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Article

Follicular Fluid Proteomic Analysis of Women Undergoing Assisted Reproduction Suggests Apolipoprotein A1 to Be a Potential Fertility Marker

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Abstract: Infertility affects millions worldwide, posing a significant global health challenge. The proteomic analysis of follicular fluid provides a comprehensive view of the complex molecular landscape within ovarian follicles, offering valuable information on the factors influencing oocyte development and on the overall reproductive health. The follicular fluid is derived from the plasma and contains various proteins that can have different roles in oocyte health and infertility and this fluid is a critical microenvironment for the developing oocytes as well. Using HPLC/MS method we investigated the protein composition of the follicular fluid and after classification we did relative quantification of the identified proteins in the pregnant (P) and non-pregnant (NP) groups. Based on the protein-protein interaction analysis ALB and ApoA1 was found to be hub proteins and the quantitative comparison of the P and NP groups resulted significantly lower concentration of ApoA1 and HDL-cholesterol in the P group. As both molecules are involved in the cholesterol transport, we also investigated their role in the development of oocytes and in the prediction of fertility.

Keywords: assisted reproductive treatment; follicular fluid; proteomics; mass spectrometry; apolipoprotein A1; HDL cholesterol; predictive value

1. Introduction

Infertility is described as a condition where a clinical pregnancy cannot be achieved after 12 months of regular, unprotected sexual intercourse. According to the WHO definition, infertility is a disease that causes disability by impairing normal reproductive functions [1]. Infertility affects 8-12% of reproductive-aged couples worldwide [2]. Since infertility affects millions of individuals, understanding the factors contributing to this disease is of paramount importance in guiding effective fertility treatments and interventions. Follicular fluid is a crucial component of the female reproductive system therefore, its analysis can provide valuable information on the microenvironment where the oocyte development occurs. By studying the composition of the follicular fluid, it is possible to find key biomarkers and factors that influence fertility offering a hope for improved diagnostic methods and personalized therapies for individuals struggling with infertility [3].

Despite the increasing use of assisted reproduction technologies (ART) during the last decades, the effectiveness of these procedures remains still under the desired level. To help pinpointing several proteins that could serve as potential biomarkers or treatment targets for achieving successful ART, pregnancy related proteomics has been extensively used [4]. Proteomics is a type of 'omics' focusing on the examination of the interactions, functions, composition and structures of proteins and their cellular activities [5]. The progress in mass spectral (MS) profiling has empowered the identification

of potential biomarkers and treatment targets in biological fluids linked to human reproduction. This advancement offers crucial information on different stages of the ART process and raises hopes for successful outcomes in high-risk ART pregnancies [4].

The follicular fluid (FF) serves as a natural environment for the oocyte, encompassing a complex network of proteins that undergoes dynamic changes. These proteins play an important role in the oocyte’s growth and development, significantly influencing its quality and potential for fertilization. Fortunately, FF is readily accessible as it is abundantly collected during ART procedures at the oocyte retrieval stage. It has been demonstrated to be a valuable substrate for proteomic analysis, facilitating the exploration of its precise composition and functions [4]. FF is a complex biological fluid found within the follicular antrum and provides a microenvironment for the developing oocytes and contains hormones, enzymes, anticoagulants, reactive oxygen species and antioxidants. It also plays a crucial role in facilitating communication among cells within the antral follicle and serves as a vital medium delivering nutrients to the oocyte [6]. The composition of follicular fluid is derived partially from the plasma by diffusion and transudation and, the wall of the follicle synthesizes metabolites that are going to be altered by granulosa and theca cells. This unique biological matrix is the only one directly linked to the oocyte, serving as the microenvironment for its growth and differentiation in vivo [7]. FF, like human serum, contains a wide array of proteins, each presented at varying concentrations [8]. The primary role of these proteins is to modulate oocyte and follicular maturation [9] and to protect the developing oocytes and other follicular cells from physical or oxidative damage. They play a crucial role in facilitating communication between somatic and germ cells, which is essential for the oocyte to acquire its competence [10]. Any changes or any disruptions in the follicular fluid microenvironment can result in altered protein function with a consequence of altered viability of the oocytes thus causing adverse pregnancy outcomes [11]. A better understanding how different molecular changes may interfere with fertility and pregnancy rates in assisted reproduction technologies would be of utmost importance and may provide new information for the future clinical practice [12].

In the present work, we planned to investigate the protein composition of the FF because proteomic analysis of FF provides a good tool to gather information on the follicular development and on the oocyte quality as well [13] providing a huge impact on the success of the ART. After identification of the FF proteins, we performed an analysis on their protein-protein interactions. Then the most potent proteins were quantified, and the literature surveyed to support the role of our best candidates and their potential significance on the quality of oocytes and on fertility as well.

2. Results

2.1. Demographic and Clinical Data of Patients

In Table 1, the demographic and clinical features of the patients enrolled in our study are shown.

Table 1. Clinical characteristics of the patients involved in our research.

	Pregnant [Mean± SD] (n=15)	Non-pregnant [Mean± SD] (n=15)
Age	36.31±5.51	32.93±4.94
BMI	26.31±5.84	25.92±5.65
Number of oocytes retrieved	12.27±7.54	11.85±8.4
Number of fertilized oocytes	4.86±3.68	3.15±2.69
Number of IVF cycles	1.89±0.9	2.07±0.75

2.2. Classification of the Proteins in the Follicular Fluids

A total of 131 individual proteins were identified in the FF samples. These proteins can be classified based on their role in biological processes, their biological classes, and according to the pathways they are involved in. The classification is illustrated in Figure 1. Based on the biological processes the identified proteins were classified as follows: 1/ *cellular processes* (20.7%), 2/ *metabolic processes* (16.7%) and 3/ *biological regulation* (14.9%). Other features such as *response to stimulus*, *localization*, *immune system processes*, *multicellular organism related processes*, *signaling* and *growth pathways* were also suggested. The proteins of the follicular fluid can be differentiated into different protein classes. The three biggest classes were the *defense/immunity proteins* (32.4%), the *protein binding activity modulator proteins* (27.9%), and the *transfer/carrier proteins* (16.2%). *Protein modifying enzyme proteins*, *metabolite interconversion enzyme proteins*, *transmembrane signal receptor proteins*, and *intercellular signal molecules* classes were classified in lower percentages. Based on the pathways the identified proteins were involved in, two main classes were distinguished. Pathways related to *blood coagulation* (44.4%) and the *plasminogen activating cascade* were the most important ones but others such as *B cell activation*, *angiogenesis*, *integrin signaling pathways*, pathways related to *vitamin D metabolism* and *serine-glycine biosynthesis* were also presented.

