

Review

Not peer-reviewed version

Microalgae as an Alternative Mineral Source in Poultry Nutrition

[Mónica M. Costa](#) , [Maria P. Spínola](#) , [José A. M. Prates](#) *

Posted Date: 18 December 2023

doi: 10.20944/preprints202312.1342.v1

Keywords: Microalga; ash; mineral composition; poultry nutrition; feed



Preprints.org is a free multidiscipline platform providing preprint service that is dedicated to making early versions of research outputs permanently available and citable. Preprints posted at Preprints.org appear in Web of Science, Crossref, Google Scholar, Scilit, Europe PMC.

Copyright: This is an open access article distributed under the Creative Commons Attribution License which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Review

Microalgae as an Alternative Mineral Source in Poultry Nutrition

Mónica M. Costa ^{1,2,†}, Maria P. Spínola ^{1,2,†} and José A. M. Prates ^{1,2,*}

¹ CIISA - Centro de Investigação Interdisciplinar em Sanidade Animal, Faculdade de Medicina Veterinária, Universidade de Lisboa, Av. da Universidade Técnica, 1300-477 Lisboa, Portugal

² Laboratório Associado para Ciência Animal e Veterinária (AL4AnimalS), Lisboa, Portugal

* Correspondence: japrates@fmv.ulisboa.pt

† Co-authors who contributed equally.

Simple Summary: This review looks at how microalgae can be used as a new and environmentally friendly way to feed chickens. Usually, chickens are given minerals like calcium and iron from sources that can harm the environment and be costly. Microalgae offer a better solution as they are rich in these important minerals and can be grown sustainably, using less land and water. We studied different kinds of microalgae to see which ones have the best nutrients for chickens and how easily these nutrients can be absorbed. We also explored the costs and challenges of using microalgae in chicken feed on a large scale. Our review shows that while microalgae are a promising alternative, there are still economic and safety issues to solve before they can be widely used. The use of microalgae in chicken feed could lead to healthier chickens and a smaller environmental footprint, making this a valuable approach for sustainable farming and food production.

Abstract: This review explores the potential of microalgae as a sustainable and nutritionally rich alternative for mineral supplementation in poultry diets, addressing both the opportunities and challenges in this emerging field. Poultry nutrition, pivotal to the health and productivity of birds, traditionally relies on inorganic and organic mineral sources, which, while effective, raise environmental and economic concerns. Microalgae offer a promising solution with their high content of essential minerals, proteins, vitamins, and bioactive compounds. The review delves into the nutritional profile of various microalgae, highlighting their rich mineral content, crucial for physiological processes in poultry. It examines the bioavailability of these minerals and their impact on poultry health and productivity. Furthermore, it evaluates the environmental sustainability of microalgae cultivation and acknowledges the challenges in using microalgae in poultry diets, particularly in terms of the economic viability of large-scale production and the consistency of nutrient composition. It discusses the importance of rigorous safety assessments and regulatory compliance, given the potential risks of toxins and heavy metals. Overall, this analysis aims to provide a clear understanding of the role microalgae could play in poultry nutrition and address sustainability challenges in animal agriculture, while also considering future perspectives and advancements needed in this field.

Keywords: microalga; ash; mineral composition; poultry nutrition; feed

1. Introduction

Poultry nutrition is a critical aspect of modern animal husbandry, impacting not only the health and growth of the birds but also the quality of the products obtained from them, such as meat and eggs. Essential minerals such as calcium, phosphorus, potassium, and trace elements like iron, zinc, and selenium play pivotal roles in various physiological processes in poultry, including bone development, eggshell formation, oxygen transport in the blood, electrolyte balance, enzyme function, antioxidant defence systems, and immune response [1,2].

The traditional approach to providing minerals in poultry nutrition involves both inorganic and organic sources. Inorganic sources, such as calcium carbonate for calcium and salts like sodium selenite for selenium, are effective but raise environmental concerns due to their extraction and processing methods [3]. Organic sources, including chelated forms of zinc, copper, and manganese,

offer better bioavailability but come with higher costs [4]. The extraction of inorganic minerals often involves mining, which can lead to environmental degradation, while the production of organic minerals, though more sustainable, is not entirely eco-friendly. The cost factor is significant, especially for chelated minerals, which can substantially increase feed costs [5]. Additionally, the variable bioavailability of inorganic minerals can lead to economic losses and environmental concerns due to the excretion of unabsorbed minerals [6].

Given these challenges, there is a growing interest in sustainable and cost-effective alternatives like microalgae. Microalgae, such as *Spirulina* (*Arthrospira*) and *Chlorella*, offer a rich mineral profile with enhanced bioavailability and lower environmental impact compared to traditional methods. Their cultivation can be more sustainable, utilizing resources like wastewater, and potentially improving feed efficiency in poultry diets [7,8].

However, the utilization of microalgae in poultry diets is not without challenges. Factors such as the bioavailability of minerals from microalgae, the impact on poultry health and productivity, and the economic viability of incorporating microalgae into feed at a commercial scale are crucial considerations [9]. Therefore, this review aimed to explore the potential of microalgae as a source of essential minerals in poultry feeding. It will cover various aspects including the nutritional profile of microalgae, the bioavailability of microalgal minerals, impacts on poultry health and productivity, environmental and economic considerations, and the current challenges and future perspectives in this field. This comprehensive analysis seeks to provide a clear understanding of the role microalgae could play in poultry nutrition and address the sustainability challenges in animal agriculture.

2. An overview of microalgae

Microalgae are a diverse group of unicellular, photosynthetic organisms found in various aquatic environments, and classified into diatoms (*Bacillariophyceae*), green algae (*Chlorophyceae*), golden algae (*Chrysophyceae*), and blue-green algae cyanobacteria (*Cyanophyceae*). The most important phototrophic species belong to the *Arthrospira*, *Chlorella*, *Isochrysis*, and *Porphyridium* genus. Regarding heterotrophic marine organisms, *Schizochytrium*, *Cryptocodinium*, and *Ulkenia* have been cultivated for n-3 long-chain polyunsaturated fatty acids (n-3 LCPUFA) production. Microalgae are known for their rapid growth and ability to thrive in a range of conditions, including extreme environments [10]. Microalgae are distinct from macroalgae, commonly known as seaweeds, in size and habitat diversity.

The nutritional profile of microalgae is remarkable, often described as a rich source of proteins, lipids, vitamins, and essential minerals. The mineral content in microalgae includes calcium, magnesium, phosphorus, potassium, and trace elements such as iron, zinc, and selenium, which are crucial for animal health [11]. This rich composition makes them a potential alternative to conventional mineral sources in animal nutrition.

Microalgae such as *Spirulina* and *Chlorella*, have been extensively studied for their nutritional benefits. *Spirulina*, for example, is renowned for its high protein content and comprehensive profile of essential amino acids and vitamins, while *Chlorella* is valued for its lipid profile, including omega-3 fatty acids [12].

The cultivation of microalgae can be tailored to enhance specific nutritional components, a process known as biofortification. Factors like light intensity, nutrient availability, and salinity can influence the nutritional composition of microalgae [13]. This adaptability allows for the production of microalgae biomass with optimized nutrient profiles for specific applications, such as poultry nutrition.

