

Review

Not peer-reviewed version

Unmet Horizons: Assessing the Challenges in the Treatment of TP53-Mutated Acute Myeloid Leukemia

[Christos Stafylidis](#) , Dimitra Vlachopoulou , Christina-Nefeli Kontandreopoulou , [Panagiotis Diamantopoulos](#)

*

Posted Date: 29 December 2023

doi: 10.20944/preprints202312.2256.v1

Keywords: acute myeloid leukemia; TP53 mutations; TP53-mutated acute myeloid leukemia; novel targeted AML therapies; AML immunotherapies



Preprints.org is a free multidiscipline platform providing preprint service that is dedicated to making early versions of research outputs permanently available and citable. Preprints posted at Preprints.org appear in Web of Science, Crossref, Google Scholar, Scilit, Europe PMC.

Copyright: This is an open access article distributed under the Creative Commons Attribution License which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Review

Unmet Horizons: Assessing the Challenges in the Treatment of *TP53*-Mutated Acute Myeloid Leukemia

Christos Stafylidis, Dimitra Vlachopoulou, Christina-Nefeli Kontandreopoulou and Panagiotis Diamantopoulos *

Hematology Unit, First Department of Internal Medicine, Laikon General Hospital, National and Kapodistrian University of Athens, Athens, Greece

* Correspondence: pandiamantopoulos@gmail.com

Abstract: Acute myeloid leukemia (AML) remains a challenging hematologic malignancy. The presence of *TP53* mutations in AML poses a therapeutic challenge, considering that standard treatments face significant setbacks in achieving meaningful responses. There is a pressing need for the development of innovative treatment modalities to overcome resistance to conventional treatments attributable to the unique biology of *TP53*-mutated (*TP53*^{mut}) AML. This review underscores the role of *TP53* mutations in AML, examines the current landscape of treatment options, and highlights novel therapeutic approaches, including targeted therapies, combination regimens, and emerging immunotherapies as well as agents being explored in preclinical studies, for their potential to address the unique hurdles posed by *TP53*^{mut} AML.

Keywords: acute myeloid leukemia; *TP53* mutations; *TP53*-mutated acute myeloid leukemia; novel targeted AML therapies; AML immunotherapies

1. Introduction

P53 is a tumor suppression protein encoded by the *TP53* gene and a vital regulator of genomic stability preservation in response to DNA damage, through the activation of DNA repair pathways or triggering of cell-cycle arrest and induction of apoptosis [1].

Acute myeloid leukemia (AML) harboring *TP53* mutations, which is now classified according to the International Consensus Classification (ICC) of myeloid neoplasms and acute leukemias as a distinct AML subtype [2], presents a redoubtable clinical challenge since it is associated with adverse prognosis [3–5]. These mutations are observed mostly in treatment-related, relapsed, and elderly AML patients, often characterized by remarkable genomic instability [3–5]. While the rate of *TP53* mutations in de novo AML is 5–10%, it is significantly increased in older patients with de novo AML, with a median age of 60–67 years, up to 25% [6]. Higher frequency rates, up to 35%, are reported in treatment-related AML (t-AML) [6], whereas the highest rates up to 70% are observed in patients with a complex karyotype and those with loss of chromosome 17/17p, 5/5q, or 7/7q [4,5,7].

Mutated *TP53* induces genomic instability, hence contributing to leukemogenesis, while it also confers unique characteristics to AML and results in evasion of apoptosis, inherent resistance to conventional chemotherapy, and poor clinical outcomes [1,3–5]. Several studies report lower complete response (CR) rates, inferior complete remission duration, and dismal overall survival (OS) among *TP53*-mutated (*TP53*^{mut}) AML patients [3–5]. Importantly, *TP53* mutations have been found to be predictors of adverse outcomes irrespective of age, chemotherapy regimen, or complex karyotype [3,4]. Moreover, AML patients with *TP53* mutations are at a higher risk of relapse and death after allogeneic stem cell transplantation (aSCT) [8]. Diagnostic approaches such as fluorescence in situ hybridization (FISH), next-generation sequencing (NGS), and in silico approaches may promptly identify these patients and may hold significant predictive value, thus facilitating decisions on treatment strategies [9,10]. Importantly, loss of *TP53* detected by FISH at diagnosis has been correlated with poor response to chemotherapy [10].

The challenging management of *TP53*-mutated AML highlights the crucial need for the development of novel therapeutic approaches. During past years, targeted agents, immunotherapy,

and combination strategies have come into the spotlight and have become the subject of intense research in this setting, in order to overcome the hurdle of the intrinsic resistance caused by *TP53* mutations. In this review, we discuss the role of *TP53* mutations in AML, outcomes with current treatment options, as well as data on innovative agents that are currently being investigated in the preclinical setting and clinical trials.

The role of TP53 in AML

TP53 is a 20-kbp tumor suppressor gene located on chromosome 17p13.1 [6]. It encodes for the transcription factor p53 and functions as the “guardian of the human genome” [6]. The p53 protein is a key transcription factor playing a pivotal role in tumor suppression through DNA repair, cell cycle arrest, differentiation, senescence, apoptosis, autophagy, metabolism, and chemosensitivity [11,12]. The protein contains five important domains: the N-terminal trans-activation domain, a proline-rich domain, a central DNA-binding domain (DBD), a C-terminal oligomerization domain, and a regulatory domain [6].

Since its first description in 1979, *TP53* has been the most frequently mutated gene across all human cancers. More than 50% of human tumors carry *TP53* mutations, whereas many others carrying wild-type *TP53* alleles exhibit decreased *TP53* activity via other mechanisms [13,14]. One of the most well-studied functions of *TP53* is its role in limiting cellular proliferation in response to aberrant oncogene expression. Therefore, *TP53* inactivation by gene deletion or mutation enhances the effect of oncogenes and plays a key role in promoting uncontrolled proliferation of cancer cells. Germline *TP53* mutations cause Li-Fraumeni syndrome (LFS), a disorder that predisposes patients to different types of cancer, including sarcomas, breast cancer, leukemias, and lymphomas [15,16].

It has been observed that a vast majority of de novo AML have intact, unaltered *TP53* alleles [17]. However, the frequency of genomic *TP53* alterations is increased in certain patients [4–7]. In AML, *TP53* mutations are mostly missense somatic substitutions, mostly heterozygous, and include those that are observed in the known hotspot sites of the gene [18]. Diverse genetic aberrations in *TP53*, such as chromosomal alterations leading to allelic gain, losses, or frameshift insertions and deletions have also been described, with the impact ranging from partial to complete loss of function (LOF) mostly in the germline LOF mutations that underlie LFS [18]. Gain of function (GOF) mutations with varied effect sizes are also present in different *TP53* mutants and are thought to mostly result from their binding to different proteins including transcription factors [19,20]. GOF *TP53* mutants have also been reported to affiliate with epigenetic pathways, e.g., binding and enhancing transcription of the methyl-transferases MLL1 and MLL2 [21]. Monoallelic *TP53* mutations frequently have co-mutations in other genes, mostly *TET2*, *SF3B1*, *ASXL1*, and *DNMT3A* and are likely to be subclonal events with varying impacts on outcomes in MDS/AML [22]. On the other hand, multihit *TP53*^{mut} MDS/AML represents a distinct disorder, with co-mutations occurring in less than 25% of cases [23]. Finally, the mutational burden of *TP53* has also arisen as a crucial prognostic factor in AML with therapy-choice implications [24]. Despite, being one of the most studied genes, *TP53* is still considered “undruggable”, so future studies are needed to ascertain the role of *TP53* mutations in myeloid malignancies.

Current treatment options for TP53-mutated AML

Intensive chemotherapy (IC) with an anthracycline and cytosine arabinoside (AraC) remains the backbone of treatment in patients with newly diagnosed (ND) AML. Eligibility for IC is largely based upon age and comorbidities, hence patients with *TP53*^{mut} AML, who are frequently elderly, may be unfit for this treatment option. Additionally, the presence of *TP53* mutations in AML patients who receive anthracycline- and cytarabine-based induction chemotherapy has been previously associated with inferior outcomes, with reported initial response rates of 20-30% and a poor OS of less than a year [4,5]. Baseline *TP53* variant allelic frequency (VAF) has been previously shown to be predictive of response to cytarabine-based treatment, with VAF ≤ 40% being associated with a superior CR and CR with incomplete hematologic recovery (CRI) rate of 79% and a median OS of 7.3 months

juxtaposed to VAF > 40%, which has been associated with a CR/CRi rate of 35% and a median OS of 4.7 months [24].

Lower-intensity therapies, including low-dose cytarabine (LDAC) monotherapy or in combination with cladribine and hypomethylating agents (HMAs), are also being used in these patients and are an attractive option since they are accompanied by significantly lower toxicity. Regarding the efficacy of lower-intensity chemotherapy regimens data are conflicting. A single-center study has demonstrated superior CR rates in *TP53*^{mut} AML patients receiving IC as compared to patients treated with lower-intensity regimens (45% vs. 14.3%), but no difference in OS (8.8 months vs. 9.4 months respectively) [25]. On the contrary, a study has demonstrated lower CR rates among patients with *TP53*^{mut} AML regardless of regimen intensity and has also shown that the intensity of therapy does not predict improved survival [3].

Azacitidine (AZA) and decitabine (DEC) are HMAs that are currently being used either alone or in combination with other agents in the management of *TP53*^{mut} AML. Although the efficacy of AZA monotherapy in AML has been previously demonstrated [26], with a reported CR/CRi rate being 28% [26], its efficacy in AML harboring *TP53* mutations is not well established. A randomized phase 3 trial comparing the impact of AZA on the survival of AML patients versus conventional care regimens (CCREs), including IC, LDAC, and best supportive care, has shown that the median OS was prolonged in *TP53*^{mut} patients treated with AZA compared to those treated with CCREs (7.2 months vs. 2.4 months respectively); however, this result did not reach statistical significance [27].

