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Review

# Application of Enzymes in Biomass Waste Management

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**Abstract:** Enzymes are biological molecules produced by living entities for carrying out biological processes. The application of enzymes for waste treatment has been gaining pace commercially to solve concerns related to agricultural residues, wastewater, replacement of synthetic processes with natural ones, etc. The application of enzymes for waste management has been an environmentally reliable and sustainable process. Treatment of waste with enzymes such as xylanase, proteases, hydrolases, cellulose, peroxidases, chitinases, laccases, etc. has been studied to be effective. These enzymes act upon the waste products and transform them into biodegradable forms that can be recycled, reused and converted to value-added products. They have wide applications and utility as it has been an effective approach, economically cheaper and sustainable techniques. Application of such enzymes for waste management would be beneficial for reducing the quantity of waste, diminishing the negative effects of waste and pollution on the environment, and would be beneficial in bio-converting the waste products into alternate sources of energy. The current chapter focuses on different types of enzymes, their applications for waste management, and their limitations. This chapter also emphasizes the usage of some prominent microorganisms, their secreted enzymes and their proposed mechanisms of action involved with the degradation of the waste products.

**Keywords:** enzymes; biomass; waste management; waste treatment; microorganisms; applications of enzymes

## 1. Introduction

Several anthropogenic activities like agriculture practices, industrialization, etc. are increasing day by day. These activities contribute to increasing the level of pollutants in the environment. These pollutants may be azo dyes, phenols, polycyclic aromatic hydrocarbons (PAHs), polychlorinated biphenyls (PCBs), pesticides, dioxins, as well as heavy metals (Bhardwaj et al. 2023a). There are several processes such as physical and chemical treatment and activated carbon adsorption that have been developed in recent years for removing pollutants from waste. Industrial wastes contain toxic organic as well as inorganic materials. The presence of organic materials in the waste may be treated biologically for the minimization of the overall treatment cost.

Enzymes can play an important role in the treatment of waste. The use of enzymes in waste treatment was first proposed in 1930. They are target-specific and can particularly attack target pollutants. They are used as an efficient catalyst and are used at a particular pH and temperature. Researchers were focusing on the development of new technology for waste treatment which will be faster, cheaper, simpler, and more reliable. Now, they have developed enzymatic processes with the help of biotechnological tools. This technique replaces costly chemicals and does not require specialized machinery. It is a highly acceptable technique in its clean/green and biodegradable nature

due to several advantages. It falls between the physicochemical and biological processes because it contains chemical processes that are based on the biological catalysis action.

Enzymatic conversion of biomass offers several advantages over chemical and physical techniques. These advantages are:

It can be operated at high as well as low concentrations of contaminant.

It could be operated over a broader range of parameters of pH, temperature, as well as salinity.

There are no shock loading effects.

There is a reduction in the volume of sludge.

The process of controlling is simple.

It is an environmentally friendly technique.

With these potential advantages, researchers have focused on the use of this technique for the treatment of wastewater, solid wastes, and hazardous wastes.

Biomass waste treatment uses enzymes from microorganisms such as bacteria or fungi. They are involved in several metabolic activities and produce different enzymes. These enzymes are specific to their work and involve a series of chemical reactions. Enzymes that are produced by aerobic bacteria (*Alcaligenes*, *Mycobacterium*, *Pseudomonas*, *Rhodococcus*, and *Sphingomonas*) are used in the degradation of pesticides and hydrocarbons while those enzymes produced by anaerobic bacteria that are used in the degradation of compounds like polychlorinated biphenyls (PCBs), dichlorination of trichloroethylene (TCE), as well as chloroform (Sharma 2012).

The pollutants including compounds containing azo dyes, phenols, PAHs, pesticides, polychlorinated compounds, heavy metals, etc. have been studied to create adverse effects of teratogenic nature, carcinogenic potential, mutagenic, as well as toxic effects on the health of humans (Liu et al. 2019; Alam et al. 2023; Bhardwaj et al. 2023b). However, the enzymatic technique is slow, and only some bacterial species are capable enough to produce specific enzymes. So, the researchers prefer genetically engineered microbes for this process. This process converts the pollutants from toxic forms to nontoxic forms (Phale et al. 2019). Thermophilic bacteria *Caldicelluloseruptor bescii* has been studied to directly convert plant biomass into bioethanol. This could be used as a potential agent in the commercial sector for bioethanol production (Chung et al., 2014). Similarly, hemicellulases, cellulases, xylanases, endoxylanases, and  $\beta$ -xylosidases extracted from a variety of ascomycetes *Trichoderma*, *Aspergillus*, etc. have been applied extensively at commercial levels to act as biocatalysts in lignocellulosic biorefineries (Ferreira et al., 2016). A recent study has shown that the co-cultivation and harvesting of microalgae and filamentous fungi were efficiently used for wastewater treatment as well as biofuel production (Chu et al., 2021).

The aim of this chapter is to focus on different types of enzymes and their applications for waste treatment.

## 2. Enzymes Used for Biomass Conversion, Degradation, and Hydrolysis

Enzymes can help in converting biomass into various useful by-products. Biomass conversion involves the catalysis of complex organic materials, such as plant fibers, feedstocks, agricultural waste, etc. into simple components. These components could further be processed into biofuels, biochemicals, and other valuable products (Demirbas 2009; Mahapatra et al. 2021). Several types of enzymes are commonly used in biomass conversion processes (Table 1). Some of the example of enzymes are given below:-

**Amylases:** They are extracted from fungal and bacterial strains that hydrolyze starch into glucose and maltose. Starch is often used as a feedstock in biomass conversion processes, and amylases are employed at a commercial scale to convert it into fermentable sugars for biofuel/bioethanol production (Castro et al., 2011).

**Cellulases:** They helps in the breakdown of cellulose into glucose moieties and cellulose is a major component of plant cell walls formed by  $\beta$ -1,4-glycosidic bonds. They consist of three main types: endoglucanases, exoglucanases (or cellobiohydrolases), and  $\beta$ -glucosidases (Dashtban et al.,

2010). Cellulases have been observed to be important in the biofuel production like cellulosic ethanol and in the conversion of biomass into various biochemicals (Siqueira et al., 2020).

