

Review

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Review

Identification of Alzheimer's Disease by Molecular Imaging: Progress and Prospects

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Abstract: Alzheimer's disease (AD), which is characterized by the development of β -amyloid plaques and neurofibrillary tangles in the brain parenchyma, can result in synaptic and neuronal loss. Clinical signs, including deteriorating memory deficiency, may lead to catastrophic effects for the elderly. Research on brain imaging and its application in the study of AD has advanced extraordinarily over the past 30 years. However, widely used imaging methods are only capable of identifying structural or metabolic alterations at the anatomical level in the brain and can only be utilized for drawing inferential distinctions. As a disease with a complex pathophysiological mechanism, AD has many different molecular pathological features and molecular biomarkers. Therefore, molecular imaging for AD pathology indicators provides a practical method for the early diagnosis and impartial assessment of AD. The present review discusses the pathogenesis of AD, as well as the detection methods and principles of molecular imaging, such as positron emission tomography (PET) and single-photon emission computed tomography (SPECT) techniques. Meanwhile, various tracers and probes applied in molecular imaging that detect A β deposits, tau protein accumulation, and neurotransmitters are also reviewed. Molecular pathological tracers provide the possibility to accurately quantify brain changes through molecular imaging.

Keywords: Alzheimer's disease; molecular imaging; pathogenesis; diagnosis; treatments

1. Introduction

As a leading contributor to dementia, Alzheimer's disease (AD) is a progressive neurodegenerative condition. The most common presentation of AD is with insidious, progressive problems centered around the episodic memory of elderly individuals, which may meet the requirements of mild cognitive impairment (MCI) at this stage. Challenges with navigation as well as multitasking and confidence issues frequently surface. A patient can be diagnosed with AD dementia when cognitive impairments develop, becoming more severe and pervasive, and begin to interfere with daily activities. As the condition progresses, increasing reliance is typical, and subsequent behavioral changes, decreased mobility, hallucinations, and seizures may appear. The average time to death after presentation is 8.5 years (1). According to the Alzheimer's Association, 60–80 percent of dementia cases globally are caused by AD. In the United States, it is estimated that

6.7 million people aged 65 or older will be diagnosed with Alzheimer's disease in 2023 (2). According to recent estimates, 44 million individuals worldwide currently have dementia. As the population ages, this is anticipated to more than triple by 2050, when the yearly cost of dementia in the USA alone may surpass USD 600 billion (3).

There is currently a lack of knowledge on the precise pathophysiological pathways underlying AD. From a neuropathological standpoint, AD is characterized by the following: (1) extracellular neuritic plaques, which are extracellular deposits of β -amyloid ($A\beta$); (2) neurofibrillary tangles (NTFs), which are neuronal aggregates of hyperphosphorylated tau; and (3) neuronal loss, particularly in the medial temporal lobe structures and the temporo-parietal association cortex (4). Along with aberrant protein deposition, various biochemical processes such as inflammation, oxidative damage, and lysosomal dysfunction occur, supporting the hypothesis that the etiological factors contributing to the disease process are heterogeneous and interact until the full disease pathway is established (5).

A diagnosis of dementia is typically determined by the patient's medical history, a pattern of cognitive deficits, and other factors evaluated through clinical investigations, such as blood testing and structural imaging of the brain, to rule out nondegenerative causes of the symptoms. The early-stage clinical diagnosis of AD is still challenging and requires a thorough medical history as well as a battery of neuropsychological tests to establish a diagnosis of dementia and to distinguish AD from other types like vascular dementia, frontotemporal dementia, and Lewy body dementia (6). The remaining functioning neurons in the AD brain cannot currently be directly measured in vivo; instead, postmortem pathological examination is used to determine the disease's definitive diagnosis (7). The amyloid cascade hypothesis states that amyloid production occurs 10 years or more before the start of clinical symptoms (8). Thus, investigating new techniques for the early and precise identification of AD in vivo is crucial for treatments to be effective.

Research on brain imaging and its application in the study of AD has advanced extraordinarily over the past 30 years (9). Currently, a number of imaging techniques have been utilized to clinically diagnose AD. Computed tomography (CT), nuclear magnetic resonance imaging (MRI), and different derivative technologies may be utilized, and these technologies can be classified in general terms. However, widely used imaging methods are only capable of identifying structural or metabolic alterations at the anatomical level in the brain, and can only be utilized for drawing inferential distinctions. As a disease with a complex pathophysiological mechanism, AD has many different molecular pathological features and molecular biomarkers. Therefore, molecular imaging for AD pathology indicators provides a practical method for the early diagnosis and impartial assessment of AD (10).

Functional neuroimaging using positron emission tomography (PET) and single-photon emission computed tomography (SPECT) has been used to study various metabolic and biochemical alterations in the brain in vivo, contributing to the early diagnosis and differential diagnosis of AD and providing essential data for the understanding of the underlying pathology. Different molecular radiotracers have been employed to estimate impairments in neurotransmitters, regional blood flow, and glucose metabolism, among other aspects of brain function (11). These molecular imaging approaches will be essential for accurate patient selection and the assessment of therapeutic response in light of new medicines. The imaging of aberrant protein deposits was made possible by recent developments in technology and radiopharmacology (12), while targeting new biochemical and molecular processes with the use of novel probes is a growing aspect of nuclear medicine imaging in the field of AD (13). Herein, we review the pathogenesis of AD, as well as the detection methods and principles of molecular imaging, such as positron emission tomography (PET) and single-photon emission computed tomography (SPECT) techniques. Meanwhile, various tracers and probes applied in molecular imaging that detect $A\beta$ deposits, tau protein accumulation, and neurotransmitters are also reviewed.

2. Pathogenesis of AD

The pathological phenomenon of AD is complex. Brain tissue examination results have shown that senile plaques (SPs) formed by β -amyloid ($A\beta$) plaques outside of brain nerve cells were common in patients with AD. Neurofibrillary tangles (NFTs), neuron loss, neurodystrophy, synaptic loss, and other pathological phenomena are formed by SPs and the abnormal phosphorylation of tau proteins [3]. However, the relevant pathogenesis is still unclear. The following section covers the pathogenesis of AD, which is summarized in Figure 1.

