



Supplementary Information

Rapid Antibacterial Activity Assessment of Chimeric Lysin

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Table S1. Information of SH3 domains used in this study.

Abbreviated name	Sequence identity to K _{SH3} (%)	Genbank Protein Full name	Genbank accession No.	Amino acid Region	Amino acid Length
L25(K _{SH3})	100	LysK [<i>Staphylococcus phage P4W</i>]	AFN38929	409-495	87
ClyC _{SH3}	94.2	Lysin [<i>Staphylococcus phage PALS_1</i>]	QDJ97474.1	123-209	87
vB _{SH3}	93.1	N-acetylmuramoyl-L-alanine amidase [<i>Staphylococcus phage vB_Sau_Clo6</i>]	ARM69344.1	409-495	87
tras14 _{SH3}	40.2	endolysin [<i>Staphylococcus phage trsa14</i>]	AWH13066.1	395-481	87
ClyO _{SH3}	11.4	PlySs2 family phage lysin [<i>Streptococcus suis</i>]	WP_024417665.1	162-245	84
Lsp _{SH3}	9.1	Lysostaphin [<i>Staphylococcus simulans</i>]	WP_013012297.1	369-452	84
2638A _{SH3}	6.8	endolysin [<i>Staphylococcus phage 2638A</i>]	YP_239818.1	401-486	86
2958 _{SH3}	4.5	endolysin [<i>Staphylococcus phage phi2958PVL</i>]	YP_002268027.1	405-484	80

Table S2. Information of lysostaphin and chimeric lysins used in this study.

Protein name	Genbank accession No.	Amino acid Region	Amino acid Length	Reference
Lysostaphin	WP_013012297.1	207-452	246	[45]
Lsp		207-368	162	
ClyC	AGO49867.1	1-171	171	[24]
	InsertVD	172-173	2	
	QDJ97474.1	102-209	108	
ClyO	WP_258391128.1	1-157	157	[33]
	InsertG	158	1	
	WP_024417665.1 (R195C)	146-245	100	

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Table S3. PCR primers used for amplification of 10 linker candidates