**Hemicellulases:** They target hemicellulose which is another complex carbohydrate present in plant cell walls. These enzymes include xylanases, mannanases, and arabinases (Zanuso et al., 2021). Bacteria and fungi produce large amounts of hemicellulases. They break down hemicellulose into smaller sugar units, such as xylose, mannose, and arabinose, which can be fermented into biofuels or used as building blocks for biochemical production (Méndez-Líter et al., 2021).

**Ligninases:** They include lignin peroxidases and manganese peroxidases and are involved in the degradation of lignin which is a complex and highly resistant polymer found in plant biomass. Lignin allows access to the cellulose and hemicellulose components for further enzymatic degradation or chemical processing (Siqueira et al. 2020; Bilal and Iqbal 2021).

**Proteases:** They are also known as peptidases and are responsible for breaking down proteins into smaller peptides and further to amino acids.

**Laccases:** They are extracellular enzymes containing multi-copper that consists of glycoproteins with dimeric, tetrameric and monomeric units characterized by bacteria, fungi, and plants (Shekher et al. 2011). Lignin as well as phenolic compounds which are found in banana peels, sawdusts, and rice bran have enhanced production of laccase (Muthukumarasamy et al. 2015).

**Table 1.** List of the Different Enzymes with Their Sources and Applications.

| S. No. | Name of Enzyme              | Sources   | Application of Enzymes  | References  |
|--------|-----------------------------|---|---|---|
| 1      | Alkylsulfatase              | <i>Pseudomonas</i> C12B   | Surfactant degradation  | Toesch et al. 2014  |
| 2      | Azoreductase                | Intestinal microflora   | Removal of azo dyes   | Sandhya 2010  |
| 3      | $\alpha$ -Amylase           | <i>Bacillus subtilis</i>  | Glucose production and hydrolysis of starch   | Kolusheva and Marinova 2007; Presečki et al. 2013           |
| 4      | Glucoamylase                |   |   |   |
| 5      | Cellulase                   | <i>Trichoderma harzianum</i> ,<br><i>Trichoderma viride</i>   | Hydrolysis of cellulose in sludges & municipal solid waste (MSW) to produce alcohol, sugars, and energy   | Champagne and Li 2009; Khan et al. 2016; Pandey et al. 2017 |
| 6      | Cellobio-hydrolase          |   |   |   |
| 7      | Cellobiose                  |   |   |   |
| 8      | Exo-1,4-b-D-glucosidase     |   |   |   |
| 9      | Chitinase                   | <i>Streptomyces anulatus</i> CS242  | Production of N-acetyl glucosamine from shellfish waste through bioconversion   | Mander et al. 2016  |
| 10     | Chloro-peroxidase           | <i>Caldariomyces fumago</i>   | Oxidation of phenolic compounds   | Sjogblad and Bollag 2021                                    |
| 11     | Cyanidase                   | <i>Pseudomonas</i> sp.  | Cyanide decay   | Akcil et al. 2003   |
| 12     | Cyanide hydratase           | <i>Fusarium lateritium</i>  | Cyanide hydrolysis  | Ebbs 2004   |
| 13     | Depolymerase                | Bacteriophage   | Bacterial exopolysaccharide   | Knecht et al. 2020  |
| 14     | Haemoglobin                 | Blood   | Removal of aromatic amines and phenols  | Pérez-Prior et al. 2014                                     |
| 15     | L-Galactono-lactone oxidase | <i>Candida norvegensis</i>  | Conversion of galactose from L-ascorbic acid  | Nicell 2003   |
| 16     | Laccase                     | <i>Pleurotus</i> ( <i>P. ostreatus</i> , <i>P. pulmonarius</i> ) and <i>Trametes</i> ( <i>T. versicolor</i> , <i>T. hirsuta</i> ) | Decolorization of kraft, removal of phenols, bleaching of paper pulp, binding of phenols and aromatic amines with humas, detoxification of wastewater | Khatami et al. 2022   |
| 17     | Lactases                    | Bacterial   | Dairy waste processing and production of value-added products   | Coughlin and Charles 2022                                   |
| 18     | Lignin peroxidase (LiP)     | <i>Phanerochaete chrysosporium</i>  | Removal of phenols and aromatic compounds decolorization of Kraft bleaching effluents   | Falade et al. 2017  |
| 19     | Lipase                      | Various sources   | Improved sludge dewatering  | Nimkande and Bafana (2022); Di et al. 2023                  |
| 20     | Lysozyme                    | Bacterial   |   |   |
| 21     | Manganese peroxidase (MnP)  | <i>Phanerochaete chrysosporium</i>  | Decolorization of synthetic dyes, removal of phenolic contaminants, removal of endocrine disruptive   | Bansal and Kanwar 2013                                      |

|    |                      |   |   |   |
|----|----------------------|---|---|---|
|    |                      |   | chemicals (EDC), degradation of chlorinated alkanes and alkenes, degradation of chlorinated dioxins |   |
| 22 | Nitrile hydratase    | Mesorhizobium sp.   | Removal of acrylonitrile  | Feng et al. 2008                        |
| 23 | Parathion hydrolase  | Azohydromonas australica                                      | Hydrolyzation of organophosphate pesticides   | Zhao et al. 2021                        |
| 24 | Pectin Lyase         | Clostridium beijerinckii                                      | Pectin degradation  | Yadav et al. 2009                       |
| 25 | Pectinmethylesterase | Aspergillus niger   |   | Kohli et al. 2015                       |
| 26 | Phosphatase          | Escherichia coli C90  | Removal of heavy metals   | Chaudhuri et al. 2013                   |
| 27 | Phosphoesterases     | Aspergillus sydowii CBMAI 935                                 | Removal of chlorpyrifos, diazinon, parathions   | Soares et al. 2021                      |
| 28 | Polyphenol oxidase   | Mushroom (Agaricus bisporus)                                  | Removal of phenolic compounds   | Li et al. 2021                          |
| 29 | Proteases            | Bacillus licheniformis, Aspergillus niger, Chlorella vulgaris | Hydrolyze or breakdown the protein molecules in meat, biodegradation of the industrial sludge       | Karn and Kumar 2015; Arslan et al. 2021 |
| 30 | Tyrosinase           | Mushroom (Agaricus bisporus)                                  | Removal of phenolic compounds   | Bayramoglu et al. 2013                  |

3. Mechanism of Treatment of Biomass:

The enzymes break down complex organic compounds into simpler components, as depicted in Figure 1. The mechanism involves the following steps:

3.1. Pretreatment of Biomass Wastes

Biomass waste like agricultural residues, wooden chips, residual crops, etc. are pretreated to enhance the enzyme’s accessibility to the complex carbohydrates present in biomass. Such techniques include combinations of physical, chemical, or biological means for example, acid or alkali treatment, fungal/bacterial/yeast treatment, milling, steam explosion, etc. These methods are capable of disrupting the biomass structure thereby making it more susceptible to enzymatic degradations (Zhang et al., 2021).