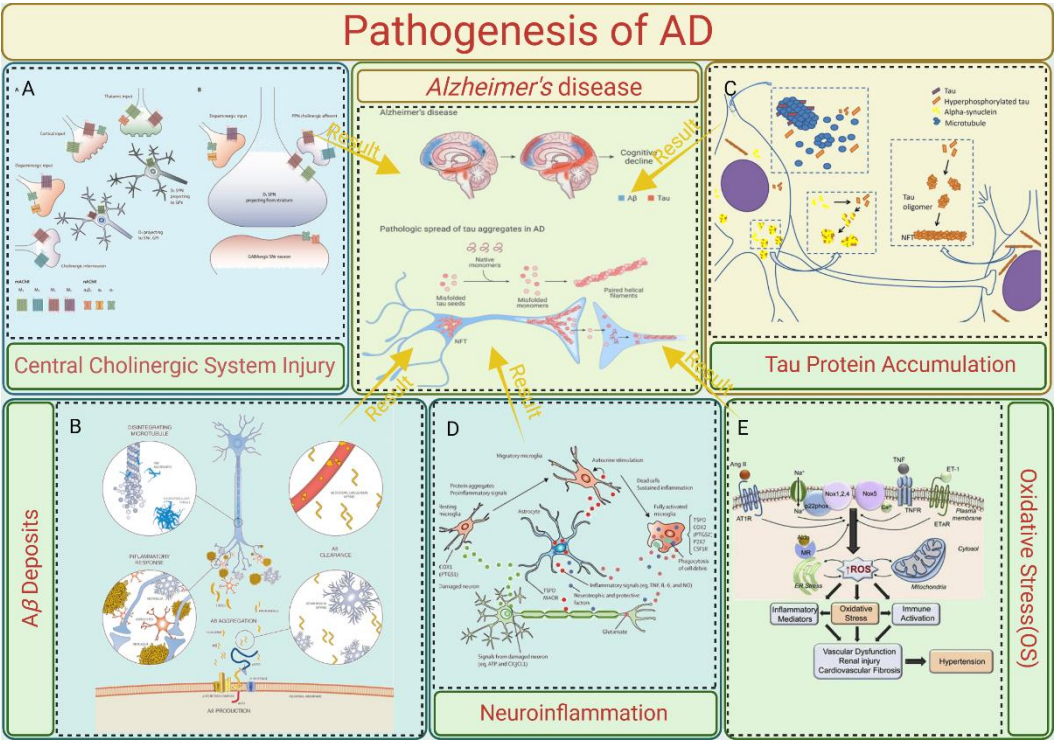


Figure 1. Summary of mechanisms for pathogenesis of AD. A. Central Cholinergic System Injury. B. $A\beta$ Deposits. C. Tau Protein Accumulation. D. Neuroinflammation. E. Oxidative Stress.

2.1. Central Cholinergic System Injury

The cholinergic system injury theory is the earliest theory about AD. Acetylcholine (ACh) is an important central excitatory neurotransmitter, which is related to a variety of advanced behaviors such as learning and memory. The central cholinergic nervous system affects the level of central acetylcholine by regulating the synthesis and release of acetylcholine (14). Early studies showed that severe neurodegeneration occurs in the basal nucleus of Meynert (mainly composed of cholinergic neurons), which is located in the basal forebrain, in patients with AD. At the same time, the presynaptic cholinergic transmitters in the cerebral cortex were severely depleted and the activity of choline acetyltransferase (ChAT) was significantly decreased (15). The above two points indicate that severe cholinergic system damage occurs in the brains of patients with AD.

Studies have shown that a cholinergic nerve imbalance can accelerate the deposition of $A\beta$, and acetylcholinesterase (AChE) can directly bind to presenilin-1 (PS-1), a key enzyme in the production of $A\beta$, to significantly enhance its expression, thus increasing the level of $A\beta$ and accelerating cognitive impairment (16). In addition, abnormal central cholinergic changes can also induce the abnormal phosphorylation of tau proteins, neuronal inflammation, apoptosis, and an imbalance in neurotransmitters and the neurohormone system (16).

2.2. $A\beta$ Deposits

$A\beta$ is a peptide produced by the cleavage of the amyloid precursor protein (APP), which is mediated by β -secretase and γ -secretase. It has 37043 amino acid residues (17). Toxic oligopeptides

generated by disrupting the assembly of APP fragments comprise 39–43 fragments, including protofibrils or fibrils. These then result in deposits that become apparent under a microscope. BACE 1 cleaves APP at β -sites Asp1 and Glu11. γ -secretase further cleaves the 99 amino acid residues that are linked to the C-terminal membrane, resulting in the production of the isoforms A β 1-42 and A β 1-40. Presenilin 1 (PS1) or presenilin 2 (PS2) are the primary forms of γ -secretase. The typical soluble isoform is A β 1-40, but if the cleavage pattern is altered, it could result in A β 1-42, which aggregates readily and forms plaque because it contains two extra amino acids: alanine and isoleucine (18). Mutations in the APP, the presenilin 1 and presenilin 2 genes, or the apolipoprotein E (APOE4) gene cause this shift in the cleavage pattern. In addition to genetic mutations, a multitude of neuropeptides may play a role in the development of plaque. For instance, low levels of somatostatin, neuropeptide Y, and corticotrophin-releasing hormone (CRH) may be linked to plaque formation, while higher levels of angiotensin II may contribute to irregular APP cleavage or poor removal of the A β 1-42 fragment (19). Different parts of the brain experience the formation and deposition of A β plaques. The brain recognizes these plaques as foreign material, which triggers an immunological and inflammatory response due to the triggering of microglia and releasing of cytokines. This ultimately results in cell death and dementia (20).

A total of 26 A β protein forms were identified from the cerebrospinal fluid of patients with AD, among which A β 1-40 and A β 1-42 with 40 and 42 amino acid residues were the main components of accumulated A β . A β 1mur42 is more hydrophobic and easier to gather (21), and it is believed to be the main culprit for the initiation of senile plaques in the brain. An increase in the level or proportion of A β 1mur42 can induce the formation of A β amyloid fibers, which develop into senile plaques in the brain and lead to neurotoxicity (22). The A β aggregate can activate cysteine protease, which can cleave tau proteins and change the conformation of normal tau proteins so that the tau proteins cannot bind to microtubules and aggregate, which induces the pathology of the tau proteins and leads to AD.

2.3. Tau Protein Accumulation

Microtubular neuronal proteins are called tau proteins. The microtubule binding domain of tau proteins is important in the stabilization and polymerization of the microtubule assembly, hence preserving the cytoskeleton's integrity. Numerous kinases, including the Fyn kinase, glycogen synthase kinase-3 β (GSK3 β), and cyclin-dependent kinase-5 (CDK5), phosphorylate the serine and/or threonine residues to control this interaction. CDK5 may be involved in the development of neurofibrillary tangles. Calpain is activated by A β , and p35, an activator of CDK5, is deregulated. Phosphorylated tau is hyperphosphorylated as a result of p35 splitting into p25 in response to an excess of cytosolic calcium. This hyperactivates CDK5 (23). The hyperphosphorylation of tau proteins causes a reduction in their affinity for microtubules. The hyperphosphorylated tau loses its ability to preserve the cell's structure when it produces NFTs and is deposited in the cytoplasm. Additionally, typical cellular functions like synaptic transmission, axonal transport, signal transduction, and progressive cell degeneration are impacted by this deposition.

The Tau protein is another element involved in the process of plaque formation. It is a factor that facilitates the tubulin protein's particular assembly process. In contrast, tubulin polymerizes and forms microtubules that form the intracellular pathway that the cell's motor proteins travel along or use as a dividing spindle during cell division. The hyperphosphorylation of the Tau protein results in the formation and deposition of neurofibrillary fibers in the pathophysiology of Alzheimer's disease (24). The loss of physiological function by a normal Tau protein, which results in the instability of microtubules, or the increase in function that is toxic to neurons, which causes apoptosis, are the two main mechanisms by which the Tau protein causes neurotoxicity (25). Numerous researchers have also demonstrated a link between the accumulation of β -amyloid and the Tau protein's aggregation, which is the last stage in the pathophysiology of illness (26).

2.4. Neuroinflammation

Microglia are the most important immune cells in the central nervous system that can recognize and remove damaged nerves, plaques, and infectious substances in the central nervous system, which is of great significance to maintain the homeostasis of the central nervous system. Microglia surface receptors can recognize damaged cells and heterogenous substances in the brain, cause microglia activation, produce a series of downstream effects, and play an immune role. However, the long-term excessive activation of microglia will release too many inflammatory factors and oxidizing substances, leading to an inflammatory reaction, nerve cell damage, and neurotoxicity (27). Bamberg et al. found that A β can bind and activate receptors on the surface of microglia, and activated microglia release cytokines, chemokines, reactive oxygen species, and other neurotoxic substances (such as NO, tumor necrosis factor, superoxide, etc.), leading to inflammation, which proves that neuroinflammation in the brain of patients with AD is closely related to A β -induced microglial activation (28).