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Primer name	Sequence (5' -3')	Usage
P T7-F	TCGAAATTAATACGACTCACTATAG	IVTC/TL
T term-R	GAGGGGATCCAGATCTTGGGAT	amplification for all
P_gCHAP-L-F	<u>CTTTAAGAAGGAGATATACATATGG-</u> <u>CATCATCTTTTAATCACC</u>	Linker & SH3 domain
P_gCHAP-L-R	<u>GTGATTAATAA-</u> <u>GATGATGCCATATGTATATCTCCTTCTTAAAG</u>	SOE-PCR for all
K _{SH3} _T-L-F	GTTGCATGGGGTGTTTTCAAAAAGGGTCATCATCAC- CATCAC	Linker & SOE-PCR
K _{SH3} _T-L-R	GTGATGGTGTATGATGACCCTTTT- GAAAACACCCCATGCAAC	for all
gCHAP_L21-L-F	<u>GCTATGCATTCATCCATTCTGCATCTACTCCGGCAAC</u>	SOE-PCR
gCHAP_L21-L-R	GTGCCGGAGTAGATGCAGA <u>ATGGATGAATGCATAGC</u>	for linker21
gCHAP_L24-L-F	<u>GCTATGCATTCATCCATAAAACTTCTTCTGCATCTAC</u>	SOE-PCR
gCHAP_L24-L-R	GTAGATGCAGAAGAAGTTTT <u>ATGGATGAATGCATAGC</u>	for linker24
gCHAP_L25-L-F	<u>GCTATGCATTCATCCATGGTAAAACCTTCTTCTGCATC</u>	SOE-PCR
gCHAP_L25-L-R	GATGCAGAAGAAGTTTT <u>ACCATGGATGAATGCATAGC</u>	for linker25
gCHAP_L26-L-F	<u>GCTATGCATTCATCCATGACGGTAAAACCTTCTTCTGC</u>	SOE-PCR
gCHAP_L26-L-R	GCAGAAGAAGTTTT <u>ACCGTCATGGATGAATGCATAGC</u>	for linker26
gCHAP_L27-L-F	<u>GCTATGCATTCATCCATAAAGACGGTAAAACCTTCTTC</u>	SOE-PCR
gCHAP_L27-L-R	GAAGAAGTTTT <u>ACCGTCTTTATGGATGAATGCATAGC</u>	for linker27
gCHAP_L28-L-F	<u>GCTATGCATTCATCCATGTTAAAGACGGTAAAACCTTC</u>	SOE-PCR
gCHAP_L28-L-R	GAAGTTTT <u>ACCGTCTTTAACATGGATGAATGCATAGC</u>	for linker28
gCHAP_L31-L-F	<u>GCTATGCATTCATCCATTCTACTGTCGTAAAGAC</u>	SOE-PCR
gCHAP_L31-L-R	GTCTTTAACGACAGTAGA <u>ATGGATGAATGCATAGC</u>	for linker31
gCHAP_L34-L-F	<u>GCTATGCATTCATCCATACTTCTTCTTCTACTGTCG</u>	SOE-PCR
gCHAP_L34-L-R	CGACAGTAGAAGAAGAAGT <u>ATGGATGAATGCATAGC</u>	for linker34
gCHAP_L35-L-F	<u>GCTATGCATTCATCCATGGTACTTCTTCTTCTACTG</u>	SOE-PCR
gCHAP_L35-L-R	CAGTAGAAGAAGAAGT <u>ACCATGGATGAATGCATAGC</u>	for linker35
gCHAP_L38-L-F	<u>GCTATGCATTCATCCATATGGACAAGGGTACTTCTTC</u>	SOE-PCR
gCHAP_L38-L-R	GAAGAAGTACCCTTGCCAT <u>ATGGATGAATGCATAGC</u>	for linker38

*Underlined letters indicate the sequence of gCHAP fragments.

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Table S4. PCR primers used for amplification 8 SH3 domain linker

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Primer name	Sequence (5' -3')	Usage
ALS2-dA-ClyC _{SH3} -L-F	<u>GAAAAACCAGTACGGTACT</u> TGGTACAAACCGGAATCG	SOE-PCR for
ALS2-dA-ClyC _{SH3} -L-R	CGATTCCGGTTTGTACCA <u>AGTACCGTACTGGTTTTTC</u>	ALS2-dA-ClyC _{SH3}
ALS2-dA-vB _{SH3} -L-F	<u>GAAAAACCAGTACGGTACT</u> TGGTACAAACCGGAAAAACG	SOE-PCR for
ALS2-dA-vB _{SH3} -L-R	CGTTTTCCGGTTTGTACCA <u>AGTACCGTACTGGTTTTTC</u>	ALS2-dA-vB _{SH3}
ALS2-dA-tras14 _{SH3} -L-F	<u>GAAAAACCAGTACGGTACT</u> TACTACATGGAAGAAAGC	SOE-PCR for
ALS2-dA-tras14 _{SH3} -L-R	GCTTTCTTCCATGTAGTA <u>AGTACCGTACTGGTTTTTC</u>	ALS2-dA-tras14 _{SH3}
ALS2-dA-ClyO _{SH3} -L-F	<u>GAAAAACCAGTACGGTACT</u> CGTTCCTATCGCGAGAC	SOE-PCR for
ALS2-dA-ClyO _{SH3} -L-R	GTCTCGCGATAGGAACGAGTACCGTACTGGTTTTTC	ALS2-dA-ClyO _{SH3}
ALS2-dA-Lsp _{SH3} -L-F	<u>GAAAAACCAGTACGGTACT</u> CTGTATAAATCTGAATCTG	SOE-PCR for
ALS2-dA-Lsp _{SH3} -L-R	CAGATTCAGATTTATACAGAGTACCGTACTGGTTTTTC	ALS2-dA-Lsp _{SH3}
ALS2-dA-2638A _{SH3} -L-F	<u>GAAAAACCAGTACGGTACT</u> TGGTATAAAGCTGAACATG	SOE-PCR for
ALS2-dA-2638A _{SH3} -L-R	CATG TTCAGCTTTATACCA <u>AGTACCGTACTGGTTTTTC</u>	ALS2-dA-2638A _{SH3}
ALS2-dA-2958 _{SH3} -L-F	<u>GAAAAACCAGTACGGTACT</u> GGAAATTATACCGTCGC	SOE-PCR for
ALS2-dA-2958 _{SH3} -L-R	GCGACGGTATAATTCC <u>AGTACCGTACTGGTTTTTC</u>	ALS2-dA-2958 _{SH3}