3.2. Enzyme Production

Microorganisms that are capable of producing enzymes suitable for biomass degradation are cultivated in a controlled environment. These microbial cultures are grown in bioreactors under specific conditions that promote enzyme production either naturally or by genetic engineering (Guo et al., 2023).

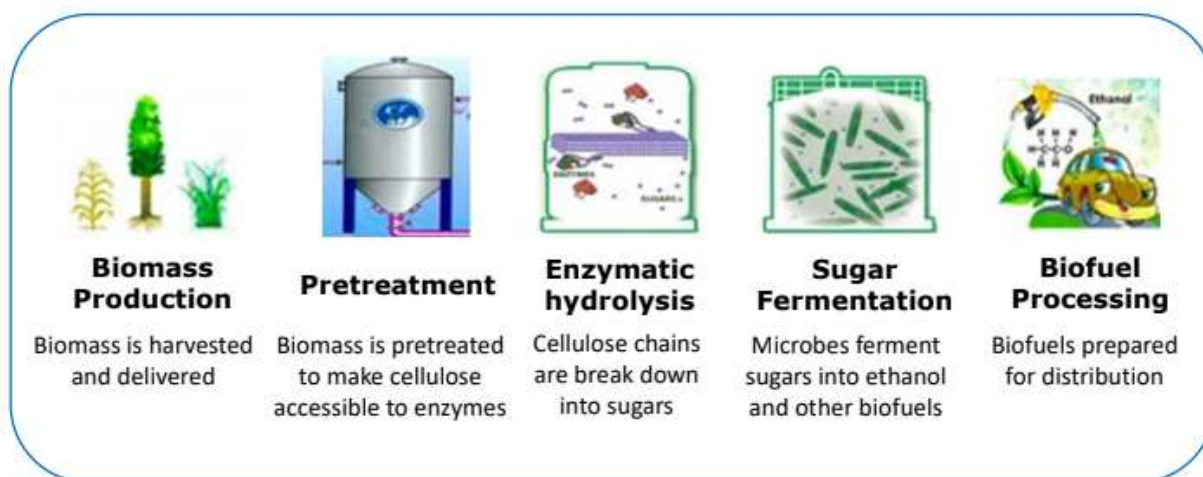
3.3. Enzymatic Hydrolysis

The pretreated biomass and enzyme solutions are mixed to initiate the hydrolysis process. Enzymes, such as cellulases, hemicellulases, lignin, act on complex carbohydrates of biomass and hydrolyze them into simpler molecules. These enzymes work synergistically to degrade different components of the biomass, releasing glucose, xylose, and other sugars (Huang et al. 2022a).

3.4. Fermentation and Further Processing

The resultant sugar-rich hydrolysate is subjected to fermentation using microorganisms, such as yeast or bacteria, to convert the sugars into desired end products. For example, ethanol-producing microorganisms can ferment the sugars to produce bioethanol, while other microorganisms can convert the sugars into various biochemicals, such as organic acids, enzymes, or bio-based materials (Huang et al. 2022a; Huang et al. 2022b).





**Figure 1.** Diagrammatic Representation of the Mechanism of Biomass Wastes Treatment.

#### 4. Application of Biomass Waste Management for Treating Contaminated Wastewater

Water is necessary for all living things (Bhardwaj and Sharma 2021a; Bhardwaj and Sharma 2021b; Bhardwaj 2022; Bhardwaj 2023). Now, the water bodies have been contaminated with the effluent of various industries, institutes, and residential areas. This effluent comprises decomposed organic waste that can generate gases with an awful odor in great amounts. This polluted water contains toxic persistent pollutants which might be hazardous to the environment (Bhardwaj and Jindal 2019; Bhardwaj and Jindal 2020; Bhardwaj et al. 2021; Bhardwaj and Jindal 2022). The usage of enzymes for the treatment of wastewater reduces the toxicity of water and makes the water fit to reuse.

Most of the pollutants act as substrates for certain enzymes. By the action of enzymes, the toxic pollutants are fragmented into smaller fragments that no longer pose a danger to the surroundings. Enzymes can be introduced directly or even mixed with microbes. The entire body of plants or their tissue culture that contains the enzymes in their normal form can be deployed in water bodies. BIO-CAT's enzymes are effectively used to treat organic waste (Sahal et al. 2023).

Certain enzymes are required to catalyze specific reactions in specified concentrations. Oxidoreductases, peroxidases, and oxygenases are used for the removal of inorganic contaminants like biphenols, chlorophenols, benzidines, methylated phenols, phenols, heterocyclic aromatic compounds, and anilines while lipase, urease, amylase, xylanases, cellulase, and protease are used for the removal of organic pollutants. These enzymes are called septic tank enzymes and are utilized for the treatment of wastewater from sewage. Oxygenases have been used for treatment of water bodies that are poisoned by the presence of fossil fuels. Peroxidases are utilized for the treatment of water that is contaminated with dyes, phenols, and hydrogen peroxide. Polyphenol oxidases are used in wastewater treatment plants for the removal of phenol from wastewater and can further be divided into Tyrosinase and Laccase.

Joutey et al. (2013) studied the degradation of persistent organic pollutants (POPs) by using enzymes from microbes and stated that this technique is environment-friendly, cost-effective, and innovative. Enzymes, for example, cytochrome P<sub>450s</sub> laccases, hydrolases, dehalogenases, dehydrogenases, proteases, and lipases, are involved in the degradation of harmful chemicals (Bhandari et al. 2021).