A key factor in the pathophysiology of AD is neuroinflammation. Acute inflammation serves as a barrier to prevent brain damage caused by conditions like A β plaque buildup. The microglia's ability to release proinflammatory cytokines is maintained despite their inability to remove plaque due to chronic activation, which leads to an imbalance between pro- and anti-inflammatory cytokines. A β deposits trigger the activation of many Toll-like receptors (TLR2, TLR4, and TLR6) and their co-receptors, which include the microglia-expressed CD36, CD14, and CD47. The immune system produces proinflammatory cytokines of the IL-1 β family, such as IL-1 β and IL-18, upon the identification of microorganisms. Upon activation, caspase-1 or caspase-8 expresses these cytokines. The activation of caspase-1 is aided by inflammatory proteins such as the PYHIN (pyrin and HIN domain-containing) and NLR (Nod-like receptor) family. The main inflammasome that is capable of detecting Ab clumps is NLRP3. These proinflammatory cytokines can interfere with the microglial clearance of A β and damage dendritic spines. Nitric oxide (NO) generation is enhanced by the neuronal and glial cell expression of inducible isoforms of NO synthase in response to proinflammatory cytokines. As a result, the peptide's capacity to aggregate is enhanced, and its ability to inhibit synaptic plasticity is strengthened (29). As previously mentioned, these cytokines cause CDKs to become activated, which increases tau hyperphosphorylation and A β plaque formation (20).

2.5. Oxidative Stress (OS)

The functional imaging of the brains of patients with AD precursors revealed the hyperactivation of neurons in the hippocampus and neocortex regions of the brain (30). Mitochondria are the energy factories of cells, and abnormal neuron excitation indicates increased mitochondrial activity. Mitochondria are the main sites for the generation of intracellular reactive oxygen species (ROS). Under normal circumstances, ROS are produced in a small amount and can function as the second messenger in cells. When neurons are abnormally excited, mitochondrial activity is enhanced and ROS production increases, and the antioxidant defense system cannot clear them in time. Excessive ROS will damage mitochondria and then damage nerve cells, resulting in abnormal neuronal death and promotion of the pathological process of AD (31). Mitochondrial defects exist in the brain tissue of AD patients. Pathological phenomena of abnormal mitochondrial function have been found through detection utilizing brain tissue samples of AD patients, experiments using AD transgenic mice, and studies on the expression of mutant APP- or A β -treated cells (32).

Mitochondrial dysfunction may result from cytochrome c oxidase levels that are too low. Furthermore, OS-induced hyperexcitation of glycogen synthase kinase (GSK-3) can change the permeability of mitochondria. This might cause ROS to be produced in excess (33). ROS can be produced by metal ions, particularly copper and zinc, binding to the A β plaque. The resulting ROS oxidize the A β peptide, making it difficult to remove, and oxidize the cell membrane's lipid and protein membrane, making it permeable and therefore prone to degeneration (34).

3. Molecular Imaging Modalities in AD

At present, there are great differences in neuroimaging results and interpretations, which are not enough to meet the needs for the early detection, disease classification, treatment evaluation, and accurate prognosis of AD. Initial factors such as pathological changes at the cellular level occur much earlier than the appearance of clinical symptoms and changes in the anatomical structure. In view of the fact that the epidemiological characteristics of AD have not been elucidated and there is a lack of disease reversal drugs, the need for early detection of molecular level changes through molecular imaging is quite urgent. Several molecular imaging modalities for detecting AD are summed up in Figure 2.

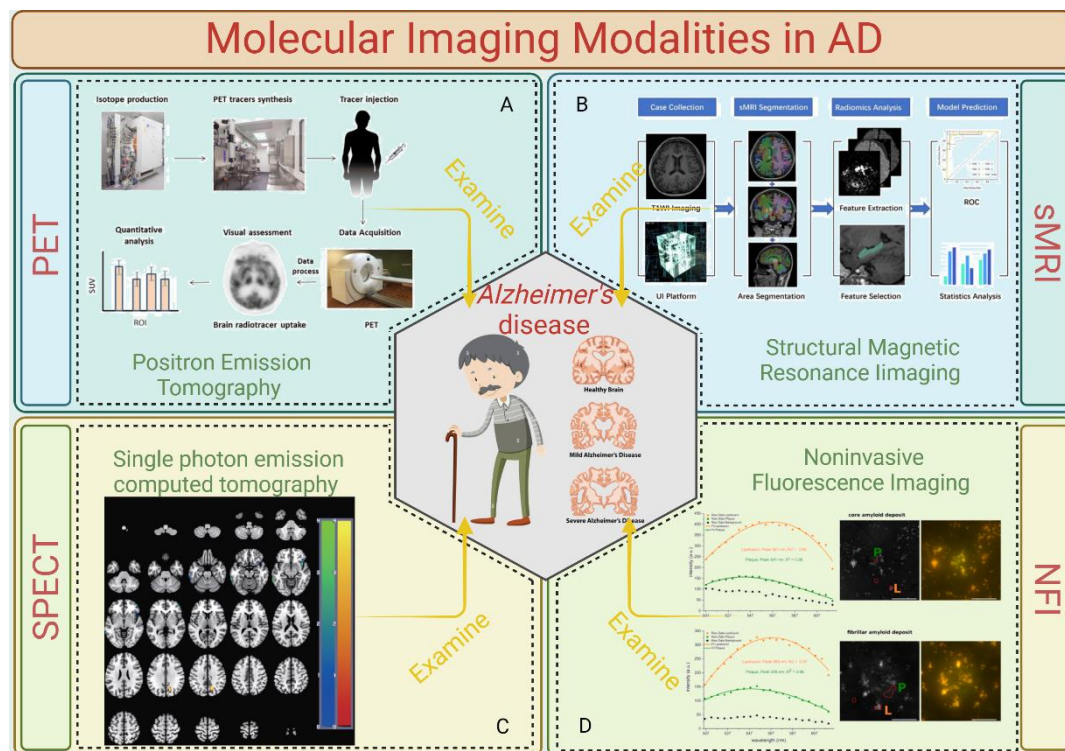


Figure 2. Molecular imaging modalities in AD. A. Reference to (35) Xie F, Peng F. Radiopharmaceuticals for Assessment of Altered Metabolism and Biometal Fluxes in Brain Aging and Alzheimer's Disease with Positron Emission Tomography. *J Alzheimers Dis.* 2017;59(2):527-536. doi:10.3233/JAD-170280. B. Reference to (36) Liu S, Jie C, Zheng W, Cui J, Wang Z. Investigation of Underlying Association Between Whole Brain Regions and Alzheimer's Disease: A Research Based on an Artificial Intelligence Model. *Front Aging Neurosci.* 2022;14:872530. Published 2022 Jun 7. doi:10.3389/fnagi.2022.872530. C. Reference to (37) Jeong HS, Park JS, Yang Y, Na SH, Chung YA, Song IU. Cerebral Perfusion Changes after Acetyl-L-Carnitine Treatment in Early Alzheimer's Disease Using Single Photon Emission Computed Tomography. *Dement Neurocogn Disord.* 2017 Mar;16(1):26-31. doi: 10.12779/dnd.2017.16.1.26. Epub 2017 Mar 31. PMID: 30906367; PMCID: PMC6427992. D. Reference to (38) Lochocki B, Boon BDC, Verheul SR, et al. Multimodal, label-free fluorescence and Raman imaging of amyloid deposits in snap-frozen Alzheimer's disease human brain tissue. *Commun Biol.* 2021;4(1):474. Published 2021 Apr 15. doi:10.1038/s42003-021-01981-x.

