Review

Optimizing the Nitrogen Use Efficiency in Vegetable Crops

Hector Valenzuela *

Posted Date: 12 December 2023

doi: 10.20944/preprints202312.0768.v1

Keywords: agroecology; nitrogen; nitrogen cycle; nitrogen use efficiency; greenhouse gas emissions; organic matter; vegetable crops

Review

Optimizing the Nitrogen Use Efficiency in Vegetable Crops

Hector Valenzuela

Department of Plant and Environmental Protection Sciences, University of Hawaii at Manoa, Honolulu, HI 96822, USA; hector@hawaii.edu

Abstract: Nitrogen (N) is the most limiting nutrient for the production of vegetable crops, but anthropogenic sources pose risks due to its transformation into several reactive forms and movement throughout the environment. To mitigate the impact of environmental losses, current research efforts aim to improve management practices to obtain a greater Nutrient Use Efficiency (NUE). From a chemical, biological, and ecophysiological perspective, at a reductionist level we have acquired a considerable mechanistic understanding about the N cycle, the varied N chemical forms, microbial interactions and physiology of crop uptake. However, at a holistic level, we are far from obtaining an adequate understanding of the myriad of multi-level cropping system, genetic, socioeconomic, environmental, and ecological interactions that have an effect on the N cycle and NUE. Here, based on a selected review of the literature, we provide a perspective on the potentials to increase the NUE of vegetable crops, by adopting a range of best management practices and by highlighting some of the N x system interactions that may be considered to better understand the complex dynamics of the N cycle, to optimize the NUE of vegetable crops, and to explore alternatives to the reliance on the use of synthetic N sources.

Keywords: agroecology; nitrogen; nitrogen cycle; Nitrogen Use Efficiency; greenhouse gas emissions; organic matter; vegetable crops

1. Introduction

While N is an essential nutrient for plant growth, and life on earth, surplus application rates, beyond the amounts needed by the crop, pose considerable health and environmental risks, at a local and global scale [1–4]. Because of its varied reactive forms, and complex movement dynamics in the soil, aquatic habitats, and atmosphere [5], as affected by management, biological, and environmental conditions– research on the N biogeochemical cycle, and its use efficiency, continues to be an active field of research [6–8]. The complexity of the N cycle at the farm level increases markedly in highly diversified subsistence agriculture and in capital intensive diversified production systems that include the use of crop rotations, a diversity of crop species in time and space, alternative production and soil management practices, the use of organic amendments, cover crops, and the particular socioeconomic conditions of the farm.

Nitrogen plays an integral role in plant nutrition, is involved in plant photosynthesis, in the production of photosynthetic assimilating area to maximize light interception, and is a key constituent of proteins, amino acids, nucleic acids, enzymes, hormones, and chlorophyll [9,10]. Nitrogen regulates root growth, flower formation, canopy development and life-span, fruit set and improves quality [11,12]. A N deficiency reduces transpiration rates and stomatal conductance, as well as carotenoid, chlorophyll and soluble sugar levels; and, reduces the activity of photosynthetic enzymes and of PSII [10]. N containing molecules, such as amino acids, polyamines and nitric oxide (NO), play physiological and signaling pathway roles that contribute to plant defense and stress response mechanisms [13–15]. Nitrate in the soil may also serve as a signal molecule increasing the formation of lateral roots to reach areas of less mobile nutrients. In the plant nitrate also functions as a signaling substance that regulates gene transcription affecting seed germination, root growth, and stomatal activity [16–18]. On the other hand, the overuse of N has resulted in adverse environmental

 doi:10.20944/preprints202312.0768.v1

impacts including atmospheric reactive N deposition and pollution of aquatic habitats, contributing to global warming, reduced biodiversity and disruption of ecosystem functions [7].

Globally about 120 million tons of N fertilizer are applied every year, representing about 60% of all the applied NPK fertilizers [19]. As high-value crops, vegetables represent a unique case, because fertilizer input costs represent a relatively smaller percentage of the total production costs, often 5-10% [20–22]. Thus, as an insurance to minimize risk, vegetable growers have traditionally over-applied fertilizers to ensure adequate yields [23,24], when compared to the rates applied on staple crops such as cereals or root crops, that have a lower per hectare cost of production value. The relative greater fertilizer application rates applied to vegetable crops may partly explain the relative low nutrient use efficiency observed globally for the production of vegetables, as compared to lower-valued crops [25].

One of the most challenging aspects to better understand the N cycle and to improve its use efficiency (NUE) is to dissect and predict the effect of the myriad ecophysiological system x N interactions that are observed on the farm. The degree of multiple-level interactions, which occur both in time and space, and which may vary from farm to farm, makes it a challenge to better understand N cycle dynamics, its impacts on N uptake, and to identify management practices to improve its use efficiency [26]. An improved NUE helps to reduce production costs, the incidence of nutrient imbalances, and environmental N losses [27]. Despite improved genetics and the adoption of improved production practices, N_20 emissions have increased by 64% over the past five decades, with agriculture accounting for 78% of the increased emissions [28], highlighting the importance to improve the NUE in agricultural systems.

Despite the wide diversity in production systems and socioeconomic background, at a global scale vegetable farmers may be divided into two major groups. A prominent group of vegetable farmers, those responsible for most of the international trade and for consumption by the middle class, represent those that rely on high chemical N inputs under both field or protected agriculture. The second group of vegetable farmers include small-scale subsistence farmers, who feed about half of the global population, and who rely more on locally available resources, than on external N inputs [29]. Thus, the different groups of farmers will require distinct approaches to either improve NUE or to improve the fertility of the soil to meet the crop uptake N demand and to minimize environmental losses. Here, building on earlier reviews [24,29–36], we present key management variables and system interactions that should be considered to improve the NUE in vegetable crops.

2. NUE in Vegetable Crops Production

2.1. Background on NUE in Vegetable Crops

Nitrogen Use Efficiency (NUE) is defined as the percent of the applied fertilizer N that is recovered by the current crop (Table 1). The global average NUE for crops ranges from about 50 to 80%, with values varying from region to region, primarily based on socioeconomic conditions [23,27,37]. For example, in China, where high N application rates are 3.3 times the global average for the production of maize, the observed NUE for maize was 25% compared to a 42% global average, and to 68% in North America [38]. Conversely, in Africa where relative low N rates are applied, NUE reaches 80% [27]. The global pooled crop NUE average is estimated to be about 55%, meaning that about 45% of the applied N may be lost to the environment, if soil N levels have already reached a steady state [2,39].

Table 1. Some of the terminology used in the literature to determine Nitrogen Use Efficiency, and as components of N management strategies.

Term	Definition	Sources
Nitrogen Use Efficiency (NUE)	Proportion of applied fertilizer N that is recovered by the current crop; A product of N uptake and utilization efficiency.	[32,40,41]
N recovery efficiency (RE_N)	N harvested in marketable product (dry weight) as a proportion of external N inputs	[19,40,42]

Preprints (www.preprints.org) | NOT PEER-REVIEWED | Posted: 12 December 2023 doi:10.20944/preprints202312.0768.v1

Percent Apparent N recovery	Total above-ground N uptake at maturity at a given N fertilizer rate minus uptake at zero N-rate, divided by the amount of N applied	[19,32,43]
Ndff	Nitrogen derived from fertilizer;	[19,44]
NFUE	N fertilizer use efficiency= (N derived from fertilizer/ applied N rate)* 100	[44,45]
Agronomic NUE	Harvestable biomass production divided by unit of available N (soils and fertilizers); Provides an economic benefits : cost ratio of added rates of N fertilizer.	[12,41,46]
N utilization efficiency (NUtE)	Amount of marketable product per unit taken up by the crop and utilized via remobilization and assimilation	[12,41,47]
N uptake efficiency (NUpE)	Unit of N taken by the plant divided by unit of N available in the soil	[41,46,47]
N harvest Index (NHI)	Ratio of N in harvestable product divided by the total crop N	[46,48]
N balance	Term used to determine the amount of fertilizer N (N_{fert}) to apply, calculated as $N_{fert}= N_{outputs} - N_{inputs}$.	[26]
N budget	Assessment of the major N inputs and outputs on a farm	[49]

In Europe, by following improved nutrient management practices for the production of cereal crops, NUE was increased over two decades from 42% to over 60%, serving as a model to improve NUE in other cropping systems and regions. Cereal yields, over that time span, were increased considerably in Europe, while chemical N applications were reduced by 25% [50]. In Nebraska, USA, when following recommended nutrient management practices, the NUE ranged from 60-70%, as compared to 40% obtained in plots receiving the highest N application rate [51].

To assess the N crop recovery rate from chemical N fertilizers, a multi-year experiment was conducted in the tropics on 13 different locations, in 8 countries, by evaluating the fate of initial ^{15}N chemical N and separate ^{15}N crop residue applications over a five-year crop rotation. During the first year of the study an average of 21% of the applied chemical N fertilizer was recovered by the crop, and 79% was derived from the soil N. After the 5-year period, an average 18% of the chemical N was recovered by the crop, compared to 40% recovered from the ^{15}N-labeled crop residues [52]. Recent ^{15}N rainfed field studies conducted with maize in central Illinois, USA showed that most of the crop N uptake (up to 89%) was provided by the soil N rather than from the applied chemical fertilizers [53]. Similarly, ^{15}N studies showed that about 20% of the applied chemical N was recovered in sugarcane at maturity [54], again highlighting the importance of soil and crop residue mineralization, as a source of N.

For vegetable crops the NUE during the 1970s in California, USA was reported to be less than 50%. During the 1980s a ^{15}N study with lettuce showed a NUE of 12% with a single pre-plant application of 180 Kg N Ha^{-1}, and a 25% NUE when receiving 60 Kg N Ha^{-1} in two split applications [55]. Globally, the estimated NUE efficiency for fruits and vegetables is estimated to be 14%, compared to 46% for maize, and 40% for both rice and wheat [56]. Improvement in NUE for vegetable crop species is attributed to either physiological or morphological plant traits [46,48,57]. Under best management practices in intensive production systems, the NUE for vegetable crops may reach 70% [58].

NUE is dependent on the physiological capacity of vegetable species to uptake, metabolize, and redistribute the necessary amount of N, during the appropriate stage of crop growth, to optimize yields. Three key components that determine the NUE of crops include the N uptake efficiency (NUpE), N utilization efficiency (NUtE), and the N harvest index (NHI) [46,47] (Table 1). These components are affected by the form of N uptake, either NO_3^- or NH_4^+, N translocation within the plant, the reduction of NO_3^-, and the assimilation of reduced N into organic compounds [57].

The ability of crops to uptake N is dependent on a number of interrelated variables such as soil fertility, cultivar, soil moisture, temperature, time of the year, N uptake pattern, the incidence of pests and diseases, farmer expertise, farm socioeconomic conditions, among other variables. In turn, physiological mechanisms that affect NUE include root growth, architecture and N uptake patterns, leaf duration and growth, and N remobilization within the plant [46]. A field study with tomato

 doi:10.20944/preprints202312.0768.v1

conducted in California, USA on an unfertilized field, and previously fallowed for two years, showed a relatively low, 3% root N uptake as a percentage of the total inorganic N available in the sampled soil profile (60 cm x 50 cm x 120 cm), attributed to a general low root density. These relative low root uptake rates indicate a relative low NUE and the potential for leaching of inorganic N, below the root zone [59].

Total N uptake values reported in the literature (Kg N Ha^{-1}) [23,60], and uptake patterns during the growing season, need to be updated on a periodical basis, to develop realistic budgets and fertilizer management recommendations, as new cultivars are introduced [49]. There are also relatively few studies with respect to nutrient uptake patterns and budgets under organic farming systems. It is thus critical that N uptake and NUE efficiency information be developed under low-input or chemical-free farming conditions [8,12,46,61].

For leafy crops, N uptake patterns correspond to the periods of biomass accumulation. In the case of head lettuce, 70-80% of the N uptake demand occurs between heading and harvest [46]. In the case of onion, 9% of the N uptake occurred during the first half of the growth cycle, and 68% of the uptake occurred from the period of bulbing to harvest [62]. The crop uptake pattern of fruit vegetables, such as peppers, follows a Mitscherlich type curve response which includes a lag phase during crop establishment, a vegetative or log phase of rapid nutrient uptake (20-30 days after transplanting), a leveling rate, a peak of maximum nutrient uptake (30-60 days), and a reproductive or senescence stage, with the most nutrient uptake occurring during the last three phases [24,63,64]. During the last stages of crop growth and maturity, both rates of crop N uptake, and of N remobilization within the plant, should be considered to determine their combined effects on NUE and crop yields [40]. Greater rates of remobilization are observed in crops with a reproductive phase. For instance, almost 50% of the N uptake in pepper is partitioned to the fruits [64,65]. It should be recognized that when comparing treatment effects, agronomic nutrient indices are more accurate, with respect to use efficiency, when the soil is in a relative steady-state with respect to nutrient content [66].

As observed in several studies, in general higher N application rates, especially at above optimum levels, results in a lower NUE [43,51,67,68]. Farmers, in general, tend to apply N fertilizers above the recommended rates, to ensure adequate yields, and as an insurance against possible losses [23]. For instance, maize farmers in Northeast China apply an average of 350 Kg N Ha^{-1}, with a range of 210–490 Kg N Ha^{-1}, which is well above the recommended rates of 190 Kg N Ha^{-1} [69]. Similarly vegetable growers in Spain and Florida tend to make N applications at above the recommended rates, under greenhouse and field conditions, resulting in N environmental contamination [70–72]. However, not only is NUE reduced [70,73], but some vegetable species experience a yield depression at application rates above the recommended levels [74], as well as reduced quality and delayed maturity [75,76]. The reported value of NUE for some staple and representative vegetable crops is presented in Table 2.

Table 2. Representative Nitrogen Use Efficiency (NUE) values reported for some vegetable and reference agronomic or staple crops.

Crop	NUE (%)	Region	Sources
Cereals	26-35	China, commercial production, under high N application rates	[38,77]
Maize	36-46	Global; and range, USA	[30,37,56,78]
Rice	29-42	Global; and range	[37,56]
Wheat, Rice	38-42	Global; and range	[37,56]
Cabbage	27-55	Average 40%, Florida	[79]
Cucumber, bell pepper, tomato	54-61	Greenhouse fertigation, Netherlands	[80]
Onion	30-40; 10-26	Netherlands; Colorado, Idaho, USA, varied by timing & method of application	[44,81]
Peppers, bell	30-50	Florida	[82]

 doi:10.20944/preprints202312.0768.v1

Potatoes	40-77; 27-37; 40-60	Range for Low N vs high N rates; Netherlands; Florida; Minnesota;	[12,68,83]
Tomato	15.4 (12 spring; 32 fall),	Sandy soils, Florida, ^{15}N study	[19,84]
Lettuce	12-25	For high (180) and low (60 kg Ha-) N application rates	[55]

2.2. Soil Quality and NUE

Improvements in soil management and fertility impact soil N levels, N soil mineralization rates, crop N uptake and NUE [85]. The interactions between soil management and soil moisture also affects microbial and fauna activity, the N cycle, and NUE. The soil organic matter (OM) contains over 90% of the top 25 cm of soil N, which may represent 1,700 to 1,800 Kg N Ha^{-1}, of which 1-3% may be mineralized on an annual basis [23]. The soil OM level and its replenishment with amendments or crop residues will thus impact the long-term soil N stocks, and mineralization rates. With some crops the soil OM may provide 60-90% of the total crop N uptake, even in fields receiving high N chemical fertilizer inputs [86]. Adequate levels of soil OM also contribute to reduce soil erosion, by helping to stabilize soil structure, with a reported 20-30% reduced erosion with a 1-3% increase in OM levels [87]. The corresponding increase in soil OM levels also contributes to increased yields [88], with a study in a temperate region showing a 12% yield increase for every 1% increase in soil OM [87].

Long-term fertilization with urea or sources of ammonium acidify the soil which may require periodic lime applications to maintain the proper soil pH and soil nutrient balance [89]. A 28-year field experiment conducted in Hunan Province, China showed that unlimed NPK fertilized fields, with urea as the N source, resulted in 15-16% lower levels of pH, OM content, NO_3^- and total N, as well as in lower NUE, as compared to the limed treatments, indicating the impact of low pH levels on soil quality and NUE [85]. In addition, soil management practices that result in soil compaction may restrict root growth, water content, N mineralization rates, and nutrient uptake, and alter the dynamics of microbial activity, resulting in potential adverse impact on NUE [90–92].

Organic, low-input, and subsistence farmers depend less on external N inputs and instead rely on enhancing the soil organic matter content and fertility of the soil to meet long-term crop N uptake requirements. Ecologically-based management practices are prescribed to improve the long-term soil fertility, organic matter content, and NUE on the farm (see Section 4.0) [8,29]. .

2.3. Effect of N Form on Crop Growth and NUE

Plants uptake N as NO_3^-or NH_4^+ and in soils NO_3^- is generally the predominant N form for plant uptake. Crops normally uptake more NO_3^- than NH_4^+. Through the process of mineralization the N found in organic compounds is converted to NO_3^- or NH_4^+. The bacterial led process of nitrification converts NH_4^+ into NO_3^-, especially under warm and well-drained soil conditions. Thus under cooler soil conditions the NH_4^+ : NO_3^- ratio may be greater than during warmer months. Ammonium may also be dominant under acidic and aerobic conditions [93], and at high levels may cause toxicity on plants [23]. Similarly, applied urea in the soil hydrolyses into NH_4^+ which may be uptaken by plants,or is converted via nitrification into NO_3^-.

For many crops a combination of NO_3^- and NH_4^+ improves crop growth and quality [23,93], and the particular optimal ratio will vary by crop species and growing conditions, impacting its NUE, as described in Table 3. To minimize leaching, especially on sandy soils, it is recommended that nitrate represents 25-50% of the applied fertilizer [68]. The NO_3^-: NH_4^+ ratio has an effect on the uptake of other mineral nutrients. Increased ratios of NH_4^+ may reduce the uptake of mineral cations and increase the uptake of mineral anions. A higher ratio of NH_4^+ may reduce the uptake of calcium, which may cause some physiological disorders such as blossom-end rot in tomato and pepper [94,95].

Most N form studies have been conducted under greenhouse conditions and with particular crop varieties, thus, further field-based studies are required to confirm results observed under protected conditions. Results on the effect of N form conducted under protected conditions don't

Preprints (www.preprints.org) | NOT PEER-REVIEWED | Posted: 12 December 2023 doi:10.20944/preprints202312.0768.v1

necessarily correspond to those obtained in the field [96]. However, evaluations of the N form under field conditions is challenging due to the inherent dynamic nature of the soil NH_4^+ : NO_3^- ratios as affected by differential crop uptake, microbial transformations, and NO_3^- movement in the soil profile [59,96]. An unfertilized field study conducted with tomato in California, using several methodologies to assess N soil dynamics, found that NO_3^- levels were 10-20 times higher than for NH_4^+ in the top 15 cm soil layer, with the available NH_4^+ being quickly nitrified into NO_3^-. The available NH_4^+ pool appeared to be mostly utilized by microbes rather than for crop uptake. Because of the relatively high levels of NO_3^- in the soil profile, the authors predicted that considerable N losses would be likely to occur due to leaching below the root-zone [59]. Recent ^{15}N field studies conducted in Illinois, USA with different N sources determined that NUE was greater for NO_3^- than for NH_4^+ and urea fertilizer sources. The greater environmental losses observed for urea, resulting in the lower NUE, was attributed to greater NH3 volatilization than for the NH_4^+ N sources [53].

Plants may also uptake organic sources of N, such as aminoacids or proteins, and the levels of soluble organic N may contribute a portion of the overall N budget [8,86,97]. The predominant available N form under organic systems may vary depending on the production system. For instance, under aged manure or pig slurry fertilizer programs the proportion of NH_4^+ in the amendments may reach 75-80%, with the balance of the total N supply for crop uptake complemented depending on the soil properties, crop rotation or field history, and management system [98].

Table 3. The effect of N form, and NO_3^- : NH_4^+ ratios, on the growth of some representative vegetable crop species.