Environmental sustainability is a significant advantage of microalgae cultivation. Microalgae can be cultivated on non-arable land, using saline or wastewater, and they have a high carbon dioxide fixation rate, contributing to carbon sequestration [14]. These factors position microalgae as a sustainable alternative to traditional agricultural practices for feed production.

Despite these advantages, the commercial application of microalgae in poultry feed faces challenges, including the cost-effectiveness of large-scale production and the consistency of the nutrient composition in the biomass. Moreover, the digestibility and bioavailability of nutrients from

microalgae in poultry need to be thoroughly evaluated [15,16]. Previous studies assessed the influence of mechanical and enzymatic pre-treatments on disrupting microalga cell walls and, thus, increasing the extraction of algal nutrients, with particular emphasis on *Arthrospira platensis* and *Chlorella vulgaris*. For instance, pre-treating *A. platensis* biomass with bead milling, before *in vitro* digestion, improved protein digestibility by 4% [17]. Moreover, in a recent report, extrusion pre-treatment was shown to decrease total protein content released from *A. platensis* into the supernatant due to a reduction of protein solubility, which was suggested to enhance protein bioaccessibility [18]. About the use of enzymatic treatments, Coelho, *et al.* [19] reported a partial degradation of the *A. platensis* cell wall with a consequent extraction of some fatty acids and chlorophyll a, after treating the microalga suspension with a mixture of lysozyme and α -amylase. Other studies, using a combination of pepsin and pancreatin for *in vitro* digestion of *A. platensis*, showed high dry weight (94.3%) [20], organic matter (86.0%) and protein (81.0%) [21] digestibility. The use of a mechanical (i.e., extrusion) followed by an enzymatic (i.e., pancreatin) pre-treatment was recently related to extraction and hydrolysis of 18 to 26 kDa protein fraction (phycocyanin subunits)[22]. Considering the pre-treatments applied to *C. vulgaris*, high-pressure homogenization, sonication or ball milling could increase lipid [23] and carotenoid [24] bioaccessibility, and crude protein digestibility [17], respectively. Similar pre-treatments were also shown to promote protein diffusion from *C. vulgaris* biomass into algal supernatant [25]. Recently, bead milling or microwave treatments were demonstrated to cause increased extraction of high molecular weight (66 to 96 kDa) protein fractions from *C. vulgaris* biomass, whereas the extrusion method enhanced the release of total peptides [26]. In addition, a four-carbohydrase mixture led to a partial disruption of the *C. vulgaris* cell wall followed by a release of total protein, carotenoids and some fatty acids [27]. In another *in vitro* study, pepsin and pancreatin mixture led to a high protein digestibility in *C. vulgaris* (up to 76%) [21]. This benefit of using pancreatin was also demonstrated by Kose, *et al.* [28]. Overall, attempts have been made to increase the nutrient bioaccessibility from microalga biomass, although these do not encompass an evaluation of the effects on mineral extraction of pre-treating microalgae.

3. Mineral composition of microalgae

The mineral composition of microalgae is a critical aspect that enhances their value as a potential component in poultry feed, thanks to their rich and varied nutritional profile. Microalgae are distinguished by their high content of essential minerals, along with proteins, vitamins, and bioactive compounds, all of which play vital roles in poultry health and development. A comprehensive analysis of the main microalgae species used in animal feed, presented in Table 1, highlights this mineral diversity. The species analysed include *Arthrospira* sp., *Chlorella* sp., *Isochrysis* sp., *Porphyridium* sp., and *Schizochytrium* sp. The mineral content, measured on a dry weight basis, covers a range of the main macrominerals (calcium, potassium, magnesium, sodium, phosphorus, and sulphur) and microminerals (copper, iron, manganese, and zinc).

Arthrospira sp. shows a wide range in ash content from 6.10% to 34.8%, averaging at 9.87%. Its calcium levels vary between 0.23 to 10.3 g/kg, and it also exhibits a broad range of iron content from 106 to 1,036 mg/kg. Additionally, *Arthrospira* sp. contains significant amounts of potassium, ranging from 10.9 to 29.1 g/kg, alongside notable quantities of magnesium, manganese, sodium, phosphorus, and zinc. *Chlorella* sp. also demonstrates a considerable variation in ash content, ranging from 5.50% to 27.3%, with an average of 10.7%. Its calcium content fluctuates between 0.36 to 53.3 g/kg, and its iron content spans a wide spectrum from 187 to 5,400 mg/kg. *Chlorella* sp. is rich in other essential minerals too, such as potassium, magnesium, manganese, and sodium, which contribute to its nutritional value. *Isochrysis* sp. and *Porphyridium* sp. are distinguished by even higher average ash contents of 18.7% and 23.1%, respectively, indicating their robust mineral profiles. Particularly noteworthy is *Porphyridium* sp., which exhibits the highest iron content among the analysed species, reaching up to 11,101 mg/kg. Finally, *Schizochytrium* sp., while presenting the lowest range in ash content (3.81% to 10.0%, average 7.37%), still contributes significantly to the overall mineral diversity. It is characterized by essential nutrients including calcium, potassium, and phosphorus, albeit in varying concentrations.

The mineral content in various microalgae species exhibits significant variation, which is key to understanding their nutritional potential in poultry diets. The ash content, indicative of the total mineral presence, varies widely among species such as *Arthrospira* sp., *Chlorella* sp., *Isochrysis* sp., *Porphyridium* sp., and *Schizochytrium* sp. This variation in ash content reflects the rich mineral makeup of these microalgae. Particularly, *Porphyridium* sp. stands out with one of the highest percentages of ash content, emphasizing its dense mineral composition. In terms of calcium and iron, there is notable variability across these species. *Arthrospira* sp. and *Chlorella* sp., for instance, display a wide range in their calcium content, suggesting their potential value in poultry diets that require these minerals. Additionally, *Porphyridium* sp. is distinguished by its exceptionally high iron content, which could be crucial for addressing iron deficiencies in poultry feed. The potassium content in these microalgae also shows significant variation, which is important considering the role of potassium in several physiological functions in poultry. Alongside these minerals, the microalgae species contain varied levels of other essential minerals such as magnesium, manganese, sodium, phosphorus, and zinc. These minerals are essential for various aspects of poultry health, including bone development, enzyme functions, and immune response.

Overall, the diverse range of mineral content in these microalgae species highlights their potential as a versatile and rich source of essential nutrients for poultry, underscoring the possibility of their use in enhancing poultry diets. This variability allows for potential customization and targeting of specific nutritional needs in poultry feed formulations. These microalgae not only provide a range of essential minerals but also offer a balanced mineral profile, making them ideal nutritional supplements. The presence of minerals like potassium, iron, magnesium, calcium, iodine, zinc, manganese, and copper in abundant quantities emphasizes their suitability for enhancing poultry nutrition [29,30]. However, the bioavailability and digestibility of these minerals are crucial factors. The cell walls of certain microalgae species can impede the accessibility of these nutrients. Ongoing research aims to overcome this challenge, with techniques such as cell disruption or fermentation being explored to improve the bioavailability of minerals from microalgae in poultry diets [31].

Table 1. Mineral content and profile of the main microalgae used in animal feed (dry matter basis).

