

Review

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Review

Hemodynamic Melody of Postnatal Cardiac and Pulmonary Development in Children with Congenital Heart Diseases

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Simple Summary: Hemodynamics is the eternal theme of the circulatory system. Abnormal hemodynamics and cardiac and pulmonary development intertwine to form the most important features of children with congenital heart diseases (CHDs), thus determining these children's long-term quality of life. However, due to the lack of neonatal rodent models of CHDs, it is unclear how CHD-associated hemodynamics shape postnatal cardiac and pulmonary development. Here, we review the varieties of hemodynamic abnormalities that exist in children with CHDs, the recently developed neonatal rodent models of CHDs, and the inspirations these models have brought us in the areas of cardiomyocyte proliferation and maturation, and in alveolar development. Furthermore, current limitations, future directions, and clinical decision-making based on these inspirations are highlighted. Understanding how CHD-associated hemodynamic scenarios shape postnatal heart and lung development may provide a novel path to improving the long-term quality of life of children with CHDs, transplantation of stem cell-derived cardiomyocytes, and cardiac regeneration.

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Keywords: volume overload; pressure overload; cardiomyocyte; maturation; proliferation; pulmonary alveolar dysplasia; pulmonary blood flow

1. Introduction

Since Harvey's discovery of the circulatory system in 1628, it has been recognized that the primary function of the circulatory system is to provide organs, including the heart and lungs, with well-oxygenated blood and nutrients, as well as to transport all types of harmful metabolites [1,2]. The hemodynamics resulting from blood flowing in the circulatory system inevitably becomes the eternal theme of the circulatory system [1,2]. Insufficient embryonic volume load is one of the main causes of left ventricular dysplasia, while adult volume overload (VO) or pressure overload (PO) leads to cardiac failure [3–7]. However, pediatric hearts and lungs are in a stage of active

development. For example, 90% of alveoli form between the ages of 0 and 7 years, and there is an extensive cardiomyocyte maturation transition [8,9]. Hemodynamic modifications of embryonic or adult hearts and lungs are well established [3–7], but leave a vacuum to be filled of our understanding of the hemodynamic shaping of pediatric hearts and lungs.

Congenital heart diseases (CHDs) represent the most prevalent birth defect in the world [10,11]. With the progress in pediatric cardiac surgery, cardiology, and cardiac intensive care, most simple CHDs can be corrected, and the overall mortality shows a significant downward trend. However, the mortality of complex CHDs, such as hypoplastic left heart syndrome, Ebstein anomaly (EA), and tetralogy of Fallot (TOF), is still very high. Most of these cannot be physiologically corrected, leaving residual hemodynamic abnormalities, such as cardiac volume overload (VO), pressure overload (PO), pulmonary congestion, and reduced pulmonary blood flow (RPF), and therefore hemodynamic abnormalities dominate postnatal cardiac and pulmonary development in children with CHDs [12–17]. As a result, the long-term life quality of children with complex CHDs is poor, with significantly reduced exercise capacity and impaired cardiac performance, and some children ultimately require heart or lung transplantation [18,19].

In addition, cardiovascular therapies that are very effective in adults have limited effectiveness or even cause harm when treating infants and young children [20,21]. A possible reason underlying this phenomenon is that the extremely small size of a neonatal mouse heart challenges the creation of neonatal surgical mouse models of cardiovascular diseases. Consequently, clinicians or scientists generally use adult mouse models to explore mechanisms and pathophysiology and to obtain targets for pediatric cardiovascular diseases.

Here we reviewed the abnormal hemodynamics in children with CHDs and the recently developed neonatal surgeries for producing CHD-hemodynamic-associated animal models, which include neonatal aortic and inferior vena cava fistula (nACF) surgery, neonatal pulmonary artery banding (nPAB) surgery, neonatal pulmonary vein banding (nPVB) surgery, and neonatal transverse aortic constriction (nTAC) surgery [22–33]. nACF produces atrial and ventricular VO and pulmonary congestion; nPAB produces right ventricular (RV) PO and RPF; nPVB produces pulmonary vein stenosis (PVS), one of the most challenging issues of pediatric CHDs [34]; and nTAC produces left ventricular (LV) PO. These neonatal surgeries greatly inspire our understanding of the hemodynamic melody of postnatal cardiac and pulmonary development, such as cardiomyocyte proliferation and maturation, and pulmonary dysplasia.

2. Abnormal Hemodynamics in Children with CHD

2.1. Abnormal Hemodynamics in Ventricles

CHD usually leads to ventricular VO and PO, specifically RV VO, LV VO, RV PO, and LV PO. Table 1 summarizes each type of abnormal hemodynamic and its corresponding CHDs.

Table 1. Abnormal hemodynamic classification and their corresponding congenital heart diseases(CHDs).

Abnormal hemodynamic classification	RV VO	LV VO	RV PO	LV PO
CHD	ASD、PDA、PAPVD 、 VSD 、 TR 、	ASD+TS/RV 、 OT 、 MR 、	PVS 、 VSD 、 PDA 、 PAH 、 MS、 AS、 TOF	BAV 、 IAA 、 CAA 、 MS 、 AS、 HCM

	EA 、 PVI 、 TOF 、 AVSD 、 PAH	MVI 、 AR 、 BAV	、 IAA 、 CAA 、 PAS、 PVS	
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There are three types of CHDs that can result in RV VO. The first type is CHD with a left-to-right shunt, which includes atrial septal defect (ASD), patent ductus arteriosus (PDA), partial anomalous pulmonary vein drainage (PAPVD), atrioventricular septal defect (AVSD), and ventricular septal defect (VSD) [35–39]. The second type is right heart valve regurgitation-associated CHDs, which include tricuspid regurgitation (TR), EA, pulmonary regurgitation, pulmonary valve insufficiency (PVI), and TOF after patch enlargement of the RV outflow tract correction [40–44]. The third includes advanced pulmonary hypertension (PH), increased returning blood (e.g., in thyrotoxicosis), and functional univentricular conditions (e.g., hypoplastic left heart syndrome and Fontan surgery) [45–47].

Right-to-left shunting and left heart valvular regurgitation can lead to LV VO. Right-to-left shunting-associated CHDs include ASD with tricuspid stenosis (TS) or RV outflow obstruction (RVOT)[48]. These defects result in a high pressure in the right atrium compared to the left atrium, leading to a right-to-left shunt. Left heart valvular regurgitation includes congenital mitral regurgitation (CMR), acquired mitral regurgitation (rheumatic or calcific mitral valve), aortic valve calcification, and bicuspid aortic valve (BAV) [49–51]. In addition, increased venous return can lead to LV VO.

PH, left cardiac defects that cause PH at later stages, and RVOT obstructive diseases may cause RV PO. CHDs that cause PH include PVS, large VSD, and PDA [34,36,39,52]. Large VSD and PDA lead to increased pulmonary blood flow, resulting in pulmonary congestion and small vessel remodeling, which in turn causes PH. CHD with a left cardiac defect that causes PH includes mitral or aortic stenosis (MS or AS), and coarctation/interruption of the aortic arch (CAA/IAA) [50,51,53]. CHDs with RVOT obstruction includes TOF, isolated pulmonary artery obstruction or embolism, pulmonary artery stenosis (PAS), and pulmonary valve stenosis (PS) [54–57].

Hypertension and left ventricular outflow tract (LVOT) obstruction leads to LV PO. Hypertension is uncommon in children. CHD with LVOT obstruction includes BAV, CAA/IAA, mitral or aortic stenosis, and hypertrophic cardiomyopathy [50,51,53,58].

It is evident from Table 1 that various hemodynamic abnormalities, including both ventricular VO and PO, usually coexist in the same type of CHD. In addition, these hemodynamic abnormalities are closely associated with the disease progression, and sometimes one type of abnormal hemodynamic abnormality dominates a particular stage of disease progression. The extent of VO and PO often determines the performance of the RV and LV, as well as surgical approaches and the prognosis of children with CHDs. Therefore, gaining a comprehensive understanding of how VO and PO shape postnatal cardiac development may help establish a theoretical foundation for enhancing the quality of life in children with CHD.

2.2. Abnormal Hemodynamics in Lungs

A left-to-right shunt or increased venous return will induce increased pulmonary blood flow (IPF), which leads to pulmonary congestion, ultimately resulting in pulmonary hypertension and congestive heart failure. RVOT obstruction, such as TOF and PS, results in RPF, which leads to pulmonary dysplasia and reduced exercise endurance, and renders patients less susceptible to COVID-19 infection [25,29,59,60]. A deeper understanding of the mechanisms by which IPF and RPF regulate postnatal lung development may help improve lung function and exercise capacity in children with CHD.

PVS is another deadly CHD [28,34]. Stenosis of the pulmonary vein leads to remodeling of small pulmonary vessels, which in turn leads to PH, and ultimately right heart failure occurs. Despite advancements in surgical techniques, interventional procedures, and postoperative monitoring, the prognosis for children with PVS remains unfavorable, with 60% of children with PVS dying within 2

years after diagnosis [34]. Moreover, due to stenosis, there may be oozing blood upstream of the pulmonary vessels, giving rise to hemoptysis and pulmonary infections, which further deteriorate the prognosis of children with PVS. Understanding how and why PVS occurs is crucial to treat children with PVS.

3. Immature Hearts and Lungs in Children

3.1. Immature Hearts in Children

Unlike adult mature hearts and lungs, children's hearts and lungs are immature and at an active stage of development [8,12].

To meet the physiological demands of pumping blood at adulthood, there are three main characteristic that immature cardiomyocytes need to develop (Figure 1) [8,61–67]: (1) sarcomere maturation, which includes sarcomere component maturation (an Myh7 to Myh6 and TnI1 to TnI3 switch), and arrangement maturation (disordered and irregular arrangement switched to a rod-like and orderly arrangement); (2) metabolism maturation, which is a shift from anaerobic glycolysis to oxidative phosphorylation due to mitochondrial maturation, with an increase in the number of mitochondrial and their ridges, as well as a closer proximity of mitochondria to sarcomeres; and (3) electrophysiological maturation, in which transverse tubules (T-tubules) form, with a gradually increased density and integrity, and are accompanied by a significant improvement in calcium handling and excitation-contraction coupling. Failure of cardiomyocyte maturation results in arrhythmias and heart failure.

