

Review

Not peer-reviewed version

Benefits and caveats in the use of retinal pigment epithelium-specific Cre mice.

[Sai Kocherlakota](#) and [Myriam Baes](#) *

Posted Date: 20 December 2023

doi: [10.20944/preprints202312.1531.v1](https://doi.org/10.20944/preprints202312.1531.v1)

Keywords: Retinal pigment epithelium; knockout; mouse; Cre mice



Preprints.org is a free multidiscipline platform providing preprint service that is dedicated to making early versions of research outputs permanently available and citable. Preprints posted at Preprints.org appear in Web of Science, Crossref, Google Scholar, Scilit, Europe PMC.

Copyright: This is an open access article distributed under the Creative Commons Attribution License which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Disclaimer/Publisher's Note: The statements, opinions, and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions, or products referred to in the content.

Review

Benefits and Caveats in the Use of Retinal Pigment Epithelium-Specific Cre Mice

Sai Kocherlakota ¹ and Myriam Baes ^{1,*}

¹ Laboratory of Cell Metabolism, Department of Pharmaceutical and Pharmacological Sciences, KU Leuven, 3000 Leuven, Belgium.

* Correspondence: myriam.baes@kuleuven.be

Abstract: The retinal pigment epithelium (RPE) is an important monolayer of cells present in the outer retina forming a major part of the blood-retina-barrier (BRB). It performs many essential tasks for the maintenance of retinal integrity and function. With increasing knowledge of the retina, it is getting clearer that both common retinal disorders, like age related macular degeneration, and rare genetic disorders originate in the RPE. This calls for a better understanding of the functions of various proteins within the RPE. In this regard, mice enabling an RPE-specific gene deletion are a powerful tool to study the role of a particular protein within the RPE cells in their native environment, simultaneously negating any potential influences of systemic changes. Moreover, since the RPE cells interact closely with the adjacent photoreceptors, these mice also provide an excellent avenue to study the importance of a particular gene function within the RPE to the retina as a whole. In this review, we outline and compare the features of various Cre mice created for this purpose, allowing researchers a well-informed decision on the choice of Cre mouse to use in relation to their research needs.

Keywords: retinal pigment epithelium; knockout; mouse; Cre mice

1. Introduction

The retina can be broadly divided into two major components: the neural retina, consisting of neuronal cells like photoreceptors, interneurons, glial cells, astrocytes and ganglion cells, and a non-neuronal retinal pigment epithelium (RPE). The RPE is a monolayer of post-mitotic hexagonally-shaped pigmented cells, interconnected by tight junctions [1]. It forms an integral part of the blood-retina-barrier (BRB), regulating the transport of nutrients and ions in and out of the underlying neural retina [1–3].

While it is the neural retina that is responsible for the generation and propagation of the nerve signal in response to a light stimulus, the RPE plays an essential role in the maintenance of the structural and functional integrity of the neural retina [1,2]. This involves carrying out varied functions including the daily phagocytosis and recycling of damaged photoreceptor outer segments and the regeneration of visual chromophore 11-cis-retinal required for phototransduction [2]. Consequently, any defects in the RPE often result in degeneration of the underlying neural retina [4–6]. Indeed, the sporadically occurring age-related macular degeneration (AMD), the leading cause of blindness among the elderly in developed nations [7], and many less common inherited retinal disorders such as Stargardt's disease and Leber congenital amaurosis, are thought to originate in the RPE [8,9]. Moreover, the RPE is a highly versatile layer, owing to its diverse functions. To carry them out most efficiently, it has evolved unique adaptations, which remain poorly studied. Therefore, it becomes increasingly important that the cell biology of the RPE and the role of various genes within these cells is well understood.

RPE biology can be studied both by *in vitro* and *in vivo* approaches. Although various *in vitro* models including primary RPE cells and differentiated pluripotent stem cells have successfully been used to reveal deregulations upon genetic mutations, it remains necessary to investigate RPE cells in their native environment [1,2,6,10]. To this end, the RPE has been studied in many mouse models with gene knockouts. However, both the intricate interplay between photoreceptors and the RPE [1,2,10], and influences on the RPE due to the systemic loss of the gene, hinder the elucidation of the

role of a particular protein in the RPE. Therefore, mice with an RPE-specific knockout of a gene of interest are required.

Studying RPE-specific knockout mice has additional advantages. Firstly, they are useful when studying the role of a gene that is lethal when knocked out globally as it may be essential for the development of the organism [11], such as the various peroxisome biogenesis genes [12] and autophagy related genes [13]. Secondly, the effect of a gene knockout specifically in the RPE, on the rest of the retina can be studied. Finally, some of these RPE-specific knockout mice allow for an inducible knockout of the gene of interest, enabling a researcher to study the importance of the gene at various stages in the life cycle of a mouse.

All RPE-specific gene knockout mice to date employ the famed Cre-loxP system. In this review, the various Cre mice available today that target the RPE will be discussed, with their benefits and caveats. These mice will be compared with regard to features like the specificity, timing, inducibility and potential toxicity of Cre expression, allowing researchers to make a well-informed decision about the right Cre mice to use in order to answer their specific research question.

2. Regulation of the Cre-loxP system

In the Cre-loxP system, the promotor driving the expression of the Cre recombinase is the crucial factor determining the spatial and temporal inactivation of the gene of interest. Another level of control on the Cre recombinase expression or function can be achieved with the use of inducible Cre-loxP systems, allowing for a temporally regulated knockout of the gene of interest. This is most commonly done using either the tamoxifen-inducible system or the tetracycline- or doxycycline-inducible system [14].

The tamoxifen inducible system involves the use of a modified Cre recombinase, which includes its fusion with the estrogen receptor containing a mutated ligand binding site [14], expressed under the control of a promoter expressed specifically in the RPE cells (Fig. 1A). This fused Cre protein is called CreERT and is normally sequestered in the cytoplasm due to its binding with the heat shock protein 90 (HSP90). Upon binding with synthetic steroids such as tamoxifen or its metabolically active form 4-hydroxytamoxifen (4-OHT), this interaction is severed and the CreERT translocates into the nucleus, acting on the LoxP sites, thereby knocking out the gene of interest. The efficiency of the system was later on improved with the development of a modified version of the CreERT, called CreERT², which is ~10 times more sensitive to tamoxifen or 4-OHT.