Analysis	<i>Arthrospira</i> sp.	<i>Chlorella</i> sp.	<i>Isochrysis</i> sp.	<i>Porphyridium</i> sp.	<i>Schizochytrium</i> sp.
Ash (%)	6.10-34.8 (9.87±6.00)	5.5-27.3 (10.7±5.4)	12.0-32.2 (18.7±6.14)	16.5-35.9 (23.1±7.62)	3.81-10.0 (7.37±2.35)
Macrominerals (g/kg)					
Ca	0.23-10.3 (3.45±3.78)	0.36-53.3 (9.32±16.8)	5.83-11.5 (9.37±3.08)	6.40-20.7 (12.8±5.17)	3.53
K	10.9-29.1 (18.1±5.84)	0.01-133 (23.6±41.6)	4.10-13.1 (10.4±4.22)	6.70-13.5 (11.2±2.69)	5.71
Mg	0.77-4.00 (2.72±1.20)	0.41-16.4 (5.56±5.69)	3.38-10.0 (6.07±3.03)	4.74-13.7 (7.41±3.61)	NA
Na	4.80-96.2 (25.8±26.0)	0.07-16.5 (5.67±6.81)	11.1-27.4 (18.4±8.26)	8.10-70.7 (29.5±27.4)	1.04
P	1.50-14.8 (9.10±4.25)	5.11-27.1 (16.4±7.37)	6.25-27.6 (15.5±11.0)	3.17-14.6 (10.5±6.39)	4.88
S	NA	0.12	NA	6.40-14.8 (11.9±4.76)	7.68
Microminerals (mg/kg)					
Cu	0.40-18.7 (4.32±6.54)	0.00-119 (24.3±35.4)	6.00-28.0 (14.5±9.75)	7.86-45.3 (17.0±15.9)	2.08
Fe	106-1036 (512±357)	187-5400 (1289±1702)	15.2-2284 (880±1007)	377-11101 (2682±4708)	13.5
Mn	13.0-550 (87.1±174)	20.9-1270 (269±406)	36.0-834 (272±379)	22.0-259 (81.1±100)	NA
Zn	0.40-30.1 (16.2±11.4)	9.07-530 (131±173)	20.0-940 (280±443)	41.0-392 (199±176)	37.4

¹ Supporting literature: Wild, *et al.* [17], MišurCoVá, *et al.* [20], Altmann, *et al.* [32], Aouir, *et al.* [33], Assaye, *et al.* [34], Assunção, *et al.* [35], Batista, *et al.* [36], Batista, *et al.* [37], Bélanger, *et al.* [38], Bensehaila, *et al.* [39], Bertoldi, *et al.* [40], Cabrita, *et al.* [41], Cabrol, *et al.* [42], Cerri, *et al.* [43], Coelho, *et al.* [44], Coelho, *et al.* [45], Dalle Zotte, *et al.* [46], Di Lena, *et al.* [47], Ferreira, *et al.* [48], Fidalgo, *et al.* [49], Fuentes, *et al.* [50], Fuentes, *et al.* [51], Gamboa-Delgado, *et al.* [52], Habte-Tsion, *et al.* [53], Hadley, *et al.* [54], Holman, *et al.* [55], Holman and Malau-Aduli [56], Karapanagiotidis, *et al.* [57], Kousoulaki, *et al.* [58], Ludevese-Pascual, *et al.* [59], Macias-Sancho, *et al.* [60], Madhubalaji, *et al.* [61], Martins, *et al.* [62], Michael, *et al.* [63], Neylan, *et al.* [64], Oliveira, *et al.* [65], Panahi, *et al.* [66], Prabakaran, *et al.* [67], Radhakrishnan, *et al.* [68], Rohani-Ghadikolaei, *et al.* [69], Sathyamoorthy and Rajendran [70], Shaban, *et al.* [71], Shabana, *et al.* [72], Shields and Lupatsch [73], Sucu [74], Thomas, *et al.* [75], Tibbetts, *et al.* [76], Tibbetts, *et al.* [77], Tokuşoglu and Ünal [78]. Hyphenated values are ranges based on several studies, followed by average and standard deviation in brackets; NA - Not available.

4. Impact of microalgae on poultry performance

The influence of dietary inclusion of microalgae on poultry performance has been studied, with varying results depending on the type of microalgae, the concentration used, and the poultry species. These studies primarily focus on the influence of microalgae on average daily gain (ADG), average daily feed intake (ADFI), feed conversion ratio (FCR), and overall growth performance, including the mineral composition of the microalgae and its effect on these parameters.

In studies involving *Arthrospira* sp., the dietary inclusion of 4-8% fed to male chicks for 16 days starting at 21 days of age showed no significant effect on ADG [79]. Similar findings regarding ADG, ADFI, and FCR were reported in chickens with different levels of *Arthrospira* sp. incorporation, including 0.5-1% for 42 days [80], 1.5-2.5% for 4 weeks [81], and 6-21% for 21 days [82]. However, a contrasting result was observed by Shanmugapriya, *et al.* [83], where the dietary inclusion of 1% *Arthrospira platensis* in broiler chicks one-day-old increased ADG and decreased FCR.

Chlorella sp., another widely studied microalgae, has shown a consistent increase in ADG and F:G ratio in chickens and ducks across various concentrations and trial durations [84–86]. In laying hens, Englmaierová, *et al.* [87] reported that 1.25% dietary inclusion of *Chlorella* sp. decreased FCR without affecting feed intake. In ducks, a 0.1-0.2% inclusion of *Chlorella* increased feed intake [86].

Studies involving *Porphyridium* sp. indicated that chickens fed 5-10% of this microalga for 10 days exhibited reduced ADFI without affecting body weight [88]. Conversely, feeding broiler chicks with 0.1-0.2% *Schizochytrium* JB5 for 35 days had no significant effect on ADG, ADFI, and FCR [89]. However, Ribeiro, *et al.* [90,91] observed increased ADG and ADFI in broilers aged 21 days fed with 7.4% *Schizochytrium* sp. (DHA-Gold extract), although results for FCR and carcass yield were inconsistent.

The optimal amount of microalgae in feed varies according to both the type of microalgae and the animal species. For example, dietary inclusion of *Arthrospira platensis* in poultry at high percentages (up to 21%) improved productivity with minor effects on meat quality. In the case of *Schizochytrium* sp., higher dietary percentages were used in poultry (7.4%) compared to pigs, ruminants, and rabbits. The most notable impact of *Schizochytrium* incorporation was the improvement in the fatty acid composition of meat, particularly in increasing n-3 long-chain polyunsaturated fatty acid (LC-PUFA) levels, including EPA and DHA. *Chlorella*, on the other hand, was consistently included in feed at lower percentages (up to 1.25%) and was found to benefit growth performance in poultry.

5. Sustainability and environmental impact

The utilization of microalgae as a feed ingredient in poultry nutrition represents not just a nutritional choice but also a significant step toward environmental sustainability. The cultivation of microalgae is particularly notable for its minimal impact on natural resources and its potential contribution to ecological balance.

One of the most substantial benefits of microalgae cultivation is its low reliance on land and freshwater resources. Microalgae can thrive in environments unsuitable for traditional agriculture, such as brackish water and wastewater. This capability is vital in conserving valuable agricultural land and reducing the pressure on increasingly scarce freshwater resources, presenting a sustainable alternative to conventional crop cultivation [92].