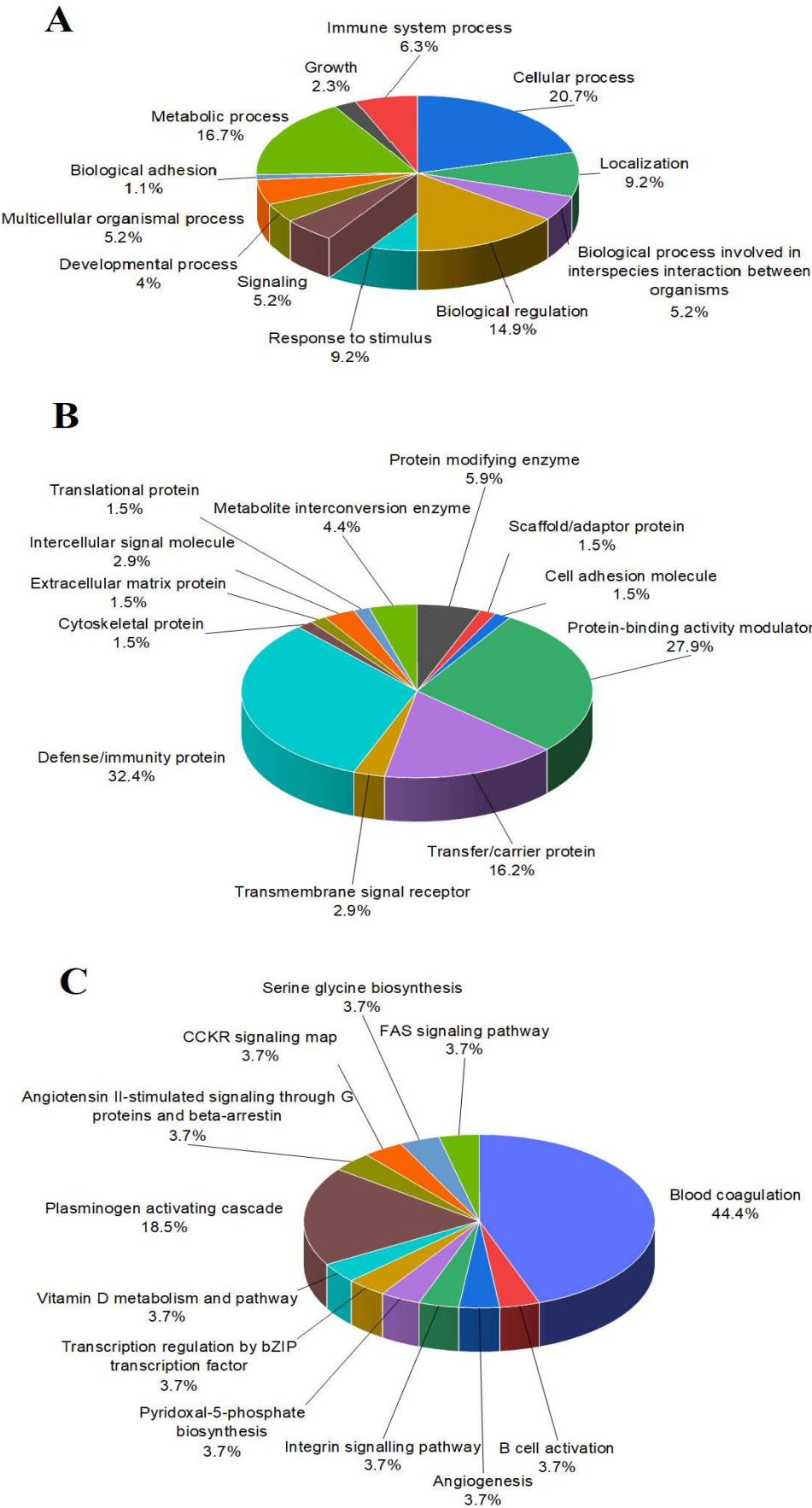


Figure 1. Gene ontology analysis of the proteins identified in human follicular fluid. Proteins were classified according to A) biological processes, B) protein classes, and C) pathways involved. Results are displayed as percentage of proteins classified to a category over the total number of class hits.

2.3. Relative Quantification of the Identified Proteins

To evaluate our results more precisely, relative quantification was used to compare the intensities of the MS signals in the pregnant (P, n=15) and non-pregnant (NP, n=15) groups. 77 proteins were found to have different levels in the two groups. Combining intensity values with protein-protein interaction and functional analysis can reveal the roles of proteins in specific biological functions or pathways. Proteins with both string interactions (altered intensity values) and relevant functions can be particularly interesting.

2.4. Protein-Protein Interactions (PPIs)

The purpose of protein-protein interaction analysis was to monitor the physical connection between identified proteins and their interactions to understand how they interact with each other and carry out various biological processes. These interactions are crucial for the functioning of biological pathways, signal transduction and the regulation of gene expression. PPI analysis can help to uncover the complex network of interactions of cellular functions and can provide insights into disease mechanisms. By adding intensity values to the general PPIs results, it can provide additional quantitative information on the interactions so for each protein P/NP intensity value ratio was calculated and illustrated as a full STRING network. The results of the PPI analysis can be seen in Figure 2. The explanation of the abbreviations is listed in Supplementary Table 1.

In Figure 2 a full STRING network of our follicular fluid samples is shown. In this analysis every node represents a protein and edges in the PPI network illustrate the interactions between the proteins. These interactions can be physical (such as direct binding) or functional indicating proteins that cooperate in the same biological process. The layout of the nodes is also an important factor when analyzing the results. The arrangement of the nodes provides information on the clusters or groups of interacting proteins. Nodes close together often have more interactions among them. Based on the P/NP intensity value ratio calculation there were some proteins that had value above 1 these were ALB, APOL1, A2M, APOC3, FGA, FGB, TTR, SERPIND1, GSN and ITIH1 meaning they had higher intensity values in the P group. SERPINC1, AGT, ApoA1, AMBP, ORM1, HPX, SHBG, LBP and ITIH2 had lower ratio values meaning the intensity value of these proteins was higher in the NP group.