Crop Species	N form	References
Cucumber, Cucurbits	NO_3^- improves growth while NH_4^+ depresses growth (likely due to a lower pH in the root zone)	[99,100]
Flowering Chinese Cabbage, *Brassica campestris* L. ssp. *chinensis* var. *utilis* Tsen et Lee	Improved yields with a combination of NO_3^- : NH_4^+ ratio; Improved yield at 10 : 0 and 9 : 1 ratios; Improved NUE at 9 : 1 ratio; An earlier study found best growth at 1 : 1 ratios and marked growth reduction at 1 : 9 ratios	[93,101]
Chinese Kale, *Brassica alboglabra* L. H. Bailey	Improved growth at NO_3^- : NH_4^+ ratio of 3:1 and 9:1; Improved NUE at 3:1 ratio; Inhibited growth at high NH_4^+ ratios.	[102]
Lettuce, leafy	Head fresh weight higher with NO_3^- : NH_4^+ ratios of 1:1 or 1:0; Greatest NUE at 1:0 or 1:1 ratios	[103]
Onion	No consensus on effect of N form on yield; An earlier study found that NO_3^- alone or in combination with NH_4^+ improved plant growth; bulb weight highest with NO_3^- : NH_4^+ ratio of 3:1 to 1:3	[81,104]
Pepper	Yields highest when NO_3^- is the predominant N form, and with increased NO_3^- : NH_4^+ ratio	[105,106]
Strawberry	Fruit yield greatest at NO_3^- : NH_4^+ ratio of 3:1 to 1:1	[107]
Taro, *Colocasia esculenta*	Improved growth with 75:25 or 100:0 NO_3^- : NH_4^+ ratios	[108]
Tomato	Trend to improved yield (but not significant) with 1:4 NO_3^- : NH_4^+ ratio; Improved fruit quality with organic N source or with 1:4 NO_3^- : NH_4^+ ratio	[16,109]

2.4. Crop Improvement for NUE

Given its global economic importance, with a trade value of over USA $59 billion [110], maize is among the most extensively researched crop species in terms of the impact of breeding, management and fertilizer practices on crop yields. The five-fold improvement in maize yields over the past century are attributed to be about 60% due to genetic improvement, and 40% to improved management practices [111]. While early yield increases in maize were achieved by increasing fertilizer rates, application rates have remained static over the past 4 decades in the USA, reflecting on an improved NUE [47]. However, NUE improvements were not a goal in maize breeding programs, but rather occurred through the adoption of improved management practices such high planting densities, or selection for increased harvest index, resulting in increased N partitioning to

Preprints (www.preprints.org) | NOT PEER-REVIEWED | Posted: 12 December 2023 doi:10.20944/preprints202312.0768.v1

the ears [46,47]. Similar improvements in NUE were reported from long-term breeding programs in wheat, perhaps due to an increased N Harvest Index, and improved management practices [112]. An evaluation of the available germplasm indicates the potential for increasing the NUE in cereal crops, through targeted crop improvement [112,113].

It has been well recognized that germplasm diversity exists within vegetable species with respect to nutrient rates and NUE [57] with respect to crops such as okra, pumpkin and tomato (Table 4) [114]. However, breeding for NUE has not been a component of improvement programs for most crops, likely because it is a complex genetic trait, controlled by multiple genes, and because of the difficulty in managing the many environmental variables that affect N form, movement, and cycles in the farm [46]. In many cases the potential to select for improved NUE among modern cultivars has been diminished as the desirable traits have been bred out, after many years of selecting for yield response to high N application rates, which may prescribe for the inclusion of a wider genetic pool, such as wild relatives and landraces, in the selection process [46,115–119]. While most research to date has focused on increasing NUE under conditions of high N fertility, future breeding work may include characterization and selection for high NUE under limited N availability [57,116], as is often the case under low-input or in organic production systems [8,120,121].

Table 4. Profile of breeding and germplasm selection on representative vegetable crops, with respect to Nitrogen Use Efficiency.

Crop	Notes	References
Cabbage	Evaluation of N harvest index, and selection criteria of growth parameters	[46]
Cassava, *Manihot esculenta*	Genotype with improved N uptake was identified among 25 accessions, under low N conditions; Molecular analysis identified mechanisms for possible improved N uptake in efficient lines	[18,122]
Eggplant	NUE efficient genotypes identified from germplasm collections and crosses	[116,123]
Lettuce	Evaluation of wild species, plasticity in response to environmental variables	[46]
Onions	NUE evaluation of landraces	[118]
Potato	Identification of growth traits associated with NUE among a range of potato cultivars; Differential cultivar responses to N rates; cultivar selection and N efficiency under organic systems	[12,46,83,124]
Spinach	Identification of QTLs for NUE	[46]
Tomato	High NUE genotype identified among 14 landrace varieties; wild relative genotype identified with increased NUE under low N rates; Genotypes with improved NUE with potential use as rootstocks; greater N uptake in drought tolerant cultivars	[115,119,125,126]

3. Strategies to Improve NUE in Vegetable Crops

3.1. Development of Fertilizer N Recommendations for Vegetable Crops

Experience over the past decades indicates that the adoption of best nutrient management practices improves the NUE for individual crop species, and at a regional level [50,51,58]. Soil N tests are often unreliable to determine N rate recommendations, because of its rapid chemical transformations, such as denitrification, immobilization, mineralization, and volatilization, especially in warm climates [19]. Nitrogen fertilizer rate recommendations are thus developed based on yield response curves, or calibration studies, based on a range of N application rates [8,19].

N application recommendations are based on an understanding of the N cycle at the farm and landscape level [8,127], on nutrient budgets, and on calibration studies to determine crop response curves to a range of N application rates (Table 5). Data required to develop adequate nutrient budgets and calibration curves include total N uptake amount over the growing season (Table 6); patterns of N uptake, based on crop phenological stage; tissue N levels during particular periods of crop growth; and N residual levels or mineralization rates for representative soils in the region.

Preprints (www.preprints.org) | NOT PEER-REVIEWED | Posted: 12 December 2023 doi:10.20944/preprints202312.0768.v1

Table 5. Approaches that are followed to develop N rate recommendations, to improve yields and Nitrogen Use Efficiency (NUE).

Research protocol	Notes	References
Studies on N cycle at a farm and landscape level	China; long term rotation, Europe; Organic systems, California, USA; Canada	[127–130]
Nutrient budget or balance studies	Oregon; Balance (organic systems); Europe; Canada; Florida	[26,49,64,131–133]
Timing of application (uptake patterns)	Oregon; Montana	[64,134]
Calibration (growth N response curves)	At experiment station or on-farm trials; Oregon	[64,135,136]
Soil and tissue analysis	Includes soil nitrate testing, Oregon; Brazil; Florida; plant monitoring, e.g. chlorophyll meters, petiole NO_3^{-1} sap analysis, canopy sensors	[26,64,137–139]
Placement of fertilizers	Montana; Europe	[24,134]
N fertigation guides	Greenhouse; Europe; India	[23,24,74,80,140]
Modeling, crop models, decision support systems (DSSs)	Onions, Brazil; Europe; Florida; Organic rotations and N budget, UK.	[24,26,138,141,142]
Use of optical sensors	Maine	[143]
Adoption of integrated or best management practices (BMPs)	Oregon; Europe; Florida	[19,24,26,50,64,82]

Table 6. Reported Nitrogen uptake rates for representative vegetable and staple crops (marketable product), as reported in the literature.

Crop	N uptake (Kg Ha^{-1})	Notes	References
Broccoli	50-90	Oregon, USA	[64]
Brussel sprouts	170	Organic, Europe	[61]
Cabbage, head	130-230	Europe	[46]
Cassava, *Manihot esculenta*	124 (55-62 roots, 202 entire plant)	Tropics	[88]
Lettuce	105; 100-110	Organic, Europe; California	[46,61]
Onions	60-120, 160	Oregon, California, USA; Brazil	[62,64,144]
Potatoes	130; 80-130; 220	Organic, Europe; Oregon, USA; California, USA	[46,61,64]
Spinach	20-90	Leafy baby & processing, California, USA	[46]
Tomato	3.9-4.4 (fruit) 51.8-72.2 (whole plant)	Florida, USA	[19]

A challenge to develop appropriate N fertilizer recommendations for vegetable crops is that calibration data or N response curves are location [136,138], crop species, cultivar, and production system specific (such as plasticulture vs. bare-ground culture, or conventional vs. organic). Calibration research should also be revisited periodically, as new cultivars are continually introduced, as production practices are adjusted, and as new technology is adopted to improve production efficiency. Fertilizer calibration data are lacking for most production areas, especially in the tropics. When available, calibration data are often only available for a few selected crops of major economic importance, or are outdated, based on field research that may be years to decades old. When local calibration studies are unavailable, extensionists and crop advisors often rely on studies conducted in other regions or countries. In practice, in most instances and locations, decisions on the rates and timing of N applications are based to some degree on calibration studies conducted elsewhere, and largely on the experience of local growers, consultants, and extension advisors (Table 7) [26]. As a consequence, it is likely that the lack of currently available calibration data, for specific

Preprints (www.preprints.org) | NOT PEER-REVIEWED | Posted: 12 December 2023 doi:10.20944/preprints202312.0768.v1

crops and production areas, results in farmers over-applying N on their farms, resulting in lower rates of NUE and in respective economic and environmental losses.

Table 7. Recommended N application rates for representative vegetable crops.

Crop	Application Rate (Kg Ha^{-1})	Comments	References
Cabbage, head	220-350	Europe	[46]
Carrots	120; 100-150	California, 110-116 day season; California (Oxnard); Florida, sandy soils	[145,146]
Celery	200-400	California	[147,148]
Cucurbits	60-100; 90-170 (summer squash); 150	Wisconsin; California; Florida	[149–151]
Lettuce	120-220	Europe	[46]
Onion	170-200; 200;	Pennsylvania and Utah; Washington State	[118,152–154]
Pepper	150-200; 200-240; 300	Georgia; Florida, higher rates with extended season; Puerto Rico	[63,82,155]
Potato	120-180; 150-250; 200-250; 250-300	California; Florida	[12,46,68,147]
Spinach	140-290	Europe	[46]
Tomato	110-225 (plus weekly maintenance applications of 10 kg for staked tomatoes); 160-200	California, fresh market; Florida and Eastern USA	[19,156–158]

3.2. Best Management Practices to Improve NUE

Research on NUE indicates that environmental losses may be reduced by 15-30% by adopting improved management practices [41]. A global meta-analysis estimates that NUE may be increased by improving nutrient (by 27%), crop (6.6%), and soil (0.6%) management practices. The analysis estimates that NUE may be improved globally by 30% by adopting improved production practices [27]. These values and improvements would vary by cropping system and region, but point out the potentials for improving crop NUE, by adopting best management practices (BMPs). Recommended practices to improve NUE under intensive commercial vegetable production systems include to follow what are considered to be BMPs such as adopting the right source, rate, timing, and placement of N fertilizers, along with management practices such as tissue testing and fertigation-based plasticulture systems [19,27,159–161]. Conversely, production factors that may decrease NUE include poor seed-bed preparation, unadapted cultivars, poor seed quality and stand establishment, improper irrigation or drainage, as well as pest and weed infestations [162].

An initial step to estimate N application rates for a specific crop with realistic yields is to develop a nutrient budget by subtracting the total amount needed by the crop by the estimated residual levels already available in the soil [71]. A management program is designed to synchronize the levels of available or applied N, with the estimated demand of the crop, during the different phenological stages of development. Tissue analyses may be conducted during the growing season to monitor the crop's nutritional status, to further fine-tune the N management program [24]. The ultimate level of NUE reached in particular locations will vary depending on farmer expertise and on field conditions. However, when particular management practices are adopted uniformly by all farmers, NUE tends to increase area-wide. Experience indicates that with many crops it is possible to increase NUE by 30-50%, with the adoption of recommended management practices [66].

While soil N tests are normally unreliable to estimate soil N levels [59], some locations, especially in temperate areas, have adopted the use of pre-side dress Nitrate-N tests, to estimate available N soil levels, prior to planting. Multiple-field trials conducted in California, USA with cool season vegetables, determined that at NO_3^- soil levels above 20 ppm, initial side-dress N applications could

 doi:10.20944/preprints202312.0768.v1

be delayed without affecting crop growth [26,163] . In addition, system-wide management practices may be adopted, such as alternative soil management or tillage; alternative cropping systems such as double cropping or intercropping; integration of livestock and crop production; alternative water management; as well as agroecological systems that may help to improve internal N cycles and use efficiency [8,164]. A profile of several recommended management practices to improve NUE is presented in Table 8.

Table 8. Profile of representative management practices to improve Nitrogen Use Efficiency in vegetable cropping systems.

Production Practice	Notes	References
Optimize N application rates based on crop demand	Improved NUE	[27,82,164]
Selection of adapted crop varieties	Potato & lettuce germplasm; Selection for roots systems with improved N uptake	[124,165,166]
Controlled- or slow-release fertilizers	Utilizing nitrification inhibitors, not cost-effective in some systems	[24,68,81,167,168]
Combined use of organic and chemical fertilizers	Improved NUE, soil fertility, and use of local resources,	[27,169]
Placement of fertilizers	Field placement including subsurface drip fertigation	[24,81,164,170,171]
Planting density, spacing	Spacing x N response interaction, onions, tomato	[172–175]
Plasticulture systems and fertigation	Improved NUE	[65,158,170,176,177]
Timing of application to synchronize with crop uptake demand	Tomato, onion, including split-applications	[19,23,45,58]
Precision farming	Assessment of yield variations across a field; N status monitoring systems; May not be cost-effective in some systems or for small-scale production	[159,178,179]
Grafting	Improved NUE, melons, tomato	[26,125,180]

3.3. Crop Rotations Effects on NUE

Crop rotations have historically been used to manage soil fertility, to manage pest and weed infestations, and to diversify the farming operation [181]. By incorporating species with complementary nutrient use patterns, root traits, and microbial rhizosphere assemblage, crop rotations help to improve the N cycle within the farm; the soil structure; organic matter, nutrient and water contents; NUE; reduce residual N losses; and, improve yields [8,24,181,182]. A ^{15}N field study conducted in Nebraska, USA, showed that a combination of low N application rates and complex rotation schemes improved internal organic N cycling while maintaining similar N mineralization rates as those obtained under high N rate treatments, showing the potential of complex rotations to reduce N losses and to improve NUE [183].

While residual N levels may remain from the previous crop in fertilized fields, the ultimate management goal, especially on sandy soils, aims to minimize N residual levels by fine-tuning chemical N applications to the actual needs of each crop in the rotation [184]. Rotation ^{15}N studies in sandy soils have found negligible levels of N recovery from the chemical N rates applied on the first crop; with the second crop in the rotation recovering about 2% of the initial ^{15}N application, to <1% recovery for further subsequent crops in the rotation [19]. In a double-crop tomato experiment conducted in northern Florida, USA, in which a ^{15}N labeled fertilizer was applied only for the first crop, the ^{15}N fertilizer Recovery Efficiency for both seasons ranged from 10.7 to 15.4%, and thus the levels of unrecovered N ranged from about 85-90%. At the highest N application rate in the experiment, about 90% resulted in environmental losses, attributed to leaching or volatilization [19]. With such potential chemical N losses, given that farmers tend to over-fertilize their crops, alternative management practices may be adopted, such as changing the fertilizer input, and including cover

 doi:10.20944/preprints202312.0768.v1

crops or catch crops in the rotation, to make better use of residual N levels and to improve soil tilth and NUE [24,120,181,185,186]. For example, a two-year legume-based rotation field and modeling study conducted in northern Florida, USA showed that N application rates could be reduced by 26% and that N leaching was reduced by 37%, without sacrificing yields, under the rotation and irrigation scheduling program [187].

The fertility of the soil, residual N availability and NUE may also be improved when legumes are incorporated into a rotation [24,120]. Six-year rotation on farm experiments conducted in Malawi showed improvements in several soil fertility variables when pigeon peas (*Cajanus cajan*) and peanuts (*Arachis hypogaea* L.) were included in the rotation, including greater levels of mineralizable and microaggregate Carbon [121], indicating the potential of legume-based rotations to improve NUE and long-term productivity.

4. Agroecological Practices to Improve NUE

4.1. Organic and Low-Input Farming Practices Effect on NUE

Nitrogen is among the most limiting production factors in organic farming with a modeling study on potatoes showing that 48% of the yield variability could be explained by the N availability [12]. Ecologically-based management strategies on farms that rely on little or no chemical N inputs, revolve around improving the long-term fertility of the soil. Suggested ecological soil management principles include to build the soil organic matter content and nutrient reserves; to increase vegetational diversity in time and space, to build synergistic and multifunctional interactions; to minimize soluble N pools that may be susceptible to environmental loss; and to develop mass N balance budgets at the farm and landscape level to better understand and improve nutrient flows in time and space [8,29].

The synchronization of soil and organic amendment N mineralization levels with the timing of crop uptake is a major challenge for organic production [8,131,188]. Variables that affect the improved synchronization of available soil N and crop uptake include the soil fertility, root growth and architecture, and the selection of crop species adapted to lower N soil conditions. Plant traits that may improve adaptation to low N soils include a higher root to shoot ratio, slower growth rates, and effective microbial associations. In turn, organically managed soils have in general lower mineral N levels, but have a greater mineralization potential due to a greater organic matter content [120].

A three-year organic rotation study with several vegetable crops, conducted in Portugal, concluded that split-applications of fast N release nutrient sources, and increasing the legume/cereal ratio of cover crops, to improve N mineralization rates, improved N uptake and NUE [131]. A survey of organic and conventional farms in Germany found less surplus N levels, subject to environmental losses, and a greater NUE in the organic farms. Overall the highest NUE and lowest surplus N levels were observed on the integrated crop-livestock organic farms [132]. A ^{15}N experiment that compared organic and conventionally managed soils in Switzerland found a greater NUE in the organically amended soils (93%) compared to the conventional ones (55%), attributed in part to the slower rate of N mineralization from the organic amendments. Soil fertility attributes that linked the organic soils to a greater NUE included microbial activity, organic matter content, and aggregate size [189]. Conversely, soils with a reduced microbial activity resulted in a 20% reduced N crop uptake, increased leaching (by 65%) and emission losses (94%) [190], which highlights the importance of soil fertility and microbial activity on the N cycle and NUE in organic farms. The importance of long-term soil ecological management under organic legume-based cover crop rotations as the sole N source, was also observed in a recent study, showing a near balance of N inputs and outputs, and improved soil N mineralization rates, likely a result of improved N cycling and NUE [191].

Organic residues and amendments can provide a considerable source of N, contribute to build up the soil organic matter and may improve the system NUE [192]. The selection on the type of amendment used on the farm depends in part on the N mineralization rates, to meet crop uptake demand during the different growth stages. N mineralization rates in temperate areas, during the first month after application range from 60% for specialty organic sources such as feather meal, to 20-

 doi:10.20944/preprints202312.0768.v1

40% for legume residues and poultry manure to less than 5% for composts. By one year after application, N mineralization rates range from 100% for specialty soluble organic sources to 50% for legume residues and poultry manure, to 5-10% for composts [64]. Alternative organic amendments, such as seaweed, may be used as N sources, which may also contain growth promoting or biostimulant properties [193–195]. Examples of other amendments, as effective sources of N on vegetable crops include composts and vermicompost; commercial manure sources such as chicken pellets; soybean meal in combination with Effective Microorganisms; and, commercial fertilizer mixes, such as a combination of feather, meat, and blood meal [144,196,197]. A combination or readily soluble N sources with organic amendments may help to better synchronize N availability with crop uptake, by immobilizing N during the early growth stages, and by mineralizing N during the later stages of high N uptake demand, minimizing environmental losses, as observed in a ^{15}N experiment with Chinese cabbage [198]. A description of some organic and agroecological management practices that may be adopted to increase NUE is presented in Table 9.

Table 9. Profile of selected agroecological management practices that may help to improve Nitrogen Use Efficiency.

Production Practice	Notes	Source
Organic systems	Less surplus N levels in organic systems, improved NUE	[132,189]
Legume-based rotations	A more balanced N budget, improved N cycles	[29,191]
Organic amendments	Locally available, source of N and improve soil organic matter; serve as a slow-release source of N	[8,24]
Cover crops	Soil fertility and improved N cycles	[24,199,200]
Intercropping systems	Improved N cycling, NUE, soil microbial interactions	[24,201–204]
Integrated crop-livestock/aquaculture systems	Improved Nutrient cycling, economic diversification, resilient systems	[29,205]
Agroforestry/alley cropping	Improved nutrient cycling, less N losses	[206–208]

4.2. System Diversification to Improve NUE

Vegetational diversification can help to improve N cycles and NUE at the farm and landscape level. Diversification consists of introducing species, in time and space, that are complementary in terms of resource use with respect to space, light, water and nutrients [209,210]. The complementary nature of the species results in a synergistic resource and nutrient use efficiency, through niche complementarity, imparting a greater level of productivity and resilience to the agroecosystem [211].

Cover crops represent an efficient and practical way to incorporate crop diversification within a farming operation, as part of a rotation program [24]. Cover crops may help to improve soil organic matter content and fertility, resulting in improved N cycles, NUE, and yields [27,121,200,212]. Cover crops may consist of legumes that contribute N via biological N fixation; grasses or other non-leguminous species that help to break disease or weed cycles and to serve as 'catch crops' of residual N; and legume-non-legume combinations, to obtain the partial benefit of both types of cover crops [8]. In addition, cover crops provide many other ecological services to the agroecosystem [200,213,214]. As is the case with the use of organic amendments, the rate and timing of N mineralization from cover crop residues is important to meet the N demand of the following crop, as well as to better synchronize the N residue mineralization rates with the demand during the different growth stages of the cash crop [215]. The C:N ratio of the cover crop residues determines their rate of mineralization. Legume residues in general have C:N ratios between 12 and 25 which promote mineralization while cereal C:N ratios above 80 lead to immobilization [120].

