding, Q. Wang, H. Wang, Y. Chen, X. Liu, C. Wang, Nuclear cGAS Functions Non-canonically to Enhance Antiviral Immunity via Recruiting Methyltransferase Prmt5., *Cell Rep.* 33 (2020) 108490. <https://doi.org/10.1016/j.celrep.2020.108490>.

97. J.L. Justice, M.A. Kennedy, J.E. Hutton, D. Liu, B. Song, B. Phelan, I.M. Cristea, Systematic profiling of protein complex dynamics reveals DNA-PK phosphorylation of IFI16 en route to herpesvirus immunity., *Sci. Adv.* 7 (2021). <https://doi.org/10.1126/sciadv.abg6680>.

98. P.E. Merkl, D.M. Knipe, Role for a Filamentous Nuclear Assembly of IFI16, DNA, and Host Factors in Restriction of Herpesviral Infection., *MBio*. 10 (2019). <https://doi.org/10.1128/mBio.02621-18>.

99. G. Dunphy, S.M. Flannery, J.F. Almine, D.J. Connolly, C. Paulus, K.L. Jönsson, M.R. Jakobsen, M.M. Nevels, A.G. Bowie, L. Unterholzner, Non-canonical Activation of the DNA Sensing Adaptor STING by ATM and IFI16 Mediates NF-κB Signaling after Nuclear DNA Damage., *Mol. Cell.* 71 (2018) 745–760.e5. <https://doi.org/10.1016/j.molcel.2018.07.034>.

100. M.S. Crow, I.M. Cristea, Human Antiviral Protein IFIX Suppresses Viral Gene Expression during Herpes Simplex Virus 1 (HSV-1) Infection and Is Counteracted by Virus-induced Proteasomal Degradation., *Mol. Cell. Proteomics.* 16 (2017) S200–S214. <https://doi.org/10.1074/mcp.M116.064741>.

101. L. Cao, S. Liu, Y. Li, G. Yang, Y. Luo, S. Li, H. Du, Y. Zhao, D. Wang, J. Chen, Z. Zhang, M. Li, S. Ouyang, X. Gao, Y. Sun, Z. Wang, L. Yang, R. Lin, P. Wang, F. You, The Nuclear Matrix Protein SAFA Surveils Viral RNA and Facilitates Immunity by Activating Antiviral Enhancers and Super-enhancers., *Cell Host Microbe.* 26 (2019) 369-384.e8. <https://doi.org/10.1016/j.chom.2019.08.010>.

102. X. Lahaye, M. Gentili, A. Silvin, C. Conrad, L. Picard, M. Jouve, E. Zueva, M. Maurin, F. Nadalin, G.J. Knott, B. Zhao, F. Du, M. Rio, J. Amiel, A.H. Fox, P. Li, L. Etienne, C.S. Bond, L. Colleaux, N. Manel, NONO Detects the Nuclear HIV Capsid to Promote cGAS-Mediated Innate Immune Activation., *Cell.* 175 (2018) 488-501.e22. <https://doi.org/10.1016/j.cell.2018.08.062>.

103. M. Morschikh, A. Cribier, R. Raffel, S. Amraoui, J. Cau, D. Severac, E. Dubois, O. Schwartz, Y. Bennasser, M. Benkirane, HEXIM1 and NEAT1 Long Non-coding RNA Form a Multi-subunit Complex that Regulates DNA-Mediated Innate Immune Response., *Mol. Cell.* 67 (2017) 387-399.e5. <https://doi.org/10.1016/j.molcel.2017.06.020>.

104. R.M. Rubio, D.P. Depledge, C. Bianco, L. Thompson, I. Mohr, RNA m(6)A modification enzymes shape innate responses to DNA by regulating interferon  $\beta$ , *Genes Dev.* 32 (2018) 1472–1484. <https://doi.org/10.1101/gad.319475.118>.

105. Q. Zheng, J. Hou, Y. Zhou, Z. Li, X. Cao, The RNA helicase DDX46 inhibits innate immunity by entrapping m(6)A-demethylated antiviral transcripts in the nucleus., *Nat. Immunol.* 18 (2017) 1094–1103. <https://doi.org/10.1038/ni.3830>.