Correspondingly, data regarding the efficacy of DEC monotherapy is conflicting [28,29]. A retrospective study has shown similar CR rates among *TP53*^{mut} AML patients treated with either LDAC or a 5-day or 10-day DAC regimen (DEC5 and DEC10 respectively) as well as comparable OS rates among all treatment arms [28]. Accordingly, a study of AML patients treated with DEC has also shown no response or survival benefit in *TP53*^{mut} patients versus *TP53* wild type (*TP53*^{wt}) ones [29]. Conversely, a single-institution trial evaluated the efficacy of DEC10 in AML patients and demonstrated exceptionally higher responses in *TP53*^{mut} patients (100% vs. 41% in the *TP53*^{wt} arm) [30]. These responses were accompanied by clearance of *TP53*^{mut} leukemic clones in most of the cases, but mutation clearance was never complete [30]. Although *TP53* VAF predicts response and OS in AML patients treated with IC, no effect has been demonstrated on response rates and OS in those treated with HMAs [24]. Moreover, despite the fact that DEC augments chemotherapy responses in *TP53*^{mut} AML, with a currently unknown underlying mechanism, these responses are not durable and do not significantly affect subclones bearing *TP53* mutations [30]. Nonetheless, this enhanced effect paves the way for the design of more combination strategies in these patients.

Recently, the combination of AZA and Venetoclax (VEN), a selective B-cell lymphoma-2 (BCL-2) inhibitor, has become the cornerstone in the treatment of elderly AML patients who are ineligible for IC [31]. First-line treatment of *TP53*^{mut} AML with poor-risk cytogenetics with AZA and VEN initially showed promising results, with a study reporting CR and CRi combined rates of 41% in the combination arm versus 17% in the AZA monotherapy arm, exceeding the historical standards of 28% CR rates [5,32]. Yet, the duration of response (DOR) and median OS were similar among both treatment arms (6.5 versus 6.7 months and 5.2 versus 4.9 months, respectively) [32]. Furthermore, a study evaluating the efficacy of DEC10 and VEN combination in patients with ND AML has shown pronouncedly inferior outcomes in *TP53*^{mut} compared to *TP53*^{wt} patients, with reported ORR and CR/CRi rates in *TP53*^{mut} patients of 66% and 57%, versus 89% and 77% respectively in the *TP53*^{wt} group [33]. Importantly, the 60-day mortality rate was higher in *TP53*^{mut} patients (26% versus 4% in *TP53*^{wt}) and OS was profoundly lower in these patients (5.2 versus 19.4 months in *TP53*^{wt}) [33]. It has been previously demonstrated that *TP53* mutations disrupt the BAX/BAK pathway and establish an elevated activation threshold in leukemic cells (LCs). Although VEN initially suppresses this effect, LCs finally avoid BCL-2 inhibition due to competitive advantage, thus conferring resistance to VEN [34]. Additionally, adaptive resistance associated with alterations in mitochondrial homeostasis and increased oxidative phosphorylation has also been observed [35,36]. Despite this resistance to VEN, its incorporation in novel combination therapies in *TP53*^{mut} AML may still be promising. Concurrent

inhibition of BCL-2 and myeloid leukemia 1 (MCL-1) can achieve long-term outcomes by increasing the early apoptotic response in TP53-deficient cells, thus making this approach highly promising [35].

Currently available combination strategies

Lately, several combination strategies have been investigated in clinical trials regarding the management of AML patients harboring *TP53* mutations. A recent cohort study has evaluated the efficacy of DEC, LDAC, aclarubicin, and granulocyte colony-stimulating factor (G-CSF) [DCAG regimen] versus standard chemotherapy in *TP53*^{mut} AML patients [36], based on the previous encouraging results of a multicentre phase 2 trial which reported 82.4% ORR and 64.7% CR rates of the DCAG regimen in elderly AML patients [37]. Although differences were not statistically significant, a trend towards higher ORR, CR, and OS rates was observed in the DCAG arm [36]. Importantly, patients with poor cytogenetics in the DCAG arm displayed superior responses with a significantly higher CR rate of 56.3% and a median OS of 7.8 months (versus CR 0% and median OS of 3 months in the standard chemotherapy arm) [36].

The combination of LDAC with clofarabine or cladribine alternating with DEC has been evaluated in the management of treatment-naïve elderly AML with reported CR and CRi rates of 59% and 7% respectively and a median OS of 12.5 months [38–40]. Long-term results from these studies have shown that among all patients, those with *TP53* mutations yielded the lowest responses with a composite complete remission (cCR) rate of 44% and a poor median OS of 5.4 months [40]. Addition of AZA prior to treatment with high-dose cytarabine (HiDAC) and mitoxantrone, considering that epigenetic priming induced by AZA before cytotoxic chemotherapy could contribute to enhanced responses has been previously examined in a phase 1 study of high-risk AML patients, demonstrating an ORR of 61% [41]. However, patients with *TP53* mutations seemed not to benefit from this regimen [41]. A study of *TP53*^{mut} AML patients has demonstrated that the combination of DEC and chidamide, a histone deacetylase inhibitor (HDACi) with a priming regimen consisting of omacetaxine mepesuccinate, which is an alkaloid herbal derivative, cytarabine and G-CSF (HAG) yields potent responses with an ORR of 71.4%, with manageable toxicity [42]. Yet, the study sample size was really small and conclusions cannot be drawn, but these promising results guarantee further investigation in the near future [42].

CPX-351, a liposomal formulation of cytarabine and daunorubicin, constitutes the contemporary treatment of AML with myelodysplasia-related changes (MRC-AML) and t-AML [43]. Real-life data from the French cohort study of CPX-351 has indicated that *TP53* mutations were the only predictive factor of inferior responses in multivariate analysis, although high-risk molecular prognosis subgroups, including patients with *ASXL1* and *RUNX1* mutations displayed higher than expected response rates [43]. In accordance, similar results have been reported by another retrospective study, demonstrating inferior responses in *TP53*^{mut} patients, who achieved lower CR and CRi rates as compared to *TP53*^{wt} patients (33% versus 62% respectively) [44]. Consistently, a post hoc analysis of a randomized phase 3 trial has also shown poor outcomes in *TP53*^{mut} patients [45]. Opposingly, a German retrospective analysis has demonstrated that the presence of *TP53* mutations did not impact responses to CPX-351 or survival [46]. Nonetheless, the role of CPX-351 in the management of *TP53*^{mut} AML needs to be further evaluated.

Novel therapeutic agents

Considerable progress has also been made regarding the development of novel agents, including mutant p53-targeted approaches and immunotherapy.

A. Targeted treatments

Novel targeted therapies incorporated into combination regimens have also been explored in the *TP53*^{mut} AML setting. Pevonedistat (PEVO), an inhibitor of the NEDD8-activating enzyme (NAE) seems to exert antiproliferative effects on LCs and preclinical data supports synergistic effects with AZA and VEN [47–49]. A phase 1b study of unfit, treatment-naïve AML patients treated with PEVO

and AZA showed improved responses with an ORR of 50%, with *TP53*^{mut} patients achieving a CR and partial response (PR) rate of 80% [47]. Based on these results, a phase 2 study consisting of *TP53*^{mut} AML patients was conducted, but failed to show enhanced CRR rates and was prematurely terminated [48]. Intriguingly, a phase 1/2 study evaluating the efficacy of combined PEVO, AZA, and VEN in ND secondary AML reported a CR/CRi rate of 64%, but a dreadful 1-year OS of 0% in *TP53*^{mut} patients, contrary to a median OS of 18 months in *TP53*^{wt} patients [49]. Moreover, the DOR differed significantly among these patients [49]. These conflicting results may be attributable to the different VAF of patients since the second study included only *TP53*^{mut} patients with a VAF of >30%. Nevertheless, although these results seem discouraging, data are scarce and derive from small studies, thus PEVO may still have a role to play in this setting.

Ibrutinib, a Bruton tyrosine kinase (BTK) inhibitor has been previously shown to impede the proliferation of human AML blasts in vitro, either alone or combined with cytarabine or daunorubicin [50]. A randomized phase 2 study evaluated the outcomes of adding ibrutinib to DEC10 versus DEC10 monotherapy in elderly, previously untreated AML patients [51]. Surprisingly, although the addition of ibrutinib did not yield favorable outcomes, *TP53*^{mut} was correlated with higher responses and CR/CRi rates of 56% [51]. However, these responses were not translated into a superior OS [51]. Nevertheless, although ibrutinib's efficacy in *TP53*^{mut} AML needs to be further validated, it remains a highly appealing approach.

Finally, bortezomib, a proteasome inhibitor, is widely investigated in the management of AML patients, since it has been associated with potent antiproliferative properties [52]. A randomized phase 2 trial of AML patients treated with either combined bortezomib and DEC10 or DEC10 alone failed to show a potential advantage of the combination in those with *TP53* mutations [52]. Moreover, the addition of bortezomib conferred no benefit to the study patients overall [52]. Conclusively, targeted therapies' efficacy remains ambiguous and warrants further exploration of these agents in *TP53*^{mut} AML through large clinical trials.

B. *TP53* targeting agents

Although p53 has traditionally been considered undruggable, efforts have been made to overcome this hurdle and have led to the development of a new, small molecule, called "p53 reactivation and induction of massive apoptosis" (PRIMA-1), that can reverse the mutant conformation of p53, which induces protein unfolding and restores wild-type functions to mutant p53, such as induction of apoptosis and promotion of cell cycle arrest [53]. Eprenetapopt (EP) or APR-246, a methylated derivative of PRIMA-1 (PRIMA-1^{MET}), is a first-in-class agent that binds covalently to cysteine residues in mutant p53 protein [53]. Preclinical studies have demonstrated that EP exerts apoptotic effects on AML cell lines and primary LCs from AML patients in a dose-dependent manner [54]. Noteworthy, the presence of *TP53* mutations did not significantly affect sensitivity to this agent [54]. Subsequent studies have shown significant synergistic cytotoxicity of EP and AZA in *TP53*^{mut} primary cells from MDS/AML patients [55]. Apart from the reported mutant p53 reactivation, preclinical data have also demonstrated that EP results in glutathione depletion and induction of ferroptosis, irrespective of the *TP53* status, thus indicating a different mechanism of action that leads to p53-independent cell death [56,57].