In lettuce, an improved NUE was observed when following a cover crop mixture of rye and hairy vetch, as compared to single species cover crops, and this was accompanied by apparently improved microbial activity, as indicated by greater microbial β-glucosidase enzyme activity, which likely regulated the mineralized N pool from the crop residues [216]. In an eight-year experiment conducted in Salinas, California consisting of organic vegetable rotations, a rye-legume cover crop

 doi:10.20944/preprints202312.0768.v1

combination also resulted in improved NUE. However, because of the relatively high levels of external inputs, the legume cover crops increased the surplus of N in the system which may result in environmental losses, while rye was more effective as a cover crop, to minimize N surplus levels from the system [217]. Based on multi-year cover crop evaluations conducted in Japan, including ^{15}N studies, a combined 2:1 hairy vetch and rye planting ratio was also found to improve the mineralization and synchronization of N release by the individual species, with the N uptake demand of tomato, along with an increase in the overall soil N pool and N cycling [218].

A ^{15}N isotopic crop residue 4-year field study conducted in Washington State, USA, determined the level of N contribution from *Brassica hirta* mustard cover crops to a following potato crop. Overall, the Brassica cover crops recovered 34 to 51% of the 56 Kg Ha^{-1} of ^{15}N applied. About 30% of the N in the Brassica cover crops was released for the following potato crop, representing a contribution of 30-40 Kg Ha^{-1} of N [219], indicating the potential on non-legume cover crops to serve as catch crops of N for the following crop in the rotation [220]. A two year field experiment conducted in Massachusetts, USA also showed that adequate yields could be reached with potatoes with reduced N application rates of 75 kg N Ha^{-1} when following a legume and of 150 Kg M Ha^{-1} when following a forage radish cover crop, showing an improved NUE with these cover crops, compared to the standard rates of 225 Kg N Ha^{-1}, required to reach the same yields without a cover crop in the rotation [221].

Additional strategies to increase vegetational diversification in vegetable agroecosystems, in time and space, with the goal of improving system resilience, soil fertility, N cycles, and NUE include intercropping [24,209], alley cropping [206], agroforestry [207], as well as crop-livestock integrated systems [8] (Table 9). Diversified systems of production are more often found in small-scale conventional or organic farms as well as in subsistence farms of the tropics, but are increasingly being adopted in some temperate regions with the goal of establishing more resilient systems that may be able to better cope with the environmental challenges and fluctuations posed by climate change.

Well designed intercropping systems may result in improved N cycling and NUE by incorporating crop species with complementary growth habits and nutrient uptake demands, such as by interplanting deep-rooted with shallow-rooted crops [24,202,222]. Effective intercropping systems may maintain productivity under a lower N soil status or application rates, through an increased NUE [223]. The species composition and design of the intercropping system, such as the planting patterns, will determine whether the complementary resource use between the species is dominated by above or below-ground interactions [224].

Intercropping systems showed an improved NUE with cabbage and faba bean (*Vicia faba* L.) intercrops [203]; an improved Land Equivalent Ratio for NUE (LER_{NUE}) in a tomato and corn intercropping system [225]; reduced N applications maintained yields in an organic cauliflower and grass-legume mix system, minimizing the risks of leaching [201]; however an organic strip intercropping system of faba bean and tomato did not provide consistent positive effects on tomato N uptake, as the N mineralization rates from the companion faba bean intercrop, fluctuated throughout the experimental period [226].

Integrated crop-livestock systems have been an integral part of many traditional cropping systems around the world for thousands of years, improving the cycling and use of nutrients at the farm or landscape level [29,159,205,227]. For example, an early analysis of nutrient flow in traditional integrated systems followed in Jiangsu Province, China showed that fish waste, from seven different species, provided over 25% of the crop N inputs, while pig waste supplied an additional 13% of the N input for crops and forage production, in addition to contributing to the soil organic matter content [227]. Aquaponic systems utilizing fish effluent as a nutrient source have been shown to improve crop growth and NUE, compared to other nutrient sources [228]. However research in Alabama, USA showed a greater NUE with the conventional fertilizer treatment, attributed to possible greater nitrification rates from the fish effluent, which indicated a need to better fine-tune their system [137]. Table 9 provides an overview of alternative diversification strategies and their effects on the N cycle [8].

 doi:10.20944/preprints202312.0768.v1

5. Nitrogen x System Interactions That Affect NUE

The uptake and use efficiency of N is affected by a myriad of multi-level production, environmental, genetic, and ecophysiological interactions. The N uptake and use efficiency may be affected by variables such as the incident light level and shade adaptation [229], soil type and fertility, planting distance, crop variety, timing of application, cropping schemes, and management practices. The bulk of the research on N applications over the past 70 years focused on yield maximization based on vegetationally simplified agricultural systems, following a reductionist research approach. New directions for research aim to better understand the ecophysiological basis of nutrient x cropping system x environmental interactions, often under diversified and chemical-free production systems, to minimize environmental losses, to improve use efficiency, and to establish more resilient systems that will be better adapted to the environmental fluctuations and extremes posed by the specter of climate change [8,29,230].

5.1. Water Use x N Interactions

Because of its solubility, transformation and mobility of the various reactive N forms in the environment, water x N interactions are among the most important determinants of potential environmental losses and its synchronization with crop uptake, to improve NUE. Crop N uptake is strongly affected by the soil water supply [43]. In general, as observed with potato, increased irrigation rates and N applications result in decreased water and NUE [231]. Excessive N application rates early in the growing season may result in considerable leaching losses from rainfall or surplus irrigation [69]; anaerobic conditions after heavy irrigations may lead to denitrification losses; and deficit soil moisture from droughts or uneven irrigation, may restrict N uptake, resulting in lower crop yields or quality. Farmer education and management guidelines to improve both water use efficiency (WUE) and NUE are important, because as with the use of fertilizers, farmers tend to over-irrigate their crops, resulting in potential excess N losses [24,232]. Furthermore, improvements in WUE and NUE are often correlated variables that tend to result in high yields, as water mobilizes nitrate in the soil, making it available for crop uptake via mass flow and diffusion via transpiration [74,233].

To maintain crop productivity while minimizing losses, current research efforts aim to improve both water and N use efficiency [74]. The type of irrigation affects several production variables and N uptake dynamics. In potato, sprinkler irrigation improved marketable tuber quality by reducing water stress, lower soil temperatures, and improved N management, as compared to furrow irrigation [231]. Irrigation can also be employed in some cases to reduce ammonia N volatilization, such as when irrigating a field after an application urea, reducing losses by 65-95%, depending on the urea formulation [234].

^{15}N studies showed that crop NUE may be improved with the integrated timing and rates of both the irrigation supply and N applications, resulting in improved yields and lower N losses [77]. Tillage and organic amendment use practices may also have a positive impact on water use and NUE. A four year experiment under rainfed agriculture showed that the combined practice of no-tillage and application of organic amendments improved soil moisture conservation, resulting in equal or improved yields, and with increased water and NUE, as compared to conventional tillage and the use of chemical fertilizers [235]. Other management practices on the farm, related to the available soil moisture content, also affect the crop response to N applications. For example, under rainfed systems, the NUE is increased when basal N applications are made after the start of the rainy season has begun [236].

The determination of the appropriate irrigation and N application rates is an active field of research, such as for the production of high-value specialty crops, with the use of organic N sources, and under protected cultivation [237]. The more precise and timing application of N may increase its use efficiency [238], but the particular rates and practices may be cultivar and location specific [231]. Research in Ghana showed that weekly drip fertigation, as compared to sprinkler irrigation or two fertigation events, improved the NUE in okra [171]. To fine-tune a tomato fertigation program, research in Florida, USA determined that a pre-plant basal N application representing 25% of the

Preprints (www.preprints.org) | NOT PEER-REVIEWED | Posted: 12 December 2023 doi:10.20944/preprints202312.0768.v1

total application rates improved yields compared to a full fertigation program. Also, a partial deficit irrigation treatment, consisting of limited watering rates during the first 4 weeks of growth after transplanting, resulted in similar yields to those obtained under standard irrigation rates [239]. Research conducted with cucumber and tomatoes in Beijing, China showed that by limiting the irrigation rate by calibrating its application with the soil matric potential did not affect yields and improved the NUE when compared to the traditional and greater irrigation rates [240]. Conversely optimal yields with field grown cucumbers grown in Egypt were obtained with the high irrigation rate, at 1.0 of crop evapotranspiration (ET), compared to lower ET levels, and with a combination of chemical and organic amendments at high N rates of 315 Kg Ha^{-1} [241].

Plastic and organic mulches may be used effectively to improve both water and NUE, along with other benefits to the production system, such as weed control [43,69]. An analysis on the effect of mulches on cereal crops showed that mulches overall increase WUE, especially under conditions of adequate fertilization [43]. A similar review of the cereal literature found that mulching increased NUE from 4-18%, compared to non-mulching treatments, especially in arid and semi-arid regions [43]. A study conducted in a rainfed arid region of central Mozambique found a differential effect in the response to organic mulch applications, depending on the soil moisture holding capacity of the soils. Crop residue applications increased the WUE and NUE in soils with a low water holding capacity, while these variables were reduced when residues with a high C:N ratio were applied on soils with a high water holding capacity, apparently fixing the available N for their microbial decomposition [242].

Vegetational diversification in time and space may be managed to improve soil water relations. Research indicates that the Water Use Efficiency and corresponding NUE of some crops may be improved with crop rotations [243], including a reduction in nutrient losses [244]. Reflecting on the current international efforts to optimize water and N use, their interaction has been recently evaluated for broccoli [245]; cucumbers [246]; African eggplant (*Solanum aethiopicum* L) [247]; onions [248,249]; and, tomato [239] (Table 10).

Table 10. The effect of selected Nitrogen x Water management interactions on Nitrogen Use Efficiency.

Interaction Variable	Notes	References
Crop responses to N x water interactions	Broccoli (tunnels), Cabbage; Greenhouse cucumbers; African eggplant (*Solanum aethiopicum* L); Onions; tomato	[239,245–247,249,250]
Cultivar selection	Selection of early maturing cultivars during drought periods	[251]
Fertigation	Improved NUE	[24,171,238]
Irrigation system selection	Effect on moisture uniformity, WUE x N x yields	[231,249]
Irrigation rates and timing	Irrigation x N interaction	[239]
Organic Mulches	Reduced evapotranspiration and erosion; moderates soil temperature and moisture; N cycling	[43,242]
Plastic mulches	Reduced evapotranspiration and erosion; moderates soil temperature and moisture	[43,69]
Rotations	Water and NUE	[243,244]
Tillage	Moisture retention under no-till farming and NUE	[235]

5.2. Nutrients x N Interactions

Nitrogen interacts with other nutrients in a way that may impact competition for nutrient uptake, crop physiology, quality and yields, and NUE. In general, high soil N levels may have an adverse effect on the uptake of potassium (K), boron (B) and zinc (Zn), while high levels of K or magnesium (Mg) may reduce the uptake of N [252]. The type of N form has a differential effect on nutrient uptake. Under high NH_4^+ levels or NO_3^- deficiency there is a reduced cation uptake of K, Mg, and Ca, while there is an increase anion uptake of phosphorus (P), sulfur (S), and micronutrients, with the reverse observed under high NO-3 levels [99,105]. While nutrient x nutrient interaction

 doi:10.20944/preprints202312.0768.v1

studies conducted in the greenhouse provide valuable insight, they should be treated with caution, as different responses may be observed under field conditions, and may vary, to a degree by environment, soil type, and cultivar, among other possible variables.

Some crops, like cassava require proportionately more K than N, to maintain adequate yields, so low K levels often become the limiting nutrient factor to reach higher yields [88,253]. In this case, adequate levels of K will be required to optimize crop growth, yields, as well as NUE. In pepper, as observed with other Brassica and solanaceous crops, calcium uptake was reduced with incremental increases of NH_4^+ as the source of N, with yield reductions observed with NH_4^+ : NO_3^- ratios greater than 50%. The lower fruit levels of Ca observed when NH_4^+ is the primary N source may also lead to a greater incidence of blossom-end rot in fruit vegetables, further reducing potential productivity and NUE [105,254]. Conversely, when supplemental Calcium was added to an NH_4^+-N source in greenhouse and field studies, NH_4^+ uptake was increased, resulting in greater top growth, root yields in radish, and NUE [255].

A greater response to phosphorus applications is observed when supplemental levels of N are provided, in N deficient soils [159,256,257]. Nitrogen may help to increase plant P levels by promoting root growth, improving uptake and translocation, and by increasing the soil pH to solubilize soil or P fertilizer [256]. For the production of celery transplants, an interaction was observed between N and P levels. Increased P levels of up to 125 ppm in the soil solution increased seedling growth only when N levels were above 250 ppm [258]. On a global scale, and as observed in Africa [257], an analysis of agricultural production on 113 countries found a close correlation between P use efficiency (PUE) and NUE, indicating that both nutrients, as well as the adoption of best nutrient management practices, are integral to maintain agricultural productivity [25].

Additional nutrient x N or NUE positive interactions have been observed for Selenium [259,260] and Sulfur [261]. Other nutrients such as molybdenum and copper are involved in N metabolism [257], and their uptake may be affected by the soil pH, as affected by the prevailing N form in the soil [256].

5.3. Environmental Stress x N Interactions

Environmental stress has a strong impact on plant growth, photosynthetic assimilating area, shoot-root interactions, nutrient uptake, yields and quality, and thus it has a strong influence on NUE. Chlorophyll fluorescence has been used as a technique to evaluate the effect of environmental stress on the photosynthetic process. Because N is intricately involved in the photosynthetic process, chlorophyll fluorescence may serve as an early indicator of N x environmental stress interactions, including of biotic and abiotic stress such from pests, nutrients, or water deficits [10,262,263]. For example, chlorophyll fluorescence may be used to identify germplasm with greater photosynthetic efficiency under temperature stress, which affects NUE, as heat stress disrupts nutrient uptake, leaf area duration, and both N metabolism and fixation [264].

While high ammonium levels are toxic to many plants, evidence indicates that at lower levels it may allow plants to overcome periods of biotic or abiotic stress, including improved tolerance to salinity, with production of bioactive compounds such as glucosinolates, and improved nutrient uptake via rhizosphere acidification [265]. While crop salinity adversely affects N uptake and metabolism, research indicates that modulation of N in the plant may help to overcome some of the effects of salt stress by either modifying N fertilizer practices, such as the form of N, or by selecting plants with an improved N metabolism. Some of the possible physiological mechanisms of N nutrition to ameliorate the impacts of salt stress include the up-regulation of salt-stress tolerant genes, activation of the antioxidant system and synergistic hormonal interactions [266].

The potential effect of N nutrition to overcome periods of abiotic stress include tolerance to drought stress through an improved NUE, WUE, and osmoregulation [267,268]. Drought tolerant tomato cultivars also have shown a greater N uptake than sensitive ones, allowing the plants to better maintain some physiological functions [126]. Nitric oxide has been found to be a mediator in the stress response to both drought and salinity stress, as a signaling molecule that enhances the activity of antioxidant defense mechanisms, moderating the plant water status and photosynthetic activity

 doi:10.20944/preprints202312.0768.v1

[269]. Mycorrhizal associations may also help to improve the plant's nutrient status and N uptake under stressful growing conditions, such as during exposure to ozone, or nutrient deficiency [270]. Nitrogen may also play a role in increasing the tolerance to high temperatures, as observed in cucumber. By playing a role in the regulation of the specific heat capacity and leaf water status, a higher N supply resulted in lower leaf temperatures through increased transpiration rates [271].

5.4. Pests x N Interactions

A balanced crop nutrition is considered a centerpiece of a pest management program [272]. Crop quality and yield losses caused by pests and weeds have a direct impact on NUE, resulting in direct economic losses for farmers. Nutrient management practices may be adjusted in terms of N form, rate and timing of application, to minimize the incidence and degree of losses from pest infestations or weed competition. By playing an integral role in the production of vegetative growth, in the plant's metabolism, and as a part of its defense system, the N plant status affects the plant's susceptibility or tolerance to pest infestations. Surplus N levels may trigger pest outbreaks, as pests and pathogens rely on N resources for their sustenance (e.g. aminoacids), while N deficits may also make the plant more susceptible to infection by diseases or to competition from weeds [160,273] (Table 11). The common practice of over fertilizing and over watering also often results in succulent plant growth and in pest outbreaks [9]. As part of the nutrient management practices that are followed to minimize the incidence of pest outbreaks, other closely interlinked production factors need to be considered as part of the overall management strategy, such as germplasm selection, soil moisture, tillage, vegetational diversification, and landscape or area-wide design.

In general, high N rates are related with increased susceptibility to biotrophic pathogens that require live living host cells for their sustenance, and to a reduced susceptibility to necrotrophs, which feed on dead or dying cells [274]. While in general a greater N:C ratio is more likely to result in pest outbreaks, N may also play a role in the plant's disease defense mechanism as it is a component of varied plant defense compounds, such as enzymes and proteins that are involved in the plant immune signaling cascade response mechanisms and resistance metabolic pathways [273–276].

Table 11. Effect of Nitrogen nutrition on selected plant pathogens.

Pathogen	Notes	References
Bacterial rots, *Pantonea* spp.; Bacterial blights, *Pseudomonas*	Low foliar N results in higher disease incidence, onion; celery	[152,277]
Damping-off, *Phytophthora, Pythium, Rhizoctonia* spp.	High N rates and planting density increase disease severity, eggplant, tomato; N stimulates plant defense compounds, potato	[275,278,279]
Early blight, *Alternaria solan*i	N deficient tomato and potato plants are more susceptible; N stimulates plant defense compounds, potato, tomato	[272,275,280]
Foliar diseases, *Mycosphaerella, Diplocarpon* spp.	Increased incidence with high N during spring applications in strawberry	[281]
Fruit rots, *Botrytis*	High N spring applications increase disease incidence, strawberry	[282]
Fusarium wilt, *Fusarium* oxysporum	High N favors the disease, tomato; N stimulates plant defense compounds, tomato	[275,278]
Leaf spots, *Botrytis fabae* and rust (*Uromyces viciae-fabae*)	High N results in greater disease incidence on faba bean, *Vicia faba* L.	[283]
Soft rots, *Erwinia*, *Pseudomonas*, *Clostridium*	Excessive N may exacerbate the disease on vegetables, affecting fruits, tubers, roots, and foliage; N stimulates plant defense compounds, tomato	[275,278,284]
Storage diseases, *Aspergillus*; Bacterial soft rot, *Pseudomonas, Erwinia, Botrytis*	Increased incidence with excess N rates, onion	[81,285]
Physiological disorders	Increased incidence of sugar ends in potato with high N rates;	[231]

Preprints (www.preprints.org) | NOT PEER-REVIEWED | Posted: 12 December 2023 doi:10.20944/preprints202312.0768.v1

Nitrogen fertilizer practices, especially under monoculture systems, also have an effect on arthropod population dynamics [2]. Higher population levels in response to high N fertilizer applications have been reported for important vegetable pests such as aphids, with differential effects on abundance, growth rates, timing of N applications, and abundance of natural enemy populations [150,286,287]; caterpillars [279,288]; higher numbers with high N rates, and lower tissue N levels found in leafhopper resistant okra varieties [279,289]; mites [279,290,291]; thrips, showing a greater feeding preference at higher N levels [70,152]; and increased numbers and fitness on whiteflies [292,293]. Greater pest numbers may also be observed in crops receiving chemical N sources, as compared to organic sources [294].

A number of management practices and variables affect the interaction between weed pressure, crop fertility, and NUE. Nitrogen contributes to vigorous top growth which can increase early competitiveness against weeds, reducing the length of the weed-free period required to maintain yield losses below economic injury levels [295]. Research conducted over several years on small farms of Zimbabwe showed that greater weed infestations resulted in a lower NUE, which could be improved with higher planting densities and improved weed management practices [296]. Research with peanuts conducted over two years in Egypt found an interaction between N fertilizer rates, the length of weed-free periods and NUE. The highest N rate resulted in the greatest yields, increased weed pressure, but a lower seed oil content and a 50% lower NUE, compared to the lower N rates. Overall, increased weed pressure resulted in lower yields and a lower NUE. With respect to the weed-free periods (WFP) a lower NUE was observed with a WFP up to 6 weeks after emergence, with increasing NUE values obtained with longer weed-free periods. A higher NUE was also observed when the fields were allowed to remain weedy only up to the first 4 weeks after emergence, indicating that most of the competition for N likely occurred during the later stages of crop growth [297]. A weed-free period x N rate experiment conducted with tomatoes in Central Sulawesi, Indonesia also found a significant weed-free period x N rate interaction. The optimal N application rates to reach commercial yields decreased, with the increased length of the weed free periods (from 0 to 8 weeks), while the greatest weed growth for each WFP was obtained with the higher N rates (of up to 150 Kg N Ha^{-1}), highlighting the importance of timely weed management to minimize the need for high N application rates [298].