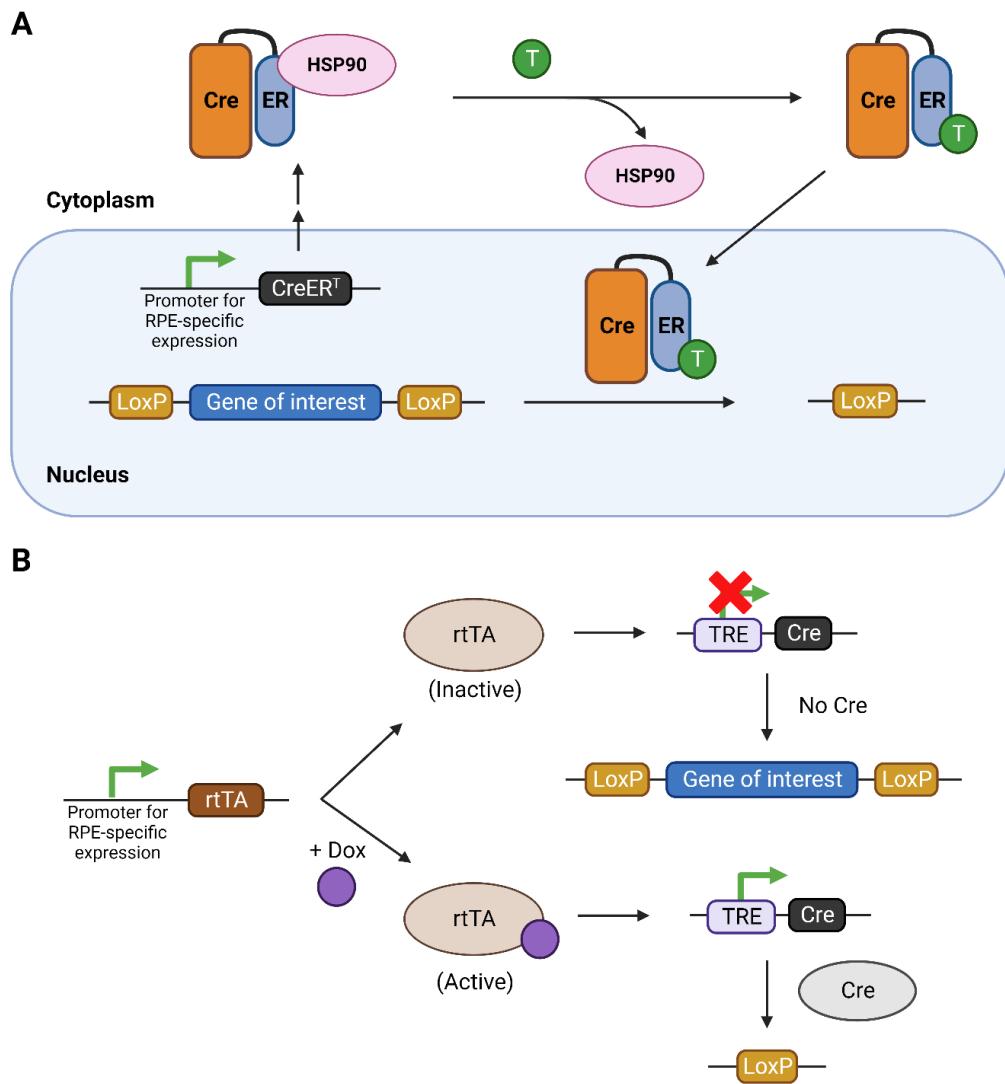


Figure 1. The inducible Cre-LoxP systems used for temporal regulation of knockout of the gene of interest. **A)** Tamoxifen-inducible Cre-LoxP system. **B)** Tetracycline- or doxycycline-inducible Cre-LoxP system. Figure created using *Biorender.com*.

The tetracycline-inducible system involves the use of a reverse tetracycline-controlled transactivator (rtTA) protein, expressed specifically in the RPE cells, and a tetracycline responsive element (TRE) in the genome, which controls the expression of the Cre recombinase (Fig. 1B) [14]. Only upon binding with tetracycline, or its more efficient and cost-effective analogue doxycycline (dox), does the rtTA get activated. This activation leads to its binding to the TRE, consequently activating the expression of Cre recombinase.

The tamoxifen system is generally preferred owing to its higher efficiency among the inducible systems. Moreover, there may be trace amounts of tetracycline in the mouse chow, leading to unintended “leaky” expression of Cre recombinase. However, tamoxifen can have many undesirable side effects owing to its own biological activity [15,16]. Since it modulates the estrogen receptor, it may be advised to thoroughly study sex-based differences in the induced mice, before making final conclusions. Furthermore, there are concerns on safety of its use as prenatal exposure to tamoxifen is known to cause developmental defects in both humans [17,18] and in mice [19,20]. Therefore, care should be taken with the dosage used to prevent any unwanted defects interfering with the phenotype to be studied, and to limit any exposure of tamoxifen to pregnant women involved in studies using these mice. The doxycycline-inducible system is relatively safer. It is to be noted that

both these systems are to some extent susceptible to inducer-independent “leaky” Cre recombinase activity [21,22] with the tetracycline-inducible system being more susceptible to it. Therefore, the Cre expression in these mice should be thoroughly validated before studies on the downstream effects of the gene knockout.

The expression of Cre recombinase specifically in the RPE of floxed mice can also be achieved by sub-retinal injection of viral vectors like lentiviruses, containing expression elements for the enzyme in their genome. The specificity to the RPE is achieved due to the presence of the outer limiting membrane [23]. A similar injection technique has also been used to generate a CRISPR-based knockout of a gene of interest [24], where the gene sequences encoding both the Cas9 enzyme and the guide RNA (gRNA), are delivered using the viral vector. The main advantage of using such injection methods is that each researcher can generate a custom strategy to knockout any specific gene according to their research interests, which is relatively easier than generating new Cre mice. However, the use of this technique is limited by the requirement of expertise for sub-retinal injections, high resource and time investment owing to the high number of mice to be injected, and relatively higher levels of variability. Since the strategies involving these techniques can be tailor-made for the needs of a researcher, this review will not discuss them further.

3. Non-inducible Cre mice targeting the RPE

A summary of the various Cre mice discussed here can be found in table 1. The selectivity and/or efficacy of the Cre mice can be monitored in 2 ways: 1) evaluating the expression levels of Cre protein by immunostaining or immunoblotting, or of Cre mRNA by (RT-q)PCR; 2) evaluating the function of the Cre enzyme by crossing the Cre mice with reporter mice like mT/mG or Rosa-lacZ [25–28], or by monitoring the knockout of the protein of interest.

Table 1. RPE-selective Cre expressing mice without an inducible Cre system.

Model	Promoter	Cre expression start	Important features	References
<i>Trp1-Cre</i>	Tyrosinase-related protein 1 (<i>Trp1</i>) promoter	E10.5	- Untargeted insertion of Cre - Cre toxicity to RPE - Ectopic Cre expression in some cells of neural retina, along with some other tissues	[29,30]
<i>Dct-Cre</i>	Dopachrome tautamerase (<i>Dct</i>) promoter	E9.5	- Untargeted insertion of Cre - Mosaic Cre expression - Cre expression also in melanocytes and in cells of telencephalon* - Ectopic Cre expression in caudal nerves and dorsal root ganglia	[31]
<i>MART-1-Cre</i>	Melanoma associated antigen recognized by T-cells (<i>MART-1</i>) promoter	E12.5	- Untargeted insertion of Cre - Uniform Cre expression - Cre expression also in all melanocytes* - Minimal ectopic Cre expression in some epidermal cells of the skin	[32]
<i>Best1-Cre</i>	Bestrophin-1 (<i>Best1</i>) promoter	P10	- Untargeted insertion of Cre - Mosaic Cre expression - Age and dosage-dependent Cre toxicity to RPE\$ - Cre expression also in testis*	[33,34]

* Cre expression in these cells is expected based on the promoter expression pattern. \$ Contradictory data on Cre toxicity were reported.

