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Review

# Synephrine and Its Derivative CpdA: Common and Specific Biological Effects

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**Abstract:** This review is focused on synephrine, the principal phytochemical found in bitter orange and other medicinal plants, and widely used as a dietary supplement for weight loss/body fat reduction. We examine different aspects of synephrine biology, delving into its established and potential molecular targets, as well as its mechanisms of action. We present an overview of the origin, chemical composition, receptors, and pharmacological properties of synephrine, including its anti-inflammatory and anti-cancer activity in various in vitro and animal models. Additionally, we conduct a comparative analysis of the molecular targets and effects of synephrine with those of its metabolite, selective glucocorticoid receptor agonist (SEGRA) Compound A (CpdA), which shares a similar chemical structure with synephrine. SEGRA, including CpdA, have been extensively studied as glucocorticoid receptor activators that have a better benefit/risk profile than glucocorticoids due to the reduced adverse effects. We discuss the potential of synephrine usage as a template for the synthesis of new generation of non-steroidal SEGRA. The review also provides insights into the safe pharmacological profile of synephrine.

**Keywords:** synephrine; glucocorticoids; selective glucocorticoid receptor activator (SEGRA); inflammation; cancer; CpdA; metabolic disorders; cardiovascular system; beta-adrenergic receptors

## 1. Introduction

This review provides a comparative analysis of two biologically active compounds that have been intensively studied in the past decades both as potential drugs.

Synephrine is found in the fruits of several trees from Rutaceae family, including Bitter orange (*Citrus aurantium*), or Seville orange, sour orange, Green orange, Zhi Shi and Kijitsu, as well as from some other citrus species such as Nova tangerines and Marrs sweet oranges [1]. Synephrine had been widely used in traditional Chinese medicine as an energy stimulant due to its beneficial effects on the cellular energetic [2]. Subsequently, it was introduced as a biologically active dietary supplement with thermogenic and weight-loss properties [3]. Since ephedrine-containing dietary supplements were banned in April 2004, ephedrine-free dietary supplements including synephrine have become widely available on the market [4]. The impact of synephrine on regulating the metabolic rate could be mainly explained by its molecular effects as a non-specific agonist of  $\beta$ -adrenergic receptors, the well-known regulators of lipolysis [5]. It provided the solid groundwork for using synephrine as a

safer analogue of ephedrine, which had been banned in several countries either as a drug or as a biologically active dietary supplement [6]. As a low-molecular-weight compound, synephrine may have multiple targets. Specifically, quantitative proteomics and bioinformatics analyses, as well as in vitro and in vivo experiments revealed synephrine to regulate several sets of genes involved in tumor development and inflammation [7–10]. Moreover, the similarity between the chemical structures of synephrine and several biologically active compounds makes it reasonable to explore the synephrine effects and their molecular mechanisms.

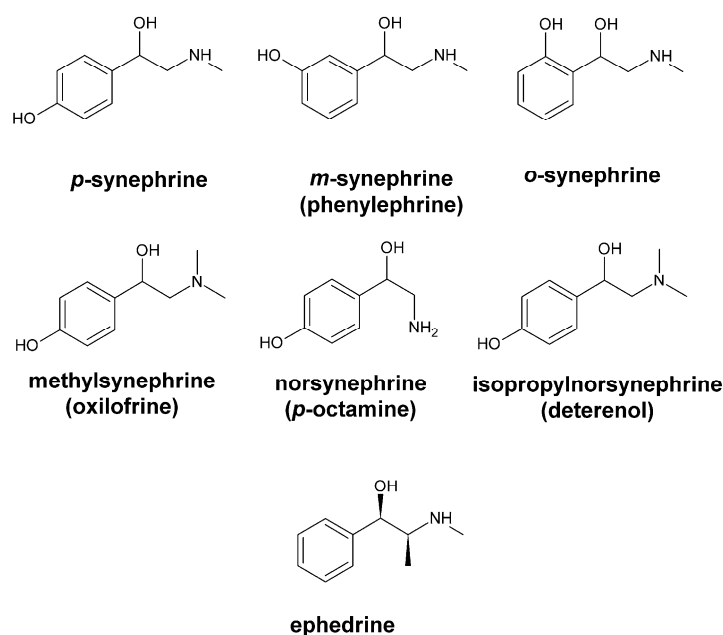
The non-steroidal selective glucocorticoid receptor agonist (SEGRA), the molecule of the natural origin, 2-(4-acetoxyphenyl)-2-chloro-N-methyl-ethylammonium chloride, also known as Compound A or CpdA, was described in the 1990s. CpdA binds the glucocorticoid receptor (GR) with the affinity comparable to that of classic glucocorticoids (GCs) [11]. However, CpdA only partially activates GR, mostly inducing therapeutically important transrepression (TR) function, negative protein-protein interaction of GR with a variety of transcription factors (TF) [12,13]. GR binding to TFs such as NF- $\kappa$ B and AP1 among others, suppresses their activity, underlying the anti-inflammatory and anti-cancer effects of GCs and CpdA [14–16]. The experimental evidence suggests that GR transactivation (TA) mediated via binding of activated GR to GC-responsive elements (GRE) in the gene promoters results in preferential transcriptional activation of the genes associated with metabolic and atrophic adverse effects of GCs [17,18]. The failure of CpdA to induce full-fledged GR activation has been linked to its non-steroidal structure and consequently its inability to promote formation of GR-GR homodimers necessary for GR binding to palindromic GREs and activation of target genes transcription [19,20]. Thus, CpdA has been extensively studied as an alternative to GCs for clinical applications. However, CpdA use in clinic is restricted by its chemical instability and decomposition to the intermediate metabolite aziridine, a well-known carcinogen [21,22]. Many attempts were made to find/generate another SEGRA by the molecular libraries screening, analysis of the components of anti-inflammatory natural extracts; directional chemical synthesis [23], or by virtual GR docking [24]. Nevertheless, this ~ 25-year effort by pharmaceutical companies and academia to identify or synthesize clinically promising, has not brought the expected results.