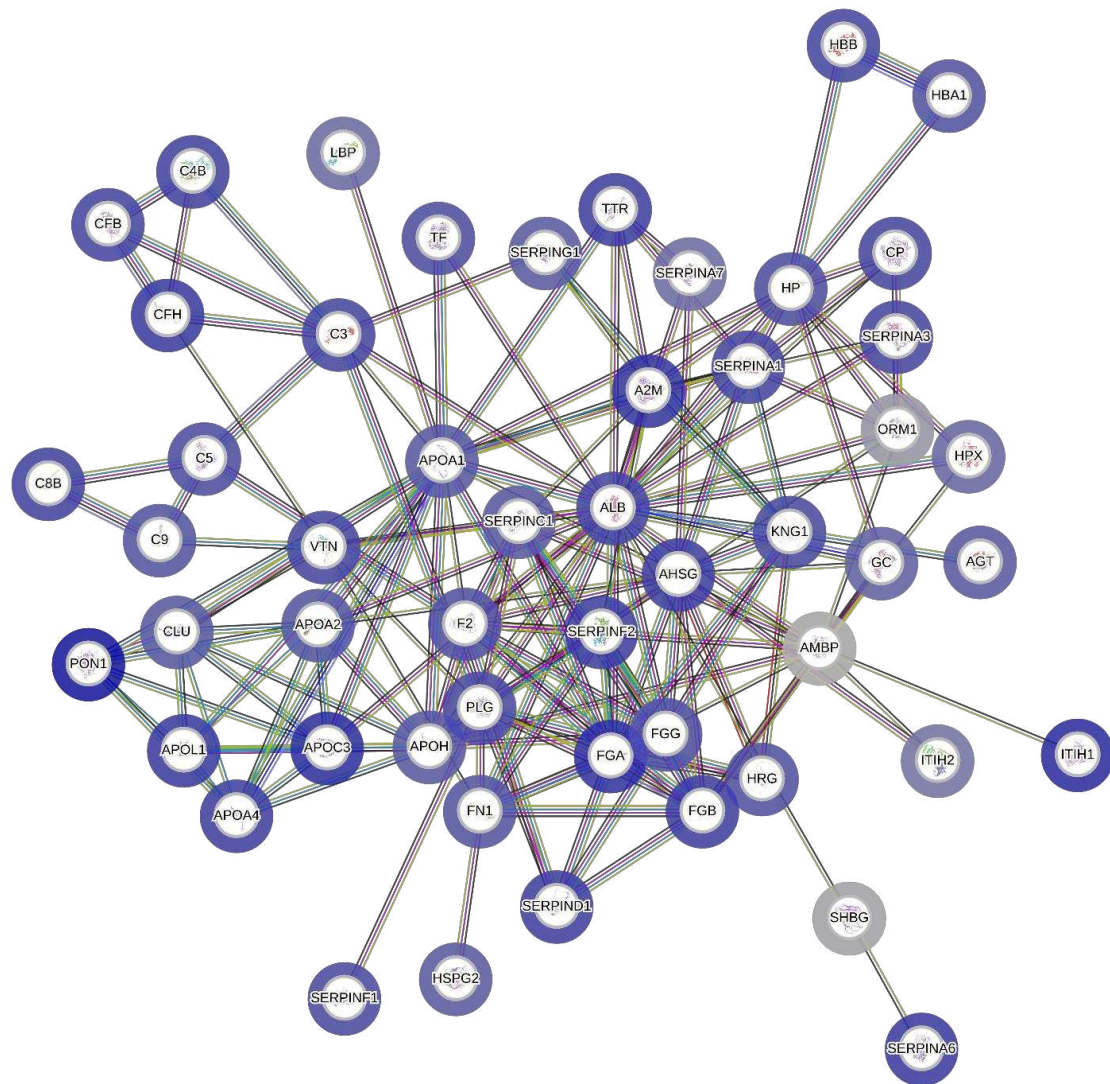


Figure 2. Protein-protein interaction network (full STRING network) of the identified proteins. The color of the lines illustrates the type of interaction evidence (e.g., turquoise=known interactions from curated databases, purple=experimentally determined, predicted interactions based on green=gene neighborhood, red=gene fusions, dark blue=gene co-occurrence, other associations such as yellow=text mining, black=co-expression and light blue=protein homology). Dark blue color of the halo represents that the intensity values of the certain protein is higher than 1 P/NP ratio value and light blue color represents lower P/NP ratio values than 1. For the minimum required interaction score 0.9 was set which provides the highest confidence.

Proteins with many connections are called hub proteins, being quite often crucial in the network. These hub proteins play important roles in biological systems and are usually involved in multiple interactions, making them crucial for maintaining the network's structure and function and these proteins may be targeted for further investigations as they are likely to be involved in critical biological functions. Node degree is the number of connections or interactions that a certain protein has with other proteins in the network. In Table 2. the node degree of proteins in our network is shown. In our study ALB had high node degree meaning that this protein interacted with 27 other proteins making it a hub protein. Another hub protein was ApoA1 that interacted with 18 other proteins.

Table 2. The node degree of proteins in our network. Proteins only with node degree over 10 are shown.

Name of node (protein)	Node degree
ALB	27
ApoA1	18
AHSG	17
FGA	14
APOH	13
PLG	13
AMBP	12
F2	12
FGB	11
FGG	11
HP	10
A2M	10
SERPINC1	10

2.5. Functional Enrichment Analysis

Following the PPI analysis, the nodes were associated with biological functions and pathways through enrichment analysis. Functional enrichment analysis is a commonly used method to identify trends in datasets and to predict functions and the involved pathways of the proteins of interest. The result of our analysis is shown in Table 3. The *Count in network* numbers show how many proteins in our network are annotated with a particular term and the second number indicates how many proteins in total have this term assigned. The significance of the enrichment results was represented in two important statistical measures namely *Strength* and *False Discovery Rate* (FDR). *Strength* quantifies how much a certain functional term or pathway is overrepresented or enriched in the dataset. Higher value indicates a stronger enrichment signal, and it suggests that proteins associated with that term are significantly overrepresented in the dataset. This value can be used to rank the enriched terms. *FDR* is a statistical value, and it estimates the proportion of false positives among the statistically significant results. The threshold is typically set to 0.05 and it determines which enriched terms or pathways are significant after multiple testing corrections. Values below 0.05 are considered as significant.

In our dataset the most significant terms were High-density lipoprotein (HDL) receptor binding, Phosphatidylcholine-sterol O-acyltransferase activator activity (Lecithin Cholesterol Acyltransferase [LCAT] activator activity) and Apolipoprotein receptor binding. These terms had the highest Strength score, and their FDR values were also below 0.05 suggesting a high level of confidence in the association between these proteins and the term presented.

Table 3. Results of the functional enrichment analysis based on intensity values of the pregnant and non-pregnant patient groups. The table represents proteins that are over-represented in a large set of proteins.

Term description (Function)	Count in network	Strength	False discovery rate
High-density lipoprotein particle receptor binding	3 of 3	2.5	0.00018
Phosphatidylcholine-sterol O-acyltransferase activator activity	3 of 6	2.2	0.00067
Apolipoprotein receptor binding	2 of 6	2.02	0.0426

Lipoprotein particle receptor binding	5 of 30	1.72	2.56E-05
Serine-type endopeptidase inhibitor activity	16 of 98	1.71	9.09E-20
Cholesterol transfer activity	3 of 23	1.62	0.0156
Endopeptidase inhibitor activity	21 of 177	1.58	2.24E-23
Complement binding	3 of 26	1.56	0.0185
Phosphatidylcholine binding	3 of 29	1.52	0.0233
Peptidase regulator activity	22 of 227	1.49	2.28E-23
Proteoglycan binding	3 of 37	1.41	0.0426
Cholesterol binding	4 of 52	1.39	0.0067
Steroid binding	7 of 101	1.34	1.62E-05
Enzyme inhibitor activity	23 of 396	1.27	4.86E-20
Chaperone binding	6 of 108	1.25	0.00044
Molecular carrier activity	4 of 72	1.25	0.0181
Glycosaminoglycan binding	13 of 245	1.23	6.10E-10
Heparin binding	9 of 173	1.22	2.34E-06
Antioxidant activity	4 of 76	1.22	0.0205
Extracellular matrix structural constituent	6 of 131	1.16	0.0012
Sulfur compound binding	10 of 272	1.07	6.86E-06
Protease binding	5 of 135	1.07	0.016
Enzyme regulator activity	28 of 1239	0.86	8.74E-15
Lipid binding	15 of 796	0.78	8.08E-06
Molecular function regulator activity	30 of 1960	0.69	1.11E-11
Signaling receptor binding	23 of 1499	0.69	3.28E-08
Protein binding	40 of 7242	0.24	0.0022

3.6. Quantitative and Statistical Analysis of the Selected Proteins

Based on the PPI and enrichment analysis we concluded that there are several proteins that should be further quantified. These proteins were ALB and ApoA1. ALB was confirmed to be a central protein in the FF based on the PPI. ApoA1 was also found to be a central protein and the functional enrichment analysis has proven that it is indeed a potentially interesting molecule in the FF. Since ApoA1 is a major component of the HDL molecule the concentration of HDL cholesterol (HDL-C) of the follicular fluid was also determined. Haptoglobin (HP) was also investigated because

it was found to be associated with ApoA1 in FF. Previous research indicated that HP has a significant influence on fertility [14]. Another study showed that HP in human follicular fluid inhibits the reverse transport of cholesterol to the bloodstream by blocking the apolipoprotein A1-induced activation of LCAT [15]. The results of the quantitative measurements are shown in Table 4.