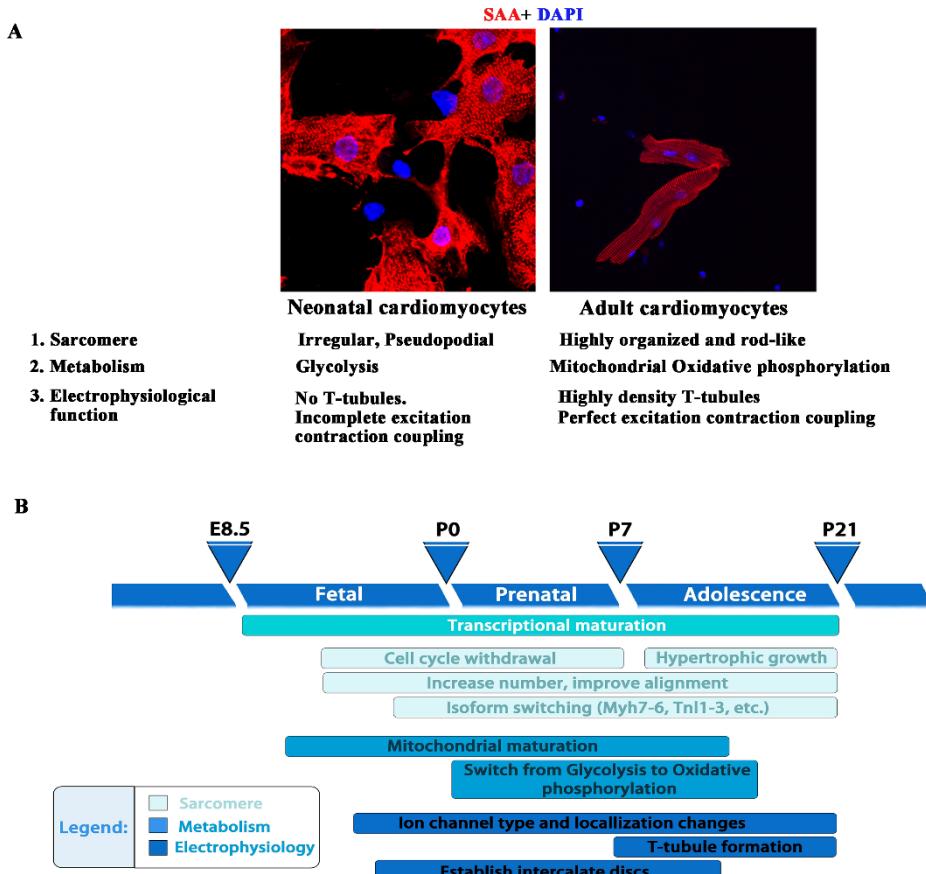


Figure 1. Illustration of three main characteristics that immature cardiomyocytes require for development. (A) Neonatal and adult cardiomyocytes have differences in sarcomere, metabolism, and electrophysiological functions. (B) Three main characteristics that immature cardiomyocytes need to develop, and the presumed key time points in rodents.

3.2. Immature Lungs in Children

There are five developmental stages of human lungs (Figure 2) [68–71]: (1) Embryonic stage (pulmonary bud stage): Occurring between the third and sixth weeks of gestation, this stage marks the appearance of precursor cells and epidermal cells known as pulmonary buds. (2) Pseudoglandular stage (bronchial stage): Taking place between the sixth and 16th weeks of gestation, this stage is characterized by bronchial formation. The pulmonary buds proliferate and migrate to the visceral mesenchyme, forming airways and peripheral acinar buds, eventually developing into alveoli. (3) Tubular phase (alveolar tubular phase): This stage occurs from 16–26 weeks of gestation and involves the differentiation of intra-airway epithelial cells, including basal cells, cupped cells, ciliated cells, and other secretory cells. (4) Vesicular phase (terminal vesicular phase): Spanning from 26–36 weeks of gestation, the differentiation of peripheral glandular alveoli characterizes this phase, including cubic type 2 alveolar (AT2) cells and squamous type 1 alveolar (AT1) cells. At this stage, alveoli gradually expand and are covered by AT1 cells, and AT2 cells differentiate to produce alveolar surfactant. (5) Alveolar phase: This phase, which extends from 36 weeks of gestation to adolescence, is the main stage of alveolar formation, with over 90% of alveoli forming at ages 0–7 years. Bronchopulmonary dysplasia, a major complication of prematurity, occurs when the immature lungs fail to develop due to various damaging factors such as hyperoxia, toxicity, and inflammation. Therefore, the lungs of children are immature and in a critical stage of alveolar development, while adult lungs have completed their development. Consequently, many drugs effective in treating adult lung diseases have limited efficacy in addressing lung diseases in infants and children [72,73].

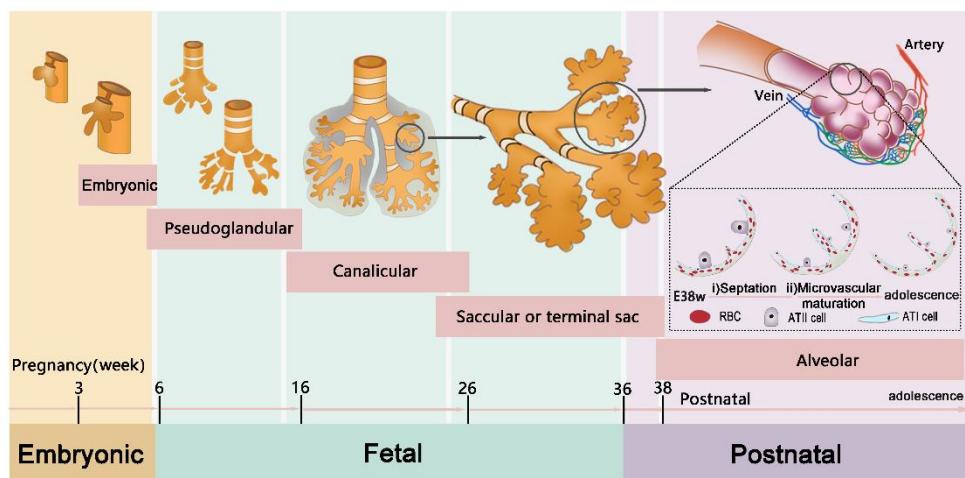


Figure 2. Five developmental stages of human lungs. Note that 90% of alveoli are formed at ages 0 – 7 years.

4. Creation of Neonatal Surgical Rodent Models of CHDs

4.1. Challenges in Constructing Neonatal Surgical Rodent Models of CHDs

The lack of neonatal rodent models for cardiac VO, PO, IPF, and RPF limits our understanding of how hemodynamics regulates postnatal heart and lung development [23,28,30,31]. Consequently, managing CHD primarily relies on surgical intervention, and thus the long-term quality of life for children with complex CHD remains poor [74,75]. The challenges of creating neonatal rodent CHD models are: (1) the size of the neonatal rodent heart is extremely small, with limited operation space; (2) the neonatal anesthesia is ice cooling, limiting the surgical time to no more than 20 min; and (3) cannibalization may account for additional post-surgical mortality in the neonatal surgery. The above three challenges require superb cardiac microsurgical skills.

As the largest pediatric heart center in the world, the heart center of Shanghai Children's Medical Center has about 4,000 open-heart surgeries per year; as a result, surgeons at Shanghai Children's Medical Center are very skillful in cardiac surgery. Thus, in recent years they have successfully

developed a series of neonatal CHD models (Figure 3), including models of nACF, nPAB, nPVB, and nTAC [23,28,30,31,76].

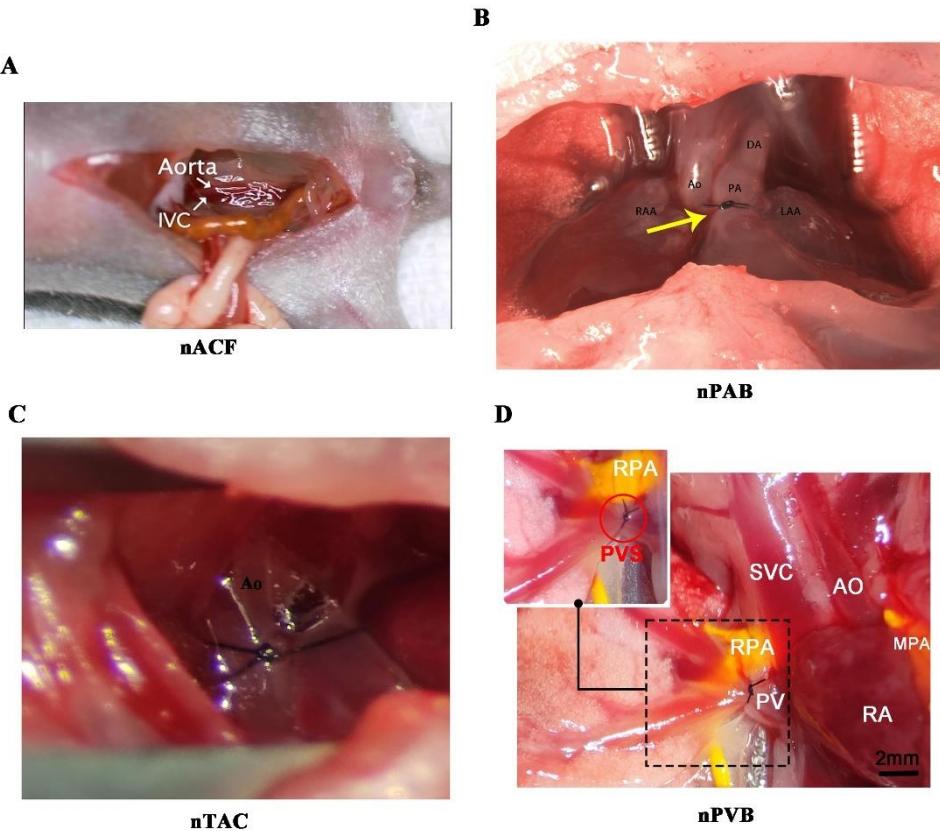


Figure 3. Neonatal surgical rodent models of CHDs. (A) nACF produces RA VO, RV VO, LA VO, LV VO, and IPF. (B) nPAB produces RV PO and RPF. (C) nTAC produces LV PO. (D) nPVB produces PVS and IPF. IVC: inferior vena cava; PA: pulmonary artery; DA: ductus arteriosus; Ao: aorta; RPA: right pulmonary artery; PV: pulmonary vein; SVC: superior vena cava; RA: right atrium; MPA: main pulmonary artery. Figure 3A was adopted from the JAHA journal (Sun S, et al. J Am Heart Assoc. 2021 Aug 17;10(16):e020854) under a Creative Commons Attribution-NonCommercial-NoDerivs License; Figure 3B was adopted from the JTCVS journal (Wang S, et al. J Thorac Cardiovasc Surg. 2017 Nov;154(5):1734-1739) with the permission of the publisher.

4.2. Key Points of Constructing Neonatal Surgical Rodent Models of CHDs

4.2.1. nACF Surgery

The nACF surgery [33] (Figure 3A) includes the following four steps: (1) anesthesia and fixation; (2) abdominal aorta (AA) and inferior vena cava (IVC) exposure; (3) fistula creation, and (4) abdominal closure. There are two key points of nACF surgery: (1) During fistula creation, a mixing of arteriovenous blood in IVC should be observed under a microscope to make sure a successful fistula was created. (2) When exposing the AA and IVC, the small intestine and large intestine should be moved with a cotton swab slowly to prevent bleeding.