Cytochrome P<sub>450</sub> is responsible for synthesizing natural products in living organisms, and it is used in biotransformation of poisonous chemicals present in our ecosystem (Li et al. 2020). This enzyme has an intrinsic capability to degrade xenobiotics (Anzenbacher and Anzenbacherova 2001; Chakraborty and Das 2016). P<sub>450</sub> BM<sub>3</sub> is used in the degradation of several organic pollutants using Pt/TiO<sub>2</sub>-Cu under solar radiation and is formed from *E. coli* BL<sub>21</sub> (Awad and Mohamed 2019).

Laccase has been observed to possess catalytic capability to degrade the aromatic amine and phenolic compounds (Chandra and Chowdhary 2015). It degrades the PAHs to carbon dioxide (CO<sub>2</sub>) (Khelifi et al. 2010) and converts acenaphthylene to 1,8-naphthalic acid and 1,2-acenaphthalenedione (Madhavi and Lele 2009). It can detoxify dyes produced by the textile industry (Sondhi et al. 2015).

Dehalogenases belong to the oxidoreductase family and are used for degradation of halogenated compounds. Reductive, oxygenolytic, and hydrolytic mechanisms are used in the breakdown of halogenated compounds (Wang et al. 2018a; Wang et al. 2018b). Halohydrin dehydrogenase (HHDH) and haloalkane dehydrogenase (HADH) are used in the degradation of haloalkane and are expressed in *E. coli* (Xue et al. 2018). 2,4,6-trichlorophenol reductive dehalogenase dechlorinates pentachlorophenol (PCP) into 3-chlorophenol and is isolated from *Desulfitobacterium frappieri* PCP-1 (Boyer et al. 2003). Alcohol dehydrogenase catalyzes the conversion process of alcohol into ketone or aldehyde. It also catalyzed NAD(P)<sup>+</sup>-dependent oxidation of aldehydes into carboxylic acids.

Polyethylene glycol dehydrogenase (PEGDH) degrades the pollutants which are released from industries. The enzyme NAD<sup>+</sup>-dependent polypropylene glycol dehydrogenase (PPGDH) is isolated from the species *Stenotrophomonas maltophilia* and plays a role in oxidization of secondary alcohols (Tachibana et al. 2008). Polyvinyl alcohol dehydrogenase (PVADD) degrades polyvinyl alcohol which is water-soluble contaminants (Hirota-Mamoto et al. 2006). Aldehyde dehydrogenase (ADH) degrades aromatic compounds and catalyzes the conversion of 1-hydroxy-2-naphthaldehyde to 1-hydroxy-2-naphthoic (Ji et al. 2020).

Hydrolases reduce pollutants toxicity by degrading the larger molecules to small molecules. Hydrolytic enzymes like amylases, proteases, lipases, esterases, nitrilases, cellulases, cutinase, as well as peroxidases can be used in the degradation of insecticides, and oil-contaminated soils. They are used in biomedical sciences, and chemical industries (Kumar and Sharma 2019). Hydrolases such as parathion hydrolase or carbamate which are isolated from *Pseudomonas*, *Achromobacter*, *Flavobacterium*, *Bacillus cereus*, and *Nocardia* have been used for converting diazinon, carbofuran, coumaphos, and carbaryl / parathion through the method of hydrolysis.

Organophosphate pesticides (OPs) degrade through hydrolysis of P-O-alkyl and P-O-aryl bonds (Singh 2014). Malathion is degraded by the action of *Brevibacillus* sp., *Alicyclobacillus tengchogenesis*, *Bacillus cereus*, and *Bacillus licheniformis* (Littlechild 2015). OP acid anhydrolases enzymes, methyl parathion hydrolases (MPH), and OP hydrolases are used for the degradation of Organophosphates (OPs) (Schenk et al. 2016). Cutinase is used for the degradation of polycaprolactone, and it is isolated from the bacterial species *Fusarium solani* f. *pisi* (Singh et al. 2016). Protease belongs to the hydrolase family and is isolated from *Amycolatopsis* sp., *Aspergillus* sp., as well as *Bacillus* sp. They are low cost and catalyzing enzymes and are used in industries of food, leather as well as wastewater treatment (Kumar and Sharma 2019). They are capable of the degradation of poly(hydroxybutyrate) (PHB) depolymerase  $\beta$ -ester bonds, lipase  $\gamma$ - $\omega$  bonds, as well as  $\alpha$ -ester bonds (Haider et al. 2019). They convert marine crustacean wastes and keratinous wastes into useful products. Keratinase is isolated from *Stenotrophomonas maltophilia* KB<sub>13</sub> and used in the degradation of chicken feathers (Bhange et al. 2016). Keratinase is used in the leather industry, and it reduces the chemical (CaO and Na<sub>2</sub>S) load in wastewater (Akhter et al. 2020). The enzyme chitinase is isolated from *Bacillus subtilis* and degrades crystalline chitin into N-acetyl-D-glucosamine (Wang et al. 2018a). *Pseudomonas fluorescens* degrades polyurethane (PU) within 4-5 days using the enzyme protease.

Lipases are a well-known biocatalyst and degrade lipids (Casas-Godoy et al. 2012). They are used in the degradation of contaminants such as petroleum, residues of oil, greasy effluents, as well as oil spills (Basheer et al. 2011; Casas-Godoy et al. 2012; Hassan et al. 2018). Lipases which are isolated from species of *Pseudomonas* used for degradation of industrial waste oil (Amara and Salem 2009). Crude lipase which is formed from the species *Bacillus subtilis* is used in soap industries to reduce phosphate-based chemicals (Saraswat et al. (2017). Lipase PL which is isolated from *Alcaligenes* sp., catalyzes the conversion of poly (L-lactide) (PLA) polymers to the monomers. Lipases which are formed from *Lactobacillus plantarum* and *Lactobacillus brevis* are used in the degradation of polycaprolactone (PCL), and polyester (Wang et al. 2022).

