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Article

Solid Phase Oligo-DNA Extraction from Complex Medium Using an Aminated Graphene/Nitrocellulose Membrane Hybrid

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Abstract: A hybrid material, consisting of commercially available nitrocellulose (NC) membrane non-covalently modified with amino-polyethylene glycol functionalized reduced graphene oxide (NH₂-PEG-rGO) nanoparticles, was successfully synthesized for oligonucleotide extraction. Fourier Transform Infrared Spectroscopy (FTIR) confirmed the modification of the NC membrane, revealing characteristic peaks of both compounds NC and NH₂-PEG-rGO. Scanning Electron Microscopy (SEM) exhibited morphological changes in the NC/NH₂-PEG-rGO hybrid membrane, marked by the introduction of NH₂-PEG-rGO particles, resulting in a distinctly smothered surface compared to the porous surface of the NC control membrane. Wettability assays revealed hydrophobic behavior for the NC/NH₂-PEG-rGO hybrid membrane, with a water contact angle exceeding 90°, contrasting with the hydrophilic behavior characterized by a 16.7° contact angle in the NC membrane. The performance of the NC/NH₂-PEG-rGO hybrid membrane was evaluated for the extraction of ssDNA with fewer than 50 nucleotides from solutions containing various ionic species (MnCl₂, MgCl₂, and MnCl₂/MgCl₂). The NC/NH₂-PEG-rGO hybrid membrane exhibited optimal performance when incubated in MgCl₂, presenting the highest fluorescence emission at 525 relative fluorescence units (r.f.u.). This corresponds to the extraction of approximately 610 pg (≈13%) of the total oligo-DNA, underscoring the efficacy of the pristine material, which extracts 286 pg (≈6%) of oligo-DNA in complex solutions.

Keywords: nitrocellulose; fluorescence quenching; DNA extraction; aminated graphene; ions

1. Introduction

In molecular biology and medical research, single-stranded deoxyribonucleic acid (ssDNA) is a fundamental component, playing an essential role in various cellular processes and disease detection [1,2]. Extracting and purifying nucleic acids (DNA and RNA) with precision and efficiency is of paramount importance as it serves as the foundation for numerous applications in diagnostics, genetic testing, gene expression analysis, and therapeutic interventions. As the demand for accurate and reliable genetic information increases, there is an arising need to develop advanced and reliable extraction methods for ssDNA. Among the various methods used for isolating nucleic acids, such as alkaline lysis, selective hybridization, and ethanol precipitation, the magnetic beads (MB) extraction technique stands out for its non-use of filters and time-efficient procedure [3–5]. During the conventional MB technique, an outer magnetic field is utilized to attract and enclose the ssDNA-

capturing particles near the tube's margin, but there is a risk of MB contamination in the eluted sample upon the addition of elution buffer when the magnetic force is turned off [4,6,7].

To address this, a solid-phase extraction method was introduced that is based on nitrocellulose (NC) membranes and offers an alternative method for enhancing the surface adsorption of ssDNA. NC membranes are known for their flexibility, high binding capacity of up to 80 $\mu\text{g}/\text{cm}^2$, as well as their stable physical and chemical properties. Upon application, ssDNA adheres to the NC membrane through a combination of van der Waals forces, electrostatic interactions, and hydrogen bonding, and can also be covalently linked via chemical cross-linkers or selectively hybridized through sequence-complementary probes. Further developments of the NC membrane extraction method include the modification of NC membranes with graphene-based materials, renowned for their ability to immobilize low concentrations of nucleic acids. [8–12].

In our prior study, the combination of NC membranes modified with graphene oxide (GO) nanoparticles was utilized as a solid phase for oligonucleotide extraction. This approach demonstrated favorable outcomes after a 60-minute incubation in αMEM , yielding a fluorescent signal equivalent to 7% (330-370 pg) of the total ssDNA, prompting the investigation to extend to other graphene variants [13]. Among the well-known graphene derivatives like pristine graphene, GO, reduced graphene oxide (rGO), graphene nanoplatelets, and graphane, there is another derivative called amino-polyethylene glycol functionalized reduced graphene oxide ($\text{NH}_2\text{-PEG-rGO}$) [14–18]. $\text{NH}_2\text{-PEG-rGO}$ serves as support material for DNA immobilization in various approaches, such as field effect transistors (FET) [19,20], electrochemical [21,22] and fluorescent based biosensors [23–25]. Among these options, fluorescent biosensors have captured the attention of researchers due to their distinctive characteristics, which encompass selectivity, sensitivity, simplicity, cost-effectiveness, and rapid response [26–28]. In a previous study, our group utilized $\text{NH}_2\text{-PEG-rGO}$ as a quencher due to its capability to function as an electron acceptor, which suppresses the fluorescence of the marked ssDNA through the process of fluorescence resonance energy transfer (FRET) [29]. The phenomenon in question involves the transfer of excited-state energy from the donor FAM-ssDNA molecule to the acceptor $\text{NH}_2\text{-PEG-rGO}$ molecule. This transfer primarily occurs through a combination of surface adsorption and electron transfer, resulting in the quenching of the FAM-ssDNA sequence, also known as the 'turn-off' effect [30–32].