4.2.2. nPAB Surgery

The nPAB surgery [30] (Figure 3B) includes the following four steps: (1) anesthesia and fixation; (2) pulmonary artery (PA) exposure; (3) PA banding; and (4) closure of the thoracic cavity. There are two key points of nPAB surgery: (1) The PA should be exposed with a small incision as much as possible to reduce the total time for the thoracotomy surgery, which is critical for the neonatal pups' recovery from anesthesia. (2) The PA should be separated from its adjacent small vessels with a blunt needle to avoid bleeding.

4.2.3. nTAC Surgery

The nTAC surgery [32] (Figure 3C) includes the following four steps: (1) anesthesia and fixation; (2) ascending aorta exposure; (3) aorta banding; and (4) closure of the thoracic cavity. There are two key points of nTAC surgery: (1) Blunt needles should be used to prevent intraoperative bleeding, which is a primary cause of postoperative mortality. (2) During the surgery, the pericardium must be carefully separated from the aortic surface to expose the aortic site for banding. The pericardium should be kept intact as much as possible to avoid postoperative adhesion.

4.2.4. nPVB Surgery

The nPVB surgery [28] (Figure 3D) includes the following four steps: (1) anesthesia and fixation; (2) pulmonary vein (PV) exposure; (3) PV banding; and (4) closure of the thoracic cavity. There are two key points of nPVB surgery: (1) The right upper and right middle PVs, which are thicker than the other PVs, are selected for banding to replicate PVS. (2) The chest is closed layer by layer to avoid postoperative pneumothorax.

In summary, nACF surgery, which does not require opening the thoracic cavity, is easier to perform than the other three neonatal surgeries. All neonatal surgeries required ice-cooling anesthesia to reduce the operation time as much as possible, which increases the rate of neonatal pup recovery from anesthesia. A blunt needle is required to avoid intraoperative bleeding. For thoracotomy surgery, preventing postoperative adhesions and pneumothorax is a key for the neonatal pups' long term survival. The surgical videos for each neonatal cardiac surgery can be found at public access websites, summarized in Table 2.

Table 2. Neonatal cardiac surgery teaching videos.

Surgery	Website	ref
nACF	https://www.ahajournals.org/doi/suppl/10.1161/JAHA.121.020854	23
nTAC	https://www.nature.com/articles/s41596-020-00434-9	32
nPAB	https://www.jtcvs.org/article/S0022-5223(17)31192-3/fulltext#supplementaryMaterial	30
nPVB	https://cellandbioscience.biomedcentral.com/articles/10.1186/s13578-023-01058-8	28

5. Hemodyn

5.1. Cardiomyocyte Proliferation

Promoting cardiomyocyte proliferation is a fundamental method for treating heart failure, which not only improves heart failure caused by myocardial infarction [77,78], but also improves pediatric heart failure caused by CHDs [79,90]. A previous study demonstrated that children with TOF have impaired RV cardiomyocyte proliferation due to reduced expression of epithelial cell transforming 2 (ECT2) [79]. The study also found that beta blockers can promote cardiomyocyte proliferation by enhancing the expression of ECT2 [79]. Thus, beta blockers have been recommended to treat TOF [79]. The New England Journal of Medicine highlighted the study, commenting that the treatment of CHDs may not rely on surgical correction; this study opened a new approach for treating CHDs [80].

However, this study lacked a nPAB model to match the TOF most important clinical feature—RVOT obstruction, which produces RV PO—and made its conclusion based on an increase in the percentage of polyploidic cardiomyocytes in children with TOF [79]. An increased percentage of polyploidic cardiomyocytes could be a consequence of cardiomyocyte proliferation (Figure 4). Increasingly more studies showed that PO promoted neonatal cardiomyocyte proliferation, in both RV and LV [31,81–85]. We also found that PO promoted RV cardiomyocyte proliferation in children

with TOF [31]. Therefore, it is inappropriate and unwise to ignore the contribution of PO to cardiomyocyte proliferation when studying pediatric TOF. Moreover, CHD is a polygenic disease, and mutations of ECT2 have rarely been reported in TOF patients [79]. Thus, it is more reasonable to open a new field for CHD treatment focused on abnormal hemodynamics than a single gene modification.

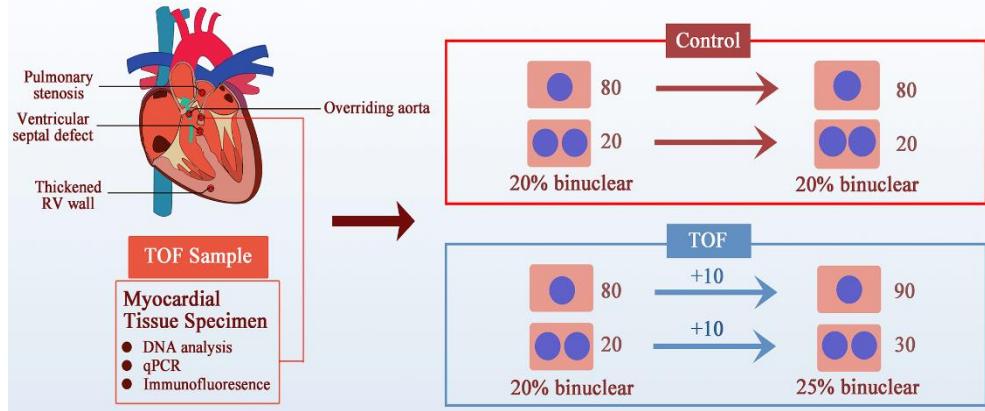


Figure 4. Possible mechanism of an increased percentage of polyploid cardiomyocytes in TOF children. (A) Four clinical features of TOF: RVOT obstruction, ventricular septal defect, thickened RV, and aortic riding, the most important of which is RVOT obstruction, which leads to RV PO and RV failure. (B) Illustration of increased binuclear cardiomyocytes generated in TOF patients. Greater numbers of binuclear cardiomyocytes do not indicate failure of cytokinesis. For example, in the beginning of a study, if both the control group and TOF group had 80 mononucleated and 20 binucleated cardiomyocytes (CMs), the proportion of binucleated CMs was 20%. Under PO conditions, both mononucleated and binucleated CMs in the TOF group increased by 10, and the proportion of binucleated CMs was 25%. Therefore, an increase in the proportion of binucleated CMs does not necessarily mean impaired cytokinesis and proliferation.

Nevertheless, the effect of PO on promoting proliferation of cardiomyocytes decreases with age, and PO cannot re-induce proliferation of differentiated and mature cardiomyocytes [31]. However, interestingly, VO re-induces proliferation of prepubertal cardiomyocytes in both the LV and RV [22,23], but does not promote proliferation of neonatal cardiomyocytes [86]. In contrast, with heart failure and at an adult stage, pressure unloading promotes cardiomyocyte proliferation [87]. These results show the complex roles of forces generated by abnormal hemodynamics on cardiomyocyte proliferation.

Force is one of the foundations of matter [88,89], and the maintenance of many life phenomena depends on a balance of forces [88,89]. How force regulates cardiac regeneration remains elusive, and perhaps changing a single mechanoreceptor can promote cardiac regeneration. In fact, a recent study demonstrated that Plxnd1 is a necessary and sufficient condition for endothelial cells to sense force for regulating cardiovascular pathophysiology [90]. We also found that Plxnd1 significantly increased in prepubertal cardiomyocytes under VO conditions (Figure 5). The introduction of neonatal ventricular PO and VO models provides a platform for us to explore the regulation of cardiac regeneration by forces, and the role of Plxnd1 may be one of the future directions.

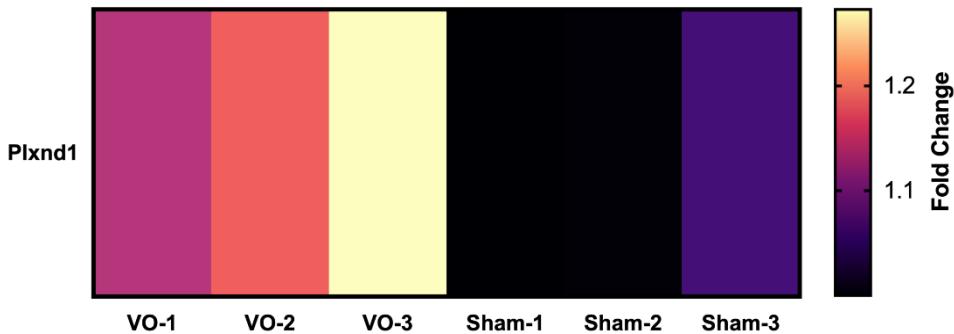


Figure 5. *Plxnd1* significantly increased in prepubertal cardiomyocytes under VO. Raw data have been deposited in NCBI's Gene Expression Omnibus database (<https://www.ncbi.nlm.nih.gov/geo>) with accession number GSE157396.

5.2. Cardiomyocyte Maturation

Maturation is the most important event of postnatal cardiomyocyte development in mammals [91,92]. Although the underlying mechanisms of cardiomyocyte maturation are largely unknown, the switch from pediatric to adult hemodynamics is undoubtedly an important contribution [91,92]. Due to the increase in body size, the preload and afterload faced by the adult heart is significantly greater than that of children [91,92]. In other words, it may be the force generated by the increasing preload and afterload that guide the maturation of cardiomyocytes.

The immaturity of cardiomyocytes derived from pluripotent stem cells (iPS-CMs) and their failure to mature *in vivo* limits their clinical use [93,94]. Therefore, *in vitro* experiments with various stimuli, including electrical rhythm, force, and 3D culture, were used to promote the maturation of iPS-CMs [93,94]. Although their maturity has increased, there is still a large gap between stimuli-enhanced iPS-CMs and mature adult cardiomyocytes [93,94]. The differences between *in vivo* and *in vitro* conditions, as well as an insufficient understanding of force, may account for the limited effect of force on cardiomyocyte maturation in studies.

Recent studies indicated that cardiomyocyte proliferation and maturation are two opposite processes [95,96]. Because VO and PO promote cardiomyocyte proliferation at neonatal or prepubertal stages [23,31], it is not surprising to find that cardiomyocyte maturation was impaired because of VO [22,23]. We also found that VO impeded cardiomyocyte maturation (Figure 6). The purpose of iPS-CM transplantation is to treat patients with heart failure, the hemodynamic characteristic of which is VO. Thus, the finding that VO impedes cardiomyocyte maturation may further deter the transplantation of iPS-CMs into patients with heart failure.