## 5. Challenges and Future Prospects

Biomass waste treatment using enzymes from microorganisms is an area of ongoing research and development and has gained attention. Researchers are continuously researching to improve enzyme efficiency, process condition optimization, development of more cost-effective and sustainable methods, etc. (Madhavan et al., 2021). Utilizing biomass resources, enzyme diversity and specificity, have been areas of active research. One of the main challenges is scaling-up the biomass waste treatment from the laboratory to industrial-scale processes (Kalak 2023). Advancement in enzyme engineering techniques, protein engineering, meta-genomics studies are required for developing enzymes with enhanced catalytic properties, stability, and specificity (Kate et al., 2022). Biomass waste management using enzymes could also be integrated with bio-refineries, an together aim to produce multiple valuable products that are more sustainable and economically viable such as biofuels, bio-based chemicals, materials, etc. (Leong et al., 2021). Analyzing the properties of biomass waste materials would be helpful for understanding the appropriate methods to utilize them for energy and thereby reducing the consumption of fossil fuels in the future (Savla et al. 2021).

## 6. Conclusion

There is a need for continuous and vigorous research on usage of enzymes for the treatment of waste. Many industries such as textiles, agro, food & beverages, solid-waste treatment, sugar-mill, etc. are using enzymes for the treatment of wastes. The use of enzymes is a cost-effective, and sustainable technique for the treatment of waste. The usage of enzymes offers many advantages in biomass waste management strategies. Enzymatic processes are generally environmentally friendly, highly specific in their actions, operated under mild conditions, produce minimal waste as compared to traditional chemical methods. Furthermore, enzymes are produced through sustainable and renewable sources, such as microbial cultures. The application of enzymes for biomass waste management on a wide scale/industrial level has challenges. Overall, the application of enzymes for biomass waste management represents a promising approach towards achieving a more sustainable and circular economy.

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## References

1. Alam, S., Bhardwaj, L.K., Mallick, R., Rai, S. (2023). Estimation of Heavy Metals and Fluoride Ion in Vegetables Grown Nearby the Stretch of River Yamuna, Delhi (NCR), India. *Indian Journal of Environmental Protection*, 43 (1): 64-73.
2. Akcil, A., Karahan, A. G., Ciftci, H., & Sagdic, O. (2003). Biological treatment of cyanide by natural isolated bacteria (*Pseudomonas* sp.). *Minerals engineering*, 16(7), 643-649.
3. Akhter, M., Wal Marzan, L., Akter, Y., & Shimizu, K. (2020). Microbial bioremediation of feather waste for keratinase production: an outstanding solution for leather dehairing in tanneries. *Microbiology insights*, 13, 1178636120913280.
4. Amara, A. A., & Salem, S. R. (2009). Degradation of castor oil and lipase production by *Pseudomonas aeruginosa*. *American-Eurasian J Agric Environ Sci*, 5(4), 556-63.
5. Anzenbacher, P., & Anzenbacherova, E. (2001). Cytochromes P450 and metabolism of xenobiotics. *Cellular and Molecular Life Sciences CMLS*, 58, 737-747.
6. Arslan, N. P., Yazici, A., Komesli, S., Esim, N., & Ortucu, S. (2021). Direct conversion of waste loquat kernels to pigments using *Monascus purpureus* ATCC16365 with proteolytic and amylolytic activity. *Biomass Conversion and Biorefinery*, 11, 2191-2199.
7. Awad, G., & Mohamed, E. F. (2019). Immobilization of P450 BM3 monooxygenase on hollow nanosphere composite: application for degradation of organic gases pollutants under solar radiation lamp. *Applied Catalysis B: Environmental*, 253, 88-95.
8. Bansal, N., & Kanwar, S. S. (2013). Peroxidase (s) in environment protection. *The Scientific World Journal*, 2013.
9. Basheer, S. M., Chellappan, S., Beena, P. S., Sukumaran, R. K., Elyas, K. K., & Chandrasekaran, M. (2011). Lipase from marine *Aspergillus awamori* BTMFW032: production, partial purification and application in oil effluent treatment. *New Biotechnology*, 28(6), 627-638.