3.1. Positron Emission Tomography (PET)

Various radioactive tracers are used depending on the intended target. Positron emission tomography (PET) imaging analyzes changes in metabolism (39). PET imaging is the simultaneous detection of gamma rays emitted by positron annihilation events from a radiotracer. Since a radioisotope is utilized to replace one of the tracer's atoms, the radioactive tracers in this case are comparable to common biological compounds like glucose, peptides, and proteins (40). After being injected into the patient, a radioactive tracer enters the targeted organs or tissues through the

circulatory system and takes part in metabolic activities there (39). Due to their instability, the radioisotopes in the tracer decay. During this decay, positrons are released and collide with the electrons of nearby atoms, causing an annihilation reaction (41). Two 511 keV gamma rays, separated by around 180 degrees, are produced by the annihilation and are absorbed by scintillation crystals, where they are transformed into low-energy visible photons (41). The light impulses are subsequently translated into electrical signals by a photosensor. Scintillators, detectors, and readout electronics are combined to make a detector that measures three parameters: the energy of the gamma ray, its position upon impact, and the moment the ray strikes the detector (42). After that, these electronic signals are processed using reconstruction and correction techniques to create an image. The capacity of PET imaging to map various signals, such as brain metabolic activity, amyloid load, or tau-tracer retention, differs depending on the radiotracer used. PET scans of AD patient and non-patient groups can be compared as a diagnostic technique to observe variations in uptake patterns.

3.2. Structural Magnetic Resonance Imaging (sMRI)

The most widely utilized brain imaging technique in the research of AD has been structural MRI. Clinically affected patients have increased ventricular and sulcal volumes; reduced gray matter or cortical thickness in other cerebral regions, such as the precuneus and posterior cingulate, parietal, and temporal cortex; and accelerated rates of decline in these and whole brain measurements over time. They also have significantly reduced hippocampal and entorhinal cortex volumes, gray matter, and cortical thicknesses (8). Hippocampal and entorhinal cortex size reductions seem to be associated with early memory decline and can be used to predict the development of more severe clinical stages, such as Alzheimer's dementia and moderate cognitive impairment (MCI) (43). In the clinical setting, structural MRI is often recommended to help rule out potentially reversible conditions.

3.3. Single-photon emission computed tomography (SPECT)

PET and SPECT are quite comparable. When used to measure cerebral perfusion in dementia patients, SPECT produces results that are similar to those from the clinical diagnostic PET scanning of glucose metabolism. SPECT research has demonstrated the ability to forecast MCI deterioration (44) and has led to similar results as those from autopsies (45). For a long time, the scarcity of PET led to a significant reliance on SPECT scanning. Nevertheless, with the increasing number of PET scanners and their usage in clinical settings for oncological purposes, PET's availability and potential for use in dementia research and care have grown.

3.4. Noninvasive Fluorescence Imaging

By verifying the fluorescence of A β , tau, and other AD-associated proteins, optical fluorescence imaging has been established as the most effective technique for diagnosing AD pathology and neural networking. Endogenous and/or exogenous fluorophores that emit light in response to laser excitation are used in fluorescence imaging. In particular, because of low light scattering, low light absorption in surrounding tissues, and negligible autofluorescence, NIR fluorescence imaging (650–1000 nm for NIR-I and 1000–1700 nm for NIR-II) can provide great sensitivity and specificity for the real-time imaging of biological systems (46). To facilitate the noninvasive imaging of AD, fluorescence imaging probes must meet three requirements: (1) they must be (pseudo)permeable to cross the pathologically vulnerable blood–brain barrier (BBB); (2) they must bind to AD-specific cells and proteins, such as tau proteins in the brain; and (3) they must have absorbance and fluorescence emission spectra in the NIR window (650–1700 nm) (46).

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4. Radioactive Tracers/Fluorescence Imaging Probes in Molecular Imaging

AD molecular imaging is used to design molecular probes or imaging agents based on indicative molecular targets. The commonly used criteria for testing and evaluating the performance of the designed tracers are high affinity, high binding specificity, high specific activity, high blood–brain barrier penetration, and low side effects. Before achieving these goals, the metabolism and mechanisms of the peripheral and central nervous systems need to be fully studied. Their emission energy and half-life should be consistent with the target application. In addition, tracers need to maintain the same biological properties after positron or single-photon isotope labeling. The application of tracers and fluorescence imaging probes to diagnose AD is summarized in Table 1.

Table 1. Summary of studies indicating tracers and probes in application on AD.

Tracers/Probes Name	Design	Conclusion	Mechanisms	References
BSB	Utilized BSB to examine postmortem tissues from individuals suffering from different neurodegenerative conditions, and contrasted the findings with conventional histochemical dyes like thioflavin S and immunohistochemistry stains that were particular to the same lesions.	Various amyloids can be monitored in vivo using radio iodinated BSB derivatives or similar ligands as imaging agents.	Targeted amyloid	β- (47)
	Using multimodality positron emission tomography (PET) data from a clinically diverse group of patients suspected of having AD, find spatial patterns of glucose metabolism measured by FDG-PET and Aβ deposition measured by ((11)C)-	Joint analysis of PiB and FDG with pICA revealed a correlated component pair, in which increased frontal and decreased PCC/precuneus PiB correlated	Targeted β-amyloid	

	<p>labeled Pittsburgh Compound B (PiB-PET) that are correlated with particular clinical presentations.</p>	<p>with decreased FDG in the frontal, occipital and temporal regions.</p> <p>Fluorometamol injection labeled with radioactive fluorine-18 in a population nearing the end of life</p>		
Flutimol	<p>Following positron emission tomography imaging and brain donation, fluometamol injection tagged with radioactive fluorine 18 (Vizamyl; GE Healthcare) was administered.</p>	<p>demonstrated high sensitivity and specificity while remaining safe. The detection of brain β-amyloid plaque density in vivo has the potential to improve diagnostic precision in patients with cognitive impairments.</p> <p>Florbetaben PET exhibits a high</p>	Targeted β -amyloid	(49)
Flubetaben	<p>Open-label, nonrandomized, multicenter, phase 3 study to validate the (18)F-labeled β-amyloid tracer florbetaben by comparing in vivo PET imaging with post-mortem histopathology.</p>	<p>degree of sensitivity and specificity in identifying neuritic β-amyloid plaques verified by histology. As such, it could serve as a useful</p>	Targeted β -amyloid	(50)

		supplement to clinical diagnosis, especially in ruling out Alzheimer's disease.		
Attach oligomer- specific antibodies onto magnetic nanostructures	Attach oligomer-specific antibodies onto magnetic nanostructures and the complex is stable to bind to A β oligomers on cells and brain tissues to give a magnetic resonance imaging signal, and intranasally administered to an Alzheimer's disease mouse model.	The complex can enter the central nervous system through intranasal administration and combine with A β oligomers to produce detectable MRI signals. BIP-NMe2 may be utilized to create cutting- edge techniques for AD progression tracking, diagnosis, and treatment.	Targeted β - amyloid	(51)
BIP-NMe2	Designed and synthesized novel A β and tau imaging probes for SPECT.	^{11}C -THK5351 is more closely related to the pre-determined distribution of tau protein, while ^{11}C -PPB is more closely related to the co- deposition of β - amyloid protein and tau	Targeted Phosphorylated Tau Protein	(52)
^{11}C -THK5351 and ^{11}C -PPB	Evaluated the binding of two chemically different tau-specific PET tracers (^{11}C -THK5351 and ^{11}C - PBB3) in a head-to-head, in vivo, multimodal design.		Targeted Phosphorylated Tau Protein	(53)

