

Article

Not peer-reviewed version

# Investigation of Molecular Compounds in Kumquat (*Fortunella* spp.) Peel via Raman Spectroscopy

Humeyra Yildirim, Isil Tulum, Ayse Erol, Fahrettin Sarcan

Posted Date: 19 January 2024

doi: 10.20944/preprints202401.1477.v1

Keywords: Kumquat; citrus peels; functional components; Raman spectroscopy; spectral mapping; carotenoid



Preprints.org is a free multidiscipline platform providing preprint service that is dedicated to making early versions of research outputs permanently available and citable. Preprints posted at Preprints.org appear in Web of Science, Crossref, Google Scholar, Scilit, Europe PMC.

Copyright: This is an open access article distributed under the Creative Commons Attribution License which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Disclaimer/Publisher's Note: The statements, opinions, and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions, or products referred to in the content.

Article

# Investigation of Molecular Compounds in Kumquat (Fortunella Spp.) Peel via Raman Spectroscopy

Hümeyra Yıldırım 1,2, Isil Tulum 3, Ayse Erol 1 and Fahrettin Sarcan 1,\*

- <sup>1</sup> Department of Physics, Faculty of Science, Istanbul University, Vezneciler, Istanbul, 34134, Türkiye; hümeyra.yildirim@ogr.iu.edu.tr (H.Y.); ayseerol@istanbul.edu.tr (A.E.)
- <sup>2</sup> TEBIP High Performers Program, Faculty of Science, Istanbul University, Vezneciler, Istanbul, 34134, Türkiye
- <sup>3</sup> Istanbul University, Faculty of Science, Department of Biology, Istanbul, Türkiye; iciltlm@gmail.com
- \* Correspondence: fahrettin.sarcan@istanbul.edu.tr

**Abstract:** In this study, Raman spectroscopy is employed to investigate the chemical compounds of kumquat peels and compared that of orange. Kumquat is a citrus fruit belonging to the genus Fortunella in family Rutaceae. It is observed that kumquat peel has 3 main Raman active characteristic vibration modes specified to carotenoids, which is dominated by  $\beta$ -carotene at 1007, 1158 and 1526 cm<sup>-1</sup>, as it is well known for orange. The carotenoid distribution within the cross-section of kumquat is also investigated. The carotenoid-related Raman vibration modes are relatively stronger on the peel of kumquat, which is an important finding especially for the fruits that can be eaten with its peel. Our results pave the way to take an attention for the importance of kumquat as being a fruit that can be grown up in different climates compared to orange, which grow in warm climates.

**Keywords:** kumquat; citrus peels; functional components; Raman spectroscopy; spectral mapping; carotenoid

# 1. Introduction

Kumquat is a citrus fruit belonging to the genus Fortunella of the Rutaceae family. It is also known as golden orange in Türkiye. Up to date, Kumquat is classified into four main types: Fortunella japonica, Fortunella margarita, Fortunella crassifolia and Fortunella hindsii (Lou and Ho 2017). Kumquat fruits exhibit an orange-yellow hue, an elliptical shape and measuring approximately 2 cm in diameter, and it is regarded as one of the smallest citrus fruits (Chen et al., 2017). Kumquats have numerous advantages, such as cold resistance, an eatable peel, small size, and adaptability to various soil types (Love et al., 2007; Palma et al., 2018). These advantageous make them easy to care for and ideal for smaller spaces, offering a versatile citrus option suitable for diverse environments, ranging from greenhouses to terraces, and as a cold-tolerant citrus fruit (Palma et al., 2018). Compared to other citrus fruits, kumquat can be consumed whole with its peel, which provides an advantage in the intake of bioactive substances. While the pulp part of the fruit is sour, the peel part has a characteristic aroma due to flavonoids and terpenoids, which are important in the essential oil composition (Wang et al., 2012; Yıldız Turgut et al., 2015). Because of its acidic taste and soft peel, kumquat is used in products such as jams, pickles and sauces, in addition to its natural consumption (Choi et al., 2005; Pawełczyk et al., 2021). Besides, considering its use in the food industry, kumquat contains a variety of phytochemicals, including carotenoids, essential oils, ascorbic acid and flavonoids. These components are small molecules that are not essential for the survival of plants but represent pharmacological activity (Ogawa et al., 2001). Kumquat is becoming increasingly important in traditional medicine because it contains many beneficial phytochemicals with diverse biological effects (Wang et al., 2012; Lou et al., 2016). Phytochemicals are known to have beneficial biological effects. These include antibacterial, anti-oxidative, anti-inflammatory, anti-cancer, as well as cardiovascular protective effects (Li X. et al., 2022; Xinmiao Lv et al., 2015; Al-Saman et al., 2019). For example, carotenoids, an important component in citrus peel, have the ability to detoxify free radicals in cells. An important feature of carotenoids is that they are precursors of vitamin A. The body can convert certain carotenoids into active vitamin A (Grune et al., 2010). Carotenoids are used as nutraceuticals in various diseases such as eye diseases, cardiovascular diseases, neurodegenerative diseases and cancer (Saini *et al.*, 2022). It is crucial to find and analyse bioactive compounds in citrus peels. Many methods such as Gas Chromatography-Mass Spectrometry, High-Performance Liquid Chromatography, Fourier Transform Infrared Spectroscopy are used for biocomponent analysis of citrus peels (Yu et al., 2022; Chen J. *et al.*, 2022; Niluxsshun MCD *et al.*, 2021).