Recently, EP's efficacy in combination with AZA has been evaluated in patients with *TP53*^{mut} MDS and AML in two phase 2 studies, one in the USA and another one in Europe [58,59]. EP was administered by an intravenous infusion, at a fixed dose, on days 1-4 of each 28-day cycle and AZA was administered subcutaneously, at the standard dose, for seven days of each 28-day cycle [58,59]. *TP53*^{mut} AML patients in the US trial have achieved ORR and CR rates of 64% and 36%, respectively, and a median OS of 10.8 months [58]. However, the sample size was significantly small and only patients with oligoblastic AML (20-30% marrow blasts) were included [58]. The European trial, additionally including *TP53*^{mut} AML patients with more than 30% marrow blasts, has demonstrated an ORR of 33% and a CR rate of 17% [59]. However, none of the patients with a high blast count achieved a CR [59]. Median OS in patients with less and more than 30% marrow blasts was 13.9 months and 3.0 months respectively [59]. Both studies have reported a significant reduction in the

TP53 VAF and p53 expression by immunochemistry in responding patients, with some patients achieving *TP53* negativity (VAF <5%) [59]. These findings indicate a promising efficacy, since ORR, CR, and OS rates are generally higher than those reported with AZA monotherapy, particularly for patients with oligoblastic AML [59]. Of note, patients with *TP53*^{mut} MDS have also yielded high response rates in both studies, with a CR rate of around 50% [58,59]. The doublet of EP and AZA has also been evaluated in a phase 2 trial of *TP53*^{mut} AML patients, as post-aSCT maintenance therapy administered for up to 12 cycles, with reported relapse-free survival and median OS being 12.5 and 20.6 months, respectively, which is quite encouraging for this high-risk population [60]. The triplet combination of EP, AZA, and VEN has also been studied recently in the *TP53*^{mut} AML setting. In a phase 1, dose-finding and expansion study, patients with ND *TP53*^{mut} AML achieved an ORR, CR, and CR/CRi rate of 64%, 38%, and 56%, respectively, whereas DOR and median OS were 4.2 and 7.3 months, respectively [61]. Importantly, the blast count did not have an impact on patients' responses [61]. Moreover, *TP53* negativity (VAF <5%) by NGS was achieved in 27% [61]. These results are highly promising, since CR rates are higher than the CR rates of 22% that have been reported in patients with previously untreated *TP53*^{mut} AML receiving AZA in combination with VEN [61]. Collectively, EP has demonstrated promising efficacy in *TP53*^{mut} AML patients and provides the basis for further investigation in randomized clinical trials in the near future.

C. Immunotherapeutic approaches

Increasing interest has also grown regarding the use of immunotherapeutic agents in *TP53*^{mut} AML. CD47 or the “don't eat me signal” is a transmembrane protein that interacts with signal-regulatory protein alpha (SIRPa), which is expressed in macrophages, and impedes macrophage-mediated phagocytosis [62]. LCs have high levels of CD47, thus escaping immune surveillance [62]. Increased CD47 expression in AML hematopoietic stem cells (HSCs) has been independently correlated with inferior outcomes, thus making the CD47/SIRPa axis an appealing therapeutic target [63]. Blockade of CD47 in AML models has resulted in the induction of phagocytosis and elimination of LCs [63,64]. Magrolimab (MAG) is a novel, first-in-class, IgG4 monoclonal antibody against CD47 that acts as a macrophage checkpoint inhibitor and has exerted synergistic effects with AZA and VEN in preclinical in vitro and in vivo studies, with the latter agents eliciting “eat me” signals by upregulating calreticulin [64,65]. A phase 1b study has evaluated the combination of MAG and AZA in patients with previously untreated AML, ineligible for IC, with the majority of patients (82.8%) having *TP53* mutations [65]. The CR rate was similar among *TP53*^{mut} and *TP53*^{wt} patients (31.9% and 32.2%, respectively), whereas OS was 9.8 months and 18.9 months, respectively [65]. A phase 1/2 study of the triplet AZA, VEN, and MAG in ND elderly AML, high-risk (HR)-AML, and relapsed/refractory (R/R) AML patients has demonstrated an ORR and a CR rate of 74% and 41%, respectively, in ND *TP53*^{mut} patients [66]. Although preliminary results were encouraging, a subsequent phase 3 trial (ENHANCE-2), evaluating MAG and AZA versus physician's choice of VEN and AZA or IC in *TP53*^{mut} AML, was prematurely terminated, since MAG failed to demonstrate a survival benefit compared to standard of care [67].

Several other agents targeting the disrupted CD47-SIRPa axis are also being explored in MDS/AML. Maplirpacept (MAP) or TTI-622 is a soluble fusion protein with anti-CD47 properties that, unlike other anti-CD47 agents, binds minimally to normal erythrocytes [68]. In vivo studies of AML xenografts have demonstrated the efficacy of TTI-622 in enhancing macrophage-mediated phagocytosis [68]. A phase 1a/1b dose-escalation and expansion trial of MAP alone or in combination with other agents in patients with advanced hematologic malignancies, including a cohort of ND *TP53*^{mut} AML patients treated with MAP and AZA is currently active (NCT03530683). Lemzoparlimab is another anti-CD47 agent that is currently being investigated in patients with HR-MDS and AML, in combination with AZA and/or VEN (NCT04202003, NCT0491206). A recent phase 1b study has evaluated the efficacy of AK117, an anti-CD47 agent, in combination with AZA as frontline treatment in AML patients and has demonstrated a CR and CR/CRi rate of 45% and 55%, respectively [69]. Evorpaccept (EVO) or ALX148 has been associated with increased LC phagocytosis in *TP53*^{mut} AML lines and mouse xenograft models, and its combination with HMA and/or VEN

confers better survival [70]. Hence, EVO entered a phase 1/2 trial, which studied its combination with VEN and AZA in patients with AML (ASPEN-05 trial, NCT04755244). However, ASPEN-05 was terminated, based on data from the ASPEN-02 trial, which was also terminated, reporting failure to achieve superior outcomes in MDS patients treated with EVO and AZA [71]. Other anti-CD47 agents that are currently being studied in AML, combined with AZA and VEN include DSP107 (NCT04937166) and SL-172154 (NCT05275439), whereas a phase 1b study (NCT04485052) of IB188 (letaplimab) plus AZA in AML was suspended.

T-cell immunoglobulin mucin-3 (TIM-3) is a cell-surface glycoprotein that is constitutively expressed on the surface of certain immune cells, such as the T-cells and acts as a co-inhibitory receptor [72,73]. When interacting with one of its ligands, such as galectin-9, TIM-3 prompts the inhibition of T-cell responses [72,73]. It has also been demonstrated that TIM-3 is overexpressed in LCs and that TIM-3⁺ AML leukemic stem cells (LSCs) secrete galectin-9 in an autocrine loop, that regulates self-renewal of these cells, via enhanced NF- κ B and β -catenin signaling [72,73]. Hence, antibodies targeting TIM-3 provide a highly appealing therapeutic opportunity. Sabatolimab (SAB) or MBG453, is a humanized, high-affinity IgG4 antibody that targets TIM-3 [74]. A phase 1b study that has evaluated SAB in combination with HMAs in patients with HR-MDS and ND AML displayed promising preliminary results, with ND AML patients exhibiting ORR and CR rates of 40% and 25%, respectively and a median duration of response of 12.6 months [74]. Importantly, durable responses have been observed in patients with adverse-risk mutations, including *TP53*, indicating that this combination may be effective in the *TP53*^{mut} setting [74]. The addition of VEN is also explored in an ongoing phase 1b trial (NCT03940352), which investigates the combination of SAB and VEN in AML and HR-MDS patients. Furthermore, a phase 2 trial (STIMULUS-AML1, NCT04150029) is currently underway, investigating the combination of SAB, AZA, and VEN in patients with ND AML.

CD123 also serves as an appealing candidate for targeting. CD123 is a component of the interleukin-3 receptor (IL-3R) that plays a multifaceted role in hematopoiesis and immune responses; it stimulates HSC proliferation through activation of the PI3K/MAPK pathway and upregulation of antiapoptotic proteins, while it also participates in the modulation of T-cell responses [75]. CD123 is widely expressed in blasts of AML patients, and its overexpression has been correlated with poor prognosis [76]. In vitro and in vivo studies have demonstrated that a novel CD123 x CD3 dual-affinity retargeting (DART) molecule mediates T-cell activation and proliferation, leading to dose-dependent elimination of AML cell lines and primary AML blasts [75]. Flotetuzumab (FLOT), is a CD123 x CD3 DART antibody that has been evaluated in a phase 1/2 study in R/R AML after primary induction failure or in early relapse, with the reported ORR being 30% [77]. Remarkably, *TP53*^{mut} patients yielded encouraging responses with a CR rate of 47% and a median OS of 10.3 months in responding patients [78]. Currently, early-phase trials are also exploring FLOT in post-transplant relapsed AML (NCT04582864, NCT05506956). Pivekimab sunirine (PVEK) or IMG632, is a first-in-class antibody-drug conjugate (ADC) with a high affinity for CD123, which has displayed synergy with AZA and/or VEN in preclinical models [79]. An ongoing multicenter, phase 1/2 study investigates PVEK as a triplet with AZA and VEN or in combination with MAG, in patients with R/R AML or ND CD123⁺ AML [80]. Preliminary data have shown that treatment with the triplet in R/R AML patients has led to an ORR and a composite CR rate (coCR) rate of 51% and 31%, respectively [79]. However, VEN-naïve patients yielded significantly higher responses, than those with prior exposure to VEN [79]. Recent data regarding patients in the ND AML cohort receiving frontline triplet treatment have reported robust responses with a CR and a coCR rate of 52% and 66%, respectively, whereas CR and coCR rates for *TP53*^{mut} patients were 13% and 47%, respectively [81]. Rapid minimal residual disease (MRD) negativity was achieved in 73% of patients achieving coCR [81]. Exceptionally, high coCR_{MRD} rates have been demonstrated among adverse risk patients, *TP53*^{mut} included [81]. Triple combination therapy has been also associated with a manageable safety profile [79,81]. A phase 1 clinical trial of PVEK in combination with fludarabine, high-dose cytarabine (HiDAC), G-CSF, and idarubicin (FLAG-Ida) for frontline treatment of ND adverse-risk AML is ongoing (NCT06034470).