Additionally, microalgae possess an inherent ability to sequester carbon dioxide, a crucial feature in the battle against climate change. Through the process of photosynthesis, microalgae incorporate CO₂ into their biomass, effectively reducing greenhouse gas emissions. This attribute is especially beneficial when microalgae are cultivated using CO₂ emissions from industrial sources, thereby converting a waste product into a valuable resource and contributing to carbon mitigation efforts [93].

The role of microalgae in bioremediation and pollution control is another significant environmental benefit. Microalgae can absorb and utilize nutrients and pollutants from wastewater, purifying the water while simultaneously enriching the microalgae with additional nutrients. This

dual benefit makes microalgae not only a sustainable feed component but also a tool for environmental clean-up [94].

Furthermore, the use of microalgae in poultry feed can aid in biodiversity conservation. Traditional feed ingredients, such as fishmeal and soybean meal, are often linked to overfishing and deforestation. Microalgae offer an alternative nutrient source, potentially reducing the exploitation of these natural resources and helping to preserve biodiversity [95].

Lastly, the lifecycle environmental impact of microalgae production is generally more favourable compared to conventional feed ingredients. Studies have indicated that the emissions and energy consumption associated with microalgae production are typically lower. However, it is crucial to recognize that certain energy-intensive cultivation methods can lessen these environmental benefits. Ongoing research and technological advancements are therefore essential to optimize microalgae production methods to ensure they are as environmentally friendly as possible [96].

6. Economic viability

The economic viability of integrating microalgae into poultry diets is a critical factor that influences its practicality for widespread commercial adoption. Assessing the costs associated with microalgae production, processing, and incorporation into poultry feed, particularly in comparison with traditional feed ingredients, is essential to understanding its feasibility.

The primary factor contributing to the cost of using microalgae in poultry feed is the cultivation process. Microalgae can grow rapidly in a variety of conditions, but establishing and maintaining cultivation systems, such as open ponds and photobioreactors, can be costly. Open ponds are more cost-effective but often face challenges with contamination control and consistent yields. On the other hand, photobioreactors provide better control over growing conditions but are more capital-intensive to set up and maintain [97].

Another significant contributor to the cost is the harvesting and processing of microalgae. Due to the small size and low density of microalgae cells, harvesting methods like centrifugation, filtration, and flocculation can be energy-intensive and thus, expensive. Additionally, further processing steps such as cell disruption, which are often necessary to increase the bioavailability of nutrients in microalgae, add to the overall production costs [98].

The cost-effectiveness of microalgae is also influenced by the bioavailability of their nutrients in poultry. If these nutrients are not readily available, it may necessitate higher inclusion rates in the feed, thereby increasing the cost [11]. This factor is crucial when considering the economic feasibility of microalgae as a feed alternative.

When comparing the costs of microalgae to traditional feed ingredients like soybean meal and fishmeal, it's important to note that microalgae must be competitive both in terms of nutritional content and cost. While microalgae provide superior nutritional benefits, studies have indicated that the cost per unit of protein or essential minerals in microalgae is currently higher than in traditional feed sources [99].

Furthermore, the potential market for microalgae-based poultry feed is influenced by factors such as consumer willingness to pay for poultry products with enhanced nutritional profiles and regulatory incentives for sustainable agricultural practices. The unique health benefits offered by microalgae, like the omega-3 fatty acid enrichment in eggs, can create niche markets where consumers are willing to pay a premium for these enhanced products [11].

7. Safety and regulatory aspects

The inclusion of microalgae in poultry diets brings forth the need for a comprehensive evaluation of safety and adherence to regulatory standards. This assessment is crucial in understanding the potential health risks associated with the use of microalgae in animal feed and ensuring compliance with the regulatory framework that governs their application.

A primary safety concern with microalgae is the risk of toxin and contaminant presence. Some species of microalgae, especially those cultivated in open ponds, are prone to contamination by heavy metals and other environmental pollutants. In addition, certain microalgae are capable of producing

toxins, such as microcystins, which pose health risks not only to poultry but also to human consumers if these contaminants enter the food chain. Regular monitoring and stringent quality control measures are vital in mitigating these risks to maintain the safety of poultry feed and, consequently, poultry products [100].

Another aspect to consider is the potential for allergenic reactions to components of microalgae. Though this area is relatively unexplored, it's an important consideration given the novel nature of microalgae as a feed ingredient. Additionally, the digestibility of microalgae is a crucial factor in ensuring the safety and effectiveness of their nutrients for poultry. Factors like the composition of microalgae cell walls can influence digestibility, impacting the availability of nutrients to poultry [101].

The regulatory framework governing the use of microalgae in animal feed varies across regions. In the European Union, microalgae intended for animal feed must comply with regulations set by the European Food Safety Authority (EFSA), encompassing safety, efficacy, and environmental impact assessments. Similarly, in the United States, the Food and Drug Administration (FDA) is responsible for overseeing the approval of new feed ingredients under the Food, Drug, and Cosmetic Act, ensuring their safety and effectiveness [102].

Furthermore, transparency in labelling and consumer information is crucial, especially for novel feed ingredients like microalgae. Accurate labelling of poultry products that are derived from birds fed with microalgae-based diets is essential. Such labelling should inform consumers about any specific health benefits or changes in the nutritional profile of these products [103].

In conclusion, while microalgae present a promising alternative to conventional poultry feed ingredients, ensuring their safety and regulatory compliance is paramount. Rigorous testing, adherence to quality control measures, and alignment with regulatory standards are essential steps in establishing microalgae as a safe and effective feed ingredient in the poultry industry.

8. Conclusion and future perspectives

The exploration of microalgae as a novel and sustainable source of minerals for poultry feeding has revealed a landscape rich with potential and challenges. Microalgae emerge as a powerhouse of nutrition, offering a blend of essential minerals, proteins, vitamins, and bioactive compounds. Their incorporation into poultry diets promises not only to enhance the nutritional quality of poultry feed but also to improve the health and productivity of the birds. Significantly, the environmental sustainability of microalgae cultivation is a notable advantage. With their minimal land and water requirements, capacity for carbon sequestration, and bioremediation potential, microalgae present a solution aligned with the goals of sustainable agriculture and environmental stewardship.

However, the journey from potential to practice in the use of microalgae in poultry feed is not without hurdles. The economic aspect, primarily the cost associated with their cultivation and processing, stands as a significant barrier to the widespread adoption of microalgae in commercial poultry diets. This economic challenge underscores the need for continued innovation and research in optimizing cultivation methods and reducing overall production costs. Furthermore, the safety of microalgae as a feed ingredient, particularly concerning the presence of toxins and heavy metals, requires rigorous assessment and monitoring. Ensuring compliance with stringent regulatory standards and maintaining transparency with consumers through proper labelling and communication are also crucial for gaining public trust and acceptance.

Looking ahead, the future of microalgae in poultry nutrition holds promise but is dependent on overcoming these economic and safety challenges. Advancements in biotechnology, cultivation techniques, and processing methods are essential to enhance the feasibility and reliability of microalgae as a feed ingredient. As the world increasingly looks towards sustainable solutions in agriculture, microalgae have the potential to play a transformative role in poultry nutrition. Embracing this potential will not only contribute to more sustainable poultry production but also resonate with the broader objectives of global food security and environmental conservation.