¹⁸ F-T807	Used metabolite-corrected arterial input functions and dynamic PET imaging to describe the pharmacokinetics of ¹⁸ F-T807 in human participants	¹⁸ F-T807 can detect tau protein quantitatively and the plasma clearance rate is high. Brain region with strong binding to muscarinic acetylcholine receptor was not correlated with the glucose metabolism intensity measured by FDG-PET, and the defect range was larger in AD patients. The compound is also considered to be used to detect cholinergic neuronal degeneration, but its specificity and sensitivity for the diagnosis of AD need to be determined	Targeted Phosphorylated Tau Protein	(54)
¹²³ I-4-IQNB	High-resolution IQNB SPECT and FDG PET scans were performed on seven dementia patients and four healthy participants.		Targeted Neurotransmitter	(55)
¹²³ I-IBVM	Verify a streamlined acquisition technique that maintains quantification accuracy using a single ¹²³ I-IBVM static scan. Comparing three acquisition times to a kinetic analysis with MRTM2 was tested.		Targeted Neurotransmitter	(56)
¹²³ I-FP-CIT	Evaluated patient files to identify 67 individuals who had undergone ¹²³ I-FP-CIT SPECT in the Department of Nuclear Medicine at University Medical	Support for the use of ¹²³ I-FP-CIT SPECT as a supplemental diagnostic technique to	Targeted Neurotransmitter	(57)

	Center Göttingen (UMG) and were recruited for the study.	enhance the identification of prodromal DLB and probable dementia with Lewy Bodies (DLB) in a group of individuals with concurrent mental health symptoms
¹¹ C-doxepin	assessed it in ten individuals with Alzheimer's disease and eleven normal people (five elderly and six young)	Low signal intensity in the frontal and parietal lobes in patients with AD. More importantly, the expression concentration of the corresponding receptors represented by the signal in specific brain regions is closely related to the severity of the disease
		Targeted Neurotransmitter (58)

4.1. Targeted β -amyloid traces/Probes

In in vitro experiments, Congo red and sulfur T were the first compounds found to bind to β -fiber cross-structures, but their ionic charges prevented them from crossing the blood–brain barrier (59). Based on the structures of these two compounds, many PET tracers with ¹¹C or ¹⁸F as radioisotopes have been developed to directly visualize A β and its distribution in vivo to evaluate neuroinflammatory plaques.

The first compound used in vivo to recognize the β -amyloid protein was BSB, which has good blood–brain barrier penetration and plaque-specific binding power. Marie L. et al. compared the results of using BSB with those using standard histochemical dyes like thioflavin S and

immunohistochemical stains specifically meant for use on the same lesions in postmortem tissues from patients with a variety of neurodegenerative diseases. The diagnostic lesions were characterized by fibrillar intra- or extracellular lesions. These findings indicate that BSB binds not only to the external amyloid beta protein but also to a variety of intracellular lesions made up of aberrant tau and synuclein proteins. This suggests that derivatives of radioiodinated BSB or comparable ligands could be helpful imaging agents to track various amyloids in vivo (47).

After testing synthetic A β fibers from AD patients and cadaveric AD specimens, 6-OH-BAT-1, or PIB, stood out among more than 100 chemicals. During the prodromal phase of AD, PIB binding increases, showing bimodal behavior, and approximately 50% of positive individuals subsequently progress to AD. In prospective studies, increased PIB ligand retention was able to identify 82% of AD patients at follow-up, with only one prediction failure, demonstrating its high sensitivity and specificity. In addition, both amygdala and hippocampus atrophy were found to be positively correlated with PIB binding, suggesting differences in susceptibility to A β toxicity in different brain regions. Although this tracer seems promising for showing an increase in A β deposits before the clinical onset of AD, there is a ceiling effect, and large multicenter studies are still needed to develop objective evaluation criteria (48).

Based on sensitivity and specificity, the U.S. Food and Drug Administration has approved three radiopharmaceuticals that bind to the fiber-aggregating form of beta-amyloid. Flutimol, as a benzothiazole derivative, can estimate plaque density with a sensitivity of 88% and a specificity greater than 80%, and no adverse reactions were observed in the phase III clinical trials (49).

Osama et al. compared in vivo PET imaging with postmortem histopathology in an open-label, nonrandomized, multicenter, phase 3 research study to validate the (18)F-labeled β -amyloid tracer florbetaben. Flubetaben demonstrated a 97.9% sensitivity and an 88.9% specificity in identifying A β plaque, as validated by histology. As a result, florbetaben PET may be a useful addition for clinical diagnosis, and especially for the exclusion of AD, as the results revealed good sensitivity and specificity for the detection of neuritic β -amyloid plaques as verified by histology (50).

Viola et al. attached oligomer-specific antibodies to magnetic nanostructures to construct a stable compound which can enter the central nervous system through intranasal administration and combine with A β oligomers to produce detectable MRI signals. Targeting neurotoxic A β oligomers, these nanostructures have the potential to be valuable in assessing the effectiveness of novel medications and, eventually, in diagnosing and treating Alzheimer's disease in its early stages (51).

4.2. Targeted Phosphorylated Tau Protein Tracers/Probes

Fluorine-containing radioactive tracers have been developed to bind tau fibers with high precision. Watanabe [11] constructed a compound called BIP-NMe2. As a tau imaging probe, BIP-NMe2 demonstrated a strong and specific affinity for tau aggregation in brain slices from AD patients. Furthermore, BIP-NMe2 showed a strong initial uptake into the normal mouse brain and a rapid washout from it, indicating that its pharmacokinetics are advantageous for identifying tau aggregation through in vivo imaging (52).

The tracers labeled with ^{11}C are ^{11}C -THK5351 and ^{11}C -PPB. Konstantinos evaluated the binding of two chemically different tau-specific PET tracers (^{11}C -THK5351 and ^{11}C -PBB3) in a head-to-head, in vivo, multimodal design. It was found that the former signal is more closely related to the pre-determined distribution of tau proteins, while the latter is more closely related to the co-deposition of β -amyloid and tau proteins (53).

A PET radiotracer called ^{18}F -T807 was created to image tau protein aggregates, which have been linked to neurologic conditions like Alzheimer's disease and traumatic brain injury (TBI). Dustin et al. used metabolite-corrected arterial input functions and dynamic PET imaging to describe the pharmacokinetics of ^{18}F -T807 in human participants. The results indicated that ^{18}F -T807 can detect tau proteins quantitatively and the plasma clearance rate is high (54).

Though their value in early diagnosis or detection is inferior to that of A β -targeted tracers, the pathology of tau proteins is more closely linked to the clinical symptoms of AD and appears relatively

late in the disease's progression, making this imaging more commonly used in clinical trials to assess how well drugs delay the disease's progression.

4.3. Targeted Neurotransmitter Tracers/Probes

The most important energy consumption site of neurons is the synapse, that is, it is the place where active electrical activity and material exchange take place (60). Neuronal dysfunction and loss of function are usually reflected in the reduction in neurotransmitters or their receptors that are produced and released (61). Neurotransmitters and their receptors are generally considered to be markers of specific types of neurons (61).

Weinberger et al. found that when ^{123}I -4-IQNB was used in SPECT, the brain region with strong binding to the muscarinic acetylcholine receptor was not correlated with the glucose metabolism intensity measured by FDG-PET, and the defect range was larger in AD patients. In patients with AD, the signal of the tracer showed differences in the expression of receptors in different brain regions (55).