10. Bayramoglu, G., Akbulut, A., & Arica, M. Y. (2013). Immobilization of tyrosinase on modified diatom biosilica: Enzymatic removal of phenolic compounds from aqueous solution. *Journal of hazardous materials*, 244, 528-536.
11. Bhandari, S., Poudel, D. K., Marahatha, R., Dawadi, S., Khadayat, K., Phuyal, S., ... & Parajuli, N. (2021). Microbial enzymes used in bioremediation. *Journal of Chemistry*, 2021, 1-17.
12. Bhange, K., Chaturvedi, V., & Bhatt, R. (2016). Feather degradation potential of *Stenotrophomonas maltophilia* KB13 and feather protein hydrolysate (FPH) mediated reduction of hexavalent chromium. *3 Biotech*, 6, 1-9.
13. Bhardwaj, L.K. (2022). Evaluation of Bis (2-ethylhexyl) Phthalate (DEHP) in the PET Bottled Mineral Water of Different Brands and Impact of Heat by GC-MS/MS. *Chemistry Africa*, 5(4), 929-942.
14. Bhardwaj, L.K., & Jindal, T. (2020). Persistent organic pollutants in lakes of Grovnes Peninsula at Larsemann Hill area, East Antarctica. *Earth Systems and Environment*, 4, 349-358.
15. Bhardwaj, L.K., & Sharma, A. (2021a). Microplastics (MPs) in Drinking Water: Uses, Sources & Transport. DOI: <https://doi.org/10.20944/preprints202104.0498.v1>
16. Bhardwaj, L. K. (2023). Occurrence of Microplastics (MPs) in Antarctica and Its Impact on the Health of Organisms. *Maritime Technology and Research*, 6(2):265418. DOI: <https://doi.org/10.33175/mtr.2024.265418>
17. Bhardwaj, L.K., & Sharma, A. (2021b). Estimation of physico-chemical, trace metals, microbiological and phthalate in PET bottled water. *Chemistry Africa*, 4(4), 981-991.
18. Bhardwaj, L. K., Sharma, S., & Jindal, T. (2021). Occurrence of polycyclic aromatic hydrocarbons (PAHs) in the lake water at Grovnes Peninsula Over East Antarctica. *Chemistry Africa*, 4, 965-980.
19. Bhardwaj, L.K., & Jindal, T. (2019). Contamination of Lakes in Broknes peninsula, East Antarctica through the Pesticides and PAHs. *Asian-Journal of Chemistry*, 31(7), 1574-1580.
20. Bhardwaj, L.K., & Jindal, T. (2022). Polar Ecotoxicology: Sources and Toxic Effects of Pollutants. *New Frontiers in Environmental Toxicology*, 9-14.
21. Bhardwaj, L.K., Kumar, D., & Kumar, A. (2023a). Phytoremediation Potential of *Ocimum Sanctum*: A Sustainable Approach for Remediation of Heavy Metals. <https://doi.org/10.20944/preprints202308.0593.v1>
22. Bhardwaj, L.K., Sharma, S., & Jindal, T. (2023b). Estimation of Physico-Chemical and Heavy Metals in the Lakes of Grovnes & Broknes Peninsula, Larsemann Hill, East Antarctica. *Chemistry Africa*, 1-18.
23. Bilal, M., & Iqbal, H. M. (2021). Ligninolysis potential of ligninolytic enzymes: a green and sustainable approach to bio-transform lignocellulosic biomass into high-value entities. *Alternative Energy Resources: The Way to a Sustainable Modern Society*, 151-171.
24. Boyer, A., Pagé-BéLanger, R., Saucier, M., Villemur, R., Lépine, F., Juteau, P., & Beaudet, R. (2003). Purification, cloning and sequencing of an enzyme mediating the reductive dechlorination of 2, 4, 6-trichlorophenol from *Desulfitobacterium frappieri* PCP-1. *Biochemical Journal*, 373(1), 297-303.
25. Casas-Godoy, L., Duquesne, S., Bordes, F., Sandoval, G., & Marty, A. (2012). Lipases: an overview. Lipases and phospholipases: methods and protocols, 3-30.
26. Castro, A. M., Castilho, L. R., & Freire, D. M. (2011). An overview on advances of amylases production and their use in the production of bioethanol by conventional and non-conventional processes. *Biomass Conversion and Biorefinery*, 1, 245-255.
27. Chakraborty, J., & Das, S. (2016). Molecular perspectives and recent advances in microbial remediation of persistent organic pollutants. *Environmental Science and Pollution Research*, 23, 16883-16903.
28. Chandra, R., & Chowdhary, P. (2015). Properties of bacterial laccases and their application in bioremediation of industrial wastes. *Environmental Science: Processes & Impacts*, 17(2), 326-342.
29. Chaudhuri, G., Dey, P., Dalal, D., Venu-Babu, P., & Thilagaraj, W. R. (2013). A novel approach to precipitation of heavy metals from industrial effluents and single-ion solutions using bacterial alkaline phosphatase. *Water, Air, & Soil Pollution*, 224, 1-11.
30. Chu, R., Li, S., Zhu, L., Yin, Z., Hu, D., Liu, C., & Mo, F. (2021). A review on co-cultivation of microalgae with filamentous fungi: Efficient harvesting, wastewater treatment and biofuel production. *Renewable and Sustainable Energy Reviews*, 139, 110689.
31. Chung, D., Cha, M., Guss, A. M., & Westpheling, J. (2014). Direct conversion of plant biomass to ethanol by engineered *Caldicellulosiruptor bescii*. *Proceedings of the National Academy of Sciences*, 111(24), 8931-8936.
32. Coughlin, R. W., & Charles, M. (2019). Applications of lactase and immobilized lactase. In *Immobilized enzymes for food processing* (pp. 153-173). CRC Press.
33. Dai, Z., Liu, L., Duan, H., Li, B., Tang, X., Wu, X., ... & Zhang, L. (2023). Improving sludge dewaterability by free nitrous acid and lysozyme pretreatment: Performances and mechanisms. *Science of the Total Environment*, 855, 158648.
34. Dashtban, M., Maki, M., Leung, K. T., Mao, C., & Qin, W. (2010). Cellulase activities in biomass conversion: measurement methods and comparison. *Critical reviews in biotechnology*, 30(4), 302-309.
35. Demirbas, M. F. (2009). Biorefineries for biofuel upgrading: a critical review. *Applied energy*, 86, S151-S161.
36. Ebbs, S. (2004). Biological degradation of cyanide compounds. *Current opinion in Biotechnology*, 15(3), 231-236.
37. Falade, A. O., Nwodo, U. U., Iweriebor, B. C., Green, E., Mabinya, L. V., & Okoh, A. I. (2017). Lignin peroxidase functionalities and prospective applications. *MicrobiologyOpen*, 6(1), e00394.
38. Feng, Y. S., Chen, P. C., Wen, F. S., Hsiao, W. Y., & Lee, C. M. (2008). Nitrile hydratase from *Mesorhizobium* sp. F28 and its potential for nitrile biotransformation. *Process Biochemistry*, 43(12), 1391-1397.
39. Ferreira, J. A., Mahboubi, A., Lennartsson, P. R., & Taherzadeh, M. J. (2016). Waste biorefineries using filamentous ascomycetes fungi: present status and future prospects. *Bioresource Technology*, 215, 334-345.
40. Guo, H., Zhao, Y., Chang, J. S., & Lee, D. J. (2023). Enzymes and enzymatic mechanisms in enzymatic degradation of lignocellulosic biomass: A mini-review. *Bioresource Technology*, 367, 128252.
41. Haider, T. P., Völker, C., Kramm, J., Landfester, K., & Wurm, F. R. (2019). Plastics of the future? The impact of biodegradable polymers on the environment and on society. *Angewandte Chemie International Edition*, 58(1), 50-62.
42. Hassan, S. W., Abd El Latif, H. H., & Ali, S. M. (2018). Production of cold-active lipase by free and immobilized marine *Bacillus cereus* HSS: application in wastewater treatment. *Frontiers in microbiology*, 9, 2377.