Tyrosinase gene family promoters

The first Cre mice with an RPE-specific gene knockout were generated in 2002 [29,31]. These mice used promoters of genes specific to all pigmented cells (melanocytes and RPE) rather than to the RPE itself. These included the promoters of the genes from the tyrosinase gene family, tyrosinase related protein 1 (*Trp1*) [29] and dopachrome tautomerase (*Dct*) [31]. Tyrosinase is the rate limiting enzyme in the synthesis of melanin pigment from tyrosine [35]. The other enzymes in the family, tyrosinase related protein 1 and dopachrome tautomerase stabilize tyrosinase and melanosomes, along with their own functions in melanin synthesis [36,37].

Under the control of the *Trp1* promoter, Cre expression was observed from embryonic day (E10.5) to postnatal day 12 (P12) in the RPE [29]. However, there was ectopic expression in other ocular tissues starting at various stages of development: ciliary margin of retina and optic stalk at E11.5, subset of cells in the ganglion cell layer (GCL) and neuroblastic layer of retina at E14.5, optic nerve, iris and ciliary body at P2-12. There was also ectopic expression in subsets of non-ocular cell types: the mesencephalon, trigeminal nerve ganglion, and dorsal root ganglia.

Interestingly, there was no expression of Cre recombinase in other pigment cells, including those of the choroid. Similarly, a Cre mouse generated using the promoter for tyrosinase (*Tyr*) expressed Cre only in the melanoblasts of the skin, but did not show expression in the RPE [38]. This led to the identification of regulatory elements that specify the expression of tyrosinase family genes *Trp1* and *Tyr* either to the RPE or to other melanocytes [39–41].

The adult *Trp1-Cre* mice showed no Cre expression in any of the tissues, suggesting that the Cre expression is only temporary in these mice. This is inconsequential as Cre mediated genetic recombination is irreversible and therefore, a transient exposure of this recombinase is sufficient to achieve a knockout of the gene of interest. Moreover, the RPE cells are post-mitotic, which means that the cells with the gene of interest knocked out cannot be replaced over the lifetime of the mice. However, despite such a transient expression, in a later study, these RPE have been shown to suffer from Cre-mediated toxicity [30] in the form of reduced pigmentation, loss of RPE characteristic hexagonal shape and reduced thickness. The underlying retina was also affected leading to reduced electroretinogram (ERG) responses and increased infiltration of microglia into the subretinal space. It is likely that this toxicity, despite a transient expression of the Cre recombinase in these cells, arises from the toxic effects of Cre during the development of the RPE cells because RPE morphological defects were already evident at P14 [30].

The *Dct-Cre* mice on the other hand showed Cre function in the optic cup, in dorsal telencephalon and in cells lateral to the neural tube, already at E9.5 [31]. Towards E12.5, the presumptive RPE, migrating melanoblasts and telencephalon showed Cre function, although not all cells of these tissues showed the reporter gene expression. Owing to its expression in melanoblasts, this Cre mouse has been used more in the context of pigment cell studies.

MART-1 promoter

A different strategy, developed a decade later, also targeted the melanocytes by driving the Cre expression using the promoter for the *MART-1* gene (melanoma antigen recognized by T cells 1) also called melan-A (*mlana*) [32]. It is thought to be essential for melanosome biogenesis [42]. These *MART-1-Cre* mice showed Cre expression in the RPE starting at E12.5 and later in the melanocyte precursors at E17.5. There was no ectopic Cre expression detected in these mice, except for some epidermal cells of the skin. No Cre toxicity was reported in these mice.

Bestrophin 1 promoter

A more suitable Cre mouse to study the effects of a gene knockout specifically in the RPE was later generated by making use of the promoter for the *Bestrophin 1* (*Best1*) gene [33], which codes for a calcium activated chloride channel present on cell membranes [43]. It was found that loss of

Bestrophin 1 function led to the Best vitelliform macular dystrophy (BVMD) in humans, which resulted in the gene being also named the *VMD2* gene. In mice, Bestrophin 1 is specifically expressed only in the RPE cells, with the exception of Sertoli cells of the testis [33,44,45]. Concurrently, Cre expression in these mice was detected only in the RPE and the testis. Therefore, it may be prudent not to use the male Cre expressing mice for breeding as their testis may be affected depending on the gene being knocked out.

Cre expression in the RPE of these mice starts from P10, with higher expression observed at P28. This circumvents the potential embryonic toxicity of Cre recombinase observed in the *Trp1-Cre* mice. However, owing to its continuous expression in the RPE cells from P10 onwards, age and dosage-dependent toxicity of Cre recombinase was later reported in the RPE of these mice [34], requiring a thorough examination of Cre-expressing controls. However, there appear to be inconsistencies in literature with regard to such toxicity as several reports were published using these mice where no Cre-mediated RPE toxicity was observed [5,6,46,47].

Similar to the *Dct-Cre* mice, not all RPE cells in these mice express Cre recombinase, leading to what is termed a 'mosaic pattern' of Cre expression, which has also been observed in other Cre mice targeting other cell types [48,49]. It was later surmised to be a side effect of non-targeted integration of the Cre recombinase gene leading to the epigenetic silencing of the repeated copies of the inserted Cre gene [45,50–52]. The extent of RPE Cre expression depends on the strain and the age of the mice. While the C57BL/6 mice expressed Cre in 50-90% of the RPE cells, mice with a mixed B6/129 background showed a consistent expression in 90% of the RPE cells. With respect to age, the percentage of RPE cells expressing Cre increased from ~15% at P10 to almost 70% by P28, peaking at ~90% at 9 weeks (9w) of age. This mosaic expression of Cre recombinase in the RPE cells can be taken advantage of as this can provide an ideal internal control when the Cre-expressing cells are identified. However, it comprises an impediment in biochemical studies like western blotting where individual Cre expressing cells cannot be identified. In such cases, the percentage of RPE cells expressing the Cre recombinase should be known. As it was shown that the recombination efficiency is similar in both eyes of a mouse, this information can be obtained by always using one eye to stain for Cre recombinase in the RPE [33,53], while the other can be used for any other experiment. This, therefore, limits the tissue availability in these mice. Despite these shortcomings, the *Best1-Cre* mice have since been extensively used to study the effects of conditional loss of various genes in the RPE [5,6,46,47,53–56].

4. Inducible Cre mice targeting the RPE

The inducible Cre systems have the advantage of letting the researcher decide when to knock out their gene of interest over the course of a mouse's life span. This can bypass not only any potential developmental defects caused by the removal of the gene of interest during RPE development, but also any potential toxicity of the Cre recombinase function in developing RPE cells, which has been discussed in the previous section. A summary of the various Cre mice discussed in this section can be found in Table 2.

Table 1. RPE-selective Cre expressing mice with an inducible Cre system.