Table 4. Results of the quantitative measurements of the selected proteins.

	Pregnant [Mean± SD] (n=15)	Non-pregnant [Mean± SD] (n=15)
Total protein (g/l)	45.37±6.21	47.61±11.14
Albumin (g/l)	32.72±4.25	34.43±8.05
ApoA1 (mg/dl)	123.54±16.51	139.96±22.27
HP (g/l)	0.55±0.373	0.43±0.24
HP/ApoA1 ratio	0.46±0.29	0.32±0.26
HDL-C (mmol/l)	0.55±0.24	0.78±0.23
HDL/ApoA1 ratio	0.17±0.07	0.21±0.04

2.6.1. Comparison of the Protein Concentrations Based on the Outcome

Based on the outcome two groups were separated and pregnant (n=15) and non-pregnant (n=15) groups were determined. There were no significant differences in the concentration of TP and ALB concentration of the P and NP groups. In the P group the concentration of the ApoA1 and HDL-C was lower and p values were 0.007 and 0.010 respectively, showing significant differences between the P and NP groups. The HDL/ApoA1 ratio was also significantly altered between the two groups (p=0.045). The concentration of HP and the HP/ApoA1 ratio did not show any significant differences (Tables 4. and 5.)

2.6.2. Comparison of Protein Concentrations Based on the Age of the Patients

In comparison, two age groups were determined. One group (younger, n=12) contained adult patients aged equal to or under 34 and the members of the other group were 35 years old or older. In this comparison none of the proteins showed significantly altered levels (Table 5).

2.6.3. Comparison of the Protein Concentrations Based on the BMI

The patients were divided into two groups according to their BMI. Patients with BMI values between 18.5 and 24.9 were considered to belong to the normal group (n=13), while in the overweighted group the BMI score was above 25 (n=16). There were no significant differences between the normal and overweight groups. (Table 5.)

Table 5. Results of the statistical analysis of measured FF proteins. The significance level was set as p < 0.05. Data significantly influencing the pregnancy outcome are highlighted in bold.

Protein name	Parameter	p value
TP	Outcome	0.595
ALB	Outcome	0.567
HP	Outcome	0.367
ApoA1	Outcome	0.007
HDL-C	Outcome	0.010
HDL-C/ApoA1 ratio	Outcome	0.045
HP/ApoA1 ratio	Outcome	0.148
TP	Age	0.232
ALB	Age	0.267

HP	Age	0.439
ApoA1	Age	0.325
HDL	Age	0.325
TP	BMI	0.846
ALB	BMI	0.559
HP	BMI	0.449
ApoA1	BMI	0.268
HDL	BMI	0.503

3. Discussion

In this article we investigated the composition of the FF and characterized its content based on PPI and enrichment network analysis. In many proteomic studies during the sample preparation the most abundant proteins such as albumin, different subclasses of immunoglobulins, haptoglobin, transferrin, fibrinogen and ApoA1 are removed because their signals tend to obscure the signal of the less abundant proteins. Their high abundance interferes with the detection of other proteins and can prevent accurate detection. By depleting them the quality and depth of the analysis can be greatly enhanced but unfortunately removing these components may result in the loss of precise detection of other follicular fluid proteins with much lower concentration. Therefore, we did not remove the abundant proteins to save all the protein contents of the FF. After analyzation of the follicular fluid, we concluded that there are several proteins that can be involved in the lipid metabolism and transport in the FF. Quantitative measurements were made to determine the concentrations of those proteins that may have important role in the fertility and in the outcome of ART. To investigate the effects of the age and the BMI of the patients on the protein pattern of the FF, statistical analysis was performed. ApoA1 showed a significantly lower concentration in the FF of the pregnant patients compared with the NP individuals and the same was found for HDL-C as well. Both molecules are involved in the transport of cholesterol that is a vital molecule in early mammalian development. It is essential for membrane fluidity making it a crucial component for the synthesis of cell membranes in rapidly dividing cells [16]. The metabolism of cholesterol is an important aspect of producing good quality embryos supported by the finding that ApoA1 is produced in the embryo because of the rapid cell division and the intense membrane synthesis [17]. Cholesterol serves as a precursor for steroid hormones, playing a pivotal role in the maturation of ovarian follicles [18]. In the follicular stage the thecal cells absorb cholesterol from blood HDL and produce androgens (androstenedione and testosterone). These androgens cross the basal membrane and stimulate estrogen production by granulosa cells within the developing follicle. Following the LH surge, the follicular basal membrane becomes more permeable to larger lipoproteins, increasing cholesterol availability to support the active production during ovulation [19]. Following ovulation, the follicle becomes vascularized, and the follicular cells utilize stored cholesterol esters while increasing the expression of lipoprotein receptors. In the corpus luteum the luteal cells employ cholesterol from both de novo biosynthesis and plasma LDL and HDL to significantly boost progesterone synthesis [20].

In our research we compared the concentration of ApoA1 in the FF of the pregnant and the non-pregnant patient groups, and we observed that the concentration of ApoA1 was significantly lower in the pregnant group. ApoA1 is a protein that is part of the lipoprotein class. It is a crucial protein for cholesterol metabolism, and it also helps in the synthesis of HDL molecules. ApoA1 makes up 70% of the HDL molecule being produced in the liver (80%) and in the gut (20%) as well [21]. Some studies reported that ApoA1 has a significant role in female reproduction and in the positive outcome of pregnancies [22]. Patients with different diseases were examined and it was revealed that there is an altered expression of ApoA1 in preeclampsia, endometriosis, PCOS and repeated implantation failure. A recent study identified elevated ApoA1 levels in women who experienced spontaneous preterm delivery [23]. To achieve a successful pregnancy, multiple other factors should be taken into consideration. There were several reports suggesting the association of ApoA1 with the quality of the embryos, their development, and viability [24]. Several past studies have linked the levels of ApoA1 to embryo quality in IVF settings associated with the success of IVF trials [25]. Regarding the

expression of the ApoA1 protein in the culture medium of day 2 to 3 embryos, it was determined that ApoA1 was less abundant in the samples resulting in pregnancy [26]. Also, based on previous research it was observed that there is a connection between successful embryo transfer and low levels of ApoA1 in the early culture medium suggesting that the internalization of ApoA1 might be a characteristic feature of viable embryos. This idea was supported by the presence of ApoA1 mRNA transcripts in day 5 blastocysts but not in day 3 embryos indicating higher ApoA1 concentrations inside the blastocysts [17]. A recent study on bovine embryos revealed a decrease in proteins related to lipid metabolism, including a 1.6-fold reduction in ApoA1 abundance from the oocyte stage to the 4-cell stage. This decline could be attributed to an increased secretion ability of the embryo into the culture medium during development [27].