Furthermore, the versatile nature of $\text{NH}_2\text{-PEG-rGO}$ extends beyond its role as a support material for ssDNA immobilization, encompassing the facile adsorption of ssDNA on the rGO lattice, which involves both covalent and non-covalent interactions, with a particular emphasis on non-covalent π -stacking interactions as the primary pathway. Conversely, double-stranded (ds) DNA exhibits a weaker binding affinity to the rGO surface compared to ssDNA, primarily because of the electrostatic repulsion from the phosphate backbone [33–36].

Building upon these findings, the current study aims to advance the investigation of ssDNA immobilization by substituting GO with $\text{NH}_2\text{-PEG-rGO}$ as the surface modification agent for NC membranes. The NC membrane modification with $\text{NH}_2\text{-PEG-rGO}$ was confirmed through Scanning Electron Microscopy (SEM), wettability assays, and Fourier Transform Infrared Spectroscopy (FTIR).

The NC/ $\text{NH}_2\text{-PEG-rGO}$ hybrid membrane obtained underwent further testing for the adsorption of FAM-ssDNA from three distinct complex solutions. These tests were conducted using αMEM as the base medium with an incubation time of 60 minutes. Since ionic concentration plays a substantial role in establishing the attachment of ssDNA, these solutions included one with MgCl_2 , another with MnCl_2 , and a third one containing a combination of MgCl_2 and MnCl_2 . Following the binding process, the membrane was washed with distilled water and then incubated in a desorption solution. This step facilitated the separation of FAM-ssDNA from the membrane, and improved affinity for extracting ssDNA.

2. Results and Discussion

2.1. Analysis of NC/NH₂-PEG-rGO Hybrid Membrane

2.1.1. FTIR Analysis

Through the molecular vibrations associated with each band, infrared spectroscopy imparts insight into the structures of the NC membrane modified with NH₂-PEG-rGO investigated in the present study. The FTIR spectra for NH₂-PEG-rGO, NC, and NC/NH₂-PEG-rGO hybrid membranes are depicted in Figure 1.

Usually, the infrared spectrum of NH₂-PEG-rGO is characterized by absorption peaks that are commonly found in the 1050-1085 cm⁻¹, 1650-1540 cm⁻¹, and 2800-3100 cm⁻¹ sections [37,38]. These spectral bands are attributed to different vibrational modes, corresponding to C-O-C (epoxy groups), N-H (amine), and C-H (ethylene groups). Furthermore, the spectral band at approximately 3300 cm⁻¹ indicates the stretching vibrations of O-H (hydroxyl), in their free and unbound states [38–41]. In our particular situation, the mentioned absorption bands are displayed at 1066, 1558, 2852, and 2924 cm⁻¹, as well as within the spectrum spanning from 3100 to 3400 cm⁻¹. The FTIR spectrum of NC exhibits three main absorption peaks situated at 1648, 1279, and 839 cm⁻¹, conceding to both symmetric and asymmetric stretching of the NO₂ group [42–44].

The FTIR analysis of the NC/NH₂-PEG-rGO hybrid membrane displays the characteristic absorption bands associated with both NH₂-PEG-rGO and NC, confirming the successful fabrication the NC/NH₂-PEG-rGO hybrid membrane. The attendance of NH₂-PEG-rGO is discernible through the faint absorption bands displayed at approximately 3300 cm⁻¹, which are linked to the O-H groups within the rGO structure, as well as the C-H bending and stretching vibrations in the range of 2800-3100 cm⁻¹, corresponding to the ethylene groups within the PEG structure [45,46]. Furthermore, the intensity of the distinctive absorption band corresponding to amino groups in the range of 1540-1600 cm⁻¹ (N-H) appears reduced, likely due to the presence of NO₂ functional groups in their proximity [37,47].

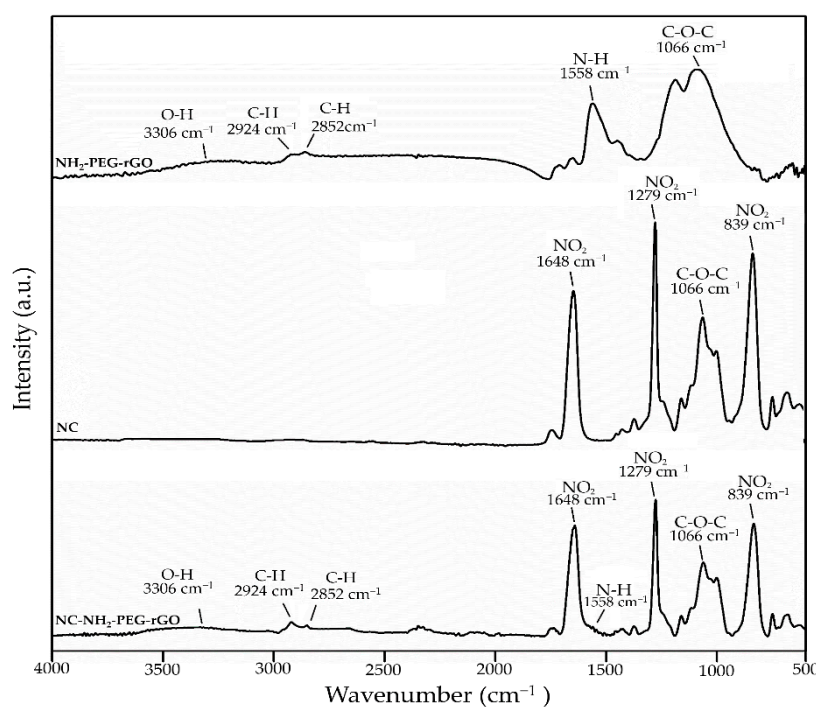


Figure 1. FTIR spectrum of NH₂-PEG-rGO, NC, and NC/NH₂-PEG-rGO hybrid membrane.

