

Review

Not peer-reviewed version

Advancements in Circulating Tumor Cell Research: Bridging Biology and Clinical Applications

[Philip Salu](#) * and [Katie M. Reindl](#) *

Posted Date: 4 March 2024

doi: 10.20944/preprints202403.0086.v1

Keywords: Circulating tumor cells; liquid biopsy; cancer research; metastasis; personalized medicine; single-cell sequencing; biomarkers.



Preprints.org is a free multidiscipline platform providing preprint service that is dedicated to making early versions of research outputs permanently available and citable. Preprints posted at Preprints.org appear in Web of Science, Crossref, Google Scholar, Scilit, Europe PMC.

Copyright: This is an open access article distributed under the Creative Commons Attribution License which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Review

Advancements in Circulating Tumor Cell Research: Bridging Biology and Clinical Applications

Philip Salu ¹ and Katie M. Reindl ^{1,2,*}

¹ North Dakota State University, Cellular and Molecular Biology, Fargo, ND, USA; philip.salu@ndsu.edu

² North Dakota State University, Department of Biological Sciences, Fargo, ND, USA; katie.reindl@ndsu.edu

* Correspondence: katie.reindl@ndsu.edu; Tel.: 701-231-9427

Simple Summary: Cancer remains one of the leading causes of death worldwide due to widespread metastasis. Circulating tumor cells (CTCs) were first detected in 1886 and have been thought of as seeds of metastasis. However, emerging evidence has shown that CTCs can be used for early disease detection and as predictors of disease prognosis and treatment response. This review discusses recent findings regarding the detection and characterization of CTC. Integrating these findings into routine clinical workflows promises to revolutionize the diagnosis and treatment of cancer.

Abstract: Circulating tumor cells (CTCs) are cells released from the primary tumor and intravasate into the blood or lymphatic vessels, where they are transported to distant sites and act as seeds that initiate cancer metastases. Recent advances in CTC research have shown their relevance as prognostic markers for early and metastatic disease detection, predictive biomarkers for relapse, and response to medical intervention or therapy. The rapidly evolving landscape of CTC biology has opened new avenues for understanding cancer progression, metastasis, and treatment response. Additionally, translating these findings into clinical applications holds promise for improving cancer diagnostics, prognosis, and personalized therapeutic strategies. This review discusses the significance of CTCs in cancer research and their associated challenges. We explore recent developments in the detection and characterization of CTCs and their implications in cancer research and clinical practice.

Keywords: Circulating tumor cells; liquid biopsy; cancer research; metastasis; personalized medicine; single-cell sequencing; biomarkers

1. Introduction

In the United States (US), cancer remains the second leading cause of death, affecting individuals of all ages and ethnic groups [1]. The estimated number of all cancer-related deaths in the US is over 609,000, about 18 % of all US deaths [1]. Cancer mortality is due to poor treatment response, metastasis, and the lack of early detection methods [2–4]. Liquid biopsies are gaining popularity as noninvasive techniques for early disease detection [5,6]. These liquid biopsies contain circulating tumor cells (CTCs) that are released into the blood by a tumor and travel through the bloodstream to other areas of the body to form metastatic niches (Figure 1) [7].

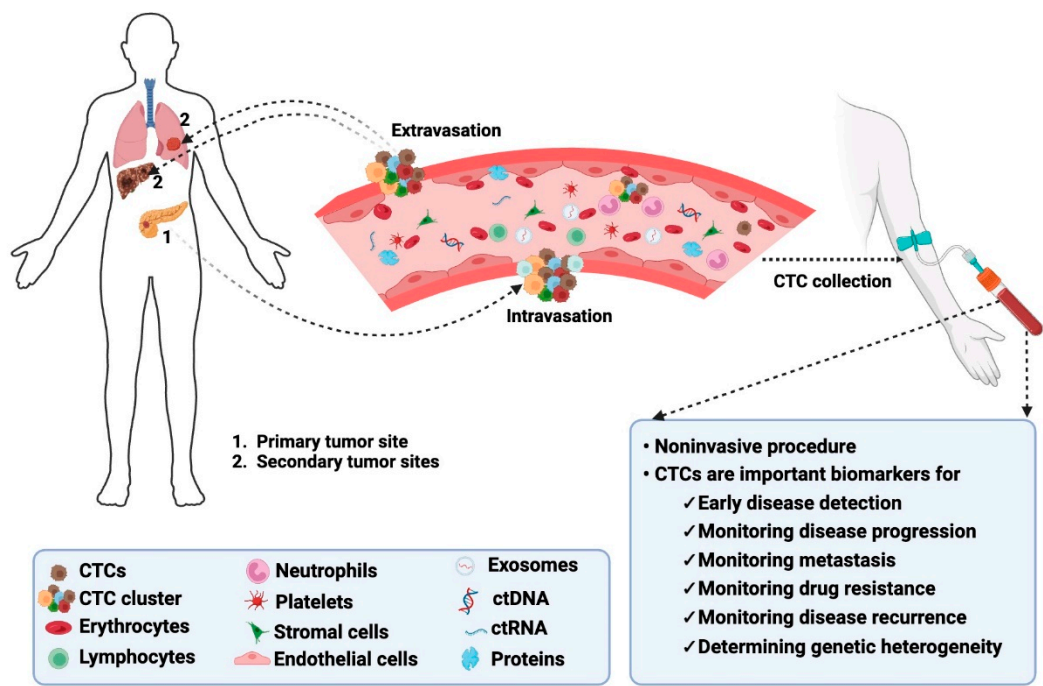


Figure 1. Biology of CTCs in cancer. CTCs detach from their primary tumors, intravasate into the bloodstream, and extravasate from the bloodstream to colonize secondary tumors or metastatic sites. CTCs can be detected following noninvasive collection procedures and serve as biomarkers for monitoring multiple disease aspects. The image was created using BioRender.com.

CTCs are proving to be essential biomarkers that can be used for early disease detection, monitoring metastasis and cancer progression, and determining genetic heterogeneity in tumors [8–10]. Compared to traditional tumor biopsies, obtaining CTCs from liquid biopsies is non-invasive and real-time [11]. This can be more acceptable to patients, paving the way for more personalized cancer medicine.

However, several challenges currently affect CTC research. One of the main issues is that they are rare and heterogeneous [12,13]. CTCs are present in very small numbers, about 1-100 per 10^9 white blood cells [14]. These scarcely populated cells are characterized by varying morphology, phenotypical characteristics, and genetic makeup, making isolation and characterization difficult [15,16]. Moreover, standardizing isolation techniques and establishing their clinical relevance presents ongoing challenges [17,18]. Ethical considerations and the need for more extensive clinical trials also pose significant obstacles to advancing CTC research [19]. Available techniques for detecting CTCs, like flow cytometry-based and microscopy-based detection methods, need refinements to improve accuracy and sensitivity [20].

Current approaches to the isolation and analysis of CTC biology involve using microfluidic devices, single-cell sequencing, and the integration of multi-omics data to determine the molecular and functional heterogeneity of CTCs [21–23]. Machine learning tools trained to interpret complex CTC data are also being developed to aid in identifying biomarkers and potential therapeutic targets [24,25]. These methods hold promise for improved detection, monitoring, and uncovering treatment-resistant or metastatic CTC subpopulations. This review discusses the significance of CTCs in cancer research, current challenges, and new approaches to characterizing CTC tumor cell biology. Additionally, we preview the latest findings in CTC research and their clinical implementations in cancer.

2. Significance of CTCs in Cancer and Clinical Implications

Circulating tumor cells hold immense significance in cancer research and clinical applications due to their potential to offer valuable insights into various aspects of cancer biology and patient care. Recent discoveries in CTC research have significantly advanced our understanding of their roles in cancer progression, metastasis, treatment response, and personalized medicine. These new developments hold significant clinical implications, potentially revolutionizing various aspects of cancer. Below, we discuss recent insights into early cancer detection, metastatic potential, prognosis, treatment monitoring, and personalized therapy based on CTC characterization and their potential clinical impacts.

2.1. Early Cancer Detection

Early detection of cancer is crucial for effective treatment and disease management to reduce cancer-related deaths and improve patient outcomes. CTCs offer the potential for liquid biopsy-based screening, allowing the identification of cancers at an earlier, more treatable stage [26]. CTCs were first detected in cancer in the eighteenth century [27]. However, the process of local invasion and extravasation of CTCs has recently been shown to occur within a short span of time [28], suggesting that CTCs can be detected in the bloodstream at early stages of cancer, potentially allowing for early diagnosis before tumors become clinically evident [29,30]. A study involving 667 participants, of which 235 were healthy individuals and 432 patients with either colorectal cancer (CRC) or adenomas, showed that CTCs can be used to distinguish healthy individuals from patients with CRC or adenomas with a detection sensitivity of up to 95.2 % even before colonoscopy [31]. Another study evaluating patients with chronic obstructive pulmonary disease (COPD) without clinically diagnosed lung cancer revealed that 5 out of 168 patients had detectable CTCs. Notably, all five individuals with detectable CTCs developed lung nodules within 1-4 years [32]. A third study involving patients with suspected prostate cancer based on high serum prostate-specific antigen (PSA) levels also revealed a high positive CTC-predicted biopsy outcome and prostate cancer aggressiveness [33]. Thus, the importance of using CTCs to predict disease occurrence cannot be underestimated. Additionally, capturing and analyzing CTCs using liquid biopsy techniques offer a non-invasive alternative to traditional tissue biopsies, providing a real-time snapshot of the tumor's genetic and phenotypic characteristics.

2.2. Prognostic Indicators

The presence of CTCs in the bloodstream provides valuable prognostic information, aiding in predicting disease outcomes and guiding treatment decisions [34–36]. Studies have shown that higher CTC counts and the presence of CTC clusters are associated with poorer outcomes across various cancer types [37,38]. In clinical settings, CTCs are commonly used as a liquid biopsy method to screen for tumors, monitor disease, and predict patients' prognoses [39]. A CTC count cutoff value of ≥ 3 CTCs/7.5 mL blood is considered unfavorable for metastatic colorectal cancer [40]. In a study involving hormone receptor-positive (HR+) metastatic breast cancer patients, the overall survival (OS) and progression-free survival (PFS) of patients with CTC count ≥ 5 per 7.5 mL blood after treatment was significantly worse than that of patients with < 5 CTCs [41]. Also, CTC counts ≥ 10 per 5 mL blood of small-cell lung cancer (SCLC) patients were closely associated with advanced stage (high lymph node metastasis and distant metastasis), indicating a more unfavorable prognosis [42]. These recent investigations have refined our understanding of CTCs as robust prognostic indicators, guiding clinicians in predicting disease outcomes and tailoring treatment strategies. Multiple detection methods have been developed to comprehensively characterize CTCs, providing valuable insights into the likely cause of disease [39,43]. CTC enumeration is increasingly being incorporated into clinical practice as a predictive tool, and CTC counts are being used to stratify patients, helping to tailor treatment plans based on individual prognoses [44].

2.3. Tumor Heterogeneity

Tumor cells obtained from liquid biopsies exhibit genetic and phenotypic diversity, reflecting the heterogeneity within the primary tumor [13,45]. Numerous studies have discovered the presence of different CTC subpopulations in cancers [13,46–48]. For instance, Freed and colleagues identified two CTC subpopulations expressing epithelial cell adhesion molecule, EpCAM ($\text{CTC}^{\text{EpCAM}}$) or fibroblast activation protein-alpha, FAP α ($\text{CTC}^{\text{FAP}\alpha}$) in pancreatic ductal adenocarcinoma (PDAC) patients. Using the ratios of $\text{CTC}^{\text{FAP}\alpha}$ to $\text{CTC}^{\text{EpCAM}}$; they were able to stratify patients as responders versus non-responders to niraparib treatment [46]. In a non-small cell lung cancer (NSCLC) study, the authors were able to detect different gene signatures related to therapy resistance (MET and HER3) and the initiation of metastasis (ALHD1) using CTCs [49]. The spectrum of cancer heterogeneity has long been under scrutiny as a factor allowing the tumor to adapt to different microenvironmental stressors with different subpopulations that may evolve to increase disease aggressiveness and decrease therapeutic response [50]. The study of CTCs, therefore, allows researchers to understand and monitor tumor heterogeneity, providing crucial information for devising effective and personalized treatment strategies.

2.4. Metastasis and Disease Progression

Metastasis accounts for more than 90% of cancer-related mortalities [51–53]. This is because treatment options available for patients with metastatic solid tumors are rarely curative [54,55]. Metastatic events in cancer involve the spread of tumor cells from the primary to secondary sites following phenotypic transition from epithelial to mesenchymal phenotype, cancer cell invasion into circulation, dormancy, and colonization at distant sites [56,57]. Research suggests that CTCs can be shed from primary tumors at the early stages of cancer, challenging the traditional view that metastasis occurs primarily in advanced disease [6]. This early dissemination contributes to the understanding of metastatic potential and the dynamics of cancer progression [6,58]. Following the injection of pancreatic cancer cells expressing fluorescent proteins into the earlobes of mice to form solid tumors, the presence of CTCs with fluorescent proteins were detected in the bloodstream of the mice at different stages of development [59]. Using quantum dots, the researchers identified cells with high metastatic potential, those expressing clusters of differentiation CD24+ and CD133+ [59]. The development of metastatic niches often signifies a significant progression in the tumor stage, and studies have linked CTC characteristics, such as the presence of CTC clusters, to increased metastatic potential, aiding in the identification of patients at higher risk [60–62]. In metastatic breast cancer, a CTC count ≥ 5 in 7.5 mL of blood correlates with worse overall survival and progression-free survival [40]. Indeed, a CTC count of less than five in patients with stage IV breast cancer is used to classify the tumor as stage IV indolent, whereas a CTC count greater than five classifies the tumor as stage IV aggressive [40]. CTC clusters exhibit stemness characteristics and can evade the immune system by recruiting immunosuppressive cells [63]. This attribute helps to prevent CTCs from being attacked by antitumor immune cells like natural killer (NK) cells, thereby increasing their metastatic potential [63–65]. Using a mass spectrometry-based untargeted metabolomics approach, human colorectal cancer CTC-derived cells were shown to have low or high metastatic potential based on metabolic features [66]. A combination of metabolites such as glutamic acid, malic acid, lactic acid, and aspartic acid, along with higher CTC counts, positively predicted the metastatic risk in patients, providing evidence of metabolic phenotype influence on the metastatic potential of cells [66]. Metabolic reprogramming of CTCs enables them to survive harsh conditions in the bloodstream and enhances their ability to establish a favorable microenvironment for metastasis, known as metastatic niches [67]. The molecular mechanisms by which CTCs influence the pre-metastatic niche (PMN), prepare distant organs for colonization, and contribute to the metastatic process are yet to be fully understood. However, CTCs acquire enhanced migratory and invasive capabilities when undergoing epithelial-mesenchymal transition (EMT) [68]. Recent discoveries have elucidated the dynamic nature of EMT in CTCs, identifying hybrid epithelial/mesenchymal states that may contribute to metastatic progression [38]. For instance, the presence of both epithelial and mesenchymal markers indicates the occurrence of intrahepatic metastasis, while the presence of mesenchymal phenotypes prompts the development of extrahepatic metastasis [38]. Expression of mesenchymal markers like

N-cadherin, vimentin, snail, and slug have also been detected in CTCs, highlighting their use in metastasis prediction [69,70]. Using CTC-based information to assess the risk of metastasis will help implement more aggressive treatment strategies for patients with a higher likelihood of developing metastatic disease.