Synephrine is the final product of CpdA metabolism and at the same time it serves as precursor for CpdA synthesis [11,25]. Given high biological activity of synephrine, the analysis of common and specific targets of CpdA and synephrine becomes particularly relevant. Here we discuss the biological activity of synephrine, its source, chemical and pharmacological features, target receptors, anti-inflammatory, anti-cancer, and other effects as well as synephrine and its possible derivatives as potential clinical benefits in comparison with the effects of GR ligands, the selective glucocorticoid receptor agonist CpdA, which is structurally similar to synephrine.

## 2. Source, Chemical and Pharmacological Features of Synephrine

Synephrine is a phenethylamine alkaloid which is 4-(2-aminoethyl)phenol substituted with a hydroxyl group and a methyl group at the amino nitrogen [4]. There are three different positional isomers of synephrine (ortho/-o-, para/-p- and meta/-m-) [5,8]. Their chemical structure is similar to ephedrine, the phenylpropylamine derivative that does not contain a substituted hydroxyl group in the phenyl (benzene) ring (Figure 1) [26].

Importantly, the presence of a hydroxyl group in the synephrine molecule as well as the lack of a methyl group on the side chain modifies its stereochemistry and impairs the interaction with proteins. The latter results in an altered pharmacokinetic profile, allowing synephrine to permeate the blood-brain barrier. For example, the lipid solubility of p-synephrine is much lower compared to ephedrine, resulting in a decreased transport of p-synephrine to the central nervous system (CNS) [27].



**Figure 1.** Structures of the synephrine isomers, derivatives and ephedrine.

The p-synephrine isomer is widely used as a dietary supplement for weight loss since it promotes fat oxidation [1,6]. The safety of p-synephrine to be used in thermogenic dietary supplements was demonstrated in multiple studies [7,26,28,29].

m-Synephrine, also called phenylephrine, is considered to be the most potent adrenergic agonist of synephrine at  $\alpha$ 1-adrenoreceptors ( $\alpha$ 1-AR) as compared to other isomers [1]. The o-synephrine isomer is not found in dietary supplements, and no pharmacological effect of it on humans was revealed [30]. Structural and stereochemical differences between p-synephrine and m-synephrine lead to different binding characteristics in relation to AR. For example, weak binding of p-synephrine to AR explains its lower deleterious cardiovascular effects as compared to m-synephrine [31]. In addition, each positional isomer can also be found in two enantiomeric forms varying in pharmacological and physiological activity [1].

Another component of a bitter orange extract with a similar structure is norsynephrine (p-octopamine), an octopamine receptor ligand, a neurotransmitter, and an adrenergic receptor ligand with low affinity [32]. A number of synephrine derivatives (such as isopropyl-norsynephrine and methylsynephrine) are thermogenic agents [33,34].

Synephrine is an optically active compound that exists as a mixture of R- and S-isomers. The R-enantiomer is the main form in in Citrus aurantium (92-96%) [35–38]. There are several lines of evidence that R-synephrine is a more potent agonist in relation to vascular  $\alpha$ -,  $\beta$ 1-, and  $\beta$ 2-AR [39]. At the same time, S-enantiomer has been suggested to block noradrenaline reuptake [40], which may elevate blood pressure and heart rate. In line with these findings, only S-synephrine has been shown to possess antidepressant-like activity [40]. Of note, CpdA enantiomers slightly differ in their biological effects from a racemic mixture. In our previous studies, we synthesized CpdA enantiomers, and in our in vitro studies S-CpdA revealed to exert stronger proapoptotic effects on leukemia cells as compared to R-CpdA [41].

### 3. Mechanism of Synephrine Effects

Synephrine is an adrenergic receptor agonist that acts predominantly through the  $\beta$ -adrenergic receptors ( $\beta$ -ARs).  $\beta$ -ARs are a subfamily of G protein-coupled receptors (GPCRs) that are expressed by most cell types in humans [9]. This subfamily consists of three members,  $\beta$ 1,  $\beta$ 2, and  $\beta$ 3-AR, the targets of endogenous catecholamines (epinephrine and norepinephrine) [42].  $\beta$ -Agonists promote receptor-mediated G-protein activation of the adenylyl cyclase enzyme and increase cyclic adenosine monophosphate (cAMP) production. Regardless of the subtype differences, all activated  $\beta$ -ARs are

typically phosphorylated by regulatory kinases such as GPCR kinases (GRKs), and signaling is then terminated by interaction with  $\beta$ -arrestin. This process is called desensitization [42,43].

p-Syneprhine and m-syneprhine differ in their biological activity due to certain stereochemical differences as separate isomers bind to receptors with different affinity [31]. Therefore, p-syneprhine has a lower ability to stimulate  $\alpha$ -1,  $\alpha$ -2,  $\beta$ -1, and  $\beta$ -2-AR, compared to the typical sympathomimetics. Moreover, p-syneprhine predominantly binds to  $\alpha$ 1-AR, with lower affinity to  $\alpha$ 2-AR and thus a much lower potency relative to  $\beta$ -AR, regardless of the subtype [44]. In vitro studies described above show that the biological effects of p-syneprhine are mediated by various mechanisms different from selective binding to specific ARs and with a limited contribution of  $\alpha$ -,  $\beta$ -1 and  $\beta$ -2, and  $\beta$ 3-AR. Analysis of the chemical structure and pharmacological action of syneprhine indicates that its molecular action is similar and involves interference with the 3',5'-cyclic adenosine monophosphate/protein kinase A (cAMP/PKA) signaling. This pathway is triggered by sympathetic activation to restore cellular homeostasis via stimulating glucose uptake, lipolysis, fatty acid oxidation, mitochondrial biogenesis, and cell proliferation [8,45].