*Underlined letters indicate the sequence of gCHAP fragments.

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Table S5. PCR primers used for amplification of pET101/D/TOPO vector

Primer name	Sequence (5' -3')	Usage
gCHAP_CACC-F	caccATGGCATCATCTTTTAATC	ALS2-dA-L25 amplification
K _{SH3} -R	TTTGAAAACACCCCATGCAAC	ALS2-dA-L25 amplification
ClyC _{SH3} -R	CTTGAATGTTCCCCAGGCAAC	ALS2-dA-ClyC _{SH3} amplification

*small case letters indicate nucleotides added artificially.

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Table S6. PCR primers used for amplification of various chimeric lysins

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Primer name	Sequence (5' -3')	Usage
P_Lsp-L-F	CTTTAAGAAGGAGATATACATatgGCGGCAACGCATGAAC	SOE-PCR for Lysostaphin
P_Lsp-L-R	G TTCATGCGTTGCCGCcatATGTATATCTCCTTCTTAAAG	
Lsp_T-L-F	CTGTGGGGTACTATCAAAAAGGGTCATCATCACCATC	SOE-PCR for Lysostaphin & Lsp_ClyC _{SH3}
Lsp_T-L-R	GATGGTGATGATGACCCTTTTTGATAGTACCCACAG	
P_ClyC-L-F	CTTTAAGAAGGAGATATACATATGCAAGCAAACTAAC	SOE-PCR for ClyC
P_ClyC-L-R	GTTAGTTTTGCTTGCATATGTATATCTCCTTCTTAAAG	
ClyC_T-L-F	CCTGGGGAACATTCAAGAAGGGTCATCATCACCATC	
ClyC_T-L-R	GATGGTGATGATGACCCTTCTTGAATGTTCCCAAG	
P_ClyO-L-F	CTTTAAGAAGGAGATATACATATGGCACTGCCTAAAACGG	SOE-PCR for ClyO
P_ClyO-L-R	CCGTTTTAGGCAGTGCCATATGTATATCTCCTTCTTAAAG	
ClyO_T-L-F	CTTGGGGTACATTTAAAAAGGGTCATCATCACCATC	
ClyO_T-L-R	GATGGTGATGATGACCCTTTTTAAATGTACCCCAAG	
Lsp_ClyC-L-F	GAAAACAAACAATATGGTACTTGGTACAAACCGGAATCG	SOE-PCR for Lsp_ClyC _{SH3}
Lsp_ClyC-L-R	CGATTCCGGTTTGTACCAAGTACCATATTTGTTAGTTTC	

*small case letters indicate nucleotides added artificially.

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Table S7. Analyses of genetic traits related to expression levels of chimeric lysins.