2.5. Treatment Response Monitoring

Changes in CTC counts and characteristics during treatment can indicate treatment response or resistance [41]. Real-time monitoring of CTC dynamics will allow for timely adjustments to therapeutic strategies. To monitor patient response to therapy, longitudinal tissue biopsies must be collected. However, this procedure is often invasive and complicated by the location and size of tumors [71]. Therefore, CTCs can serve as dynamic biomarkers for monitoring treatment response and assessing the efficacy of therapies. Indeed, CTCs from patients with either chemosensitive or chemorefractory SCLC were tumorigenic in immune-compromised mice, and their resultant CTC-derived explants mirrored the donor patients' response to platinum and etoposide chemotherapy [72]. Molecular alterations that affect a tumor cell's response to specific drugs can also be determined through molecular profiling of CTCs. For example, the expression level of cyclin D1 (CCND1) was shown to be significantly reduced in patients with head and neck squamous cell carcinoma (HNSCC) following nivolumab treatment [73]. In contrast, the expression of NANOG increased significantly [73]. Changes in CTC count or specific genetic alterations can, thus, indicate whether a treatment effectively targets the tumor, enabling timely adjustments to the therapeutic approach.

2.6. Minimal Residual Disease Monitoring

CTCs can be used to monitor minimal residual disease (MRD) after surgery or other treatments, helping identify potential disease recurrence early. Li and colleagues used preoperative CTC concentration to predict postoperative recurrence or metastasis in patients with NSCLC [74]. Detection of CTCs pre- and post-adjuvant chemotherapy can also serve as prognostic indicators of early relapse [34]. These indicators allow for investigations into novel therapeutic approaches to curb relapse and drug resistance. CTC-based MRD monitoring has also been explored as a valuable tool in post-treatment surveillance, offering insights into the effectiveness of interventions and informing decisions regarding additional therapies [75]. CTCs can enter a state of dormancy, temporarily evading the immune system and therapeutic interventions [76,77]. Recent studies have shed light on the mechanisms governing CTC dormancy and the factors that lead to their reactivation, contributing to a deeper understanding of cancer relapse [78–80]. Molecular characterization of CTCs can also help identify and analyze drug-tolerant clones within the tumor microenvironment (TME) [81].

2.7. Personalized Medicine

Precision oncology aims to treat patients according to the molecular characteristics of their tumors, adjusting these treatment strategies as the tumors evolve [82]. Molecular characterization of CTCs contributes to identifying actionable mutations and potential therapeutic targets. Whole exome sequencing (WES) of patient CTC samples has been shown to reflect the genomic characteristics of their corresponding solid tumors more accurately [83–85]. This genomic information can be crucial in developing personalized treatment plans. Functional mutations in driver genes like EGFR, KRAS, and TP53 can be determined quickly and used to stratify patients for therapy [85]. Mutational characterization of CTCs can also help physicians decipher if and when a patient is developing resistance to a particular chemotherapeutic agent, thereby allowing for proactive treatment decisions [86]. In this regard, CTC-based liquid biopsies facilitate the development of personalized treatment plans, allowing for the selection of targeted therapies based on the specific genetic profile of the tumor. Furthermore, CTC-based liquid biopsies are increasingly integrated into clinical trials and treatment decision-making.

2.8. Clinical Trials and Drug Development

CTCs offer a tool for assessing drug response in preclinical and clinical trials, aiding in developing novel cancer therapeutics. Current CTC-based clinical trials aim to use CTC-positivity and dynamics to predict clinical outcomes and therapy responses in patients [19]. Following the STIC CTC randomized clinical trial (NCT0170605), researchers determined that CTC count was a reliable biomarker for choosing between chemotherapy and a single-agent endocrine therapy as a first-line treatment in hormone receptor-positive ERBB2-negative metastatic breast cancer [87]. Data from clinical trials can also be used to model the prognostic impact of a tumor and the design of specific targeting agents [88]. Therefore, CTC-based studies provide a platform for evaluating the effectiveness of new treatments and understanding drug resistance mechanisms.

In summary, CTCs have a significant role in cancer research and clinical applications. They act as dynamic biomarkers, providing insights into the complex nature of cancer. With technological advancements, CTC-based approaches can be integrated into regular clinical practice. This can potentially revolutionize cancer diagnosis, treatment, and patient outcomes.

3. Challenges of using CTCs in Research and Clinical Applications

Studying circulating tumor cells presents several challenges, primarily stemming from their rarity in the bloodstream and the technical complexities associated with their isolation and characterization. Despite all the limitations associated with CTCs, rapidly evolving technology is paving the way for cutting-edge detection and characterization methods. Table 1 outlines some key challenges related to the use of CTCs.

Table 1. Challenges associated with detection, isolation, and characterization of CTCs.

Challenges	Description/ Effects	Ref.
Low Frequency in Bloodstream	<ul style="list-style-type: none">- Extremely rare compared to other circulating cells (e.g. blood cells)- Ranges from a few to a few hundred CTCs per milliliter of blood- Low frequency makes detection and isolation difficult	[89,90]
Heterogeneity	<ul style="list-style-type: none">- Exhibits both genetic and phenotypic variabilities- Reflects cellular diversity within the primary tumor- Heterogeneity complicates efforts to capture a representative sample of CTCs for analysis	[91–93]
Cell Viability	<ul style="list-style-type: none">- CTCs are fragile and can be damaged during isolation processes- Isolation and analysis of viable CTCs are crucial for meaningful downstream studies- Lack of viable cells may affect subsequent functional assays	[94]
Dynamic Changes in CTC Numbers	<ul style="list-style-type: none">- Number of CTCs in the bloodstream vary over time- Number of CTCs is affected by tumor size, treatment effects, and stage of the disease- Variable CTC numbers add complexity to studying CTCs longitudinally	[14,95]
Contamination from Normal Cells	<ul style="list-style-type: none">- Isolation of CTCs can be complicated by contamination from normal blood cells such as leukocytes- Presence of other cells reduces the purity of CTC samples- Contaminating cells can interfere with downstream analyses	[96]
Technical Limitations	<ul style="list-style-type: none">- Traditional methods for CTC isolation, such as density gradient centrifugation, do not efficiently capture CTCs due to their similar size and density compared to other blood cells	[97,98]

	- Newer technologies, like microfluidic devices and immunomagnetic separation, need refinements to achieve high purity and recovery rates	
Lack of Standardization	- Lack of standardized protocols for CTC isolation and characterization - Different isolation methods and technologies may yield varying results - Lack of standardization makes it challenging to compare data across studies, leading to potential discrepancies in the interpretation of findings	[26,99,100]
Ethical and Consent Issues	- Obtaining blood samples for CTC analysis requires informed consent - Collecting longitudinal samples to monitor disease progression may have psychological impacts on patients	[101]
Clinical Relevance	- Establishing the clinical relevance of CTCs and their role as prognostic or predictive biomarkers requires large-scale clinical validation - Research findings are not integrated with routine clinical applications	[102,103]
Cost and Accessibility	- Advanced technologies for CTC isolation and analysis are expensive - The cost associated with isolation and analysis methods limit their widespread implementation and use - Advanced technologies needed are not readily accessible in all healthcare settings	[104,105]

4. Circulating Tumor Cell Detection Strategies- Pros and Cons

The study of CTCs presents an opportunity to gain valuable insights in advancing cancer research and improving patient care. However, addressing the challenges with CTC research starts with effectively detecting CTCs in liquid biopsies. As such, there is a need to refine existing techniques and develop standardized protocols while making the process cost-effective and accessible to patients [106]. Admittedly, the methodologies used to detect or isolate CTCs have evolved significantly, becoming progressively sophisticated and sensitive [107–110]. Current CTC detecting methods can be categorized into label-dependent (affinity-based) and label-independent [12]. The latter involves traditional methods that separate cells based on size, deformability, and other biophysical properties [111,112]. These methods range from density gradient centrifugation, inertia focusing, filtration, and dielectrophoresis [113,114]. The challenge is that these methods lack the sensitivity and specificity needed to isolate CTCs efficiently [97].

Label-dependent methods detect cells based on the expression of specific markers like EpCAM, vimentin, and N-cadherin (positive selection) or the lack thereof of markers like CD45 (negative selection) and the binding affinity to specific antigens, RNA, and DNA sequences [97,115,116]. Such methods include EpCAM enrichment, immunomagnetic separation, and microfluidic devices [117–119]. By far, the cell surface adhesion molecule, EpCAM, is the most common marker used for CTC enrichment (increasing CTC concentration for their subsequent detection) [120]. Recent studies have focused on improving CTC enrichment through highly engineered microfluidic devices and selective CTC markers [108,120]. To date, label-dependent methods offer improved sensitivity and specificity, enabling the capture of rare CTCs while minimizing contamination from normal blood cells [121–123]. Nonetheless, the lack of universal markers for detecting heterogeneous CTCs coupled with the unavailability of validated and standardized approaches means most of these approaches need to be validated and standardized in the pre-analytical, analytical, and post-analytical phases [124]. Table 2

displays some methods and their principles for detecting CTCs and their advantages and disadvantages.

Table 2. CTC isolation methods—advantages and disadvantages.

Method	Principle	Pros	Cons	Refs
Density Gradient Centrifugation	Differential centrifugation separates blood components based on their density, allowing for the isolation of CTCs	Simple, cost-effective	- Limited specificity - Contamination from normal blood cells	[125]
Filtration Techniques	Filters with defined pore sizes are used to physically separate CTCs from blood cells based on size	Simple, cost-effective	- Risk of clogging - Loss of smaller CTCs	[126 – 128]
Epithelial Cell Adhesion Molecule (EpCAM) Enrichment	EpCAM, a cell surface marker often expressed in epithelial cancers, is targeted for CTC enrichment	Commonly used, FDA-approved platforms	- EpCAM-negative CTCs may be missed - Loss of CTC heterogeneity	[116]
Immunomagnetic Separation	Antibodies specific to tumor-associated antigens are used to capture CTCs by attaching to magnetic beads	High specificity, potential for enrichment of viable CTCs	- Limited by the availability of specific antigens - Loss of CTC viability	[129, 130]
Microfluidic Devices	Microscale devices use various mechanisms, such as size-based filtration or antibody-coated surfaces, to isolate CTCs from blood	High throughput, potential for single-cell analysis, and minimal sample processing	- Requires efficient capture to avoid CTC damage - Requires standardization of devices	[121, 131]
Fluorescent-Activated Cell Sorting (FACS)	Fluorescently tagged cells are separated by flow cytometry	High specificity and throughput for CTC enrichment	- Requires a high number of input cells	[132, 133]

Commercialized single-cell isolation strategies such as fluorescence-activated cell sorting and droplet-based systems isolate single cells randomly, making isolating rare single cells difficult [16]. The Food and Drugs Authority (FDA) has approved CELLSEARCH for the enumeration of CTCs of epithelial origin (EpCAM+, CD45-, and cytokeratins 8, 18+, and/or 19+) in whole blood [134]. Other systems such as DEPArray [135], RareCytes CyterFinder [136], ALS Collector [137], and VyCap Puncher [138] can isolate rare single cells but lack the capacity for single-cell enrichment. A study by Yoshino and colleagues showed that target cells could be encapsulated by confocal laser-scanning microscopy following the introduction of a photopolymerized hydrogel, polyethylene glycol diacrylate (PEGDA) into cell entrapped on a microcavity array (MCA) [139]. In another study utilizing the MCA/gel-based cell manipulation (GCM), a size-selective CTC enrichment and a surface-independent CTC detection method (based on CellTracker Green-positive and CD45-negative) showed an increased recovery and effective detection of gastric cancer CTCs [140]. This GCM-based single-cell manipulation method is advantageous for extremely low-volume reactions due to the minimal water composition of hydrogel-encapsulated single cells [140]. These efforts are

necessary to decrease cell loss during sample handling, decrease white blood cell contamination, and increase the recovery of CTCs in cancer.

5. Advances in Characterizing CTC Biology and Clinical Implications

The study of CTCs is inherently challenging due to their rarity in the bloodstream. To overcome this challenge and make the most use of CTCs as valuable biomarkers, advanced technologies are essential for their efficient capture and precise analysis. Herein, we discuss recent progress that has been made in characterizing CTC biology.

5.1. Polymerase Chain Reaction (PCR)

Liquid biopsies contain CTCs and other tumor-derived materials shed into the bloodstream, such as circulating tumor nucleic acids, including DNA (ctDNA), RNA (ctRNA), exosomes, and other microvesicles [141–143]. ctDNA, derived from apoptotic or necrotic tumor cells and CTCs, can be used to predict changes to therapeutic response [144]. The FDA approved the Cobas EGFR mutational test for the management of therapy in lung cancer [145]. This was a result of the detection and mutational testing (Cobas EGFR mutation test) of ctDNA from NSCLC patients that showed the importance of ctDNA as a biomarker in cancer diagnosis and prognosis [146,147]. Different types of PCR methods have been developed with the aim of detecting and characterizing CTCs and ctDNA [148–151]. These methods continue to be improved to increase sensitivity and allow for accurate and quantitative analysis of CTCs [107]. The development of droplet digital PCR (ddPCR) has allowed for the partitioning of a sample into many “droplets,” enabling the amplification of rare mutations [149,152]. The ddPCR method offers several advantages over conventional quantitative and real-time PCR [153–156]. It has improved sensitivity [154], is less susceptible to PCR inhibitors [153], and allows for absolute quantification without the need for an external calibration curve [156]. CTCs can also be quantified independent of nucleic acid extraction using real-time quantitative PCR (Q-PCR) [151]. In the article by Mei and colleagues, they isolated CTCs using tag-DNA-modified CK19 antibody and magnetic beads conjugated with EpCAM antibody. They reported a detection rate of 92.3% in the clinical tumor blood samples, and by quantifying the tag-DNA that had immobilized on the tumor cells, they showed a correlation between CTC counts and tumor stage/ status [151]. As PCR methods improve, single-cell and next-generation sequencing can be integrated to enhance CTC characterization and their roles in cancer.