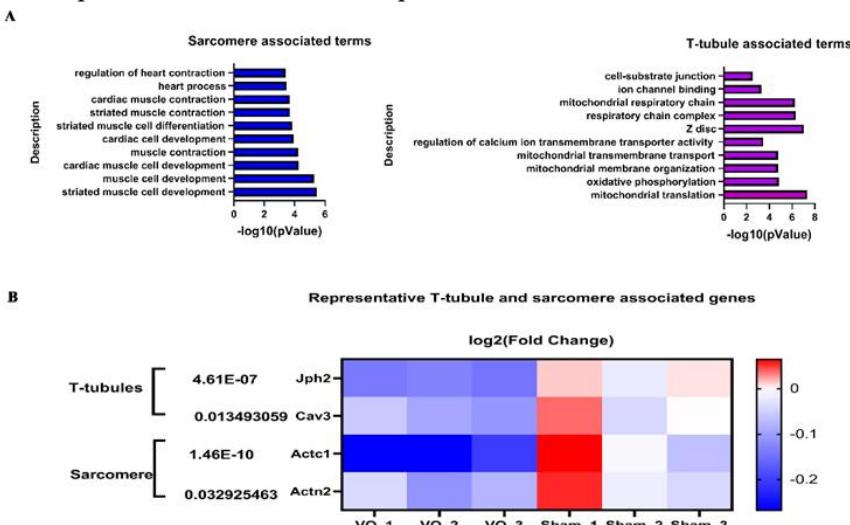


Figure 6. VO impeded cardiomyocyte maturation. (A) Enrichment of sarcomere- and T-tubule-associated terms in the Gene Ontology (GO) analysis of downregulated genes under VO. (B) Heat

map of representative sarcomere and T-tubule genes. Raw data have been deposited in NCBI's Gene Expression Omnibus database (<https://www.ncbi.nlm.nih.gov/geo>) with accession number GSE186968.

Another issue is whether VO or PO affects cardiomyocyte maturation temporarily or permanently. In other words, when VO or PO is released, can the cardiomyocytes still mature? This is the situation that exists in children with CHD. When the structural defects of the heart of children with CHDs are corrected, will the maturity of their adult cardiomyocytes be affected? Clinical investigations have found that even with perfect anatomic correction in childhood, patients with TOF are still at a high risk of arrhythmia [97], one feature of cardiomyocyte immaturity, suggesting that cardiomyocyte maturation may be permanently impaired.

In summary, some force is helpful for cardiomyocyte maturation *in vitro*, and force generated by age-increased preload and afterload may guide cardiomyocyte maturation *in vivo*. However, excessive force due to VO and PO impairs postnatal cardiomyocyte maturation. A key question is how force regulates cardiomyocyte maturation. Does maturation share the same mechanoreceptor(s) used in proliferation? After mechanoreceptor activation, different pathways involved in maturation and proliferation may be subsequently activated. In fact, we found that VO initiated an immune response at neonatal and prepubertal stages in both the RV and LV, including macrophage activation [22,23,86]. Consistently, activation of the mechanoreceptor Plxnd1 in endothelial cells led to expression of macrophage chemokines, which recruited macrophages from the peripheral blood to the heart [90]. These interesting studies are a good foundation for us to understand how force, a basic component of the material world, affects cardiovascular pathophysiology.

5.3. Lung Development

The interplay of the heart and lung profoundly, functionally, and anatomically determine a person's quality of life [98–100]. About half a century ago, an autopsy study revealed a decrease in lung volume and pulmonary dysfunction in patients with RPF-associated CHDs [101]. However, the underlying mechanisms remained elusive until recent nPAB surgery was developed, which showed that RPF caused alveolar dysplasia, angiogenesis impairment, and inflammation [28,29]. RPF also impaired cell–cell communication and axon guidance, two critical events of late alveolar formation [29,102]. Axon guidance is required for the coronavirus infection, which may explain why children with RPF-associated CHDs are relatively insensitive to COVID-19 infection [29,103]. RPF reduces the intravascular pressure and gas exchange rate, which means a reduced force in the vessels of the lung. It is possible that force is the basic cause of RPF-induced pulmonary dysplasia. Interestingly, RPF also induces inflammation [25], similar to that of VO-induced cardiomyocyte proliferation, further suggesting that there may be a force-mediated regulation.

Current bronchopulmonary dysplasia (BPD) animal models for premature infants include models of hyperoxia, pulmonary ventilation, and lipopolysaccharide [104,105], all of which aim to induce inflammation, yet yield poor targets for improving lung development of premature infants [106,107]. This may be because inflammation is not the initiating factor. Complications of premature infants often include PH and PDA [107,108], both of which induce RPF. Thus, RPF may account for premature BPD, and the nPAB model provide a new window into the study of BPD.

In contrast to RPF, IPF, which increases intravascular pressure, leads to pulmonary congestion and thickening of pulmonary small blood vessels, a characteristic of PH, which in turn increases intravascular pressure, and ultimately a vicious circle forms. A nPVB model showed a similar presentation as children with PVS, which included PV thickening, pulmonary small vessels thickening, pulmonary congestion, PH, and RV failure [28]. Consistently, the nACF model causing IPF also showed thickening of pulmonary small blood vessels, but to a lesser extent [33]. The mechanoreceptors in lungs are less studied in lungs than in hearts, including their regulation of force in lungs.

6. Summary and Prospects

Force, a basic component of our world and generated by abnormal hemodynamics in CHDs, produces profound effects on postnatal heart and lung development, which we were previously unaware of due to the lack of neonatal rodent animal models. Nevertheless, we now know that both VO and PO promote prepubertal cardiomyocyte proliferation and impede cardiomyocyte maturation, and we also know that RPF leads to pulmonary dysplasia and IPF leads to thickening of pulmonary blood vessels.

6.1. Clinical Decision Making

VO- and PO-impaired cardiomyocyte maturation are associated with arrhythmias and weakened cardiac systolic function [93,94], suggesting that childhood correction of VO or PO or the promotion of cardiomyocyte maturation may improve adult cardiac performance. However, VO and PO promote prepubertal cardiomyocyte proliferation, which is fundamental to heart failure treatment [96,97], suggesting that VO and PO should be enhanced. Clearly, these two suggestions are contradictory. An in-depth understanding of how force regulates cardiomyocyte proliferation and maturation is a prerequisite for us to precisely regulate proliferation and maturation via VO and PO for improving the quality of life of children with CHD.

RPF and IPF are both detrimental to pulmonary performance, suggesting that it should be corrected as early as possible. If RPF or IPF cannot be corrected, mechanoreceptor blockers, axon guidance molecules, or vessel thickening inhibitors are suggested to be used as early as possible to improve pulmonary performance of children with CHD.

6.2. Limitations and Future Directions

Currently, the neonatal surgical rodent models only help us obtain a very primitive observation, leaving an abundance of unanswered questions: (1) Do mechanoreceptors for VO or PO promote cardiomyocyte proliferation or impede cardiomyocyte maturation? Apart from Plxnd1, other mechanoreceptors have also been revealed. Sdc4 is upregulated, while Itga11 is downregulated in the neonatal PO RV (Figure 7). The expression pattern of neonatal Sdc4 and Itga11 is different from that of adults, in whom both are upregulated [109,110]. How PO or VO regulates cardiomyocyte proliferation and maturation via Plxnd1, Sdc4, and Itga11 might be a future direction of research. Whether there are other mechanoreceptors that play a crucial role in regulating postnatal heart and lung development also needs to be explored. (2). Metabolic reprogramming also occurred under the condition of neonatal PO and VO (Figure 8). Is metabolic reprogramming the cause or result of cardiomyocyte proliferation? How do PO and VO initiate metabolic reprogramming of cardiomyocytes? (3) The chromatin openness of many genes associated with heart and lung development has been changed greatly by PO, VO, IPF, and RPF (data not shown). Epigenetics of hemodynamics should also be a future direction.

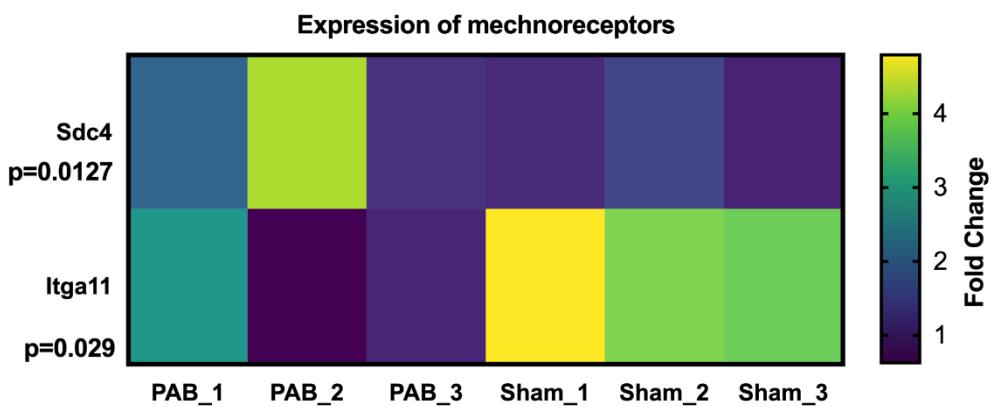


Figure 7. Sdc4 is upregulated and Itga11 is downregulated in the neonatal PO RV. Raw data have been deposited in NCBI's Gene Expression Omnibus database (<https://www.ncbi.nlm.nih.gov/geo>) with accession number GSE139561.

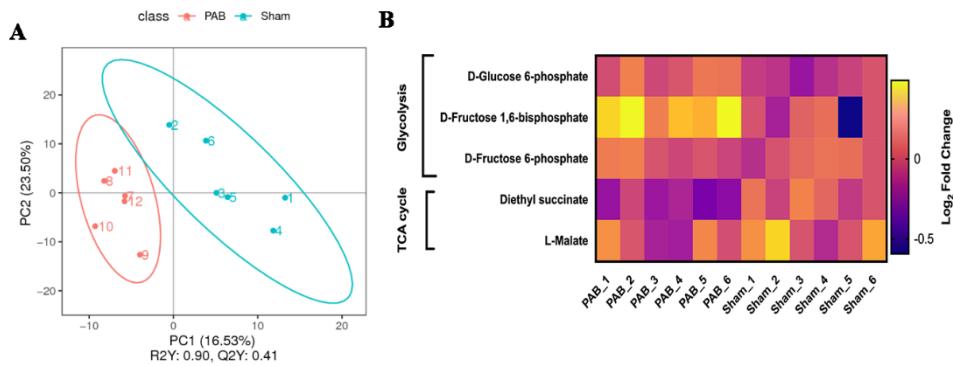


Figure 8. Metabolic reprogramming occurred with neonatal PO or VO. (A) Principal component analysis of RV metabolic reprogramming under neonatal PO. (B) Heat map of metabolites under neonatal PO. Note that glycolysis was upregulated, while the TCA cycle was downregulated under PO. Raw data have been deposited in NCBI's Gene Expression Omnibus database (<https://www.ncbi.nlm.nih.gov/geo>) with accession number GSE139561.

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Data Availability Statement: We encourage all authors of articles published in MDPI journals to share their research data. In this section, please provide details regarding where data supporting reported results can be found, including links to publicly archived datasets analyzed or generated during the study. Where no new data were created, or where data is unavailable due to privacy or ethical restrictions, a statement is still required. Suggested Data Availability Statements are available in section "MDPI Research Data Policies" at <https://www.mdpi.com/ethics>.