2.1.2. Morphological Characterization

Before and after the adsorption procedure, morphological assessments were carried out on both, the NC and NC/NH₂-PEG-rGO hybrid membranes.

Prior to the experiment, the SEM micrograph from Figure 2A shows the NC membranes, which possessed a sponge-like structure with high porosity and interconnected open pores, characterized by surface irregularities. The observed homogeneity is consistent with the inherent porous nature of nitrocellulose, providing a baseline representation of the pristine membrane. Figure 2B captures the morphological changes in the NC membrane following the experimental incubation in the used media, i.e. aMEM, sodium dodecyl sulfate (SDS), bovine serum albumin (BSA), FAM-ssDNA, and various ionic particles (MnCl₂, MgCl₂, and MnCl₂/MgCl₂). The flattening of the NC membrane surface indicates moderate stability in aqueous media, leading to a more uniform surface morphology suggesting the efficient immobilization of FAM-ssDNA.

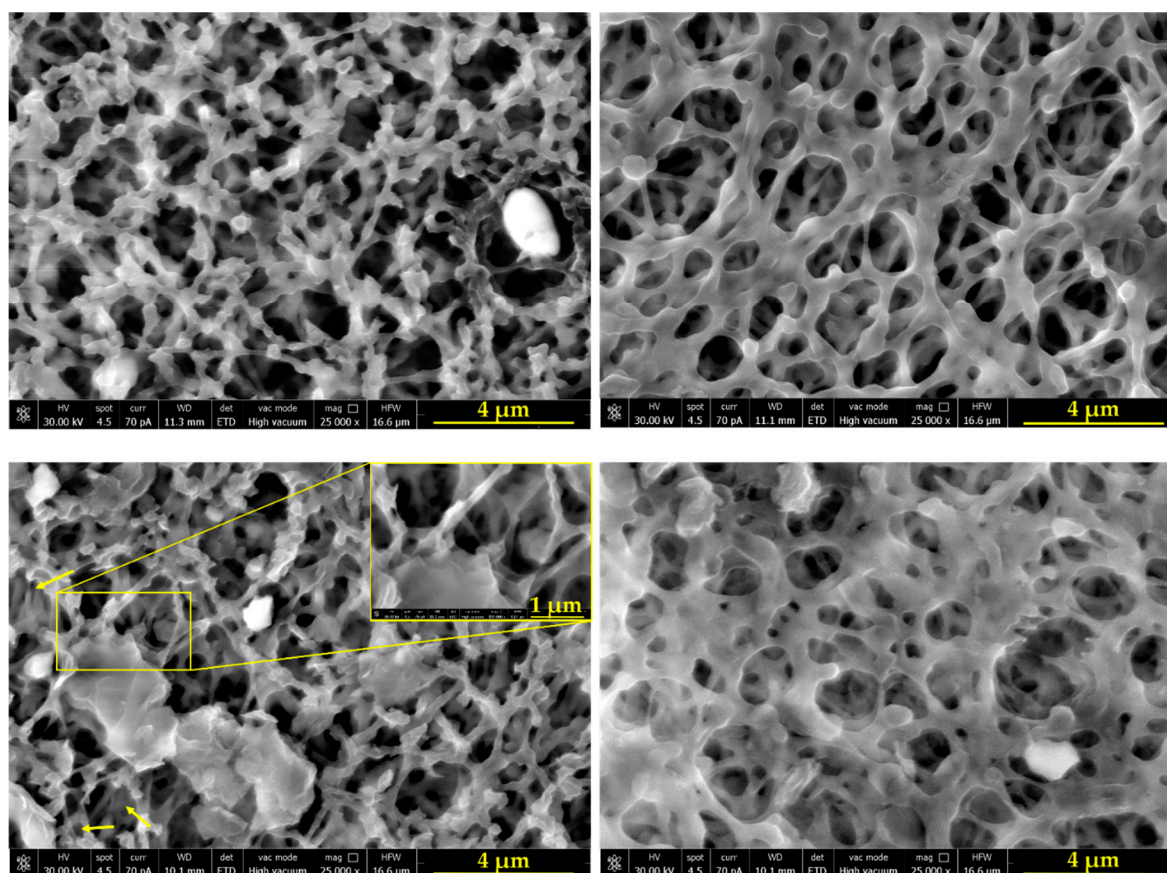


Figure 2. SEM micrographs that show the NC membrane before (A) and after (B) the experiment, and the NC/NH₂-PEG-rGO membrane before (C) and after (D) the experiment, all at a 4 μm scale bar.