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Chimeric lysin	RI	Frequency of amino acid (%)		Translation stalling codon pair ^c	Free energy ^d (kcal/mol)
		Low expression-related amino acid ^a /codon ^b	high expression-related amino acid		
ALS2-dA-L21	1.00	52.3 (11/21)/ 14.2 (3/21)	52.3% (11/21)	PP, PG	-14.4/-213.2
ALS2-dA-L24	0.35	50.0 (12/24)/12.5 (3/24)	50.0% (12/24)	PP, PG	-16.3/-213.6
ALS2-dA-L25	0.95	52.0 (13/25)/16.0 (4/25)	52.0% (13/25)	PP, PG	-16.5/-219.2
ALS2-dA-L26	0.93	50.0 (13/26)/15.3 (4/26)	50.0% (13/26)	PP, PG	-15.1/-214.8
ALS2-dA-L27	0.52	48.1 (13/27)/14.8 (4/27)	48.1% (13/27)	PP, PG	-15.2/-213.9
ALS2-dA-L28	0.76	46.4 (13/28)/14.2 (4/28)	46.4% (13/28)	PP, PG	-16.3/-216.0
ALS2-dA-L31	0.46	45.1 (14/31)/12.9 (4/31)	45.1% (14/31)	PP, PG	-23.4/-222.7
ALS2-dA-L34	0.23	47.0 (16/34)/11.7 (4/34)	47.0% (16/34)	PP, PG	-25.6/-221.7
ALS2-dA-L35	0.57	48.5 (17/35)/14.2 (5/35)	48.5% (17/35)	PP, PG	-28.5/-227.0
ALS2-dA-L38	0.33	44.7 (17/38)/13.1 (5/38)	44.7% (17/38)	PP, PG	-29.6/-227.8
ALS2-dA-L25	0.32	54.0 (47/87) 19.5 (17/87)	11.5% (10/87)	PP, PG	-85.7/-219.2
ALS2-dA-ClyC _{SH3}	0.40	54.0 (47/87) 16.0 (14/87)	11.5% (10/87)	-	-66.0/-203.4
ALS2-dA-vB _{SH3}	0.36	55.1 (48/87) 18.3 (16/87)	11.5% (10/87)	-	-64.3/-197.8
ALS2-dA-tras14 _{SH3}	0.57	48.2 (42/87) 16.0 (14/87)	18.4% (16/87)	-	-76.9/-210.1
ALS2-dA-ClyO _{SH3}	0.64	44.0 (37/84) 8.3 (7/84)	15.4% (13/84)	-	-81.7/-208.3
ALS2-dA-Lsp _{SH3}	0.91	46.4 (39/84) 15.4 (13/84)	19.0% (16/84)	-	-59.6/-187.5
ALS2-dA-2638A _{SH3}	1.00	40.6 (35/86) 9.3 (8/86)	30.2% (26/86)	-	-54.4/-187.3
ALS2-dA-2958 _{SH3}	0.09	50.0 (40/80) 12.5 (10/80)	10.0% (8/80)	-	-56.1/-184.9
ALS2-dA-L25	0.29	50.6% (115/227)/16.7% (38/227)	13.2% (30/227)	PP, PG SH3	-1.3/-219.2
Lysostaphin	1.00	46.9% (116/247)/15.4% (38/247)	19.0% (47/247)	-	-4.6/-214.4
Lsp-ClyC _{SH3}	0.10	49.6% (124/250)/15.6% (39/250)	16.4% (41/250)	-	-4.6/-224.8
ClyC	0.17	47.6% (134/281)/11.7% (33/281)	19.9% (56/281)	-	-3.6/-241.8
ClyO	2.08	46.9% (121/258)/13.2% (34/258)	15.9% (41/258)	-	-10.0/-245.5