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References

1. Aird WC. Discovery of the cardiovascular system: from Galen to William Harvey. *J Thromb Haemost*. 2011 Jul;9 Suppl 1:118-29. doi: 10.1111/j.1538-7836.2011.04312.x. PMID: 21781247.
2. Bynum B, Bynum H. William Harvey's demonstration rod. *Lancet*. 2015 Nov 14;386(10007):1933. doi: 10.1016/S0140-6736(15)00819-3. Epub 2015 Nov 13. PMID: 26841739.
3. Dewan S, Krishnamurthy A, Kole D, Conca G, Kerckhoffs R, Puchalski MD, Omens JH, Sun H, Nigam V, McCulloch AD. Model of Human Fetal Growth in Hypoplastic Left Heart Syndrome: Reduced Ventricular Growth Due to Decreased Ventricular Filling and Altered Shape. *Front Pediatr*. 2017 Feb 22;5:25. doi: 10.3389/fped.2017.00025. PMID: 28275592; PMCID: PMC5319967.
4. Feit LR, Copel JA, Kleinman CS. Foramen ovale size in the normal and abnormal human fetal heart: an indicator of transatrial flow physiology. *Ultrasound Obstet Gynecol*. 1991 Sep 1;1(5):313-9. doi: 10.1046/j.1469-0705.1991.01050313.x. PMID: 12797035.
5. Tanai E, Frantz S. Pathophysiology of Heart Failure. *Compr Physiol*. 2015 Dec 15;6(1):187-214. doi: 10.1002/cphy.c140055. PMID: 26756631.
6. Thandavarayan RA, Chitturi KR, Guha A. Pathophysiology of Acute and Chronic Right Heart Failure. *Cardiol Clin*. 2020 May;38(2):149-160. doi: 10.1016/j.ccl.2020.01.009. Epub 2020 Mar 5. PMID: 32284093.
7. Nagalingam RS, Chattopadhyaya S, Al-Hattab DS, Cheung DYC, Schwartz LY, Jana S, Aroutiounova N, Ledingham DA, Moffatt TL, Landry NM, Bagchi RA, Dixon IMC, Wigle JT, Oudit GY, Kassiri Z, Jassal DS, Czubryt MP. Scleraxis and fibrosis in the pressure-overloaded heart. *Eur Heart J*. 2022 Dec 1;43(45):4739-4750. doi: 10.1093/eurheartj/ehac362. PMID: 36200607.

8. Burri PH. Fetal and postnatal development of the lung. *Annu Rev Physiol.* 1984;46:617-28. doi: 10.1146/annurev.ph.46.030184.003153. PMID: 6370120.
9. Whitsett JA, Wert SE, Trapnell BC. Genetic disorders influencing lung formation and function at birth. *Hum Mol Genet.* 2004 Oct 1;13 Spec No 2:R207-15. doi: 10.1093/hmg/ddh252. PMID: 15358727.
10. Pedra CA, Haddad J, Pedra SF, Peirone A, Pilla CB, Marin-Neto JA. Paediatric and congenital heart disease in South America: an overview. *Heart.* 2009 Sep;95(17):1385-92. doi: 10.1136/heart.2008.152017. Epub 2009 Jan 27. PMID: 19174420.
11. Fernandes PS, Magalhães LR, Pezzini TR, de Sousa Santos EF, Calderon MG. Congenital heart diseases trends in São Paulo State, Brazil: a national live birth data bank analysis. *World J Pediatr.* 2022 Jul;18(7):472-481. doi: 10.1007/s12519-022-00543-3. Epub 2022 Mar 26. PMID: 35338440.
12. Galdos FX, Guo Y, Paige SL, VanDusen NJ, Wu SM, Pu WT. Cardiac Regeneration: Lessons From Development. *Circ Res.* 2017 Mar 17;120(6):941-959. doi: 10.1161/CIRCRESAHA.116.309040. PMID: 28302741; PMCID: PMC5358810.
13. GBD 2017 Congenital Heart Disease Collaborators. Global, regional, and national burden of congenital heart disease, 1990-2017: a systematic analysis for the Global Burden of Disease Study 2017. *Lancet Child Adolesc Health.* 2020;4(3):185-200. doi:10.1016/S2352-4642(19)30402-X.
14. Tsao CW, Aday AW, Almarzooq ZI, Anderson CAM, Arora P, Avery CL, Baker-Smith CM, Beaton AZ, Boehme AK, Buxton AE, Commodore-Mensah Y, Elkind MSV, Evenson KR, Eze-Nliam C, Fugar S, Generoso G, Heard DG, Hiremath S, Ho JE, Kalani R, Kazi DS, Ko D, Levine DA, Liu J, Ma J, Magnani JW, Michos ED, Mussolino ME, Navaneethan SD, Parikh NI, Poudel R, Rezk-Hanna M, Roth GA, Shah NS, St-Onge MP, Thacker EL, Virani SS, Voeks JH, Wang NY, Wong ND, Wong SS, Yaffe K, Martin SS; American Heart Association Council on Epidemiology and Prevention Statistics Committee and Stroke Statistics Subcommittee. Heart Disease and Stroke Statistics-2023 Update: A Report From the American Heart Association. *Circulation.* 2023 Feb 21;147(8):e93-e621. doi: 10.1161/CIR.0000000000001123.
15. Corporan D, Segura A, Padala M. Ultrastructural Adaptation of the Cardiomyocyte to Chronic Mitral Regurgitation. *Front Cardiovasc Med.* 2021;8:714774. Published 2021 Oct 18. doi:10.3389/fcvm.2021.714774.
16. Offen SM, Baker D, Puranik R, Celermajer DS. Right ventricular volume and its relationship to functional tricuspid regurgitation. *Int J Cardiol Heart Vasc.* 2021 Dec 30;38:100940. doi: 10.1016/j.ijcha.2021.100940. PMID: 35024430; PMCID: PMC8728462.
17. Otani H, Kagaya Y, Yamane Y, Chida M, Ito K, Namiuchi S, Shiba N, Koseki Y, Ninomiya M, Ikeda J, Saito H, Maruoka S, Fujiwara T, Ido T, Ishide N, Shirato K. Long-term right ventricular volume overload increases myocardial fluorodeoxyglucose uptake in the interventricular septum in patients with atrial septal defect. *Circulation.* 2000 Apr 11;101(14):1686-92. doi: 10.1161/01.cir.101.14.1686. PMID: 10758051.
18. Goldberg SW, Fisher SA, Wehman B, Mehra MR. Adults with congenital heart disease and heart transplantation: optimizing outcomes. *J Heart Lung Transplant.* 2014 Sep;33(9):873-7. doi: 10.1016/j.healun.2014.05.001. Epub 2014 Jun 2. PMID: 25110322.
19. Palomo-López N, Escalona-Rodríguez S, Martín-Villén L, Herruzo-Avilés Á, Hinojosa-Pérez R, Escoresca-Ortega A, Porras-López M, Corcia-Palomo Y, Adsuar-Gómez A. Transplantation in Congenital Heart Disease: A Challenge. *Transplant Proc.* 2020 Mar;52(2):577-579. doi: 10.1016/j.transproceed.2019.12.027. Epub 2020 Feb 8. PMID: 32046860.
20. van der Bom T, Winter MM, Bouma BJ, Groenink M, Vliegen HW, Pieper PG, van Dijk AP, Sieswerda GT, Roos-Hesselink JW, Zwintzman AH, Mulder BJ. Effect of valsartan on systemic right ventricular function: a double-blind, randomized, placebo-controlled pilot trial. *Circulation.* 2013 Jan 22;127(3):322-30. doi: 10.1161/CIRCULATIONAHA.112.135392. Epub 2012 Dec 17. PMID: 23247302.
21. Mathur K, Hsu DT, Lamour JM, Aydin SI. Safety of Enalapril in Infants: Data from the Pediatric Heart Network Infant Single Ventricle Trial. *J Pediatr.* 2020 Dec;227:218-223. doi: 10.1016/j.jpeds.2020.07.058. Epub 2020 Aug 5. PMID: 32768465.
22. Hu Y, Li D, Zhou C, Xiao Y, Sun S, Jiang C, Chen L, Liu J, Zhang H, Li F, Hong H, Ye L. Molecular Changes in Prepubertal Left Ventricular Development Under Experimental Volume Overload. *Front Cardiovasc Med.* 2022 Apr 12;9:850248. doi: 10.3389/fcvm.2022.850248. PMID: 35497975; PMCID: PMC9039316.
23. Sun S, Hu Y, Xiao Y, Wang S, Jiang C, Liu J, Zhang H, Hong H, Li F, Ye L. Postnatal Right Ventricular Developmental Track Changed by Volume Overload. *J Am Heart Assoc.* 2021 Aug 17;10(16):e020854. doi: 10.1161/JAHA.121.020854. Epub 2021 Aug 13. PMID: 34387124; PMCID: PMC8475045.
24. Wang S, Jiang C, Zhao L, Sun S, Xiao Y, Ye L, Sun Q, Li J. Metabolic maturation during postnatal right ventricular development switches to heart-contraction regulation due to volume overload. *J Cardiol.* 2022 Jan;79(1):110-120. doi: 10.1016/j.jcc.2021.08.025. Epub 2021 Sep 10. PMID: 34518077.
25. Li DB, Xu XX, Hu YQ, Cui Q, Xiao YY, Sun SJ, Chen LJ, Ye LC, Sun Q. Congenital heart disease-associated pulmonary dysplasia and its underlying mechanisms. *Am J Physiol Lung Cell Mol Physiol.* 2023 Feb 1;324(2):L89-L101. doi: 10.1152/ajplung.00195.2022. Epub 2022 Dec 6. PMID: 36472329; PMCID: PMC9925164.

26. Zhou C, Li D, Cui Q, Sun Q, Hu Y, Xiao Y, Jiang C, Qiu L, Zhang H, Ye L, Sun Y. Ability of the Right Ventricle to Serve as a Systemic Ventricle in Response to the Volume Overload at the Neonatal Stage. *Biology (Basel)*. 2022 Dec 15;11(12):1831. doi: 10.3390/biology11121831. PMID: 36552341; PMCID: PMC9775952.

27. Zhou C, Sun S, Hu M, Xiao Y, Yu X, Ye L, Qiu L. Downregulated developmental processes in the postnatal right ventricle under the influence of a volume overload. *Cell Death Discov*. 2021 Aug 7;7(1):208. doi: 10.1038/s41420-021-00593-y. PMID: 34365468; PMCID: PMC8349357.

28. Li D, Qiu L, Hong H, Chen H, Zhao P, Xiao Y, Zhang H, Sun Q, Ye L. A neonatal rat model of pulmonary vein stenosis. *Cell Biosci*. 2023 Jun 19;13(1):112. doi: 10.1186/s13578-023-01058-8. PMID: 37337290; PMCID: PMC10278335.