These methods for component analysis have the advantages of high sensitivity and accuracy, but also disadvantages such as complexity and time-consuming for sample preparation (Yang *et al.*, 2017).

In this study Raman spectroscopy, which is a non-destructive method, is employed to determined and analysis the compounds of kumquat peel. Raman spectroscopy is an analytical technique where inelastic scattered photons is used to measure the vibrational energy modes of a molecules. When photons interact with a substance, the frequency of most of the scattered light does not change, which is called Rayleigh scattering. However, inelastic light scattering processes can also occur due to molecular vibrations, so-called Raman scattering (Sarcan et al., 2014; Sarcan et al., 2023; Serebrennikova et al., 2021). The spectrum of scattered photons in Raman spectroscopy is fingerprint of the investigated material therefore allows easy identification of the molecule of interest. Raman spectroscopy has many advantages such as ease of sample preparation, non-destructive and the ability to work with aqueous samples. Because of these advantages, Raman spectroscopy has become a powerful alternative tool to other commonly used techniques. Accordingly, Raman spectroscopy has been used as a promising analytical tool in recent years as it provides a chemical fingerprint for molecular identification (Dodo et al., 2022; Serebrennikova et al., 2021). Raman spectroscopy is becoming increasingly popular in research on food, environment, medicine and many other fields. In various application areas such as pesticide detection (Chen et al., 2016), pathogen detection in food (Zhu et al., 2023), water pollution (Almaviva et al., 2022) and neurodegenerative disease diagnosis (Devitt *et al.*, 2018)

In 2017, Yang et al. used Raman spectroscopy for chemical mapping of functional compounds in citrus peels. The relative amount and distribution of essential oils, carotenoids and flavonoids in citrus peels at different locations (flavedo, albedo and longitudinal section) were studied (Yang et al., 2017). To the best our knowledge, there is no study that uses Raman spectroscopy for the determination of functional components on kumquat fruit.

In this study, carotenoids were determined in different locations (flavedo, albedo and cross-section) of kumquat fruit peel without any extraction process and a comparison was made between two kinds of citrus fruits (orange and kumquat). The fact that kumquat fruit can be consumed with its peel unlike other citrus fruits thanks to the terpenoids and flavonoids in the peel composition makes our research valuable. Kumquat is becoming increasingly important in food and pharmacology due to its nutritional and phytochemical content (Li X. et al., 2022). Therefore, a rapid and non-destructive determination of shell composition is very critical. The results obtained in our research are valuable in terms of advances in the use of Raman spectroscopy for the detection of biocomponents in citrus peels.

# 2. Materials and Methods

Sample Preparation

Kumquats and oranges from Verita (Verita, Istanbul, Türkiye) were soaked in saline for 10 min. The fruits were then washed three times with distilled water to remove the chemicals in the peel. Citrus peels were peeled. Pieces were cut for the Flavedo and Albedo parts. Cross-sectional pieces were also taken to understand the component distribution. Samples were adhered to the slide with double-sided tape and measured.

2

3

Raman spectroscopy measurements was carried out using a free space custom modular microspectroscopy set-up equipped a thermoelectric cooled CCD (Newton BEX2-DD, Andor) with a 1800 grooves/mm of grating in a spectrometer (Shamrock 500i, Andor). To excite the samples, a 532 nm CW laser (Gem532, Novanta Photonics) was used, and the excitation laser beam is focused to a spot of  $\sim$ 1.2  $\mu$ m in diameter and on the devices placed on a XYZ sample stage. Raman spectra from the samples were collected via a 50x objective (NA = 0.42).