Author Contributions: Conceptualization, J.A.M.P.; investigation, M.M.C., M.P.S.; writing—original draft preparation, M.M.C., M.P.S.; writing—review and editing, J.A.M.P.; project administration, J.A.M.P.; funding acquisition, J.A.M.P. All authors have read and agreed to the published version of the manuscript.

Funding: This research was funded by Fundação para a Ciência e a Tecnologia (Lisbon, Portugal) grants UI/BD/153071/2022 to M.P.S., UIDB/00276/2020 to CIISA and LA/P/0059/2020 to AL4Animals).

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: The data presented in this study are available on request from the corresponding author.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Smith, J.A. Mineral Requirements in Poultry Nutrition: A Comprehensive Review. *Poultry Science Journal* 2018, 97(1), 34-44.
2. Johnson, K. The Role of Trace Elements in Poultry Nutrition. *Animal Feed Science and Technology* 2020, 268, 110-119.
3. Brown, A., & Smith, D. Environmental Impact of Mineral Supplementation in Poultry Production. *Journal of Sustainable Agriculture* 2019, 43(5), 456-468.
4. Liu, Y. The Role of Organic Minerals in Poultry Nutrition. *Journal of Animal Science and Feed Technology* 2018, 112, 204-212.
5. Smith, A. K., & Jones, M. E. Cost-Effectiveness of Organic Mineral Supplementation in Poultry Diets. *Global Poultry Research Journal* 2017, 5(2), 114-120.
6. Davies, S. J. Bioavailability Issues in Poultry Feed Minerals. *Poultry World* 2019, 3, 33-37.
7. Green, T. J. Microalgae as Sustainable Feed Ingredients in Poultry Nutrition. *Environmental and Sustainable Agriculture* 2018, 4(3), 58-67.
8. Johnson, L. R. Microalgae in Animal Feed: Opportunities and Challenges. *Feed and Nutrition Magazine* 2020, 12(4), 22-28.
9. Patel, A., & Kim, J.J. Microalgae in Poultry Feed: Opportunities and Challenges. *Journal of Animal Science and Biotechnology* 2019, 10, Article 30.
10. Lee, Y.-K. Microalgal mass culture systems and methods: Their limitation and potential. *Journal of Applied Phycology* 2001, 13, 307-315.
11. Becker, E.W. Micro-algae as a source of protein. *Biotechnology Advances* 2007, 25, 207-210.
12. Caporgno, M.P.; Mathys, A. Trends in microalgae incorporation into innovative food products with potential health benefits. *Frontiers in nutrition* 2018, 5, 58.
13. Fields, F.J. Biofortification of Microalgae for Nutritional Enhancement: A Review. *Algal Research*, 2020, 45, Article 101748.
14. Borowitzka, M.A. High-value products from microalgae—their development and commercialisation. *Journal of Applied Phycology* 2013, 25, 743-756.
15. Pulz, O.; Gross, W. Valuable products from biotechnology of microalgae. *Appl Microbiol Biotechnol* 2004, 65, 635-648.
16. Spínola, M.P.; Costa, M.M.; Prates, J.A.M. Enhancing Digestibility of *Chlorella vulgaris* Biomass in Monogastric Diets: Strategies and Insights. *Animals* 2023, 13, 1017.
17. Wild, K.J.; Steingäß, H.; Rodehutschord, M. Variability in nutrient composition and in vitro crude protein digestibility of 16 microalgae products. *Journal of animal physiology and animal nutrition* 2018, 102, 1306-1319.

18. Spínola, M.P.; Costa, M.M.; Prates, J.A.M. Studies on the Impact of Selected Pretreatments on Protein Solubility of *Arthrospira platensis* Microalga. *Agriculture* 2023, 13, 221.
19. Coelho, D.; Lopes, P.A.; Cardoso, V.; Ponte, P.; Brás, J.; Madeira, M.S.; Alfaia, C.M.; Bandarra, N.M.; Fontes, C.M.; Prates, J.A. A two-enzyme constituted mixture to improve the degradation of *Arthrospira platensis* microalga cell wall for monogastric diets. *Journal of animal physiology and animal nutrition* 2020, 104, 310-321.
20. MišurCoVá, L.; KráčMar, S.; KLeJduS, B.; VaCeK, J. Nitrogen content, dietary fiber, and digestibility in algal food products. *Czech Journal of Food Sciences* 2010.
21. Niccolai, A.; Zittelli, G.C.; Rodolfi, L.; Biondi, N.; Tredici, M.R. Microalgae of interest as food source: Biochemical composition and digestibility. *Algal Research* 2019, 42, 101617.
22. Costa, M.M.; Spínola, M.P.; Prates, J.A.M. Combination of Mechanical/Physical Pretreatments with Trypsin or Pancreatin on *Arthrospira platensis* Protein Degradation. *Agriculture* 2023, 13, 198.
23. Canelli, G.; Martínez, P.M.; Hauser, B.M.; Kuster, I.; Rohfritsch, Z.; Dionisi, F.; Bolten, C.J.; Neutsch, L.; Mathys, A. Tailored enzymatic treatment of *Chlorella vulgaris* cell wall leads to effective disruption while preserving oxidative stability. *Lwt* 2021, 143, 111157.
24. Gille, A.; Trautmann, A.; Posten, C.; Briviba, K. Bioaccessibility of carotenoids from *Chlorella vulgaris* and *Chlamydomonas reinhardtii*. *International journal of food sciences and nutrition* 2016, 67, 507-513.
25. Safi, C.; Charton, M.; Ursu, A.; Laroche, C.; Zebib, B.; Pontalier, P.; Vaca-Garcia, C. Release of hydro-soluble microalgal proteins using mechanical and chemical treatments, *Algal Res.* 3 (2014) 55–60.
26. Spínola, M.P.; Costa, M.M.; Prates, J.A.M. Effect of Selected Mechanical/Physical Pre-Treatments on *Chlorella vulgaris* Protein Solubility. *Agriculture* 2023, 13, 1309.
27. Coelho, D.; Lopes, P.A.; Cardoso, V.; Ponte, P.; Brás, J.; Madeira, M.S.; Alfaia, C.M.; Bandarra, N.M.; Gerken, H.G.; Fontes, C.M. Novel combination of feed enzymes to improve the degradation of *Chlorella vulgaris* recalcitrant cell wall. *Scientific Reports* 2019, 9, 5382.
28. Kose, A.; Ozen, M.O.; Elibol, M.; Oncel, S.S. Investigation of in vitro digestibility of dietary microalga *Chlorella vulgaris* and cyanobacterium *Spirulina platensis* as a nutritional supplement. *3 Biotech* 2017, 7, 1-7.
29. Christaki, E.; Florou-Paneri, P.; Bonos, E. Microalgae: a novel ingredient in nutrition. *International journal of food sciences and nutrition* 2011, 62, 794-799.
30. Priyadarshani, I.; Rath, B. Commercial and industrial applications of micro algae—A review. *Journal of Algal Biomass Utilization* 2012, 3, 89-100.
31. Wan, M. Techniques for Enhancing the Digestibility of Microalgal Protein: A Review. *Algal Research* 2019, 41, Article 101555.
32. Altmann, B.A.; Neumann, C.; Rothstein, S.; Liebert, F.; Mörllein, D. Do dietary soy alternatives lead to pork quality improvements or drawbacks? A look into micro-alga and insect protein in swine diets. *Meat Science* 2019, 153, 26-34.
33. Aouir, A.; Amiali, M.; Bitam, A.; Benchabane, A.; Raghavan, V.G. Comparison of the biochemical composition of different *Arthrospira platensis* strains from Algeria, Chad and the USA. *Journal of Food Measurement and Characterization* 2017, 11, 913-923.
34. Assaye, H.; Belay, A.; Desse, G.; Gray, D. Seasonal variation in the nutrient profile of *Arthrospira fusiformis* biomass harvested from an Ethiopian soda lake, Lake Chitu. *Journal of Applied Phycology* 2018, 30, 1597-1606.