Several studies show that p-syneprhine can bind to and activate  $\beta$ 3-AR, contributing to thermogenesis, lipolysis, glucose, and cholesterol metabolism, and possibly reducing food intake.  $\beta$ 3-AR is a G protein-coupled receptor functionally linked to Gas and G $\beta\gamma$  subunits. Stimulating  $\beta$ 3-AR increases intracellular cAMP levels via adenylyl cyclase activation. Consequently, cAMP triggers two major signaling cascades in adipocytes: protein kinase A (PKA) signaling as well as Exchange Protein directly activated by cAMP/Ras-related protein (EPAC/RAP) pathway. PKA catalyzes the phosphorylation of hormone-sensitive lipase (HSL) and perilipin stimulating lipolysis [46,47] and induces thermogenic gene transcription through activating transcription factor 2/cAMP-responsive element binding protein (ATF-2/CREB) TA [48–51]. EPAC function as guanine nucleotide exchange factors (GEFs) for the Ras-like small GTPases, or Ras proximate proteins, Rap1 and Rap2. EPAC acts synergistically with the cAMP-dependent protein kinase PKA via Rap [52,53].

In invertebrates, p-syneprhine binds to octopamine receptor subtypes, homologous to vertebrate adrenergic receptors. However, the affinity of p-syneprhine, m-syneprhine, and norepinephrine binding to octopamine receptors varies considerably and has a non-linear correlation with their AR binding characteristics. Therefore, p-syneprhine affinity to octopamine receptors cannot be extrapolated to human and other vertebrate ARs [54]. There is some evidence that syneprhine also has weak affinity to 5-hydroxytryptamine (serotonin) receptors (5-HT2A and 5-HT1D) and may interact with trace amine-associated receptor 1 (TAAR1) [44]. TAAR1 is essential for regulating neurotransmission in dopamine, norepinephrine and serotonin neurons in the CNS and mediates the mechanisms involved in mood changes, attention, memory, and anxiety disorders. Moreover, TAAR1 has recently been described to play mediate or modulate immune dysregulation [55,56]. TAARs represent another mechanism underlying p-syneprhine action either as a neurotransmitter precursor or as a neuromodulator [44].

B-agonists have also been reported to affect GR function. Forskolin, which boosts intracellular cAMP production thus mimicking  $\beta$ 2-AR activation, elevates GR expression in rat hepatocellular carcinoma cells and potentiates the production of dexamethasone-induced neurotensin, regulating analgesia, thermoregulation, reward, arousal, blood pressure, and ultimately modulating feeding behaviors and body weight [57]. Forskolin can also antagonize the negative autoregulation of GRs induced by dexamethasone. These effects indirectly suggest that  $\beta$ -receptor agonists may enhance GR function by increasing the intracellular cAMP levels [58].

#### 4. Chemical and Pharmacological Features of CpdA and Its Target Receptors

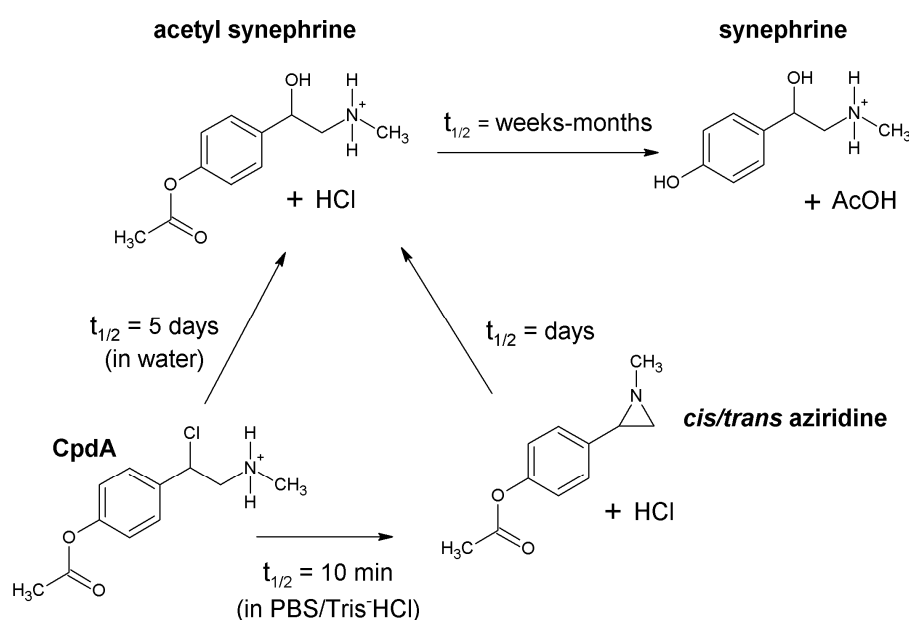
The biological effects of both GCs and the selective glucocorticoid receptor agonist CpdA are mediated by GR, a transcription factor from the superfamily of nuclear hormone receptors [59]. GR regulates protein, lipid and carbohydrate metabolism, the cell cycle, immune, stress and inflammatory responses, affecting thousands of genes [60]. The effects of receptor activation upon binding to GC or CpdA are mediated by DNA-independent transrepression (TR), which underlies its therapeutic, anti-inflammatory, and anti-cancer activities. Upon transactivation (TA), GR dimers



bind to GC-responsive elements (GRE), followed by a transcriptional activation of the genes, associated with an elevated risk of adverse effects [17]. The adverse effects including diabetes mellitus, peptic ulcer disease, Cushing's syndrome, osteoporosis, hyperglycemia, skin and muscle atrophy, psychosis, glaucoma, and many other complications are mainly induced by TA and are partially irreversible [18,61].

CpdA, the well-characterized SEGRA, is a synthetic analog of a natural compound found in *Salsola tuberculatiformis* Botschantzev and exerts GR-dependent anti-inflammatory and anti-cancer effects [16,41,62,63]. CpdA appears to be a promising alternative to GCs with fewer side effects [18,41,51,62,64–66].

In water solution CpdA decomposes directly to acetyl synephrine in about 5 days, followed by ester hydrolysis to synephrine after a few weeks or months. In contrast, in buffer solutions, CpdA forms a mixture of optical isomers of aziridine within a few minutes. With a half-life ( $t_{1/2}$ ) of several days, aziridines are subsequently hydrolyzed to acetyl synephrine and then to synephrine [1,22] (Figure 2). It remains to be elucidated whether synephrine and its derivatives could exhibit the SEGRA activity inducing only GR TR without TA-related adverse effects.