106. C.R. Hesser, D. Walsh, YTHDF2 Is Downregulated in Response to Host Shutoff Induced by DNA Virus Infection and Regulates Interferon-Stimulated Gene Expression., *J. Virol.* 97 (2023) e0175822. <https://doi.org/10.1128/jvi.01758-22>.

107. F. Qin, B. Cai, J. Zhao, L. Zhang, Y. Zheng, B. Liu, C. Gao, Methyltransferase-Like Protein 14 Attenuates Mitochondrial Antiviral Signaling Protein Expression to Negatively Regulate Antiviral Immunity via N(6)-methyladenosine Modification., *Adv. Sci. (Weinheim, Baden-Wurttemberg, Ger.)* 8 (2021) e2100606. <https://doi.org/10.1002/advs.202100606>.

108. N. Li, H. Hui, B. Bray, G.M. Gonzalez, M. Zeller, K.G. Anderson, R. Knight, D. Smith, Y. Wang, A.F. Carlin, T.M. Rana, METTL3 regulates viral m6A RNA modification and host cell innate immune responses during SARS-CoV-2 infection., *Cell Rep.* 35 (2021) 109091. <https://doi.org/10.1016/j.celrep.2021.109091>.

109. W. Qiu, Q. Zhang, R. Zhang, Y. Lu, X. Wang, H. Tian, Y. Yang, Z. Gu, Y. Gao, X. Yang, G. Cui, B. Sun, Y. Peng, H. Deng, H. Peng, A. Yang, Y.-G. Yang, P. Yang, N(6)-methyladenosine RNA modification suppresses antiviral innate sensing pathways via reshaping double-stranded RNA., *Nat. Commun.* 12 (2021) 1582. <https://doi.org/10.1038/s41467-021-21904-y>.

110. Z. Feng, Q. Li, R. Meng, B. Yi, Q. Xu, METTL3 regulates alternative splicing of MyD88 upon the lipopolysaccharide-induced inflammatory response in human dental pulp cells., *J. Cell. Mol. Med.* 22 (2018) 2558–2568. <https://doi.org/10.1111/jcmm.13491>.

111. L. Yang, G. Wu, Q. Wu, L. Peng, L. Yuan, METTL3 overexpression aggravates LPS-induced cellular inflammation in mouse intestinal epithelial cells and DSS-induced IBD in mice., *Cell Death Discov.* 8 (2022) 62. <https://doi.org/10.1038/s41420-022-00849-1>.

112. K. Karikó, M. Buckstein, H. Ni, D. Weissman, Suppression of RNA recognition by Toll-like receptors: the impact of nucleoside modification and the evolutionary origin of RNA, *Immunity.* 23 (2005) 165–175.

113. J. Tong, X. Wang, Y. Liu, X. Ren, A. Wang, Z. Chen, J. Yao, K. Mao, T. Liu, F.-L. Meng, W. Pan, Q. Zou, J. Liu, Y. Zhou, Q. Xia, R.A. Flavell, S. Zhu, H.-B. Li, Pooled CRISPR screening identifies m(6)A as a positive regulator of macrophage activation., *Sci. Adv.* 7 (2021). <https://doi.org/10.1126/sciadv.abd4742>.

114. H. Wang, X. Hu, M. Huang, J. Liu, Y. Gu, L. Ma, Q. Zhou, X. Cao, Mettl3-mediated mRNA m6A methylation promotes dendritic cell activation, *Nat. Commun.* 10 (2019) 1898. <https://doi.org/10.1038/s41467-019-09903-6>.

115. S. Geng, W. Zheng, Y. Zhao, T. Xu, METTL3-Mediated m6A Modification of TRIF and MyD88 mRNAs Suppresses Innate Immunity in Teleost Fish, *Miichthys miiuy.*, *J. Immunol.* 211 (2023) 130–139. <https://doi.org/10.4049/jimmunol.2300033>.