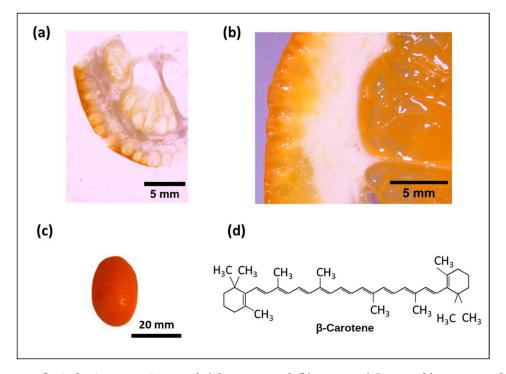
#### 3. Results and Discussion

#### Results

The flavedo and albedo structures of kumquat and orange fruits were observed by optical microscope (Figure 2). It was noted that the flavedo layer of the kumquat ( $N\sim1,5$  mm) was 2.5 times thinner than that of the orange ( $N\sim4$  mm).

The thinner flavedo part makes the kumquat fruit eatable with its peel. In addition to that the thinner flavedo layer of kumquat compared to orange is important for obtaining a higher concentration of beneficial components from a smaller part of the fruit.

Figure 3 shows the Raman spectra of kumquat and orange peel. The strong Raman peaks were observed at 1007, 1158 and 1526 cm<sup>-1</sup> as well as the relatively weak peaks were observed at 872, 963, 1188, 1212 and 1263 cm<sup>-1</sup> (Figure 3). Peaks at 1007, 1158 and 1526 cm<sup>-1</sup> are assigned to carotenoids, a vitamin A precursor according to previous literature data (Yang *et al.*, 2017).  $\beta$ -carotene containing 9 conjugated double bonds (Figure 1d) has been detected by Raman spectroscopy in foods such as tomatoes, carrots and pumpkin in previous studies. The bands attributed to carotenoids in the studies are shown in the Table 1.



**Figure 1.** Optical microscope image of **a**) kumquat and **(b)** orange. **c**) Image of kumquat and **d**) Chemical structures of *β* -*carotene*.

4

 $\label{prop:cond} \textbf{Figure 2. Cross-sectional optical microscope images of a) orange and \textbf{b)} \ kumquat.$ 

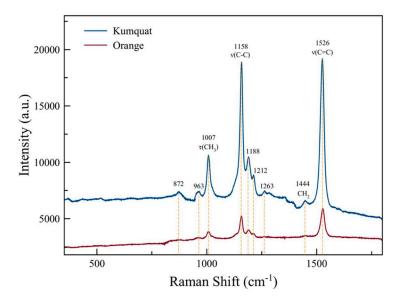


Figure 3. Raman Spectra of Orange and Kumquat.

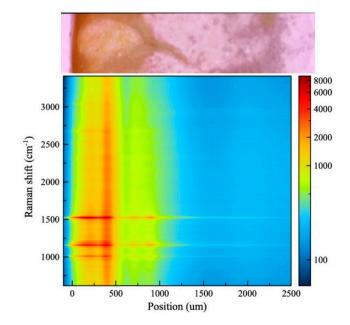


Figure 4. Raman spectrum mapping of transverse section of kumquat peel.

**Table 1.** Wavenumbers of  $v_1$ ,  $v_2$  and  $v_3$  modes (cm<sup>-1</sup>) of the predominant carotenoids obtained from several products by Raman spectroscopy.

Sample	ν <sub>1</sub> (C=C)	v <sub>2</sub> (C–C)	ν <sub>3</sub> (C–CH <sub>3</sub> )	
Citrus	1528	1156	1010	Yang et al. 2017
Tomato	1510	1156	1005	Baranska et al. 2006
Carrot	1520	1156	1007	Schulz et al. 2005
Pumpkin	1527	1157	1008	Oliveira et al. 2009
Kumquat	1526	1158	1007	This work

Raman spectrum of carrot showed strong bands at 1520, 1156 and 1007 cm<sup>-1</sup> assigned to  $\nu$ (C=C),  $\nu$ (C-C) and  $\tau$ (CH<sub>3</sub>) of  $\beta$  -carotene , respectively; in the Raman spectrum of tomato puree,  $\beta$  -carotene was observed with three intense bands at 1510 ( $\nu$ <sub>1</sub>), 1156 ( $\nu$ <sub>2</sub>) and 1005 cm<sup>-1</sup> ( $\nu$ <sub>3</sub>) (Schulz *et al.* 2005; Baranska *et al.* 2006).