Because weeds are often more competitive for N than crops, in well fertilized soils, several management strategies may be considered to improve the crop N competitiveness [299]. Management practices to reduce weed competition for N include proper placement, such as side-dressing applications vs. broadcasting; the timing of applications to better synchronize crop uptake demands; cultivar selection; planting densities, as well as weed-management strategies such as mulching or crop rotations [299]. In organic systems where tillage is often practiced for weed control, it is important to consider the effect of tillage on potential soil N mineralization, which could result in environmental losses during periods of low N uptake demand [120]. Thus, the timing of tillage operations to reduce weed pressure below economic injury levels, and to better synchronize it with periods of higher N crop demand may minimize the potential of crop N losses from leaching or volatilization. Research conducted in Tehran, Iran illustrates the need to recognize the important water x weed x N use interactions, when evaluating aspects of NUE, especially in semi-arid regions under irrigation. A two-year experiment with maize showed increased yield responses to N with a higher irrigation rate; greater N environmental losses with the highest N rate (450 Kg N Ha^{-1}); greater N losses under no weed pressure than in weedy plots; a greater NUE in treatments with a greater irrigation rate; a greater NUE at the lower N rates (150 Kg N Ha^{-1}) compared to the highest N rate; and, a greater level of inputs in weedy plots resulted in proportionally greater yield losses, highlighting the need to use inputs moderately to optimize their use efficiency [300].

5.5. Soil Biota x N Interactions

As a driver of the N cycle, involved in the regulation of soil ecological functions, the soil biota and crop NUE are closely interlinked [8,190,204]. A diverse microbial community, involved in six distinct N transformation processes such as N mineralization, ammonification, and fixation,

 doi:10.20944/preprints202312.0768.v1

contribute to the N cycle and to the overall soil N pool. High chemical N fertilizer inputs, common in commercial operations, tend to decrease the soil microbial activity and diversity [43]. Current research efforts aim to identify nutrient and management practices, such as vegetational diversity, grazing, tillage and application of organic amendments, that reduce the reliance on chemical N inputs, reduce N losses, and that improve the soil microbial activity, which has a positive impact on the system NUE [86,190,301].

Several alternative management practices may be followed to improve the soil fertility and microbial activity. A two-year greenhouse and field experiment conducted in Hokkaido, Japan found a 47.5% increased lettuce NUE with rye (*Secale cereale*)/hairy vetch and 27% with sole hairy vetch cover crop rotations, compared to the controls. In terms of N, hairy vetch appeared to be the most valuable component of the cover crops, by increasing the concentration of soil inorganic N. Both cover crop treatments also had a greater microbial β-glucosidase enzyme (BG) activity, responsible for residues degradation, and a greater soil microbial biomass than the controls, indicating their possible role in improving N mineralization and NUE [216]. A similar stimulation of soil microbial and invertebrate activity from the use of legume-grass mixtures and non-legume cover crops on organic systems were reported from a six-year study with cool season vegetables in Salinas, California [302]. Organic mulches also help to improve microbial activity, with reported improvements of 42% in microbial biomass, compared to non-mulch treatments, resulting in improved soil quality and yields. A review of the literature indicated that in cereal crops mulching increased NUE by 4-17% compared to non-mulched treatments, attributed primarily to an increased rhizosphere microbial activity [43]. Intercropping systems, given the differential root distribution and exudate pattern of the component intercrops, also have a positive impact on the diversity and activity of microbial activity, contributing to improved internal nutrient cycles and NUE [204].

Fungal mycorrhizae form symbiotic associations with crops, providing several benefits including overcoming periods of stress and improving nutrient uptake. Mycorrhizae may form synergistic associations with N fixing rhizobia bacteria, resulting in increased N fixation rates [303]; N may be transported via mycorrhizal networks [304]; N uptake and metabolism may improve during periods of stress [270,305]; resulting in improved crop growth and improved N uptake, as observed under varied levels of stress with legumes, eggplant, and tomato [262,305,306].

Microbial inoculants, such as from *Bacillus*, *Pseudomonas*, *Streptomyces*, *Trichoderma* and mycorrhizal species, are increasingly being used under commercial production, as biofertilizers or for plant growth promotion which may have a positive effect on N uptake [307]. Plant growth promoting bacterial inoculants that are applied in some commercial operations include from *Pseudomonads*, *Azoarcus*, *Beijerinckia*, and *Cyanobacteria* species. While research indicates that microbial inoculants, applied in consortia, may have growth-promoting effects, and may improve water use and nutrient uptake [308,309], further research is required to validate their effects under a varied set of production systems and environmental conditions [310].

Earthworms, among the most important soil invertebrates, are considered to be drivers of the global food system as soil ecological engineers, contributing to residue decomposition, soil structure, water conservation, soil microbial activity, and organic matter and nutrient cycling [311–313]. As decomposers of crop residues earthworms help to release N for crop uptake, among other benefits, resulting in an estimated 23% increased top growth and in 25% increased global crop yields, under conditions of no high external N inputs, highlighting their potential contribution to N uptake and NUE under low-input, subsistence, or organic systems [314]. A ^{15}N residue-labeled study evaluated the intricate interactions between earthworms, microbial activity and mycorrhizae with respect to crop N uptake. Earthworms enhanced the residue decomposition by regulating soil microbial activity, and with the combined presence of *Glomus* mycorrhizae resulted in a synergistic effect, improving crop N uptake, from the residue derived N mineralization [315].

6. Nitrogen Losses Management

Given the wider recognition about the environmental risks posed by reactive N forms in the environment, N fertilizer recommendations aim to maintain crop quality and productivity, while

 doi:10.20944/preprints202312.0768.v1

minimizing losses [36]. In commercial operations, it is estimated that farm losses represent up to 70% of the applied N fertilizer, and that these losses may be reduced by 15-30% by adopting improved management practices [41]. Studies with potato have reported total N losses of 35%, N leaching losses of 20-30%, up to 60% under excess irrigation, and <0.5% losses from volatilization [71,141]. In field-grown tomato, N losses reached almost 90% under high ^{15}N application rates, while under hydroponic tomatoes N volatilization can reach up to 15% of the applied N [19,316]. Under chemical-free or organic farming conditions asynchrony between the timing of N mineralization and crop uptake may result in N losses, such as in cases of early season precipitation, as observed in sweet corn [8,188].

Mitigation strategies have been proposed to reduce N emissions, both from the manufacture of chemical fertilizers and by following best management practices that involve the proper N source, application rates, timing, and placement [4,317]. Solutions to address problems of N pollution are not only agronomic, but have a strong social component, based on the particular socioeconomic conditions of the production system. Thus, policy, economic, market, and structural barriers need to be identified to design educational and implementation programs that meet the needs of particular production regions [318,319].

Examples of management strategies to minimize losses by improving the N cycle on the farm [8], include to develop a baseline crop N budget analysis [49,141]; proper irrigation rates, placement and scheduling [232,250] ; increased vegetational diversity [320]; conservation practices such as the planting of vegetative buffers [321]; slow-release fertilizers [73]; N fertigation, especially in arid regions [322]; and cover crops in the rotation showing an average of 63% reduction in nitrate leaching [323].

7. Conclusions

Nitrogen is the most limiting nutrient for the production of vegetable crops. Historically, farmers have applied high N rates, to ensure adequate yields, resulting in significant economic and environmental losses, and on possible adverse impacts on the long-term fertility of the soil.

Because the N cycle is affected by so many interacting system variables, an understanding and predictions about its movement is highly location, season, and cropping system specific. Thus, only broad generalizations can be made about its movement in the environment, based on general biophysical mechanisms and known microbial and chemical transformations. Because vegetable crops are considered specialty or minor crops, and because they represent hundreds of species, individual species have historically received considerably less research attention compared to the major cereal staple crops, with respect to N fertilization practices. However, because vegetables are relatively high-value crops often grown under capital and input-intensive systems, such as under protected agriculture, it is important to evaluate mitigation strategies to minimize potential impacts from the extensive use of N fertilizers.

Steps to improve N use recommendations include to develop a budget to estimate total N outputs minus inputs, based on realistic yield estimates and to develop diagnostic tools such as N tissue analysis, based on N rate growth-response curves, to fine-tune N recommendations. Improved chemical N fertilizer practices include adjustments on the total application rates, the timing, and placement of application. Alternative practices that can be explored to improve the internal N cycle include the use of cover crops, rotations, alternative tillage, integrated crop-livestock systems, vegetational diversification, a greater reliance on organic amendments, as well as the use of crop cultivars with improved NUE traits under the different types of cropping systems and growing environments [8].

Because vegetable crops are grown under such a wide-range of environmental settings, ranging from small-scale production as cash-crops under subsistence agriculture, to organic systems, large-scale intensive monocultures, to capital- and input-intensive protected agriculture, it is imperative to consider the particular socioeconomic conditions of the production system, to better formulate potential recommended practices that may help to improve system NUE. When considering the human-factor or local socioeconomic conditions, the level of complexity rises considerably when

conducting research, outreach or educational activities to improve the management of the N cycle on the farm, and agricultural productivity.

Funding: This research received no external funding.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Tan, Y.; Xu, C.; Liu, D.; Wu, W.; Lal, R.; Meng, F. Effects of optimized N fertilization on greenhouse gas emission and crop production in the North China Plain. Field Crops Res. 2017, 205, 135-146. http://dx.doi.org/10.1016/j.fcr.2017.01.003
2. Martínez-Dalmau, J.; Berbel, J.; Ordóñez-Fernández, R. Nitrogen fertilization: A review of the risks associated with the inefficiency of its use and policy responses. Sustainability 2021, 13, 5625. https://doi.org/10.3390/ su13105625
3. Schulte-Uebbing, L.F.; Beusen, A.H.W.; Bouwman, A.F.; De Vries, W. From planetary to regional boundaries for agricultural nitrogen pollution. Nature 2022, 610, 7932, 507-512. https://doi.org/10.1038/s41586-022-05158-2
4. Gao, Y.; Cabrera Serrenho, A. Greenhouse gas emissions from nitrogen fertilizers could be reduced by up to one-fifth of current levels by 2050 with combined interventions. Nat. Food, 2023, 4, 170-178, https://doi.org/10.1038/s43016-023-00698-w
5. Schomberg, A.C.; Bringezu, S.; Beusen, A.W. Water quality footprint of agricultural emissions of nitrogen, phosphorus and glyphosate associated with German bioeconomy. Commun. Earth Environ. 2023, 4, 404. https://doi.org/10.1038/s43247-023-01054-3
6. Morris, T.F.; Murrell, T.S.; Beegle, D.B.; Camberato, J.J.; Ferguson, R.B.; Grove, J.; Ketterings, Q.; Kyveryga, P.M.; Laboski, C.A.; McGrath, J.M.; Meisinger, J.J. Strengths and limitations of nitrogen rate recommendations for corn and opportunities for improvement. Agron. J. 2018, 110, 1-37. doi:10.2134/agronj2017.02.0112
7. Li, S.L.; Liu, X.; Yue, F.J.; Yan, Z.; Wang, T.; Li, S.; Liu, C.Q. Nitrogen dynamics in the critical zones of China. Progress in Physical Geography: Earth and Environment, 2022, 46, 869-888. DOI: 10.1177/03091333221114732
8. Valenzuela, H. Ecological management of the nitrogen cycle in organic farms. Nitrogen 2023, 4, 58-84. https://doi.org/10.3390/ nitrogen4010006
9. Devi, H.M.; Pongener, N.; Devi, H.S.; Hemanta, L. Effect of plant nutrition on development of plant diseases. In: A. Mahapatra, et al. (Eds.) *Advances in Agriculture and Allied technologies*. AkiNik Pub. New Delhi. 2022, pp.155-163. https://doi.org/10.22271/ed.book.1919
10. Kalaji, H.M.; Jajoo, A.; Oukarroum, A.; Brestic, M.; Zivcak, M.; Samborska, I.A.; Cetner, M.D.; Łukasik, I.; Goltsev, V.; Ladle, R.J. Chlorophyll a fluorescence as a tool to monitor physiological status of plants under abiotic stress conditions. Acta Physiol. Plant. 2016, 38, 1-11. DOI 10.1007/s11738-016-2113-y
11. Patra, S.; Mandal, A. Significance of Plant E3 ubiquitin ligases in NPK homeostasis: A review. Plant Stress 2023, 100207. https://doi.org/10.1016/j.stress.2023.100207
12. Tiemens-Hulscher, M.; Lammerts van Bueren, E.T.; Struik, P.C. Identifying nitrogen-efficient potato cultivars for organic farming. Euphytica 2014, 199, 137-154. DOI 10.1007/s10681-014-1143-z
13. Samari, E.; Sharifi, M.; Ghanati, F.; Fuss, E.; Ahmadian Chashmi, N. Chitosan-induced phenolics production is mediated by nitrogenous regulatory molecules: NO and PAs in Linum album hairy roots. Plant Cell, Tissue and Organ Culture (PCTOC), 2020, 140, 563-576. https://doi.org/10.1007/s11240-019-01753-w
14. Gauthier, K.; Pankovic, D.; Nikolic, M.; Hobert, M.; Germeier, C.U.; Ordon, F.; Perovic, D.; Niehl, A. Nutrients and soil structure influence furovirus infection of wheat. Front. Plant Sci. 2023, 14. DOI 10.3389/fpls.2023.1200674
15. Hancock, J.T. Nitric oxide signaling in plants. Plants 2020, 9, 1550. doi:10.3390/plants9111550
16. Römheld, V.; Jiménez-Becker, S.; Neumann, G.; Gweyi-Onyango, J.P.; Puelschen, L.; Spreer, W.; Bangerth, F. Non-nutritional fertigation effects as a challenge for improved production and quality in horticulture. In: P. Imas and M.R. Price (Eds.) *Fertigation Proceedings: Optimizing the Utilization of Water and Nutrients*, Beijing, 20-24 September 2005, International Potash Institute, Horgen, Switzerland, 2008, pp.103-115. DOI 10.3235/978-3-9523243-8-7
17. Gao, Y.; Qi S.; Wang, Y. Nitrate signaling and use efficiency in crops. Plant Comm. 2022, 3, 100353, https://doi.org/10.1016/j.xplc.2022.100353
18. Liang, Q.; Dong, M.; Gu, M.; Zhang, P.; Ma, Q.; He, B. MeNPF4. 5 improves cassava nitrogen use efficiency and yield by regulating nitrogen uptake and allocation. Front. Plant Sci. 2022, 13, 866855. doi: 10.3389/fpls.2022.866855

19. Jalpa, L.; Mylavarapu, R.S.; Hochmuth, G.; Wright, A.; van Santen, E. Recovery efficiency of applied and residual nitrogen fertilizer in tomatoes grown on sandy soils using the ^{15}N technique. Sci. Hortic. 2021, 278, 109861. https://doi.org/10.1016/j.scienta.2020.109861
20. Engindeniz, S.; Gül, A. Economic analysis of soilless and soil-based greenhouse cucumber production in Turkey. Sci. Agric. 2009, 66, 606-614.
21. Duhan, P.K. Cost benefit analysis of tomato production in protected and open farm. Intern. J. Adv. Res. Management and Social Sciences (IJARMSS) 2016, 5, 140-148.
22. Univ. Massachusetts Extension. Crop production budgets. Center for Agriculture, Food, and the Environment. Accessed Oct. 12, 2023 from, https://ag.umass.edu/vegetable/fact-sheets/crop-production-budgets
23. Gianquinto, G.; Muñoz, P.; Pardossi, A.; Ramazzotti.; Savvas, D. Soil fertility and plant nutrition. In: W. Baudoin et al. (Eds.) *Good agricultural practices for greenhouse vegetable crops: Principles for mediterranean climate areas.* Food and Agriculture Organization (FAO). Plant Production and Protection paper 217. Rome, Italy. 2013. pp. 205-270.
24. Tei, F.; De Neve, S.; de Haan, J.; Kristensen, H.L. Nitrogen management of vegetable crops. Agric. Water Manag. 2020, 240, 106316. https://doi.org/10.1016/j.agwat.2020.106316
25. Zou T.; Zhang X.; Davidson E.A. Global trends of cropland phosphorus use and sustainability challenges. Nature 2022, 611, 81-87. https://doi.org/10.1038/s41586-022-05220-z
26. De Pascale, S.; Rouphael, Y.; Gallardo, M.; Thompson, R.B. Water and fertilization management of vegetables: State of art and future challenges. Eur. J. Hortic. Sci, 2018, 83, 306-318. https://doi.org/10.17660/eJHS.2018/83.5.4
27. You, L.; Ros, G.H.; Chen, Y.; Shao, Q.; Young, M.D.; Zhang, F.; de Vries, W. Global mean nitrogen recovery efficiency in croplands can be enhanced by optimal nutrient, crop and soil management practices. Nat. Commun. 2023, 14, 5747. https://doi.org/10.1038/s41467-023-41504-2
28. Feng, R.; Li, Z. Current investigations on global N_2O emissions and reductions: Prospect and outlook. Environ. Pollut. 2023, 122664. https://doi.org/10.1016/j.envpol.2023.122664
29. Drinkwater, L.E.; Snapp, S.S. Advancing the science and practice of ecological nutrient management for smallholder farmers. Front. Sustain. Food Syst. 2022, 6, 921216. DOI 10.3389/fsufs.2022.921216
30. Cassman, K.G.; Dobermann A.; Walters, D.T. Agroecosystems, nitrogen-use efficiency, and nitrogen management. Ambio 2002, 31, 132-140. doi:10.1579/0044-7447-31.2.132
31. Fageria, N.; Baligar, V. Enhancing nitrogen use efficiency in crop plants. Adv. Agron. 2005, 88, 97-185. https://doi.org/10.1016/S0065-2113(05)88004-6
32. Hirel, B.; Tétu, T.; Lea, P.J.; Dubois, F. Improving nitrogen use efficiency in crops for sustainable agriculture. Sustainability 2011, 3, 1452-1485. doi:10.3390/su3091452
33. Scholberg, J. M. S.; Zotarelli, L.; Dukes, M.D.; Ozores-Hampton, M.; Liu, G.D. Enhancing fertilizer efficiency in high input cropping systems in Florida. Sustain. Agric. Rev. 2013, 12, 143-174. https://doi.org/10.1007/978-94-007-5961-9_5
34. Han, M.; Okamoto, M.; Beatty, P.H.; Rothstein, S.J.; Good, A.G. The genetics of nitrogen use efficiency in crop plants. Annu. Rev. Genet. 2015, 49, 269-289. doi:10.1146/annurev-genet-112414-055037
35. Liu, Q.; Wu, K.; Song, W.; Zhong, N.; Wu, Y.; Fu, X. Improving crop nitrogen use efficiency toward sustainable green revolution. Annu. Rev. Plant Biol. 2022b, 73, 523-551. https://doi.org/10.1146/annurev-arplant-070121-015752
36. Govindasamy, P.; Muthusamy, S.K.; Bagavathiannan, M.; Mowrer, J.; Jagannadham, P.T.K.; Maity, A.; Halli, H.M.; GK, S.; Vadivel, R.; TK, D.; Raj, R. Nitrogen use efficiency- a key to enhance crop productivity under a changing climate. Front. Plant Sci. 2023, 14, 1121073. DOI 10.3389/fpls.2023.1121073
37. He, G.; Liu, X.; Cui, Z. Achieving global food security by focusing on nitrogen efficiency potentials and local production. Glob. Food Sec. 2021, 29, 100536. https://doi.org/10.1016/j.gfs.2021.100536
38. Tian, X.; Zhuang, M.; Yin, Y.; Cong, J.; Ying, H.; Wang, Y.; Cui, Z. Improved mapping of nitrogen loss and surplus in China's maize belt. Agron. J. 2022, 114, 2811-2821. DOI: 10.1002/agj2.21172
39. Zhang, X. A plan for efficient use of nitrogen fertilizers. Nature 2017, 543, 322-323. https://doi.org/10.1038/543322a
40. Ciampitti, I.A.; Vyn, T.J. Physiological perspectives of changes over time in maize yield dependency on nitrogen uptake and associated nitrogen efficiencies: A review. Field Crops Res. 2012, 133, 48-67, doi:10.1016/j.fcr.2012.03.008
41. Anas, M.; Liao, F.; Verma, K.K.; Sarwar, M.A.; Mahmood, A.; Chen, Z.L.; Li, Q.; Zeng, X.P.; Liu, Y.; Li, Y.R. Fate of nitrogen in agriculture and environment: Agronomic, eco-physiological and molecular approaches to improve nitrogen use efficiency. Biol. Res. 2020, 53 (1), 1-20. https://doi.org/10.1186/s40659-020-00312-4
42. Conant, R.T.; Berdanier, A.B.; Grace, P.R. Patterns and trends in nitrogen use and nitrogen recovery efficiency in world agriculture. Global Biogeochem. Cycles 2013, 27, 558-566. doi:10.1002/gbc.20053, 2013