29. Li D, Wang J, Fang Y, Hu Y, Xiao Y, Cui Q, Jiang C, Sun S, Chen H, Ye L, Sun Q. Impaired cell-cell communication and axon guidance because of pulmonary hypoperfusion during postnatal alveolar development. *Respir Res*. 2023 Jan 11;24(1):12. doi: 10.1186/s12931-023-02319-3. PMID: 36631871; PMCID: PMC9833865.

30. Wang S, Ye L, Hong H, Tang C, Li M, Zhang Z, Liu J. A neonatal rat model of increased right ventricular afterload by pulmonary artery banding. *J Thorac Cardiovasc Surg*. 2017 Nov;154(5):1734-1739. doi: 10.1016/j.jtcvs.2017.06.016. Epub 2017 Jun 13. PMID: 28697895.

31. Ye L, Wang S, Xiao Y, Jiang C, Huang Y, Chen H, Zhang H, Zhang H, Liu J, Xu Z, Hong H. Pressure Overload Greatly Promotes Neonatal Right Ventricular Cardiomyocyte Proliferation: A New Model for the Study of Heart Regeneration. *J Am Heart Assoc*. 2020 Jun 2;9(11):e015574. doi: 10.1161/JAHA.119.015574. Epub 2020 May 30. PMID: 32475201; PMCID: PMC7429015.

32. Malek Mohammadi M, Abouissa A, Heineke J. A surgical mouse model of neonatal pressure overload by transverse aortic constriction. *Nat Protoc*. 2021 Feb;16(2):775-790. doi: 10.1038/s41596-020-00434-9. Epub 2020 Dec 16. PMID: 33328612.

33. Sun S, Zhu H, Wang S, Xu X, Ye L. Establishment and Confirmation of a Postnatal Right Ventricular Volume Overload Mouse Model. *J Vis Exp*. 2023 Jun 9;(196). doi: 10.3791/65372. PMID: 37358276.

34. McLennan DI, Solano ECR, Handler SS, Lincoln J, Mitchell ME, Kirkpatrick EC. Pulmonary Vein Stenosis: Moving From Past Pessimism to Future Optimism. *Front Pediatr*. 2021 Oct 5;9:747812. doi: 10.3389/fped.2021.747812. PMID: 34676188; PMCID: PMC8524035..

35. Otani H, Kagaya Y, Yamane Y, Chida M, Ito K, Namiuchi S, Shiba N, Koseki Y, Ninomiya M, Ikeda J, Saito H, Maruoka S, Fujiwara T, Ido T, Ishide N, Shirato K. Long-term right ventricular volume overload increases myocardial fluorodeoxyglucose uptake in the interventricular septum in patients with atrial septal defect. *Circulation*. 2000 Apr 11;101(14):1686-92. doi: 10.1161/01.cir.101.14.1686. PMID: 10758051.

36. Abu-Shaweesh JM, Almidani E. PDA: Does it matter? *Int J Pediatr Adolesc Med*. 2020 Mar;7(1):9-12. doi: 10.1016/j.ijpam.2019.12.001. Epub 2019 Dec 3. PMID: 32373696; PMCID: PMC7193069.

37. Hegde M, Manjunath SC, Usha MK. Isolated Partial Anomalous Pulmonary Venous Connection: Development of Volume Overload and Elevated Estimated Pulmonary Pressure in Adults. *J Clin Imaging Sci*. 2019 Jun 14;9:29. doi: 10.25259/JCIS-8-2019. PMID: 31508264; PMCID: PMC6712552.

38. Calkoen EE, Hazekamp MG, Blom NA, Elders BB, Gittenberger-de Groot AC, Haak MC, Bartelings MM, Roest AA, Jongbloed MR. Atrioventricular septal defect: From embryonic development to long-term follow-up. *Int J Cardiol*. 2016 Jan 1;202:784-95. doi: 10.1016/j.ijcard.2015.09.081. Epub 2015 Sep 26. PMID: 26476030.

39. Grosse-Brockhoff F, Loogen F. Ventricular septal defect. *Circulation*. 1968 Jul;38(1 Suppl):13-20. doi: 10.1161/01.cir.38.1s5.v-13. PMID: 5712380.

40. Hahn RT. Tricuspid Regurgitation. *N Engl J Med*. 2023 May 18;388(20):1876-1891. doi: 10.1056/NEJMra2216709. PMID: 37195943.

41. Holst KA, Connolly HM, Dearani JA. Ebstein's Anomaly. *Methodist Debakey Cardiovasc J*. 2019 Apr-Jun;15(2):138-144. doi: 10.14797/mdcj-15-2-138. PMID: 31384377; PMCID: PMC6668741.

42. Bouzas B, Kilner PJ, Gatzoulis MA. Pulmonary regurgitation: not a benign lesion. *Eur Heart J*. 2005 Mar;26(5):433-9. doi: 10.1093/eurheartj/ehi091. Epub 2005 Jan 7. PMID: 15640261.

43. Tatewaki H, Shiose A. Pulmonary valve replacement after repaired Tetralogy of Fallot. *Gen Thorac Cardiovasc Surg*. 2018 Sep;66(9):509-515. doi: 10.1007/s11748-018-0931-0. Epub 2018 May 19. PMID: 29779123.

44. Said SM, Mainwaring RD, Ma M, Tacy TA, Hanley FL. Pulmonary Valve Repair for Patients With Acquired Pulmonary Valve Insufficiency. *Ann Thorac Surg*. 2016 Jun;101(6):2294-301. doi: 10.1016/j.athoracsur.2016.01.035. Epub 2016 Apr 12. PMID: 27083251.

45. Sanz J, Sánchez-Quintana D, Bossone E, Bogaard HJ, Naeije R. Anatomy, Function, and Dysfunction of the Right Ventricle: JACC State-of-the-Art Review. *J Am Coll Cardiol*. 2019 Apr 2;73(12):1463-1482. doi: 10.1016/j.jacc.2018.12.076. PMID: 30922478.

46. Sharma A, Stan MN. Thyrotoxicosis: Diagnosis and Management. *Mayo Clin Proc*. 2019 Jun;94(6):1048-1064. doi: 10.1016/j.mayocp.2018.10.011. Epub 2019 Mar 25. PMID: 30922695.

47. 47.Metcalf MK, Rychik J. Outcomes in Hypoplastic Left Heart Syndrome. *Pediatr Clin North Am.* 2020 Oct;67(5):945-962. doi: 10.1016/j.pcl.2020.06.008. PMID: 32888691.

48. 48.Cohen ML, Spray T, Gutierrez F, Barzilai B, Bauwens D. Congenital tricuspid valve stenosis with atrial septal defect and left anterior fascicular block. *Clin Cardiol.* 1990 Jul;13(7):497-9. doi: 10.1002/clc.4960130713. PMID: 2364584.

49. 49.Enriquez-Sarano M, Akins CW, Vahanian A. Mitral regurgitation. *Lancet.* 2009 Apr 18;373(9672):1382-94. doi: 10.1016/S0140-6736(09)60692-9. Epub 2009 Apr 6. PMID: 19356795.

50. 50.Cramariuc D, Bahlmann E, Gerdts E. Grading of Aortic Stenosis: Is it More Complicated in Women? *Eur Cardiol.* 2022 Nov 1;17:e21. doi: 10.15420/ecr.2022.13. PMID: 36643071; PMCID: PMC9820123.

51. 51.Bravo-Jaimes K, Prakash SK. Genetics in bicuspid aortic valve disease: Where are we? *Prog Cardiovasc Dis.* 2020 Jul-Aug;63(4):398-406. doi: 10.1016/j.pcad.2020.06.005. Epub 2020 Jun 27. PMID: 32599026; PMCID: PMC7530017.

52. 52.Mandras SA, Mehta HS, Vaidya A. Pulmonary Hypertension: A Brief Guide for Clinicians. *Mayo Clin Proc.* 2020 Sep;95(9):1978-1988. doi: 10.1016/j.mayocp.2020.04.039. PMID: 32861339.

53. 53.Chetan D, Mertens LL. Challenges in diagnosis and management of coarctation of the aorta. *Curr Opin Cardiol.* 2022 Jan 1;37(1):115-122. doi: 10.1097/HCO.0000000000000934. PMID: 34857719.

54. 54.Dickey J, Phelan C. Unrepaired Tetralogy of Fallot in Adulthood. *N Engl J Med.* 2020 Jun 18;382(25):e97. doi: 10.1056/NEJMcm1912128. PMID: 32558471.

55. 55.Rali PM, Criner GJ. Submassive Pulmonary Embolism. *Am J Respir Crit Care Med.* 2018 Sep 1;198(5):588-598. doi: 10.1164/rccm.201711-2302CI. PMID: 29672125.

56. 56.Patel AB, Ratnayaka K, Bergersen L. A review: Percutaneous pulmonary artery stenosis therapy: state-of-the-art and look to the future. *Cardiol Young.* 2019 Feb;29(2):93-99. doi: 10.1017/S1047951118001087. Epub 2018 Dec 27. PMID: 30587259.

57. 57.Marchini F, Meossi S, Passarini G, Campo G, Pavasini R. Pulmonary Valve Stenosis: From Diagnosis to Current Management Techniques and Future Prospects. *Vasc Health Risk Manag.* 2023 Jun 30;19:379-390. doi: 10.2147/VHRM.S380240. PMID: 37416511; PMCID: PMC10320808.

58. 58.Maron BJ, Maron MS. Hypertrophic cardiomyopathy. *Lancet.* 2013 Jan 19;381(9862):242-55. doi: 10.1016/S0140-6736(12)60397-3. Epub 2012 Aug 6. PMID: 22874472.

59. 59.Abassi H, Gavotto A, Picot MC, Bertet H, Matecki S, Guillaumont S, Moniotte S, Auquier P, Moreau J, Amedro P. Impaired pulmonary function and its association with clinical outcomes, exercise capacity and quality of life in children with congenital heart disease. *Int J Cardiol.* 2019 Jun 15;285:86-92. doi: 10.1016/j.ijcard.2019.02.069. Epub 2019 Mar 1. PMID: 30857849.

60. 60.Diller GP, Dimopoulos K, Okonko D, Li W, Babu-Narayan SV, Broberg CS, Johansson B, Bouzas B, Mullen MJ, Poole-Wilson PA, Francis DP, Gatzoulis MA. Exercise intolerance in adult congenital heart disease: comparative severity, correlates, and prognostic implication. *Circulation.* 2005 Aug 9;112(6):828-35. doi: 10.1161/CIRCULATIONAHA.104.529800. Epub 2005 Aug 1. PMID: 16061735.