Tagraxofusp (TAG) is a CD123-targeted immunotoxin and has been evaluated as monotherapy in a phase 1 trial of AML and MDS patients, with reported responses being modest [82]. However,

recent data have supported that AZA, when combined with TAG, overcomes TAG resistance and restores TAG sensitivity, thus providing a rationale for the combination of these two agents [83]. A phase 1b trial of TAG with AZA and/or VEN in AML and MDS patients is ongoing and preliminary results indicate promising efficacy [84]. Remarkably, *TP53*^{mut} patients have achieved a CR/CRi/morphologic leukemia-free state (MLFS) rate of 54%, with a CR rate of 31% [84]. Early-phase studies include the use of TAG as maintenance therapy for post-transplant AML patients (NCT05233618), for ND secondary AML after previous exposure to HMA (NCT05442216), and in combination with gemtuzumab ozogamicin for R/R AML (NCT05716009). Vibecotamab or XmAb14045, a CD3-CD123 bispecific T-cell engaging (BiTE) antibody is currently being investigated in the treatment of R/R AML, with preliminary data reporting modest ORR rates of 14% [85]. Vibecotamab has also been associated with cytokine release syndrome (CRS), which is manageable with premedication [85]. Other CD123-targeting agents that are in early clinical development include APV0436, MGD024, and CD123 chimeric antigen receptor T-cell (CAR-T) therapy [86]. In summary, these results suggest that these agents may have a role to play in AML patients, *TP53*^{mut} included, and research in this field continues to uncover new insights into potential applications of CD123.

Immune-checkpoint inhibitor-based approaches have also been studied in AML. Ipilimumab, an antibody targeting cytotoxic T-lymphocyte-associated protein 4 (CTLA-4), has yielded a CR rate of 42% in patients with post-aSCT relapsed AML [87]. Nivolumab (NIVO), a programmed cell death protein 1 (PD-1) inhibitor, has been evaluated as first-line AML therapy, in combination with idarubicin and AraC and has yielded encouraging responses in *TP53*^{mut} patients [88]. NIVO has also been studied in R/R AML patients, in combination with AZA, with a modest ORR of 33% and an ORR of 13% in *TP53*^{mut} patients [89]. A recent phase 2 trial of R/R AML patients receiving pembrolizumab, a PD-1 inhibitor, with HiDAC has demonstrated promising clinical activity in *TP53*^{mut} patients, reporting a CR rate of 40% [90]. A randomized phase 2 trial of AZA with or without durvalumab (DURV), a PD-L1 inhibitor, as first-line treatment for elderly AML patients, has failed to show a potential benefit, since the addition of DURV did not enhance clinical outcomes and recorded ORR and OS were similar among both treatment arms [91]. Interestingly, responses were similar between *TP53*^{mut} and *TP53*^{wt} patients (ORR 35% and 34%, respectively) [91]. Nonetheless, the use of CTLA-4, PD1, and PD-L1 inhibitors in AML necessitates further research for strong conclusions to be drawn.

Leukocyte-associated immunoglobulin-like receptor 1 (LAIR-1) is an immune inhibitory receptor which is present on most immune cell subsets and is implicated in immunosuppressive responses [92]. It has been demonstrated that LAIR-1 is highly expressed in AML blasts and LSCs and is responsible for the inhibition of intracellular downstream survival signals and blast proliferation, while its expression is relatively lower in normal HSCs, thus rendering LAIR-1 an ideal anti-leukemic target [92]. NC525 is a humanized monoclonal antibody that binds specifically to AML blasts and LSCs, while sparing normal hematopoiesis, and induces apoptosis through a unique signaling pathway, without evidence of immunomodulatory effects on other immune subsets [92]. Furthermore, it has been shown that NC525 displays synergistic activity when combined with AZA and VEN and results in leukemic cell destruction in patients who are refractory to VEN-AZA [92]. A phase 1 trial investigating the safety and tolerability of NC525 in patients with advanced HMs, including R/R AML is underway (NCT05787496).

D. Other agents

Murine double minute protein 2 (MDM2) is an E3 ubiquitin ligase that negatively regulates the activity of p53 [93]. MDM2 interacts with p53 and promotes its degradation via ubiquitination [93]. Inhibition of MDM2 mediates antileukemic effects in *TP53*^{wt} AML through an increase in p53 levels [93]. A phase 1/1b study has evaluated the use of idasanutlin (IDASA), an oral MDM2 inhibitor (MDM2i), either alone or in combination with AraC, in unfit for IC patients with R/R or ND AML and has demonstrated a coCR rate of 18.9% and 35.6% in patients receiving monotherapy or combination treatment, respectively [94]. A subsequent randomized, double-blind, phase 3 trial (MIRROS trial), evaluating IDASA combined with AraC or placebo in R/R AML patients, has failed

to show an improvement in OS, although the overall remission rate was enhanced by the addition of IDAS [95]. Although MDM2i require wt-p53 to be effective, hence being unable to act directly in *TP53*^{mut} AML, they indirectly induce degradation of MCL-1, which is associated with VEN resistance, thus providing a rationale for the combined use of MDM2i and VEN, even in *TP53*^{mut} patients, in order to overcome VEN resistance [96]. Milademetan, an MDM2i, in combination with LDAC, with or without VEN, has been recently explored in AML with discouraging responses and significant gastrointestinal toxicity [97]. A phase 1b trial of IDASA and VEN in R/R AML patients has shown modest responses, with *TP53* mutations having been associated with unfavorable outcomes [98]. A concern regarding the use of MDM2i is whether they select for the outgrowth of *TP53*^{mut} clones since studies have reported emergent *TP53* mutations in some patients [97,98]. Nevertheless, further studies are needed in order to assess the safety and efficacy of these agents in this setting.

Various agents for *TP53*^{mut} treatment are currently in early clinical development. Arsenic trioxide (ATO) has been shown to inactivate *TP53*, by inducing proteasomal degradation of mutant p53 and upregulating *TP53*^{wt} functions [99]. Therefore, it can lead to inactivation of proliferation of LCs and apoptosis promotion. Atorvastatin, is a potent destabilizing agent of mutant p53, since it has been shown that it effectively induces degradation for conformational or misfolded p53 mutants, via inhibition of the mevalonate pathway, with minimal effects on wt-p53 and DNA contact mutants [100]. Collectively, these findings provide insight into exploring arsenic compound-based and statin-based therapies for AML harboring *TP53* mutations. A trial of combined ATO and DEC to treat *TP53*^{mut} AML/MDS (PANDA-T0 trial, NCT03855371) and a pilot trial of atorvastatin in *TP53*^{mut} and *TP53*^{wt} malignancies (NCT03560882) are currently enrolling.

E. Novel treatments in *TP53*^{mut} AML: does a promising future await?

In brief, targeted treatments, including those targeting mutant p53, along with immunotherapeutic agents have yielded vastly different response rates in *TP53*^{mut} AML patients, as is seen in Table 1. [45–50,56–59,63,64,72,76,79,82,86–89,96] However, these responses have not been translated into a survival benefit, since the reported median OS was less than a year in the majority of the studies. [45–50,56–59,63,64,72,76,79,82,86–89,96] Although these results may be discouraging, they derive mostly from small studies, hence further study is required and these agents may still hold promise for this challenging clinical setting, particularly in combination with HMAs.

Table 1. Available current data from studies of novel agents in *TP53*-mutated acute myeloid leukemia.

Agent	Study type	Regimen	Population	<i>TP53</i> ^{mut} patients (n)	Response	OS (months)	Ref
Pevonedistat	Open-label, phase 1B, multicenter	PEVO + AZA	Unfit, untreated AML patients	8	CR/CRi/PR 75%	NR	47
	Open-label, phase 2, multicenter	PEVO + AZA	≥ 60y, untreated, <i>TP53</i> ^{mut} AML patients	10	CR/CRi 0%	mOS 6.2m	48
	Phase 1/2 single-center	PEVO + AZA + VEN	Unfit ND secondary AML patients, MDS and CMML patients after failure of HMAs	11	CR/CRi 64%	mOS 8.1m	49
Ibrutinib	Randomized, phase 2, multicenter	Ibrutinib + DEC10 vs. DEC10 monotherapy	Elderly, unfit, untreated AML patients	27	CR/CRi 56% in both arms	Inferior OS compared to <i>TP53</i> ^{wt} patients	51
Bortezomib	Randomized, phase 2, multicenter	Bortezomib + DEC10 vs.	Elderly, ND AML patients	12 in combination	CR 17% in combination	1-year OS 17% in combination	52

		DEC10 monotherapy		arm and 14 in DEC10 arm	arm vs. 21% in DEC10 arm	vs. 21% in DEC10 arm	
Eprenetapopt (APR-246)	Open-label, phase 1b/2, multicenter	APR-246 + AZA	≥ 18y, <i>TP53</i> ^{mut} , HMA-naïve MDS, MDS/MPN, CMML, oligoblastic (20-30% blasts) AML patients	11	ORR 64% CR 36%	mOS 10.8m	58
	Open-label, phase 2, multicenter	APR-246 + AZA	≥ 18y, <i>TP53</i> ^{mut} , HMA-naïve MDS, CMML, oligoblastic and > 30% blasts AML patients	18	ORR 33% CR 17% CR 27% in oligoblastic AML CR 0% in AML with >30% blasts	mOS 10.4m mOS 13.4m in oligoblastic AML mOS 3m in AML with >30% blasts	59
	Open-label, phase 2, multicenter	APR-246 + AZA as maintenance treatment after HCT	≥ 18y, <i>TP53</i> ^{mut} , MDS or AML patients post-HCT	14	NA	mRFS 12.5m mOS 20.6m (for all patients)	60
	Open-label, phase 1, multicenter	APR-246 + AZA + VEN	≥ 18y, <i>TP53</i> ^{mut} , untreated AML patients	43	ORR 64% CR 38% CR/CRi 56%	mOS 7.3m	61
	Open-label, phase 1b, multicenter	MAG + AZA	≥ 18y, unfit, untreated AML patients	72	ORR 47% CR 32%	mOS 9.8m	65
Magrolimab	Open-label, phase 1b/2, multicenter	MAG + AZA + VEN	≥ 18y, unfit, ND or untreated secondary and VEN-naïve or VEN- exposed R/R AML patients	27 in the ND and untreated secondary AML cohort	ORR 74% CR 86% CR/CRi 63%	1-year OS 53%	66
Sabatolimab (MBG453)	Open-label, phase 1b, multicenter	SAB + HMA	Unfit, ND or R/R HMA-naïve AML, high risk HMA- naïve MDS and CMML patients	NR	ORR 53.8% in ND AML patients with at least 1 ELN adverse-risk mutation, including <i>TP53</i>	NR	74
Flotetuzumab	Open-label, phase 1/2, multicenter	FLOT monotherapy	R/R AML/MDS patients	15 in the R/R AML cohort	ORR 60% CR 47%	mOS 10.3m	78
Pivekimab sunirine (IMGN632)	Open-label, phase 1b/2, multicenter	PVEK + AZA + VEN	ND, CD123 ⁺ AML patients	19	CR 13% coCR 47%	NR	81
Tagraxofusp	Open-label, phase 1b, multicenter	TAG + AZA +/- VEN	Unfit, ND, CD123 ⁺ AML patients	13	CR 31% CR/CRi/MLFS 54%	mOS 9.5m mPFS 5.1m	84
Nivolumab	Open-label, phase 1/2, single-center	NIVO + idarubicin + AraC	Fit for IC, ND AML patients	8	CR/CRi/CRp 50% for <i>TP53</i> ^{mut} patients CR 67% for patients with poor-risk mutation	NR	88