43. Wang, X.; Fan, J.; Xing, Y.; Xu, G.; Wang, H.; Deng, J.; Wang, Y.; Zhang, F.; Li, P.; Li, Z. The effects of mulch and nitrogen fertilizer on the soil environment of crop plants. Adv. Agron. 2019, 153, 121-173. https://doi.org/10.1016/bs.agron.2018.08.003
44. Halvorson, A.D.; Follett, R.F.; Bartolo, M.E.; Schweissing, F.C. Nitrogen fertilizer use efficiency of furrow-irrigated onion and corn. Agron. J. 2002, 94, 442-449. https://doi.org/10.2134/agronj2002.4420
45. de Jesus, H.I.; da Silva, A.L.B.R.; Cassity-Duffey, K.; Coolong, T. Estimating fertilizer nitrogen-use efficiency in transplanted short-day onion. Nitrogen 2023a, 4, 286–295. https://doi.org/10.3390/nitrogen4030021
46. Lammerts van Bueren, E.T.; Struik, P.C. Diverse concepts of breeding for nitrogen use efficiency: A review. Agron. Sustain. Dev. 2017, 37, 1-24. DOI 10.1007/s13593-017-0457-3
47. Haegele, J.W.; Cook, K.A.; Nichols, D.M.; Below, F.E. Changes in nitrogen use traits associated with genetic improvement for grain yield of maize hybrids released in different decades. Crop Sci. 2013, 53,1256-1268. doi: 10.2135/cropsci2012.07.0429
48. Fageria, N.K.; Nitrogen harvest index and its association with crop yields, J. Plant Nutr. 2014, 37, 795-810. DOI: 10.1080/01904167.2014.881855
49. Prasad, R.; Hochmuth, G.J. How to calculate a partial nitrogen mass budget for potato. Univ. Florida Coop. Ext. Serv, SL401, Gainesville, Florida, 2020, 6 pp.
50. Lammel, J. Fertilizer best management practices in the context of product stewardship. In: *Fertilizer Best Management Practices,* Papers presented at the IFA International Workshop on Fertilizer Best Management Practices, 7-9 March 2007, Brussels, Belgium. International Fertilizer Industry Association (IFA), Paris, 2007. pp. 71-76.
51. Wortmann, C.S.; Tarkalson, D.D.; Shapiro, C.A.; Dobermann, A.R.; Ferguson, R.B.; Hergert, G.W.; Walters, D. Nitrogen use efficiency of irrigated corn for three cropping systems in Nebraska. Agron. J. 2011, 103, 76-84. doi:10.2134/agronj2010.0189
52. Dourado-Neto, D.; Powlson, D.; Bakar, R.A.; Bacchi, O.O.S.; Basanta, M.D.V.; Cong, P.T.; Keerthisinghe, G.; Ismaili, M.; Rahman, S.M.; Reichardt, K.; Safwat, M.S.A. Multiseason recoveries of organic and inorganic nitrogen-15 in tropical cropping systems. Soil Sci. Soc. of America J. 2010, 74, 139-152. doi:10.2136/sssaj2009.0192
53. Griesheim, K.L.; Mulvaney, R.L.; Smith, T.J.; Nunes, V.L.; Hertzberger, A.J. Isotopic comparison of ammonium and nitrate sources applied in-season to corn. Soil Sci. Soc. Amer. J. (SSSAJ) 2023, 87, 555-571. DOI: 10.1002/saj2.20531
54. Vieira-Megda, M.X.; Mariano, E.; Leite, J.M.; Franco, H.C.J.; Vitti, A.C.; Megda, M.M.; Khan, S.A.; Mulvaney, R.L.; Trivelin, P.C.O. Contribution of fertilizer nitrogen to the total nitrogen extracted by sugarcane under Brazilian field conditions. Nutr. Cycling Agroecosyst. 2015, 101, 241-257. DOI 10.1007/s10705-015-9676-7
55. Anon. Nitrogen fertilization of lettuce. Vegetable growers update newsletter. Colorado State University Coop. Ext. Serv. Spring, 1985, 1, 2, Fort Collins, Colorado.
56. Zhang, X.; Davidson, E.A.; Mauzerall, D.L.; Searchinger, T.D.; Dumas, P.; Shen, Y. Managing nitrogen for sustainable development. Nature 2015b, 528, 51-59. doi:10.1038/nature15743
57. Barker, A.V. Genotypic responses of vegetable crops to nitrogen nutrition. HortScience 1989, 24, 584-591. DOI: https://doi.org/10.21273/HORTSCI.24.4.584
58. Hochmuth, G.; Hanlon, E.A. Summary of N, P, and K research with tomato in Florida. Univ Florida Coop. Ext. Serv. SL 355. Gainesville, FL. 2020a
59. Jackson, L.E.; Bloom, A.J. Root distribution in relation to soil nitrogen availability in field-grown tomatoes. Plant Soil 1990, 128, 115-126. https://doi.org/10.1007/BF00011100
60. Fox, R.; Valenzuela, H.R. Vegetables grown under tropical and subtropical conditions. In: W. Wichmann (Ed.) *IFA World Fertilizer Use Manual*. International Fertilizer Industry Assoc. Germany. 1992. pp. 293-338.
61. de Haan, J.J.; F.G. Wijnands. In: De Haan, J.J. (Ed.) *Integrated and ecological nutrient management.* VEGINECO Project Report No. 3. Applied Plant Research. Wageningen, Netherlands. 2002, p. 8-23.
62. Zink, F. Studies on the growth rate and nutrient absorption of onion. Hilgardia 1966, 37, 203-218. DOI:10.3733/hilg.v37n08p203.
63. Crespo-Ruiz, M.; Goyal, M.; Baez, C.C.D.; Rivera, L.E. Nutrient uptake and growth characteristics of nitrogen fertigated sweet peppers under drip irrigation and plastic mulch. J. Agric. Univ. Puerto Rico, 1988, 4, 575-584. https://doi.org/10.46429/jaupr.v72i4.8006
64. Sullivan, D.M.; Peachey, R.E.; Heinrich, A.L.; Brewer, L.J. Nutrient management for sustainable vegetable cropping systems in Western Oregon. Oregon State Univ.Coop. Ext. Serv. EM 9165. Corvallis, Oregon, USA. 2017, 25 pp
65. Bowen, P.; Frey, B. Response of plasticultured bell pepper to staking, irrigation frequency, and fertigated nitrogen rate. HortScience 2002, 37, 95-100. https://doi.org/10.21273/HORTSCI.37.1.95

66. Dobermann, A. Nutrient use efficiency–measurement and management. In: *Fertilizer Best Management Practices*, Papers presented at the IFA International Workshop on Fertilizer Best Management Practices, 7-9 March 2007, Brussels, Belgium, International Fertilizer Industry Association (IFA), Paris, 2007, pp. 1-28.
67. Zandvakili, O.R.; Barker, A.V.; Hashemi, M.; Autio, W.R.; Etemadi, F.; Sadeghpour, A. Influence of nitrogen source and rate on lettuce yield and quality. Agron. J. 2022, 114, 1401-1414. DOI: 10.1002/agj2.20966
68. Zotarelli, L.; Wade, T.; England, G.; Christensen, C. Nitrogen fertilization guidelines for potato production in Florida. Univ. Florida Coop. Ext. Serv, EDIS, HS1429. Gainesville, Florida. 2021b
69. Wang, M.; Wang, L.; Cui, Z.; Chen, X.; Xie, J.; Hou, Y. Closing the yield gap and achieving high N use efficiency and low apparent N losses. Field Crops Res. 2017, 209, 39-46. http://dx.doi.org/10.1016/j.fcr.2017.03.016
70. Buckland, K.R. Evaluating fertilizer rate, crop rotation and trap crops for effects on onion growth and yield, soil health, thrips densities and iris yellow spot virus incidence. MS Thesis, Utah State Univ. Paper 980, 2011, 133 pp. http://digitalcommons.usu.edu/etd/980
71. Prasad, R.; Hochmuth, G.J. Environmental nitrogen losses from commercial crop production systems in the Suwannee River Basin of Florida. PLoS One 2016, 11, e0167558. doi:10.1371/journal.pone.0167558
72. Grasso, R.; Peña-Fleitas, M.T.; de Souza, R.; Rodríguez, A.; Thompson, R.B.; Gallardo, M.; Padilla, F.M. Nitrogen effect on fruit quality and yield of muskmelon and sweet pepper cultivars. Agronomy 2022, 12, 2230. https://doi.org/ 10.3390/agronomy12092230
73. Cui, Z.; Dou, Z.; Chen, X.; Ju, X.; Zhang, F. Managing agricultural nutrients for food security in China: past, present, and future. Agron. J., 2014, 106, 191-198. doi:10.2134/agronj2013.0381
74. Casals,J.; Martí,M.; Rull, A.; Pons, C. Sustainable transfer of tomato landraces to modern cropping systems: The effects of environmental conditions and management practices on long-shelf-life tomatoes. Agronomy 2021, 11, 533. https://doi.org/10.3390/agronomy11030533
75. Knuteson, D.L.; Stevenson, W.R.; Wyman, J.A.; Bussan, A.J.; Colquhoun, J.B.; Laboski, C.A.M.; Silva, EM. BioIPM Pepper Workbook. A3844,Univ. Wisc. Coop. Ext. Serv. Madison, 2010. 76 pp.
76. Lan, G.; Jiao, C.; Wang, G.; Sun, Y.; Sun, Y. Effects of dopamine on growth, carbon metabolism, and nitrogen metabolism in cucumber under nitrate stress. Sci. Hortic. 2020, 260, 108790. https://doi.org/10.1016/j.scienta.2019.108790
77. Cao, Y.; Yin, B. Effects of integrated high-efficiency practice versus conventional practice on rice yield and N fate. Agric. Ecosyst. Environ. 2015, 202, 1-7. http://dx.doi.org/10.1016/j.agee.2015.01.001
78. Subedi, K.D.; Ma, B.L. Dry matter and nitrogen partitioning patterns in Bt and non-Bt near-isoline maize hybrids. Crop Sci. 2007, 47, 1186-1192. doi: 10.2135/cropsci2006.09.0593
79. Zotarelli, L.; Barrett, C.E.; da Silva, A.L.B.R.; Christensen, C.T.; England, G.K. Nitrogen fertilization guidelines for bare-ground and plastic mulch cabbage production in Florida. Univ. FL Coop. Ext. Serv. HS1428, Gainesville, Florida. 2021a. https://doi.org/10.32473/edis-HS1428-2021
80. Voogt, W. Fertigation in Greenhouse Production. In: P. Imas and M.R. Price (Eds.) *Fertigation: Optimizing the utilization of water and nutrients*, Fertigation Proceedings: Selected Papers of the IPI-NATESC-CAU-CAAS International Symposium on Fertigation Beijing/China, 20-24 September 2005. International Potash Institute, Horgen, Switzerland, 2008. pp. 116-129. DOI 10.3235/978-3-9523243-8-7
81. Geisseler, D.; Ortiz, R.S.; Diaz, J. Nitrogen nutrition and fertilization of onions (*Allium cepa* L.) - A literature review. Sci. Hortic. 2022, 291, 110591. https://doi.org/10.1016/j.scienta.2021.110591
82. Hochmuth, G.; Hanlon, E.A. Summary of N, P, and K research with pepper in Florida. Univ. Florida Coop. Ext. Serv. SL334. Gainesville, FL. 2020b
83. Jones, C.R.; Michaels, T.E.; Schmitz Carley, C.; Rosen, C.J.; Shannon, L.M. Nitrogen uptake and utilization in advanced fresh-market red potato breeding lines. Crop Sci. 2021, 61, 878-895, DOI: 10.1002/csc2.20297
84. Jalpa, L.; Mylavarapu, R.S.; Hochmuth, G.J.; Wright, A.L.; van Santen, E. Apparent recovery and efficiency of nitrogen fertilization in tomato grown on sandy soils. HortTechnology 2020, 30, 204-211. https://doi.org/10.21273/HORTTECH04480-19
85. Daba,N.A.; Li,D.; Huang, J.; Han, T.; Zhang, L.; Ali, S.; Khan, M.N.; Du, J.; Liu, S.; Legesse, T.G.; et al. Long-term fertilization and lime-induced soil pH changes affect nitrogen use efficiency and grain yields in acidic soil under wheat-maize rotation. Agronomy 2021, 11, 2069. https://doi.org/10.3390/ agronomy11102069
86. Grandy, A.S.; Daly, A.B.; Bowles, T.M.; Gaudin, A.C.; Jilling, A.; Leptin, A.; McDaniel, M.D.; Wade, J.; Waterhouse, H. The nitrogen gap in soil health concepts and fertility measurements. Soil Biol. Biochem. 2022, 175, 108856. https://doi.org/10.1016/j.soilbio.2022.108856
87. Carlson, M.; Hauber, R; Krause, L.; Kretz, R. WCROC Farm sustainability project: Considering online decision support tools. Humphrey School Capstone Report, The Hubert H. Humphrey School of Public Affairs, The University of Minnesota, Minneapolis, 2021, 46 pp.
88. Howeler, R.H. Long-term effect of cassava cultivation on soil productivity. Field Crops Res. 1991, 26, 1-18. https://doi.org/10.1016/0378-4290(91)90053-X
89. Bolin, P.; Brandenberger, L. eds. Cucurbit integrated crop management. Univ. Oklahoma Cooperative Extension Service E-853. Stillwater, OK. 2001. 96 pp.

90. Johansen, T.J.; Thomsen, M.G.; Loes, A.-K.; Riley, H. Root development in potato and carrot crops - influences of soil compaction. Acta Agr. Scand. 2015, 65, 182-192. https://doi.org/10.1080/09064710.2014.977942
91. Breland, T.A.; Hansen, S. Nitrogen mineralization and microbial biomass as affected by soil compaction. Soil Biol. Biochem. 1996, 28 , 655-663. https://doi.org/10.1016/0038-0717(95)00154-9
92. Xu, Y.; Jeanne, T.; Hogue, R.; Shi, Y.; Ziadi, N.; Parent, L.E. Soil bacterial diversity related to soil compaction and aggregates sizes in potato cropping systems. Appl. Soil Ecol. 2021, 168, 104147. https://doi.org/10.1016/j.apsoil.2021.104147
93. Zhu, Y.; Qi, B.; Hao, Y.; Liu, H.; Sun, G.; Chen, R.; Song, S. Appropriate NH4+/NO3- ratio triggers plant growth and nutrient uptake of flowering Chinese cabbage by optimizing the pH value of nutrient solution. Front. Plant Sci. 2021, 12, 656144. doi: 10.3389/fpls.2021.656144
94. Bar-Tal, A.; Aloni, B. Effects of fertigation regime on blossom end rot of vegetable fruits. In: P. Imas and M.R. Price (Eds.) *Fertigation Proceedings: Optimizing the Utilization of Water and Nutrients*, Beijing, 20-24 September 2005, International Potash Institute, Horgen, Switzerland, 2008, pp. 130-145. DOI 10.3235/978-3-9523243-8-7
95. Topcu, Y.; Nambeesan, S.U.; van der Knaap, E. Blossom-end rot: a century-old problem in tomato (*Solanum lycopersicum* L.) and other vegetables. Mol. hortic. 2022, 2, 1. https://doi.org/10.1186/s43897-021-00022-9
96. Goos, R.J.; Schimelfenig, J.A.; Bock, B.R.; Johnson, B.E. Response of spring wheat to nitrogen fertilizers of different nitrification rates. Agron. J. 1999, 91, 287-293. https://doi.org/10.2134/agronj1999.00021962009100020018x
97. Yamagata, M.; Ae, N. Direct acquisition of organic nitrogen by crops. Japan Agric. Res. Quart., JARQ 1999, 33, 15-21.
98. Petersen, S.O.; Schjønning, P.; Olesen, J.E.; Christensen, S.; Christensen, B.T. Sources of nitrogen for winter wheat in organic cropping systems. Soil Sci. Soc. Amer. J. (SSSAJ) 2013, 77, 155-165, doi:10.2136/sssaj2012.0147
99. Alan, R. The effect of nitrogen nutrition on growth, chemical composition and response of cucumbers (*Cucumis sativus* L.) to nitrogen forms in solution culture. J. Hort. Sci. 1989, 64 (4), 467-474. DOI: 10.1080/14620316.1989.11515979
100. Nerson, H. Mineral nutrition of cucurbit crops. Dynamic Soil, Dynamic Plant 2008, 2, 23-32.
101. Zhang, F.C.; Kang, S.Z.; Li, F.S.; Zhang, J.H. Growth and major nutrient concentrations in *Brassica campestris* supplied with different NH4+/NO3− ratios. J. Integr. Plant Biol. 2007, 49, 455-462. doi: 10.1111/j.1672-9072.2006.00373.x
102. Wang, Y.; Zhang, X.; Liu, H.; Sun, G.; Song, S.; Chen, R. High NH4+ /NO3- ratio inhibits the growth and nitrogen uptake of chinese kale at the late growth stage by ammonia toxicity. Horticulturae 2022b, 8, 8. https://doi.org/10.3390/ horticulturae8010008
103. Urlić, B.; Jukić Špika, M.; Becker, C.; Kläring, H.P.; Krumbein, A.; Goreta Ban, S.; Schwarz, D. Effect of NO3 and NH4 concentrations in nutrient solution on yield and nitrate concentration in seasonally grown leaf lettuce. Acta Agric. Scand. B Soil Plant Sci. 2017, 67, 748-757. https://doi.org/10.1080/09064710.2017.1347704
104. Gamiely, S.; Randle, W.M.; Mills, H.A.; Smittle, D.A.; Banna, G.I. Onion plant growth, bulb quality, and water uptake following ammonium and nitrate nutrition. HortScience, 1991, 26, 1061-1063. DOI: https://doi.org/10.21273/HORTSCI.26.8.1061
105. Marti, H.R.; Mills, H.A. Calcium uptake and concentration in bell pepper plants as influenced by nitrogen form and stages of development. J. Plant Nutr. 1991, 14, 1177-1185. DOI: 10.1080/01904169109364276
106. Bar-Tal, A.; Aloni, B.; Karni, L.; Rosenberg, R. Nitrogen nutrition of greenhouse pepper. II. Effects of nitrogen concentration and NO3: NH4 ratio on growth, transpiration, and nutrient uptake. HortScience 2001, 36(7), 1252-1259. https://doi.org/10.21273/HORTSCI.36.7.1252
107. Tabatabaei, S.J.; Yusefi, M.; Hajiloo, J. Effects of shading and NO3: NH4 ratio on the yield, quality and N metabolism in strawberry. Sci. Hortic. 2008, 116, 264-272. doi:10.1016/j.scienta.2007.12.008
108. Osorio, N.W.; Shuai, X.; Miyasaka, S.; Wang, B.; Shirey, R.L.; Wigmore, W.J. Nitrogen level and form affect taro growth and nutrition. HortScience 2003, 38, 36-40. https://doi.org/10.21273/HORTSCI.38.1.36
109. Heeb, A.; Lundegårdh, B.; Ericsson, T.; Savage, G.P. Nitrogen form affects yield and taste of tomatoes. J. Sci. Food Agric. 2005, 85, 1405-1414. DOI: 10.1002/jsfa.2127
110. FAOStat. 2023. Accessed Nov. 9, 2023, https://www.fao.org/faostat/en/#data.
111. Woli, K.P.; Boyer, M.J.; Elmore, R.W.; Sawyer, J.E.; Abendroth, L.J.; Barker, D.W. Corn era hybrid response to nitrogen fertilization. Agron. J. 2016, 108, 473-486. doi:10.2134/agronj2015.0314
112. Brancourt-Hulmel, M.; Doussinault, G.; Lecomte, C.; Bérard, P.; Le Buanec, B.;Trottet, M. Genetic improvement of agronomic traits of winter wheat cultivars released in France from 1946 to 1992. Crop Sci. 2003, 43, 37-45. https://doi.org/10.2135/cropsci2003.3700
113. Andorf, C.; Beavis, W.D.; Hufford, M.; Smith, S.; Suza, W.P.; Wang, K.; Woodhouse, M.; Yu, J.; Lübberstedt, T. Technological advances in maize breeding: past, present and future. Theor. Appl. Genet. 2019, 132, 817-849. https://doi.org/10.1007/s00122-019-03306-3