Other factors such as endometrial receptivity are important to achieve successful pregnancy. Endometrial receptivity refers to the capability of the uterine lining to receive and accommodate an implanting embryo leading to a successful pregnancy. The successful implantation process relies on the precise coordination of protein networks that play a crucial role in facilitating communication between the developing embryo and the endometrial lining [22]. It was reported that ApoA1 plays a role in endometrial receptivity, and it was recognized as a potential anti-implantation factor released by the differentiating endometrium. ApoA1 potentially plays a role in influencing the receptivity of the endometrium. In a recent study comparing endometrial biopsies between facing recurrent implantation failure and fertile controls, it was discovered that the levels of ApoA1 were 4.2 times higher in women experiencing recurrent failure. The ApoA1 expression in the endometrium of infertile women was lower during the midsecretory phase compared to the proliferative phase and the hormone hCG was found to strongly inhibit ApoA1 expression in the human endometrium. Additionally, this hCG-mediated regulation of ApoA1 appeared to be disrupted in patients with pelvic endometriosis. This implies that if ApoA1 acts as a signal from the implanting embryo, reducing ApoA1 levels in the endometrium could potentially enhance its responsiveness to this signal by minimizing background ApoA1 interference [28]. Other investigations confirmed that patients with PCOS seem to have elevated levels of ApoA1 suggesting ApoA1 as a biomarker for nonreceptive endometrium [29].

The other protein that was present in lower concentration in the pregnant group was HDL-C. HDL is the smallest and most compact class of lipoproteins [30]. In the FF, which is only permeable to proteins with a molecular weight up to 300 kDa, HDL is the sole lipoprotein that can be detected there [31]. The HDL in the FF is primarily sourced from the plasma and compared to the plasma it varies in both size and composition having lower cholesterol content and higher levels of phospholipids [32]. HDL-C may play a crucial role in fertility because it is a main class of lipoproteins present in significant quantities in the FF. It is believed that follicular HDL may transport essential lipids to follicular cells or facilitate the removal of cholesterol from these cells. A key process for delivering HDL-C to ovarian cells, especially those involved in steroid production, is known as selective lipid uptake and it is facilitated by the HDL receptor SR-B1. Vital knowledge about the connection between HDL metabolism and female fertility has been gained from studies involving SR-B1 knock-out (KO) mice. These mice have high levels of cholesterol-rich HDL in their bloodstream leading to female infertility. Interestingly, this infertility can be reversed by administering cholesterol-reducing medication. Oocytes released during ovulation in SR-B1 KO females display dysfunction and accumulate excessive cholesterol. However, the precise mechanisms by which the FF HDL contributes to maintaining oocytes' cholesterol balance are not yet understood [33].

In a review it was suggested that FF HDL plays a role in maintaining balanced cholesterol levels in developing oocytes and eggs by removing excess cholesterol from the plasma membrane. If there is a disruption in the reverse cholesterol transport process caused by abnormal FF HDL, there is an accumulation of unesterified cholesterol in the egg. This imbalance can interfere with crucial processes such as oocyte maturation, meiosis arrest and egg activation following fertilization. Consequently, disruptions in HDL-C metabolism could impair FF HDL function leading to infertility due to dysfunctional and unstable eggs [34].

Because of its anti-inflammatory and antioxidant properties [35], as well as its presence in the FF that directly surrounds the developing oocyte, HDL is believed to have a significant role in fostering oocyte competence and enhancing embryo quality [36]. In a recent study it was found that FF HDL exhibited positive anti-inflammatory properties, correlating with the developmental potential of the oocyte. It was also reported that there is a negative correlation between the concentration and particle size of HDL-C in FF and the outcomes of IVF, as well as parameters related to the quality of preimplantation embryos such as fragmentation and symmetry [34]. It was demonstrated that HDL molecules have a significant role in the developmental potential of the oocytes during early embryogenesis. HDL but also ApoA1 levels in the follicular fluid show a negative association with embryo fragmentation. This implies that the oocyte relies on HDL even before fertilization and embryonic development [37].

The outcome of ART can be unfavorably affected by a woman's increasing age. The declines in fertility - generally the likelihood of a successful pregnancy - among individuals begin at around the age from 31 to 35. By the age of 40, at least half of the patients may experience infertility. The exact reasons behind the women's age-related decline in fertility are not fully understood. However, several factors may contribute to this phenomenon, including a reduction in ovarian reserve, poorer quality of oocytes, lower rates of successful implantation, ovulatory issues stemming from changes in hormonal patterns and potential uterine problems [38]. Previous studies have also shown that the composition of follicular fluid changes with age, and these changes could be associated with the quality of eggs [11]. Both age and obesity are linked to systemic oxidative stress and in this physiological context, elevated levels of specific lipids could potentially trigger harmful processes that negatively impact reproductive function [39]. It was previously presented that the higher BMI can be associated with fertility problems and adverse pregnancy outcomes [40]. Also, higher BMI and obesity are associated with changes in the composition of the FF including alterations in its lipid profile. Increased levels of triglycerides, cholesterol and inflammatory markers in FF have been observed in obese individuals [41]. The BMI-dependent concentration of HDL-C in the serum was significantly associated with those of the FF concentrations and this suggests that changes in serum cholesterol levels could influence the quality of the oocytes. Additionally, the article implied that the number of HDL particles in FF might adversely affect the developmental capacity of the oocyte, even if fertilization occurred normally. Notably, there's a proposition that the combined impact of FF HDL-C and ApoA1 has a protective role in oocyte health and embryo development by reducing embryo fragmentation [42]. Interestingly, our results did not show any significant alteration between the follicular fluid ApoA1 and HDL-C concentration of the younger/older groups and normal weight/obese groups.

The field of reproductive biology is quite complex and is under continuous exploration and discovery. Despite the interactions occurring within the ovarian microenvironment, there is a notable gap in our understanding of the potential influence of HDL and its major component, ApoA1 on the development, health of the oocyte and the pregnancy outcomes. Surprisingly, scientific literature is relatively scarce in providing comprehensive information on the role of HDL and ApoA1 in these fundamental biological processes. This paucity of research is particularly striking given the critical functions that can be attributed to HDL and ApoA1 in lipid metabolism, inflammation modulation and antioxidant defense. These two molecules have been extensively studied in the context of cardiovascular health where their roles in cholesterol transport and cardiovascular risk reduction is well-documented. However, their potential impact on the female reproductive health, especially concerning oocyte maturation, fertilization and pregnancy outcomes remains an unexplored territory. Understanding the interplay between the HDL, ApoA1 and the ovarian microenvironment has importance not only for advancing our knowledge of reproductive biology but also for unrevealing potential therapeutic avenues to enhance fertility treatments and to improve pregnancy success rates. This knowledge gap could start a new research field that can contribute to the development of more effective interventions and personalized strategies for couples struggling with infertility.

When fully validated on a much larger sample size the proteins analyzed in our study may prove to be clinically relevant for predicting the outcome of the ART. Protein, lipid, and lipoprotein analysis, which are commonly available in clinical laboratories could offer timely information before and during IVF treatment. We would like to emphasize that molecular studies – especially proteomics - of FF in relationship with ART are still few which makes interpretation of the results a bit difficult.