61. 61.Talman V, Teppo J, Pöhö P, Movahedi P, Vaikkinen A, Karhu ST, Trošt K, Suvitaival T, Heikkonen J, Pahikkala T, Kotiaho T, Kostiainen R, Varjosalo M, Ruskoaho H. Molecular Atlas of Postnatal Mouse Heart Development. *J Am Heart Assoc.* 2018 Oct 16;7(20):e010378. doi: 10.1161/JAHA.118.010378. PMID: 30371266; PMCID: PMC6474944.

62. 62.Guo Y, Pu WT. Cardiomyocyte Maturation: New Phase in Development. *Circ Res.* 2020 Apr 10;126(8):1086-1106. doi: 10.1161/CIRCRESAHA.119.315862. Epub 2020 Apr 9. PMID: 32271675; PMCID: PMC7199445.

63. 63.Papanicolaou KN, Kikuchi R, Ngoh GA, Coughlan KA, Dominguez I, Stanley WC, Walsh K. Mitofusins 1 and 2 are essential for postnatal metabolic remodeling in heart. *Circ Res.* 2012 Sep 28;111(8):1012-26. doi: 10.1161/CIRCRESAHA.112.274142. Epub 2012 Aug 17. PMID: 22904094; PMCID: PMC3518037.

64. 64.Piquereau J, Novotova M, Fortin D, Garnier A, Ventura-Clapier R, Veksler V, Joubert F. Postnatal development of mouse heart: formation of energetic microdomains. *J Physiol.* 2010 Jul 1;588(Pt 13):2443-54. doi: 10.1113/jphysiol.2010.189670. Epub 2010 May 17. PMID: 20478976; PMCID: PMC2915519.

65. 65.Reynolds JO, Chiang DY, Wang W, Beavers DL, Dixit SS, Skapura DG, Landstrom AP, Song LS, Ackerman MJ, Wehrens XH. Junctophilin-2 is necessary for T-tubule maturation during mouse heart development. *Cardiovasc Res.* 2013 Oct 1;100(1):44-53. doi: 10.1093/cvr/cvt133. Epub 2013 May 27. PMID: 23715556; PMCID: PMC3778955.

66. 66.Hong T, Yang H, Zhang SS, Cho HC, Kalashnikova M, Sun B, Zhang H, Bhargava A, Grabe M, Olgin J, Gorelik J, Marbán E, Jan LY, Shaw RM. Cardiac BIN1 folds T-tubule membrane, controlling ion flux and limiting arrhythmia. *Nat Med.* 2014 Jun;20(6):624-32. doi: 10.1038/nm.3543. Epub 2014 May 18. PMID: 24836577; PMCID: PMC4048325.

67. 67.Ong LP, Bargehr J, Knight-Schrijver VR, Lee J, Colzani M, Bayraktar S, Bernard WG, Marchiano S, Bertero A, Murry CE, Gambardella L, Sinha S. Epicardially secreted fibronectin drives cardiomyocyte maturation in 3D-engineered heart tissues. *Stem Cell Reports.* 2023 Mar 27;S2213-6711(23)00062-0. doi: 10.1016/j.stemcr.2023.03.002. Epub ahead of print. PMID: 37001515.

68. 68.Thébaud B, Goss KN, Laughon M, Whitsett JA, Abman SH, Steinhorn RH, Aschner JL, Davis PG, McGrath-Morrow SA, Soll RF, Jobe AH. Bronchopulmonary dysplasia. *Nat Rev Dis Primers*. 2019 Nov 14;5(1):78. doi: 10.1038/s41572-019-0127-7. PMID: 31727986; PMCID: PMC6986462.

69. 69.Salaets T, Aertgeerts M, Gie A, Vignero J, de Winter D, Regin Y, Jimenez J, Vande Velde G, Allegaert K, Deprest J, Toelen J. Preterm birth impairs postnatal lung development in the neonatal rabbit model. *Respir Res*. 2020 Feb 21;21(1):59. doi: 10.1186/s12931-020-1321-6. PMID: 32085773; PMCID: PMC7035772.

70. 70.Jobe AH. Animal Models, Learning Lessons to Prevent and Treat Neonatal Chronic Lung Disease. *Front Med (Lausanne)*. 2015 Aug 7;2:49. doi: 10.3389/fmed.2015.00049. PMID: 26301222; PMCID: PMC4528292.

71. 71.Zepp JA, Morley MP, Loebel C, Kremp MM, Chaudhry FN, Basil MC, Leach JP, Liberti DC, Niethamer TK, Ying Y, Jayachandran S, Babu A, Zhou S, Frank DB, Burdick JA, Morrisey EE. Genomic, epigenomic, and biophysical cues controlling the emergence of the lung alveolus. *Science*. 2021 Mar 12;371(6534):eabc3172. doi: 10.1126/science.abc3172. PMID: 33707239; PMCID: PMC8320017.

72. 72.Frank BS, Ivy DD. Pediatric Pulmonary Arterial Hypertension. *Pediatr Clin North Am*. 2020 Oct;67(5):903-921. doi: 10.1016/j.pcl.2020.06.005. PMID: 32888689.

73. 73.vy DD, Abman SH, Barst RJ, Berger RM, Bonnet D, Fleming TR, Haworth SG, Raj JU, Rosenzweig EB, Schulze Neick I, Steinhorn RH, Beghetti M. Pediatric pulmonary hypertension. *J Am Coll Cardiol*. 2013 Dec 24;62(25 Suppl):D117-26. doi: 10.1016/j.jacc.2013.10.028. PMID: 24355636.

74. 74.Marino BS, Cassedy A, Brown KL, Franklin R, Gaynor JW, Cvetkovic M, Laker S, Levinson K, MacGloin H, Mahony L, McQuillan A, Mussatto K, O'Shea D, Newburger J, Sykes M, Teele SA, Wernovsky G, Wray J. Long-Term Quality of Life in Congenital Heart Disease Surgical Survivors: Multicenter Retrospective Study of Surgical and ICU Explanatory Factors. *Pediatr Crit Care Med*. 2023 May 1;24(5):391-398. doi: 10.1097/PCC.0000000000003190. Epub 2023 Feb 21. PMID: 37140331.

75. 75.Wang T, Chen L, Yang T, Huang P, Wang L, Zhao L, Zhang S, Ye Z, Chen L, Zheng Z, Qin J. Congenital Heart Disease and Risk of Cardiovascular Disease: A Meta-Analysis of Cohort Studies. *J Am Heart Assoc*. 2019 May 21;8(10):e012030. doi: 10.1161/JAHA.119.012030. PMID: 31070503; PMCID: PMC6585327.

76. 76.Shi B, Zhang X, Song Z, Dai Z, Luo K, Chen B, Zhou Z, Cui Y, Feng B, Zhu Z, Zheng J, Zhang H, He X. Targeting gut microbiota-derived kynurenone to predict and protect the remodeling of the pressure-overloaded young heart. *Sci Adv*. 2023 Jul 14;9(28):eadg7417. doi: 10.1126/sciadv.adg7417. Epub 2023 Jul 14. PMID: 37450589; PMCID: PMC10348671.

77. 77.Du J, Zheng L, Gao P, Yang H, Yang WJ, Guo F, Liang R, Feng M, Wang Z, Zhang Z, Bai L, Bu Y, Xing S, Zheng W, Wang X, Quan L, Hu X, Wu H, Chen Z, Chen L, Wei K, Zhang Z, Zhu X, Zhang X, Tu Q, Zhao SM, Lei X, Xiong JW. A small-molecule cocktail promotes mammalian cardiomyocyte proliferation and heart regeneration. *Cell Stem Cell*. 2022 Apr 7;29(4):545-558.e13. doi: 10.1016/j.stem.2022.03.009. PMID: 35395187.

78. 78.Wu Y, Zhou L, Liu H, Duan R, Zhou H, Zhang F, He X, Lu D, Xiong K, Xiong M, Zhuang J, Liu Y, Li L, Liang D, Chen YH. LRP6 downregulation promotes cardiomyocyte proliferation and heart regeneration. *Cell Res*. 2021 Apr;31(4):450-462. doi: 10.1038/s41422-020-00411-7. Epub 2020 Sep 24. PMID: 32973339; PMCID: PMC8114926.

79. 79.Liu H, Zhang CH, Ammanamanchi N, Suresh S, Lewarchik C, Rao K, Uys GM, Han L, Abrial M, Yimlamai D, Ganapathy B, Guillermier C, Chen N, Khaladkar M, Spaethling J, Eberwine JH, Kim J, Walsh S, Choudhury S, Little K, Francis K, Sharma M, Viegas M, Bais A, Kostka D, Ding J, Bar-Joseph Z, Wu Y, Yechoor V, Moulik M, Johnson J, Weinberg J, Reyes-Múgica M, Steinhauser ML, Kühn B. Control of cytokinesis by β -adrenergic receptors indicates an approach for regulating cardiomyocyte endowment. *Sci Transl Med*. 2019 Oct 9;11(513):eaaw6419. doi: 10.1126/scitranslmed.aaw6419. PMID: 31597755; PMCID: PMC8132604.

80. 80.Yutzey KE. Cytokinesis, Beta-Blockers, and Congenital Heart Disease. *N Engl J Med*. 2020 Jan 16;382(3):291-293. doi: 10.1056/NEJMcb1913824. PMID: 31940705.

81. 81.Gu J, Chen X, Jin Y, Liu M, Xu Q, Liu X, Luo Z, Ling S, Liu N, Liu S. A Neonatal Mouse Model for Pressure Overload: Myocardial Response Corresponds to Severity. *Front Cardiovasc Med*. 2021 May 21;8:660246. doi: 10.3389/fcvm.2021.660246. PMID: 34095250; PMCID: PMC8175619.

82. 82.Malek Mohammadi M, Abouissa A, Azizah I, Xie Y, Cordero J, Shirvani A, Gigina A, Engelhardt M, Trogisch FA, Geffers R, Dobreva G, Bauersachs J, Heineke J. Induction of cardiomyocyte proliferation and angiogenesis protects neonatal mice from pressure overload-associated maladaptation. *JCI Insight*. 2019 Jul 23;5(16):e128336. doi: 10.1172/jci.insight.128336. PMID: 31335322; PMCID: PMC6777810.

83. 83.Ding X, Wang S, Wang Y, Yang J, Bao N, Liu J, Zhang Z. Neonatal Heart Responds to Pressure Overload With Differential Alterations in Various Cardiomyocyte Maturation Programs That Accommodate Simultaneous Hypertrophy and Hyperplasia. *Front Cell Dev Biol*. 2020 Nov 19;8:596960. doi: 10.3389/fcell.2020.596960. PMID: 33330485; PMCID: PMC7710899.

84. 84.Liu X, Pu W, He L, Li Y, Zhao H, Li Y, Liu K, Huang X, Weng W, Wang QD, Shen L, Zhong T, Sun K, Ardehali R, He B, Zhou B. Cell proliferation fate mapping reveals regional cardiomyocyte cell-cycle activity

in subendocardial muscle of left ventricle. *Nat Commun.* 2021 Oct 1;12(1):5784. doi: 10.1038/s41467-021-25933-5. PMID: 34599161; PMCID: PMC8486850.