116. M. Balzarolo, S. Engels, A.J. de Jong, K. Franke, T.K. van den Berg, M.F. Gulen, A. Ablasser, E.M. Janssen, B. van Steensel, M.C. Wolkers, m6A methylation potentiates cytosolic dsDNA recognition in a sequence-specific manner., *Open Biol.* 11 (2021) 210030. <https://doi.org/10.1098/rsob.210030>.

117. J. Song, L. Zhang, C. Li, M. Maimaiti, J. Sun, J. Hu, L. Li, X. Zhang, C. Wang, H. Hu, m(6)A-mediated modulation coupled with transcriptional regulation shapes long noncoding RNA repertoire of the cGAS-STING signaling., *Comput. Struct. Biotechnol. J.* 20 (2022) 1785–1797. <https://doi.org/10.1016/j.csbj.2022.04.002>.

118. Y. Ge, T. Ling, Y. Wang, X. Jia, X. Xie, R. Chen, S. Chen, S. Yuan, A. Xu, Degradation of WTAP blocks antiviral responses by reducing the m(6) A levels of IRF3 and IFNAR1 mRNA., *EMBO Rep.* 22 (2021) e52101. <https://doi.org/10.15252/embr.202052101>.

119. H. Imam, G.-W. Kim, S.A. Mir, M. Khan, A. Siddiqui, Interferon-stimulated gene 20 (ISG20) selectively degrades N6-methyladenosine modified Hepatitis B Virus transcripts., *PLoS Pathog.* 16 (2020) e1008338. <https://doi.org/10.1371/journal.ppat.1008338>.

120. G.-W. Kim, H. Imam, M. Khan, S.A. Mir, S.-J. Kim, S.K. Yoon, W. Hur, A. Siddiqui, HBV-Induced Increased N6 Methyladenosine Modification of PTEN RNA Affects Innate Immunity and Contributes to HCC., *Hepatology*. 73 (2021) 533–547. <https://doi.org/10.1002/hep.31313>.

121. Y. Zhang, X. Wang, X. Zhang, J. Wang, Y. Ma, L. Zhang, X. Cao, RNA-binding protein YTHDF3 suppresses interferon-dependent antiviral responses by promoting FOXO3 translation., *Proc. Natl. Acad. Sci. U. S. A.* 116 (2019) 976–981. <https://doi.org/10.1073/pnas.1812536116>.

122. N. Tirumuru, B.S. Zhao, W. Lu, Z. Lu, C. He, L. Wu, N 6-methyladenosine of HIV-1 RNA regulates viral infection and HIV-1 Gag protein expression, *Elife*. 5 (2016) e15528.

123. C.-L. Tsai, C.-Y. Pan, Y.-T. Tseng, F.-C. Chen, Y.-C. Chang, T.-C. Wang, Acute effects of high-intensity interval training and moderate-intensity continuous exercise on BDNF and irisin levels and neurocognitive performance in late middle-aged and older adults., *Behav. Brain Res.* 413 (2021) 113472. <https://doi.org/10.1016/j.bbr.2021.113472>.

124. S. Chen, S. Kumar, C.E. Espada, N. Tirumuru, M.P. Cahill, L. Hu, C. He, L. Wu, N 6-methyladenosine modification of HIV-1 RNA suppresses type-I interferon induction in differentiated monocytic cells and primary macrophages, *PLoS Pathog.* 17 (2021) e1009421.

125. D.G. Courtney, E.M. Kennedy, R.E. Dumm, H.P. Bogerd, K. Tsai, N.S. Heaton, B.R. Cullen, Epitranscriptomic Enhancement of Influenza A Virus Gene Expression and Replication., *Cell Host Microbe*. 22 (2017) 377–386.e5. <https://doi.org/10.1016/j.chom.2017.08.004>.

126. G. Lichinchi, B.S. Zhao, Y. Wu, Z. Lu, Y. Qin, C. He, T.M. Rana, Dynamics of Human and Viral RNA Methylation during Zika Virus Infection., *Cell Host Microbe*. 20 (2016) 666–673. <https://doi.org/10.1016/j.chom.2016.10.002>.