Drugs targeting vesicular ACh transporters are also under development; it has been found that the signal intensity of ^{123}I -IBVM, as a representative, is inversely proportional to age and symptom severity. In addition, the decrease in affinity was limited to the parietal lobe and occipital lobe in patients with Parkinson's disease (PD), but decreased in the whole cortex in patients with PD and AD. Therefore, this compound is also considered to be used to detect cholinergic neuronal degeneration, but its specificity and sensitivity for the diagnosis of AD need to be determined (56).

Dopaminergic neurons are widely studied in diseases such as PD, schizophrenia, and drug abuse. Tracers targeting dopamine transporters, such as ^{123}I -fluoropropyl carboxyl-methoxydeoxane (^{123}I -FP-CIT), help to distinguish between PD dementia, Lewy body dementia, and AD dementia. Niels advocated for the use of ^{123}I -FP-CIT SPECT as a supplemental diagnostic technique to enhance the identification of prodromal DLB and probable dementia with Lewy bodies (DLB) in a group of individuals with concurrent mental health symptoms. When DLB patients arrive with nigrostriatal dysfunction, psychiatric onset is more common than MCI onset. This suggests the value of extensive clinical phenotyping in memory clinics, which includes psychopathology assessment (57).

Higuchi et al. developed ^{11}C -doxepin, a radioligand for H(1) receptors, and using positron emission tomography, assessed cerebral histamine H(1) receptor binding in vivo in ten individuals with Alzheimer's disease and eleven without (five elderly and six young). The results showed a low signal intensity in the frontal and parietal lobes in patients with AD. More importantly, the expression concentration of the corresponding receptors represented by the signal in specific brain regions is closely related to the severity of the disease (58).

5. Summary and Conclusions

Molecular pathological tracers provide a possibility to accurately quantify brain changes through molecular imaging. The absence of diagnostic criteria to detect AD early is the main obstacle to theranostics in this field. As evidenced by our explanation of the pathophysiology of AD, various brain imaging modalities, and radioactive tracers/fluorescence probes, molecular imaging enhances our understanding of AD by establishing more crucial links between various patient groups at different stages of the disease. Typically, at least one type of imaging technique can lead to an AD diagnosis or the exclusion of dementia through the identification of intracranial causes that demonstrate structural alterations in the brain. Subjective vision is dependent on technology and software and has limitations when used to assess hippocampus volume or medial temporal lobe atrophy. In the future, the issues with conventional imaging techniques may be resolved by fusing automatic computer-aided techniques with different tracers or multimodal imaging.

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References

1. Jost BC: Grossberg GT. The Natural History of Alzheimer's Disease: A Brain Bank Study. *Journal of the American Geriatrics Society* (1995) 43(11):1248-55. Epub 1995/11/01. doi: 10.1111/j.1532-5415.1995.tb07401.x.
2. 2023 Alzheimer's Disease Facts and Figures. *Alzheimer's & dementia : the journal of the Alzheimer's Association* (2023) 19(4):1598-695. Epub 2023/03/16. doi: 10.1002/alz.13016.
3. Lane CA, Hardy J, Schott JM. Alzheimer's Disease. *European journal of neurology* (2018) 25(1):59-70. Epub 2017/09/06. doi: 10.1111/ene.13439.
4. Blennow K, de Leon MJ, Zetterberg H. Alzheimer's Disease. *Lancet (London, England)* (2006) 368(9533):387-403. Epub 2006/08/01. doi: 10.1016/s0140-6736(06)69113-7.
5. Chen YG. Research Progress in the Pathogenesis of Alzheimer's Disease. *Chinese medical journal* (2018) 131(13):1618-24. Epub 2018/06/27. doi: 10.4103/0366-6999.235112.
6. Dubois B, Hampel H, Feldman HH, Scheltens P, Aisen P, Andrieu S, et al. Preclinical Alzheimer's Disease: Definition, Natural History, and Diagnostic Criteria. *Alzheimer's & dementia : the journal of the Alzheimer's Association* (2016) 12(3):292-323. Epub 2016/03/26. doi: 10.1016/j.jalz.2016.02.002.
7. Sacks D, Baxter B, Campbell BCV, Carpenter JS, Cognard C, Dippel D, et al. Multisociety Consensus Quality Improvement Revised Consensus Statement for Endovascular Therapy of Acute Ischemic Stroke. *International journal of stroke : official journal of the International Stroke Society* (2018) 13(6):612-32. Epub 2018/05/23. doi: 10.1177/1747493018778713.
8. Jack CR, Jr., Knopman DS, Jagust WJ, Shaw LM, Aisen PS, Weiner MW, et al. Hypothetical Model of Dynamic Biomarkers of the Alzheimer's Pathological Cascade. *The Lancet Neurology* (2010) 9(1):119-28. Epub 2010/01/20. doi: 10.1016/s1474-4422(09)70299-6.
9. Valotassiou V, Malamitsi J, Papatriantafyllou J, Dardiotis E, Tsougos I, Psimadas D, et al. Spect and Pet Imaging in Alzheimer's Disease. *Annals of nuclear medicine* (2018) 32(9):583-93. Epub 2018/08/22. doi: 10.1007/s12149-018-1292-6.
10. Kim J, Jeong M, Stiles WR, Choi HS. Neuroimaging Modalities in Alzheimer's Disease: Diagnosis and Clinical Features. *International journal of molecular sciences* (2022) 23(11). Epub 2022/06/11. doi: 10.3390/ijms23116079.
11. Valotassiou V, Wozniak G, Sifakis N, Demakopoulos N, Georgoulas P. Radiopharmaceuticals in Neurological and Psychiatric Disorders. *Current clinical pharmacology* (2008) 3(2):99-107. Epub 2008/08/12. doi: 10.2174/157488408784293679.
12. Valotassiou V, Archimandritis S, Sifakis N, Papatriantafyllou J, Georgoulas P. Alzheimer's Disease: Spect and Pet Tracers for Beta-Amyloid Imaging. *Current Alzheimer research* (2010) 7(6):477-86. Epub 2010/05/12. doi: 10.2174/156720510792231757.
13. Mier W, Mier D. Advantages in Functional Imaging of the Brain. *Frontiers in human neuroscience* (2015) 9:249. Epub 2015/06/05. doi: 10.3389/fnhum.2015.00249.
14. Wang H, Zhang H. Reconsideration of Anticholinesterase Therapeutic Strategies against Alzheimer's Disease. *ACS chemical neuroscience* (2019) 10(2):852-62. Epub 2018/12/07. doi: 10.1021/acscemneuro.8b00391.
15. Whitehouse PJ, Price DL, Struble RG, Clark AW, Coyle JT, Delon MR. Alzheimer's Disease and Senile Dementia: Loss of Neurons in the Basal Forebrain. *Science (New York, NY)* (1982) 215(4537):1237-9. Epub 1982/03/05. doi: 10.1126/science.7058341.
16. Campanari ML, García-Ayllón MS, Belbin O, Galcerán J, Lleó A, Sáez-Valero J. Acetylcholinesterase Modulates Presenilin-1 Levels and γ -Secretase Activity. *Journal of Alzheimer's disease : JAD* (2014) 41(3):911-24. Epub 2014/04/05. doi: 10.3233/jad-140426.
17. Qiu X, Xu J, Cardoso Dos Santos M, Hildebrandt N. Multiplexed Biosensing and Bioimaging Using Lanthanide-Based Time-Gated Förster Resonance Energy Transfer. *Accounts of chemical research* (2022) 55(4):551-64. Epub 2022/01/28. doi: 10.1021/acs.accounts.1c00691.