43. Hirota-Mamoto, R., Nagai, R., Tachibana, S., Yasuda, M., Tani, A., Kimbara, K., & Kawai, F. (2006). Cloning and expression of the gene for periplasmic poly (vinyl alcohol) dehydrogenase from *Sphingomonas* sp. strain 113P3, a novel-type quinoxaemoprotein alcohol dehydrogenase. *Microbiology*, 152(7), 1941-1949.
44. Huang, C., Jiang, X., Shen, X., Hu, J., Tang, W., Wu, X., ... & Yong, Q. (2022b). Lignin-enzyme interaction: A roadblock for efficient enzymatic hydrolysis of lignocellulosics. *Renewable and Sustainable Energy Reviews*, 154, 111822.
45. Huang, C., Li, R., Tang, W., Zheng, Y., & Meng, X. (2022a). Improve Enzymatic Hydrolysis of Lignocellulosic Biomass by Modifying Lignin Structure via Sulfite Pretreatment and Using Lignin Blockers. *Fermentation*, 8(10), 558.
46. Ji, D., Mao, Z., He, J., Peng, S., & Wen, H. (2020). Characterization and genomic function analysis of phenanthrene-degrading bacterium *Pseudomonas* sp. Lphe-2. *Journal of Environmental Science and Health, Part A*, 55(5), 549-562.
47. Joutey, N. T., Bahafid, W., Sayel, H., & El Ghachtouli, N. (2013). Biodegradation: involved microorganisms and genetically engineered microorganisms. *Biodegradation-life of science*, 1, 289-320.
48. Khan, M. N., Luna, I. Z., Islam, M. M., Sharmeen, S., Salem, K. S., Rashid, T. U., ... & Rahman, M. M. (2016). Cellulase in waste management applications. In *New and future developments in microbial biotechnology and bioengineering* (pp. 237-256). Elsevier.
49. Khatami, S. H., Vakili, O., Movahedpour, A., Ghesmati, Z., Ghasemi, H., & Taheri-Anganeh, M. (2022). Laccase: Various types and applications. *Biotechnology and Applied Biochemistry*, 69(6), 2658-2672.
50. Khlifi, R., Belbahri, L., Woodward, S., Ellouz, M., Dhoub, A., Sayadi, S., & Mechichi, T. (2010). Decolourization and detoxification of textile industry wastewater by the laccase-mediator system. *Journal of Hazardous Materials*, 175(1-3), 802-808.
51. Knecht, L. E., Veljkovic, M., & Fieseler, L. (2020). Diversity and function of phage encoded depolymerases. *Frontiers in microbiology*, 10, 2949.
52. Kohli, P., Kalia, M., & Gupta, R. (2015). Pectin methylesterases: a review. *Journal of Bioprocessing & Biotechniques*, 5(5), 1.
53. Kumar, A., & Sharma, S. (2019). Microbes and enzymes in soil health and bioremediation (pp. 353-366). Singapore: Springer.
54. Leong, H. Y., Chang, C. K., Khoo, K. S., Chew, K. W., Chia, S. R., Lim, J. W., ... & Show, P. L. (2021). Waste biorefinery towards a sustainable circular bioeconomy: a solution to global issues. *Biotechnology for Biofuels*, 14(1), 1-15.
55. Li, S., Zhong, L., Wang, H., Li, J., Cheng, H., & Ma, Q. (2021). Process optimization of polyphenol oxidase immobilization: Isotherm, kinetic, thermodynamic and removal of phenolic compounds. *International Journal of Biological Macromolecules*, 185, 792-803.
56. Li, Z., Jiang, Y., Guengerich, F. P., Ma, L., Li, S., & Zhang, W. (2020). Engineering cytochrome P450 enzyme systems for biomedical and biotechnological applications. *Journal of Biological Chemistry*, 295(3), 833-849.
57. Littlechild, J. A. (2015). Archaeal enzymes and applications in industrial biocatalysts. *Archaea*, 2015.
58. Liu, L., Bilal, M., Duan, X., & Iqbal, H. M. (2019). Mitigation of environmental pollution by genetically engineered bacteria—current challenges and future perspectives. *Science of The Total Environment*, 667, 444-454.
59. Madhavan, A., Arun, K. B., Binod, P., Sirohi, R., Tarafdar, A., Reshmy, R., ... & Sindhu, R. (2021). Design of novel enzyme biocatalysts for industrial bioprocess: Harnessing the power of protein engineering, high throughput screening and synthetic biology. *Bioresource Technology*, 325, 124617.
60. Madhavi, V., & Lele, S. S. (2009). Laccase: properties and applications. *BioResources*, 4(4).
61. Mahapatra, S., Kumar, D., Singh, B., & Sachan, P. K. (2021). Biofuels and their sources of production: A review on cleaner sustainable alternative against conventional fuel, in the framework of the food and energy nexus. *Energy Nexus*, 4, 100036.
62. Mander, P., Cho, S. S., Choi, Y. H., Panthi, S., Choi, Y. S., Kim, H. M., & Yoo, J. C. (2016). Purification and characterization of chitinase showing antifungal and biodegradation properties obtained from *Streptomyces anulatus* CS242. *Archives of pharmacal research*, 39, 878-886.
63. Méndez-Líter, J. A., de Eugenio, L. I., Nieto-Domínguez, M., Prieto, A., & Martínez, M. J. (2021). Hemicellulases from *Penicillium* and *Talaromyces* for lignocellulosic biomass valorization: A review. *Bioresource Technology*, 324, 124623.
64. Muthukumarasamy, N. P., Jackson, B., Joseph Raj, A., & Sevanan, M. (2015). Production of extracellular laccase from *Bacillus subtilis* MTCC 2414 using agroresidues as a potential substrate. *Biochemistry research international*, 2015.
65. Nicell, J. A. (2003). Enzymatic treatment of waters and wastes. In *Chemical Degradation Methods for Wastes and Pollutants* (pp. 395-441). CRC Press.
66. Nimkande, V. D., & Bafana, A. (2022). A review on the utility of microbial lipases in wastewater treatment. *Journal of Water Process Engineering*, 46, 102591.
67. Pandey, K., Singh, B., Pandey, A. K., Badruddin, I. J., Pandey, S., Mishra, V. K., & Jain, P. A. (2017). Application of microbial enzymes in industrial waste water treatment. *International Journal of Current Microbiology and Applied Sciences*, 6(8), 1243-1254.
68. Kalak, T. (2023). Potential Use of Industrial Biomass Waste as a Sustainable Energy Source in the Future. *Energies*, 16(4), 1783.
69. Pérez-Prior, M. T., González-Sánchez, M. I., & Valero, E. (2014). Removal of aromatic compounds from wastewater by hemoglobin soluble and immobilized on Eupergit® CM. *Environmental Engineering & Management Journal (EEMJ)*, 13(10).
70. Phale, P. S., Sharma, A., & Gautam, K. (2019). Microbial degradation of xenobiotics like aromatic pollutants from the terrestrial environments. In *Pharmaceuticals and personal care products: waste management and treatment technology* (pp. 259-278). Butterworth-Heinemann.
71. Presečki, A. V., Blažević, Z. F., & Vasić-Rački, Đ. (2013). Complete starch hydrolysis by the synergistic action of amylase and glucoamylase: impact of calcium ions. *Bioprocess and biosystems engineering*, 36, 1555-1562.