127. N.S. Gokhale, A.B.R. McIntyre, M.J. McFadden, A.E. Roder, E.M. Kennedy, J.A. Gandara, S.E. Hopcraft, K.M. Quicke, C. Vazquez, J. Willer, O.R. Ilkayeva, B.A. Law, C.L. Holley, M.A. Garcia-Blanco, M.J. Evans, M.S. Suthar, S.S. Bradrick, C.E. Mason, S.M. Horner, N6-Methyladenosine in Flaviviridae Viral RNA Genomes Regulates Infection., *Cell Host Microbe*. 20 (2016) 654–665. <https://doi.org/10.1016/j.chom.2016.09.015>.

128. G.-W. Kim, A. Siddiqui, The role of N6-methyladenosine modification in the life cycle and disease pathogenesis of hepatitis B and C viruses, *Exp. Mol. Med.* 53 (2021) 339–345. <https://doi.org/10.1038/s12276-021-00581-3>.

129. G.-W. Kim, H. Imam, M. Khan, A. Siddiqui, N (6)-Methyladenosine modification of hepatitis B and C viral RNAs attenuates host innate immunity via RIG-I signaling., *J. Biol. Chem.* 295 (2020) 13123–13133. <https://doi.org/10.1074/jbc.RA120.014260>.

130. M. Lu, M. Xue, H.-T. Wang, E.L. Kairis, S. Ahmad, J. Wei, Z. Zhang, Q. Liu, Y. Zhang, Y. Gao, D. Garcin, M.E. Peebles, A. Sharma, S. Hur, C. He, J. Li, Nonsegmented Negative-Sense RNA Viruses Utilize N(6)-Methyladenosine (m(6)A) as a Common Strategy To Evade Host Innate Immunity., *J. Virol.* 95 (2021). <https://doi.org/10.1128/JVI.01939-20>.

131. M. Xue, Y. Zhang, H. Wang, E.L. Kairis, M. Lu, S. Ahmad, Z. Attia, O. Harder, Z. Zhang, J. Wei, P. Chen, Y. Gao, M.E. Peebles, A. Sharma, P. Boyaka, C. He, S. Hur, S. Niewiesk, J. Li, Viral RNA N6-methyladenosine modification modulates both innate and adaptive immune responses of human respiratory syncytial virus., *PLoS Pathog.* 17 (2021) e1010142. <https://doi.org/10.1371/journal.ppat.1010142>.

132. J. Liu, Y.-P. Xu, K. Li, Q. Ye, H.-Y. Zhou, H. Sun, X. Li, L. Yu, Y.-Q. Deng, R.-T. Li, M.-L. Cheng, B. He, J. Zhou, X.-F. Li, A. Wu, C. Yi, C.-F. Qin, The m(6)A methylome of SARS-CoV-2 in host cells., *Cell Res.* (2021). <https://doi.org/10.1038/s41422-020-00465-7>.

133. H.M. Burgess, D.P. Depledge, L. Thompson, K.P. Srinivas, R.C. Grande, E.I. Vink, J.S. Abebe, W.P. Blackaby, A. Hendrick, M.R. Albertella, T. Kouzarides, K.A. Stapleford, A.C. Wilson, I. Mohr, Targeting the m(6)A RNA modification pathway blocks SARS-CoV-2 and HCoV-OC43 replication., *Genes Dev.* 35 (2021) 1005–1019. <https://doi.org/10.1101/gad.348320.121>.

134. K. Liu, Y.-Z. Zhang, H. Yin, L.-L. Yu, J.-J. Cui, J.-Y. Yin, C.-H. Luo, C.-X. Guo, Identification of SARS-CoV-2 m6A modification sites correlate with viral pathogenicity., *Microbes Infect.* 26 (2024) 105228. <https://doi.org/10.1016/j.micinf.2023.105228>.

135. H. Imam, M. Khan, N.S. Gokhale, A.B.R. McIntyre, G.-W. Kim, J.Y. Jang, S.-J. Kim, C.E. Mason, S.M. Horner, A. Siddiqui, N6-methyladenosine modification of hepatitis B virus RNA differentially regulates the viral life cycle., *Proc. Natl. Acad. Sci. U. S. A.* 115 (2018) 8829–8834. <https://doi.org/10.1073/pnas.1808319115>.