114. Valenzuela, H.R. Ecologically-based practices for vegetable crops production in the Tropics. HortReviews 2000, 24, 139-228. DOI: 10.1002/9780470650776.ch4
115. Abenavoli, M.R.; Longo, C.; Lupini, A.; Miller, A.J.; Araniti, F.; Mercati, F.; Princi, M.P.; Sunseri, F. Phenotyping two tomato genotypes with different nitrogen use efficiency. Plant Physiol. Biochem. 2016, 107, 21-32. http://dx.doi.org/10.1016/j.plaphy.2016.04.021
116. Villanueva,G.; Rosa-Martínez, E.; Sahin, A.; García-Fortea, E.; Plazas, M.; Prohens, J.; Vilanova, S. Evaluation of advanced backcrosses of eggplant with *Solanum elaeagnifolium* introgressions under low N conditions. Agronomy 2021, 11, 1770. https://doi.org/10.3390/ agronomy11091770
117. Ayadi, S.; Jallouli, S.; Chamekh, Z.; Zouari, I.; Landi, S.; Hammami, Z.; Ben Azaiez, F.E.; Baraket, M.; Esposito, S.; Trifa, Y. Variation of grain yield, grain protein content and nitrogen use efficiency components under different nitrogen rates in mediterranean durum wheat genotypes. Agriculture 2022, 12, 916. https://doi.org/10.3390/ agriculture12070916
118. Di Miceli, G.; Farruggia, D.; Iacuzzi, N.; Bacarella, S.; La Bella, S.; Consentino, B.B. Planting date and different N-fertilization rates differently modulate agronomic and economic traits of a Sicilian onion landrace and of a commercial variety. Horticulturae 2022, 8, 454. https://doi.org/10.3390/ horticulturae8050454
119. Renau-Morata, B., Cebolla-Cornejo, J., Carrillo, L., Gil-Villar, D., Martí, R., Jiménez-Gómez, J.M., Granell, A., Monforte, A.J., Medina, J., Molina, R.V. and Nebauer, S.G., 2024. Identification of *Solanum pimpinellifolium* genome regions for increased resilience to nitrogen deficiency in cultivated tomato. Scientia Horticulturae, 323, p.112497. https://doi.org/10.1016/j.scienta.2023.112497
120. Dawson, J.C.; Huggins, D.R.; Jones, S.S. Characterizing nitrogen use efficiency in natural and agricultural ecosystems to improve the performance of cereal crops in low-input and organic agricultural systems. Field Crops Res. 2008, 107, 89-101. doi:10.1016/j.fcr.2008.01.001
121. Witcombe, A.M.; Tiemann, L.K.; Chikowo, R.; Snapp, S.S. Diversifying with grain legumes amplifies carbon in management-sensitive soil organic carbon pools on smallholder farms. Agric. Ecosyst. Environ. 2023, 356, 108611. https://doi.org/10.1016/j.agee.2023.108611
122. Liang, K.A.N.G.; Liang, Q.Y.; Jiang, Q.; Yao, Y.H.; Dong, M.M.; Bing, H.E.; Gu, M.H. Screening of diverse cassava genotypes based on nitrogen uptake efficiency and yield. J. Integr. Agric. 2020, 19, 965-974. doi: 10.1016/S2095-3119(19)62746-2
123. Mauceri, A.; Bassolino, L.; Lupini, A.; Badeck, F.; Rizza, F.; Schiavi, M.; Toppino, L.; Abenavoli, M.R.; Rotino, G.L.; Sunseri, F. Genetic variation in eggplant for nitrogen use efficiency under contrasting NO3- supply. J. Integr. Plant Biol. 2020, 62, 487-508. doi: 10.1111/jipb.12823
124. Ospina, C.A.; Lammerts van Bueren, E.T.; Allefs, J.J.H.M.; Engel, B.V.; Van der Putten, P.E.L.; Van der Linden, C.G.; Struik, P.C. Diversity of crop development traits and nitrogen use efficiency among potato cultivars grown under contrasting nitrogen regimes. Euphytica 2014, 199, 13-29. DOI 10.1007/s10681-014-1203-4
125. Adam, M.B.; Ulas, A. Vigorous rootstocks improve nitrogen efficiency of tomato by inducing morphological, physiological and biochemical responses. Gesunde Pflanzen 2023, 75, 565-575, https://doi.org/10.1007/s10343-022-00819-8
126. Machado, J.; Fernandes, A.P.G.; Fernandes, T.R.; Heuvelink, E.; Vasconcelos, M.W.; Carvalho, S.M.P. Drought and nitrogen stress effects and tolerance mechanisms in tomato: A review. In: *Plant nutrition and food security in the era of climate change*, Academic Press, Elsevier: Amsterdam, The Netherlands, 2022, pp.315-359. https://doi.org/10.1016/B978-0-12-822916-3.00014-7
127. Bowles, T.M.; Hollander, A.D.; Steenwerth, K,; Jackson L.E. Tightly-coupled plant-soil nitrogen cycling: Comparison of organic farms across an agricultural landscape. PLoS ONE 2015, 10, e0131888. doi:10.1371/journal.pone.0131888
128. Guo, P.; Wang, C.; Jia, Y.; Wang, Q.; Han, G.; Tian, X. Responses of soil microbial biomass and enzymatic activities to fertilizations of mixed inorganic and organic nitrogen at a subtropical forest in East China. Plant Soil 2011, 338, 355-366. DOI 10.1007/s11104-010-0550-8
129. Hobley, E.U.; Honermeier, B.; Don, A.; Gocke, M.I.; Amelung, W.; Kögel-Knabner, I. Decoupling of subsoil carbon and nitrogen dynamics after long-term crop rotation and fertilization. Agric. Ecosyst. Environ. 2018, 265, 363-373. https://doi.org/10.1016/j.agee.2018.06.021
130. Congreves, K.A.; Van Eerd, L.L. Nitrogen cycling and management in intensive horticultural systems. Nutr. Cycling Agroecosyst. 2015, 102, 299-318. DOI 10.1007/s10705-015-9704-7
131. Pinto, R.; Brito, L.M.; Mourão, I.; Coutinho, J. Nitrogen balance in organic horticultural rotations. Acta Hort. 2020, 1286, 127-134. DOI 10.17660/ActaHortic.2020.1286.18
132. Chmelíková, L.; Schmid, H.; Anke, S.; Hülsbergen, K.J. Nitrogen-use efficiency of organic and conventional arable and dairy farming systems in Germany. Nutr. Cycling Agroecosyst. 2021, 119, 337-354. https://doi.org/10.1007/s10705-021-10126-9
133. Karimi, R.; Janzen, H.H.; Smith, E.G.; Ellert, B.H.; Kröbel, R. Nitrogen balance in century-old wheat experiments. Can. J. Soil Sci. 2017, 9, 580-591. dx.doi.org/10.1139/cjss-2016-0118

134. Jones, C.; Jacobsen, J. Fertilizer placement and timing. Nutrient management module, 11. Montana State Univ. Coop. Ext. Serv. 4449-11. Bozeman, MT, USA. 2009.16 pp.
135. Greenwood, D.J.; Cleaver, T.J.; Turner, M.K.; Hunt, J.; Niendorf, K.B.; Loquens, S.M.H. Comparison of the effects of nitrogen fertilizer on the yield, nitrogen content and quality of 21 different vegetable and agricultural crops. J. Agric. Sci. 1980, 95, 471-485. doi:10.1017/S0021859600039514
136. Gao, Q.; Li, C.; Feng, G.; Wang, J.; Cui, Z.; Chen, X.; Zhang, F. Understanding yield response to nitrogen to achieve high yield and high nitrogen use efficiency in rainfed corn. Agron. J., 2012, 104, 165-168. doi:10.2134/agronj2011.0215
137. Pickens, J.M.; Danaher, J.J.; Sibley, J.L.; Chappell, J.A.; Hanson, T.R. Integrating greenhouse cherry tomato production with biofloc tilapia production. Horticulturae 2020, 6, 44. doi:10.3390/horticulturae6030044
138. Hahn, L.; Kurtz, C.; Paula, B.V.; Feltrim, A.L.; Higashikawa, F.S.; Moreira, C.; Rozane, D.E.; Brunetto, G.; Parent, L.É. Customized nutrient management of onion (*Allium cepa*) agroecosystems. Res. Sq. 2023, March 18. DOI:https://doi.org/10.21203/rs.3.rs-2556545/v1
139. Hochmuth, G.J.; Hanlon, E. Plant tissue analysis and interpretation for vegetable crops in Florida. Fla. Coop. Ext. Serv. HS964. Gainesville, Florida, USA. 50 pp. 2022a
140. Jat, R.A.; Wani, S.P.; Sahrawat, K.L.; Singh, P.; Dhaka, P.L. Fertigation in vegetable crops for higher productivity and resource use efficiency. Indian Journ. Fert. 2011, 7, 22-37.
141. Prasad R.; Hochmuth G.J.; Boote K.J. Estimation of nitrogen pools in irrigated potato production on sandy soil using the model SUBSTOR. PLoS One 2015, 10, e0117891. doi: 10.1371/journal.pone.
142. Smith, L.G.; Tarsitano, D.; Topp, C.F.; Jones, S.K.; Gerrard, C.L.; Pearce, B.D.; Williams, A.G.; Watson, C.A. Predicting the effect of rotation design on N, P, K balances on organic farms using the NDICEA model. Renew. Agric. Food Syst. 2016, 31, 471-484. DOI:10.1017/S1742170515000381
143. Zaeen, A.A. Improving nitrogen management in potatoes with active optical sensors. Ph.D. Thesis, The University of Maine. Orono. 2020, 200 pp.
144. Higashikawa, F.S.; Cantú, R.R.; Jindo, K.; Kurtz, C.; de Souza Gonçalves, P.A.; Vieira Neto, J. Use of compost in onion cultivation under no-tillage system: Effect on nutrient uptake. Commun. Soil Sci. Plant Anal. 2023, 54,1215-1238. https://doi.org/10.1080/00103624.2022.2139388
145. Fennimore, S.A.; Richard, S.J.; Flewelling, N.L. Crop Profile for Carrots in California. Univ. Calif. Coop. Ext. Serv. USDA OPMP & Pesticide Assessment program (PIAP), Washington D.C. 2000.
146. Anon. Crop Profile for carrots in Florida. Univ. Florida Coop. Ext. Serv. USDA Office of Pest Management Policy (OPMP) & Pesticide Assessment program (PIAP), Washington D.C. 1999
147. Tyler. K.B. Nitrogen fertilization of California vegetable crops. Univ. Calif. Vegetable Briefs, Jan. 8, 1975, Issue No. 175, Davis, California, 1975
148. Takele, E.; Daugovish, O.; Vue, M. Costs and profitability analysis for celery production in the Oxnard Plain, Ventura County, 2012–2013. Univ. California Coop. Ext., Ventura, CA., 2013.
149. Knuteson, D.L.; Groves, R.L.; Colquhoun, J.B.; Ruark, M.; Gevens, A.J.; Bussan, A.J. BioIPM Vine Crops Workbook. Univ. Wisc. Coop. Ext. Serv. Madison, Wisconsin. 2012, 118 pp.
150. Anon. Crop profile for squash in California, USDA Office of Pest Management Policy (OPMP) & Pesticide Assessment program (PIAP), Washington D.C. 2000.
151. Hochmuth, G.; Hanlon, E.A. Summary of N, P, and K research with watermelon in Florida: Univ. Florida Coop. Ext. Serv. SL325, Gainesville, Florida, USA, 20 pp. 2020c
152. Pfeufer, E.E. Sources of inoculum, epidemiology, and integrated management of bacterial rots of onion (*Allium cepa*) with a focus on center rot, caused by *Pantoea ananatis* and *Pantoea agglomerans*. Ph.D. Thesis. The Pennsylvania State University. State College, PA., 2014. 149 pp.
153. Alston, D.G. Onion thrips (*Thrips tabaci*). Utah State Univ. Coop. Ext. Serv. Fact Sheet. ENT-117-08PR. Logan, Utah. 2008, 7 pp.
154. Waters, T.; Wohleb, C. Thrips. Washington State Coop. Ext. Serv. Fact Sheet, FS126E, Pullman, Wash., USA. 2014.
155. Coolong, T.; da Silva, A.L.B.R.; Shealey, J. Fertilizer program impacts yield and blossom end rot in bell pepper. HortTechnology 2019, 29, 163-169. https://doi.org/10.21273/HORTTECH04249-18
156. Le Strange, M.; Schrader, W.; Hartz, T. Fresh-market tomato production in California. Univ. Calif. Coop. Ext. Serv., Publication 8017. UCANR Publications. Berkeley, California. 2000.
157. Gatton, H.; Nessler, S.; Kuhar, T.; Jennings, K.; King, S.; Monks, D.; Rideout, S.; Troth, S.; Waldenmeire, C.; Weaver, M.; Wison, H. Pest management strategic plan for tomato in Virginia, North Carolina and Delaware. Southern Region IPM Center, Virginia Tech, North Carolina State University, University of Delaware. 2007.
158. Jalpa, L.; Mylavarapu, R. Current tomato production practices and their effects on plant and soil carbon and nitrogen dynamics. J. Plant Nutr. 2023, 16, 3905–3917. https://doi.org/10.1080/01904167.2023.2220708
159. Goulding, K.; Jarvis, S.; Whitmore, A. Optimizing nutrient management for farm systems. Philos. Trans. R. Soc. Lond. B Biol. Sci. 2008, 363, 667-680. doi:10.1098/rstb.2007.2177

160. Martinez, D.A.; Loening, U.E.; Graham, M.C.; Gathorne-Hardy, A. When the medicine feeds the problem; Do nitrogen fertilisers and pesticides enhance the nutritional quality of crops for their pests and pathogens? Front. Sustain. Food Syst. 2021, 5, 701310. doi: 10.3389/fsufs.2021.701310
161. Hochmuth, G.; Hanlon, E. The four Rs of fertilizer management. Fla. Coop. Ext. Serv. SL411. Gainesville, Florida, USA. 5 pp. 2022b
162. Ahmad, N. Fertilizer best management practices in Pakistan. In: *Fertilizer best management practices*, papers presented at the IFA international workshop on fertilizer best management practices, 7-9 March 2007, Brussels, Belgium. International Fertilizer Industry Association (IFA), Paris, 2007, pp. 213-220.
163. Hartz, T. Nitrogen management, evaluating practices for coastal production of Brassica crops, Vegetables West Mag. 2001, May, pg. 11
164. Anon. Nitrogen pollution and the European environment implications for air quality policy. Science for Environment Policy, European Commission. Prepared by the Science Communication Unit, University of the West of England, Bristol. September. 2013, 28 pp.
165. Overeem, R. Physiology and genetic variation of nitrogen use efficiency in spinach (*Spinacia oleracea* L.). MS Thesis. Wageningen University, Wageningen, Netherlands 2015, 86 pp.
166. Rao, I.M.; Miles, J.W.; Beebe, S.E.; Horst, W.J. Root adaptations to soils with low fertility and aluminium toxicity. Ann. Bot. 2016, 118, 593-605. doi:10.1093/aob/mcw073
167. Zhang, M.; Fan, C.H.; Li, Q.L.; Li, B.; Zhu, Y.Y.; Xiong, Z.Q. A 2-yr field assessment of the effects of chemical and biological nitrification inhibitors on nitrous oxide emissions and nitrogen use efficiency in an intensively managed vegetable cropping system. Agric. Ecosyst. Environ. 2015a, 201, 43-50. http://dx.doi.org/10.1016/j.agee.2014.12.003
168. Dodgson, J.; Weston, A.K.; Marks, D.J. Use of stabilised amine nitrogen (SAN) reduces required nitrogen input and increases yield of onions (*Allium cepa* L.). Crops 2023, 3, 148–157. https://doi.org/10.3390/crops3020015
169. Das, D.; Dwivedi, B.S.; Meena, M.C.; Singh, V.K.; Tiwari, K.N. Integrated nutrient management for improving soil health and crop productivity. Indian J. Fert. 2015, 11, 64-83
170. Zotarelli, L.; Dukes, M.D.; Scholberg, J.M.; Hanselman, T.; Le Femminella, K.; Munoz-Carpena, R. Nitrogen and water use efficiency of zucchini squash for a plastic mulch bed system on a sandy soil. Sci. Hortic. 2008, 116, 8-16. doi:10.1016/j.scienta.2007.10.029
171. Danso, E.O.; Abenney-Mickson, S.; Sabi, E.B.; Plauborg, F.; Abekoe, M.; Kugblenu, Y.O.; Jensen, C.R.; Andersen, M.N. Effect of different fertilization and irrigation methods on nitrogen uptake, intercepted radiation and yield of okra (*Abelmoschus esculentum* L.) grown in the Keta Sand Spit of Southeast Ghana. Agric. Water Manag. 2015, 147, 34-42. http://dx.doi.org/10.1016/j.agwat.2014.07.029
172. Russo, V.M. Plant density and nitrogen fertilizer rate on yield and nutrient content of onion developed from greenhouse-grown transplants. HortScience 2008, 43, 1759-1764. https://doi.org/10.21273/HORTSCI.43.6.1759
173. Kirimi, J.K.; Itulya, F.M.; Mwaja, V.N. Effects of nitrogen and spacing on fruit yield of tomato. African J. Hort. Sci. 2011, 5, 50-60.
174. Gastal, F.; Lemaire, G.; Durand, J.L.; Louarn, G. Quantifying crop responses to nitrogen and avenues to improve nitrogen-use efficiency. In V.O. Sadras and D. Calderini (Eds.) *Crop physiology.* Academic Press. London. 2015. pp. 161-206. DOI: 10.1016/B978-0-12-417104-6.00008-X
175. Vidigal, S.M.; Moreira, M.A.; Paes, J.; Pedrosa, M.W. Does high onion plant density increase nitrogen demand?. Rev. Caatinga 2023, 36, 381-389. http://dx.doi.org/10.1590/1983-21252023v36n215rc
176. Liu, R.; Yang, Y.; Wang, Y.S.; Wang, X.C.; Rengel, Z.; Zhang, W.J.; Shu, L.Z. Alternate partial root-zone drip irrigation with nitrogen fertigation promoted tomato growth, water and fertilizer-nitrogen use efficiency. Agric. Water Manag. 2020, 233, 106049. https://doi.org/10.1016/j.agwat.2020.106049
177. Plaza, B.M.; Lao, M.T.; Jiménez-Becker, S. Fertigation strategies to alleviate fertilizer contamination generated by tomato crops under plastic greenhouses. Agronomy 2021, 11, 444. https://doi.org/10.3390/agronomy11030444
178. Huang, Y.; Brown, M. Advancing to the next generation of precision agriculture. Pp. 285-314. In: R. Serraj and P. Pingali (Eds.) *Agriculture & Food Systems to 2050: Global Trends, Challenges and Opportunities*. World Scientific Pub. Singapore. 2019.
179. Padilla, F.M.; Farneselli, M.; Gianquinto, G.; Tei, F.; Thompson, R.B. Monitoring nitrogen status of vegetable crops and soils for optimal nitrogen management. Agric. Water Manag. 2020, 241, 106356. https://doi.org/10.1016/j.agwat.2020.106356
180. Colla, G.; Suárez, C.M.C.; Cardarelli, M.; Rouphael, Y. Improving nitrogen use efficiency in melon by grafting. HortScience 2010, 45, 559-565. https://doi.org/10.21273/HORTSCI.45.4.559
181. Liu, C.; Plaza-Bonilla, D.; Coulter, J.A.; Kutcher, H.R.; Beckie, H.J.; Wang, L.; Floc'h, J.B.; Hamel, C.; Siddique, K.H.; Li, L.; Gan, Y. Diversifying crop rotations enhances agroecosystem services and resilience. Adv. Agron. 2022a, 173, 299-335. DOI: 10.1016/bs.agron.2022.02.007