**Figure 2.** Decomposition of CpdA to aziridine derivatives and synephrine depends on the solvent and pH [22].

It is well known that CpdA acts as a GR ligand. It binds to GR with high affinity causing moderate GR nuclear translocation, but no GR dimerization necessary for GR binding to palindromic GRE sequences in GR target gene promoters/enhancers [67]. Furthermore, CpdA does not induce GR phosphorylation at Ser211 and hence does not enhance GR TA activity [68]. In contrast, CpdA and classic GCs have remarkably similar TR profiles, suppressing the activity of many pro-proliferative and anti-apoptotic transcription factors: NF- $\kappa$ B, AP-1, Ets-1, Elk-1, SRF, and NFATc [69]. Apart from its SEGRA function, CpdA acts as an antagonist of the androgen receptor. Although CpdA has lower affinity for the androgen receptor compared to active androgens, CpdA induces considerable androgen receptor nuclear translocation, suppresses the interaction between the NH(2)- and COOH-terminal domains of the androgen receptor that critical for its function, and inhibits both constitutive and dihydrotestosterone (DHT)-inducible androgen receptor transcription activity [62,70]. CpdA does not significantly affect the activity of other steroid hormone receptors such as mineralocorticoid, progesterone, and estrogen receptors [15].

The beta-adrenergic system plays a crucial role in the body's response to stress and is involved in various physiological reactions to stress, including increasing cardiovascular output,

bronchodilation, and mobilization of energy resources via increased breakdown of glycogen into glucose (glycogenolysis) and via lipolysis -breakdown of stored fat into free fatty acids which could be used as an energy source. In turn, GCs that are often referred as stress hormones also mobilize energy resources and increase glucose blood levels via induction of protein catabolism and lipolysis needed for gluconeogenesis. This overlap in functions of GCs and beta-adrenergic agonists inspired efforts to identify the potential cross-talk between down-stream signaling mediated by GR and  $\beta$ -AR and even at the level of ligand cross-binding.

A crosstalk between GR and  $\beta$ -AR has been described in the literature based on two mechanisms. GCs regulate  $\beta$ -AR binding to G proteins, followed by adenylyl cyclase activation. There are several phosphorylation pathways, including  $\beta$ -adrenergic kinase ( $\beta$ -ARK), and they represent the first step in the development of drug tolerance. The impact of GCs helps restore the function of the receptor, returning  $\beta$ -AR to a sensitized state. GCs can also enhance the suppressed  $\beta$ -AR function following the chronic agonist exposure. Reduced  $\beta$ -AR function is characterized by the internalization and degradation of  $\beta$ -AR which require their rapid resynthesis. Binding of the activated GR to GRE in the promoter region of the  $\beta$  receptor gene increases the gene transcription rate and hence the protein level [58]. Synephrine effects on GR remain unknown.

On the other hand, it is well documented that in asthma, the clinical efficacy of inhaled GCs is enhanced by  $\beta_2$ -adrenoceptor agonists, and the mechanisms underlying their effects on GR function were extensively studied in bronchial epithelial cells (pHBECs) and airway smooth muscle cells [71,72]. Surprisingly, it was shown that GR-mediated transcription could be significantly increased by  $\beta$ -agonists without changes in GR phosphorylation, nuclear translocation and increased loading on GREs as assayed by ChIP [73]. As many GR-target genes are independently induced by  $\beta$ -agonists, gene-specific control by GR- and  $\beta$ -agonist-activated transcription factors may explain the increased activation of GR in the presence of GCs and  $\beta$ -agonists

## 5. Overlapping Targets and Activities of Synephrine, SEGRA and Glucocorticoids in Pathological Conditions

### 5.1. Anti-Inflammatory Effects

The anti-inflammatory effects of synephrine have been studied in various models of inflammation, but a detailed mechanism of action has not been fully established. In the lipopolysaccharide (LPS)-stimulated RAW264.7 murine macrophages, p-synephrine inhibited the production of pro-inflammatory cytokines and nitric oxide. This effect was associated with the suppressed p38 mitogen-activated protein kinase (p38 MAPK) and nuclear factor kappa B (NF- $\kappa$ B) signaling pathways, and was mediated by  $\beta$ -AR [9]. In the acute lung injury model, the anti-inflammatory effects of p-synephrine were accompanied by a decreased activity of pro-inflammatory tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ) and interleukin-6 (IL-6) genes, the increased activity of interleukin-10 (IL-10) and superoxide dismutase (SOD), as well as suppressed reactive oxygen species (ROS) generation, reduced myeloperoxidase activity, and the inhibited NF- $\kappa$ B phosphorylation [74,75]. Moreover, synephrine reduced the degradation of the NF- $\kappa$ B inhibitor, nuclear factor of kappa light polypeptide gene enhancer in B-cells inhibitor, alpha (I $\kappa$ B $\alpha$ ) [75]. Under normal physiological conditions, NF- $\kappa$ B is present in the cytoplasm in an inactive form in a complex with I $\kappa$ B $\alpha$ . Intranasal administration of LPS leads to activation of NF- $\kappa$ B and degradation of I $\kappa$ B $\alpha$  in the lung tissue. However, pre-treatment with p-synephrine prevented these processes in vivo [75,76].

In addition, p-synephrine significantly inhibited the proliferation of neutrophils and macrophages in the bronchoalveolar lavage fluid, leading to the anti-inflammatory effect [75]. In the murine model of systemic inflammatory response, p-synephrine inhibits serum levels of pro-inflammatory cytokines and reduces the inflammation [9]. In studies harnessing NIH/3T3 murine fibroblasts and normal human fibroblasts, p-synephrine dose-dependently inhibited IL-4-induced expression of eotaxin-1 via suppression of a signal transducer and activator of transcription (STAT6). STAT6 is critical for activating cytokine gene expression and cytokine signaling in immune cells and target tissue cells. As eotaxin-1 is a chemoattractant and mediator of eosinophilic inflammation development, p-synephrine also inhibited eosinophil recruitment [26].