profile, TP53 included							
	Open-label, phase 2, single-center	NIVO + AZA	R/R AML patients	16	ORR 28%	NR	89
Pembrolizumab	Open-label, phase 2, two-center	PEMBRO + HiDAC	R/R AML patients	5	CR 40%	NR	90
Durvalumab	Randomized, open-label, phase 2, multicenter	DURV + AZA vs. AZA monotherapy	Elderly, unfit, ND AML patients	21 in the combination arm, 17 in the monotherapy arm	ORR 34% in TP53 ^{mut} AML vs. ORR 33% in TP53 ^{wt} AML for both treatment arms	NR	91
Idasanutlin	Open-label, phase 1, multicenter	IDASA + VEN	Unfit, ND sAML or R/R AML patients	10	CR/CRi/CRp 20%	mOS 3.7m	98

.PEV: pevonedistat; AZA, azacitidine; AML, acute myeloid leukemia; CR, complete remission; CR, complete remission with incomplete count recovery; PR, partial remission; NR, not reported; mOS, median overall survival; VEN, venetoclax; ND, newly diagnosed; MDS, myelodysplastic syndrome; CMML, chronic myelomonocytic leukemia; HMA, hypomethylating agent; DEC10, 10days decitabine treatment; MDS/MPN, myelodysplastic syndrome/myeloproliferative neoplasm; ORR, overall response rate; mRFS, median relapse free survival; HCT, hematopoietic cell transplant; NA, not applicable; MAG, Magrolimab; R/R, relapsed/refractory; SAB, sabatolimab; ELN, European Leukemia Net; FLOT, flotetuzumab; PVEK, pivekimab sunirine; coCR, composite complete remission; TAG, tagraxofusp; MLFS, morphologic leukemia-free state; PFS, progression free survival; NIVO, nivolumab; AraC, cytarabine; IC, intensive chemotherapy; CRp, complete remission with incomplete platelet recovery; PEMBRO, pembrolizumab; HiDAC, high dose cytarabine; DURV, durvalumab; IDASA, idasanutlin; sAML, secondary acute myeloid leukemia.

Novel agents in preclinical studies

Despite growth in understanding AML pathobiology, therapeutic progress is still inadequate. The required improvement has yielded development of novel drugs targeting various molecularly defined AML entities, including p53-based therapies. Cells with mutant or deleted TP53 frequently have a defective G1 checkpoint and are more dependent on the G2 checkpoint to repair DNA damage; the G2 checkpoint allows p53-deficient AML cells to repair genetic lesions and continue through the cell cycle. Consistent with this finding, inhibition of kinases involved in the G2 checkpoint, such as aurora kinase A (AURKA) and, aurora kinase B (AURKB) has induced mitotic catastrophe and p53-independent cell death in TP53^{mut} cancer cells [101]. TP-0903, a small molecule originally developed as an AXL inhibitor, is a multikinase inhibitor with activity against AURKA/B, Chk1/2, and other cell cycle regulators and has activity in models of drug-resistant AML with both WT and mutated TP53 [101]. Xpo7, a putative nuclear/cytoplasmic transporter, was recently identified as a factor necessary for the survival of Trp53-knockout (KO) AML cells with the performance of genome-wide CRISPR-Cas9 screens using Trp53-KO and WT mouse AML cells, indicating a synthetic lethal relationship between TP53 and XPO7 [102]. TP53^{mut} targeted therapy aims to abolish TP53^{mut} cancer cells or to rescue p53 mutational inactivation. The pharmacological strategies are directed toward regaining p53^{wt}-like conformation and p35^{mut} tumor-suppressive functions, abrogating distinct mechanisms underlying p53^{mut} GOF, and promoting p53^{mut} degradation [103–109]. On the other hand, dysfunctional p53^{wt} targeted therapy aims to rescue p53^{wt} by addressing various AML-related p53^{wt} inactivating mechanisms. As aforementioned, one such strategy involves MDM2i that disrupts WTp53-MDM2 interactions. [93] Table 2 summarizes the available preclinical studies targeting TP53/p53 in AML in vitro and in vivo models. Finally, agents that can help overcome resistance to currently available therapies have also been investigated. Targeting mitochondrial metabolism with novel antimitochondrial agents, including electron transport chain complex inhibitors, pyruvate

dehydrogenase inhibitors, and mitochondrial ClpP protease agonists has led to enhanced sensitivity of leukemic cells to combination treatment with VEN and AraC and substantially delayed relapse. [110]

Table 2. TP53/p53 targeting in preclinical studies.

Compound	Target	Model	Mechanism of action	Combination with other therapy	Ref
Compounds that restore p53 wildtype function					
PK7088	Y220C	Cell lines	Selective induction of caspase 3/7 in p53-Y220C cells and restoration of p53 ^{wt} conformation	NA	
PhiKan083	Y220C	In silico	BAX nuclear export induction to the mitochondria, and restoration of p53 nontranscriptional apoptosis	NA	105
NSC319726 (ZMC1)	R175H, R172H	In silico	Zinc chelator, providing optimal zinc concentration for mut p53-R175H proper folding; induction of ROS formation	NA	106
			Restoration of p53 ^{wt} conformation and activity with MDM2-dependent degradation		
PEITC	R175H	Cell lines	Sensitization of p53 ^{mut} to proteasome-mediated degradation and further restoration of p53 ^{wt} conformation and transactivation	NA	107
COTI-2	R175H, R273H	Cell lines	Restoration of p53 ^{wt} activity by targeting and binding to misfolded p53 mutant	NA	109
Compounds that induce degradation of mutant p53					
PU-H71 (Zelavespib)	R248W	Molm13 and K562 cells	Induction of cell death in <i>TP53</i> ^{wt} , <i>TP53</i> -KO, and <i>TP53</i> ^{mut} cells	VEN enhanced the killing of both <i>TP53</i> ^{wt} and <i>TP53</i> ^{mut} cells by PU-H71	108
Compounds with miscellaneous targets					
TP-0903 (Duberminatinib)	Multikinase inhibitor	Cell lines	AURKA/B inhibition in <i>TP53</i> ^{mut} AML G2/M arrest and apoptosis in <i>TP53</i> ^{mut} AML cells Chk1/2 inhibition in <i>TP53</i> ^{mut} AML cells DNA damage response through upregulation of p53	Combination of TP-0903 and DEC is active in vitro demonstrating an additive effect TP-0903/DEC prolongs survival in vivo in a HL-60 xenograft model	101
XPO7		Mouse cell lines	<i>Trp53</i> -KO cells are vulnerable to XPO7 depletion, while XPO7 functions as a <i>Trp53</i> -dependent tumor suppressor in <i>Trp53</i> ^{wt} AML cells	NA	102

Synthetic lethal relationship between <i>TP53</i> and <i>XPO7</i>					
RETRA	mutp53-p73 binding	Mouse cell lines	Increase in the expression level of p73, and release of p73 from the blocking complex with p53 ^{mut} , which produces tumor-suppressor effects similar to the functional reactivation of p53. RETRA is active against tumor cells expressing a variety of p53 mutants and does not affect normal cells.	NA	103

p53, protein 53; mut, mutant; BAX, Bcl-2 associated X protein; TP53, tumor protein 53; KO, knockout; CHIP, carboxyl terminus of Hsc70 interacting protein; AML, acute myeloid leukemia; AURKA/B, aurora kinase A/B; CHK1/2, checkpoint kinase 1/2; pH2AX, phosphor-histone h2AX; HL-60; human leukemia cell line 60; XPO7, exportin 7; RETRA, reactivation of transcriptional reporter activity; p73, protein 73; VEN, venetoclax; DEC, decitabine; NA, not applicable.

2. Conclusions

In conclusion, the management of *TP53*^{mut} AML remains a formidable clinical challenge and current therapeutic approaches yield suboptimal outcomes, thus denoting the urgency for tailored strategies addressing the molecular landscape of *TP53* mutations along with the inherent resistance and aggressive nature of the disease. Although the armamentarium of promising approaches keeps expanding, most novel agents have not been met with satisfactory efficacy, with survival rates similar to current treatments. However, these data derive mostly from quite small studies, so strong conclusions cannot be drawn. Among novel treatments, immunotherapeutic agents such as pevonedistat, nivolumab, and flotetuzumab have displayed promising efficacy and warrant rigorous investigation through large clinical trials. Preclinically, agents that target *TP53*/p53 have also yielded encouraging responses, thus necessitating their study in the clinical setting. What is certain, is that as we delve deeper into the molecular landscape of AML, the significance of *TP53* mutations becomes increasingly apparent, thus requiring a paradigm shift to our clinical strategies, with hopes of fostering a brighter future for patients with *TP53*^{mut} AML.

Funding: This research received no funding.

Conflicts of Interest statement: CS reports personal fees from Abbvie, outside of the submitted work. PD reports personal fees from Novartis, Amgen, Janssen, Abbvie, BMS, and Roche, outside of the submitted work.