18. Begcevic I, Brinc D, Brown M, Martinez-Morillo E, Goldhardt O, Grimmer T, et al. Brain-Related Proteins as Potential Csf Biomarkers of Alzheimer's Disease: A Targeted Mass Spectrometry Approach. *Journal of proteomics* (2018) 182:12-20. Epub 2018/04/24. doi: 10.1016/j.jprot.2018.04.027.
19. Guo T, Zhang D, Zeng Y, Huang TY, Xu H, Zhao Y. Molecular and Cellular Mechanisms Underlying the Pathogenesis of Alzheimer's Disease. *Molecular neurodegeneration* (2020) 15(1):40. Epub 2020/07/18. doi: 10.1186/s13024-020-00391-7.
20. Khan S, Barve KH, Kumar MS. Recent Advancements in Pathogenesis, Diagnostics and Treatment of Alzheimer's Disease. *Current neuropharmacology* (2020) 18(11):1106-25. Epub 2020/06/03. doi: 10.2174/1570159x18666200528142429.
21. Montalto MC, Farrar G, Hehir CT. Fibrillar and Oligomeric Beta-Amyloid as Distinct Local Biomarkers for Alzheimer's Disease. *Annals of the New York Academy of Sciences* (2007) 1097:239-58. Epub 2007/04/07. doi: 10.1196/annals.1379.023.
22. Kametani F, Hasegawa M. Reconsideration of Amyloid Hypothesis and Tau Hypothesis in Alzheimer's Disease. *Frontiers in neuroscience* (2018) 12:25. Epub 2018/02/15. doi: 10.3389/fnins.2018.00025.
23. Crews L, Masliah E. Molecular Mechanisms of Neurodegeneration in Alzheimer's Disease. *Human molecular genetics* (2010) 19(R1):R12-20. Epub 2010/04/24. doi: 10.1093/hmg/ddq160.
24. Guan PP, Cao LL, Wang P. Elevating the Levels of Calcium Ions Exacerbate Alzheimer's Disease Via Inducing the Production and Aggregation of B-Amyloid Protein and Phosphorylated Tau. *International journal of molecular sciences* (2021) 22(11). Epub 2021/06/03. doi: 10.3390/ijms22115900.
25. Sebastián-Serrano Á, de Diego-García L, Díaz-Hernández M. The Neurotoxic Role of Extracellular Tau Protein. *International journal of molecular sciences* (2018) 19(4). Epub 2018/03/28. doi: 10.3390/ijms19040998.
26. Sies H, Berndt C, Jones DP. Oxidative Stress. *Annual review of biochemistry* (2017) 86:715-48. Epub 2017/04/26. doi: 10.1146/annurev-biochem-061516-045037.
27. Karch CM, Goate AM. Alzheimer's Disease Risk Genes and Mechanisms of Disease Pathogenesis. *Biological psychiatry* (2015) 77(1):43-51. Epub 2014/06/22. doi: 10.1016/j.biopsych.2014.05.006.
28. Das P, Howard V, Loosbrock N, Dickson D, Murphy MP, Golde TE. Amyloid-Beta Immunization Effectively Reduces Amyloid Deposition in Fcrgamma-/- Knock-out Mice. *The Journal of neuroscience : the official journal of the Society for Neuroscience* (2003) 23(24):8532-8. Epub 2003/09/19. doi: 10.1523/jneurosci.23-24-08532.2003.
29. Heneka MT, Golenbock DT, Latz E. Innate Immunity in Alzheimer's Disease. *Nature immunology* (2015) 16(3):229-36. Epub 2015/02/18. doi: 10.1038/ni.3102.
30. Zott B, Busche MA, Sperling RA, Konnerth A. What Happens with the Circuit in Alzheimer's Disease in Mice and Humans? *Annual review of neuroscience* (2018) 41:277-97. Epub 2018/07/10. doi: 10.1146/annurev-neuro-080317-061725.
31. Lambert AJ, Brand MD. Reactive Oxygen Species Production by Mitochondria. *Methods in molecular biology (Clifton, NJ)* (2009) 554:165-81. Epub 2009/06/11. doi: 10.1007/978-1-59745-521-3_11.
32. Eckert A, Hauptmann S, Scherping I, Rhein V, Müller-Spahn F, Götz J, et al. Soluble Beta-Amyloid Leads to Mitochondrial Defects in Amyloid Precursor Protein and Tau Transgenic Mice. *Neuro-degenerative diseases* (2008) 5(3-4):157-9. Epub 2008/03/07. doi: 10.1159/000113689.
33. Luca M, Di Mauro M, Di Mauro M, Luca A. Gut Microbiota in Alzheimer's Disease, Depression, and Type 2 Diabetes Mellitus: The Role of Oxidative Stress. *Oxidative medicine and cellular longevity* (2019) 2019:4730539. Epub 2019/06/11. doi: 10.1155/2019/4730539.
34. Cheignon C, Tomas M, Bonnefont-Rousselot D, Faller P, Hureau C, Collin F. Oxidative Stress and the Amyloid Beta Peptide in Alzheimer's Disease. *Redox biology* (2018) 14:450-64. Epub 2017/10/29. doi: 10.1016/j.redox.2017.10.014.
35. Xie F, Peng F. Radiopharmaceuticals for Assessment Of altered Metabolism and Biometal Fluxes In brain Aging and Alzheimer's Disease With positron Emission Tomography. *Journal of Alzheimer's disease : JAD* (2017) 59(2):527-36. Epub 2017/07/04. doi: 10.3233/jad-170280.
36. Liu S, Jie C, Zheng W, Cui J, Wang Z. Investigation of Underlying Association between Whole Brain Regions and Alzheimer's Disease: A Research Based on an Artificial Intelligence Model. *Frontiers in aging neuroscience* (2022) 14:872530. Epub 2022/06/25. doi: 10.3389/fnagi.2022.872530.
37. Jeong HS, Park JS, Yang Y, Na SH, Chung YA, Song IU. Cerebral Perfusion Changes after Acetyl-L-Carnitine Treatment in Early Alzheimer's Disease Using Single Photon Emission Computed Tomography. *Dementia and neurocognitive disorders* (2017) 16(1):26-31. Epub 2017/03/01. doi: 10.12779/dnd.2017.16.1.26.
38. Lochocki B, Boon BDC, Verheul SR, Zada L, Hoozemans JJM, Ariese F, et al. Multimodal, Label-Free Fluorescence and Raman Imaging of Amyloid Deposits in Snap-Frozen Alzheimer's Disease Human Brain Tissue. *Communications biology* (2021) 4(1):474. Epub 2021/04/17. doi: 10.1038/s42003-021-01981-x.
39. Kapoor V, McCook BM, Torok FS. An Introduction to Pet-Ct Imaging. *Radiographics : a review publication of the Radiological Society of North America, Inc* (2004) 24(2):523-43. Epub 2004/03/18. doi: 10.1148/rg.242025724.