Inflammation is strongly associated with oxidative stress. The antioxidant properties of p-synephrine were shown in the murine model of diabetes mellitus developed by alloxan injection. p-Synephrine significantly increased the activity of SOD, catalase (CAT), and glutathione (GSH) and reduced the level of malondialdehyde (MDA) in the blood serum of diabetic mice. In addition, p-synephrine administration inhibited kidney inflammation by downregulating TNF- $\alpha$ , IL-6, and IL-1 $\beta$  gene expression levels. The mechanism of anti-inflammatory effects may include the suppression of NF- $\kappa$ B activation and MAPK phosphorylation [7].

The structural similarity of synephrine and CpdA allows proposing generalized mechanisms of anti-inflammatory effects [6,9]. CpdA exerts its anti-inflammatory function via reducing p65 DNA-binding activity in vivo as well as via inhibiting NF- $\kappa$ B transactivation potential [14]. Furthermore, CpdA is partially responsible for the GC-dependent inhibition of inflammation by inhibiting the activator protein-1 (AP-1) involved in the transcription of pro-inflammatory cytokine genes [17,48,50,77,78], suppressing MAPK [79,80], and STAT signaling resulting in the downregulated production of cytokine including IL-1 $\beta$ , IL-2, IL-6, and TNF [58,79], a reduction in the phospholipase A<sub>2</sub> activity, as well as a decrease in cyclooxygenase-2 (COX-2) expression [81,82] (Table 1).

Moreover, the regular use of  $\beta$ 2-agonists has been suggested to cause adverse effects on asthma control due to the crosstalk between the cAMP responsive element binding (CREB) proteins and GRs [83]. The cross-talk between GR and  $\beta$ -AR ligands may rely on the cAMP-dependent activation of the nuclear transcription factor CREB, which further binds to cAMP response elements (CREs) in the promoters of target genes. This involves competitive binding of GRs and protein cofactors, such as CREB or related P300, required for activating transcription factor response elements. Furthermore, there is evidence for mutual inhibition of nuclear transcription factors by  $\beta$ -AR agonists and GCs due to the interaction between GR and CREB induced by the high concentrations of  $\beta$ 2-AR agonists [84] (Table 1).

**Table 1.** Overlapping targets and activities of synephrine and CpdA in pathological conditions.

Possible molecular mechanisms	Effects	Model description	Substance
Anti-inflammatory effects			
In vitro			
Downregulated p38 MAPK and NF-κB signaling pathway; Inhibited expression of pro-inflammatory cytokines: IL-8, IL-6, TNF-α [9]		LPS-stimulated RAW264.7 cells	Synephrine
Inhibited IL-4-induced expression of eotaxin-1 via suppression of STAT6 [26]		NIH/3T3 mouse fibroblasts	
Attenuated expression of TNF-α, iNOS, and IL-1, but increased expression of anti-inflammatory IL-10; Induced macrophage differentiation towards M2 anti-inflammatory phenotype [85]		Immortalized murine macrophage cell line RAW 264.7	CpdA
Inhibited NF-κB activity and IKK phosphorylation; Induced IκB-α accumulation; decreased IL-1β expression [86,87]		Synovial fibroblasts from patients with rheumatoid arthritis	
In vivo			
Reduced TNF-α, IL-6 and increased IL-10 activity; elevated SOD activity and suppressed ROS generation; Reduced MPO activity; attenuated histological changes; Inhibited NF-κB phosphorylation and IκB degradation [75]	Inhibited pulmonary edema; Reduced histological changes	LPS-induced ALI, mice	Synephrine



Reduced serum levels of proinflammatory cytokines [9]	Improved survival rate	LPS-induced systemic inflammatory response syndrome, a mouse model	
Increased activity of SOD, CAT, and GSH; reduced MDA content; Downregulation of TNF- $\alpha$ , IL-6, and IL-1 $\beta$ gene expression levels [7]		The mouse model of diabetes mellitus	
Suppressed NF- $\kappa$ B activity and nuclear translocation; Inhibited STAT6 activity and nuclear translocation; Reduced expression of Th2-cytokines: IL-4, IL-5, and IL-13 [88]	Reduced inflammatory cell infiltration in lungs, cytokine production, mucus and Ig production; Reduced development of airway hyperresponsiveness	Ovalbumin-induced Th2-driven asthma model	
Decreased NF- $\kappa$ B activity and downregulation of pro-inflammatory cytokines: IL-8, IL-6, and E-selectin [14]	Decreased swelling	Zymosan-induced inflamed paw	CpdA
Inhibited pro-inflammatory cytokines: IL-1 $\beta$ , TNF- $\alpha$ , IL-6; Upregulation of anti-inflammatory cytokines: IL-4 and IL-10 [89]	Protected from the development of diabetes; Modulated peripheral immune response (switching from Th1/Th17 towards anti-inflammatory T-regulatory/ Treg response)	Streptozotocin-induced model of type 1 diabetes	
<b>Anti-cancer effects</b>			
<b>In vitro</b>			
Reduced expression of p-AKT, AKT, p-ERK and ERK [90]	Suppressed proliferation	Esophageal squamous cell carcinoma	
Increased ROS formation; Increased activity of the antioxidant molecules glutathione and catalase [8]	Revealed no cytotoxic effect	Human colon adenocarcinoma (Caco-2) cells	
Induced DNA damage and apoptosis; hyperproduction of intracellular ROS [91,92]		Human hepatocellular carcinoma (HepG2)	Syneprhine
Increased expression of Bax and p53 at the mRNA and protein levels; Suppressed PI3K/AKT/mTOR signaling pathway [10]		Lung cancer cells (H460)	
Inhibited several transcription factors, including nuclear factor kappa B (NF- $\kappa$ B), AP-1, Ets-1, Elk-1; Induced caspase-dependent apoptosis [69]	Decreased growth	Highly malignant androgen-independent DU145 and PC3 cells	
	Strongly inhibited growth and viability [63]	CEM T-cell acute lymphoblastic leukemia; K562 chronic myeloid leukemia cells	CpdA
Inhibited NF- $\kappa$ B signaling [16]		Murine L929sa fibrosarcoma cells	
Increased GR-GR dimerization; a decreased number of MR-GR heterodimers [93]		Rat pheochromocytoma PC12 cells	
<b>In vivo</b>			
Reduced level of glucose metabolism genes, G6Pase and PEPCK [90]	Reduced glucose production	Human ESCC xenografts in nude mice	Syneprhine
Induced apoptosis in cancer cells via the upregulation of pro-apoptotic members		P2 rat pups	CpdA