5.2. Single-Cell Analysis

Genomic and phenotypic characteristics of cells differ within a patient’s tumor (intratumor heterogeneity) and between tumors of the same type (intertumor heterogeneity) [157]. These heterogeneous differences contribute to treatment failure and disease recurrence as there is no one-size-fits-all treatment approach [81]. Evidence has shown that CTCs detected in patients represent an array of cell states with varying genetic and phenotypic profiles [93,158]. Therefore, advanced technologies like single-cell RNA sequencing (scRNA-seq), single-cell protein analysis, and imaging techniques are needed [159]. These techniques allow for detailed molecular and phenotypic characterization with the potential to reveal deep insights into intratumor heterogeneity and cellular plasticity and help to identify pathways activated in different cell states, details often masked by profiling pooled cells [159]. Traditional methods of CTC analyses have targeted the use of epithelial biomarkers whose expression may not be universal across all tumors [116]. Recent improvements in single-cell analysis have focused on enhancing sensitivity, reducing technical variabilities, increasing throughput, and upgrading data analysis tools [160–162]. Upgrades on single-cell analysis packages such as Seurat and Uniform Manifold Approximation and Projection (UMAP) have enabled a better clustering analysis and classification of CTCs into multiple subtypes [163,164]. Single-cell sequencing analysis of CTCs from SCLC patients showed a global increase in intratumoral heterogeneity (ITH), including a heterogeneous expression of potential therapeutic resistance pathways, like EMT, between different cellular subpopulations following treatment resistance [48]. Indeed, serial profiling

of the CTCs post-relapse confirmed an increased ITH, suggesting that treatment resistance was characterized by coexisting subpopulations of cells with heterogeneous gene expression, leading to multiple concurrent resistance mechanisms [48]. This finding effectively highlights the need for clinical efforts targeted at developing combination therapies for treatment-naïve SCLC tumors in order to increase initial response and counteract the emergence of ITH and diverse resistance mechanisms [48]. Another analysis of prostate CTCs revealed heterogeneity in signaling pathways like non-canonical Wnt signaling that could contribute to treatment failure or therapeutic resistance (antiandrogen resistance) [165]. By integrating single-cell and batch RNA sequencing, Zhang et al. found gene markers associated with CD8⁺ T cell infiltration (LGR5, CCR7, STC2, DEFB1, TYK2, SCARB1, AICDA, and ULBP2) in HNSCC, which can serve as predictors of disease prognosis and clinical treatment indicators [166]. These findings may change the way cancer patients are staged and risk-stratified and provide new prognostic approaches to the identification of targeted therapy and resistance mechanisms.

5.3. Next-Generation Sequencing (NGS)

High-throughput sequencing of CTC-derived DNA or RNA can allow for a comprehensive genomic and transcriptomic analysis to detect heterogeneous and shared mutational signatures within and between different cancer types [167,168]. This could provide better insight into a patient's genetic makeup, which in turn can guide clinicians for better disease management. The challenges in sequencing the CTC genome and transcriptome include low abundance in the bloodstream, making their isolation difficult [169], significant genetic heterogeneity compared to the primary tumor, which can affect the capture of the full spectrum of mutations present [15], and the need for sophisticated bioinformatics tools and computational resources for data analysis and interpretation [170]. However, improvements in the isolation and capture of CTCs, the ease of use of different analytical tools, and the removal of biases introduced during sample and sequencing library preparation are increasing the possibility of detecting rare mutations in CTCs [171,172]. By targeting mutational hotspots, a combination of whole genome amplification, PCR, and Sanger sequencing was used to examine point mutations in the KRAS, BRAF, and PIK3CA genes in CTCs of patients with CRC. The authors detected single-cell mutations in the genes. Interestingly, these mutations were not found in the corresponding tumors [173]. Gkoutela and colleagues used a comparative analysis of the methylation landscape of single and CTC clusters from breast cancer patients and mouse models to reveal similarity patterns to embryonic stem cells and identify pharmacological agents that can target clustering, blunt metastasis spreading, and suppress stemness [60]. They observed hypomethylation in the binding sites for OCT4, NANOG, SOX2, and SIN3A of CTC clusters similar to embryonic stem cells and identified an FDA-approved Na⁺/P⁺ ATPase inhibitor that enabled the dissociation of CTC clusters into single cells. Thus, they established a link between specific DNA methylation changes and the promotion of cancer stemness and metastasis and pointed to cluster-targeting compounds to suppress the spread of cancer [60]. In a study investigating somatic mutations present in CTCs from colorectal cancer patients, the frequencies of somatic mutations present in the CTCs correlated with prognostic markers and resembled mutational signatures present in the primary tumor [174]. These findings have resulted from significant improvements in the isolation and enrichment of CTCs and efficient whole genome amplification (WGA) and NGS workflows that reduce bias with improved coverage, sensitivity, and specificity [175,176]. As more targeted cancer panels become increasingly available, the possibility of detecting more actionable mutations will increase with lower costs and minimal requirements for specialized equipment [177].

5.4. In Situ Analysis

Techniques that enable in situ analysis of CTCs directly within the bloodstream or in patient-derived samples provide a real-time understanding of their behavior. The advantage of in situ analysis is that challenges with traditional CTC isolation methods that compromise CTC integrity are avoided. These approaches also eliminate potential biases introduced by ex vivo isolation and culture, offering a more accurate representation of CTC biology. CellCollector CANCER01 (DC01) is

an approach developed for the in vivo isolation of CTCs from the peripheral blood of cancer patients [178]. The DC01 device enables in vivo isolation and enumeration of CTCs by capturing EpCAM-positive CTCs through a detector placed in the peripheral vein of cancer patients via a cannula [179]. This device has been shown to be tolerated for 30 minutes in vivo by patients without any side effects [180]. In a study evaluating the feasibility of CTC detection and monitoring in high-risk non-metastatic prostate cancer (PCa) patients undergoing radiotherapy, the authors show that CTCs could be detected before and after radiotherapy using the DC01 device in vivo [180]. Figure 2a below shows images of the captured CTCs. The CTCs were positive for pan-cytokeratin (pan-CK; green) and negative for CD45-red (Figure 2a). This study used the human prostate cancer cell line LNCaP as a positive control (Figure 2b), with non-specific binding lymphocytes as an internal negative control (Figure 2b,c). Three CTC clusters, each containing 2-3 CTCs, were detected in samples from two patients having a total CTC count of 10 and 15, respectively (Figure 2d).

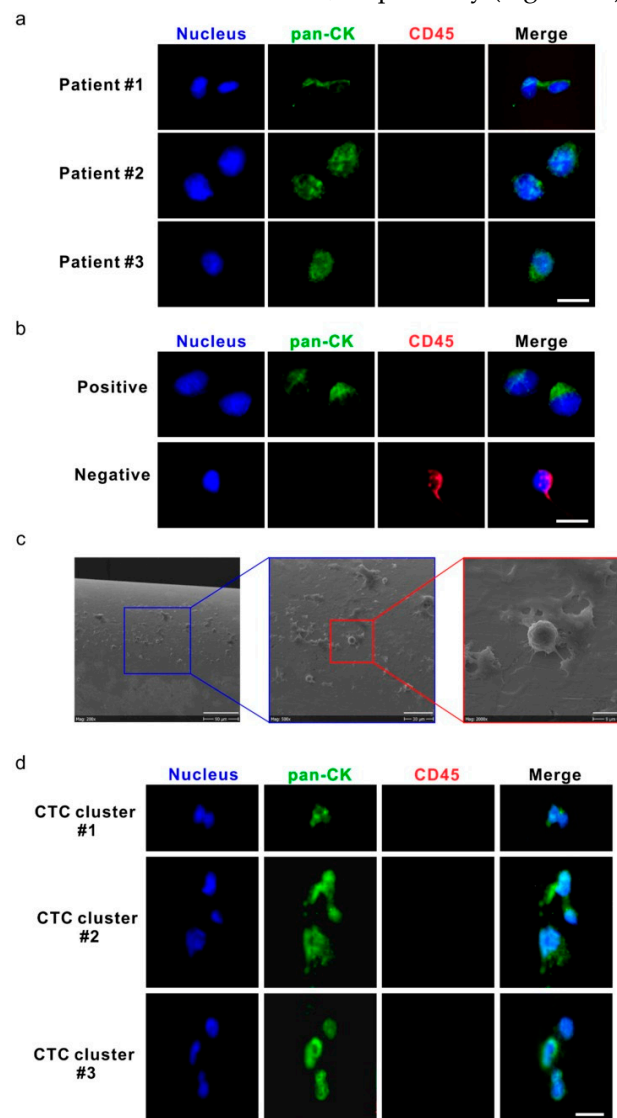


Figure 2. Circulating tumor cells (CTCs) and CTC clusters were detected by the DC01 device in patients with high-risk non-metastatic prostate cancer (PCa). (a) Micrographs of five CTCs on the DC01 detected from three patients; CTCs were defined as being positive for nucleic staining and pan-CK (pan-cytokeratin), and negative for CD45 by immunofluorescence staining (DNA (blue) and pan-CK (green)). Scale bar: 20 μ m. (b) Images of control samples for CTC staining panel, upper panel: LNCaP cells captured on the DC01 as positive control for CTC staining panel by immunofluorescence staining (DNA (blue) and pan-CK (green)); lower panel: leukocytes attached to the DC01 served as negative controls for the CTC staining panel by immunofluorescence staining (DNA (blue) and CD45 (red)). Scale bar: 20 μ m. (c) Scanning electron microscope image showing the surface of DC01 and

blood components, including unspecific binding of leucocytes onto DC01. Scale bar (from left to right): 90 μm , 30 μm , and 9 μm . (d) Three CTC clusters on the DC01 detected from two patients; CTC clusters were found as a cluster of CTCs being positive for nucleic staining and pan-CK, and negative for CD45 by immunofluorescence staining (DNA (blue) and pan-CK (green)). All CTCs were negative for PSA, so this channel is not shown. Exposure time for each channel: nucleus DNA, 800~1000 ms; pan-CK, 3000 ms; CD45, 6000 ms. Scale bar: 20 μm . Source: Reprinted from Chen, S., et al., *In Vivo Detection of Circulating Tumor Cells in High-Risk Non-Metastatic Prostate Cancer Patients Undergoing Radiotherapy*. *Cancers* (Basel), 2019. **11**(7) [180] under an open-access Creative Commons CC BY 4.0 license.

In the medical community, simple, easy, and sensitive methods to monitor tumor growth are essential for patient care [181]. These may come in the form of improvements or modifications to existing technology, allowing for economical and effective characterization of CTCs. For instance, confocal line scanning, which can be implemented in any confocal microscope system, helped detect CTC and CTC clusters in vivo in the blood vessels of live mice [182]. Kuo and colleagues also used multi-photon microscopy and antibody-conjugated quantum dots to image CTCs in real-time in living animals [59]. They detected different subpopulations of CTCs, especially those with high metastatic potential—CD24⁺ and CD133⁺ CTCs [59]. These simple and direct noninvasive strategies are needed to detect and investigate the mechanistic underpinnings of tumor metastasis and functionally characterize CTCs.

5.5. Functional Assays

Ex vivo culture of CTCs and CTC-derived animal models are gaining increasing popularity as tools to study functional properties, such as drug response and metastatic potential [14,183,184]. These models provide insights into CTC behavior beyond molecular characterization [98]. Additionally, functional characterization of CTCs provides insights into the potential treatment options [184]. However, there is a lack of standardized protocols for culturing CTCs of different cancer cell types, and optimal conditions are undefined [185,186]. Current culture conditions involve a cocktail of growth factors like epidermal growth factor (EGF) and fibroblast growth factors (FGF2 and FGF10), cytokines, hormones, tissue organ extracts, and insulin/insulin-like growth factor 1 (IGF-1) [187,188]. In a recent study optimizing cell culture conditions using a model of primary ovarian cells, Cobalt(II) chloride (CoCl₂) was used as a hypoxia-mimicking agent [14]. A CoCl₂ concentration between 100 and 150 μM in culture media was shown to lead to a significant increase in cell counts, demonstrative of a sustainable proliferative activity. However, when the concentration of CoCl₂ was increased to 200 μM , there was a significant cell cycle arrest at G0/G1 [14]. Efforts like this highlight the importance of refining cell culture conditions to ensure optimal cell growth. Regardless, once successful cultures have been established, the cells can be characterized to provide insight into their biology. Figure 3 shows a phenotypic characterization of CTCs after ~2 weeks in culture [183]. Expression of epithelial markers was observed in 18.18% of samples (number of samples, n = 22), while 47% were vimentin-positive (n = 17) [183]. These findings corroborate existing knowledge on the loss of epithelial features and the gain of mesenchymal properties following cell culture [189–191].

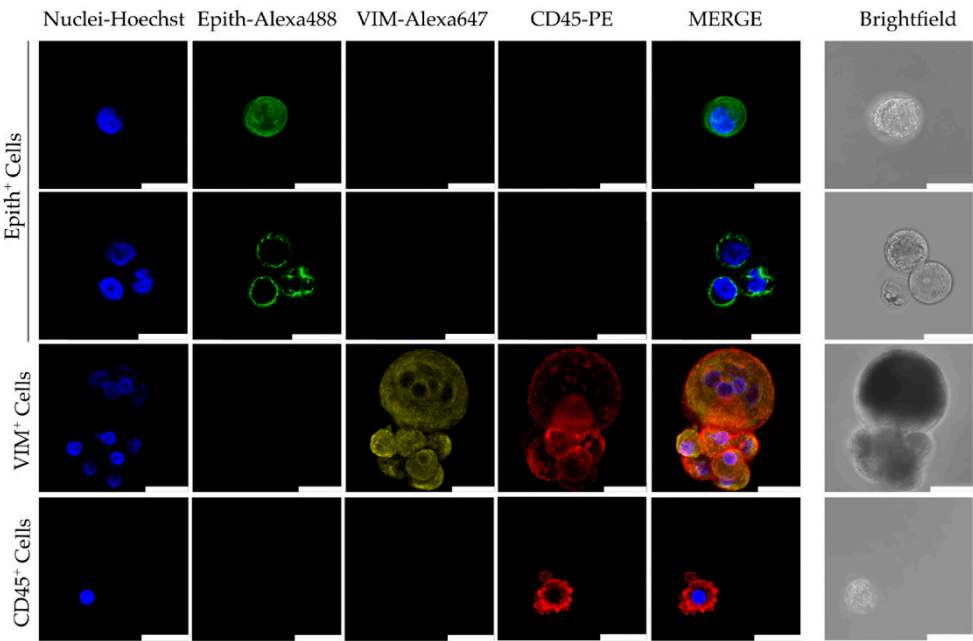


Figure 3. Representative images by confocal microscopy of Immunofluorescence characterization of Circulating Tumor Cells (CTCs) after culture. Immunofluorescent staining was performed using a combination of anti-human Epithelial markers (Epith: EpCAM, E-Cadh, and PanCK) (in green), anti-Vimentin (VM, in yellow), and anti-CD45 (in red). Scale bar represents 25 μm . Source: Reprinted from Carmona-Ule, N., et al., *Short-Term Ex Vivo Culture of CTCs from Advance Breast Cancer Patients: Clinical Implications*. Cancers (Basel), 2021. 13(11) [183] under an open-access Creative Commons CC BY 4.0 license.