Model	Promoter	Induction by	Important features	References
<i>Inducible VMD2-Cre</i>	Vitelliform macular dystrophy-2 (<i>VMD2</i>), promoter	Tetracycline/Doxycline	- Untargeted insertion of Cre. - "Leaky" Cre expression - Mosaic Cre expression - Cre recombinase undetectable by immunostaining - Weak ectopic Cre expression in the optic nerve	[57–59]

<i>Inducible MCT3-Cre</i>	Monocarboxylate transporter 3 (<i>Mct3</i>) promoter	Tamoxifen	- Untargeted insertion of Cre - Mosaic Cre activity. Only 5-20% of RPE show Cre activity - Cre activity also in the choroid plexus epithelium of brain*	[4]
<i>Inducible Trp1-Cre</i>	Tyrosinase-related protein 1 (<i>Trp1</i>) promoter	Tamoxifen	- Untargeted insertion of Cre. - Mosaic Cre activity - Ectopic Cre activity in some cells of neural retina, iris, ciliary body and optic nerve	[60]
<i>Inducible Tyr-Cre</i>	Tyrosinase (<i>Tyr</i>) promoter	Tamoxifen	- Untargeted insertion of Cre - Mosaic Cre activity with better expression in embryonic RPE - Cre activity also observed in the ciliary body* - Weak ectopic Cre function observed in inner nuclear layer without any cell-type specificity	[61]
<i>Inducible Best1-Cre</i>	Bestrophin-1 (<i>Best1</i>) promoter	Tamoxifen	- Targeted insertion of Cre gene into the Rosa26 locus - Cre function also in the testis* - Cre activity in >90% of RPE cells - Minimal/negligible (<1%) ectopic Cre function in Muller glia	[45]
<i>Inducible RPE65-Cre</i>	Retinal pigment epithelium-specific 65 kDa protein (<i>RPE65</i>) promoter	Tamoxifen	- Targeted knock-in of sequence for P2A-CreERT2 fused in-frame with RPE65 gene - Cre activity in >90% of RPE cells - Levels of Cre recombinase undetectable by immunoblotting	[52]
<i>Inducible Pmel-Cre</i>	Premelanosome protein (<i>Pmel</i>) promoter	Tetracycline/Doxycline	- Untargeted insertion of Cre. - No mosaic Cre expression - Cre expression also in most melanocytes* - Ectopic Cre expression in lung and heart mesothelial cells	[62]

* Cre expression in these cells is expected based on the promoter expression pattern.

Tetracycline/doxycycline-inducible VMD2 promoter

The first Cre mouse facilitating inducible RPE-selective gene knockout was the inducible *VMD2-Cre* model [57], which is regulated by tetracycline. Cre function could be induced as early as E9 until P60, with highest expression observed at P4. This covers the entire span of RPE development and therefore enables studying the importance of a gene during various stages of the process. However, these mice suffer from several drawbacks. Firstly, to achieve the maximum efficiency, they required tetracycline administration through gavage, which can only be done from P3 and needs relevant expertise. Secondly, the expression and function of Cre recombinase is not entirely inducer-dependent, meaning there is “leaky” expression of Cre recombinase observed even when the mice are not administered tetracycline or its analogue, dox. Thirdly, these mice exhibited mosaic Cre expression that was revealed using reporter mice. This particular limitation is especially significant in these mice, as the levels of Cre recombinase could not be detected using immunohistochemical techniques. While these low expression levels could be good to avoid any potential dosage-dependent Cre-induced toxicity to RPE cells, the inability to stain the Cre recombinase precludes

assessing the percentage of Cre positive RPE cells. As pointed out before, the latter is important when using such mosaic Cre expressing RPE for biochemical studies. Finally, these mice also showed some ectopic Cre recombinase function in the optic nerve cells. Despite these shortcomings, the *VMD2-Cre* mice were quite useful at the time, being one of the few models that did not suffer from Cre-induced RPE toxicity, with intact RPE and retinal integrity and function even up to 10 months of age.

To tackle some of the drawbacks of these mice, the authors later evaluated the efficacy of administering dox via intravitreal injections [59]. This allowed for better efficiency of Cre expression and function with ~60% of the RPE showing Cre activity starting from 15 days post injection. No Cre levels were detected at 4 months post injection, which is beneficial in avoiding any potential Cre-induced toxicity. Indeed, the retinal morphology was preserved even until 12 months after dox induction. This also avoided the ectopic expression of Cre recombinase in the optic nerve, which was observed when dox was administered orally. However, certain drawbacks remain with regard to the technical expertise required for intravitreal injections, especially in the neonates, and the fact that this is not possible to perform in embryonic mice.

Tamoxifen-inducible MCT3 promoter

Around the same time, a tamoxifen- inducible RPE-selective gene knockout mouse was generated using the promoter for the *Slc16a8* gene, which codes for the monocarboxylate transporter 3 (MCT3) [4]. The MCT3 protein is responsible for transport of several monocarboxylate substrates including lactate, pyruvate and some ketone bodies, across the cell membrane. It is only expressed in the RPE and in the choroid plexus epithelium of the brain [63,64]. Concurrently, the *MCT3-Cre* mice exhibited Cre function only in these two cell types without any unexpected ectopic expression. In neonates, Cre function was detected in the RPE 7 days post injection of 4-OHT. However, they also suffer from mosaic Cre activity, with only ~20% of the RPE cells exhibiting Cre function, which further fell to ~5% when older mice were induced. Owing to the very low efficiency, these mice were not widely used.

Tamoxifen-inducible Trp1 promoter

Soon after, another inducible Cre mouse was generated using the promoter for the *Trp1* gene paired with the tamoxifen-inducible Cre system. A week after induction with tamoxifen, Cre activity was detected in the RPE in a mosaic pattern, with ~40-80% of the RPE exhibiting Cre mediated recombination. Interestingly, in contrast to the *Trp1-Cre* mice [60], no choroidal or extra-ocular Cre recombinase activity was detected using these inducible *Trp1-Cre* mice. However, there was still some ectopic Cre function detected in some cells of the neural retina, iris, ciliary body and optic nerve.

Tamoxifen-inducible hsp70 promoter with RPE-specific Cns-2 enhancer

Schneider *et al.* generated an inducible RPE-selective Cre mouse by expressing the tamoxifen-inducible Cre-ER^{T2} under the control of the ubiquitous *hsp70 minimal promoter* and the RPE-specific *Tyr enhancer* (*Cns-2*) [61]. The RPE specificity is conferred by the *Tyr enhancer* element *Cns-2*, which was shown to drive tyrosinase expression specifically in the RPE cells [41].

Cre activity was only detected upon induction with 4-OHT in the RPE and to some extent in the ciliary body (~10%). Some ectopic Cre activity was detected in the inner nuclear layer (INL) of the neural retina, but this was not specific to any cell-type in the retina. No extra-ocular Cre activity was detected in these mice. In adult mice, after 5 consecutive days of tamoxifen treatment, ~47-70% of the RPE exhibited Cre recombinase activity, with better efficiency observed in the central RPE compared to the peripheral RPE. Similar to inducible *MCT3-Cre* mice, the efficiency of tamoxifen induction increased when embryonic mice were induced at E9.5 (through the mother) for 5 days, with the percentage of RPE showing Cre activity rising to ~83%. This difference in efficacy was attributed to the possible differences in the effective dosage of tamoxifen to the RPE and a possibly higher efficacy of recombination in the developing RPE as opposed to a fully differentiated adult RPE. Interestingly, some sex-based differences in the efficacy of the tamoxifen administration method (Intraperitoneal

(IP) vs gavage) were also reported with the IP route being more efficient in males than in females. No Cre toxicity was reported even until 3 months post induction. However, this was only assessed in adult mice induced with tamoxifen. Any potential toxicity of Cre expression when induced in embryonic RPE was not evaluated and thus, needs to be thoroughly examined before making conclusions using these mice.