of the B-cell lymphoma (Bcl-2) family [93]			
Metabolic and anti-diabetic effects			
In vitro			
Reduced level of glucose metabolism genes, G6Pase and PEPCK [94]	Reduced glucose production	Rat liver cells (H4IIE)	Synephrine
Acted as partial GR antagonist [95]		Immortalized murine keratinocytes	CpdA
In vivo			
Suppressed gene expression levels of TNF- $\alpha$ , IL-6, IL-1 $\beta$ ; Activated enzymes of the antioxidant system; Inhibited oxidative stress via suppressing the NF- $\kappa$ B and MAPK pathways [7]	Prevented alloxan-induced changes in body weight, organ parameters, serum uric acid and serum creatinine, and improved lipid profile	Alloxan-induced diabetes mellitus in mice	Synephrine
Increased metabolic rate via agonistic activity on $\beta$ -3 adrenoreceptors; Exhibited hypoglycemic and insulin-stimulating properties; Stimulated translocation of the glucose-4 transporter protein [96]	Decreased blood glucose levels; Increased insulin levels; Decreased insulin resistance	Gliclazide-treated rats and rabbits	
Did not induce gluconeogenesis enzymes in the liver [87]	Did not increase in blood glucose levels; Did not induce hyperinsulinemia	Collagen-induced arthritis in mice	CpdA
Inhibited endogenous GR signaling due to CpdA antagonistic effect on GR activity [95]	Revealed anti-inflammatory and atrophogenic effects	Model of contact dermatitis in mice	

5.2. Anti-Cancer Effects

The anti-cancer effects of synephrine are largely unknown. However, several in vitro studies on synephrine described its anti-cancer activity. Synephrine hydrochloride inhibits the proliferation of esophageal squamous cell carcinoma (ESCC) cells in a dose-dependent manner, significantly suppresses cell migration and invasion, and reduces the growth of human ESCC xenografts in nude mice. In addition, synergistic anti-cancer effects of synephrine with 5-FU, one of the commonly used chemotherapy drugs, are demonstrated in ESCC cells [90].

As described in the pertinent literature, synephrine is involved in the regulation of AKT and extracellular signal-regulated kinase (ERK) signaling. The levels of expression of p-AKT, AKT, p-ERK, and ERK in ESCC cells treated with synephrine are reduced in a dose-dependent manner. Synephrine downregulates galectin-3, an activator of the AKT and ERK signaling pathways which plays a crucial role in cell survival and malignization [97]. Therefore, the studies highlight the therapeutic potential of synephrine as an agent that inactivates the AKT and ERK signaling that have been previously described as targets for anti-cancer therapy [90].

Recent studies in lung cancer cells H460 demonstrate the ability of synephrine to reduce cell viability by increasing the expression of Bax and p53. In addition, synephrine induces a significant decrease in the mRNA and protein levels of PI3K, AKT, and mTOR in H460 cells, followed by apoptosis induction [10].

Meanwhile, synephrine has no significant cytotoxic effect on immortalized normal esophageal epithelial cells and does not alter the levels of serum alanine transaminase (ALT) and aspartate transaminase (AST) in nude mice [90]. These data indicate that synephrine may provide a safer therapeutic profile.

However, a recent study shows that at a concentration of 25–5000  $\mu$ M, p-synephrine reveals no cytotoxic effect in human colon adenocarcinoma (Caco-2) cells. Furthermore, at the reduced concentration of 2–200  $\mu$ M, synephrine stimulates ROS generation followed by the enhanced activity

of the antioxidant enzymes GSH and CAT. In addition, synephrine activates MAPK1, the guanine nucleotide binding protein, the alpha-stimulating activity polypeptide (GNAS), the protein kinase cAMP-activated catalytic subunit alpha (PRKACA), and the AKT signaling pathway in Caco-2 cells [8]. More specifically, synephrine increases the expression of the PRKACA and the protein kinase cAMP-dependent type II regulatory subunit alpha (PRKAR2A) genes. These genes encode the catalytic subunits of PKA, the regulator of MAPK-dependent cell proliferation and differentiation [98,99]. In addition, synephrine significantly increases the expression of AKT1 (serine/threonine kinase 1) gene encoding the pro-proliferative signaling regulator. Furthermore, synephrine triggers GNAS upregulation with the subsequent production of the alpha subunits of the G protein, a key component in adenylate cyclase signaling and GPCR-dependent cellular responses [8]. These signaling pathways engage multiple targets for compounds including ephedrine, norepinephrine and synephrine [100].

Studies in human hepatocellular carcinoma (HepG2) cell line demonstrate that the combination of synephrine and caffeine induces DNA damage and apoptosis at the concentrations of 3:60, 3:90, and 3:600  $\mu$ M respectively. These results are supported by the upregulation of the genes associated with DNA repair and apoptosis, specifically caspase-9 and downregulation of the genes associated with cell cycle control [91]. However, treating HepG2 cells with synephrine causes the overproduction of intracellular ROS [92]. This could be related to a rapid transformation of synephrine by mitochondrial monoamine oxidases (MAOs) predominantly in liver cells as well as MAO-dependent ROS production [101,102]. The second possible mechanism is mediated by the stimulation of AMPKs [103], which act as the central regulators of energy metabolism [104]. Their excessive activation leads to an impaired electron transport in the respiratory chain and a subsequent mitochondrial overproduction of ROS [105]. Nevertheless, the synephrine-induced ROS overproduction produces a cytostatic effect but does not cause significant DNA damage and does not induce chromosomal instability in HepG2 cells in vitro. These results indicate that synephrine causes acute oxidative stress that is however insufficient to sustain ROS overproduction over a period of time [91].