In Figure 2C, the morphology of the NC membrane can be observed under a very thin layer of NH₂-PEG-rGO, showing features in some areas that indicate the natural flexibility of single layers of graphene, as indicated by the yellow arrows, but also occasional agglomeration of NH₂-PEG-rGO (inset, Figure 2C). Following the experiment (Figure 2D), a flattening of the membrane morphology is observed, which occurred because of the FAM-ssDNA sticking to the membrane, making it appear denser and less transparent compared to the NC/NH₂-PEG-rGO membrane and also to the pure NC membrane before the experiment. Furthermore, after the experiment, the surfaces of pure NC and NC/NH₂-PEG-rGO appear distinct, with collapsed pores observed in pure NC and a clogged appearance evident in places on the NC/NH₂-PEG-rGO hybrid membrane, indicating probably a greater affinity for the modified membrane to immobilize ssDNA compared to pure NC.

2.1.3. Wettability Characteristics

The wettability characteristics of the NC and NC/NH₂-PEG-rGO hybrid membranes are investigated by the water contact angle analysis and are depicted in Figure 3. The measurements indicate distinct surface characteristics for the NC and the NC/NH₂-PEG-rGO hybrid membrane. The NC membrane's surface is hydrophilic, having a water contact angle of 16.5° (Figure 3A), indicating significant spreading of water droplets. This behavior suggests favorable wetting characteristics, highlighting the membrane's hydrophilic nature[48,49].

In contrast, the NC/NH₂-PEG-rGO hybrid membrane (Figure 3B) exhibits hydrophobic features, as shown by the water contact angles that exceed 90°. NH₂-PEG-rGO modifies the surface of the NC membrane, resulting in reduced water attraction, with droplets resisting spreading, leading to a shift towards a more hydrophobic nature[50].

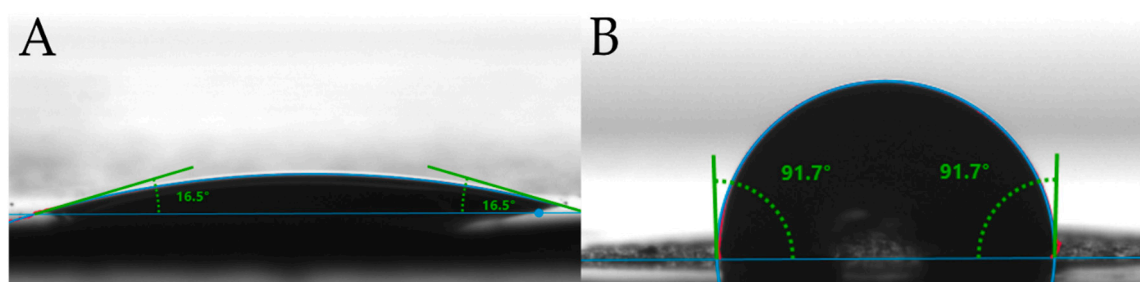


Figure 3. Water contact angles of (A) NC and (B) NC/NH₂-PEG-rGO hybrid membrane.

The characteristics exhibited by the surface of the NC/NH₂-PEG-rGO hybrid membrane have a beneficial influence on how biomolecules engage with, and respond to the membrane. The hydrophobic nature of NC/NH₂-PEG-rGO facilitates hydrophobic interactions with biomolecules, such as lipid tails or aromatic rings, enhancing the membrane's affinity for specific molecules. These interactions extend to molecules like ssDNA, where their hydrophobic regions, such as its nucleotide bases, can engage in favorable interactions with the NC/NH₂-PEG-rGO, thus enhancing the membrane's affinity for ssDNA. Moreover, the NH₂-PEG component adds specific chemical functionality to the surface, with amino (-NH₂) groups facilitating covalent or electrostatic bonding with complementary chemical groups or charges from ssDNA structure [51].

2.2. Oligo DNA Adsorption, Detection and Extraction Using NC/NH₂-PEG-rGO Hybrid Membranes in Complex Media

The efficiency of NC and NC/NH₂-PEG-rGO hybrid membranes in detecting and extracting ssDNA was assessed by measuring fluorescence intensity after a 60-minute incubation of the membranes in various ionic complex solutions containing MnCl₂, MgCl₂, or MnCl₂/MgCl₂. The resulting data is presented in Figure 4. Based on the results, the fluorescence intensity of the NC membrane in all ionic complex solutions does not exhibit any significant difference in immobilizing ssDNA, with the highest fluorescence recorded at approximately 250 r.f.u. for MgCl₂ ionic solution.

In the context of the NC/NH₂-PEG-rGO hybrid membrane, the measurements indicate a significantly greater affinity for immobilizing ssDNA in the complex solution containing MgCl₂, resulting in a fluorescence intensity of approximately 525 r.f.u. In contrast, the complex solution containing the MnCl₂ concentration demonstrates lower affinity, yielding a fluorescence intensity of around 330 r.f.u. While this affinity is lower than that observed with MgCl₂, it still demonstrates the higher membrane capacity to interact with ssDNA in different conditions when compared with the NC membrane. The divalent cations Mn²⁺ and Mg²⁺ from the complex solution enhance the interactions between ssDNA and the NC/NH₂-PEG-rGO membrane. This can occur because divalent cations can promote the condensation of ssDNA, reducing its exposure to the surrounding solvent and promoting interactions with hydrophobic surfaces [52,53]. The positively charged cations can interact with the negatively charged phosphate groups in the ssDNA backbone, neutralizing the

negative charges and reducing electrostatic repulsion between ssDNA and the membrane surface, thereby facilitating its immobilization[54,55].