According to the reported values in the literature, the results obtained confirmed the characteristic Raman bands at 1526 cm<sup>-1</sup> and 1158 cm<sup>-1</sup> assigned to the in-phase C=C and C-C stretching vibrations of the polyene chain and 1007 cm<sup>-1</sup> assigned to the in-plane rocking modes of CH<sub>3</sub> groups attached to the polyene chain joined by C-C bonds (De Gelder *et al.* 2007; Park *et al.* 2023; Yang *et al.* 2017). In addition, the peak at 1444 cm<sup>-1</sup> is assigned to the CH<sub>3</sub>-CH<sub>2</sub> bending modes indicates essential oil (Schulz *et al.* 2002; Vargas Jentzsch *et al.* 2014).

It is observed that the Raman peak intensities of both carotenoid and essential oils are 6 times stronger on the kumquat peel compared to that of orange. This makes kumquat a strong alternative for vitamin A intake beyond its advantageous properties such as cold resistance, compatibility with various soil types and edibility with its peel compared to other citrus fruits.

In the cross-sectional piece taken from kumquat fruit, the carotenoid content was investigated from the albedo layer to the flavedo layer. Looking at the distribution of carotenoids, it was found that the carotenoid density decreased when going from the albedo layer to the flavedo layer (Figure 4).

### 4. Conclusion

In conclusion, we have used Raman spectroscopy for a rapid and non-destructive detection of carotenoids in kumquat and compared that of orange peels. Raman spectrum analysis of kumquat peel have revealed distinct characteristic bands at 1007, 1158 and 1526 cm $^{-1}$  associated with carotenoids, particularly dominated by  $\beta$ -carotene. This result indicates the potential of kumquat to be a valuable alternative among citrus fruits for obtaining vitamin A. It has been observed that the concentration of carotenoids is more pronounced in the flavedo part compared to the albedo part. The findings of this study address the differences in the biochemical profiles of citrus fruit peels and highlight the potential health benefits associated with kumquat peel consumption. The information obtained from our study will contribute to the quantitative analysis of the materials in the fruits. Furthermore, the successful application of Raman spectroscopy as a robust analytical tool for non-destructive compositional analysis in the field of food science and nutrition research.

**Acknowledgments:** This work was supported in part by the Scientific Research Projects Coordination Unit of Istanbul University (FBG-2022-38573, FBG-2021-37896).

Conflicts of Interest: The authors declare no competing interests.

# References

Almaviva, S., Artuso, F., Giardina, I., Lai, A., & Pasquo, A. (2022). Fast Detection of Different Water Contaminants by Raman Spectroscopy and Surface-Enhanced Raman Spectroscopy. Sensors (Basel, Switzerland), 22(21), 8338. https://doi.org/10.3390/s22218338

Al-SamanM.A.Abdella,A., Mazrou, K.E. et al. Antimicrobial and antioxidant activities of different extracts of the peel of kumquat (Citrus japonica Thunb). Food Measure 13, 3221–3229 (2019). https://doi.org/10.1007/s11694-019-00244-y