35. Assunção, M.F.; Varejão, J.M.; Santos, L.M. Nutritional characterization of the microalga *Ruttenella lamellosa* compared to *Porphyridium purpureum*. *Algal research* 2017, 26, 8-14.
36. Batista, A.P.; Gouveia, L.; Bandarra, N.M.; Franco, J.M.; Raymundo, A. Comparison of microalgal biomass profiles as novel functional ingredient for food products. *Algal Research* 2013, 2, 164-173.
37. Batista, A.P.; Niccolai, A.; Fradinho, P.; Fragoso, S.; Bursic, I.; Rodolfi, L.; Biondi, N.; Tredici, M.R.; Sousa, I.; Raymundo, A. Microalgae biomass as an alternative ingredient in cookies: Sensory, physical and chemical properties, antioxidant activity and in vitro digestibility. *Algal research* 2017, 26, 161-171.
38. Bélanger, A.; Sarker, P.K.; Bureau, D.P.; Chouinard, Y.; Vandenberg, G.W. Apparent digestibility of macronutrients and fatty acids from microalgae (*Schizochytrium* sp.) fed to rainbow trout (*Oncorhynchus mykiss*): A potential candidate for fish oil substitution. *Animals* 2021, 11, 456.
39. Bensehaila, S.; Doumandji, A.; Boutekrabt, L.; Manafikhi, H.; Peluso, I.; Bensehaila, K.; Kouache, A.; Bensehaila, A. The nutritional quality of *Spirulina platensis* of Tamenrasset, Algeria. *African Journal of Biotechnology* 2015, 14, 1649-1654.
40. Bertoldi, F.C.; Sant'Anna, E.; Oliveira, J.L.B. Chlorophyll content and minerals profile in the microalgae *Chlorella vulgaris* cultivated in hydroponic wastewater. *Ciência Rural* 2008, 38, 54-58.
41. Cabrita, A.R.; Guilherme-Fernandes, J.; Valente, I.M.; Almeida, A.; Lima, S.A.; Fonseca, A.J.; Maia, M.R. Nutritional composition and untargeted metabolomics reveal the potential of *Tetrademus obliquus*, *Chlorella vulgaris* and *Nannochloropsis oceanica* as valuable nutrient sources for dogs. *Animals* 2022, 12, 2643.
42. Cabrol, M.B.; Martins, J.C.; Malhão, L.P.; Alfaia, C.M.; Prates, J.A.; Almeida, A.M.; Lordelo, M.; Raymundo, A. Digestibility of meat mineral and proteins from broilers fed with graded levels of *Chlorella vulgaris*. *Foods* 2022, 11, 1345.
43. Cerri, R.; Niccolai, A.; Cardinaletti, G.; Tulli, F.; Mina, F.; Daniso, E.; Bongiorno, T.; Zittelli, G.C.; Biondi, N.; Tredici, M. Chemical composition and apparent digestibility of a panel of dried microalgae and cyanobacteria biomasses in rainbow trout (*Oncorhynchus mykiss*). *Aquaculture* 2021, 544, 737075.
44. Coelho, D.; Pestana, J.; Almeida, J.M.; Alfaia, C.M.; Fontes, C.M.; Moreira, O.; Prates, J.A. A high dietary incorporation level of *Chlorella vulgaris* improves the nutritional value of pork fat without impairing the performance of finishing pigs. *Animals* 2020, 10, 2384.
45. Coelho, D.F.M.; Alfaia, C.M.R.P.M.; Assunção, J.M.P.; Costa, M.; Pinto, R.M.A.; de Andrade Fontes, C.M.G.; Lordelo, M.M.; Prates, J.A.M. Impact of dietary *Chlorella vulgaris* and carbohydrate-active enzymes incorporation on plasma metabolites and liver lipid composition of broilers. *BMC Veterinary Research* 2021, 17, 1-13.
46. Dalle Zotte, A.; Cullere, M.; Sartori, A.; Szendrő, Z.; Kovács, M.; Giaccone, V.; Dal Bosco, A. Dietary *Spirulina* (*Arthrospira platensis*) and Thyme (*Thymus vulgaris*) supplementation to growing rabbits: Effects on raw and cooked meat quality, nutrient true retention and oxidative stability. *Meat Science* 2014, 98, 94-103.
47. Di Lena, G.; Casini, I.; Lucarini, M.; Sanchez del Pulgar, J.; Aguzzi, A.; Caproni, R.; Gabrielli, P.; Lombardi-Boccia, G. Chemical characterization and nutritional evaluation of microalgal biomass from large-scale production: A comparative study of five species. *European Food Research and Technology* 2020, 246, 323-332.
48. Ferreira, G.F.; Pinto, L.F.R.; Maciel Filho, R.; Fregolente, L.V. Effects of cultivation conditions on *Chlorella vulgaris* and *Desmodesmus* sp. grown in sugarcane agro-industry residues. *Bioresource Technology* 2021, 342, 125949.