4. Materials and Methods

4.1. Patient Enrollment

The study was conducted between May 2021 and March 2023 at the Department of Obstetrics and Gynecology (FF sampling), and at the National Laboratory on Human Reproduction (analytical studies), University of Pécs, Hungary. Detailed information was given to all patients or their next-of-kin regarding our study protocol while written consent was obtained from all. Exclusion criteria were patients under 18 years of age, unobtainable or withdrawn consent and autoimmune diseases. Patients with endometriosis or polycystic ovary syndrome (PCOS) were not included in this study. The study protocol was approved by the Regional Research Ethics Committee of the University of Pécs (no. 5273-2/2012/EHR) conforming to the 7th revision of the Helsinki Declarations (2013). All the 30 patients enrolled in this study received assisted reproductive treatment (ART). The inclusion criteria were either male infertility or female infertility caused by tubal problems. The obtained patient data were based on the medical records of the hospital informational system.

4.2. Follicular Fluid Sample Collection

The GnRh agonist triptorelin was used in both long and short protocols and cetrorelix was applied in the antagonist protocols. Depending on the maturity of the follicles individual doses of rFSH ranging from 150 to 250 IU per day were used for stimulation. The starting dose was determined based on BMI and age. A maximum daily dose of 300 IU was given to those with a previously determined low response. The stimulation was supplemented with rLH or human menopausal gonadotropin (hMG) individually, according to the patient's age or response. From the 6th day of the cycle the follicular maturity was monitored by ultrasound every consecutive day. Gonadotropin was administered individually according to the size of the follicles. When at least two follicles exceeded 17 mm in diameter, 250 µg (6500 IU) of recombinant human chorionic gonadotropin was given parenterally to induce final oocyte maturation. Aspiration was performed 36 hours later using an ultrasound-guided transvaginal puncture under routine intravenous sedation. The follicular fluid collection was performed during oocyte retrieval procedure and the samples were centrifuged immediately at 6700 g for 10 minutes at room temperature to remove the erythrocytes and white blood cells. The supernatant was collected and stored at -80°C for further analyses.

4.3. Chemicals

To avoid protein loss during the sample preparation Protein LoBind Eppendorf tubes (Eppendorf AG, Hamburg, Germany) were used. For the protein separation ice-cold chloroform/methanol solution was applied (50/50, v/v, Merck Life Science, Darmstadt, Germany). After precipitation of the proteins, rehydration buffer (6 M urea, Bio-Rad Laboratories, Hercules, CA, USA; 100 mM TRIS, Bio-Rad Laboratories, Hercules, CA, USA) at pH 7.8 was used. For the determination of the protein concentration micro-BCA (bicinchoninic acid) assay (ThermoFisher Scientific, Darmstadt, Germany) was utilized. The overnight enzymatic digestion was performed by Trypsin/Lys-c enzyme mixture (Promega Corporation, Madison, USA). For sonication Sonics & Materials, Inc VCX 130 ultrasonic homogenizer (53 Church Hill Road, Newtown, CT, USA) was applied. Purification of the digested peptides was performed by an Oasis HLB 96-SPE well-plate (WAT058951; Waters, Milford, USA) instrument. The eluted samples were concentrated with an Eppendorf Concentrator Plus system (Eppendorf AG, Hamburg, Germany). The purified peptides were redissolved in 100 µL of water containing 0.1% formic acid (Merck Life Science, Darmstadt,

Germany). During the HPLC separation Solvent A and Solvent B were used. Solvent A consisted of water/formic acid (99.9/0.1, v/v), (Merck Life Science, Darmstadt, Germany) while solvent B was composed of water/formic acid/acetonitrile (4.9/0.1/95, v/v/v), (Merck Life Science, Darmstadt, Germany).

4.3.1. Sample Preparation for HPLC/MS Measurement

The thawed samples (300 μ L) were transferred into Protein LoBind Eppendorf tubes and 1000 μ L of ice-cold chloroform/methanol solution was added to each sample. After vortexing, the samples were centrifuged for 10 min at 10,000 \times g at 4°C, and the separated protein pellets were collected into new tubes. The precipitated proteins were dried for 10 min using the Eppendorf Concentrator Plus system and after that the proteins were resuspended in a rehydration buffer supported by ultrasonication. Samples were individually sonicated on ice for 1 minute using a 20% power setting on the micro-tip probe and a 50% pulse. The total protein concentrations were measured with a micro-BCA assay and all samples were normalized to a protein mass of 150 μ g before proceeding with the enzymatic digestion. The overnight in-solution enzymatic digestion was performed with a Trypsin/Lys-C enzyme mixture. Before digestion the protein extracts were reduced and alkylated according to the manufacturer's technical bulletin (V5071). The digested peptides were purified using an Oasis HLB 96-SPE well-plate and the eluted samples were concentrated with an Eppendorf Concentrator Plus system. The purified peptides were redissolved in 100 μ L of water containing 0.1% formic acid.

4.3.2. Parameters of the HPLC/MS Method

Nano separation-based proteomic analysis was performed by Bruker EASY-nLC equipment coupled with Bruker Maxis 4G UHR-QTOF instrument (Bruker Daltonics, Bremen, Germany). Five μ L aliquots of the samples were injected and separated by a homemade C18 analytical column (3 μ m, 75 μ m \times 150 mm) using a 120 min long multistep gradient elution at a flow rate of 250 nL min⁻¹. Two different solvents (Solvent A and B) were used for the separation. The mass spectrometer was operated in positive ion mode, and the scanning range was set to 300–2200 m/z. The flow rate of nebulizer gas was 2 L/min at a pressure of 0.6 bar, and the temperature was set to 160°C. The capillary voltage was 4.5 kV, and the 30 most intensive peptide peaks were selected for collision-induced dissociation (CID) fragmentation.

4.4. Measurement of ApoA1, Haptoglobin and HDL Cholesterol

Apolipoprotein A1 and haptoglobin levels were measured by immunoturbidimetric assays on a Cobas Integra 400 Plus analyzer while HDL cholesterol was determined by a homogeneous enzymatic colorimetric test on the c502/c702 modules of a Cobas 8000 analyzer (both instruments from Roche Diagnostics GmbH, Mannheim, Germany) at the Department of Laboratory Medicine (University of Pécs, Hungary; accreditation registration number: NAH-9-0008/2021).

4.5. Data Analysis

The obtained data from the MS measurements were processed using Bruker Data Analysis 4.4 software (Bruker Daltonik, Bremen, Germany). The protein identification was carried out using Mascot server (V2.4.1, Matrix Sciences Inc., Boston, MA, USA). Searching parameters were set to allow one missed cleavage site, accepting a 50 ppm mass error in MS1 and 0.3 Da in MS2 mode. The significance threshold was set to $p \leq 0.05$ for reliable identification. The classification of the identified proteins was performed by PANTHER classification system (v.17.0, <https://www.pantherdb.org/>). The protein-protein interaction network analysis of the identified proteins was performed by using STRING database (v.12.0, <https://string-db.org/>). For relative quantification MaxQuant software (v2.2.0.0, Max-Planck-Institute of Biochemistry, Munchen, Germany) was utilized in which a label-free quantification (LFQ) method was set. LFQ techniques rely on measuring the peak intensity or

area under the curve of the identified peptide ions to enable the comparative quantification of peptides across various samples (Mehta et al., 2020).

The statistical analysis of the obtained results was performed using SPSS (version 28.0.0.0, IBM SPSS Statistics, Armonk, New York, USA) and OriginPro 2023b (OriginLab, Northampton, Massachusetts, USA) programs. To determine the differences between the groups, Mann-Whitney U-test was performed. Values of $p < 0.05$ were considered statistically significant.