Tamoxifen-inducible Best1 promoter in Rosa26 locus

Since most mice with RPE-selective gene knockout suffer from mosaic Cre activity, Chen *et al.* tackled this by targeting the insertion of *Best1* promoter-controlled transgenic Cre-ERT^{T2} gene into the *Rosa26* locus of C57BL/6J mice [45]. Despite such targeted insertion, there was still some mosaic activity of Cre recombinase, suggesting that the mosaicism could arise from an intrinsic feature of the *Best1* promoter. Cre recombinase activity was most efficiently induced by 4 consecutive daily IP injections of 4-OHT beginning at P14, with no leaky expression observed. While ~90% of RPE cells in males exhibited Cre activity, the percentage was lower in females at ~85%. Robust induction was also observed in adult mice at 7w of age with ~85% percent of RPE showing Cre activity in both the genders. However, this required a double dosage than was used at P14. No Cre activity was detected in any other tissue except for the testis, which is expected based on the expression pattern of *Best1* gene in mice. Minimal/negligible Cre activity was detected in the Müller glia of the retina, which never exceeded 0.6% of these cells. This is not entirely surprising because the *Best1* promoter has been shown to drive Cre expression in the Müller cells before [58]. Moreover, no RPE or retinal toxicity was observed even until 7 months of age in the mice induced at P14. However, the same could not be said regarding the homozygous Cre mice, which show RPE morphological abnormalities at 8 months. The heterozygotes were normal until 2 yrs and therefore, it is judicious to only use these. This dose-dependent difference in Cre toxicity is in line with previous observations seen in the non-inducible *Best1-Cre* mice [33,34]. The consistently high percentage of RPE cells expressing Cre recombinase in these mice eliminates the requirement for the use of one eye to assess the percentage of Cre expressing RPE cells in a given mouse, leaving both the eyes for experiments of choice to the researcher.

Tamoxifen-inducible Cre recombinase gene conjugated to Rpe65 gene

During the time of development of the inducible *Best1-Cre* mice, the generation of another Cre mouse was under works, which largely gets rid of many of the limitations of the previously generated mice. This mouse takes advantage of the expression pattern of the native *Rpe65* gene [52], which encodes one of the most important visual cycle proteins specific to the RPE cells. It catalyzes the conversion of all-*trans*-retinyl esters to 11-*cis*-retinol, which is then converted to regenerate 11-*cis*-retinal, the visual chromophore crucial for vision [2]. Instead of creating a traditional transgenic mouse, the authors knocked-in (KI) a P2A-CreERT^{T2} sequence, fused in-frame after the last coding exon of the native *Rpe65* gene. P2A is a nucleotide sequence that causes efficient ribosome skipping during protein translation [65,66] allowing the production of 2 different proteins from a single transcript: the RPE65-P2A fusion protein, appended on its C-terminal end with non-native 21 amino acids and the Cre-ERT^{T2} protein [52]. Unfortunately, the addition of C-terminal non-native amino acids to the RPE65 protein led to its reduced stability as observed from ~40% reduction in the levels of RPE65 in these mice. This reduction was much more pronounced (~99%) in homozygous KI mice. Therefore, it is again advisable to only use the heterozygous mice. To test the efficacy of RPE65-Cre-ERT^{T2} mice, tamoxifen was administered either by IP injections (5 consecutive days) or via chow (3 weeks) at P21 and P50. Negligible Cre activity was detected in the absence of tamoxifen induction. In the induced mice, ~99% of the RPE cells showed Cre activity, with the non-expressing cells predominantly present in the peripheral retina. No ectopic expression of Cre recombinase was detected. The levels of Cre recombinase were below the detection limit for immunoblotting. Concurrently, there was no Cre induced toxicity detected in the RPE and the retina of these mice even until the age of 4 months. However, in view of the reduced expression of RPE65 protein, the retinal function was tested. This was found to be unaffected at P30, but the rate of 11-*cis*-retinal regeneration

was reduced and all-*trans*-retinyl esters accumulated, especially in mice carrying the less active M450 allele of *Rpe65* gene as compared to the more efficient L450 allele. While this was not significant at P30, any potential changes in the retinyl esters or 11-*cis*-retinal regeneration towards later ages were not reported. However, any such potential changes towards later ages can be presumed to be minimal as no significant differences were observed in the ERG of 4 month-old mice, showing that the retinal function was intact. One limitation with these mice involves the native RPE65 expression, which only starts after P4 in rats, likely being the case in mice as well [67]. Therefore, induction of gene knockout before this time is likely not possible using these mice. Moreover, the authors only induced Cre activity from P21 onwards, by when the RPE is fully developed. This likely circumvents any potential toxicity of Cre function to the developing RPE. However, if the Cre function is to be induced at time points before P21, it is prudent to thoroughly characterize the integrity of the RPE and the retina of these mice.

Tetracycline/doxycycline-inducible Pmel promoter

Most recently, another Cre mouse model was generated using the tetracycline-inducible *Pmel* gene promoter which codes for the premelanosome protein [62] that is important in the early stages of melanosome biogenesis. These mice were generated with the purpose of obtaining an inducible gene-knockout not just specifically in the RPE, but in all the melanocytes. Dox was administered to pregnant mice through drinking water for 3 days at 10.5 days post coitus. The embryos were assessed for Cre activity at E15.5. Cre activity was detected in all the pigment cells as expected from the expression pattern of the *Pmel* gene. There was some ectopic expression in the mesothelial cells of the heart and the lungs, which was independent of exposure to dox ("leaky expression"). Interestingly, the authors did not report mosaic Cre expression in these mice, despite a non-targeted integration approach to insert the transgene. Very little is reported on the RPE of these mice and since they have not yet been used for studies involving RPE-specific gene knockout, the advantages of these mice over others for such a purpose is less clear.

5. Conclusion

The availability of the various Cre mice that enable an RPE-specific gene knockout provides several options for researchers to achieve their research goals. The inevitable venture of CRISPR-Cas9 methodologies, which are faster and more effective, will only expand this portfolio. However, it is important to note that in order to study the intricate molecular pathways and interactions between various macromolecules at the subcellular level, and to study various functional parameters such as the mitochondrial oxygen consumption rate, researchers will need to adopt the usage of *in vitro* models. Nevertheless, the use of RPE-specific gene knockout mice provides an excellent first step towards understanding and narrowing down the cellular pathways a particular gene is involved in, serving as a reference as to what to study and expect when using such *in vitro* models.

Author Contributions: SK and MB searched the literature, wrote the manuscript and made figures.

Funding: M.B. is supported by grants from the KU Leuven (C14/18/088) and from the Research Foundation-Flanders (FWO G0A8619N).