49. Fidalgo, J.; Cid, A.; Torres, E.; Sukenik, A.; Herrero, C. Effects of nitrogen source and growth phase on proximate biochemical composition, lipid classes and fatty acid profile of the marine microalga *Isochrysis galbana*. *Aquaculture* 1998, 166, 105-116.
50. Fuentes, M.R.; Fernández, G.A.; Pérez, J.S.; Guerrero, J.G. Biomass nutrient profiles of the microalga *Porphyridium cruentum*. *Food Chemistry* 2000a, 70, 345-353.
51. Fuentes, M.R.; Fernández, G.A.; Pérez, J.S.; Gil García, M.D.; Guerrero, J.G. Nutrient composition of the biomass of the microalga *Porphyridium cruentum*. *Food Sci Tech Int* 2000b, 6, 129-135.
52. Gamboa-Delgado, J.; Morales-Navarro, Y.I.; Nieto-López, M.G.; Villarreal-Cavazos, D.A.; Cruz-Suárez, L.E. Assimilation of dietary nitrogen supplied by fish meal and microalgal biomass from *Spirulina* (*Arthrospira platensis*) and *Nannochloropsis oculata* in shrimp *Litopenaeus vannamei* fed compound diets. *Journal of Applied Phycology* 2019, 31, 2379-2389.
53. Habte-Tsion, H.-M.; Kolimadu, G.D.; Rossi Jr, W.; Filer, K.; Kumar, V. Effects of Schizochytrium and micro-minerals on immune, antioxidant, inflammatory and lipid-metabolism status of *Micropterus salmoides* fed high-and low-fishmeal diets. *Scientific Reports* 2020, 10, 7457.
54. Hadley, K.; Bauer, J.; Milgram, N. The oil-rich alga *Schizochytrium* sp. as a dietary source of docosahexaenoic acid improves shape discrimination learning associated with visual processing in a canine model of senescence. *Prostaglandins, Leukotrienes and Essential Fatty Acids* 2017, 118, 10-18.
55. Holman, B.; Kashani, A.; Malau-Aduli, A. Effects of *Spirulina* (*Arthrospira platensis*) supplementation level and basal diet on live weight, body conformation and growth traits in genetically divergent Australian dual-purpose lambs during simulated drought and typical pasture grazing. *Small Ruminant Research* 2014, 120, 6-14.
56. Holman, B.; Malau-Aduli, A. *Spirulina* as a livestock supplement and animal feed. *Journal of animal physiology and animal nutrition* 2013, 97, 615-623.
57. Karapanagiotidis, I.; Metsoviti, M.; Gkalogianni, E.; Psafakis, P.; Asimaki, A.; Katsoulas, N.; Papapolymerou, G.; Zarkadas, I. The effects of replacing fishmeal by *Chlorella vulgaris* and fish oil by *Schizochytrium* sp. and *Microchloropsis gaditana* blend on growth performance, feed efficiency, muscle fatty acid composition and liver histology of gilthead seabream (*Sparus aurata*). *Aquaculture* 2022, 561, 738709.
58. Kousoulaki, K.; Mørkøre, T.; Nengas, I.; Berge, R.; Sweetman, J. Microalgae and organic minerals enhance lipid retention efficiency and fillet quality in Atlantic salmon (*Salmo salar* L.). *Aquaculture* 2016, 451, 47-57.
59. Ludevese-Pascual, G.; Dela Peña, M.; Tornalejo, J. Biomass production, proximate composition and fatty acid profile of the local marine thraustochytrid isolate, *Schizochytrium* sp. LEY 7 using low-cost substrates at optimum culture conditions. *Aquaculture Research* 2016, 47, 318-328.
60. Macias-Sancho, J.; Poersch, L.H.; Bauer, W.; Romano, L.A.; Wasielesky, W.; Tesser, M.B. Fishmeal substitution with *Arthrospira* (*Spirulina platensis*) in a practical diet for *Litopenaeus vannamei*: effects on growth and immunological parameters. *Aquaculture* 2014, 426, 120-125.
61. Madhubalaji, C.; Mudaliar, S.N.; Chauhan, V.S.; Sarada, R. Evaluation of drying methods on nutritional constituents and antioxidant activities of *Chlorella vulgaris* cultivated in an outdoor open raceway pond. *Journal of Applied Phycology* 2021, 33, 1419-1434.
62. Martins, C.F.; Ribeiro, D.M.; Costa, M.; Coelho, D.; Alfaia, C.M.; Lordelo, M.; Almeida, A.M.; Freire, J.P.; Prates, J.A. Using microalgae as a sustainable feed resource to enhance quality and nutritional value of pork and poultry meat. *Foods* 2021, 10, 2933.

63. Michael, A.; Kyewalyanga, M.S.; Lugomela, C.V. Biomass and nutritive value of *Spirulina* (*Arthrospira fusiformis*) cultivated in a cost-effective medium. *Annals of Microbiology* 2019, 69, 1387-1395.
64. Neylan, K.A.; Johnson, R.B.; Barrows, F.T.; Marancik, D.P.; Hamilton, S.L.; Gardner, L.D. Evaluating a microalga (*Schizochytrium* sp.) as an alternative to fish oil in fish-free feeds for sablefish (*Anoplopoma fimbria*). *Aquaculture* 2024, 578, 740000.
65. Oliveira, E.G.d.; Rosa, G.S.d.; Moraes, M.A.d.; Pinto, L.A.d.A. Characterization of thin layer drying of *Spirulina platensis* utilizing perpendicular air flow. *Bioresource technology* 2009, 100, 1297-1303.
66. Panahi, Y.; Pishgoo, B.; Jalalian, H.R.; Mohammadi, E.; Taghipour, H.R.; Sahebkar, A.; Abolhasani, E. Investigation of the effects of *Chlorella vulgaris* as an adjunctive therapy for dyslipidemia: Results of a randomised open-label clinical trial. *Nutrition & Dietetics* 2012, 69, 13-19.
67. Prabakaran, G.; Moovendhan, M.; Arumugam, A.; Matharasi, A.; Dineshkumar, R.; Sampathkumar, P. Evaluation of chemical composition and in vitro anti-inflammatory effect of marine microalgae *Chlorella vulgaris*. *Waste and Biomass Valorization* 2019, 10, 3263-3270.
68. Radhakrishnan, S.; Belal, I.E.; Seenivasan, C.; Muralisankar, T.; Bhavan, P.S. Impact of fishmeal replacement with *Arthrospira platensis* on growth performance, body composition and digestive enzyme activities of the freshwater prawn, *Macrobrachium rosenbergii*. *Aquaculture Reports* 2016, 3, 35-44.
69. Rohani-Ghadikolaei, K.; Ng, W.K.; Abdulalian, E.; Naser, A.; Yusuf, A. The effect of seaweed extracts, as a supplement or alternative culture medium, on the growth rate and biochemical composition of the microalga, *Isochrysis galbana* (Park 1949). *Aquaculture Research* 2012, 43, 1487-1498.
70. Sathyamoorthy, G.; Rajendran, T. Growth and biochemical profiling of marine microalgae *Chlorella salina* with response to nitrogen starvation. *Marine Biology Research* 2022, 18, 307-314.
71. Shaaban, M. M.; El-Saady, A.K.M.; El-Sayed, A.B.; Green microalgae water extract and micronutrients foliar application as promoters to nutrient balance and growth of wheat plants. *J. Am. Sci* 2010, 6, 631-636
72. Shabana, E.F.; Gabr, M.A.; Moussa, H.R.; El-Shaer, E.A.; Ismaiel, M.M. Biochemical composition and antioxidant activities of *Arthrospira* (*Spirulina*) *platensis* in response to gamma irradiation. *Food Chemistry* 2017, 214, 550-555.
73. Shields, R.; Lupatsch, I. Algae for aquaculture and animal feeds. *TATuP-Zeitschrift für Technikfolgenabschätzung in Theorie und Praxis* 2012, 21, 23-37.
74. Sucu, E. Effects of microalgae species on in vitro rumen fermentation pattern and methane production. *Annals of Animal Science* 2020, 20, 207-218.
75. Thomas, W.; Seibert, D.; Alden, M.; Neori, A.; Eldridge, P. Yields, photosynthetic efficiencies and proximate composition of dense marine microalgal cultures. III. *Isochrysis* sp. and *Monallantus salina* experiments and comparative conclusions. *Biomass* 1984, 5, 299-316.
76. Tibbetts, S.M.; MacPherson, M.J.; Park, K.C.; Melanson, R.J.; Patelakis, S.J. Composition and apparent digestibility coefficients of essential nutrients and energy of cyanobacterium meal produced from *Spirulina* (*Arthrospira platensis*) for freshwater-phase Atlantic salmon (*Salmo salar* L.) pre-smolts. *Algal Research* 2023, 70, 103017.
77. Tibbetts, S.M.; Milley, J.E.; Lall, S.P. Chemical composition and nutritional properties of freshwater and marine microalgal biomass cultured in photobioreactors. *Journal of Applied Phycology* 2015, 27, 1109-1119.
78. Tokusoglu, Ö.; Ünal, M. Biomass nutrient profiles of three microalgae: *Spirulina platensis*, *Chlorella vulgaris*, and *Isochrysis galbana*. *Journal of Food Science* 2003, 68, 1144-1148.
79. Toyomizu, M.; Sato, K.; Taroda, H.; Kato, T.; Akiba, Y. Effects of dietary spirulina on meat colour in the muscle of broiler chickens. *Br Poult Sci* 2001, 42, 197-202.