Supplementary Materials: The following supporting information can be downloaded at the website of this paper posted on Preprints.org.

Author Contributions: Conceptualization, T.K., C.K., and G.L.K.; methodology, T.K., C.K., J.S., Z. H-S., P.M., K.G.; software, C.K., and J.S.; validation, T.K., C.K., and G.L.K.; formal analysis, T.K., C.K., and J.S.; investigation, C.K.; resources, T.K., Á.V., G.L.K., and J.S.; data curation, T.K. and C.K.; writing—original draft preparation, T.K. and C.K.; writing—review and editing, T.K., C.K., and G.L.K.; visualization, C.K.; supervision, T.K. and G.L.K.; project administration, C.K.; funding acquisition, T.K. and G.L.K. All authors have read and agreed to the published version of the manuscript.

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Data Availability Statement: The data that support the findings of this study are available from the corresponding author upon reasonable request.

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Reference:

1. F. Zegers-Hochschild *et al.*, "The International Glossary on Infertility and Fertility Care, 2017," *Fertil Steril*, vol. 108, no. 3, pp. 393–406, Sep. 2017, doi: 10.1016/j.fertnstert.2017.06.005.
2. M. Vander Borgh and C. Wyns, "Fertility and infertility: Definition and epidemiology," *Clinical Biochemistry*, vol. 62, Elsevier Inc., pp. 2–10, Dec. 01, 2018. doi: 10.1016/j.clinbiochem.2018.03.012.
3. G. Collodel *et al.*, "Follicular Fluid Components in Reduced Ovarian Reserve, Endometriosis, and Idiopathic Infertility," *Int J Mol Sci*, vol. 24, no. 3, Feb. 2023, doi: 10.3390/ijms24032589.
4. I. Kosteria, A. K. Anagnostopoulos, C. Kanaka-Gantenbein, G. P. Chrousos, and G. T. Tsangaris, "The use of proteomics in assisted reproduction," *In Vivo*, vol. 31, no. 3. International Institute of Anticancer Research, pp. 267–283, May 01, 2017. doi: 10.21873/invivo.11056.
5. S. Al-Amrani, Z. Al-Jabri, A. Al-Zaabi, J. Alshekaili, and M. Al-Khabori, "Proteomics: Concepts and applications in human medicine," *World J Biol Chem*, vol. 12, no. 5, pp. 57–69, Sep. 2021, doi: 10.4331/wjbc.v12.i5.57.
6. M. L. Hennet and C. M. H. Combelles, "The antral follicle: A microenvironment for oocyte differentiation," *International Journal of Developmental Biology*, vol. 56, no. 10–12, pp. 819–831, 2012, doi: 10.1387/ijdb.120133cc.
7. A. T. Brinca *et al.*, "Follicular Fluid: A Powerful Tool for the Understanding and Diagnosis of Polycystic Ovary Syndrome," *Biomedicines*, vol. 10, no. 6. MDPI, Jun. 01, 2022. doi: 10.3390/biomedicines10061254.
8. A. E. Lewandowska, K. Macur, P. Czaplewska, J. Liss, K. Łukaszuk, and S. Ołdziej, "Qualitative and Quantitative Analysis of Proteome and Peptidome of Human Follicular Fluid Using Multiple Samples from Single Donor with LC-MS and SWATH Methodology," *J Proteome Res*, vol. 16, no. 8, pp. 3053–3067, Aug. 2017, doi: 10.1021/acs.jproteome.7b00366.
9. L. Bianchi *et al.*, "Protein pathways working in human follicular fluid: The future for tailored IVF?," *Expert Reviews in Molecular Medicine*, vol. 18. Cambridge University Press, Jan. 18, 2016. doi: 10.1017/erm.2016.4.

10. G. Mariani and J. Bellver, "Proteomics and metabolomics studies and clinical outcomes," in *Reproductomics: The -Omics Revolution and Its Impact on Human Reproductive Medicine*, Elsevier, 2018, pp. 147–170. doi: 10.1016/B978-0-12-812571-7.00010-1.
11. X. Zhang, T. Wang, J. Song, J. Deng, and Z. Sun, "Study on follicular fluid metabolomics components at different ages based on lipid metabolism," *Reproductive Biology and Endocrinology*, vol. 18, no. 1, May 2020, doi: 10.1186/s12958-020-00599-8.
12. M. G. Da Broi, V. S. I. Giorgi, F. Wang, D. L. Keefe, D. Albertini, and P. A. Navarro, "Influence of follicular fluid and cumulus cells on oocyte quality: Clinical implications," *J Assist Reprod Genet*, vol. 35, no. 5, pp. 735–751, Mar. 2018, doi: 10.1007/s10815-018-1143-3.
13. M. Hashemitabar, M. Bahmanzadeh, A. Mostafaei, M. Orazizadeh, M. Farimani, and R. Nikbakht, "A proteomic analysis of human follicular fluid: Comparison between younger and older women with normal FSH levels," *Int J Mol Sci*, vol. 15, no. 10, pp. 17518–17540, Sep. 2014, doi: 10.3390/ijms151017518.
14. E. C. A. Porta, M. Balestrieri, M. Bianco, R. Picone, C. De Stefano, and P. Abrescia, "Haptoglobin transport into human ovarian follicles and its binding to apolipoprotein A-1," *Zygote*, vol. 7, pp. 67–77, 1999.
15. M. Balestrieri, L. Cigliano, M. L. De Simone, B. Dale, and P. Abrescia, "Haptoglobin inhibits lecithin-cholesterol acyltransferase in human ovarian follicular fluid," *Mol Reprod Dev*, vol. 59, no. 2, pp. 186–191, 2001, doi: 10.1002/mrd.1021.
16. T. E. Willnow, A. Hammes, and S. Eaton, "Lipoproteins and their receptors in embryonic development: More than cholesterol clearance," *Development*, vol. 134, no. 18, pp. 3239–3249, Sep. 2007. doi: 10.1242/dev.004408.
17. L. M. Mains, L. Christenson, B. Yang, A. E. T. Sparks, S. Mathur, and B. J. Van Voorhis, "Identification of apolipoprotein A1 in the human embryonic secretome," *Fertil Steril*, vol. 96, no. 2, 2011, doi: 10.1016/j.fertnstert.2011.05.049.
18. R. L. Stouffer, F. Xu, and D. M. Duffy, "Molecular control of ovulation and luteinization in the primate follicle," 2007.
19. M. Cherian-Shaw, M. Puttabyatappa, E. Greason, A. Rodriguez, C. A. Vandevoort, and C. L. Chaffin, "Expression of scavenger receptor-BI and low-density lipoprotein receptor and differential use of lipoproteins to support early steroidogenesis in luteinizing macaque granulosa cells," *Endocrinology*, vol. 150, no. 2, pp. 957–965, Feb. 2009, doi: 10.1210/en.2008-0619.
20. N. Argov, U. Moallem, and D. Sklan, "Lipid transport in the developing bovine follicle: Messenger RNA expression increases for selective uptake receptors and decreases for endocytosis receptors," *Biol Reprod*, vol. 71, no. 2, pp. 479–485, Aug. 2004, doi: 10.1095/biolreprod.104.028092.
21. A. S. Bhale and K. Venkataraman, "Leveraging knowledge of HDLs major protein ApoA1: Structure, function, mutations, and potential therapeutics," *Biomedicine and Pharmacotherapy*, vol. 154. Elsevier Masson s.r.l., Oct. 01, 2022. doi: 10.1016/j.biopha.2022.113634.
22. F. Amjadi, R. Aflatoonian, S. H. Javanmard, B. Saifi, M. Ashrafi, and M. Mehdizadeh, "Apolipoprotein A1 as a novel anti-implantation biomarker in polycystic ovary syndrome: A case-control study," *Journal of Research in Medical Sciences*, vol. 20, no. 11, pp. 1039–1045, Nov. 2015, doi: 10.4103/1735-1995.172813.
23. X. Chen, T. O. Scholl, T. P. Stein, R. A. Steer, and K. P. Williams, "Maternal circulating lipid profile during early pregnancy: Racial/ethnic differences and association with spontaneous preterm delivery," *Nutrients*, vol. 9, no. 1, Jan. 2017, doi: 10.3390/nu9010019.
24. P. Verma *et al.*, "High Level of APOA1 in Blood and Maternal Fetal Interface Is Associated With Early Miscarriage," *Reproductive Sciences*, vol. 26, no. 5, pp. 649–656, May 2019, doi: 10.1177/1933719118783266.
25. T. Rødgaard, P. M. H. Heegaard, and H. Callesen, "Non-invasive assessment of in-vitro embryo quality to improve transfer success," *Reproductive BioMedicine Online*, vol. 31, no. 5. Elsevier Ltd, pp. 585–592, Nov. 01, 2015. doi: 10.1016/j.rbmo.2015.08.003.
26. J. Nyalwidhe *et al.*, "The search for biomarkers of human embryo developmental potential in IVF: A comprehensive proteomic approach," *Mol Hum Reprod*, vol. 19, no. 4, pp. 250–263, Apr. 2013, doi: 10.1093/molehr/gas063.