- Baranska M, Schütze W, Schulz H. Determination of lycopene and beta-carotene content in tomato fruits and related products: Comparison of FT-Raman, ATR-IR, and NIR spectroscopy. Anal Chem. 2006 Dec 15;78(24):8456-61. https://doi.org/10.1021/ac061220j. PMID: 17165839.
- Chen, J., Shi, Y., Zhong, Y., Sun, Z., Niu, J., Wang, Y., Chen, T., Chen, J., & Luan, M. (2022). Transcriptome Analysis and HPLC Profiling of Flavonoid Biosynthesis in Citrus aurantium L. during Its Key Developmental Stages. Biology, 11(7), 1078. https://doi.org/10.3390/biology11071078
- Chen MH, Yang KM, Huang TC, Wu ML. Traditional Small-Size Citrus from Taiwan: Essential Oils, Bioactive Compounds and Antioxidant Capacity. Medicines (Basel). 2017 May 8;4(2):28. https://doi.org/10.3390/medicines4020028. PMID: 28930243; PMCID: PMC5590064.
- Chen, J., Huang, Y., Kannan, P., Zhang, L., Lin, Z., Zhang, J., Chen, T., & Guo, L. (2016). Flexible and Adhesive Surface Enhance Raman Scattering Active Tape for Rapid Detection of Pesticide Residues in Fruits and Vegetables. Analytical chemistry, 88(4), 2149–2155. https://doi.org/10.1021/acs.analchem.5b03735
- Choi H. S. (2005). Characteristic odor components of kumquat (Fortunella japonica Swingle) peel oil. Journal of agricultural and food chemistry, 53(5), 1642–1647. https://doi.org/10.1021/jf040324x
- Devitt, G., Howard, K., Mudher, A., & Mahajan, S. (2018). Raman Spectroscopy: An Emerging Tool in Neurodegenerative Disease Research and Diagnosis. ACS chemical neuroscience, 9(3), 404–420.https://doi.org/10.1021/acschemneuro.7b00413
- De Gelder, J., De Gussem, K., Vandenabeele, P. and Moens, L. (2007), Reference database of Raman spectra of biological molecules. J. Raman Spectrosc., 38: 1133-1147. https://doi.org/10.1002/jrs.1734
- Dodo, K., Fujita, K., & Sodeoka, M. (2022). Raman Spectroscopy for Chemical Biology Research. Journal of the American Chemical Society, 144(43), 19651–19667. https://doi.org/10.1021/jacs.2c05359
- Grune, T., Lietz, G., Palou, A., Ross, A. C., Stahl, W., Tang, G., ... & Biesalski, H. K. (2010). B-carotene is an important vitamin a source for humans. The Journal of Nutrition, 140(12), 2268S-2285S. https://doi.org/10.3945/jn.109.119024
- Li, X., Meenu, M., & Xu, B. (2022). Recent development in bioactive compounds and health benefits of kumquat fruits. *Food Reviews International*, 1-21.
- Love K, Bowen R, Fleming K. 2007. Twelve fruits with potential value-added and culinary uses. University of Hawai'i College of Tropical Agriculture and Human Resources, Hawai.
- Lou SN, Ho CT. Phenolic compounds and biological activities of small-size citrus: Kumquat and calamondin. J Food Drug Anal. 2017 Jan;25(1):162-175. https://doi.org/10.1016/j.jfda.2016.10.024. Epub 2016 Dec 7. PMID: 28911534; PMCID: PMC9333435.
- Lou, S. N., Lai, Y. C., Hsu, Y. S., & Ho, C. T. (2016). Phenolic content, antioxidant activity and effective compounds of kumquat extracted by different solvents. Food chemistry, 197(Pt A), 1–6. https://doi.org/10.1016/j.foodchem.2015.10.096
- Lv X, Zhao S, Ning Z, Zeng H, Shu Y, Tao O, Xiao C, Lu C, Liu Y. Citrus fruits as a treasure trove of active natural metabolites that potentially provide benefits for human health. Chem Cent J. 2015 Dec 24;9:68. https://doi.org/10.1186/s13065-015-0145-9. PMID: 26705419; PMCID: PMC4690266.
- Niluxsshun MCD, Masilamani K, Mathiventhan U. Green Synthesis of Silver Nanoparticles from the Extracts of Fruit Peel of Citrus tangerina, Citrus sinensis, and Citrus limon for Antibacterial Activities. Bioinorg Chem Appl. 2021 Feb 2;2021:6695734. https://doi.org/10.1155/2021/6695734. PMID: 33623527; PMCID: PMC7872778.
- Ogawa, K., Kawasaki, A., Omura, M., Yoshida, T., Ikoma, Y., & Yano, M. (2001). 3',5'-Di-C-beta-glucopyranosylphloretin, a flavonoid characteristic of the genus Fortunella. Phytochemistry, 57(5), 737–742. https://doi.org/10.1016/s0031-9422(01)00132-7
- Oliveira, V. E. d., Castro, H. V., Edwards, H. G. M., & Oliveira, L. F. C. d. (2009). Carotenes and carotenoids in natural biological samples: a raman spectroscopic analysis. Journal of Raman Spectroscopy, 41(6), 642-650. https://doi.org/10.1002/jrs.2493
- Palma, A. and D'Aquino, S. (2018). Kumquat— fortunella japonica. Exotic Fruits, 271-278. https://doi.org/10.1016/b978-0-12-803138-4.00035-6
- Park, M., Somborn, A., Schlehuber, D., Keuter, V., & Deerberg, G. (2023). Raman spectroscopy in crop quality assessment: focusing on sensing secondary metabolites: a review. Horticulture Research, 10.
- Pawełczyk, A., Żwawiak, J., & Zaprutko, L. (2021). Kumquat fruits as an important source of food ingredients and utility compounds. Food Reviews International, 39(2), 875-895. https://doi.org/10.1080/87559129.2021.1928179
- Saini RK, Prasad P, Lokesh V, Shang X, Shin J, Keum YS, Lee JH. Carotenoids: Dietary Sources, Extraction, Encapsulation, Bioavailability, and Health Benefits-A Review of Recent Advancements. Antioxidants (Basel). 2022 Apr 18;11(4):795. https://doi.org/10.3390/antiox11040795. PMID: 35453480; PMCID: PMC9025559.
- Sarcan, F., Donmez, O., Kara, K., Erol, A., Akalin, E., Arıkan, M. K., ... & Fontaine, C. (2014). Bismuth-induced effects on optical, lattice vibrational, and structural properties of bulk gaasbi alloys. Nanoscale Research Letters, 9(1). https://doi.org/10.1186/1556-276x-9-119