Furthermore, MnCl_2 and MgCl_2 can create specific binding sites on the membrane surface through their coordination chemistry. These binding sites act as anchor points for ssDNA, enhancing its immobilization in a site-specific manner. MnCl_2 and MgCl_2 interact with the DNA molecule, stabilizing its secondary and tertiary structures. This stabilization makes the ssDNA more rigid and structured, increasing its affinity for binding to the membrane. Additionally, divalent cations enhance ssDNA stability by strengthening hydrogen bonding, while hybrid membrane amino groups engage in specific hydrogen bonding interactions with ssDNA nitrogenous bases, thereby enhancing overall adsorption onto the membrane [56–58].

The divalent ions (Mg^{2+} and Mn^{2+}) in the complex solutions demonstrates a higher affinity for immobilized ssDNA compared to the previous study, where the Na^+ ions did not exceed values higher than 300 r.f.u. [13]. According to the Manning-Oosawa theory, approximately 88% of the DNA surface charge is neutralized by divalent counterions (Mg^{2+} , Mn^{2+}), while in the case of monovalent ions (Na^+), this value is reduced to 76%, resulting in a lower charge density [59]. The lower charge density of monovalent ions means that they may not neutralize DNA's negative charges as effectively as Mg^{2+} ions. The higher charge density of Mg^{2+} ions allow for more effective charge neutralization of DNA, reducing electrostatic repulsion and facilitating DNA-protein binding [59,60].

Various components, including serum proteins found within αMEM , particularly BSA and SDS, play a crucial role in reducing the non-specific binding of oligo ssDNA to diverse surfaces in the medium while simultaneously promoting its adherence to the membrane. BSA acts as a protective barrier, protecting the negatively charged oligo ssDNA molecules against repulsive electrostatic interactions with NC/ NH_2 -PEG-rGO, while SDS deactivates nucleases and regulates non-specific adsorption on the surface of the NC/ NH_2 -PEG-rGO hybrid membrane [61–65]. Moreover, αMEM incorporates glucose and other carbohydrates, leading to an increase in the solution's osmotic pressure. This heightened osmotic pressure has the potential to induce a flow of water from the surrounding medium into the membrane, thereby enhancing the adsorption of oligo ssDNA onto the membrane's surface [66,67].

The role of pH in modulating the interaction between ssDNA and the membrane is also important for understanding the binding dynamics. At Tris-HCl pH 8, the environment is slightly alkaline, resulting in partial deprotonation of both ssDNA and the amino groups ($-\text{NH}_2$) on the membrane's surface. This exposes negatively charged phosphate groups on the ssDNA backbone and positively charged amino groups on the membrane, promoting attractive electrostatic interactions between ssDNA and the membrane, facilitating ssDNA adsorption. On the other hand, at Tris-HCl pH 7 the environment is slightly acidic. This pH level can influence the charge state of both ssDNA and the membrane. The phosphate groups on the ssDNA backbone and the amino groups ($-\text{NH}_2$) on the membrane surface are both partially protonated, resulting in a reduced net charge on both molecules. This reduction in charge weakens the electrostatic interactions between ssDNA and the membrane, making it easier for ssDNA to detach or desorb from the membrane [68–71].

However, considering the aforementioned phenomena, the immobilization of ssDNA to the surface of the modified membrane seems to be controlled by a series of factors which need to be specifically modulated: components in the ionic solution, ionic valence and pH.

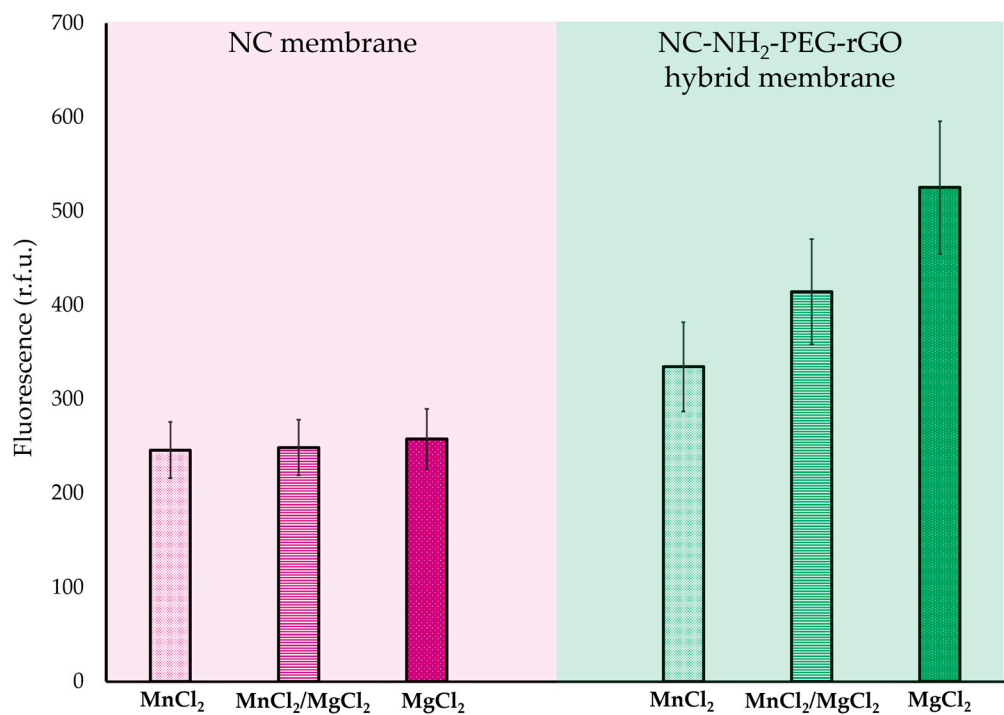


Figure 4. Measurement of FAM-ssDNA fluorescence intensity after 60 minutes of incubation on NC and NC/NH₂-PEG-rGO membranes in complex solutions with various (MnCl₂, MnCl₂/MgCl₂, and MgCl₂).