References

1. Lakin ND, Jackson SP. Regulation of p53 in response to DNA damage. *Oncogene*. 1999;18(53):7644-7655. doi:10.1038/sj.onc.1203015
2. Döhner H, Wei AH, Appelbaum FR, et al. Diagnosis and management of AML in adults: 2022 recommendations from an international expert panel on behalf of the ELN. *Blood*. 2022;140(12):1345-1377. doi:10.1182/blood.2022016867
3. Kadia TM, Jain P, Ravandi F, et al. TP53 mutations in newly diagnosed acute myeloid leukemia: Clinicomolecular characteristics, response to therapy, and outcomes. *Cancer*. 2016;122(22):3484-3491. doi:10.1002/cncr.30203
4. Bowen D, Groves MJ, Burnett AK, et al. TP53 gene mutation is frequent in patients with acute myeloid leukemia and complex karyotype, and is associated with very poor prognosis. *Leukemia*. 2009;23(1):203-206. doi:10.1038/leu.2008.173
5. Rücker FG, Schlenk RF, Bullinger L, et al. TP53 alterations in acute myeloid leukemia with complex karyotype correlate with specific copy number alterations, monosomal karyotype, and dismal outcome. *Blood*. 2012;119(9):2114-2121. doi:10.1182/blood-2011-08-375758
6. George B, Kantarjian H, Baran N, et al. TP53 in Acute Myeloid Leukemia: Molecular Aspects and Patterns of Mutation. *Int J Mol Sci*. 2021;22(19):10782. doi:10.3390/ijms221910782
7. Hunter AM, Sallman DA. Current status and new treatment approaches in TP53 mutated AML. *Best Pract Res Clin Haematol* 2019;32:134–44.

8. Della Porta MG, Galli A, Bacigalupo A, et al. Clinical Effects of Driver Somatic Mutations on the Outcomes of Patients With Myelodysplastic Syndromes Treated With Allogeneic Hematopoietic Stem-Cell Transplantation. *J Clin Oncol.* 2016;34(30):3627-3637. doi:10.1200/JCO.2016.67.3616
9. Madarász K, Mótýán JA, Bedekovics J, et al. Deep Molecular and In Silico Protein Analysis of p53 Alteration in Myelodysplastic Neoplasia and Acute Myeloid Leukemia. *Cells.* 2022;11(21):3475. doi:10.3390/cells11213475
10. Tavor S, Rothman R, Golan T, et al. Predictive value of TP53 fluorescence in situ hybridization in cytogenetic subgroups of acute myeloid leukemia. *Leuk Lymphoma.* 2011;52(4):642-647. doi:10.3109/10428194.2010.551571
11. Levine AJ. p53: 800 million years of evolution and 40 years of discovery. *Nat Rev Cancer* 2020;20:471–80.
12. Ferreira CG, Tolis C, Giaccone G. p53 and chemosensitivity. *Ann Oncol.* 1999;10:1011–1021.
13. Vogelstein B, Lane D, Levine AJ. Surfing the p53 network. *Nature.* 2000;408:307-310
14. Hainaut P, Hernandez T, Robinson A, et al. IARC database of p53 gene mutations in human tumors and cell lines: updated compilation, revised formats and new visualisation tools. *Nucleic Acids Res.* 1998;26:205-213
15. Olivier M, Hollstein M, Hainaut P. TP53 mutations in human cancers: origins, consequences, and clinical use. *Cold Spring Harb Perspect Biol* 2010;2:a001008.
16. Srivastava S, Zou Z, Pirolo K, et al. Germline transmission of a mutated p53 gene in a cancer-prone family with Li-Fraumeni syndrome. *Nature.* 1990;348:747-749
17. Ley TJ, Ding L, Raphael BJ, et al. Genomic and epigenomic landscapes of adult de novo acute myeloid leukemia the cancer genome atlas research network. *N Engl J Med.* 2013;368:2059-2074
18. Barbosa K, Li S, Adams PD, Deshpande AJ. The role of TP53 in acute myeloid leukemia: challenges and opportunities. *Genes Chromosomes Cancer* 2019;58:875–88
19. Brosh R, Rotter V. When mutants gain new powers: news from the mutant p53 field. *Nat Rev Cancer.* 2009;9:701-713
20. Shetzer Y, Molchadsky A, Rotter V. Oncogenic mutant p53 gain of function nourishes the vicious cycle of tumor development and cancer stem-cell formation. *Cold Spring Harb Perspect Med.* 2016;6(10):a026203.
21. Zhu J, Sammons MA, Donahue G, et al. Gain-of-function p53 mutants co-opt chromatin pathways to drive cancer growth. *Nature.* 2015;525:206-211
22. Bernard E, Nannya Y, Hasserjian RP, et al. Implications of TP53 allelic state for genome stability, clinical presentation and outcomes in myelodysplastic syndromes. *Nat Med* 2020;26:1549–56
23. Sill H, Zebisch A, Haase D. Acute myeloid leukemia and myelodysplastic syndromes with TP53 aberrations: a distinct stem cell disorder. *Clin Cancer Res* 2020;26:5304–9.
24. Short NJ, Montalban-Bravo G, Hwang H, et al. Prognostic and therapeutic impacts of mutant TP53 variant allelic frequency in newly diagnosed acute myeloid leukemia. *Blood Adv.* 2020;4(22):5681-5689. doi:10.1182/bloodadvances.2020003120
25. Bewersdorf JP, Shallis RM, Gowda L, et al. Clinical outcomes and characteristics of patients with TP53-mutated acute myeloid leukemia or myelodysplastic syndromes: a single center experience. *Leuk Lymphoma.* 2020;61(9):2180-2190. doi:10.1080/10428194.2020.1759051
26. Dombret H, Seymour JF, Butrym A, et al. International phase 3 study of azacitidine vs conventional care regimens in older patients with newly diagnosed AML with >30% blasts. *Blood.* 2015;126(3):291-299. doi:10.1182/blood-2015-01-621664
27. Döhner H, Dolnik A, Tang L, et al. Cytogenetics and gene mutations influence survival in older patients with acute myeloid leukemia treated with azacitidine or conventional care. *Leukemia.* 2018;32(12):2546-2557. doi:10.1038/s41375-018-0257-z
28. Boddu P, Kantarjian H, Ravandi F, et al. Outcomes with lower intensity therapy in TP53-mutated acute myeloid leukemia. *Leuk Lymphoma.* 2018;59(9):2238-2241. doi:10.1080/10428194.2017.1422864
29. Middeke JM, Teipel R, Röllig C, et al. Decitabine treatment in 311 patients with acute myeloid leukemia: outcome and impact of TP53 mutations - a registry based analysis. *Leuk Lymphoma.* 2021;62(6):1432-1440. doi:10.1080/10428194.2020.1864354
30. Welch JS, Petti AA, Miller CA, et al. TP53 and Decitabine in Acute Myeloid Leukemia and Myelodysplastic Syndromes. *N Engl J Med.* 2016;375(21):2023-2036. doi:10.1056/NEJMoa1605949
31. DiNardo CD, Jonas BA, Pullarkat V, et al. Azacitidine and Venetoclax in Previously Untreated Acute Myeloid Leukemia. *N Engl J Med.* 2020;383(7):617-629. doi:10.1056/NEJMoa2012971

32. Pollyea DA, Pratz KW, Wei AH, et al. Outcomes in Patients with Poor-Risk Cytogenetics with or without TP53 Mutations Treated with Venetoclax and Azacitidine. *Clin Cancer Res.* 2022;28(24):5272-5279. doi:10.1158/1078-0432.CCR-22-1183
33. Kim K, Maiti A, Loghavi S, et al. Outcomes of TP53-mutant acute myeloid leukemia with decitabine and venetoclax. *Cancer.* 2021;127(20):3772-3781. doi:10.1002/cncr.33689
34. Thijssen R, Diepstraten ST, Moujalled D, et al. Intact TP-53 function is essential for sustaining durable responses to BH3-mimetic drugs in leukemias. *Blood.* 2021;137(20):2721-2735. doi:10.1182/blood.2020010167
35. Nechiporuk T, Kurtz SE, Nikolova O, et al. The TP53 Apoptotic Network Is a Primary Mediator of Resistance to BCL2 Inhibition in AML Cells. *Cancer Discov.* 2019;9(7):910-925. doi:10.1158/2159-8290.CD-19-0125
36. Chen SS, Sun Q, Cao L, et al. Efficacy and safety of decitabine combined with low-dose cytarabine, aclarubicin, and granulocyte colony-stimulating factor compared with standard therapy in acute myeloid leukemia patients with TP53 mutation. *Chin Med J (Engl).* 2020;134(12):1477-1479. doi:10.1097/CM9.0000000000001316
37. Li J, Chen Y, Zhu Y, et al. Efficacy and safety of decitabine in combination with G-CSF, low-dose cytarabine and aclarubicin in newly diagnosed elderly patients with acute myeloid leukemia. *Oncotarget.* 2015;6(8):6448-6458. doi:10.18632/oncotarget.3361
38. Kadia TM, Faderl S, Ravandi F, et al. Final results of a phase 2 trial of clofarabine and low-dose cytarabine alternating with decitabine in older patients with newly diagnosed acute myeloid leukemia. *Cancer.* 2015;121(14):2375-2382. doi:10.1002/cncr.29367
39. Kadia TM, Cortes J, Ravandi F, et al. Cladribine and low-dose cytarabine alternating with decitabine as front-line therapy for elderly patients with acute myeloid leukaemia: a phase 2 single-arm trial. *Lancet Haematol.* 2018;5(9):e411-e421. doi:10.1016/S2352-3026(18)30132-7
40. Kadia TM, Ravandi F, Borthakur G, et al. Long-term results of low-intensity chemotherapy with clofarabine or cladribine combined with low-dose cytarabine alternating with decitabine in older patients with newly diagnosed acute myeloid leukemia. *Am J Hematol.* 2021;96(8):914-924. doi:10.1002/ajh.26206
41. Cahill KE, Karimi YH, Karrison TG, et al. A phase 1 study of azacitidine with high-dose cytarabine and mitoxantrone in high-risk acute myeloid leukemia. *Blood Adv.* 2020;4(4):599-606. doi:10.1182/bloodadvances.2019000795
42. Zhang B, Pei Z, Wang H, et al. Chidamide and Decitabine in Combination with a HAG Priming Regimen for Acute Myeloid Leukemia with TP53 Mutation. *Acta Med Okayama.* 2022;76(1):63-70. doi:10.18926/AMO/63213
43. Chiche E, Rahmé R, Bertoli S, et al. Real-life experience with CPX-351 and impact on the outcome of high-risk AML patients: a multicentric French cohort. *Blood Adv.* 2021;5(1):176-184. doi:10.1182/bloodadvances.2020003159
44. Aaron D Goldberg, Chetasi Talati, Pinkal Desai et al. TP53 Mutations Predict Poorer Responses to CPX-351 in Acute Myeloid Leukemia. *Blood* 2018; 132 (Supplement 1): 1433.
45. Cortes JE, Lin TL, Asubonteng K, Faderl S, Lancet JE, Prebet T. Efficacy and safety of CPX-351 versus 7 + 3 chemotherapy by European LeukemiaNet 2017 risk subgroups in older adults with newly diagnosed, high-risk/secondary AML: post hoc analysis of a randomized, phase 3 trial. *J Hematol Oncol.* 2022;15(1):155. doi:10.1186/s13045-022-01361-w
46. Rautenberg C, Stölzel F, Röhlig C, et al. Real-world experience of CPX-351 as first-line treatment for patients with acute myeloid leukemia. *Blood Cancer J.* 2021;11(10):164. doi:10.1038/s41408-021-00558-5
47. Swords RT, Coutre S, Maris MB, et al. Pevonedistat, a first-in-class NEDD8-activating enzyme inhibitor, combined with azacitidine in patients with AML. *Blood.* 2018;131(13):1415-1424. doi:10.1182/blood-2017-09-805895
48. Saliba AN, Kaufmann SH, Stein EM, et al. Pevonedistat with azacitidine in older patients with TP53-mutated AML: a phase 2 study with laboratory correlates. *Blood Adv.* 2023;7(11):2360-2363. doi:10.1182/bloodadvances.2022008625
49. Short NJ, Muftuoglu M, Ong F, et al. A phase 1/2 study of azacitidine, venetoclax and pevonedistat in newly diagnosed secondary AML and in MDS or CMML after failure of hypomethylating agents. *J Hematol Oncol.* 2023;16(1):73. doi:10.1186/s13045-023-01476-8