Expansion of CTCs ex vivo via cell cultures also provides avenues for drug sensitivity testing and allows for functional assays like proliferation, migration, invasion, and apoptosis to be performed. In a study performing drug sensitivity profiling of cultured CTCs (treatment naïve) from patients with SCLC, the authors reported a concordance between response to standard-care chemotherapy (cisplatin and etoposide) administered to the patients and the sensitivities of corresponding cultured CTCs treated with cisplatin and etoposide [184]. Following treatment, response to chemotherapy can also be assessed. CTCs can be evaluated for features of viability, early apoptosis, and late-stage apoptosis/necrosis [192]. In breast cancer, CTCs with features of early apoptosis are prognostic of metastasis-free survival and correlate with neoadjuvant chemotherapy response, while the presence of late-stage apoptotic CTCs are associated with poor response to neoadjuvant chemotherapy and metastasis-free survival [192]. Such findings support the feasibility of predicting treatment outcomes in patients using CTCs.

6. Future Perspectives

CTCs are present in very low numbers compared to other blood cells, making their isolation akin to finding a needle in a haystack. Therefore, advanced technologies with high sensitivity are required to capture and detect these rare cells efficiently. Real-time monitoring of CTC dynamics, capturing changes in CTC numbers and characteristics over time, is crucial for understanding the temporal aspects of cancer progression, treatment response, and the emergence of resistance. The development of new high-throughput technologies allows for the rapid and automated processing of large volumes of blood, enhancing the efficiency of CTC isolation. This is particularly important for clinical applications where timely results are essential. Integrating CTC research findings into routine clinical practice, however, requires standardization, validation, and collaboration between researchers and healthcare professionals. These initiatives should be focused on the validation of CTC-based assays, ensuring their reliability and reproducibility. Additionally, making CTC analysis

clinically relevant requires the adaptability of existing or new technologies to routine clinical practice. Accordingly, user-friendly platforms are needed to be integrated with existing diagnostic workflows and point-of-care systems. The integration of advanced analytical platforms, such as single-cell sequencing and next-generation sequencing, will enable comprehensive profiling of CTCs and increase accessibility while significantly decreasing time and associated costs. Thus, the use of CTCs in cancer diagnosis, prognosis, and monitoring treatment outcomes has the potential to revolutionize our approach to cancer detection and treatment.

In summary, recent advances in CTC research have deepened our understanding of their multifaceted roles in cancer progression, metastasis, and treatment resistance, contributing to developing more effective diagnostic and therapeutic strategies. These breakthroughs are translating into practical applications with direct benefits for cancer patients. As these findings become more integrated into routine clinical practice, the promise of CTC-based approaches lies in improving cancer diagnosis, prognosis, and treatment outcomes for individuals across diverse cancer types.

Author Contributions: Literature review, writing—original draft preparation, P.S.; review, editing, and funding acquisition, K.M.R. All authors have read and agreed to the published version of the manuscript.

Funding: This research was funded by NIH, grant numbers 1R15CA249714-01 and 1P20GM109024-01A1.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Conflicts of Interest: The authors declare no conflicts of interest.

References

1. Siegel, R.L.; Miller, K.D.; Wagle, N.S.; Jemal, A. Cancer statistics, 2023. *CA Cancer J Clin* **2023**, *73*, 17-48, doi:10.3322/caac.21763.
2. Vasan, N.; Baselga, J.; Hyman, D.M. A view on drug resistance in cancer. *Nature* **2019**, *575*, 299-309, doi:10.1038/s41586-019-1730-1.
3. Fares, J.; Fares, M.Y.; Khachfe, H.H.; Salhab, H.A.; Fares, Y. Molecular principles of metastasis: a hallmark of cancer revisited. *Signal Transduct Target Ther* **2020**, *5*, 28, doi:10.1038/s41392-020-0134-x.
4. Schiffman, J.D.; Fisher, P.G.; Gibbs, P. Early detection of cancer: past, present, and future. *Am Soc Clin Oncol Educ Book* **2015**, 57-65, doi:10.14694/EdBook_AM.2015.35.57.
5. Connal, S.; Cameron, J.M.; Sala, A.; Brennan, P.M.; Palmer, D.S.; Palmer, J.D.; Perlow, H.; Baker, M.J. Liquid biopsies: the future of cancer early detection. *J Transl Med* **2023**, *21*, 118, doi:10.1186/s12967-023-03960-8.
6. Ried, K.; Eng, P.; Sali, A. Screening for Circulating Tumour Cells Allows Early Detection of Cancer and Monitoring of Treatment Effectiveness: An Observational Study. *Asian Pac J Cancer Prev* **2017**, *18*, 2275-2285, doi:10.22034/APJCP.2017.18.8.2275.
7. Suvilesh, K.N.; Manjunath, Y.; Pantel, K.; Kaifi, J.T. Preclinical models to study patient-derived circulating tumor cells and metastasis. *Trends Cancer* **2023**, *9*, 355-371, doi:10.1016/j.trecan.2023.01.004.
8. Marcuello, M.; Vymetalkova, V.; Neves, R.P.L.; Duran-Sanchon, S.; Vedeld, H.M.; Tham, E.; van Dalum, G.; Flugen, G.; Garcia-Barberan, V.; Fijneman, R.J.; et al. Circulating biomarkers for early detection and clinical management of colorectal cancer. *Mol Aspects Med* **2019**, *69*, 107-122, doi:10.1016/j.mam.2019.06.002.
9. Castro-Giner, F.; Aceto, N. Tracking cancer progression: from circulating tumor cells to metastasis. *Genome Med* **2020**, *12*, 31, doi:10.1186/s13073-020-00728-3.
10. Micalizzi, D.S.; Maheswaran, S.; Haber, D.A. A conduit to metastasis: circulating tumor cell biology. *Genes Dev* **2017**, *31*, 1827-1840, doi:10.1101/gad.305805.117.
11. Liu, J.; Lian, J.; Chen, Y.; Zhao, X.; Du, C.; Xu, Y.; Hu, H.; Rao, H.; Hong, X. Circulating Tumor Cells (CTCs): A Unique Model of Cancer Metastases and Non-invasive Biomarkers of Therapeutic Response. *Front Genet* **2021**, *12*, 734595, doi:10.3389/fgene.2021.734595.
12. Ju, S.; Chen, C.; Zhang, J.; Xu, L.; Zhang, X.; Li, Z.; Chen, Y.; Zhou, J.; Ji, F.; Wang, L. Detection of circulating tumor cells: opportunities and challenges. *Biomark Res* **2022**, *10*, 58, doi:10.1186/s40364-022-00403-2.
13. Asante, D.B.; Mohan, G.; Acheampong, E.; Ziman, M.; Calapre, L.; Meniawy, T.M.; Gray, E.S.; Beasley, A.B. Genetic analysis of heterogeneous subsets of circulating tumour cells from high grade serous ovarian carcinoma patients. *Sci Rep* **2023**, *13*, 2552, doi:10.1038/s41598-023-29416-z.

14. Mohamed, B.M.; Ward, M.P.; Bates, M.; Spillane, C.D.; Kelly, T.; Martin, C.; Gallagher, M.; Heffernan, S.; Norris, L.; Kennedy, J.; et al. Ex vivo expansion of circulating tumour cells (CTCs). *Sci Rep* **2023**, *13*, 3704, doi:10.1038/s41598-023-30733-6.
15. Menyailo, M.E.; Tretyakova, M.S.; Denisov, E.V. Heterogeneity of Circulating Tumor Cells in Breast Cancer: Identifying Metastatic Seeds. *Int J Mol Sci* **2020**, *21*, doi:10.3390/ijms21051696.
16. Edd, J.F.; Mishra, A.; Smith, K.C.; Kapur, R.; Maheswaran, S.; Haber, D.A.; Toner, M. Isolation of circulating tumor cells. *iScience* **2022**, *25*, 104696, doi:10.1016/j.isci.2022.104696.
17. Wen, Z.; Li, Z.; Yong, P.; Liang, D.; Xie, D.; Chen, H.; Yang, Y.; Wu, S.; Li, C.; Cheng, Z. Detection and clinical significance of circulating tumor cells in patients with nasopharyngeal carcinoma. *Oncol Lett* **2019**, *18*, 2537-2547, doi:10.3892/ol.2019.10560.
18. Jiang, M.; Jin, S.; Han, J.; Li, T.; Shi, J.; Zhong, Q.; Li, W.; Tang, W.; Huang, Q.; Zong, H. Detection and clinical significance of circulating tumor cells in colorectal cancer. *Biomark Res* **2021**, *9*, 85, doi:10.1186/s40364-021-00326-4.
19. Schochter, F.; Friedl, T.W.P.; deGregorio, A.; Krause, S.; Huober, J.; Rack, B.; Janni, W. Are Circulating Tumor Cells (CTCs) Ready for Clinical Use in Breast Cancer? An Overview of Completed and Ongoing Trials Using CTCs for Clinical Treatment Decisions. *Cells* **2019**, *8*, doi:10.3390/cells8111412.
20. Muchlinska, A.; Smentoch, J.; Zaczek, A.J.; Bednarz-Knoll, N. Detection and Characterization of Circulating Tumor Cells Using Imaging Flow Cytometry-A Perspective Study. *Cancers (Basel)* **2022**, *14*, doi:10.3390/cancers14174178.
21. Yang, J.; Huang, X.; Gan, C.; Yuan, R.; Xiang, Y. Highly specific and sensitive point-of-care detection of rare circulating tumor cells in whole blood via a dual recognition strategy. *Biosens Bioelectron* **2019**, *143*, 111604, doi:10.1016/j.bios.2019.111604.
22. Thiele, J.A.; Pitule, P.; Hicks, J.; Kuhn, P. Single-Cell Analysis of Circulating Tumor Cells. *Methods Mol Biol* **2019**, *1908*, 243-264, doi:10.1007/978-1-4939-9004-7_17.
23. Chen, J.Y.; Chou, H.H.; Lim, S.C.; Huang, Y.J.; Lai, K.C.; Guo, C.L.; Tung, C.Y.; Su, C.T.; Wang, J.; Liu, E.; et al. Multiomic characterization and drug testing establish circulating tumor cells as an ex vivo tool for personalized medicine. *iScience* **2022**, *25*, 105081, doi:10.1016/j.isci.2022.105081.
24. Guo, Z.; Lin, X.; Hui, Y.; Wang, J.; Zhang, Q.; Kong, F. Circulating Tumor Cell Identification Based on Deep Learning. *Front Oncol* **2022**, *12*, 843879, doi:10.3389/fonc.2022.843879.
25. He, B.; Lu, Q.; Lang, J.; Yu, H.; Peng, C.; Bing, P.; Li, S.; Zhou, Q.; Liang, Y.; Tian, G. A New Method for CTC Images Recognition Based on Machine Learning. *Front Bioeng Biotechnol* **2020**, *8*, 897, doi:10.3389/fbioe.2020.00897.
26. Lawrence, R.; Watters, M.; Davies, C.R.; Pantel, K.; Lu, Y.J. Circulating tumour cells for early detection of clinically relevant cancer. *Nat Rev Clin Oncol* **2023**, *20*, 487-500, doi:10.1038/s41571-023-00781-y.
27. Ashworth, T.R. A Case of Cancer in Which Cells Similar to Those in the Tumours Were Seen in the Blood after Death. *The Medical Journal of Australia* **1869**, *14*, 146-147.
28. Massague, J.; Obenauf, A.C. Metastatic colonization by circulating tumour cells. *Nature* **2016**, *529*, 298-306, doi:10.1038/nature17038.
29. Franes, J.W.; Basar, O.; Kadayifci, A.; Yuksel, O.; Choz, M.; Kulkarni, A.S.; Tai, E.; Vo, K.D.; Arora, K.S.; Desai, N.; et al. Improved Detection of Circulating Epithelial Cells in Patients with Intraductal Papillary Mucinous Neoplasms. *Oncologist* **2018**, *23*, 1260, doi:10.1634/theoncologist.2017-0234erratum.
30. Lu, S.H.; Tsai, W.S.; Chang, Y.H.; Chou, T.Y.; Pang, S.T.; Lin, P.H.; Tsai, C.M.; Chang, Y.C. Identifying cancer origin using circulating tumor cells. *Cancer Biol Ther* **2016**, *17*, 430-438, doi:10.1080/15384047.2016.1141839.
31. Tsai, W.S.; You, J.F.; Hung, H.Y.; Hsieh, P.S.; Hsieh, B.; Lenz, H.J.; Idos, G.; Friedland, S.; Yi-Jiun Pan, J.; Shao, H.J.; et al. Novel Circulating Tumor Cell Assay for Detection of Colorectal Adenomas and Cancer. *Clin Transl Gastroenterol* **2019**, *10*, e00088, doi:10.14309/ctg.0000000000000088.
32. Ilie, M.; Hofman, V.; Long-Mira, E.; Selva, E.; Vignaud, J.M.; Padovani, B.; Mouroux, J.; Marquette, C.H.; Hofman, P. "Sentinel" circulating tumor cells allow early diagnosis of lung cancer in patients with chronic obstructive pulmonary disease. *PLoS One* **2014**, *9*, e111597, doi:10.1371/journal.pone.0111597.
33. Xu, L.; Mao, X.; Grey, A.; Scandura, G.; Guo, T.; Burke, E.; Marzec, J.; Abdu, S.; Stankiewicz, E.; Davies, C.R.; et al. Noninvasive Detection of Clinically Significant Prostate Cancer Using Circulating Tumor Cells. *J Urol* **2020**, *203*, 73-82, doi:10.1097/JU.0000000000000475.