Table 1 illustrates the mass (pg) of ssDNA desorbed from NC and NC/NH₂-PEG-rGO hybrid membrane after 60 minutes of incubation in the three complex media containing MnCl₂, MnCl₂/MgCl₂, and MgCl₂. In the case of the NC membrane, the results indicate that the desorbed mass does not significantly differ among the ionic complex used, ranging from 285 pg for MnCl₂ to 300 pg for MgCl₂. Conversely, for NC/NH₂-PEG-rGO hybrid membranes, the highest mass detachment was observed in the case of MgCl₂, approximately 610 pg, while the lowest was noted in the case of MnCl₂, with approximately 390 pg. These findings suggest that, on average, a greater quantity of ssDNA is desorbed from NC/NH₂-PEG-rGO hybrid membranes compared to the NC membrane.

The difference in the amount of ssDNA desorbed from the NC/NH₂-PEG-rGO hybrid membranes in all the media used can be attributed to the fact that Mg²⁺ ions are smaller than Mn²⁺ ions, enabling them to neutralize the negative charges more effectively on the ssDNA molecule. Moreover, the smaller size of Mg²⁺ allows them to access and interact with binding sites on the membrane more effectively. Thus, smaller ions can fit into tighter spaces and reach sites that might be less accessible to larger ions, enhancing their binding efficiency [72,73].

Furthermore, both Mg²⁺ and Mn²⁺ ions are divalent cations, but Mg²⁺ ions have a higher charge density compared to Mn²⁺ ions. This higher charge density results in stronger electrostatic interactions with negatively charged functional groups on the membrane's surface, such as oxygen atoms or other electronegative elements. This enhanced electrostatic attraction makes Mg²⁺ ions more likely to bind to the membrane. In addition to the charge density, Mg²⁺ ions are known to form stable coordination complexes with phosphate groups, which are abundant in ssDNA and RNA molecules. These complexes can enhance the binding of DNA to surfaces. While Mn²⁺ ions are capable of forming similar complexes, they may be less effective in binding ssDNA to the membrane due to their lower charge density [74–76].

Table 1. Data of ssDNA mass (pg) and standard deviation (s.d.) after 60 minutes of incubation on the NC and NC/NH₂-PEG-rGO Hybrid Membranes in complex media with various ionic particles (MnCl₂, MnCl₂/MgCl₂, and MgCl₂).

Ionic particles in complex media	Desorption of ssDNA from the NC Membrane measured in pg, with standard deviation (s.d.)		Desorption of ssDNA from the NC/NH ₂ -PEG-rGO Membrane measured in pg, with standard deviation (s.d.)	
	Time	60 minutes		
U/M	pg	s.d.	pg	s.d.
MnCl ₂	286.41	±34.75	389.32	±54.98
MnCl ₂ /MgCl ₂	289.67	±34.4	482.35	±65.23
MgCl ₂	300.25	±37.1	611.12	±82.03

U/M stands for Unit of Measurement.

Compared to our previous study where the NC-GO hybrid membrane emerged to extract approximately 335 pg of ssDNA with NaCl, in this case, the NC/NH₂-PEG-rGO membrane implies a much higher extraction yield, with over 600 pg of ssDNA in the case of MgCl₂ [13]. This effect can be attributed to the presence of amino (NH₂) and polyethylene glycol (PEG) functional groups on the NC/NH₂-PEG-rGO membrane that can enhance its ability to interact with and immobilized ssDNA molecules. These functional groups can provide additional binding sites and alter the surface charge of the membrane, potentially making it more favorable for ssDNA adsorption while leaving the graphene surface available for π - π interactions with ssDNA. The specific chemical composition and functional groups on the NC/NH₂-PEG-rGO membrane may also favor interactions with Mg²⁺ ions compared to the interactions that occur between the NC-GO membrane and Na⁺ ions.

Although the NC/NH₂-PEG-rGO membrane effectively extracts a higher quantity of ssDNA from these complex media containing various ionic particles, we recognize the necessity for further refinement. Undertaking additional research endeavors will facilitate the enhancement of both the efficiency and dependability of our technique, thereby expanding its versatility in the extraction of diverse biomolecule types from various sample sources. This advancement holds promise for the enrichment of molecular biology applications and other fields that depend on precise biomolecule extraction methods.