50. Rushworth SA, Murray MY, Zaitseva L, et al. Identification of Bruton's tyrosine kinase as a therapeutic target in acute myeloid leukemia. *Blood*. 2014;123(8):1229-1238. doi:10.1182/blood-2013-06-511154
51. Huls G, Chitu DA, Pabst T, et al. Ibrutinib added to 10-day decitabine for older patients with AML and higher risk MDS. *Blood Adv*. 2020;4(18):4267-4277. doi:10.1182/bloodadvances.2020002846
52. Roboz GJ, Mandrekar SJ, Desai P, et al. Randomized trial of 10 days of decitabine ± bortezomib in untreated older patients with AML: CALGB 11002 (Alliance). *Blood Adv*. 2018;2(24):3608-3617. doi:10.1182/bloodadvances.2018023689
53. Zhang Q, Bykov VJN, Wiman KG, et al. APR-246 reactivates mutant p53 by targeting cysteines 124 and 277 [published correction appears in *Cell Death Dis*. 2019 Oct 10;10(10):769]. *Cell Death Dis*. 2018;9(5):439. doi:10.1038/s41419-018-0463-7
54. Ali D, Jönsson-Videsäter K, Deneberg S, et al. APR-246 exhibits anti-leukemic activity and synergism with conventional chemotherapeutic drugs in acute myeloid leukemia cells. *Eur J Haematol*. 2011;86(3):206-215. doi:10.1111/j.1600-0609.2010.01557.x
55. Maslah N, Salomao N, Drevon L, et al. Synergistic effects of PRIMA-1Met (APR-246) and 5-azacitidine in TP53-mutated myelodysplastic syndromes and acute myeloid leukemia. *Haematologica*. 2020;105(6):1539-1551. doi:10.3324/haematol.2019.218453
56. Fujihara KM, Zhang BZ, Jackson TD, et al. Eprenetapopt triggers ferroptosis, inhibits NFS1 cysteine desulfurase, and synergizes with serine and glycine dietary restriction. *Sci Adv*. 2022;8(37):eabm9427. doi:10.1126/sciadv.abm9427
57. Birsén R, Larrue C, Decroocq J, et al. APR-246 induces early cell death by ferroptosis in acute myeloid leukemia. *Haematologica*. 2022;107(2):403-416. doi:10.3324/haematol.2020.259531
58. Sallman DA, DeZern AE, Garcia-Manero G, et al. Eprenetapopt (APR-246) and Azacitidine in TP53-Mutant Myelodysplastic Syndromes. *J Clin Oncol*. 2021;39(14):1584-1594. doi:10.1200/JCO.20.02341
59. Cluzeau T, Sebert M, Rahmé R, et al. Eprenetapopt Plus Azacitidine in TP53-Mutated Myelodysplastic Syndromes and Acute Myeloid Leukemia: A Phase II Study by the Groupe Francophone des Myélodysplasies (GFM). *J Clin Oncol*. 2021;39(14):1575-1583. doi:10.1200/JCO.20.02342
60. Mishra A, Tamari R, DeZern AE, et al. Eprenetapopt Plus Azacitidine After Allogeneic Hematopoietic Stem-Cell Transplantation for TP53-Mutant Acute Myeloid Leukemia and Myelodysplastic Syndromes. *J Clin Oncol*. 2022;40(34):3985-3993. doi:10.1200/JCO.22.00181
61. Garcia-Manero G, Goldberg AD, Winer ES, et al. Eprenetapopt combined with venetoclax and azacitidine in TP53-mutated acute myeloid leukaemia: a phase 1, dose-finding and expansion study. *Lancet Haematol*. 2023;10(4):e272-e283. doi:10.1016/S2352-3026(22)00403-3
62. Jaiswal S, Jamieson CH, Pang WW, et al. CD47 is upregulated on circulating hematopoietic stem cells and leukemia cells to avoid phagocytosis. *Cell*. 2009;138(2):271-285. doi:10.1016/j.cell.2009.05.046
63. Majeti R, Chao MP, Alizadeh AA, et al. CD47 is an adverse prognostic factor and therapeutic antibody target on human acute myeloid leukemia stem cells. *Cell*. 2009;138(2):286-299. doi:10.1016/j.cell.2009.05.045
64. Liu J, Wang L, Zhao F, et al. Pre-Clinical Development of a Humanized Anti-CD47 Antibody with Anti-Cancer Therapeutic Potential. *PLoS One*. 2015;10(9):e0137345. doi:10.1371/journal.pone.0137345
65. Daver N, Vyas P, Kambhampati S, et al. Tolerability and Efficacy of the Anticlust of Differentiation 47 Antibody Magrolimab Combined With Azacitidine in Patients With Previously Untreated AML: Phase Ib Results. *J Clin Oncol*. 2023;41(31):4893-4904. doi:10.1200/JCO.22.02604
66. Daver N, Senapati J, Maiti A, et al. Phase I/II Study of Azacitidine (AZA) with Venetoclax (VEN) and Magrolimab (Magro) in Patients (pts) with Newly Diagnosed (ND) Older/Unfit or High-Risk Acute Myeloid Leukemia (AML) and Relapsed/Refractory (R/R) AML. *Blood* 2022; 140 (Supplement 1): 141-144
67. Gilead statement on the discontinuation of magrolimab study in AML with TP53 mutations. News release. September 26, 2023. Available online: <https://www.gilead.com/news-and-press/company-statements/gilead-statement-on-the-discontinuation-of-magrolimab-study-in-aml-with-tp53-mutations>. Accessed December 04, 2023
68. Petrova PS, Viller NN, Wong M, et al. TTI-621 (SIRPαFc): A CD47-Blocking Innate Immune Checkpoint Inhibitor with Broad Antitumor Activity and Minimal Erythrocyte Binding. *Clin Cancer Res*. 2017;23(4):1068-1079. doi:10.1158/1078-0432.CCR-16-1700
69. Miao M, Qingliang T, Depei W, et al. AK117 (anti-CD47 monoclonal antibody) in Combination with Azacitidine for Newly Diagnosed Higher Risk Myelodysplastic Syndrome (HR-MDS): AK117-103 Phase 1b Results. *Blood* 2023; 142 (Supplement 1): 1865.