136. T. Murata, S. Iwahori, Y. Okuno, H. Nishitsuji, Y. Yanagi, K. Watashi, T. Wakita, H. Kimura, K. Shimotohno, N6-methyladenosine Modification of Hepatitis B Virus RNA in the Coding Region of HBx., *Int. J. Mol. Sci.* 24 (2023). <https://doi.org/10.3390/ijms24032265>.

137. G.-W. Kim, H. Imam, A. Siddiqui, The RNA Binding Proteins YTHDC1 and FMRP Regulate the Nuclear Export of N(6)-Methyladenosine-Modified Hepatitis B Virus Transcripts and Affect the Viral Life Cycle., *J. Virol.* 95 (2021) e0009721. <https://doi.org/10.1128/JVI.00097-21>.

138. A. Kostyusheva, S. Brezgin, D. Glebe, D. Kostyushev, V. Chulanov, Host-cell interactions in HBV infection and pathogenesis: the emerging role of m6A modification., *Emerg. Microbes Infect.* 10 (2021) 2264–2275. <https://doi.org/10.1080/22221751.2021.2006580>.

139. Z. Zhang, W. Gao, Z. Liu, S. Yu, H. Jian, Z. Hou, P. Zeng, Comprehensive analysis of m6A regulators associated with immune infiltration in Hepatitis B virus-related hepatocellular carcinoma., *BMC Gastroenterol.* 23 (2023) 259. <https://doi.org/10.1186/s12876-023-02873-6>.

140. A.M. Price, K.E. Hayer, A.B.R. McIntyre, N.S. Gokhale, J.S. Abebe, A.N. Della Fera, C.E. Mason, S.M. Horner, A.C. Wilson, D.P. Depledge, M.D. Weitzman, Direct RNA sequencing reveals m(6)A modifications on adenovirus RNA are necessary for efficient splicing., *Nat. Commun.* 11 (2020) 6016. <https://doi.org/10.1038/s41467-020-19787-6>.

141. Z. Hajikhezri, Y. Kaira, E. Schubert, M. Darweesh, C. Svensson, G. Akusjärvi, T. Punga, Fragile X-Related Protein FXR1 Controls Human Adenovirus Capsid mRNA Metabolism., *J. Virol.* 97 (2023) e0153922. <https://doi.org/10.1128/jvi.01539-22>.

142. Z. Feng, F. Zhou, M. Tan, T. Wang, Y. Chen, W. Xu, B. Li, X. Wang, X. Deng, M.-L. He, Targeting m6A modification inhibits herpes virus 1 infection., *Genes Dis.* (2021). <https://doi.org/https://doi.org/10.1016/j.gendis.2021.02.004>.

143. R.J.J. Jansens, A. Olarerin-George, R. Verhamme, A. Mirza, S. Jaffrey, H.W. Favoreel, Alphaherpesvirus-mediated remodeling of the cellular transcriptome results in depletion of m6A-containing transcripts., *IScience.* 26 (2023) 107310. <https://doi.org/10.1016/j.isci.2023.107310>.

144. W. Wang, W. Ye, S. Chen, Y. Tang, D. Chen, Y. Lu, Z. Wu, Z. Huang, Y. Ge, METTL3-mediated m(6)A RNA modification promotes corneal neovascularization by upregulating the canonical Wnt pathway during HSV-1 infection., *Cell. Signal.* 109 (2023) 110784. <https://doi.org/10.1016/j.cellsig.2023.110784>.

145. J. Xu, Y. Qi, Q. Ju, Promotion of the resistance of human oral epithelial cells to herpes simplex virus type I infection via N6-methyladenosine modification., *BMC Oral Health.* 23 (2023) 121. <https://doi.org/10.1186/s12903-023-02744-2>.

146. K. Walther, L.N. Schulte, The role of lncRNAs in innate immunity and inflammation., *RNA Biol.* 18 (2021) 587–603. <https://doi.org/10.1080/15476286.2020.1845505>.

**Disclaimer/Publisher's Note:** The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.