85. 85.Bossers GPL, Günthel M, van der Feen DE, Hagedorn QAJ, Koop AC, van Duijvenboden K, Barnett P, Borgdorff MAJ, Christoffels VM, Silljé HHW, Berger RMF, Bartelds B. Neuregulin-1 enhances cell-cycle activity, delays cardiac fibrosis, and improves cardiac performance in rat pups with right ventricular pressure load. *J Thorac Cardiovasc Surg.* 2022 Dec;164(6):e493-e510. doi: 10.1016/j.jtcvs.2021.10.045. Epub 2021 Nov 3. PMID: 34922752.

86. 86.Cui Q, Sun S, Zhu H, Xiao Y, Jiang C, Zhang H, Liu J, Ye L, Shen J. Volume Overload Initiates an Immune Response in the Right Ventricle at the Neonatal Stage. *Front Cardiovasc Med.* 2021 Nov 16;8:772336. doi: 10.3389/fcvm.2021.772336. PMID: 34869688; PMCID: PMC8635051.

87. 87.Canseco DC, Kimura W, Garg S, Mukherjee S, Bhattacharya S, Abdisalaam S, Das S, Asaithamby A, Mammen PP, Sadek HA. Human ventricular unloading induces cardiomyocyte proliferation. *J Am Coll Cardiol.* 2015 Mar 10;65(9):892-900. doi: 10.1016/j.jacc.2014.12.027. Epub 2015 Jan 21. PMID: 25618530; PMCID: PMC4488905.

88. 88.Davoodianidali M, Punzmann H, Kellay H, Xia H, Shats M, Francois N. Fluctuation-Induced Interaction in Turbulent Flows. *Phys Rev Lett.* 2022 Jan 14;128(2):024503. doi: 10.1103/PhysRevLett.128.024503. PMID: 35089756.

89. 89.Butcher DT, Alliston T, Weaver VM. A tense situation: forcing tumour progression. *Nat Rev Cancer.* 2009 Feb;9(2):108-22. doi: 10.1038/nrc2544. PMID: 19165226; PMCID: PMC2649117.

90. 90.Mehta V, Pang KL, Rozbesky D, Nather K, Keen A, Lachowski D, Kong Y, Karia D, Ameismeier M, Huang J, Fang Y, Del Rio Hernandez A, Reader JS, Jones EY, Tzima E. The guidance receptor plexin D1 is a mechanosensor in endothelial cells. *Nature.* 2020 Feb;578(7794):290-295. doi: 10.1038/s41586-020-1979-4. Epub 2020 Feb 5.

91. 91.Guo Y, Pu WT. Cardiomyocyte Maturation: New Phase in Development. *Circ Res.* 2020 Apr 10;126(8):1086-1106. doi: 10.1161/CIRCRESAHA.119.315862. Epub 2020 Apr 9. PMID: 32271675; PMCID: PMC7199445.

92. 92.Karbassi E, Fenix A, Marchiano S, Muraoka N, Nakamura K, Yang X, Murry CE. Cardiomyocyte maturation: advances in knowledge and implications for regenerative medicine. *Nat Rev Cardiol.* 2020 Jun;17(6):341-359. doi: 10.1038/s41569-019-0331-x. Epub 2020 Feb 3. PMID: 32015528; PMCID: PMC7239749.

93. 93.Wang Y, Yu M, Hao K, Lei W, Tang M, Hu S. Cardiomyocyte Maturation-the Road is not Obstructed. *Stem Cell Rev Rep.* 2022 Dec;18(8):2966-2981. doi: 10.1007/s12015-022-10407-y. Epub 2022 Jul 5. PMID: 3578883.

94. 94.Kannan S, Kwon C. Regulation of cardiomyocyte maturation during critical perinatal window. *J Physiol.* 2020 Jul;598(14):2941-2956. doi: 10.1113/JP276754. Epub 2019 Jan 15. PMID: 30571853; PMCID: PMC7682257.

95. 95.Zhao MT, Ye S, Su J, Garg V. Cardiomyocyte Proliferation and Maturation: Two Sides of the Same Coin for Heart Regeneration. *Front Cell Dev Biol.* 2020 Oct 15;8:594226. doi: 10.3389/fcell.2020.594226. PMID: 33178704; PMCID: PMC7593613.

96. 96.Wang WE, Li L, Xia X, Fu W, Liao Q, Lan C, Yang D, Chen H, Yue R, Zeng C, Zhou L, Zhou B, Duan DD, Chen X, Houser SR, Zeng C. Dedifferentiation, Proliferation, and Redifferentiation of Adult Mammalian Cardiomyocytes After Ischemic Injury. *Circulation.* 2017 Aug 29;136(9):834-848. doi: 10.1161/CIRCULATIONAHA.116.024307. Epub 2017 Jun 22. PMID: 28642276; PMCID: PMC5575972.

97. 97.Krieger EV, Zeppenfeld K, DeWitt ES, Duarte VE, Egbe AC, Haeffele C, Lin KY, Robinson MR, Sillman C, Upadhyay S; American Heart Association Adults With Congenital Heart Disease Committee of the Council on Lifelong Congenital Heart Disease and Heart Health in the Young and Council on Clinical Cardiology. Arrhythmias in Repaired Tetralogy of Fallot: A Scientific Statement From the American Heart Association. *Circ Arrhythm Electrophysiol.* 2022 Nov;15(11):e000084. doi: 10.1161/HAE.0000000000000084. Epub 2022 Oct 20. PMID: 36263773.

98. 98.Badagliacca R, Papa S, Valli G, Pezzuto B, Poscia R, Reali M, Manzi G, Giannetta E, Berardi D, Sciomber S, Palange P, Fedele F, Naeije R, Vizza CD. Right ventricular dyssynchrony and exercise capacity in idiopathic pulmonary arterial hypertension. *Eur Respir J.* 2017 Jun 1;49(6):1601419. doi: 10.1183/13993003.01419-2016. PMID: 28572119.

99. 99.Van Aerde N, Meersseman P, Debaveye Y, Wilmer A, Casaer MP, Gunst J, Wauters J, Wouters PJ, Goetschalckx K, Gosselink R, Van den Berghe G, Hermans G. Aerobic exercise capacity in long-term survivors of critical illness: secondary analysis of the post-EPaNIC follow-up study. *Intensive Care Med.* 2021 Dec;47(12):1462-1471. doi: 10.1007/s00134-021-06541-9. Epub 2021 Nov 8. PMID: 34750648; PMCID: PMC8575347.

100. 100.McKenzie DC. Respiratory physiology: adaptations to high-level exercise. *Br J Sports Med.* 2012 May;46(6):381-4. doi: 10.1136/bjsports-2011-090824. Epub 2012 Jan 20. PMID: 22267571.

101. 101.De Troyer A, Yernault JC, Englert M. Lung hypoplasia in congenital pulmonary valve stenosis. *Circulation.* 1977 Oct;56(4 Pt 1):647-51. doi: 10.1161/01.cir.56.4.647. PMID: 902390.

102. 102. Beauchemin KJ, Wells JM, Kho AT, Philip VM, Kamir D, Kohane IS, Graber JH, Bult CJ. Temporal dynamics of the developing lung transcriptome in three common inbred strains of laboratory mice reveals multiple stages of postnatal alveolar development. *PeerJ*. 2016 Aug 9;4:e2318. doi: 10.7717/peerj.2318. PMID: 27602285; PMCID: PMC4991849.
103. 103. Cantuti-Castelvetri L, Ojha R, Pedro LD, Djannatian M, Franz J, Kuivanen S, van der Meer F, Kallio K, Kaya T, Anastasina M, Smura T, Levanov L, Szirovicza L, Tobi A, Kallio-Kokko H, Österlund P, Joensuu M, Meunier FA, Butcher SJ, Winkler MS, Mollenhauer B, Helenius A, Gokce O, Teesalu T, Hepojoki J, Vapalahti O, Stadelmann C, Balistreri G, Simons M. Neuropilin-1 facilitates SARS-CoV-2 cell entry and infectivity. *Science*. 2020 Nov 13;370(6518):856-860. doi: 10.1126/science.abd2985. Epub 2020 Oct 20. PMID: 33082293; PMCID: PMC7857391.
104. 104. Namba F. An experimental animal model of bronchopulmonary dysplasia: Secondary publication. *Pediatr Int*. 2021 May;63(5):504-509. doi: 10.1111/ped.14612. Epub 2021 May 7. PMID: 33465831.
105. 105. Jobe AH. Animal Models, Learning Lessons to Prevent and Treat Neonatal Chronic Lung Disease. *Front Med (Lausanne)*. 2015 Aug 7;2:49. doi: 10.3389/fmed.2015.00049. PMID: 26301222; PMCID: PMC4528292.
106. 106. Lee JW, Davis JM. Future applications of antioxidants in premature infants. *Curr Opin Pediatr*. 2011 Apr;23(2):161-6. doi: 10.1097/MOP.0b013e3283423e51. PMID: 21150443; PMCID: PMC3289059.
107. 107. Thébaud B, Goss KN, Laughon M, Whitsett JA, Abman SH, Steinhorn RH, Aschner JL, Davis PG, McGrath-Morrow SA, Soll RF, Jobe AH. Bronchopulmonary dysplasia. *Nat Rev Dis Primers*. 2019 Nov 14;5(1):78. doi: 10.1038/s41572-019-0127-7. PMID: 31727986; PMCID: PMC6986462.
108. 108. Mourani PM, Sontag MK, Younoszai A, Miller JI, Kinsella JP, Baker CD, Poindexter BB, Ingram DA, Abman SH. Early pulmonary vascular disease in preterm infants at risk for bronchopulmonary dysplasia. *Am J Respir Crit Care Med*. 2015 Jan 1;191(1):87-95. doi: 10.1164/rccm.201409-1594OC. PMID: 25389562; PMCID: PMC4299632.
109. 109. Herum KM, Lunde IG, Skrbic B, Louch WE, Hasic A, Boye S, Unger A, Brorson SH, Sjaastad I, Tønnessen T, Linke WA, Gomez MF, Christensen G. Syndecan-4 is a key determinant of collagen cross-linking and passive myocardial stiffness in the pressure-overloaded heart. *Cardiovasc Res*. 2015 May 1;106(2):217-26. doi: 10.1093/cvr/cvv002. Epub 2015 Jan 12. PMID: 25587045.
110. 110. Yu C, Qiu M, Zhang Z, Song X, Du H, Peng H, Li Q, Yang L, Xiong X, Xia B, Hu C, Chen J, Jiang X, Yang C. Transcriptome sequencing reveals genes involved in cadmium-triggered oxidative stress in the chicken heart. *Poult Sci*. 2021 Mar;100(3):100932. doi: 10.1016/j.psj.2020.12.029. Epub 2021 Jan 23. PMID: 33652545; PMCID: PMC7936198.

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