34. Matikas, A.; Kotsakis, A.; Apostolaki, S.; Politaki, H.; Perraki, M.; Kalbakis, K.; Nikolaou, M.; Economopoulou, P.; Hatzidaki, D.; Georgoulas, V. Detection of circulating tumour cells before and following adjuvant chemotherapy and long-term prognosis of early breast cancer. *Br J Cancer* **2022**, *126*, 1563-1569, doi:10.1038/s41416-022-01699-5.
35. Geng, N.; Chen, S.; Liu, J.; Cao, W.; Zhang, D.; Feng, C. Circulating tumor cells in blood as a prognostic biomarker in tongue squamous cell carcinoma. *Oral Surg Oral Med Oral Pathol Oral Radiol* **2022**, *134*, 213-219, doi:10.1016/j.oooo.2021.12.129.
36. Guan, Y.; Xu, F.; Tian, J.; Gao, K.; Wan, Z.; Wang, Y.; Gao, M.; Wang, Z.; Chong, T. The prognostic value of circulating tumour cells (CTCs) and CTC white blood cell clusters in patients with renal cell carcinoma. *BMC Cancer* **2021**, *21*, 826, doi:10.1186/s12885-021-08463-7.
37. Murlidhar, V.; Reddy, R.M.; Fouladdel, S.; Zhao, L.; Ishikawa, M.K.; Grabauskienė, S.; Zhang, Z.; Lin, J.; Chang, A.C.; Carrott, P.; et al. Poor Prognosis Indicated by Venous Circulating Tumor Cell Clusters in Early-Stage Lung Cancers. *Cancer Res* **2017**, *77*, 5194-5206, doi:10.1158/0008-5472.CAN-16-2072.
38. Qi, L.N.; Xiang, B.D.; Wu, F.X.; Ye, J.Z.; Zhong, J.H.; Wang, Y.Y.; Chen, Y.Y.; Chen, Z.S.; Ma, L.; Chen, J.; et al. Circulating Tumor Cells Undergoing EMT Provide a Metric for Diagnosis and Prognosis of Patients with Hepatocellular Carcinoma. *Cancer Res* **2018**, *78*, 4731-4744, doi:10.1158/0008-5472.CAN-17-2459.
39. Guan, Y.; Xu, F.; Tian, J.; Chen, H.; Yang, C.; Huang, S.; Gao, K.; Wan, Z.; Li, M.; He, M.; et al. Pathology of circulating tumor cells and the available capture tools (Review). *Oncol Rep* **2020**, *43*, 1355-1364, doi:10.3892/or.2020.7533.
40. Cristofanilli, M.; Pierga, J.Y.; Reuben, J.; Rademaker, A.; Davis, A.A.; Peeters, D.J.; Fehm, T.; Nole, F.; Gisbert-Criado, R.; Mavroudis, D.; et al. The clinical use of circulating tumor cells (CTCs) enumeration for staging of metastatic breast cancer (MBC): International expert consensus paper. *Crit Rev Oncol Hematol* **2019**, *134*, 39-45, doi:10.1016/j.critrevonc.2018.12.004.
41. Magbanua, M.J.M.; Savenkov, O.; Asmus, E.J.; Ballman, K.V.; Scott, J.H.; Park, J.W.; Dickler, M.; Partridge, A.; Carey, L.A.; Winer, E.P.; et al. Clinical Significance of Circulating Tumor Cells in Hormone Receptor-positive Metastatic Breast Cancer Patients who Received Letrozole with or Without Bevacizumab. *Clin Cancer Res* **2020**, *26*, 4911-4920, doi:10.1158/1078-0432.CCR-20-1329.
42. Wang, P.-P.; Liu, S.-H.; Chen, C.-T.; Lv, L.; Li, D.; Liu, Q.-Y.; Liu, G.-L.; Wu, Y. Circulating tumor cells as a new predictive and prognostic factor in patients with small cell lung cancer. *Journal of Cancer* **2020**, *11*, 2113.
43. Beasley, A.B.; Isaacs, T.W.; Vermeulen, T.; Freeman, J.; DeSousa, J.L.; Bhikoo, R.; Hennessy, D.; Reid, A.; Chen, F.K.; Bentel, J.; et al. Analysis of Circulating Tumour Cells in Early-Stage Uveal Melanoma: Evaluation of Tumour Marker Expression to Increase Capture. *Cancers (Basel)* **2021**, *13*, doi:10.3390/cancers13235990.
44. Beasley, A.; Isaacs, T.; Khattak, M.A.; Freeman, J.B.; Allcock, R.; Chen, F.K.; Pereira, M.R.; Yau, K.; Bentel, J.; Vermeulen, T.; et al. Clinical Application of Circulating Tumor Cells and Circulating Tumor DNA in Uveal Melanoma. *JCO Precis Oncol* **2018**, *2*, doi:10.1200/PO.17.00279.
45. Vasantharajan, S.S.; Eccles, M.R.; Rodger, E.J.; Pattison, S.; McCall, J.L.; Gray, E.S.; Calapre, L.; Chatterjee, A. The Epigenetic landscape of Circulating tumour cells. *Biochim Biophys Acta Rev Cancer* **2021**, *1875*, 188514, doi:10.1016/j.bbcan.2021.188514.
46. Freed, I.M.; Kasi, A.; Fateru, O.; Hu, M.; Gonzalez, P.; Weatherington, N.; Pathak, H.; Hyter, S.; Sun, W.; Al-Rajabi, R.; et al. Circulating Tumor Cell Subpopulations Predict Treatment Outcome in Pancreatic Ductal Adenocarcinoma (PDAC) Patients. *Cells* **2023**, *12*, doi:10.3390/cells12182266.
47. Cappelletti, V.; Verzoni, E.; Ratta, R.; Vismara, M.; Silvestri, M.; Montone, R.; Miodini, P.; Reduzzi, C.; Claps, M.; Sepe, P.; et al. Analysis of Single Circulating Tumor Cells in Renal Cell Carcinoma Reveals Phenotypic Heterogeneity and Genomic Alterations Related to Progression. *Int J Mol Sci* **2020**, *21*, doi:10.3390/ijms21041475.
48. Stewart, C.A.; Gay, C.M.; Xi, Y.; Sivajothi, S.; Sivakamasundari, V.; Fujimoto, J.; Bolisetty, M.; Hartsfield, P.M.; Balasubramanian, V.; Chalisehar, M.D.; et al. Single-cell analyses reveal increased intratumoral heterogeneity after the onset of therapy resistance in small-cell lung cancer. *Nat Cancer* **2020**, *1*, 423-436, doi:10.1038/s43018-019-0020-z.
49. Hanssen, A.; Wagner, J.; Gorges, T.M.; Taenzer, A.; Uzunoglu, F.G.; Driemel, C.; Stoecklein, N.H.; Knoefel, W.T.; Angenendt, S.; Hauch, S.; et al. Characterization of different CTC subpopulations in non-small cell lung cancer. *Sci Rep* **2016**, *6*, 28010, doi:10.1038/srep28010.

50. Salu, P.; Reindl, K.M. Advancements in Preclinical Models of Pancreatic Cancer. *Pancreas* **2024**, *53*, e205-e220, doi:10.1097/MPA.0000000000002277.
51. Seyfried, T.N.; Huysentruyt, L.C. On the origin of cancer metastasis. *Crit Rev Oncog* **2013**, *18*, 43-73, doi:10.1615/critrevoncog.v18.i1-2.40.
52. Dillekas, H.; Rogers, M.S.; Straume, O. Are 90% of deaths from cancer caused by metastases? *Cancer Med* **2019**, *8*, 5574-5576, doi:10.1002/cam4.2474.
53. Suhail, Y.; Cain, M.P.; Vanaja, K.; Kurywachak, P.A.; Levchenko, A.; Kalluri, R.; Kshitiz. Systems Biology of Cancer Metastasis. *Cell Syst* **2019**, *9*, 109-127, doi:10.1016/j.cels.2019.07.003.
54. Parsons, S.; Maldonado, E.B.; Prasad, V. Comparison of Drugs Used for Adjuvant and Metastatic Therapy of Colon, Breast, and Non-Small Cell Lung Cancers. *JAMA Netw Open* **2020**, *3*, e202488, doi:10.1001/jamanetworkopen.2020.2488.
55. Ganesh, K.; Massague, J. Targeting metastatic cancer. *Nat Med* **2021**, *27*, 34-44, doi:10.1038/s41591-020-01195-4.
56. Lambert, A.W.; Pattabiraman, D.R.; Weinberg, R.A. Emerging Biological Principles of Metastasis. *Cell* **2017**, *168*, 670-691, doi:10.1016/j.cell.2016.11.037.
57. Massague, J.; Ganesh, K. Metastasis-Initiating Cells and Ecosystems. *Cancer Discov* **2021**, *11*, 971-994, doi:10.1158/2159-8290.CD-21-0010.
58. Qayyumi, B.; Bharde, A.; Aland, G.; D'Souza, A.; Jayant, S.; Singh, N.; Tripathi, S.; Badave, R.; Kale, N.; Singh, B.; et al. Circulating tumor cells as a predictor for poor prognostic factors and overall survival in treatment naive oral squamous cell carcinoma patients. *Oral Surg Oral Med Oral Pathol Oral Radiol* **2022**, *134*, 73-83, doi:10.1016/j.oooo.2022.02.018.
59. Kuo, C.W.; Chueh, D.Y.; Chen, P. Real-time in vivo imaging of subpopulations of circulating tumor cells using antibody conjugated quantum dots. *J Nanobiotechnology* **2019**, *17*, 26, doi:10.1186/s12951-019-0453-7.
60. Gkoutela, S.; Castro-Giner, F.; Szczerba, B.M.; Vetter, M.; Landin, J.; Scherrer, R.; Krol, I.; Scheidmann, M.C.; Beisel, C.; Stirnimann, C.U.; et al. Circulating Tumor Cell Clustering Shapes DNA Methylation to Enable Metastasis Seeding. *Cell* **2019**, *176*, 98-112 e114, doi:10.1016/j.cell.2018.11.046.
61. Aceto, N.; Bardia, A.; Miyamoto, D.T.; Donaldson, M.C.; Wittner, B.S.; Spencer, J.A.; Yu, M.; Pely, A.; Engstrom, A.; Zhu, H.; et al. Circulating tumor cell clusters are oligoclonal precursors of breast cancer metastasis. *Cell* **2014**, *158*, 1110-1122, doi:10.1016/j.cell.2014.07.013.
62. Jansson, S.; Bendahl, P.O.; Larsson, A.M.; Aaltonen, K.E.; Ryden, L. Prognostic impact of circulating tumor cell apoptosis and clusters in serial blood samples from patients with metastatic breast cancer in a prospective observational cohort. *BMC Cancer* **2016**, *16*, 433, doi:10.1186/s12885-016-2406-y.
63. Yu, M. Metastasis Stemming from Circulating Tumor Cell Clusters. *Trends Cell Biol* **2019**, *29*, 275-276, doi:10.1016/j.tcb.2019.02.001.
64. Wang, X.; Sun, Q.; Liu, Q.; Wang, C.; Yao, R.; Wang, Y. CTC immune escape mediated by PD-L1. *Med Hypotheses* **2016**, *93*, 138-139, doi:10.1016/j.mehy.2016.05.022.
65. Liu, Q.; Liao, Q.; Zhao, Y. Myeloid-derived suppressor cells (MDSC) facilitate distant metastasis of malignancies by shielding circulating tumor cells (CTC) from immune surveillance. *Med Hypotheses* **2016**, *87*, 34-39, doi:10.1016/j.mehy.2015.12.007.
66. Zhang, W.; Xu, F.; Yao, J.; Mao, C.; Zhu, M.; Qian, M.; Hu, J.; Zhong, H.; Zhou, J.; Shi, X.; et al. Single-cell metabolic fingerprints discover a cluster of circulating tumor cells with distinct metastatic potential. *Nat Commun* **2023**, *14*, 2485, doi:10.1038/s41467-023-38009-3.
67. Bergers, G.; Fendt, S.M. The metabolism of cancer cells during metastasis. *Nat Rev Cancer* **2021**, *21*, 162-180, doi:10.1038/s41568-020-00320-2.
68. Jie, X.X.; Zhang, X.Y.; Xu, C.J. Epithelial-to-mesenchymal transition, circulating tumor cells and cancer metastasis: Mechanisms and clinical applications. *Oncotarget* **2017**, *8*, 81558-81571, doi:10.18632/oncotarget.18277.
69. Nagel, A.; Markiewicz, A.; Szade, J.; Majewska, H.; Skokowski, J.; Seroczynska, B.; Welnicka-Jaskiewicz, M.; Zaczek, A. Expression of stem cell and mesenchymal markers in circulating tumor cells is associated with poor prognosis of early breast cancer patients. *Annals of Oncology* **2017**, *28*, vii32.
70. Werner, S.; Stenzl, A.; Pantel, K.; Todenhofer, T. Expression of Epithelial Mesenchymal Transition and Cancer Stem Cell Markers in Circulating Tumor Cells. *Adv Exp Med Biol* **2017**, *994*, 205-228, doi:10.1007/978-3-319-55947-6_11.