As mentioned above, synephrine is a stable hydrolysis product of CpdA [26], the SEGRA with well-known anti-cancer properties. The anti-cancer effects of both CpdA and GCs are partially mediated by apoptosis induction in cancer cells via the following processes: up-regulation of the pro-apoptotic members of the B-cell lymphoma (Bcl-2) family, such as Bim, Bid and Bad, and suppression of the anti-apoptotic members, such as Bcl-2, Mcl-1 and Bcl-xL [93,106,107], as well as inhibition of pro-proliferative signaling including AP-1 and NF- $\kappa$ B [16]. GCs (rather than CpdA) can also inhibit proliferation by suppressing c-MYC [108,109]. Furthermore, GR regulates the expression of multiple miRNAs, and, in particular, suppression of the miR-17-92 cluster correlates with apoptosis [110]. In contrast to CpdA, GCs inhibit cell migration/invasion in the in vitro models through a roster of mechanisms including downregulation of the Ras homolog family member A (RhoA) [111], matrix metalloproteinases 2 and 9 (MMP2,9), and IL-6 [112], or inducing E-cadherin [113], which inhibits angiogenesis by suppressing the pro-angiogenic factors, including IL-8 and vascular endothelial growth factor (VEGF) [114] (Table 1).

### 5.3. The Effects on Diabetes Mellitus and Obesity

The anti-diabetic effect of p-synephrine has been described in literature in nuance. Therefore, p-synephrine reduces glucose production by rat hepatocytes H4IIE in vitro, as well as the expression of glucose metabolism genes, glucose-6-phosphatase (G6Pase) and phosphoenolpyruvate carboxykinase (PEPCK) [115], at concentrations 1-100  $\mu$ M in a dose-dependent manner. Importantly, p-synephrine inhibits the activity of  $\alpha$ -amylase,  $\alpha$ -glycosidase, acetylcholinesterase, butyrylcholinesterase, and carbonic anhydrase [44]. Compounds with the described properties are considered for potential use for reducing postprandial blood glucose levels [115].

In the murine model of alloxan-induced diabetes mellitus, p-synephrine prevents alloxan-induced changes in body weight, organ parameters, serum uric acid, and serum creatinine levels as well as improves the lipid profile. The described effects are associated with the activation of enzymes

of the antioxidant system, such as SOD and CAT, the inhibition of NF- $\kappa$ B and MAPK signaling, as well as changes in GSH content [7] (Table 1).

Importantly, the effect of p-syneprine on the pharmacodynamics and pharmacokinetics of gliclazide, a hypoglycemic agent from the group of sulfonylurea derivatives, has been described. The repeated administration of p-syneprine in the presence of gliclazide results in a significant decrease in blood glucose levels as well as in an increase in insulin levels and activation of pancreatic  $\beta$ -cell function in rats and rabbits [96]. The observed changes can be explained by the agonistic activity of syneprine on  $\beta$ 3-AR, leading to an increase in the metabolic rate as well as by syneprine ability to exhibit hypoglycemic and insulin-stimulating properties. p-Syneprine was shown to stimulate the translocation of the glucose-4 transporter protein from the cytoplasm to the cytoplasmic membrane, leading to a decrease in insulin resistance [116].

Furthermore, syneprine modulates obesity development and lipid metabolism disorders and is widely used as a component of weight loss supplements [117]. Numerous studies revealed the favorable therapeutic profile of p-syneprine: (1) an increase of resting metabolic rate and energy expenditure; (2) inhibition of glucose production [94]; (3) thermogenic activity [118]; (4) influence on enzymes activity [115,119]; (5) lipolytic activity; (6) catabolic activity; (7) the influence on the differentiation of beige adipocytes [44].

Beige adipocytes are known to play an important role in increasing the energy expenditure through differentiation. p-Syneprine was found to be an active compound that increases the level of the uncoupling protein 1 (UCP1) mRNA in stromal vascular fraction (SVF) cells cultured in the beige adipocyte differentiation medium. p-Syneprine induces morphological changes specific to beige adipocytes in a dose-dependent manner. Similar effects were also observed in stromal vascular fraction cells obtained from leptin receptor-deficient db/db mice. These effects are mainly mediated by the activation of  $\beta$ 3-AdR and partially insulin signaling, leading to the stimulation of beige adipocyte differentiation, UCP1 expression and an increase in mTORC1 activity. In line with these findings, the combination of p-syneprine and espidulin, a flavone of natural origin, significantly inhibited expression of the genes encoding adipogenic proteins including MAPKs (ERK, ERK, JNK, and P38), C/EBP $\alpha$ , C/EBP $\beta$ , PPAR $\gamma$ , and GR [120]. In addition, p-syneprine regulates food intake and energy homeostasis exerting effects on the central nervous system. Specifically, it activates Neuromedin U2 receptor (NMU2R) in a dose-dependent manner [121]. NMUR2 is expressed in the hypothalamic regions of the brain and is involved in regulating energy balance, food intake, nociception, and stress [120]. All data presented above demonstrate the potential of p-syneprine for preventing and treating obesity and related diseases [100].

Considering syneprine as an alternative to GR ligands, it is important to highlight the adverse effects of GCs on metabolic processes. The metabolic effects of GCs are associated with physiological mechanisms of peripheral insulin resistance, hyperglycemia, and dyslipidemia. In contrast, CpdA, a non-steroidal SEGRA, does not induce the expression of genes associated with metabolism and metabolic disorders [61,62,122,123]. Specifically, CpdA does not elevate hepatic gluconeogenesis, adipocyte lipolysis and proteolysis, an increase in the circulation of free fatty acids and lipid accumulation in skeletal muscles and liver, which leads to the development of insulin resistance in these tissues [124,125]. Direct GC-related insulin release inhibition is demonstrated in the *in vivo* study on transgenic mice overexpressing GR in  $\beta$ -cells [126]. The mechanisms of GC-dependent inhibition of insulin release in  $\beta$ -cells are likely to involve the changes in the expression of TA-related subsets of genes important for glucose sensitivity and insulin secretion [122,127]. In contrast to GC, CpdA does not induce the stress-induced mTOR inhibitor DDIT4, a major molecular target of GC adverse effects including atrophic and metabolic changes [123,128–131]. Moreover, it has also been demonstrated that the targeted genetic deletion of DDIT4, treatment with SEGRA such as ZK245186/mapracorate or with the inhibitors of DDIT4 expression shift the function of GC-activated GR towards TR [123,129–131].