40. Omami G, Tamimi D, Branstetter BF. Basic Principles and Applications of (18)F-Fdg-Pet/Ct in Oral and Maxillofacial Imaging: A Pictorial Essay. *Imaging science in dentistry* (2014) 44(4):325-32. Epub 2014/12/05. doi: 10.5624/isd.2014.44.4.325.
41. Jiang W, Chalich Y, Deen MJ. Sensors for Positron Emission Tomography Applications. *Sensors (Basel, Switzerland)* (2019) 19(22). Epub 2019/11/21. doi: 10.3390/s19225019.
42. Spanoudaki V, Levin CS. Photo-Detectors for Time of Flight Positron Emission Tomography (ToF-Pet). *Sensors (Basel, Switzerland)* (2010) 10(11):10484-505. Epub 2010/01/01. doi: 10.3390/s101110484.
43. Jack CR, Jr., Shiung MM, Weigand SD, O'Brien PC, Gunter JL, Boeve BF, et al. Brain Atrophy Rates Predict Subsequent Clinical Conversion in Normal Elderly and Amnesic Mci. *Neurology* (2005) 65(8):1227-31. Epub 2005/10/26. doi: 10.1212/01.wnl.0000180958.22678.91.
44. Johnson KA, Jones K, Holman BL, Becker JA, Spiers PA, Satlin A, et al. Preclinical Prediction of Alzheimer's Disease Using Spect. *Neurology* (1998) 50(6):1563-71. Epub 1998/06/20. doi: 10.1212/wnl.50.6.1563.
45. Jagust W, Thisted R, Devous MD, Sr., Van Heertum R, Mayberg H, Jobst K, et al. Spect Perfusion Imaging in the Diagnosis of Alzheimer's Disease: A Clinical-Pathologic Study. *Neurology* (2001) 56(7):950-6. Epub 2001/04/11. doi: 10.1212/wnl.56.7.950.
46. Choi HS, Kim HK. Multispectral Image-Guided Surgery in Patients. *Nature biomedical engineering* (2020) 4(3):245-6. Epub 2020/03/14. doi: 10.1038/s41551-020-0536-7.
47. Schmidt ML, Schuck T, Sheridan S, Kung MP, Kung H, Zhuang ZP, et al. The Fluorescent Congo Red Derivative, (Trans, Trans)-1-Bromo-2,5-Bis-(3-Hydroxycarbonyl-4-Hydroxy)Styrylbenzene (Bsb), Labels Diverse Beta-Pleated Sheet Structures in Postmortem Human Neurodegenerative Disease Brains. *The American journal of pathology* (2001) 159(3):937-43. Epub 2001/09/11. doi: 10.1016/s0002-9440(10)61769-5.
48. Laforce R, Jr., Tosun D, Ghosh P, Lehmann M, Madison CM, Weiner MW, et al. Parallel Ica of Fdg-Pet and Pib-Pet in Three Conditions with Underlying Alzheimer's Pathology. *NeuroImage Clinical* (2014) 4:508-16. Epub 2014/05/13. doi: 10.1016/j.nicl.2014.03.005.
49. Curtis C, Gamez JE, Singh U, Sadowsky CH, Villena T, Sabbagh MN, et al. Phase 3 Trial of Flutemetamol Labeled with Radioactive Fluorine 18 Imaging and Neuritic Plaque Density. *JAMA neurology* (2015) 72(3):287-94. Epub 2015/01/27. doi: 10.1001/jamaneurol.2014.4144.
50. Sabri O, Sabbagh MN, Seibyl J, Barthel H, Akatsu H, Ouchi Y, et al. Florbetaben Pet Imaging to Detect Amyloid Beta Plaques in Alzheimer's Disease: Phase 3 Study. *Alzheimer's & dementia : the journal of the Alzheimer's Association* (2015) 11(8):964-74. Epub 2015/04/01. doi: 10.1016/j.jalz.2015.02.004.
51. Viola KL, Sbarboro J, Sureka R, De M, Bicca MA, Wang J, et al. Towards Non-Invasive Diagnostic Imaging of Early-Stage Alzheimer's Disease. *Nature nanotechnology* (2015) 10(1):91-8. Epub 2014/12/23. doi: 10.1038/nnano.2014.254.
52. Watanabe H. [Development of Spect Probes for in Vivo Imaging of B-Amyloid and Tau Aggregates in the Alzheimer's Disease Brain]. *Yakugaku zasshi : Journal of the Pharmaceutical Society of Japan* (2017) 137(11):1361-5. Epub 2017/11/03. doi: 10.1248/yakushi.17-00156.
53. Chiotis K, Stenkrona P, Almkvist O, Stepanov V, Ferreira D, Arakawa R, et al. Dual Tracer Tau Pet Imaging Reveals Different Molecular Targets for (11)C-Thk5351 and (11)C-Pbb3 in the Alzheimer Brain. *European journal of nuclear medicine and molecular imaging* (2018) 45(9):1605-17. Epub 2018/05/13. doi: 10.1007/s00259-018-4012-5.
54. Wooten DW, Guehl NJ, Verwer EE, Shoup TM, Yokell DL, Zubcevik N, et al. Pharmacokinetic Evaluation of the Tau Pet Radiotracer (18)F-T807 ((18)F-Av-1451) in Human Subjects. *Journal of nuclear medicine : official publication, Society of Nuclear Medicine* (2017) 58(3):484-91. Epub 2016/09/24. doi: 10.2967/jnumed.115.170910.
55. Weinberger DR, Jones D, Reba RC, Mann U, Coppola R, Gibson R, et al. A Comparison of Fdg Pet and Iqnb Spect in Normal Subjects and in Patients with Dementia. *The Journal of neuropsychiatry and clinical neurosciences* (1992) 4(3):239-48. Epub 1992/01/01. doi: 10.1176/jnp.4.3.239.
56. Mazère J, Mayo W, Pariscoat G, Schulz J, Allard M, Fernandez P, et al. Simplified Quantification Method for in Vivo Spect Imaging of the Vesicular Acetylcholine Transporter with 123i-Iodobenzovesamicol. *Journal of nuclear medicine : official publication, Society of Nuclear Medicine* (2015) 56(6):862-8. Epub 2015/04/25. doi: 10.2967/jnumed.114.147074.
57. Hansen N, Lange C, Timäus C, Wiltfang J, Bouter C. Assessing Nigrostriatal Dopaminergic Pathways Via 123i-Fp-Cit Spect in Dementia with Lewy Bodies in a Psychiatric Patient Cohort. *Frontiers in aging neuroscience* (2021) 13:672956. Epub 2021/07/09. doi: 10.3389/fnagi.2021.672956.
58. Higuchi M, Yanai K, Okamura N, Meguro K, Arai H, Itoh M, et al. Histamine H(1) Receptors in Patients with Alzheimer's Disease Assessed by Positron Emission Tomography. *Neuroscience* (2000) 99(4):721-9. Epub 2000/09/07. doi: 10.1016/s0306-4522(00)00230-x.
59. Hayne DJ, Lim S, Donnelly PS. Metal Complexes Designed to Bind to Amyloid-B for the Diagnosis and Treatment of Alzheimer's Disease. *Chemical Society reviews* (2014) 43(19):6701-15. Epub 2014/03/29. doi: 10.1039/c4cs00026a.

60. Jang S, Nelson JC, Bend EG, Rodríguez-Laureano L, Tueros FG, Cartagena L, et al. Glycolytic Enzymes Localize to Synapses under Energy Stress to Support Synaptic Function. *Neuron* (2016) 90(2):278-91. Epub 2016/04/14. doi: 10.1016/j.neuron.2016.03.011.
61. Traver S, Salthun-Lassalle B, Marien M, Hirsch EC, Colpaert F, Michel PP. The Neurotransmitter Noradrenaline Rescues Septal Cholinergic Neurons in Culture from Degeneration Caused by Low-Level Oxidative Stress. *Molecular pharmacology* (2005) 67(6):1882-91. Epub 2005/03/24. doi: 10.1124/mol.104.007864.

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