70. Chen A, Harrabi O, Fong AP, et al. Alx148 enhances the depth and durability of response to multiple aml therapies. *Blood* 2020, 136, 15–16
71. ALX Oncology Reports Second Quarter 2023 Financial Results and Provides Clinical Program Update. August 10, 2023. Available online: <https://ir.alxoncology.com/news-releases/news-release-details/alx-oncology-reports-second-quarter-2023-financial-results-and/>. Accessed December 05, 2023
72. Asayama T, Tamura H, Ishibashi M, et al. Functional expression of Tim-3 on blasts and clinical impact of its ligand galectin-9 in myelodysplastic syndromes. *Oncotarget*. 2017;8(51):88904-88917. doi:10.18632/oncotarget.21492
73. Kikushige Y, Miyamoto T, Yuda J, et al. A TIM-3/Gal-9 Autocrine Stimulatory Loop Drives Self-Renewal of Human Myeloid Leukemia Stem Cells and Leukemic Progression. *Cell Stem Cell*. 2015;17(3):341-352. doi:10.1016/j.stem.2015.07.011
74. Brunner AM, Esteve J, Porkka K, et al. Efficacy and safety of sabatolimab (MBG453) in combination with hypomethylating agents (HMAs) in patients (Pts) with very high/high-risk myelodysplastic syndrome (vHR/HR-MDS) and acute myeloid leukemia (AML): final analysis from a phase Ib study. *Blood* 2021;138:244
75. Al-Hussaini M, Rettig MP, Ritchey JK, et al. Targeting CD123 in acute myeloid leukemia using a T-cell-directed dual-affinity retargeting platform. *Blood*. 2016;127(1):122-131. doi:10.1182/blood-2014-05-575704
76. Testa U, Riccioni R, Miliati S, et al. Elevated expression of IL-3Ralpha in acute myelogenous leukemia is associated with enhanced blast proliferation, increased cellularity, and poor prognosis. *Blood*. 2002;100(8):2980-2988. doi:10.1182/blood-2002-03-0852
77. Uy GL, Aldoss I, Foster MC, et al. Flotetuzumab as salvage immunotherapy for refractory acute myeloid leukemia. *Blood*. 2021;137(6):751-762. doi:10.1182/blood.2020007732
78. Vadakekolathu J, Lai C, Reeder S, et al. TP53 abnormalities correlate with immune infiltration and associate with response to flotetuzumab immunotherapy in AML. *Blood Adv*. 2020;4(20):5011-5024. doi:10.1182/bloodadvances.2020002512
79. Daver N, Montesinos P, Aribi A, et al. Broad Activity for the Pivekimab Sunirine (PVEK, IMGN632), Azacitidine, and Venetoclax Triplet in High-Risk Patients with Relapsed/Refractory Acute Myeloid Leukemia (AML). *Blood* 2022; 140 (Supplement 1): 145–149.
80. Daver N, Montesinos P, Montesinos P, Aribi A, et al. A phase 1b/2 study of pivekimab sunirine (PVEK, IMGN632) in combination with venetoclax/azacitidine or magrolimab for patients with CD123-positive acute myeloid leukemia (AML). *Journal of Clinical Oncology* 2023 41:16_suppl, TPS7073
81. Daver N, Montesinos P, Altman J, et al. Pivekimab Sunirine (PVEK, IMGN632), a CD123-Targeting Antibody-Drug Conjugate, in Combination with Azacitidine and Venetoclax in Patients with Newly Diagnosed Acute Myeloid Leukemia. *Blood* 2023; 142 (Supplement 1): 2906.
82. Frankel A, Liu J.S, Rizzieri D, et al. Phase I clinical study of diphtheria toxin-interleukin 3 fusion protein in patients with acute myeloid leukemia and myelodysplasia. *Leuk. Lymphoma* 2008;49(3):543-553. doi:10.1080/10428190701799035
83. Togami K, Pastika T, Stephansky J, et al. DNA methyltransferase inhibition overcomes diphthamide pathway deficiencies underlying CD123-targeted treatment resistance. *J Clin Invest*. 2019;129(11):5005-5019. doi:10.1172/JCI128571
84. Lane A, Garcia J, Raulston E, et al. Tagraxofusp in Combination with Azacitidine and Venetoclax in Newly Diagnosed CD123+ Acute Myeloid Leukemia, Expansion Cohort of a Phase 1b Multicenter Trial. *Blood* 2023; Volume 142, Supplement 1, 2023, p. 4277 <https://doi.org/10.1182/blood-2023-180010>
85. Ravandi F, Bashey A, Foran J, et al. Phase 1 study of vibecotamab identifies an optimized dose for treatment of relapsed/refractory acute myeloid leukemia. *Blood Adv*. 2023;7(21):6492-6505. doi:10.1182/bloodadvances.2023010956
86. Pelosi E, Castelli G, Testa U. CD123 a Therapeutic Target for Acute Myeloid Leukemia and Blastic Plasmacytoid Dendritic Neoplasm. *Int J Mol Sci*. 2023;24(3):2718. doi:10.3390/ijms24032718
87. Davids MS, Kim HT, Bachireddy P, et al. Ipilimumab for Patients with Relapse after Allogeneic Transplantation. *N Engl J Med*. 2016;375(2):143-153. doi:10.1056/NEJMoa1601202
88. Ravandi F, Assi R, Daver N, et al. Idarubicin, cytarabine, and nivolumab in patients with newly diagnosed acute myeloid leukaemia or high-risk myelodysplastic syndrome: a single-arm, phase 2 study. *Lancet Haematol*. 2019;6(9):e480-e488. doi:10.1016/S2352-3026(19)30114-0

89. Daver N, Garcia-Manero G, Basu S, et al. Efficacy, Safety, and Biomarkers of Response to Azacitidine and Nivolumab in Relapsed/Refractory Acute Myeloid Leukemia: A Nonrandomized, Open-Label, Phase II Study. *Cancer Discov.* 2019;9(3):370-383. doi:10.1158/2159-8290.CD-18-0774
90. Zeidner JF, Vincent BG, Ivanova A, et al. Phase II Trial of Pembrolizumab after High-Dose Cytarabine in Relapsed/Refractory Acute Myeloid Leukemia. *Blood Cancer Discov.* 2021;2(6):616-629. doi:10.1158/2643-3230.BCD-21-0070
91. Zeidan AM, Boss I, Beach CL, et al. A randomized phase 2 trial of azacitidine with or without durvalumab as first-line therapy for older patients with AML. *Blood Adv.* 2022;6(7):2219-2229. doi:10.1182/bloodadvances.2021006138
92. Lovewell RR, Hong J, Kundu S, et al. LAIR-1 agonism as a therapy for acute myeloid leukemia. *J Clin Invest.* 2023;133(22):e169519. doi:10.1172/JCI169519
93. Tovar C, Rosinski J, Filipovic Z, et al. Small-molecule MDM2 antagonists reveal aberrant p53 signaling in cancer: implications for therapy. *Proc Natl Acad Sci U S A.* 2006;103(6):1888-1893. doi:10.1073/pnas.0507493103
94. Yee K, Papayannidis C, Vey N, et al. Murine double minute 2 inhibition alone or with cytarabine in acute myeloid leukemia: Results from an idasanutlin phase 1/1b study small star, filled. *Leuk. Res.* 2021, 100, 106489
95. Konopleva MY, Röhlig C, Cavenagh J, et al. Idasanutlin plus cytarabine in relapsed or refractory acute myeloid leukemia: results of the MIRROS trial. *Blood Adv.* 2022;6(14):4147-4156. doi:10.1182/bloodadvances.2021006303
96. Pan R, Ruvolo V, Mu H, et al. Synthetic Lethality of Combined Bcl-2 Inhibition and p53 Activation in AML: Mechanisms and Superior Antileukemic Efficacy. *Cancer Cell.* 2017;32(6):748-760.e6. doi:10.1016/j.ccell.2017.11.003
97. Senapati J, Muftuoglu M, Ishizawa J, et al. A Phase I study of Milademetan (DS3032b) in combination with low dose cytarabine with or without venetoclax in acute myeloid leukemia: Clinical safety, efficacy, and correlative analysis. *Blood Cancer J.* 2023;13(1):101. doi:10.1038/s41408-023-00871-1
98. Daver N, Dail M, Garcia JS, et al. Venetoclax and idasanutlin in relapsed/refractory AML: a nonrandomized, open-label phase 1b trial. *Blood.* 2023;141(11):1265-1276. doi:10.1182/blood.2022016362
99. Chen S, Wu JL, Liang Y, et al. Arsenic Trioxide Rescues Structural p53 Mutations through a Cryptic Allosteric Site. *Cancer Cell.* 2021;39(2):225-239.e8. doi:10.1016/j.ccell.2020.11.013
100. Parrales A, Ranjan A, Iyer SV, et al. DNAJA1 controls the fate of misfolded mutant p53 through the mevalonate pathway. *Nat Cell Biol.* 2016;18(11):1233-1243. doi:10.1038/ncb3427
101. Eisenmann ED, Stromatt JC, Fobare S, et al. TP-0903 Is Active in Preclinical Models of Acute Myeloid Leukemia with TP53 Mutation/Deletion. *Cancers (Basel).* 2022;15(1):29. doi:10.3390/cancers15010029
102. Semba Y, Yamauchi T, Nakao F, et al. CRISPR-Cas9 Screen Identifies XPO7 As a Potential Therapeutic Target for TP53-Mutated AML. *Blood* 2019; 134 (Supplement_1): 3784.
103. Kravchenko JE, Ilyinskaya GV, Komarov PG, et al. Small-molecule RETRA suppresses mutant p53-bearing cancer cells through a p73-dependent salvage pathway. *Proc Natl Acad Sci U S A.* 2008;105(17):6302-6307. doi:10.1073/pnas.0802091105
104. Liu Y, Chen C, Xu Z, et al. Deletions linked to TP53 loss drive cancer through p53-independent mechanisms. *Nature.* 2016;531(7595):471-475. doi:10.1038/nature17157
105. Boeckler FM, Joerger AC, Jaggi G, Rutherford TJ, Veprintsev DB, Fersht AR. Targeted rescue of a destabilized mutant of p53 by an in silico screened drug. *Proc Natl Acad Sci U S A.* 2008;105(30):10360-10365. doi:10.1073/pnas.0805326105
106. Blanden AR, Yu X, Loh SN, Levine AJ, Carpizo DR. Reactivating mutant p53 using small molecules as zinc metallochaperones: awakening a sleeping giant in cancer [published correction appears in *Drug Discov Today.* 2016 Oct;21(10):1728]. *Drug Discov Today.* 2015;20(11):1391-1397. doi:10.1016/j.drudis.2015.07.006
107. Gao N, Budhreja A, Cheng S, et al. Phenethyl isothiocyanate exhibits antileukemic activity in vitro and in vivo by inactivation of Akt and activation of JNK pathways. *Cell Death Dis.* 2011;2(4):e140. doi:10.1038/cddis.2011.22
108. Carter BZ, Mak PY, Muftuoglu M, et al. Epichaperome inhibition targets TP53-mutant AML and AML stem/progenitor cells. *Blood.* 2023;142(12):1056-1070. doi: 10.1182/blood.2022019047.

109. Lindemann A, Patel AA, Silver NL, et al. COTI-2, A Novel Thiosemicarbazone Derivative, Exhibits Antitumor Activity in HNSCC through p53-dependent and -independent Mechanisms. *Clin Cancer Res.* 2019;25(18):5650-5662. doi:10.1158/1078-0432.CCR-19-0096
110. Bosc C, Saland E, Bousard A, et al. Mitochondrial inhibitors circumvent adaptive resistance to venetoclax and cytarabine combination therapy in acute myeloid leukemia. *Nat Cancer.* 2021;2(11):1204-1223. doi:10.1038/s43018-021-00264-y

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.