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Lakkaraju, A.; Umapathy, A.; Tan, L.X.; Daniele, L.; Philp, N.J.; Boesze-Battaglia, K.; Williams, D.S. The cell biology of the retinal pigment epithelium. *Prog Retin Eye Res* **2020**, *100846*, doi:10.1016/j.preteyeres.2020.100846.
2. Strauss, O. The retinal pigment epithelium in visual function. *Physiol Rev* **2005**, *85*, 845-881, doi:10.1152/physrev.00021.2004.
3. Yang, S.; Zhou, J.; Li, D. Functions and Diseases of the Retinal Pigment Epithelium. *Front Pharmacol* **2021**, *12*, 727870, doi:10.3389/fphar.2021.727870.

4. Longbottom, R.; Fruttiger, M.; Douglas, R.H.; Martinez-Barbera, J.P.; Greenwood, J.; Moss, S.E. Genetic ablation of retinal pigment epithelial cells reveals the adaptive response of the epithelium and impact on photoreceptors. *Proc Natl Acad Sci U S A* **2009**, *106*, 18728-18733, doi:10.1073/pnas.0902593106.
5. Zhao, C.; Yasumura, D.; Li, X.; Matthes, M.; Lloyd, M.; Nielsen, G.; Ahern, K.; Snyder, M.; Bok, D.; Dunaeif, J.L.; et al. mTOR-mediated dedifferentiation of the retinal pigment epithelium initiates photoreceptor degeneration in mice. *J Clin Invest* **2011**, *121*, 369-383, doi:10.1172/JCI44303.
6. Kocherlakota, S.; Das, Y.; Swinkels, D.; Vanmunster, M.; Callens, M.; Vinckier, S.; Vaz, F.M.; Sinha, D.; Van Veldhoven, P.P.; Fransen, M.; et al. The murine retinal pigment epithelium requires peroxisomal beta-oxidation to maintain lysosomal function and prevent dedifferentiation. *Proc Natl Acad Sci U S A* **2023**, *120*, e2301733120, doi:10.1073/pnas.2301733120.
7. Guymer, R.H.; Campbell, T.G. Age-related macular degeneration. *Lancet* **2023**, *401*, 1459-1472, doi:10.1016/S0140-6736(22)02609-5.
8. Tsang, S.H.; Sharma, T. Leber Congenital Amaurosis. *Adv Exp Med Biol* **2018**, *1085*, 131-137, doi:10.1007/978-3-319-95046-4_26.
9. Lenis, T.L.; Hu, J.; Ng, S.Y.; Jiang, Z.; Sarfare, S.; Lloyd, M.B.; Esposito, N.J.; Samuel, W.; Jaworski, C.; Bok, D.; et al. Expression of ABCA4 in the retinal pigment epithelium and its implications for Stargardt macular degeneration. *Proc Natl Acad Sci U S A* **2018**, *115*, E11120-E11127, doi:10.1073/pnas.1802519115.
10. Hurley, J.B. Retina Metabolism and Metabolism in the Pigmented Epithelium: A Busy Intersection. *Annu Rev Vis Sci* **2021**, *7*, 665-692, doi:10.1146/annurev-vision-100419-115156.
11. Liptak, N.; Gal, Z.; Biro, B.; Hiripi, L.; Hoffmann, O.I. Rescuing lethal phenotypes induced by disruption of genes in mice: a review of novel strategies. *Physiol Res* **2021**, *70*, 3-12, doi:10.33549/physiolres.934543.
12. Baes, M.; Van Veldhoven, P.P. Mouse models for peroxisome biogenesis defects and beta-oxidation enzyme deficiencies. *Biochim Biophys Acta* **2012**, *1822*, 1489-1500, doi:10.1016/j.bbadiis.2012.03.003.
13. Mizushima, N.; Levine, B. Autophagy in mammalian development and differentiation. *Nat Cell Biol* **2010**, *12*, 823-830, doi:10.1038/ncb0910-823.
14. Kim, H.; Kim, M.; Im, S.K.; Fang, S. Mouse Cre-LoxP system: general principles to determine tissue-specific roles of target genes. *Lab Anim Res* **2018**, *34*, 147-159, doi:10.5625/lar.2018.34.4.147.
15. Ye, R.; Wang, Q.A.; Tao, C.; Vishvanath, L.; Shao, M.; McDonald, J.G.; Gupta, R.K.; Scherer, P.E. Impact of tamoxifen on adipocyte lineage tracing: Inducer of adipogenesis and prolonged nuclear translocation of Cre recombinase. *Mol Metab* **2015**, *4*, 771-778, doi:10.1016/j.molmet.2015.08.004.
16. Donocoff, R.S.; Teteloshvili, N.; Chung, H.; Shoulson, R.; Creusot, R.J. Optimization of tamoxifen-induced Cre activity and its effect on immune cell populations. *Sci Rep* **2020**, *10*, 15244, doi:10.1038/s41598-020-72179-0.
17. Tewari, K.; Bonebrake, R.G.; Asrat, T.; Shanberg, A.M. Ambiguous genitalia in infant exposed to tamoxifen in utero. *Lancet* **1997**, *350*, 183, doi:10.1016/S0140-6736(97)24029-8.
18. Braems, G.; Denys, H.; De Wever, O.; Cocquyt, V.; Van den Broecke, R. Use of tamoxifen before and during pregnancy. *Oncologist* **2011**, *16*, 1547-1551, doi:10.1634/theoncologist.2011-0121.
19. Ved, N.; Curran, A.; Ashcroft, F.M.; Sparrow, D.B. Tamoxifen administration in pregnant mice can be deleterious to both mother and embryo. *Lab Anim* **2019**, *53*, 630-633, doi:10.1177/0023677219856918.
20. Sun, M.R.; Steward, A.C.; Sweet, E.A.; Martin, A.A.; Lipinski, R.J. Developmental malformations resulting from high-dose maternal tamoxifen exposure in the mouse. *PLoS One* **2021**, *16*, e0256299, doi:10.1371/journal.pone.0256299.
21. Liu, Y.; Suckale, J.; Masjkur, J.; Magro, M.G.; Steffen, A.; Anastassiadis, K.; Solimena, M. Tamoxifen-independent recombination in the RIP-CreER mouse. *PLoS One* **2010**, *5*, e13533, doi:10.1371/journal.pone.0013533.
22. Lewis, K.T.; Oles, L.R.; MacDougald, O.A. Tetracycline response element driven Cre causes ectopic recombinase activity independent of transactivator element. *Mol Metab* **2022**, *61*, 101501, doi:10.1016/j.molmet.2022.101501.
23. Housset, M.; Samuel, A.; Ettache, M.; Bemelmans, A.; Beby, F.; Billon, N.; Lamonerie, T. Loss of Otx2 in the adult retina disrupts retinal pigment epithelium function, causing photoreceptor degeneration. *J Neurosci* **2013**, *33*, 9890-9904, doi:10.1523/JNEUROSCI.1099-13.2013.
24. Holmgard, A.; Askou, A.L.; Benckendorff, J.N.E.; Thomsen, E.A.; Cai, Y.; Bek, T.; Mikkelsen, J.G.; Corydon, T.J. In Vivo Knockout of the Vegfa Gene by Lentiviral Delivery of CRISPR/Cas9 in Mouse Retinal Pigment Epithelium Cells. *Mol Ther Nucleic Acids* **2017**, *9*, 89-99, doi:10.1016/j.omtn.2017.08.016.
25. Muzumdar, M.D.; Tasic, B.; Miyamichi, K.; Li, L.; Luo, L. A global double-fluorescent Cre reporter mouse. *Genesis* **2007**, *45*, 593-605, doi:10.1002/dvg.20335.
26. Soriano, P. Generalized lacZ expression with the ROSA26 Cre reporter strain. *Nat Genet* **1999**, *21*, 70-71, doi:10.1038/5007.
27. Novak, A.; Guo, C.; Yang, W.; Nagy, A.; Lobe, C.G. Z/EG, a double reporter mouse line that expresses enhanced green fluorescent protein upon Cre-mediated excision. *Genesis* **2000**, *28*, 147-155.