182. Sainju, U.M.; Ghimire, R.; Pradhan, G.P. Nitrogen fertilization II: Management practices to sustain crop production and soil and environmental quality. In: E. Rigobelo and P.A. Serra (Eds.) N*itrogen Fixation*. IntechOpen., London, 2019, pp. 1-23. DOI: http://dx.doi.org/10.5772/intechopen.86646
183. Breza, L.C.; Mooshammer, M.; Bowles, T.M.; Jin, V.L.; Schmer, M.R.; Thompson, B.; Grandy, A.S. Complex crop rotations improve organic nitrogen cycling. Soil Biol. Biochem. 2023, 177, 108911. https://doi.org/10.1016/j.soilbio.2022.108911
184. Simonne E.; Hochmuth G. Double cropping vegetables grown with plasticulture in Florida in the BMP Era. Univ. Florida Coop. Ext. Serv. HS908/HS165, IFAS, EDIS. Gainesville, Florida. 2003. https://doi.org/10.32473/edis-hs165-2003.
185. Matsumura, A.; Hirosawa, K.; Masumoto, H.; Daimon, H. Effects of maize as a catch crop on subsequent garland chrysanthemum and green soybean production in soil with excess nitrogen. Sci. Hortic. 2020, 273, 109640. https://doi.org/10.1016/j.scienta.2020.109640
186. Ma, Y.; Kang, L.; Li, Y.; Zhang, X.; Cardenas, L.M.; Chen, Q. Is sorghum a promising summer catch crop for reducing nitrate accumulation and enhancing eggplant yield in intensive greenhouse vegetable systems? Plant Soil 2023, 11, 1-13. https://doi.org/10.1007/s11104-023-05923-w
187. Re, M.I.Z.; Rath, S.; Dukes, M.D.; Graham, W. Water and nitrogen budget dynamics for a maize-peanut rotation in Florida. Trans. ASABE 2020, 63, 2003-2020. https://doi.org/10.13031/trans.13916
188. West, J.R.; Ruark, M.D.; Bussan, A.J.; Colquhoun, J.B.; Silva, E.M. Nitrogen and weed management for organic sweet corn production on loamy sand. Agron. J. 2016, 108, 758-769. doi:10.2134/agronj2015.0393
189. Toda, M.; Walder, F.; van der Heijden, M.G. Organic management and soil health promote nutrient use efficiency. J. Sust. Agric. Environ. (JSAE) 2023, 2, 215-224. DOI: 10.1002/sae2.12058
190. Bender, S.F.; Schulz, S.; Martínez-Cuesta, R.; Laughlin, R.J.; Kublik, S.; Pfeiffer-Zakharova, K.; Vestergaard, G.; Hartman, K.; Parladé, E.; Römbke, J.; Watson, C.J. Simplification of soil biota communities impairs nutrient recycling and enhances above-and belowground nitrogen losses. New Phytol. 2023, 240: 2020–2034, doi: 10.1111/nph.19252
191. Bressler, A.; Blesh, J. A grass-legume cover crop maintains nitrogen inputs and nitrous oxide fluxes from an organic agroecosystem. Ecosphere 2023, 14, p.e4428. DOI: 10.1002/ecs2.4428
192. Sinha, R.K.; Agarwal, S.; Chauhan, K.; Valani, D. The wonders of earthworms & its vermicompost in farm production: Charles Darwin's 'friends of farmers', with potential to replace destructive chemical fertilizers. Agric. Sci. 2010, 1, 76-94. doi:10.4236/as.2010.12011
193. Coppens, J.; Grunert, O.; Van Den Hende, S.; Vanhoutte, I.; Boon, N.; Haesaert, G.; De Gelder, L. The use of microalgae as a high-value organic slow-release fertilizer results in tomatoes with increased carotenoid and sugar levels. J. Appl. Phycol. 2016, 28, 2367-2377. DOI 10.1007/s10811-015-0775-2
194. Hassan,S.M.; Ashour,M.; Soliman, A.A.F.; Hassanien, H.A.; Alsanie, W.F.; Gaber, A.; Elshobary, M.E. The potential of a new commercial seaweed extract in stimulating morpho-agronomic and bioactive properties of *Eruca vesicaria* (L.) Cav. Sustainability 2021, 13, 4485. https://doi.org/10.3390/su13084485
195. Braun, J.C.; Colla, L.M. Use of microalgae for the development of biofertilizers and biostimulants. BioEnergy Res. 2023, 16, 289-310. https://doi.org/10.1007/s12155-022-10456-8
196. Libutti, A.; Russo, D.; Lela, L.; Ponticelli, M.; Milella, L.; Rivelli, A.R. Enhancement of yield, phytochemical content and biological activity of a leafy vegetable *(Beta vulgaris* L. var. cycla) by using organic amendments as an alternative to chemical fertilizer. Plants, 2023,12, 569. https://doi.org/ 10.3390/plants12030569
197. Youssef, M.A.; Yousef, A.F.; Ali, M.M.; Ahmed, A.I.; Lamlom, S.F.; Strobel, W.R.; Kalaji, H.M. Exogenously applied nitrogenous fertilizers and effective microorganisms improve plant growth of stevia (*Stevia rebaudiana* Bertoni) and soil fertility. AMB Express, 2021, 11, 1-10. https://doi.org/10.1186/s13568-021-01292-8
198. Choi, W.J.; Ro, H.M.; Chang, S.X. Recovery of fertilizer-derived inorganic-15 N in a vegetable field soil as affected by application of an organic amendment. Plant Soil 2004, 263, 191-201. https://doi.org/10.1023/B:PLSO.0000047726.09394.d3
199. Koudahe, K.; Allen, S.C.; Djaman, K. Critical review of the impact of cover crops on soil properties. Int. Soil Water Conserv. Res. (ISWCR) 2022, 10, 343-354. https://doi.org/10.1016/j.iswcr.2022.03.003
200. Quintarelli, V.; Radicetti, E.; Allevato, E.; Stazi, S.R.; Haider, G.; Abideen, Z.; Bibi, S.; Jamal, A.; Mancinelli, R. Cover crops for sustainable cropping systems: A review. Agriculture 2022, 12, 2076. https://doi.org/10.3390/agriculture12122076
201. Xie, Y.; Kristensen, H.L. Overwintering grass-clover as intercrop and moderately reduced nitrogen fertilization maintain yield and reduce the risk of nitrate leaching in an organic cauliflower (*Brassica oleracea* L. var. *botrytis*) agroecosystem. Sci. Hortic. 2016, 206, 71-79. http://dx.doi.org/10.1016/j.scienta.2016.04.034
202. Yang, H.; Zhang, W.; Li, L. Intercropping: Feed more people and build more sustainable agroecosystems. Front. Agric. Sci. Eng. 2021, 8, 373-386. https://doi.org/10.15302/J-FASE-2021398
203. Shanmugam, S.; Hefner, M.; Pelck, J.S.; Labouriau, R.; Kristensen, H.L. Complementary resource use in intercropped faba bean and cabbage by increased root growth and nitrogen use in organic production. Soil Use Manag. 2022, 38, 729-740. DOI: 10.1111/sum.12765

204. Lai, H.; Gao, F.; Su, H.; Zheng, P.; Li, Y.; Yao, H. Nitrogen distribution and soil microbial community characteristics in a legume–cereal intercropping system: A review. Agronomy 2022, 12,1900. https://doi.org/10.3390/ agronomy12081900
205. Muhie, S.H. Novel approaches and practices to sustainable agriculture. J. Agric. Food Res. 2022, 100446. https://doi.org/10.1016/j.jafr.2022.100446
206. Wolz, K.J.; Branham, B.E.; DeLucia, E.H. Reduced nitrogen losses after conversion of row crop agriculture to alley cropping with mixed fruit and nut trees. Agric. Ecosyst. Environ. 2018, 258, 172-181. https://doi.org/10.1016/j.agee.2018.02.024
207. Rosati, A.; Borek, R.; Canali, S. Agroforestry and organic agriculture. Agrofor. Syst. 2021, 95, 805-821. https://doi.org/10.1007/s10457-020-00559-6
208. Shao, G.; Martinson, G.O.; Corre, M.D.; Luo, J.; Niu, D.; Bischel, X.; Veldkamp, E. Impacts of monoculture cropland to alley cropping agroforestry conversion on soil N2O emissions. GCB Bioenergy. 2023, 15, 58-71. DOI: 10.1111/gcbb.13007
209. Brooker, R.W.; Bennett, A.E.; Cong, W.F.; Daniell, T.J.; George, T.S.; Hallett, P.D.; Hawes, C.; Iannetta, P.P.; Jones, H.G.; Karley, A.J.; Li, L. Improving intercropping: a synthesis of research in agronomy, plant physiology and ecology. New Phytol. 2015, 206, 107-117. doi: 10.1111/nph.13132
210. Li, H.; Hill, N.; Wallace, J. A perennial living mulch system fosters a more diverse and balanced soil bacterial community. PloS one, 2023a, 18, p.e0290608. https://doi.org/10.1371/journal. Pone.0290608
211. van Ruijven, J.; Berendse, F. Diversity-productivity relationships: initial effects, long-term patterns, and underlying mechanisms. PNAS 2005, 102, 695-700. www.pnas.org/cgi/doi/10.1073/pnas.0407524102
212. Doran, J. W.; & Smith, M. S. Role of cover crops in nitrogen cycling. In W. L. Hargrove (Ed.), *Cover crops for clean water*, Soil and Water Conservation Society. Ankeny, IA, 1991, pp. 85-90
213. Finney, D.M.; White, C.M.; Kaye, J.P. Biomass production and carbon/nitrogen ratio influence ecosystem services from cover crop mixtures. Agron. J., 2016, 108, 39-52. doi:10.2134/agronj15.0182
214. Van Eerd, L.L.; Chahal, I.; Peng, Y.; Awrey, J.C. Influence of cover crops at the four spheres: A review of ecosystem services, potential barriers, and future directions for North America. Sci. Total Environ. 2023, 858, 159990. http://dx.doi.org/10.1016/j.scitotenv.2022.159990
215. Kuo, S.; Sainju, U.M. Nitrogen mineralization and availability of mixed leguminous and non-leguminous cover crop residues in soil. Biol. Fert. Soils 1988, 26, 346-353. https://doi.org/10.1007/s003740050387
216. Chinta, Y.D.; Uchida, Y.; Araki, H. Availability of nitrogen supply from cover crops during residual decomposition by soil microorganisms and its utilization by lettuce (*Lactuca sativa* L.). Sci. Hortic. 2020, 270, 109415. https://doi.org/10.1016/j.scienta.2020.109415
217. White, K.E.; Brennan, E.B.; Cavigelli, M.A.; Smith, R.F. Winter cover crops increased nitrogen availability and efficient use during eight years of intensive organic vegetable production. PLoS One 2022, 17, e0267757. https://doi.org/ 10.1371/journal.pone.0267757
218. Muchanga, R.A.; Araki, H. Cover crop residue management for effective use of mineralized nitrogen in greenhouse tomato production. In: Ohyama, T.; Inubushi, K. (Eds.) *Nitrogen in Agriculture-Physiological, Agricultural and Ecological Aspects*. IntechOpen. London. 2021, pp. 1-18. DOI: 10.5772/intechopen.95359
219. Collins, H.P.; Delgado, J.A.; Alva, A.K.; Follett, R.F. Use of Nitrogen-15 isotopic techniques to estimate nitrogen cycling from a mustard cover crop to potatoes. Agron. J. 2007, 99, 27-35. doi:10.2134/agronj2005.0357
220. Delgado, J.A.; Mosquera, V.H.B.; Alwang, J.R.; Villacis-Aveiga, A.; Ayala, Y.E.C.; Neer, D.; Monar, C.; López, L.O.E. Potential use of cover crops for soil and water conservation, nutrient management, and climate change adaptation across the tropics. Adv. Agron. 2021, 165, 175-247. https://doi.org/10.1016/bs.agron.2020.09.003
221. Jahanzad, E.; Barker, A.V.; Hashemi, M.; Sadeghpour, A.; Eaton, T. Forage radish and winter pea cover crops outperformed rye in a potato cropping system. Agron. J. 2017, 109, 646-653. doi:10.2134/agronj2016.06.0342
222. Feng, L.; Yang, W.T.; Zhou, Q.; Tang, H.Y.; Ma, Q.Y.; Huang, G.Q.; Wang, S.B. Effects of interspecific competition on crop yield and nitrogen utilisation in maize-soybean intercropping system. Plant Soil Environ. 2021, 67, 460-467. https://doi.org/10.17221/665/2020-PSE
223. Chen, P.; Du, Q.; Liu, X.; Zhou, L.I.; Hussain, S.; Lei, L.U.; Song, C.; Wang, X.; Liu, W.; Yang, F.; Shu, K. Effects of reduced nitrogen inputs on crop yield and nitrogen use efficiency in a long-term maize-soybean relay strip intercropping system. PloS ONE 2017, 12, e0184503. https://doi. org/10.1371/journal.pone.0184503
224. Yang, F.; Liao, D.; Wu, X.; Gao, R.; Fan, Y.; Raza, M.A.; Wang, X.; Yong, T.; Liu, W.; Liu, J.; Du, J. Effect of aboveground and belowground interactions on the intercrop yields in maize-soybean relay intercropping systems. Field Crops Res. 2017, 203, 16-23. http://dx.doi.org/10.1016/j.fcr.2016.12.007
225. Chen, N.; Li, X.; Šimůnek, J.; Shi, H.; Zhang, Y.; Hu, Q. Quantifying inter-species nitrogen competition in the tomato-corn intercropping system with different spatial arrangements. Agric. Syst. 2022, 201, 103461, https://doi.org/10.1016/j.agsy.2022.103461

226. Warren Raffa, D.; Migliore, M.; Campanelli, G.; Leteo, F.; Trinchera, A. Effects of faba bean strip cropping in an outdoor organic tomato system on soil nutrient availability, production, and N budget under different fertilizations. Agronomy 2022, 12, 1372. https://doi.org/10.3390/ agronomy12061372
227. Guo, J.Y.; Bradshaw, A.D. The flow of nutrients and energy through a Chinese farming system. J. Appl. Ecol. 1993, 30, 86-94. https://doi.org/10.2307/2404273
228. Fruscella, L.; Kotzen, B.; Paradelo, M.; Milliken, S. Investigating the effects of fish effluents as organic fertilisers on onion (*Allium cepa*) yield, soil nutrients, and soil microbiome. Sci. Hortic. 2023, 321, 112297. https://doi.org/10.1016/j.scienta.2023.112297
229. Valenzuela, H.R.; Schaffer, B.; S.K. O'Hair, S.K. Shade and nitrogen influence gas exchange and growth of cocoyam (*Xanthosoma sagittifolium*). J. Amer. Soc. Hort. Sci. 1990, 115, 1014-1018. https://doi.org/10.21273/JASHS.115.6.1014
230. Robertson, G.P.; Bruulsema, T.W.; Gehl, R.J.; Kanter, D.; Mauzerall, D.L.; Rotz, C.A.; Williams, C.O. Nitrogen-climate interactions in US agriculture. Biogeochemistry 2013, 114, 41-70. DOI 10.1007/s10533-012-9802-4
231. Djaman, K.; Irmak, S.; Koudahe, K.; Allen, S. Irrigation management in potato (*Solanum tuberosum* L.) production: A review. Sustainability 2021, 13, 1504. https://doi.org/10.3390/su13031504
232. Wu, Y.; Yan, S.; Fan, J.; Zhang, F.; Xiang, Y.; Zheng, J.; Guo, J. Responses of growth, fruit yield, quality and water productivity of greenhouse tomato to deficit drip irrigation. Sci. Hortic. 2021, 275, 109710. https://doi.org/10.1016/j.scienta.2020.109710
233. Nayak, H.S.; Silva, J.V.; Parihar, C.M.; Kakraliya, S.K.; Krupnik, T.J.; Bijarniya, D.; Jat, M.L.; Sharma, P.C.; Jat, H.S.; Sidhu, H.S.; Sapkota, T.B. Rice yield gaps and nitrogen-use efficiency in the Northwestern Indo-Gangetic Plains of India: Evidence based insights from heterogeneous farmers' practices. Field Crops Res. 2022, 275, 108328. https://doi.org/10.1016/j.fcr.2021.108328
234. Schmid, C.J. Influence of nutrient management and soil pH on anthracnose severity of annual bluegrass putting green turf (Doctoral dissertation, Rutgers University-Graduate School-New Brunswick). New Brunswick, New Jersey. 2016, 298 pp.
235. Plaza-Bonilla, D.; Cantero-Martínez, C.; Bareche, J.; Arrúe, J.L.; Lampurlanés, J.; Álvaro-Fuentes, J. Do no-till and pig slurry application improve barley yield and water and nitrogen use efficiencies in rainfed Mediterranean conditions? Field Crops Res. 2017, 203, 74-85. http://dx.doi.org/10.1016/j.fcr.2016.12.008
236. Jagtap, S.S.; Abamu, F.J. Matching improved maize production technologies to the resource base of farmers in a moist savanna. Agric. Syst. 2003, 76, 1067-1084. https://doi.org/10.1016/S0308-521X(02)00040-9
237. Mu, T.; Yue, X.; Zang, Z.; Wang, H.; Liang, J.; Yang, Q.; Guo, J.; Li, N.; Liu, X.; You, Q. Coupling effect of water and soluble organic fertilizer on yield and quality of *Panax notoginseng* under micro-sprinkler irrigation in Southwest China. Agronomy 2023, 13, 1742. https://doi.org/10.3390/ agronomy13071742
238. Cassman, K.G. Ecological intensification of agriculture and implications for improved water and nutrient management. In: P. Imas.; M.R. Price (Eds.) *Fertigation Proceedings: Optimizing the Utilization of Water and Nutrients*, Beijing, 20-24 September 2005, International Potash Institute, Horgen, Switzerland, 2008, pp. 23-34, 2005, DOI 10.3235/978-3-9523243-8-7
239. Ayankojo, I.T.; Morgan, K.T. Optimizing tomato growth and productivity using nitrogen and irrigation application timing. Agronomy 2021, 11, 1968. https://doi.org/10.3390/agronomy11101968
240. Li, S.; Tan, D.; Wu, X.; Degré, A.; Long, H.; Zhang, S.; Lu, J.; Gao, L.; Zheng, F.; Liu, X.; Liang, G. Negative pressure irrigation increases vegetable water productivity and nitrogen use efficiency by improving soil water and NO3-N distributions. Agric. Water Manag. 2021, 251, 106853. https://doi.org/10.1016/j.agwat.2021.106853
241. Amer, K.H.; Midan, S.A.; Hatfield, J.L. Effect of deficit irrigation and fertilization on cucumber. Agron. J., 2009, 101, 1556-1564. https://doi.org/10.2134/agronj2009.0112
242. Nhantumbo, N.S. Residue management strategies for the rainfed N-deprived maize-legume cropping systems of central Mozambique. Ph.D. Thesis. University of Queensland, Brisbane, Australia. 2016, 205 pp.
243. Anderson, R.L. Are some crops synergistic to following crops? Agron. J., 2005, 97, 7-10. https://doi.org/10.2134/agronj2005.0007a
244. Tomer, M.D.; Liebman, M. Nutrients in soil water under three rotational cropping systems, Iowa, USA. Agric. Ecosyst. Environ. 2014, 186, 105-114. http://dx.doi.org/10.1016/j.agee.2014.01.025
245. Kishan, K.; Rondla, S.K.; Kumar, K.S.; Naik, S. Effect of Irrigation and N-fertigation Levels on Broccoli Performance in a Polynet House. Int. J. Environ. Clim. 2021, 11, 261-267. DOI: 10.9734/IJECC/2021/v11i1230576
246. Cui, B.J.; Niu, W.Q.; Du, Y.D.; Zhang, Q. Response of yield and nitrogen use efficiency to aerated irrigation and N application rate in greenhouse cucumber. Sci. Hortic. 2020, 265,109220, https://doi.org/10.1016/j.scienta.2020.109220
247. Mwinuka, P.R.; Mbilinyi, B.P.; Mbungu, W.B.; Mourice, S.K.; Mahoo, H.F.; Schmitter, P. Optimizing water and nitrogen application for neglected horticultural species in tropical sub-humid climate areas: A case of