^a The frequency of L, S, R, A, P, G, C, W, and I in linker, SH3 domain, insert gene and initial~16codon. 39^b The frequency of AGG (R), CGU (R), CGA (R), AGU (S), GGU (G), AGA (R), UGU (C), CGG (R), CCC (P), AUA (I) in linker, SH3 domain and complete coding region. 40^c Ribosome stalling codon pairs: RR (CGG-AGG, AGG-CGG, CGA-CGG, CGG-AGG), PP (CCA-CCC, CCG-CCA, CCT-CCG, CCG-CCG, CCT-CCT, CCA-CCG), KR (AAG-AGG), LR (CTG-AGG), PG (CCG-GGT, CCG-GGG), VG (GTG-GGA), and QD (CAG-GAT). 41^d Predicted free energy of mRNA secondary structure of chimeric lysin by RNA fold. Partial regions: linker, SH3 domain, and first 16-dodons. 42

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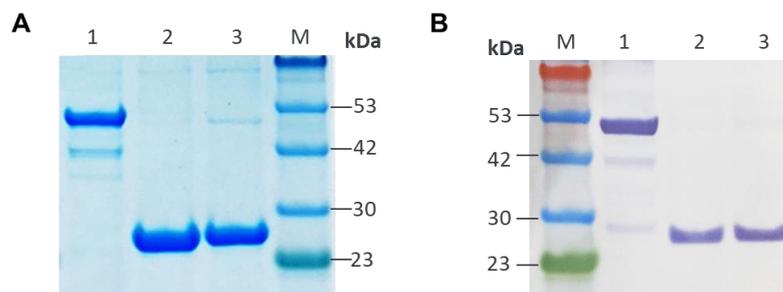


Figure S1. SDS-PAGE and Western blotting of *E. coli* expressed chimeric lysins. **(A)** SDS-PAGE analysis of purified protein using 4-12% polyacrylamide gel. Lane M: protein molecular weight marker; Lane 1: purified protein of ALS2; Lane 2: purified protein of ALS2-dA-L25; Lane 3: ALS2-dA-ClyC_{SH3}. For SDS-PAGE analysis, 5 μ L of each purified protein, **(B)** Western blot analysis of expressed protein by six histidine antibody. The molecular weight of the protein inserted into the pET/D/101 vector is approximately 52.4kDa and 28.31kDa. Lane M: protein molecular weight marker (PM2700 ExcelBand™ 3-color Broad Range Protein Marker); Lane 1: Protein expressed ALS2; Lane 2: Protein expressed ALS2-dA-L25; Lane 3: Protein expressed ALS2-dA-ClyC_{SH3}. For western blot analysis, 20 μ L of each induced protein.

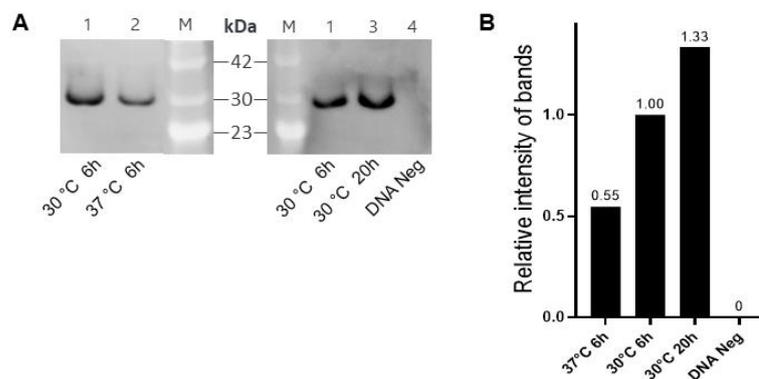


Figure S2. Comparison of expression levels of Lsp-ClyC_{SH3} in different temperature and incubation times. **(A)** Western blotting of Lsp-ClyC_{SH3} using anti-histidine antibody and ECL. Lane M: protein molecular weight marker; Lane 1: Expressed at 30°C for 6h; Lane 2: Expressed at 37°C for 6h; Lane 3: Expressed at 30°C for 20h; Lane 4: DNA negative sample. **(B)** The relative intensity of the Lsp-ClyC_{SH3} protein bands. The ratio of protein expression compared to Lsp-ClyC_{SH3} expressed at 30°C for 6h was measured using Image J 1.53