28. Lobe, C.G.; Koop, K.E.; Kreppner, W.; Lomeli, H.; Gertsenstein, M.; Nagy, A. Z/AP, a double reporter for cre-mediated recombination. *Dev Biol* **1999**, *208*, 281-292, doi:10.1006/dbio.1999.9209.
29. Mori, M.; Metzger, D.; Garnier, J.M.; Chambon, P.; Mark, M. Site-specific somatic mutagenesis in the retinal pigment epithelium. *Invest Ophthalmol Vis Sci* **2002**, *43*, 1384-1388.
30. Thanos, A.; Morizane, Y.; Murakami, Y.; Giani, A.; Mantopoulos, D.; Kayama, M.; Roh, M.I.; Michaud, N.; Pawlyk, B.; Sandberg, M.; et al. Evidence for baseline retinal pigment epithelium pathology in the Trp1-Cre mouse. *Am J Pathol* **2012**, *180*, 1917-1927, doi:10.1016/j.ajpath.2012.01.017.
31. Guyonneau, L.; Rossier, A.; Richard, C.; Hummler, E.; Beermann, F. Expression of Cre recombinase in pigment cells. *Pigment Cell Res* **2002**, *15*, 305-309, doi:10.1034/j.1600-0749.2002.02039.x.
32. Aydin, I.T.; Beermann, F. A mart-1::Cre transgenic line induces recombination in melanocytes and retinal pigment epithelium. *Genesis* **2011**, *49*, 403-409, doi:10.1002/dvg.20725.
33. Iacobelli, J.; Zhao, C.; Wolkow, N.; Veldman, P.; Gollomp, K.; Ojha, P.; Lukinova, N.; King, A.; Feiner, L.; Esumi, N.; et al. Generation of Cre transgenic mice with postnatal RPE-specific ocular expression. *Invest Ophthalmol Vis Sci* **2011**, *52*, 1378-1383, doi:10.1167/iovs.10-6347.
34. He, L.; Marioutina, M.; Dunaief, J.L.; Marneros, A.G. Age- and gene-dosage-dependent cre-induced abnormalities in the retinal pigment epithelium. *Am J Pathol* **2014**, *184*, 1660-1667, doi:10.1016/j.ajpath.2014.02.007.
35. Zolghadri, S.; Bahrami, A.; Hassan Khan, M.T.; Munoz-Munoz, J.; Garcia-Molina, F.; Garcia-Canovas, F.; Saboury, A.A. A comprehensive review on tyrosinase inhibitors. *J Enzyme Inhib Med Chem* **2019**, *34*, 279-309, doi:10.1080/14756366.2018.1545767.
36. Korner, A.M.; Pawelek, J. Dopachrome conversion: a possible control point in melanin biosynthesis. *J Invest Dermatol* **1980**, *75*, 192-195, doi:10.1111/1523-1747.ep12522650.
37. Lapedriza, A.; Petratou, K.; Kelsh, R.N. Chapter 14 - Neural Crest Cells and Pigmentation. In *Neural Crest Cells*, Trainor, P.A., Ed.; Academic Press: Boston, 2014; pp. 287-311.
38. Delmas, V.; Martinozzi, S.; Bourgeois, Y.; Holzenberger, M.; Larue, L. Cre-mediated recombination in the skin melanocyte lineage. *Genesis* **2003**, *36*, 73-80, doi:10.1002/gene.10197.
39. Murisier, F.; Guichard, S.; Beermann, F. A conserved transcriptional enhancer that specifies Tyrp1 expression to melanocytes. *Dev Biol* **2006**, *298*, 644-655, doi:10.1016/j.ydbio.2006.05.011.
40. Murisier, F.; Beermann, F. Genetics of pigment cells: lessons from the tyrosinase gene family. *Histol Histopathol* **2006**, *21*, 567-578, doi:10.14670/HH-21.567.
41. Murisier, F.; Guichard, S.; Beermann, F. Distinct distal regulatory elements control tyrosinase expression in melanocytes and the retinal pigment epithelium. *Dev Biol* **2007**, *303*, 838-847, doi:10.1016/j.ydbio.2006.11.038.
42. Hoashi, T.; Watabe, H.; Muller, J.; Yamaguchi, Y.; Vieira, W.D.; Hearing, V.J. MART-1 is required for the function of the melanosomal matrix protein PMEL17/GP100 and the maturation of melanosomes. *J Biol Chem* **2005**, *280*, 14006-14016, doi:10.1074/jbc.M413692200.
43. Kunzelmann, K. TMEM16, LRRC8A, bestrophin: chloride channels controlled by Ca(2+) and cell volume. *Trends Biochem Sci* **2015**, *40*, 535-543, doi:10.1016/j.tibs.2015.07.005.
44. Masuda, T.; Esumi, N. SOX9, through interaction with microphthalmia-associated transcription factor (MITF) and OTX2, regulates BEST1 expression in the retinal pigment epithelium. *J Biol Chem* **2010**, *285*, 26933-26944, doi:10.1074/jbc.M110.130294.
45. Chen, M.; Kim, L.; Lu, C.W.; Zeng, H.; Vollrath, D. An efficient inducible RPE-Selective cre transgenic mouse line. *Exp Eye Res* **2021**, *202*, 108370, doi:10.1016/j.exer.2020.108370.
46. Go, Y.M.; Zhang, J.; Fernandes, J.; Litwin, C.; Chen, R.; Wensel, T.G.; Jones, D.P.; Cai, J.; Chen, Y. mTOR-initiated metabolic switch and degeneration in the retinal pigment epithelium. *FASEB J* **2020**, *34*, 12502-12520, doi:10.1096/fj.202000612R.
47. Huang, J.; Gu, S.; Chen, M.; Zhang, S.J.; Jiang, Z.; Chen, X.; Jiang, C.; Liu, G.; Radu, R.A.; Sun, X.; et al. Abnormal mTORC1 signaling leads to retinal pigment epithelium degeneration. *Theranostics* **2019**, *9*, 1170-1180, doi:10.7150/thno.26281.
48. Heffner, C.S.; Herbert Pratt, C.; Babiuk, R.P.; Sharma, Y.; Rockwood, S.F.; Donahue, L.R.; Eppig, J.T.; Murray, S.A. Supporting conditional mouse mutagenesis with a comprehensive cre characterization resource. *Nat Commun* **2012**, *3*, 1218, doi:10.1038/ncomms2186.
49. Frenz, S.; Rak, K.; Volker, J.; Jurgens, L.; Scherzad, A.; Schendzielorz, P.; Radeloff, A.; Jablonka, S.; Hansen, S.; Mlynksi, R.; et al. Mosaic pattern of Cre recombinase expression in cochlear outer hair cells of the Brn3.1 Cre mouse. *Neuroreport* **2015**, *26*, 309-313, doi:10.1097/WNR.0000000000000336.
50. Garrick, D.; Fiering, S.; Martin, D.I.; Whitelaw, E. Repeat-induced gene silencing in mammals. *Nat Genet* **1998**, *18*, 56-59, doi:10.1038/ng0198-56.
51. Cain-Hom, C.; Splinter, E.; van Min, M.; Simonis, M.; van de Heijning, M.; Martinez, M.; Asghari, V.; Cox, J.C.; Warming, S. Efficient mapping of transgene integration sites and local structural changes in Cre transgenic mice using targeted locus amplification. *Nucleic Acids Res* **2017**, *45*, e62, doi:10.1093/nar/gkw1329.