27. D. R. Deutsch *et al.*, "Stage-specific proteome signatures in early bovine embryo development," *J Proteome Res*, vol. 13, no. 10, pp. 4363–4376, Oct. 2014, doi: 10.1021/pr500550t.
28. J. J. Brosens *et al.*, "Proteomic analysis of endometrium from fertile and infertile patients suggests a role for apolipoprotein A-I in embryo implantation failure and endometriosis," *Mol Hum Reprod*, vol. 16, no. 4, pp. 273–285, Dec. 2009, doi: 10.1093/molehr/gap108.
29. J. Zheng, Q. Yin, J. Cao, and B. Zhang, "Obesity contributes more to increasing ApoB/ApoA1 ratio than hyperandrogenism in PCOS women aged 20-38 years in China," *Exp Ther Med*, vol. 13, no. 4, pp. 1337–1342, Apr. 2017, doi: 10.3892/etm.2017.4094.
30. K. Kim, M. S. Bloom, R. W. Browne, E. M. Bell, R. M. Yucel, and V. Y. Fujimoto, "Associations between follicular fluid high density lipoprotein particle components and embryo quality among in vitro fertilization patients," *J Assist Reprod Genet*, vol. 34, no. 1, pp. 1–10, Jan. 2017, doi: 10.1007/s10815-016-0826-x.
31. B. Jaspard, X. Collet, R. Barbaras, J. Manent, C. Vieu, and J. Parinaud, "Biochemical Characterization of Pre-1 High-Density Lipoprotein from Human Ovarian Follicular Fluid: Evidence for the Presence of a Lipid Core \dagger , \ddagger ," 1996. [Online]. Available: <https://pubs.acs.org/sharingguidelines>
32. Jaspard B *et al.*, "Structural and functional comparison of HDL from homologous human plasma and follicular fluid. A model for extravascular fluid," *Arterioscler Thromb Vasc Biol.*, pp. 1605–13, 1997, doi: 10.1161/01.atv.17.8.1605.
33. A. Yesilaltay *et al.*, "Effects of hepatic expression of the high-density lipoprotein receptor SR-BI on lipoprotein metabolism and female fertility," *Endocrinology*, vol. 147, no. 4, pp. 1577–1588, Apr. 2006, doi: 10.1210/en.2005-1286.
34. A. Arias, A. Quiroz, N. Santander, E. Morselli, and D. Busso, "Implications of High-Density Cholesterol Metabolism for Oocyte Biology and Female Fertility," *Frontiers in Cell and Developmental Biology*, vol. 10. Frontiers Media S.A., Sep. 14, 2022. doi: 10.3389/fcell.2022.941539.
35. B. J. Ansell *et al.*, "Inflammatory/Anti-inflammatory Properties of High-Density Lipoprotein Distinguish Patients from Control Subjects Better Than High-Density Lipoprotein Cholesterol Levels and Are Favorably Affected by Simvastatin Treatment," *Circulation*, vol. 108, no. 22, pp. 2751–2756, Dec. 2003, doi: 10.1161/01.CIR.0000103624.14436.4B.
36. V. Y. Fujimoto, J. P. Kane, B. Y. Ishida, M. S. Bloom, and R. W. Browne, "High-density lipoprotein metabolism and the human embryo," *Human Reproduction Update*, vol. 16, no. 1. pp. 20–38, Aug. 28, 2009. doi: 10.1093/humupd/dmp029.
37. R. W. Browne, M. S. Bloom, W. B. Shelly, A. J. Ocque, H. G. Huddleston, and V. Y. Fujimoto, "Follicular fluid high density lipoprotein-associated micronutrient levels are associated with embryo fragmentation during IVF," *J Assist Reprod Genet*, vol. 26, no. 11–12, pp. 557–560, 2009, doi: 10.1007/s10815-009-9367-x.
38. T. Y. Tan, M. S. K. Lau, S. F. Loh, and H. H. Tan, "Female ageing and reproductive outcome in assisted reproduction cycles," *Singapore Med J*, vol. 55, no. 6, pp. 305–309, 2014, doi: 10.11622/smedj.2014081.
39. W. Wei, X. Zhang, B. Zhou, B. Ge, J. Tian, and J. Chen, "Effects of female obesity on conception, pregnancy and the health of offspring," *Frontiers in Endocrinology*, vol. 13. Frontiers Media S.A., Aug. 11, 2022. doi: 10.3389/fendo.2022.949228.
40. A. Pirnat, L. A. Deroo, R. Skjærven, and N. H. Morken, "Women's prepregnancy lipid levels and number of children: A Norwegian prospective population-based cohort study," *BMJ Open*, vol. 8, no. 6, Jun. 2018, doi: 10.1136/bmjopen-2017-021188.
41. J. Song, S. Xiang, C. Pang, J. Guo, and Z. Sun, "Metabolomic alternations of follicular fluid of obese women undergoing in-vitro fertilization treatment," *Sci Rep*, vol. 10, no. 1, Dec. 2020, doi: 10.1038/s41598-020-62975-z.
42. S. D. M. Valckx *et al.*, "BMI-related metabolic composition of the follicular fluid of women undergoing assisted reproductive treatment and the consequences for oocyte and embryo quality," *Human Reproduction*, vol. 27, no. 12, pp. 3531–3539, 2012, doi: 10.1093/humrep/des350.

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