3. Materials and Methods

3.1. Reagents

Aminated Graphene Amino-PEG covalently linked, CAS No.: 7782-42-5, was procured from ACS-Materials (Pasadena, CA, USA). FAM-ssDNA was bought from Integrated DNA Technologies, Inc. (Coralville, IA, USA) and consists of the following base series: 5'-TTTCAACATCAGTCTGATAAGCTATCTCCC-3', with labeling at the final primer using 6-carboxyfluorescein. The acquisition of the NC membranes characterized by a 47 mm diameter and an 8.0 μ m pore size was carried out from Sartorius (Gottingen, Niedersachsen, Germany). α MEM, magnesium chloride (MgCl₂), sodium dodecyl sulfate (SDS) with the chemical formula CH₃(CH₂)₁₁OSO₃Na, tris hydrochloride (Tris-HCl), and bovine serum albumin (BSA) were all sourced from Sigma-Aldrich, based in St. Louis, MO, USA. Manganese chloride (MnCl₂) was procured from SILAL (Bucharest, Romania).

3.2. Preparation of Aminated Graphene Dispersion

The aminated graphene dispersion was obtained at a concentration of 1 mg/mL after 2 hours of ultrasonication in an ice bath, using a VC×750 sonicator (Sonics & Materials, Inc., Newtown, CT, USA). This equipment operated at 10-second pulse followed by a 5-second pause, and a frequency of 20 kHz.

3.3. NC/NH₂-PEG-rGO Hybrid Membrane Fabrication

NC membranes were prepared with a 5 mm diameter and an approximate mass of 1.1 mg using a conventional paper hole punch. Non-covalent modification of the NC membrane was carried out using an approach akin to the dot blot technique. Initially, a diluted dispersion of NH₂-PEG-rGO at a concentration of 400 µg/mL was prepared, and 5 µL of this dispersion was subsequently applied to the NC membrane through drop casting. Prior to their utilization, the resulting hybrid NC/NH₂-PEG-rGO membranes were left to air-dry overnight and cleaned with deionized water.

3.4. Adsorption, Extraction, Detection of FAM-Labeled ssDNA in Ionic Solutions from NC/NH₂-PEG-rGO Hybrid Membrane

Initially, three distinct ionic solutions were prepared for oligonucleotide immobilization using αMEM, each consisting of 100mM MnCl₂, 100mM MgCl₂, and a combination of MnCl₂/MgCl₂. These solutions were supplemented with 0.1 mg/mL BSA, 10 mM Tris-HCl at pH 8.0, 0.1% SDS, and 16 nM of FAM-ssDNA. The samples were spread at a volume of 100 µL per well into black Costar 96-well flat-bottomed plates.

In the case of adsorption approach, nine NC/NH₂-PEG-rGO hybrid membranes were utilized (Phase 2 from Figure 5). Each membrane was incubated for 60 minutes in the prepared ionic solutions to facilitate the binding of FAM-ssDNA (Phase 3 according to Figure 5).

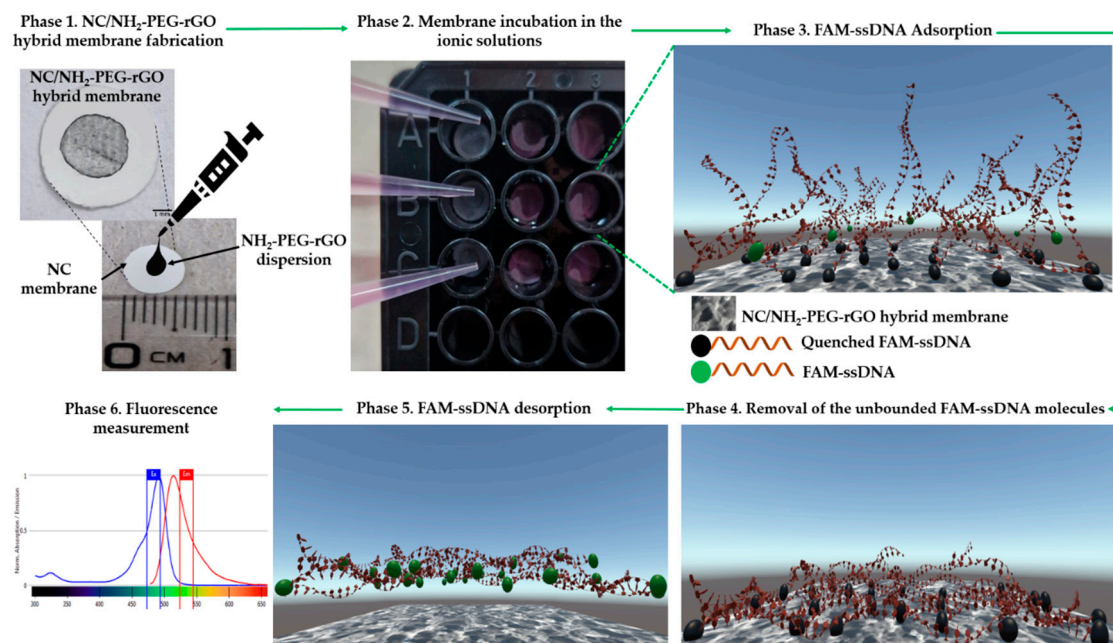


Figure 5. (Phase 1) Fabrication of NC/NH₂-PEG-rGO hybrid membrane by drop casting a dispersion of NH₂-PEG-rGO on the NC membrane; phase 2 entails placing membranes into Costar 96-well flat-bottomed plates, while phase 3 involves immersing the membranes in solution containing FAM-ssDNA, phase 4 involves removing the unbounded FAM-ssDNA molecules from the membrane surface, while phase 5 is dedicated to FAM-ssDNA desorption from NC/NH₂-PEG-rGO hybrid membrane surface and fluorescence emission intensity measurements.