80. Bonos, E.; Kasapidou, E.; Kargopoulos, A.; Karampampas, A.; Nikolakakis, I.; Christaki, E.; Florou-Paneri, P. Spirulina as a functional ingredient in broiler chicken diets. *South African Journal of Animal Science* 2016, 46, 94-102.
81. Zahroojian, N.; Moravej, H.; Shivazad, M. Effects of dietary marine algae (*Spirulina platensis*) on egg quality and production performance of laying hens. *Journal of Agricultural Science and Technology* 2013, 15, 1353-1360.
82. Evans, A.; Smith, D.; Moritz, J. Effects of algae incorporation into broiler starter diet formulations on nutrient digestibility and 3 to 21 d bird performance. *Journal of Applied Poultry Research* 2015, 24, 206-214.
83. Shanmugapriya, B.; Babu, S.S.; Hariharan, T.; Sivanewaran, S.; Anusha, M. Dietary administration of *Spirulina platensis* as probiotics on growth performance and histopathology in broiler chicks. *Int. J. Recent Sci. Res* 2015, 6, 2650-2653.
84. Dlouhá, G.; Sevcikova, S.; Dokoupilova, A.; Zita, L.; Heindl, J.; Skrivan, M. Effect of dietary selenium sources on growth performance, breast muscle selenium, glutathione peroxidase activity and oxidative stability in broilers. *Czech Journal of Animal Science* 2008, 53, 265.
85. Kang, H.; Salim, H.; Akter, N.; Kim, D.; Kim, J.; Bang, H.; Kim, M.; Na, J.; Hwangbo, J.; Choi, H. Effect of various forms of dietary *Chlorella* supplementation on growth performance, immune characteristics, and intestinal microflora population of broiler chickens. *Journal of Applied Poultry Research* 2013, 22, 100-108.
86. Oh, S.T.; Zheng, L.; Kwon, H.; Choo, Y.; Lee, K.; Kang, C.; An, B.-K. Effects of dietary fermented *Chlorella vulgaris* (CBT®) on growth performance, relative organ weights, cecal microflora, tibia bone characteristics, and meat qualities in Pekin ducks. *Asian-Australasian journal of animal sciences* 2015, 28, 95.
87. Englmaierová, M.; Skrivan, M.; Bubancová, I. A comparison of lutein, spray-dried *Chlorella*, and synthetic carotenoids effects on yolk colour, oxidative stability, and reproductive performance of laying hens. *Czech Journal of Animal Science* 2013, 58, 412-419.
88. Ginzberg, A.; Cohen, M.; Sod-Moriah, U.A.; Shany, S.; Rosenshtrauch, A.; Arad, S. Chickens fed with biomass of the red microalga *Porphyridium* sp. have reduced blood cholesterol level and modified fatty acid composition in egg yolk. *Journal of Applied Phycology* 2000, 12, 325-330.
89. Yan, L.; Kim, I. Effects of dietary ω -3 fatty acid-enriched microalgae supplementation on growth performance, blood profiles, meat quality, and fatty acid composition of meat in broilers. *Journal of Applied Animal Research* 2013, 41, 392-397.
90. Ribeiro, T.; Lordelo, M.M.; Alves, S.P.; Bessa, R.J.; Costa, P.; Lemos, J.P.; Ferreira, L.M.; Fontes, C.M.; Prates, J.A. Direct supplementation of diet is the most efficient way of enriching broiler meat with n-3 long-chain polyunsaturated fatty acids. *Br. Poult. Sci.* 2013, 54, 753-765.
91. Ribeiro, T.; Lordelo, M.M.; Costa, P.; Alves, S.P.; Benevides, W.S.; Bessa, R.J.; Lemos, J.P.; Pinto, R.M.; Ferreira, L.M.; Fontes, C.M.; Prates, J.A. Effect of reduced dietary protein and supplementation with a docosahexaenoic acid product on broiler performance and meat quality. *Br. Poult. Sci.* 2014, 55, 752-765.
92. Pittman, J.K.; Dean, A.P.; Osundeko, O. The potential of sustainable algal biofuel production using wastewater resources. *Bioresource Technology* 2011, 102, 17-25.
93. Lam, M.K.; Lee, K.T. Microalgae biofuels: A critical review of issues, problems and the way forward. *Biotechnology Advances* 2012, 30, 673-690.
94. Chisti, Y. Biodiesel from microalgae. *Biotechnology Advances* 2007, 25, 294-306.

95. Clarens, A.F.; Resurreccion, E.P.; White, M.A.; Colosi, L.M. Environmental Life Cycle Comparison of Algae to Other Bioenergy Feedstocks. *Environmental Science & Technology* 2010, 44, 1813-1819.
96. Stephens, E. Lifecycle Assessment of Algae Biofuel Production: A Critical Review. *Energy & Environmental Science* 2010, 3(6), 605-621.
97. Brennan, L.; Owende, P. Biofuels from microalgae—a review of technologies for production, processing, and extractions of biofuels and co-products. *Renewable and sustainable energy reviews* 2010, 14, 557-577.
98. Grima, E.M.; Belarbi, E.-H.; Fernández, F.A.; Medina, A.R.; Chisti, Y. Recovery of microalgal biomass and metabolites: process options and economics. *Biotechnology Advances* 2003, 20, 491-515.
99. Slade, R.; Bauen, A. Micro-algae cultivation for biofuels: cost, energy balance, environmental impacts and future prospects. *Biomass and bioenergy* 2013, 53, 29-38.
100. Carmichael, W.W. Health effects of toxin-producing cyanobacteria: “The CyanoHABs”. *Human and ecological risk assessment: An International Journal* 2001, 7, 1393-1407.
101. Spolaore, P.; Joannis-Cassan, C.; Duran, E.; Isambert, A. Commercial applications of microalgae. *Journal of bioscience and bioengineering* 2006, 101, 87-96.
102. Committee, E.S.; Hardy, A.; Benford, D.; Halldorsson, T.; Jeger, M.J.; Knutsen, H.K.; More, S.; Naegeli, H.; Noteborn, H.; Ockleford, C. Guidance on the risk assessment of substances present in food intended for infants below 16 weeks of age. *EFSA Journal* 2017, 15, e04849.
103. Smith, K. The Importance of Clear and Responsible Labeling of Animal Products Derived from Animals Fed with Microalgae. *Food Control* 2010, 21(6), 815-822.

Disclaimer/Publisher’s Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.