52. Choi, E.H.; Suh, S.; Einstein, D.E.; Leinonen, H.; Dong, Z.; Rao, S.R.; Fliesler, S.J.; Blackshaw, S.; Yu, M.; Peachey, N.S.; et al. An inducible Cre mouse for studying roles of the RPE in retinal physiology and disease. *JCI Insight* **2021**, *6*, doi:10.1172/jci.insight.146604.
53. Swarup, A.; Samuels, I.S.; Bell, B.A.; Han, J.Y.S.; Du, J.; Massenzio, E.; Abel, E.D.; Boesze-Battaglia, K.; Peachey, N.S.; Philip, N.J. Modulating GLUT1 expression in retinal pigment epithelium decreases glucose levels in the retina: impact on photoreceptors and Muller glial cells. *Am J Physiol Cell Physiol* **2019**, *316*, C121-C133, doi:10.1152/ajpcell.00410.2018.
54. Yao, J.; Jia, L.; Khan, N.; Lin, C.; Mitter, S.K.; Boulton, M.E.; Dunaief, J.L.; Klionsky, D.J.; Guan, J.L.; Thompson, D.A.; et al. Deletion of autophagy inducer RB1CC1 results in degeneration of the retinal pigment epithelium. *Autophagy* **2015**, *11*, 939-953, doi:10.1080/15548627.2015.1041699.
55. Sethna, S.; Scott, P.A.; Giese, A.P.J.; Duncan, T.; Jian, X.; Riazuddin, S.; Randazzo, P.A.; Redmond, T.M.; Bernstein, S.L.; Riazuddin, S.; et al. CIB2 regulates mTORC1 signaling and is essential for autophagy and visual function. *Nat Commun* **2021**, *12*, 3906, doi:10.1038/s41467-021-24056-1.
56. Bullock, J.; Polato, F.; Abu-Asab, M.; Bernardo-Colon, A.; Aflaki, E.; Agbaga, M.P.; Becerra, S.P. Degradation of Photoreceptor Outer Segments by the Retinal Pigment Epithelium Requires Pigment Epithelium-Derived Factor Receptor (PEDF-R). *Invest Ophthalmol Vis Sci* **2021**, *62*, 30, doi:10.1167/iovs.62.2.30.
57. Le, Y.Z.; Zheng, W.; Rao, P.C.; Zheng, L.; Anderson, R.E.; Esumi, N.; Zack, D.J.; Zhu, M. Inducible expression of cre recombinase in the retinal pigmented epithelium. *Invest Ophthalmol Vis Sci* **2008**, *49*, 1248-1253, doi:10.1167/iovs.07-1105.
58. Ueki, Y.; Ash, J.D.; Zhu, M.; Zheng, L.; Le, Y.Z. Expression of Cre recombinase in retinal Muller cells. *Vision Res* **2009**, *49*, 615-621, doi:10.1016/j.visres.2009.01.012.
59. Fu, S.; Zhu, M.; Wang, C.; Le, Y.Z. Efficient induction of productive Cre-mediated recombination in retinal pigment epithelium. *Mol Vis* **2014**, *20*, 480-487.
60. Mori, M.; Gargowitsch, L.; Bornert, J.M.; Garnier, J.M.; Mark, M.; Chambon, P.; Metzger, D. Temporally controlled targeted somatic mutagenesis in mouse eye pigment epithelium. *Genesis* **2012**, *50*, 828-832, doi:10.1002/dvg.22044.
61. Schneider, S.; Hotaling, N.; Campos, M.; Patnaik, S.R.; Bharti, K.; May-Simera, H.L. Generation of an inducible RPE-specific Cre transgenic-mouse line. *PLoS One* **2018**, *13*, e0207222, doi:10.1371/journal.pone.0207222.
62. Nasrin, M.; Ahmed, O.; Han, X.; Nojebuzzaman, M.; Abo-Ahmed, A.I.; Yazawa, S.; Osawa, M. Generation of Pmel-dependent conditional and inducible Cre-driver mouse line for melanocytic-targeted gene manipulation. *Pigment Cell Melanoma Res* **2023**, *36*, 53-70, doi:10.1111/pcmr.13074.
63. Philip, N.J.; Yoon, H.; Grollman, E.F. Monocarboxylate transporter MCT1 is located in the apical membrane and MCT3 in the basal membrane of rat RPE. *Am J Physiol* **1998**, *274*, R1824-1828, doi:10.1152/ajpregu.1998.274.6.R1824.
64. Philip, N.J.; Yoon, H.; Lombardi, L. Mouse MCT3 gene is expressed preferentially in retinal pigment and choroid plexus epithelia. *Am J Physiol Cell Physiol* **2001**, *280*, C1319-1326, doi:10.1152/ajpcell.2001.280.5.C1319.
65. Donnelly, M.L.L.; Luke, G.; Mehrotra, A.; Li, X.; Hughes, L.E.; Gani, D.; Ryan, M.D. Analysis of the aphthovirus 2A/2B polyprotein 'cleavage' mechanism indicates not a proteolytic reaction, but a novel translational effect: a putative ribosomal 'skip'. *J Gen Virol* **2001**, *82*, 1013-1025, doi:10.1099/0022-1317-82-5-1013.
66. Kim, J.H.; Lee, S.R.; Li, L.H.; Park, H.J.; Park, J.H.; Lee, K.Y.; Kim, M.K.; Shin, B.A.; Choi, S.Y. High cleavage efficiency of a 2A peptide derived from porcine teschovirus-1 in human cell lines, zebrafish and mice. *PLoS One* **2011**, *6*, e18556, doi:10.1371/journal.pone.0018556.
67. Mori, M.; Metzger, D.; Picaud, S.; Hindelang, C.; Simonutti, M.; Sahel, J.; Chambon, P.; Mark, M. Retinal dystrophy resulting from ablation of RXR alpha in the mouse retinal pigment epithelium. *Am J Pathol* **2004**, *164*, 701-710, doi:10.1016/s0002-9440(10)63157-4.

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.