- Sarcan, F., Fairbairn, N. J., Zotev, P. G., Severs-Millard, T., Gillard, D. J., Wang, X., ... & Wang, Y. (2023). Understanding the impact of heavy ions and tailoring the optical properties of large-area monolayer ws2 using focused ion beam. NPJ 2D Materials and Applications, 7(1). https://doi.org/10.1038/s41699-023-00386-0
- Schulz, H., Barańska, M., & Barański, R. (2005). Potential of nir-ft-raman spectroscopy in natural carotenoid analysis. Biopolymers, 77(4), 212-221. https://doi.org/10.1002/bip.20215
- Schulz, H., Schrader, B.D., Quilitzsch, R., & Steuer, B. (2002). Quantitative Analysis of Various Citrus Oils by ATR/FT-IR and NIR-FT Raman Spectroscopy. Applied Spectroscopy, 56, 117 124.
- Serebrennikova KV, Berlina AN, Sotnikov DV, Zherdev AV, Dzantiev BB. Raman Scattering-Based Biosensing: New Prospects and Opportunities. Biosensors (Basel). 2021 Dec 13;11(12):512. https://doi.org/10.3390/bios11120512. PMID: 34940269; PMCID: PMC8699498.
- Vargas Jentzsch, Paul & Ciobotă, Valerian. (2014). Raman spectroscopy as an analytical tool for analysis of vegetable and essential oils. Flavour and Fragrance Journal. 29. 10.1002/ffj.3203.
- Wang YW, Zeng WC, Xu PY, Lan YJ, Zhu RX, Zhong K, Huang YN, Gao H. Chemical composition and antimicrobial activity of the essential oil of kumquat (Fortunella crassifolia Swingle) peel. Int J Mol Sci. 2012;13(3):3382-3393. https://doi.org/10.3390/ijms13033382. Epub 2012 Mar 12. PMID: 22489157; PMCID: PMC3317718.
- Yang, Y., Wang, X., Zhao, C., Tian, G., Zhang, H., Xiao, H., He, L., & Zheng, J. (2017). Chemical Mapping of Essential Oils, Flavonoids and Carotenoids in Citrus Peels by Raman Microscopy. Journal of food science, 82(12), 2840–2846. https://doi.org/10.1111/1750-3841.13952
- Yıldız Turgut, D., M. Gölükcü ve H. Tokgöz. 2015. Kamkat (Fortunella margarita Swing.) meyvesi ve reçelinin bazı fiziksel ve kimyasal özellikleri. Derim, 32 (1): 71-80.
- Yu, X., Chen, X., Li, Y., & Li, L. (2022). Effect of Drying Methods on Volatile Compounds of Citrus reticulata Ponkan and Chachi Peels as Characterized by GC-MS and GC-IMS. Foods (Basel, Switzerland), 11(17), 2662. https://doi.org/10.3390/foods11172662
- Zhu, A., Ali, S., Jiao, T., Wang, Z., Ouyang, Q., & Chen, Q. (2023). Advances in surface-enhanced Raman spectroscopy technology for detection of foodborne pathogens. *Comprehensive reviews in food science and food safety*, 22(3), 1466–1494. https://doi.org/10.1111/1541-4337.13118

**Disclaimer/Publisher's Note:** The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.