71. Hirahata, T.; Ul Quraish, R.; Quraish, A.U.; Ul Quraish, S.; Naz, M.; Razzaq, M.A. Liquid Biopsy: A Distinctive Approach to the Diagnosis and Prognosis of Cancer. *Cancer Inform* **2022**, *21*, 11769351221076062, doi:10.1177/11769351221076062.
72. Hodgkinson, C.L.; Morrow, C.J.; Li, Y.; Metcalf, R.L.; Rothwell, D.G.; Trapani, F.; Polanski, R.; Burt, D.J.; Simpson, K.L.; Morris, K.; et al. Tumorigenicity and genetic profiling of circulating tumor cells in small-cell lung cancer. *Nat Med* **2014**, *20*, 897-903, doi:10.1038/nm.3600.
73. Tada, H.; Takahashi, H.; Kawabata-Iwakawa, R.; Nagata, Y.; Uchida, M.; Shino, M.; Ida, S.; Mito, I.; Matsuyama, T.; Chikamatsu, K. Molecular phenotypes of circulating tumor cells and efficacy of nivolumab treatment in patients with head and neck squamous cell carcinoma. *Sci Rep* **2020**, *10*, 21573, doi:10.1038/s41598-020-78741-0.
74. Li, Z.; Xu, K.; Tartarone, A.; Santarpia, M.; Zhu, Y.; Jiang, G. Circulating tumor cells can predict the prognosis of patients with non-small cell lung cancer after resection: a retrospective study. *Transl Lung Cancer Res* **2021**, *10*, 995-1006, doi:10.21037/tlcr-21-149.
75. Mu, H.; Zuo, D.; Chen, J.; Liu, Z.; Wang, Z.; Yang, L.; Shi, Q.; Hua, Y. Detection and surveillance of circulating tumor cells in osteosarcoma for predicting therapy response and prognosis. *Cancer Biol Med* **2022**, *19*, 1397-1409, doi:10.20892/j.issn.2095-3941.2022.0279.
76. Neophytou, C.M.; Kyriakou, T.C.; Papageorgis, P. Mechanisms of Metastatic Tumor Dormancy and Implications for Cancer Therapy. *Int J Mol Sci* **2019**, *20*, doi:10.3390/ijms20246158.
77. Leone, K.; Poggiana, C.; Zamarchi, R. The Interplay between Circulating Tumor Cells and the Immune System: From Immune Escape to Cancer Immunotherapy. *Diagnostics (Basel)* **2018**, *8*, doi:10.3390/diagnostics8030059.
78. Yadav, A.S.; Pandey, P.R.; Butti, R.; Radharani, N.N.V.; Roy, S.; Bhalara, S.R.; Gorain, M.; Kundu, G.C.; Kumar, D. The Biology and Therapeutic Implications of Tumor Dormancy and Reactivation. *Front Oncol* **2018**, *8*, 72, doi:10.3389/fonc.2018.00072.
79. Pradhan, S.; Sperduto, J.L.; Farino, C.J.; Slater, J.H. Engineered In Vitro Models of Tumor Dormancy and Reactivation. *J Biol Eng* **2018**, *12*, 37, doi:10.1186/s13036-018-0120-9.
80. Dudgeon, C.; Harris, C.R.; Chen, Y.; Ghaddar, B.; Sharma, A.; Shah, M.M.; Roberts, A.I.; Casabianca, A.; Collisson, E.A.; Balachandran, V.P. A novel model of pancreatic cancer dormancy reveals mechanistic insights and a dormancy gene signature with human relevance. *BioRxiv* **2020**, 2020.2004.2013.037374.
81. Dagogo-Jack, I.; Shaw, A.T. Tumour heterogeneity and resistance to cancer therapies. *Nat Rev Clin Oncol* **2018**, *15*, 81-94, doi:10.1038/nrclinonc.2017.166.
82. Aboulkheyr Es, H.; Montazeri, L.; Aref, A.R.; Vosough, M.; Baharvand, H. Personalized Cancer Medicine: An Organoid Approach. *Trends Biotechnol* **2018**, *36*, 358-371, doi:10.1016/j.tibtech.2017.12.005.
83. Tyner, J.W.; Tognon, C.E.; Bottomly, D.; Wilmot, B.; Kurtz, S.E.; Savage, S.L.; Long, N.; Schultz, A.R.; Traer, E.; Abel, M.; et al. Functional genomic landscape of acute myeloid leukaemia. *Nature* **2018**, *562*, 526-531, doi:10.1038/s41586-018-0623-z.
84. Dai, W.; Zheng, H.; Cheung, A.K.; Tang, C.S.; Ko, J.M.; Wong, B.W.; Leong, M.M.; Sham, P.C.; Cheung, F.; Kwong, D.L.; et al. Whole-exome sequencing identifies MST1R as a genetic susceptibility gene in nasopharyngeal carcinoma. *Proc Natl Acad Sci U S A* **2016**, *113*, 3317-3322, doi:10.1073/pnas.1523436113.
85. Chang, Y.; Wang, Y.; Li, B.; Lu, X.; Wang, R.; Li, H.; Yan, B.; Gu, A.; Wang, W.; Huang, A.; et al. Whole-Exome Sequencing on Circulating Tumor Cells Explores Platinum-Drug Resistance Mutations in Advanced Non-small Cell Lung Cancer. *Front Genet* **2021**, *12*, 722078, doi:10.3389/fgene.2021.722078.
86. Rupp, B.; Ball, H.; Wuchu, F.; Nagrath, D.; Nagrath, S. Circulating tumor cells in precision medicine: challenges and opportunities. *Trends Pharmacol Sci* **2022**, *43*, 378-391, doi:10.1016/j.tips.2022.02.005.
87. Bidard, F.C.; Jacot, W.; Kiavue, N.; Dureau, S.; Kadi, A.; Brain, E.; Bachelot, T.; Bourgeois, H.; Goncalves, A.; Ladoire, S.; et al. Efficacy of Circulating Tumor Cell Count-Driven vs Clinician-Driven First-line Therapy Choice in Hormone Receptor-Positive, ERBB2-Negative Metastatic Breast Cancer: The STIC CTC Randomized Clinical Trial. *JAMA Oncol* **2021**, *7*, 34-41, doi:10.1001/jamaoncol.2020.5660.
88. Gerratana, L.; Pierga, J.Y.; Reuben, J.M.; Davis, A.A.; Wehbe, F.H.; Dirix, L.; Fehm, T.; Nole, F.; Gisbert-Criado, R.; Mavroudis, D.; et al. Modeling the Prognostic Impact of Circulating Tumor Cells Enumeration in Metastatic Breast Cancer for Clinical Trial Design Simulation. *Oncologist* **2022**, *27*, e561-e570, doi:10.1093/oncolo/oyac045.
89. Hanssen, A.; Riebenschahm, C.; Mohme, M.; Joosse, S.A.; Velthaus, J.L.; Berger, L.A.; Bernreuther, C.; Glatzel, M.; Loges, S.; Lamszus, K.; et al. Frequency of Circulating Tumor Cells (CTC) in Patients with Brain

- Metastases: Implications as a Risk Assessment Marker in Oligo-Metastatic Disease. *Cancers (Basel)* **2018**, *10*, doi:10.3390/cancers10120527.
90. Payne, K.; Brooks, J.; Batis, N.; Khan, N.; El-Asrag, M.; Nankivell, P.; Mehanna, H.; Taylor, G. Feasibility of mass cytometry proteomic characterisation of circulating tumour cells in head and neck squamous cell carcinoma for deep phenotyping. *Br J Cancer* **2023**, *129*, 1590-1598, doi:10.1038/s41416-023-02428-2.
 91. Sun, Y.F.; Wu, L.; Liu, S.P.; Jiang, M.M.; Hu, B.; Zhou, K.Q.; Guo, W.; Xu, Y.; Zhong, Y.; Zhou, X.R.; et al. Dissecting spatial heterogeneity and the immune-evasion mechanism of CTCs by single-cell RNA-seq in hepatocellular carcinoma. *Nat Commun* **2021**, *12*, 4091, doi:10.1038/s41467-021-24386-0.
 92. Rossi, T.; Angeli, D.; Tebaldi, M.; Fici, P.; Rossi, E.; Rocca, A.; Palleschi, M.; Maltoni, R.; Martinelli, G.; Fabbri, F.; et al. Dissecting Molecular Heterogeneity of Circulating Tumor Cells (CTCs) from Metastatic Breast Cancer Patients through Copy Number Aberration (CNA) and Single Nucleotide Variant (SNV) Single Cell Analysis. *Cancers (Basel)* **2022**, *14*, doi:10.3390/cancers14163925.
 93. Rothe, F.; Venet, D.; Peeters, D.; Rouas, G.; Rediti, M.; Smeets, D.; Dupont, F.; Campbell, P.; Lambrechts, D.; Dirix, L.; et al. Interrogating breast cancer heterogeneity using single and pooled circulating tumor cell analysis. *NPJ Breast Cancer* **2022**, *8*, 79, doi:10.1038/s41523-022-00445-7.
 94. Tretyakova, M.S.; Menyailo, M.E.; Schegoleva, A.A.; Bokova, U.A.; Larionova, I.V.; Denisov, E.V. Technologies for Viable Circulating Tumor Cell Isolation. *Int J Mol Sci* **2022**, *23*, doi:10.3390/ijms232415979.
 95. Ortiz-Otero, N.; Marshall, J.R.; Lash, B.; King, M.R. Chemotherapy-induced release of circulating-tumor cells into the bloodstream in collective migration units with cancer-associated fibroblasts in metastatic cancer patients. *BMC Cancer* **2020**, *20*, 873, doi:10.1186/s12885-020-07376-1.
 96. Cheng, Y.H.; Chen, Y.C.; Lin, E.; Brien, R.; Jung, S.; Chen, Y.T.; Lee, W.; Hao, Z.; Sahoo, S.; Min Kang, H.; et al. Hydro-Seq enables contamination-free high-throughput single-cell RNA-sequencing for circulating tumor cells. *Nat Commun* **2019**, *10*, 2163, doi:10.1038/s41467-019-10122-2.
 97. Habli, Z.; AlChamaa, W.; Saab, R.; Kadara, H.; Khraiche, M.L. Circulating Tumor Cell Detection Technologies and Clinical Utility: Challenges and Opportunities. *Cancers (Basel)* **2020**, *12*, doi:10.3390/cancers12071930.
 98. Sharma, S.; Zhuang, R.; Long, M.; Pavlovic, M.; Kang, Y.; Ilyas, A.; Asghar, W. Circulating tumor cell isolation, culture, and downstream molecular analysis. *Biotechnol Adv* **2018**, *36*, 1063-1078, doi:10.1016/j.biotechadv.2018.03.007.
 99. Banko, P.; Lee, S.Y.; Nagygyorgy, V.; Zrinyi, M.; Chae, C.H.; Cho, D.H.; Telekes, A. Technologies for circulating tumor cell separation from whole blood. *J Hematol Oncol* **2019**, *12*, 48, doi:10.1186/s13045-019-0735-4.
 100. Agrawal, L.; Engel, K.B.; Greytak, S.R.; Moore, H.M. Understanding preanalytical variables and their effects on clinical biomarkers of oncology and immunotherapy. *Semin Cancer Biol* **2018**, *52*, 26-38, doi:10.1016/j.semcancer.2017.12.008.
 101. Jiang, C.Y.; Niu, Z.; Green, M.D.; Zhao, L.; Raupp, S.; Pannecouk, B.; Brenner, D.E.; Negrath, S.; Ramnath, N. It's not 'just a tube of blood': principles of protocol development, sample collection, staffing and budget considerations for blood-based biomarkers in immunotherapy studies. *J Immunother Cancer* **2021**, *9*, doi:10.1136/jitc-2021-003212.
 102. Vasseur, A.; Kiavue, N.; Bidard, F.C.; Pierga, J.Y.; Cabel, L. Clinical utility of circulating tumor cells: an update. *Mol Oncol* **2021**, *15*, 1647-1666, doi:10.1002/1878-0261.12869.
 103. Sidaway, P. New data confirm clinical utility of ctDNA. *Nat Rev Clin Oncol* **2023**, *20*, 63, doi:10.1038/s41571-022-00716-z.
 104. Alexandrou, G.; Mantikas, K.T.; Allsopp, R.; Yapeter, C.A.; Jahin, M.; Melnick, T.; Ali, S.; Coombes, R.C.; Toumazou, C.; Shaw, J.A.; et al. The Evolution of Affordable Technologies in Liquid Biopsy Diagnostics: The Key to Clinical Implementation. *Cancers (Basel)* **2023**, *15*, doi:10.3390/cancers15225434.
 105. Radfar, P.; Aboulkheyr Es, H.; Salomon, R.; Kulasinghe, A.; Ramalingam, N.; Sarafraz-Yazdi, E.; Thiery, J.P.; Warkiani, M.E. Single-cell analysis of circulating tumour cells: enabling technologies and clinical applications. *Trends Biotechnol* **2022**, *40*, 1041-1060, doi:10.1016/j.tibtech.2022.02.004.
 106. Qiu, J.; Xu, J.; Zhang, K.; Gu, W.; Nie, L.; Wang, G.; Luo, Y. Refining Cancer Management Using Integrated Liquid Biopsy. *Theranostics* **2020**, *10*, 2374-2384, doi:10.7150/thno.40677.
 107. Vidlarova, M.; Rehulkova, A.; Stejskal, P.; Prokopova, A.; Slavik, H.; Hajduch, M.; Srovnal, J. Recent Advances in Methods for Circulating Tumor Cell Detection. *Int J Mol Sci* **2023**, *24*, doi:10.3390/ijms24043902.