African eggplant (*Solanum aethiopicum* L.). Sci. Hortic. 2021, 276, 109756. https://doi.org/10.1016/j.scienta.2020.109756
248. Halvorson, A.D.; Bartolo, M.E.; Reule, C.A.; Berrada, A. Nitrogen effects on onion yield under drip and furrow irrigation. Agron. J., 2008, 100, 1062-1069. doi:10.2134/agronj2007.0377
249. Piri, H.; Naserin, A. Effect of different levels of water, applied nitrogen and irrigation methods on yield, yield components and IWUE of onion. Sci. Hortic. 2020, 268, 109361. https://doi.org/10.1016/j.scienta.2020.109361
250. Hamad, A.A.A.; Wei, Q.; Wan, L.; Xu, J.; Hamoud, Y.A.; Li, Y.; Shaghaleh, H. Subsurface drip irrigation with emitters placed at suitable depth can mitigate N2O emissions and enhance chinese cabbage yield under greenhouse cultivation. Agronomy 2022, 12, 745. https://doi.org/10.3390/ agronomy12030745
251. Ohnishi, M.; Horie, T.; Homma, K.; Supapoj, N.; Takano, H.; Yamamoto, S. Nitrogen management and cultivar effects on rice yield and nitrogen use efficiency in Northeast Thailand. Field Crops Res. 1999, 64, 109-120. DOI:10.1016/S0378-4290(99)00054-4
252. Marschner, H. Mineral nutrition of higher plants. Second edition. London. Academic press. London, UK. 1995, 889 pp.
253. Sanginga, D.A. Effects of potassium and magnesium on cassava vegetative and root yield. MS Thesis, University of Eldoret. Eldoret, Kenya. 2022, 101 pp.
254. Gholamnejad, S.; Haghighi, M.; Etemadi, N.; Pessarakli, M. The effects of N-NO3: N-NH4 ratios and calcium concentration of the nutrient solution on the growth parameters and partitioning of nitrogen and calcium in tomato plants (*Solanum lycopersicum* L.). J. Plant Nutr. 2023, 46, 2827-2840. https://doi.org/10.1080/01904167.2022.2160753
255. Fenn, L.B.; Taylor, R.M. Calcium stimulation of ammonium absorption in radish. Agron. J. 1990, 82, 81-84. https://doi.org/10.2134/agronj1990.00021962008200010018x
256. Fageria, V.D. Nutrient interactions in crop plants. J. Plant Nutr. 2001, 24, 1269-1290. http://dx.doi.org/10.1081/PLN-100106981
257. Sanginga, N.; Woomer, P.L. Eds. Integrated soil fertility management in Africa: Principles, practices, and developmental process. Tropical Soil Biology and Fertility Institute of the International Centre for Tropical Agriculture, CIAT. Nairobi, Kenya. 2009, 263 pp.
258. Dufault, R.J. Relationship among nitrogen, phosphorus, and potassium fertility regimes on celery transplant growth. HortScience 1985, 20, 1104-1106. https://doi.org/10.21273/HORTSCI.20.6.1104
259. Sabatino, L.; La Bella, S.; Ntatsi, G.; Iapichino, G.; D'Anna, F.; De Pasquale, C.; Consentino, B.B.; Rouphael, Y. Selenium biofortification and grafting modulate plant performance and functional features of cherry tomato grown in a soilless system. Sci. Hortic. 2021, 285, 110095. https://doi.org/10.1016/j.scienta.2021.110095
260. Carucci, F.;Gatta, G.; Gagliardi, A.; De Vita, P.; Bregaglio, S.; Giuliani, M.M. Agronomic strategies to improve N efficiency indices in organic durum wheat grown in Mediterranean area. Plants 2021, 10, 2444. https://doi.org/10.3390/ plants10112444
261. Przygocka-Cyna, K.; Barłóg, P.; Grzebisz, W.; Spiżewski, T. Onion (*Allium cepa* L.) yield and growth dynamics response to in-season patterns of nitrogen and sulfur uptake. Agronomy 2020, 10, 1146. doi:10.3390/agronomy10081146
262. El Amerany, F.; Rhazi, M.; Wahbi, S.; Taourirte, M.; Meddich, A. The effect of chitosan, arbuscular mycorrhizal fungi, and compost applied individually or in combination on growth, nutrient uptake, and stem anatomy of tomato. Sci. Hortic. 2020, 261, 109015. https://doi.org/10.1016/j.scienta.2019.109015
263. Moustaka, J.; Moustakas, M. Early-stage detection of biotic and abiotic stress on plants by chlorophyll fluorescence imaging analysis. Biosensors 2023, 13, 796. https://doi.org/10.3390/ bios13080796
264. Bhardwaj, R.; Lone, J.K.; Pandey, R.; Mondal, N.; Dhandapani, R.; Meena, S.K.; Khan, S. Insights into morphological and physio-biochemical adaptive responses in mungbean (*Vigna radiata* L.) under heat stress. Front. Genet. 2023, 14, 1206451, doi: 10.3389/fgene.2023.1206451
265. Marino, D.; Moran, J.F. Can ammonium stress be positive for plant performance? Front. Plant Sci. 2019, 10, 1103. doi: 10.3389/fpls.2019.01103
266. Nazir, F.; Mahajan, M.; Khatoon, S.; Albaqami, M.; Ashfaque, F.; Chhillar, H.; Chopra, P.; Khan, M.I.R. Sustaining nitrogen dynamics: A critical aspect for improving salt tolerance in plants. Front. Plant Sc. 2023, 14, 1087946. doi: 10.3389/fpls.2023.1087946
267. Shangguan, Z.P.; Shao, M.A.; Dyckmans, J. Nitrogen nutrition and water stress effects on leaf photosynthetic gas exchange and water use efficiency in winter wheat. Environ. Exp. Bot. 2000, 44,141-149. DOI:10.1016/S0098-8472(00)00064-2
268. Sassi-Aydi, S.; Aydi, S.; Abdelly, C. Inorganic nitrogen nutrition enhances osmotic stress tolerance in *Phaseolus vulgaris*: lessons from a drought-sensitive cultivar. HortScience 2014, 49, 550-555. https://doi.org/10.21273/HORTSCI.49.5.550

269. Zayed, O.; Hewedy, O.A.; Abdelmoteleb, A.; Ali, M.; Youssef, M.S.; Roumia, A.F.; Seymour, D.; Yuan, Z.-C. Nitrogen journey in plants: From uptake to metabolism, stress response, and microbe interaction. Biomolecules 2023, 13, 1443. https://doi.org/10.3390/ biom13101443
270. Khan, Y.; Shah, S.; Tian, H. The roles of arbuscular mycorrhizal fungi in influencing plant nutrients, photosynthesis, and metabolites of cereal crops- A review. Agronomy 2022, 12, 2191. https://doi.org/ 10.3390/agronomy12092191
271. Wang, R.; Zeng, J.; Chen, K.; Ding, Q.; Shen, Q.; Wang, M.; Guo, S. Nitrogen improves plant cooling capacity under increased environmental temperature. Plant Soil 2022a, 472, 329–344. https://doi.org/10.1007/s11104-021-05244-w
272. Tripathi, R.; Tewari, R.; Singh, K.P.; Keswani, C.; Minkina, T.; Srivastava, A.K.; De Corato, U.; Sansinenea. E. Plant mineral nutrition and disease resistance: A significant linkage for sustainable crop protection. Front. Plant Sci. 2022, 13, 883970. doi: 10.3389/fpls.2022.883970
273. Aigu, Y.; Daval, S.; Gazengel, K.; Marnet, N.; Lariagon, C.; Laperche, A.; Legeai, F.; Manzanares-Dauleux, M.J.; Gravot, A. Multi-omic investigation of low-nitrogen conditional resistance to clubroot reveals *Brassica napus* genes involved in nitrate assimilation. Front. Plant Sci. 2022, 13, 790563. doi: 10.3389/fpls.2022.790563
274. Reglinski, T.; Havis, N.; Rees, H.J.; de Jong, H. The practical role of induced resistance for crop protection. Phytopath. 2023, 113, 719-731. https://doi.org/10.1094/PHYTO-10-22-0400-IA
275. Sun, Y.; Wang, M.; Mur, L.A.J.; Shen, Q.; Guo, S. Unravelling the roles of nitrogen nutrition in plant disease defenses. Int. J. Mol. Sci. 2020, 21, 572. doi:10.3390/ijms21020572
276. Van de Waal, D.B.; White, L.A.; Everett, R.; Asik, L.; Borer, E.T.; Frenken, T.; González, A.L.; Paseka, R.; Seabloom, E.W.; Strauss, A.T.; Peace, A. Reconciling contrasting effects of nitrogen on host immunity and pathogen transmission using stoichiometric models. Ecology 2023, e4170. https://doi.org/10.1002/ecy.4170
277. Simone, G.W. Disease control in celery (*Apium graveolens* var. dulce). Univ. Florida Coop. Ext. Serv. PDMG-V3-36, Gainesville, Florida. 1999.
278. Howard, R.J.; Garland, J.A.; Seaman, W.L. Greenhouse tomato. Greenhouse tomato. In: Howard, R. J.; Garland, J. A.; & Seaman, W. L. (Eds). *Diseases and pests of vegetable crops in Canada: an illustrated compendium*. The Canadian Phytopathological Society, M.O.M. Printing Ltd., Ottawa, 1994, pp. 474-497.
279. FAO. Eggplant integrated pest management-an ecological guide. FAO Intercountry Programme for Integrated Pest Management in Vegetables in South and South East Asia. Rome. 2003, 182 pp.
280. Abuley, I.K.; Nielsen, B.J.; Hansen, H.H. The influence of timing the application of nitrogen fertilizer on early blight (*Alternaria solani*). Pest Manag. Sci. 2019, 75, 1150-1158. DOI 10.1002/ps.5236
281. Ries, S.M. Strawberry leaf diseases. Illinois Coop. Ext. Serv. RPD No. 702, November. University of Illinois at Urbana-Champaign, 1996, 6 pp.
282. Harrington, E.; Good, G. Crop Profile for Strawberries in New York. USDA, National IPM database. USDA-CSREES-Pest Management Alternatives Program. Washington D.C. 2000.
283. Guo, Z.; Luo, C.; Dong, Y.; Dong, K.; Zhu, J.; Ma, L. Effect of nitrogen regulation on the epidemic characteristics of intercropping faba bean rust disease primarily depends on the canopy microclimate and nitrogen nutrition. Field Crops Res. 2021, 274, 108339. https://doi.org/10.1016/j.fcr.2021.108339
284. Kucharek, T; Bartz, J. Bacterial soft rots of vegetables and agronomic crops. Univ. Florida Coop. Ext. Serv. PP-12, Gainesville, Florida, 1999.
285. Schwartz, H.F. Botrytis, downy mildew, and purple blotch of onion. Colorado State Univ. Coop. Ext. Serv. Crop series, Diseases no. 2.941. Fort Collins, CO., 2004.
286. Jansson, R.K.; Smilowitz, Z. Influence of nitrogen on population parameters of potato insects: abundance, population growth, and within-plant distribution of the green peach aphid, *Myzus persicae* (Homoptera: Aphididae). Environ. Entom. 1986, 15, 49-55, https://doi.org/10.1093/ee/15.1.49
287. Chen, Y.; Ruberson, J.R. Impact of variable nitrogen fertilisation on arthropods in cotton in Georgia, USA. Agric. Ecosyst. Environ. 2008, 126, 281-288. doi:10.1016/j.agee.2008.02.011
288. Jansson, R.K.; Leibee, G.L.; Sanchez, C.A.; Lecrone, S.H. Effects of nitrogen and foliar biomass on population parameters of cabbage insects. Entomol. Exp. Appl. 1991, 61, 7-16. https://doi.org/10.1111/j.1570-7458.1991.tb02390.x
289. Barroga, G.F.; Bernardo, E.N. Biology, feeding behavior and damage of the cotton leafhopper (*Amrasca biguttula* (Ishida)) on susceptible and resistant varieties of okra (*Abelmoschus esculentus* (L.) Moench.). Philipp. Entomol. 1993, 9, 186-200.
290. Leroux, E.J. 1952. The effect of various levels of nitrogen, phosphorus and potassium on the fecundity of the two-spotted spider mite *Tetranychus bimaculatus* Harvey. MS Thesis, McGill University, Montreal. 1952, 118 pp. https://escholarship.mcgill.ca/concern/theses/0k225f384
291. Alizade, M.; Hosseini, M.; Awal, M.M.; Goldani, M.; Hosseini, A. Effects of nitrogen fertilization on population growth of two-spotted spider mite. System. Applied Acarol. (SAA) 2016, 21(7), 947-956. http://www.bioone.org/doi/full/10.11158/saa.21.7.8
292. Hilje, L.; Costa, H.S.; Stansly, P.A. Cultural practices for managing Bemisia tabaci and associated viral diseases. Crop Prot. 2001, 20, 801-812. https://doi.org/10.1016/S0261-2194(01)00112-0

293. Perring, T.M.; Stansly, P.A.; Liu, T.X.; Smith, H.A.; Andreason, S.A. Whiteflies: Biology, ecology, and management. In: Wakil, W.; Brust, G.E.; Perring, T.M. (Eds.) *Sustainable management of arthropod pests of tomato*. Academic press, Elsevier. London. 2018, pp. 73-110. DOI: 10.1016/B978-0-12-802441-6.00004-8
294. Hu, X.F.; Cheng, C.; Luo, F.; Chang, Y.Y.; Teng, Q.; Men, D.Y.; Liu, L.; Yang, M.Y. Effects of different fertilization practices on the incidence of rice pests and diseases: A three-year case study in Shanghai, in subtropical southeastern China. Field Crops Res. 2016, 196, 33-50. http://dx.doi.org/10.1016/j.fcr.2016.06.004
295. Knezevic, S.Z.; Evans, S.P.; Blankenship, E.E.; Van Acker, R.C.; Lindquist, J.L. Critical period for weed control: The concept and data analysis. Weed Sci. 2002, 50, 773-786. https://doi.org/10.1614/0043-1745(2002)050[0773:CPFWCT]2.0.CO;2
296. Whitbread, A.M.; Robertson, M.J.; Carberry, P.S.; Dimes, J.P. How farming systems simulation can aid the development of more sustainable smallholder farming systems in southern Africa. Europ. J. Agron. 2010, 32, 51-58. doi:10.1016/j.eja.2009.05.004
297. Mekdad, A.A.; El-Enin, M.M.A.; Rady, M.M.; Hassan, F.A.; Ali, E.F.; Shaaban, A. Impact of level of nitrogen fertilization and critical period for weed control in peanut (*Arachis hypogaea* L.). Agronomy 2021, 11, 909. https://doi.org/10.3390/agronomy11050909
298. Laude, S. Competitiveness of tomato (*Lycopersicon esculentum* Mill.) with weeds at various nitrogen doses and weed free periods. In: *The 3rd International Conference On Environmental Ecology Of Food Security*, IOP Conference Series: Earth and Environmental Science, 2023, 1253 (1), 012030. IOP Publishing. Bristol, UK. doi:10.1088/1755-1315/1253/1/012030
299. Di Tomaso, J.M. Approaches for improving crop competitiveness through the manipulation of fertilization strategies. Weed Sci. 1995, 43, 491-497. doi:10.1017/S0043174500081522
300. Gholamhoseini, M.; AhaAlikhani, M.; Sanavy, S.M.; Mirlatifi, S.M. Interactions of irrigation, weed and nitrogen on corn yield, nitrogen use efficiency and nitrate leaching. Agric. Water Manag. 2013, 126, 9-18. http://dx.doi.org/10.1016/j.agwat.2013.05.002
301. Li, Y.; Zou, N.; Liang, X.; Zhou, X.; Guo, S.; Wang, Y.; Qin, X.; Tian, Y.; Lin, J. Effects of nitrogen input on soil bacterial community structure and soil nitrogen cycling in the rhizosphere soil of *Lycium barbarum* L. Front. Microbiol. 2023b, 13, 1070817. doi: 10.3389/fmicb.2022.1070817
302. Brennan, E.B.; Acosta-Martinez, V. Cover cropping frequency is the main driver of soil microbial changes during six years of organic vegetable production. Soil Biol. Biochem. 2017, 109, 188-204. http://dx.doi.org/10.1016/j.soilbio.2017.01.014
303. Pacovsky, R.S.; Fuller, G.; Stafford, A.E.; Paul, E.A. Nutrient and growth interactions in soybeans colonized with *Glomus fasciculatum* and *Rhizobium japonicum*. Plant Soil 1986, 92, 37-45. https://doi.org/10.1007/BF02372264
304. He, X.H.; Critchley, C.; Bledsoe, C. Nitrogen transfer within and between plants through common mycorrhizal networks (CMNs). CRC Crit. Rev. Plant Sci. 2003, 22, 531-567. http://dx.doi.org/10.1080/713608315
305. Loo, W.T.; Chua, K.-O.; Mazumdar, P.; Cheng, A.; Osman, N.; Harikrishna, J.A. Arbuscular mycorrhizal symbiosis: A strategy for mitigating the impacts of climate change on tropical legume crops. Plants 2022, 11, 2875. https:// doi.org/10.3390/plants11212875
306. Badr, M.A.; El-Tohamy, W.A.; Abou-Hussein, S.D.; Gruda, N.S. Deficit irrigation and arbuscular mycorrhiza as a water-saving strategy for eggplant production. Horticulturae 2020, 6 (3), 45. doi:10.3390/horticulturae6030045
307. Alori, E.T.; Dare, M.O.; Babalola, O.O. Microbial inoculants for soil quality and plant health. In: E. Lichtfouse (Ed.) *Sustainable agriculture reviews*, 22, Springer, Cham, Switzerland, 2017. Pp. 281-307. DOI 10.1007/978-3-319-48006-0_9
308. Adesemoye, A.O.; Torbert, H.A.; Kloepper, J.W. Enhanced plant nutrient use efficiency with PGPR and AMF in an integrated nutrient management system. Can. J. Microbiol. 2008, 54 (10), 876-886. doi:10.1139/W08-081
309. Ntsefong, G.N.; Toukam, G.M.S.; Mbi, K.T.; Getachew, S.E.; Yaouba, A.; Ali, S.M.; Kinge, T.R.; Teke, G.N. Unleashing the power of beneficial microorganisms for advancing crop improvement: An in-depth review. Preprints. 2023, 2 Oct. doi:10.20944/preprints202310.0050.v1
310. Canfora, L.; Costa, C.; Pallottino, F.; Mocali, S. Trends in soil microbial inoculants research: A science mapping approach to unravel strengths and weaknesses of their application. Agriculture 2021, 11, 158. https://doi.org/10.3390/ agriculture11020158
311. Scheu, S.; Schlitt, N.; Tiunov, A.V.; Newington, J.E.; Jones, H.T. Effects of the presence and community composition of earthworms on microbial community functioning. Oecologia 2002, 133, 254-260. DOI 10.1007/s00442-002-1023-4
312. Valenzuela, H. Earthworms in the farm. Hanai'Ai, The Food Provider, CTAHR Sustainable Agriculture Newsletter. Univ. Hawaii Coop. Ext. Serv. Honolulu, USA. Fall. 2010.
313. Fonte, S.J.; Hsieh, M.; Mueller, N.D. Earthworms contribute significantly to global food production. Nat. Commun. 2023, 14, 5713. https://doi.org/10.1038/s41467-023-41286-7

314. Van Groenigen, J.W.; Lubbers, I.M.; Vos, H.M.; Brown, G.G.; De Deyn, G.B.; Van Groenigen, K.J. Earthworms increase plant production: a meta-analysis. Sci. Rep. 2014, 4, 6365. DOI: 10.1038/srep06365
315. Li, H.; Wang, C.; Li, X.; Christie, P.; Dou, Z.; Zhang, J.; Xiang, D. Impact of the earthworm *Aporrectodea trapezoides* and the arbuscular mycorrhizal fungus *Glomus intraradices* on 15 N uptake by maize from wheat straw. Biol. Fertil. Soils 2013, 49, 263-271. DOI 10.1007/s00374-012-0716-z
316. Sanjuan-Delmás, D.; Josa, A.; Muñoz, P.; Gassó, S.; Rieradevall, J.; Gabarrell, X. Applying nutrient dynamics to adjust the nutrient-water balance in hydroponic crops. A case study with open hydroponic tomato crops from Barcelona. Sci. Hortic. 2020, 261, 108908.https://doi.org/10.1016/j.scienta.2019.108908
317. Rosa, L.; Gabrielli, P. Achieving net-zero emissions in agriculture: a review. Environ. Res. Lett. 2023, 18, 063002. https://doi.org/10.1088/1748-9326/acd5e8
318. Davidson, E.A.; Suddick, E.C.; Rice, C.W.; Prokopy, L.S. More food, low pollution (Mo Fo Lo Po): a grand challenge for the 21st century. J. Environ. Qual. 2015, 44, 305-311. doi:10.2134/jeq2015.02.0078
319. Reimer, A.; Doll, J.E.; Boring, T.J.; Zimnicki, T. Scaling up conservation agriculture: An exploration of challenges and opportunities through a stakeholder engagement process. J. Environ. Qual. 2022, 52, 465-475. DOI: 10.1002/jeq2.20317
320. Oelmann, Y.; Lange, M.; Leimer, S.; Roscher, C.; Aburto, F.; Alt, F.; Bange, N.; Berner, D.; Boch, S.; Boeddinghaus, R.S.; Buscot, F. Above-and belowground biodiversity jointly tighten the P cycle in agricultural grasslands. Nat. Commun. 2021, 12, 4431. https://doi.org/10.1038/s41467-021-24714-4
321. Stuart, D.; Shennan, C.; Brown, M. Food safety versus environmental protection on the Central California coast: Exploring the science behind an apparent conflict. Univ. of California at Santa Cruz, The Center for Agroecology & Sustainable Food Systems. Research Brief No. 10, Santa Cruz, California, 2006, 12 pp.
322. Zhu, Y.; Zhang, H.; Li, R.; Zhu, W.; Kang, Y. Nitrogen fertigation affects crop yield, nitrogen loss and gaseous emissions: a meta-analysis. Nutr. Cycl. Agroecosyst. 2023, 1-15. https://doi.org/10.1007/s10705-023-10318-5
323. Nouri, A.; Lukas, S.; Singh, S.; Singh, S.; Machado, S. When do cover crops reduce nitrate leaching? A global meta-analysis. Glob. Chang. Biol. 2022, 28, 4736-4749. DOI: 10.1111/gcb.16269