To eliminate any unbounded ssDNA the membranes were washed with distilled water following the incubation (phase 4 from Figure 5). Subsequently, the membranes were immersed in a 10 mM Tris-HCl solution at pH 7.0 for 45 minutes to initiate the FAM-ssDNA desorption process (phase 5 according to Figure 5). Following the removal of the NC/NH₂-PEG-rGO hybrid membrane, the fluorescence of the desorbed solution was quantified using a microplate reader at a wavelength of 535 nm. The measurements were performed in triplicate for each membrane used, and the results

were reported as average fluorescence values with corresponding SD. Utilizing Equation (1) the resulted values were transformed into weight units and showcased in Table 1.

$$M_f = \frac{F_f * M_i}{F_i} \quad (1)$$

where, M_f denotes the final mass of the released FAM–ssDNA in picograms (pg), F_f signifies the final fluorescence in relative fluorescence units (r.f.u.) following the desorption procedure. M_i is employed to represent the initial mass of the FAM–ssDNA in the complex solution (approximately 4420 pg), which was determined using the "DNA molecular weight and conversion" available through ThermoFisher [77]. Lastly, F_i denotes the solution fluorescence in r.f.u. after the introduction of the FAM–ssDNA sequence (around 3800 r.f.u.).

3.5. Spectrofluorimeter Assay

The fluorescence emission intensity was determined utilizing TECAN Spark Fluorescence microplate reader (Tecan Trading AG., Männedorf, Switzerland) at a room temperature of 23°C, with five readings recorded for each well at a wavelength of 535 nm.

3.6. Membrane Characterization

The FTIR studies to investigate the interaction between NH₂-PEG-rGO and NC were carried out using the SHIMADZU 8900 equipment (Kyoto, Japan). Each FTIR spectrum was obtained within a range of 400–4000 cm⁻¹, with 32 measurements taken for each sample, and a resolution of 4 cm⁻¹.

NC membranes surface morphology was investigated using the FEI Quanta F250 scanning electron microscope. This examination was conducted both before and after the application of NH₂-PEG-rGO particles, as well as before and after immersing the membranes in the complex solution. Beforehand to performing the SEM analysis, a thin stratum of gold-palladium was applied to increase the conductivity of the membranes.

Assessing the hydrophilicity of the membranes involved the application of the sessile drop method and the Krüss Scientific Drop Shape Analyzer-DSA100 (Hamburg, Germany). Investigating the impact of NH₂-PEG-rGO on the hydrophilic characteristics of NC membranes included conducting static water contact angle measurements at ambient temperature. Capturing the configuration of the deionized water droplet on the specimen surface utilized a CF03 digital camera over a 5-second interval following the deposition of a 2 µL droplet. The determination of water contact angle values involved using the DSA3 software and constituted the mean of three measurements for each specimen. The interpretation of outcomes employed the Young–Laplace equation [78].

4. Conclusions

In the present study, we conducted an investigation using an NC/NH₂-PEG-rGO membrane for oligonucleotide extraction. The structural and morphological features obtained by SEM and FTIR investigations demonstrated the successful fabrication of the hybrid membrane. SEM microscopy unveiled a sponge-like structure under a very thin layer of graphene with seldom agglomerations, showcasing the effective dispersion and uniform coating of NH₂-PEG-rGO on the NC membrane. The wettability characteristics revealed that the NC/NH₂-PEG-rGO hybrid membrane displayed a significantly higher hydrophobic water contact angle at 91.7° compared to the NC control membrane, which exhibited a contact angle of 16.5°. The higher hydrophobic character of the membrane is believed to have a beneficial effect on nucleic acids adsorption.

The fluorescence emission intensity results demonstrate that the NC/NH₂-PEG-rGO hybrid membrane consistently adsorbed and desorbed oligonucleotides across three complex media with different ionic compositions (MnCl₂, MnCl₂/MgCl₂, MgCl₂). Notably, the medium containing MgCl₂ exhibited the most favorable outcome.

In the case of the control, the NC membrane exhibited a reduced capacity to adsorb ssDNA, resulting in the lowest fluorescent intensity of approximately 250 r.f.u. in the media with MnCl_2 , which corresponds to a quantity of 286 pg. The data obtained suggests that modifying the NC membrane with NH_2 -PEG-rGO substantially improves its interaction with nucleic acids, resulting in a significantly higher binding affinity.

After incubating the NC/ NH_2 -PEG-rGO hybrid membrane in complex media with MgCl_2 , the highest fluorescent intensity displayed a signal above 520 r.f.u., corresponding to a quantity of 611 pg of ssDNA. However, when incubated in complex media with MnCl_2 , the fluorescent signal decreased to 330 r.f.u., corresponding to 290 pg. Moreover, in any type of media used, the NC/ NH_2 -PEG-rGO can immobilize a higher amount of oligonucleotides than the NC membrane.

Hence, our research outcomes strongly indicate that when the NC membrane is modified with NH_2 -PEG-rGO enables a substantially increased immobilization capacity for oligo-DNA on the membrane surface. This significant enhancement in oligo-DNA immobilization underscores the potential of NH_2 -PEG-rGO as a superior choice for membrane modification, with profound implications for applications demanding efficient oligo-DNA immobilization and extraction.

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