108. Cayrefourcq, L.; De Roeck, A.; Garcia, C.; Stoebner, P.E.; Fichel, F.; Garima, F.; Perriard, F.; Daures, J.P.; Meunier, L.; Alix-Panabieres, C. S100-EPISPOT: A New Tool to Detect Viable Circulating Melanoma Cells. *Cells* **2019**, *8*, doi:10.3390/cells8070755.
109. Danila, D.C.; Samoila, A.; Patel, C.; Schreiber, N.; Herkal, A.; Anand, A.; Bastos, D.; Heller, G.; Fleisher, M.; Scher, H.I. Clinical Validity of Detecting Circulating Tumor Cells by AdnaTest Assay Compared With Direct Detection of Tumor mRNA in Stabilized Whole Blood, as a Biomarker Predicting Overall Survival for Metastatic Castration-Resistant Prostate Cancer Patients. *Cancer J* **2016**, *22*, 315-320, doi:10.1097/PPO.0000000000000220.
110. Gorges, T.M.; Stein, A.; Quidde, J.; Hauch, S.; Rock, K.; Riethdorf, S.; Joosse, S.A.; Pantel, K. Improved Detection of Circulating Tumor Cells in Metastatic Colorectal Cancer by the Combination of the CellSearch(R) System and the AdnaTest(R). *PLoS One* **2016**, *11*, e0155126, doi:10.1371/journal.pone.0155126.
111. Lu, R.; Chen, Q.; Liu, X.; Shen, S.; Pan, Z.; Shi, C. Detection of circulating stage III-IV gastric cancer tumor cells based on isolation by size of epithelial tumor: using the circulating tumor cell biopsy technology. *Transl Cancer Res* **2019**, *8*, 1342-1350, doi:10.21037/tcr.2019.07.32.
112. Hvichia, G.E.; Parveen, Z.; Wagner, C.; Janning, M.; Quidde, J.; Stein, A.; Muller, V.; Loges, S.; Neves, R.P.; Stoecklein, N.H.; et al. A novel microfluidic platform for size and deformability based separation and the subsequent molecular characterization of viable circulating tumor cells. *Int J Cancer* **2016**, *138*, 2894-2904, doi:10.1002/ijc.30007.
113. Russo, G.I.; Musso, N.; Romano, A.; Caruso, G.; Petralia, S.; Lanzano, L.; Broggi, G.; Camarda, M. The Role of Dielectrophoresis for Cancer Diagnosis and Prognosis. *Cancers (Basel)* **2021**, *14*, doi:10.3390/cancers14010198.
114. Low, W.S.; Wan Abas, W.A. Benchtop technologies for circulating tumor cells separation based on biophysical properties. *Biomed Res Int* **2015**, *2015*, 239362, doi:10.1155/2015/239362.
115. Peng, H.; Lu, L.; Zhou, Z.; Liu, J.; Zhang, D.; Nan, K.; Zhao, X.; Li, F.; Tian, L.; Dong, H.; et al. CNV Detection from Circulating Tumor DNA in Late Stage Non-Small Cell Lung Cancer Patients. *Genes (Basel)* **2019**, *10*, doi:10.3390/genes10110926.
116. Eslami, S.Z.; Cortes-Hernandez, L.E.; Alix-Panabieres, C. Epithelial Cell Adhesion Molecule: An Anchor to Isolate Clinically Relevant Circulating Tumor Cells. *Cells* **2020**, *9*, doi:10.3390/cells9081836.
117. Lozar, T.; Jesenko, T.; Kloboves Prevodnik, V.; Cemazar, M.; Hosta, V.; Jericevic, A.; Nolde, N.; Grasic Kuhar, C. Preclinical and Clinical Evaluation of Magnetic-Activated Cell Separation Technology for CTC Isolation in Breast Cancer. *Front Oncol* **2020**, *10*, 554554, doi:10.3389/fonc.2020.554554.
118. Hyun, K.A.; Lee, T.Y.; Lee, S.H.; Jung, H.I. Two-stage microfluidic chip for selective isolation of circulating tumor cells (CTCs). *Biosens Bioelectron* **2015**, *67*, 86-92, doi:10.1016/j.bios.2014.07.019.
119. Wang, Z.; Wu, Z.; Sun, N.; Cao, Y.; Cai, X.; Yuan, F.; Zou, H.; Xing, C.; Pei, R. Antifouling hydrogel-coated magnetic nanoparticles for selective isolation and recovery of circulating tumor cells. *J Mater Chem B* **2021**, *9*, 677-682, doi:10.1039/d0tb02380a.
120. Zhao, S.; Yang, H.; Zhang, M.; Zhang, D.; Liu, Y.; Liu, Y.; Song, Y.; Zhang, X.; Li, H.; Ma, W.; et al. Circulating tumor cells (CTCs) detected by triple-marker EpCAM, CK19, and hMAM RT-PCR and their relation to clinical outcome in metastatic breast cancer patients. *Cell Biochem Biophys* **2013**, *65*, 263-273, doi:10.1007/s12013-012-9426-2.
121. Descamps, L.; Le Roy, D.; Deman, A.L. Microfluidic-Based Technologies for CTC Isolation: A Review of 10 Years of Intense Efforts towards Liquid Biopsy. *Int J Mol Sci* **2022**, *23*, doi:10.3390/ijms23041981.
122. Ribeiro-Samy, S.; Oliveira, M.I.; Pereira-Veiga, T.; Muinelo-Romay, L.; Carvalho, S.; Gaspar, J.; Freitas, P.P.; Lopez-Lopez, R.; Costa, C.; Dieguez, L. Fast and efficient microfluidic cell filter for isolation of circulating tumor cells from unprocessed whole blood of colorectal cancer patients. *Sci Rep* **2019**, *9*, 8032, doi:10.1038/s41598-019-44401-1.
123. Po, J.W.; Roohullah, A.; Lynch, D.; DeFazio, A.; Harrison, M.; Harnett, P.R.; Kennedy, C.; de Souza, P.; Becker, T.M. Improved ovarian cancer EMT-CTC isolation by immunomagnetic targeting of epithelial EpCAM and mesenchymal N-cadherin. *J Circ Biomark* **2018**, *7*, 1849454418782617, doi:10.1177/1849454418782617.
124. Ramos-Medina, R.; Lopez-Tarruella, S.; Del Monte-Millan, M.; Massarrah, T.; Martin, M. Technical Challenges for CTC Implementation in Breast Cancer. *Cancers (Basel)* **2021**, *13*, doi:10.3390/cancers13184619.
125. Zhang, J.; Chen, K.; Fan, Z.H. Circulating Tumor Cell Isolation and Analysis. *Adv Clin Chem* **2016**, *75*, 1-31, doi:10.1016/bs.acc.2016.03.003.

126. Hofman, V.J.; Ilie, M.I.; Bonnetaud, C.; Selva, E.; Long, E.; Molina, T.; Vignaud, J.M.; Flejou, J.F.; Lantuejoul, S.; Piaton, E.; et al. Cytopathologic detection of circulating tumor cells using the isolation by size of epithelial tumor cell method: promises and pitfalls. *Am J Clin Pathol* **2011**, *135*, 146-156, doi:10.1309/AJCP9X8OZBEIQVVI.
127. Dolfus, C.; Piton, N.; Toure, E.; Sabourin, J.C. Circulating tumor cell isolation: the assets of filtration methods with polycarbonate track-etched filters. *Chin J Cancer Res* **2015**, *27*, 479-487, doi:10.3978/j.issn.1000-9604.2015.09.01.
128. Krebs, M.G.; Hou, J.M.; Sloane, R.; Lancashire, L.; Priest, L.; Nonaka, D.; Ward, T.H.; Backen, A.; Clack, G.; Hughes, A.; et al. Analysis of circulating tumor cells in patients with non-small cell lung cancer using epithelial marker-dependent and -independent approaches. *J Thorac Oncol* **2012**, *7*, 306-315, doi:10.1097/JTO.0b013e31823c5c16.
129. Vu-Dinh, H.; Do Quang, L.; Chang, C.C.; Nhu, C.N.; Thanh, H.T.; Bui, T.T.; Duc, T.C.; Jen, C.P. Immunomagnetic separation in a novel cavity-added serpentine microchannel structure for the selective isolation of lung adenocarcinoma cells. *Biomed Microdevices* **2021**, *23*, 51, doi:10.1007/s10544-021-00589-6.
130. Sun, C.; Hsieh, Y.P.; Ma, S.; Geng, S.; Cao, Z.; Li, L.; Lu, C. Immunomagnetic separation of tumor initiating cells by screening two surface markers. *Sci Rep* **2017**, *7*, 40632, doi:10.1038/srep40632.
131. Khamenehfar, A.; Li, P.C. Microfluidic Devices for Circulating Tumor Cells Isolation and Subsequent Analysis. *Curr Pharm Biotechnol* **2016**, *17*, 810-821, doi:10.2174/1389201017666160301103509.
132. Hu, P.; Zhang, W.; Xin, H.; Deng, G. Single Cell Isolation and Analysis. *Front Cell Dev Biol* **2016**, *4*, 116, doi:10.3389/fcell.2016.00116.
133. Bhagwat, N.; Dulmage, K.; Pletcher, C.H., Jr.; Wang, L.; DeMuth, W.; Sen, M.; Balli, D.; Yee, S.S.; Sa, S.; Tong, F.; et al. An integrated flow cytometry-based platform for isolation and molecular characterization of circulating tumor single cells and clusters. *Sci Rep* **2018**, *8*, 5035, doi:10.1038/s41598-018-23217-5.
134. Riethdorf, S.; O'Flaherty, L.; Hille, C.; Pantel, K. Clinical applications of the CellSearch platform in cancer patients. *Adv Drug Deliv Rev* **2018**, *125*, 102-121, doi:10.1016/j.addr.2018.01.011.
135. Polzer, B.; Medoro, G.; Pasch, S.; Fontana, F.; Zorzino, L.; Pestka, A.; Andergassen, U.; Meier-Stiegen, F.; Czyz, Z.T.; Alberter, B.; et al. Molecular profiling of single circulating tumor cells with diagnostic intention. *EMBO Mol Med* **2014**, *6*, 1371-1386, doi:10.15252/emmm.201404033.
136. van der Toom, E.E.; Groot, V.P.; Glavaris, S.A.; Gemenetzi, G.; Chalfin, H.J.; Wood, L.D.; Wolfgang, C.L.; de la Rosette, J.; de Reijke, T.M.; Pienta, K.J. Analogous detection of circulating tumor cells using the AccuCyte((R)) -CyteFinder((R)) system and ISET system in patients with locally advanced and metastatic prostate cancer. *Prostate* **2018**, *78*, 300-307, doi:10.1002/pros.23474.
137. Donato, C.; Kunz, L.; Castro-Giner, F.; Paasinen-Sohns, A.; Strittmatter, K.; Szczerba, B.M.; Scherrer, R.; Di Maggio, N.; Heusermann, W.; Biehlmair, O.; et al. Hypoxia Triggers the Intravasation of Clustered Circulating Tumor Cells. *Cell Rep* **2020**, *32*, 108105, doi:10.1016/j.celrep.2020.108105.
138. Stevens, M.; Oomens, L.; Broekmaat, J.; Weersink, J.; Abali, F.; Swennenhuis, J.; Tibbe, A. VyCAP's puncher technology for single cell identification, isolation, and analysis. *Cytometry A* **2018**, *93*, 1255-1259, doi:10.1002/cyto.a.23631.
139. Yoshino, T.; Tanaka, T.; Nakamura, S.; Negishi, R.; Hosokawa, M.; Matsunaga, T. Manipulation of a Single Circulating Tumor Cell Using Visualization of Hydrogel Encapsulation toward Single-Cell Whole-Genome Amplification. *Anal Chem* **2016**, *88*, 7230-7237, doi:10.1021/acs.analchem.6b01475.
140. Negishi, R.; Yamakawa, H.; Kobayashi, T.; Horikawa, M.; Shimoyama, T.; Koizumi, F.; Sawada, T.; Oboki, K.; Omuro, Y.; Funasaka, C.; et al. Transcriptomic profiling of single circulating tumor cells provides insight into human metastatic gastric cancer. *Commun Biol* **2022**, *5*, 20, doi:10.1038/s42003-021-02937-x.
141. Goldstein, A.; Toro, P.V.; Lee, J.; Silberstein, J.L.; Nakazawa, M.; Waters, I.; Cravero, K.; Chu, D.; Cochran, R.L.; Kim, M.; et al. Detection fidelity of AR mutations in plasma derived cell-free DNA. *Oncotarget* **2017**, *8*, 15651-15662, doi:10.18632/oncotarget.14926.
142. Antonarakis, E.S.; Lu, C.; Luber, B.; Wang, H.; Chen, Y.; Zhu, Y.; Silberstein, J.L.; Taylor, M.N.; Maughan, B.L.; Denmeade, S.R.; et al. Clinical Significance of Androgen Receptor Splice Variant-7 mRNA Detection in Circulating Tumor Cells of Men With Metastatic Castration-Resistant Prostate Cancer Treated With First- and Second-Line Abiraterone and Enzalutamide. *J Clin Oncol* **2017**, *35*, 2149-2156, doi:10.1200/JCO.2016.70.1961.

143. Vitale, S.R.; Helmijr, J.A.; Gerritsen, M.; Coban, H.; van Dessel, L.F.; Beije, N.; van der Vlugt-Daane, M.; Vigneri, P.; Sieuwerts, A.M.; Dits, N.; et al. Detection of tumor-derived extracellular vesicles in plasma from patients with solid cancer. *BMC Cancer* **2021**, *21*, 315, doi:10.1186/s12885-021-08007-z.
144. Ricciuti, B.; Jones, G.; Severgnini, M.; Alessi, J.V.; Recondo, G.; Lawrence, M.; Forshew, T.; Lydon, C.; Nishino, M.; Cheng, M.; et al. Early plasma circulating tumor DNA (ctDNA) changes predict response to first-line pembrolizumab-based therapy in non-small cell lung cancer (NSCLC). *J Immunother Cancer* **2021**, *9*, doi:10.1136/jitc-2020-001504.
145. Kwapisz, D. The first liquid biopsy test approved. Is it a new era of mutation testing for non-small cell lung cancer? *Ann Transl Med* **2017**, *5*, 46, doi:10.21037/atm.2017.01.32.
146. Mabert, K.; Cojoc, M.; Peitzsch, C.; Kurth, I.; Souchelnytskyi, S.; Dubrovskaya, A. Cancer biomarker discovery: current status and future perspectives. *Int J Radiat Biol* **2014**, *90*, 659-677, doi:10.3109/09553002.2014.892229.
147. Bettgowda, C.; Sausen, M.; Leary, R.J.; Kinde, I.; Wang, Y.; Agrawal, N.; Bartlett, B.R.; Wang, H.; Lubner, B.; Alani, R.M.; et al. Detection of circulating tumor DNA in early- and late-stage human malignancies. *Sci Transl Med* **2014**, *6*, 224ra224, doi:10.1126/scitranslmed.3007094.
148. Gezer, U.; Bronkhorst, A.J.; Holdenrieder, S. The Clinical Utility of Droplet Digital PCR for Profiling Circulating Tumor DNA in Breast Cancer Patients. *Diagnostics (Basel)* **2022**, *12*, doi:10.3390/diagnostics12123042.
149. Pfützner, C.; Schroder, I.; Scheungraber, C.; Dogan, A.; Runnebaum, I.B.; Durst, M.; Hafner, N. Digital-Direct-RT-PCR: a sensitive and specific method for quantification of CTC in patients with cervical carcinoma. *Sci Rep* **2014**, *4*, 3970, doi:10.1038/srep03970.
150. Hinz, S.; Hendricks, A.; Wittig, A.; Schafmayer, C.; Tepel, J.; Kalthoff, H.; Becker, T.; Roder, C. Detection of circulating tumor cells with CK20 RT-PCR is an independent negative prognostic marker in colon cancer patients - a prospective study. *BMC Cancer* **2017**, *17*, 53, doi:10.1186/s12885-016-3035-1.
151. Mei, T.; Lu, X.; Sun, N.; Li, X.; Chen, J.; Liang, M.; Zhou, X.; Fang, Z. Real-time quantitative PCR detection of circulating tumor cells using tag DNA mediated signal amplification strategy. *J Pharm Biomed Anal* **2018**, *158*, 204-208, doi:10.1016/j.jpba.2018.06.009.
152. Taylor, S.C.; Laperriere, G.; Germain, H. Droplet Digital PCR versus qPCR for gene expression analysis with low abundant targets: from variable nonsense to publication quality data. *Sci Rep* **2017**, *7*, 2409, doi:10.1038/s41598-017-02217-x.
153. Sedlak, R.H.; Kuypers, J.; Jerome, K.R. A multiplexed droplet digital PCR assay performs better than qPCR on inhibition prone samples. *Diagn Microbiol Infect Dis* **2014**, *80*, 285-286, doi:10.1016/j.diagmicrobio.2014.09.004.
154. Choi, C.H.; Kim, E.; Yang, S.M.; Kim, D.S.; Suh, S.M.; Lee, G.Y.; Kim, H.Y. Comparison of Real-Time PCR and Droplet Digital PCR for the Quantitative Detection of *Lactiplantibacillus plantarum* subsp. *plantarum*. *Foods* **2022**, *11*, doi:10.3390/foods11091331.
155. Feng, J.; Cui, X.; Du, B.; Zhao, H.; Feng, Y.; Cui, J.; Yan, C.; Gan, L.; Fan, Z.; Fu, T.; et al. Detection and Quantification of *Klebsiella pneumoniae* in Fecal Samples Using Digital Droplet PCR in Comparison with Real-Time PCR. *Microbiol Spectr* **2023**, *11*, e0424922, doi:10.1128/spectrum.04249-22.
156. Zhao, Y.; Xia, Q.; Yin, Y.; Wang, Z. Comparison of Droplet Digital PCR and Quantitative PCR Assays for Quantitative Detection of *Xanthomonas citri* Subsp. *citri*. *PLoS One* **2016**, *11*, e0159004, doi:10.1371/journal.pone.0159004.
157. Cros, J.; Raffenne, J.; Couvelard, A.; Pote, N. Tumor Heterogeneity in Pancreatic Adenocarcinoma. *Pathobiology* **2018**, *85*, 64-71, doi:10.1159/000477773.
158. Yu, T.; Wang, C.; Xie, M.; Zhu, C.; Shu, Y.; Tang, J.; Guan, X. Heterogeneity of CTC contributes to the organotropism of breast cancer. *Biomed Pharmacother* **2021**, *137*, 111314, doi:10.1016/j.biopha.2021.111314.
159. Keller, L.; Pantel, K. Unravelling tumour heterogeneity by single-cell profiling of circulating tumour cells. *Nat Rev Cancer* **2019**, *19*, 553-567, doi:10.1038/s41568-019-0180-2.
160. Heumos, L.; Schaar, A.C.; Lance, C.; Litnetskaya, A.; Drost, F.; Zappia, L.; Lucken, M.D.; Strobl, D.C.; Henao, J.; Curion, F.; et al. Best practices for single-cell analysis across modalities. *Nat Rev Genet* **2023**, *24*, 550-572, doi:10.1038/s41576-023-00586-w.
161. Hafemeister, C.; Satija, R. Normalization and variance stabilization of single-cell RNA-seq data using regularized negative binomial regression. *Genome Biol* **2019**, *20*, 296, doi:10.1186/s13059-019-1874-1.