72. Sahal, S., Khaturia, S., & Joshi, N. (2023). Application of Microbial Enzymes in Wastewater Treatment. *Genomics Approach to Bioremediation: Principles, Tools, and Emerging Technologies*, 209-227.
73. Savla, N., Pandit, S., Mathuriya, A. S., Gupta, P. K., Khanna, N., Babu, R. P., & Kumar, S. (2021). Recent advances in bioelectricity generation through the simultaneous valorization of lignocellulosic biomass and wastewater treatment in microbial fuel cell. *Sustainable Energy Technologies and Assessments*, 48, 101572.
74. Sandhya, S. (2010). Biodegradation of azo dyes under anaerobic condition: role of azoreductase. *Biodegradation of azo dyes*, 39-57.
75. Saraswat, R., Verma, V., Sistla, S., & Bhushan, I. (2017). Evaluation of alkali and thermotolerant lipase from an indigenous isolated *Bacillus* strain for detergent formulation. *Electronic Journal of Biotechnology*, 30, 33-38.
76. Schenk, G., Mateen, I., Ng, T. K., Pedroso, M. M., Mitić, N., Jafelicci Jr, M., ... & Ollis, D. L. (2016). Organophosphate-degrading metallohydrolases: Structure and function of potent catalysts for applications in bioremediation. *Coordination Chemistry Reviews*, 317, 122-131.
77. Sharma, S. (2012). Bioremediation: features, strategies and applications. *Asian Journal of Pharmacy and Life Science*, 2231, 4423.
78. Shekher, R., Sehgal, S., Kamthania, M., & Kumar, A. (2011). Laccase: microbial sources, production, purification, and potential biotechnological applications. *Enzyme research*, 2011.
79. Singh, B. (2014). Review on microbial carboxylesterase: general properties and role in organophosphate pesticides degradation. *Biochem Mol Biol*, 2, 1-6.
80. Singh, R., Kumar, M., Mittal, A., & Mehta, P. K. (2016). Microbial enzymes: industrial progress in 21st century. *3 Biotech*, 6, 1-15.
81. Siqueira, J. G. W., Rodrigues, C., de Souza Vandenberghe, L. P., Woiciechowski, A. L., & Soccol, C. R. (2020). Current advances in on-site cellulase production and application on lignocellulosic biomass conversion to biofuels: a review. *Biomass and Bioenergy*, 132, 105419.
82. Sjöblad, R. D., & Bollag, J. M. (2021). Oxidative coupling of aromatic compounds by enzymes from soil microorganisms. In *Soil biochemistry* (pp. 113-152). CRC Press.
83. Soares, P. R. S., Birolli, W. G., Ferreira, I. M., & Porto, A. L. M. (2021). Biodegradation pathway of the organophosphate pesticides chlorpyrifos, methyl parathion and profenofos by the marine-derived fungus *Aspergillus sydowii* CBMAI 935 and its potential for methylation reactions of phenolic compounds. *Marine Pollution Bulletin*, 166, 112185.
84. Sondhi, S., Sharma, P., George, N., Chauhan, P. S., Puri, N., & Gupta, N. (2015). An extracellular thermo-alkali-stable laccase from *Bacillus tequilensis* SN4, with a potential to bioleach softwood pulp. *3 Biotech*, 5, 175-185.
85. Tachibana, S., Naka, N., Kawai, F., & Yasuda, M. (2008). Purification and characterization of cytoplasmic NAD<sup>+</sup>-dependent polypropylene glycol dehydrogenase from *Stenotrophomonas maltophilia*. *FEMS microbiology letters*, 288(2), 266-272.
86. Toesch, M., Schober, M., & Faber, K. (2014). Microbial alkyl- and aryl-sulfatases: mechanism, occurrence, screening and stereoselectivities. *Applied microbiology and biotechnology*, 98, 1485-1496.
87. Wang, D., Li, A., Han, H., Liu, T., & Yang, Q. (2018a). A potent chitinase from *Bacillus subtilis* for the efficient bioconversion of chitin-containing wastes. *International journal of biological macromolecules*, 116, 863-868.
88. Wang, Y., Huang, J., Liang, X., Wei, M., Liang, F., Feng, D., ... & Zou, H. (2022). Production and waste treatment of polyesters: Application of bioresources and biotechniques. *Critical Reviews in Biotechnology*, 1-18.
89. Wang, Y., Feng, Y., Cao, X., Liu, Y., & Xue, S. (2018b). Insights into the molecular mechanism of dehalogenation catalyzed by D-2-haloacid dehalogenase from crystal structures. *Scientific Reports*, 8(1), 1-9.
90. Xue, F., Ya, X., Tong, Q., Xiu, Y., & Huang, H. (2018). Heterologous overexpression of *Pseudomonas umsongensis* halohydrin dehalogenase in *Escherichia coli* and its application in epoxide asymmetric ring opening reactions. *Process Biochemistry*, 75, 139-145.
91. Yadav, S., Yadav, P. K., Yadav, D., & Yadav, K. D. S. (2009). Pectin lyase: a review. *Process Biochemistry*, 44(1), 1-10.
92. Zanuso, E., Gomes, D. G., Ruiz, H. A., Teixeira, J. A., & Domingues, L. (2021). Enzyme immobilization as a strategy towards efficient and sustainable lignocellulosic biomass conversion into chemicals and biofuels: current status and perspectives. *Sustainable Energy & Fuels*, 5(17), 4233-4247.
93. Zhang, H., Han, L., & Dong, H. (2021). An insight to pretreatment, enzyme adsorption and enzymatic hydrolysis of lignocellulosic biomass: Experimental and modeling studies. *Renewable and sustainable energy reviews*, 140, 110758.
94. Zhao, S., Xu, W., Zhang, W., Wu, H., Guang, C., & Mu, W. (2021). In-depth biochemical identification of a novel methyl parathion hydrolase from *Azohydromonas australica* and its high effectiveness in the degradation of various organophosphorus pesticides. *Bioresource Technology*, 323, 124641.

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