162. Shiau, C.K.; Lu, L.; Kieser, R.; Fukumura, K.; Pan, T.; Lin, H.Y.; Yang, J.; Tong, E.L.; Lee, G.; Yan, Y.; et al. High throughput single cell long-read sequencing analyses of same-cell genotypes and phenotypes in human tumors. *Nat Commun* **2023**, *14*, 4124, doi:10.1038/s41467-023-39813-7.
163. Stuart, T.; Butler, A.; Hoffman, P.; Hafemeister, C.; Papalexi, E.; Mauck, W.M., 3rd; Hao, Y.; Stoeckius, M.; Smibert, P.; Satija, R. Comprehensive Integration of Single-Cell Data. *Cell* **2019**, *177*, 1888-1902 e1821, doi:10.1016/j.cell.2019.05.031.
164. Zheng, G.X.; Terry, J.M.; Belgrader, P.; Ryvkin, P.; Bent, Z.W.; Wilson, R.; Ziraldo, S.B.; Wheeler, T.D.; McDermott, G.P.; Zhu, J.; et al. Massively parallel digital transcriptional profiling of single cells. *Nat Commun* **2017**, *8*, 14049, doi:10.1038/ncomms14049.
165. Miyamoto, D.T.; Zheng, Y.; Wittner, B.S.; Lee, R.J.; Zhu, H.; Broderick, K.T.; Desai, R.; Fox, D.B.; Brannigan, B.W.; Trautwein, J.; et al. RNA-Seq of single prostate CTCs implicates noncanonical Wnt signaling in antiandrogen resistance. *Science* **2015**, *349*, 1351-1356, doi:10.1126/science.aab0917.
166. Zhang, S.; Zhang, W.; Zhang, J. 8-Gene signature related to CD8(+) T cell infiltration by integrating single-cell and bulk RNA-sequencing in head and neck squamous cell carcinoma. *Front Genet* **2022**, *13*, 938611, doi:10.3389/fgene.2022.938611.
167. Wang, Q.; Zhao, L.; Han, L.; Tuo, X.; Ma, S.; Wang, Y.; Feng, X.; Liang, D.; Sun, C.; Wang, Q.; et al. The Discordance of Gene Mutations between Circulating Tumor Cells and Primary/Metastatic Tumor. *Mol Ther Oncolytics* **2019**, *15*, 21-29, doi:10.1016/j.omto.2019.08.006.
168. Auwal, A.; Hossain, M.M.; Pronoy, T.U.H.; Rashel, K.; Nurujjaman, M.; Lam, A.K.; Islam, F. Clinical significance of genomic sequencing of circulating tumour cells (CTCs) in cancer. *The Journal of Liquid Biopsy* **2023**, 100135.
169. Poellmann, M.J.; Bu, J.; Kim, D.; Iida, M.; Hong, H.; Wang, A.Z.; Wheeler, D.L.; Kimple, R.J.; Hong, S. Circulating tumor cell abundance in head and neck squamous cell carcinoma decreases with successful chemoradiation and cetuximab treatment. *Cancer Lett* **2023**, *562*, 216187, doi:10.1016/j.canlet.2023.216187.
170. Guan, Y.; Xu, F.; Wang, Y.; Tian, J.; Wan, Z.; Wang, Z.; Chong, T. Identification of key genes and functions of circulating tumor cells in multiple cancers through bioinformatic analysis. *BMC Med Genomics* **2020**, *13*, 140, doi:10.1186/s12920-020-00795-w.
171. Farhang Ghahremani, M.; Seto, K.K.Y.; Cho, W.; Miller, M.C.; Smith, P.; Englert, D.F. Novel method for highly multiplexed gene expression profiling of circulating tumor cells (CTCs) captured from the blood of women with metastatic breast cancer. *J Transl Med* **2023**, *21*, 414, doi:10.1186/s12967-023-04242-z.
172. Ge, Z.; Helmijr, J.C.A.; Jansen, M.; Boor, P.P.C.; Noordam, L.; Peppelenbosch, M.; Kwekkeboom, J.; Kraan, J.; Sprengers, D. Detection of oncogenic mutations in paired circulating tumor DNA and circulating tumor cells in patients with hepatocellular carcinoma. *Transl Oncol* **2021**, *14*, 101073, doi:10.1016/j.tranon.2021.101073.
173. Nomura, M.; Miyake, Y.; Inoue, A.; Yokoyama, Y.; Noda, N.; Kouda, S.; Hata, T.; Ogino, T.; Miyoshi, N.; Takahashi, H.; et al. Single-Cell Analysis of Circulating Tumor Cells from Patients with Colorectal Cancer Captured with a Dielectrophoresis-Based Micropore System. *Biomedicines* **2023**, *11*, doi:10.3390/biomedicines11010203.
174. Kong, S.L.; Liu, X.; Suhaimi, N.M.; Koh, K.J.H.; Hu, M.; Lee, D.Y.S.; Cima, I.; Phyto, W.M.; Lee, E.X.W.; Tai, J.A.; et al. Molecular characterization of circulating colorectal tumor cells defines genetic signatures for individualized cancer care. *Oncotarget* **2017**, *8*, 68026-68037, doi:10.18632/oncotarget.19138.
175. Liu, H.E.; Triboulet, M.; Zia, A.; Vuppapalaty, M.; Kidess-Sigal, E.; Coller, J.; Natu, V.S.; Shokoohi, V.; Che, J.; Renier, C.; et al. Workflow optimization of whole genome amplification and targeted panel sequencing for CTC mutation detection. *NPJ Genom Med* **2017**, *2*, 34, doi:10.1038/s41525-017-0034-3.
176. Singh, R.R. Next-Generation Sequencing in High-Sensitive Detection of Mutations in Tumors: Challenges, Advances, and Applications. *J Mol Diagn* **2020**, *22*, 994-1007, doi:10.1016/j.jmoldx.2020.04.213.
177. Biezuner, T.; Raz, O.; Amir, S.; Milo, L.; Adar, R.; Fried, Y.; Ainbinder, E.; Shapiro, E. Comparison of seven single cell whole genome amplification commercial kits using targeted sequencing. *Sci Rep* **2021**, *11*, 17171, doi:10.1038/s41598-021-96045-9.
178. Saucedo-Zeni, N.; Mewes, S.; Niestroj, R.; Gasiorowski, L.; Murawa, D.; Nowaczyk, P.; Tomasi, T.; Weber, E.; Dworacki, G.; Morgenthaler, N.G.; et al. A novel method for the in vivo isolation of circulating tumor cells from peripheral blood of cancer patients using a functionalized and structured medical wire. *Int J Oncol* **2012**, *41*, 1241-1250, doi:10.3892/ijo.2012.1557.

179. El-Heliebi, A.; Hille, C.; Laxman, N.; Svedlund, J.; Haudum, C.; Ercan, E.; Kroneis, T.; Chen, S.; Smolle, M.; Rossmann, C.; et al. In Situ Detection and Quantification of AR-V7, AR-FL, PSA, and KRAS Point Mutations in Circulating Tumor Cells. *Clin Chem* **2018**, *64*, 536-546, doi:10.1373/clinchem.2017.281295.
180. Chen, S.; Tauber, G.; Langsenlehner, T.; Schmolzer, L.M.; Potscher, M.; Riethdorf, S.; Kuske, A.; Leitinger, G.; Kashofer, K.; Czyz, Z.T.; et al. In Vivo Detection of Circulating Tumor Cells in High-Risk Non-Metastatic Prostate Cancer Patients Undergoing Radiotherapy. *Cancers (Basel)* **2019**, *11*, doi:10.3390/cancers11070933.
181. Shreffler, J.; Huecker, M.R. Diagnostic testing accuracy: Sensitivity, specificity, predictive values and likelihood ratios. **2020**.
182. Hu, Y.; Tang, W.; Cheng, P.; Zhou, Q.; Tian, X.; Wei, X.; He, H. Monitoring circulating tumor cells in vivo by a confocal microscopy system. *Cytometry A* **2019**, *95*, 657-663, doi:10.1002/cyto.a.23702.
183. Carmona-Ule, N.; Gonzalez-Conde, M.; Abuin, C.; Cueva, J.F.; Palacios, P.; Lopez-Lopez, R.; Costa, C.; Davila-Ibanez, A.B. Short-Term Ex Vivo Culture of CTCs from Advance Breast Cancer Patients: Clinical Implications. *Cancers (Basel)* **2021**, *13*, doi:10.3390/cancers13112668.
184. Lee, H.L.; Chiou, J.F.; Wang, P.Y.; Lu, L.S.; Shen, C.N.; Hsu, H.L.; Burnouf, T.; Ting, L.L.; Chou, P.C.; Chung, C.L.; et al. Ex Vivo Expansion and Drug Sensitivity Profiling of Circulating Tumor Cells from Patients with Small Cell Lung Cancer. *Cancers (Basel)* **2020**, *12*, doi:10.3390/cancers12113394.
185. Kapeleris, J.; Kulasinghe, A.; Warkiani, M.E.; O'leary, C.; Vela, I.; Leo, P.; Sternes, P.; O'Byrne, K.; Punyadeera, C. Ex vivo culture of circulating tumour cells derived from non-small cell lung cancer. *Translational lung cancer research* **2020**, *9*, 1795.
186. Shimada, Y.; Sudo, T.; Akamatsu, S.; Sunada, T.; Myomoto, A.; Okano, K.; Shimizu, K. Cell Lines of Circulating Tumor Cells: What Is Known and What Needs to Be Resolved. *J Pers Med* **2022**, *12*, doi:10.3390/jpm12050666.
187. Kulasinghe, A.; Perry, C.; Warkiani, M.E.; Blick, T.; Davies, A.; O'Byrne, K.; Thompson, E.W.; Nelson, C.C.; Vela, I.; Punyadeera, C. Short term ex-vivo expansion of circulating head and neck tumour cells. *Oncotarget* **2016**, *7*, 60101-60109, doi:10.18632/oncotarget.11159.
188. Yu, M.; Bardia, A.; Aceto, N.; Bersani, F.; Madden, M.W.; Donaldson, M.C.; Desai, R.; Zhu, H.; Comaills, V.; Zheng, Z.; et al. Cancer therapy. Ex vivo culture of circulating breast tumor cells for individualized testing of drug susceptibility. *Science* **2014**, *345*, 216-220, doi:10.1126/science.1253533.
189. Hamilton, G.; Hochmair, M.; Rath, B.; Klameth, L.; Zeillinger, R. Small cell lung cancer: Circulating tumor cells of extended stage patients express a mesenchymal-epithelial transition phenotype. *Cell Adh Migr* **2016**, *10*, 360-367, doi:10.1080/19336918.2016.1155019.
190. Wang, R.; Chu, G.C.Y.; Mrdenovic, S.; Annamalai, A.A.; Hendifar, A.E.; Nissen, N.N.; Tomlinson, J.S.; Lewis, M.; Palanisamy, N.; Tseng, H.R.; et al. Cultured circulating tumor cells and their derived xenografts for personalized oncology. *Asian J Urol* **2016**, *3*, 240-253, doi:10.1016/j.ajur.2016.08.005.
191. Khoo, B.L.; Lee, S.C.; Kumar, P.; Tan, T.Z.; Warkiani, M.E.; Ow, S.G.; Nandi, S.; Lim, C.T.; Thiery, J.P. Short-term expansion of breast circulating cancer cells predicts response to anti-cancer therapy. *Oncotarget* **2015**, *6*, 15578-15593, doi:10.18632/oncotarget.3903.
192. Grigoryeva, E.S.; Tashireva, L.A.; Alifanov, V.V.; Savelieva, O.E.; Vtorushin, S.V.; Zavvalova, M.V.; Cherdyntseva, N.V.; Perelmuter, V.M. The Novel Association of Early Apoptotic Circulating Tumor Cells with Treatment Outcomes in Breast Cancer Patients. *Int J Mol Sci* **2022**, *23*, doi:10.3390/ijms23169475.

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.