#### 5.4. The Effect on the Cardiovascular System

Sympathomimetics vary widely in their ability to activate or inhibit different ARs. As it was previously described, p-synephrine is chemically similar to sympathomimetic agents such as ephedrine. However, p-synephrine does not cause an increase in blood pressure or tachycardia, since it has a lower affinity for  $\alpha_1$  and  $\alpha_2$ -AR, as well as  $\beta_1$  and  $\beta_2$ -AR, compared with classic sympathomimetics [118].

In a recent placebo-controlled, double-blind study, it was shown that, p-synephrin dramatically reduces the diastolic blood pressure and the mean arterial pressure when a person was in a quite sitting position, while p-synephrine did not affect the systolic blood pressure or heart rate. Adding p-synephrine to caffeine also did not increase the systolic blood pressure or a heart rate. This indicates that p-synephrine does not affect the cardiovascular system [132].

Another double-blind, placebo-controlled crossover study in 18 healthy subjects showed that p-synephrine does not lead to any significant changes in electrocardiograms, heart rate, systolic blood pressure, blood chemistry, or blood cell count in 30 min – 8 h and thereafter within 15 days after the treatment [28]. Furthermore, the results of both published and unpublished clinical trials, involving approximately 360 subjects, show that p-synephrine alone or in combination with caffeine neither appears to cause significant side effects on the cardiovascular system nor poses a risk to human health when it is used in doses normally taken orally [54,133].

Importantly, GCs-induced side effects on the cardiovascular system are mediated by the interaction with AR. GCs increase  $\beta$ -adrenergic reactivity [83]. In lung cells, corticosteroids could increase the  $\beta_2$ -receptor numbers [134]. ARs such as  $\alpha$ -AR and  $\beta$ -AR are involved in the regulation of cardiovascular functions.  $\alpha$ -AR activation leads to an elevation in the blood pressure and heart rate, whereas  $\beta$ -AR activation increases cardiovascular function and oxygen demand. CpdA effects on the cardiovascular system have not yet been described in pertinent literature.

## 6. Conclusions

In this review, we have described a wide range of biological effects of synephrine, its identified and potential molecular targets and possible mechanisms of action. In addition, we compared the molecular targets and effects of synephrine, the selective glucocorticoid receptor agonist (SEGRA) CpdA with the similar chemical structure, as well as classic GCs. We have also considered synephrine as potentially useful in revealing SEGRA properties, when using it as the template for synthesis of putative SEGRAs. The studies of SEGRAs demonstrate that SEGRAs, including CpdA, could be effective alternatives to GCs in the treatment of inflammatory diseases, cancer, metabolic and cardiovascular diseases, using CpdA as an example. However, the degradation of CpdA can lead to toxic metabolites being formed. Therefore, searching for synephrine derivatives we overview here can open up new avenues in solving this problem. In addition, the review provides information on the safe pharmacological profile of synephrine. Overall, due to these properties synephrine and its potential derivatives, are considered to be promising for further preclinical studies.

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## References

1. Ruiz-Moreno, C.; Del Coso, J.; Giráldez-Costas, V.; González-García, J.; Gutiérrez-Hellín, J. Effects of P-Synephrine during Exercise: A Brief Narrative Review. *Nutrients* **2021**, *13*, 233, doi:10.3390/nu13010233.
2. Maldonado, M.R.; Bracht, L.; de Sá-Nakanishi, A.B.; Corrêa, R.C.G.; Comar, J.F.; Peralta, R.M.; Bracht, A. Actions of P-Synephrine on Hepatic Enzyme Activities Linked to Carbohydrate Metabolism and ATP Levels in Vivo and in the Perfused Rat Liver. *Cell Biochem. Funct.* **2018**, *36*, 4–12, doi:10.1002/cbf.3311.
3. Gutiérrez-Hellín, J.; Del Coso, J. Acute P-synephrine Ingestion Increases Fat Oxidation Rate during Exercise. *Br. J. Clin. Pharmacol.* **2016**, *82*, 362–368, doi:10.1111/bcp.12952.



4. Pellati, F.; Benvenuti, S. Fast High-Performance Liquid Chromatography Analysis of Phenethylamine Alkaloids in Citrus Natural Products on a Pentafluorophenylpropyl Stationary Phase. *J. Chromatogr. A* **2007**, *1165*, 58–66, doi:10.1016/j.chroma.2007.07.041.
5. De Jonge, M.L.L.; Kieviet, L.C.; Sierts, M.; Egberink, L.B.; Van Der Heyden, M.A.G. Review of Case Reports on Adverse Events Related to Pre-Workout Supplements Containing Synephrine. *Cardiovasc. Toxicol.* **2023**, *23*, 1–9, doi:10.1007/s12012-022-09777-z.
6. Huang, Y.-C.; Li, J.-M.; Chen, B.-Z.; Zhang, X.-M.; Wu, R.-H.; Wu, P.-P.; Li, C.; Chen, W.-H. Recent Advance in the Biological Activity of Synephrine in Citri Reticulatae Pericarpium. *Eur. J. Med. Chem. Rep.* **2022**, *5*, 100061, doi:10.1016/j.ejmcr.2022.100061.
7. Wang, Y.-L.; Lin, S.-X.; Wang, Y.; Liang, T.; Jiang, T.; Liu, P.; Li, X.-Y.; Lang, D.-Q.; Liu, Q.; Shen, C.-Y. P -Synephrine Ameliorates Alloxan-Induced Diabetes Mellitus through Inhibiting Oxidative Stress and Inflammation via Suppressing the NF-Kappa B and MAPK Pathways. *Food Funct.* **2023**, *14*, 1971–1988, doi:10.1039/D2FO03003A.
8. Ribeiro, D.L.; Machado, A.R.T.; Machado, C.; Ferro Aissa, A.; Dos Santos, P.W.; Barcelos, G.R.M.; Antunes, L.M.G. P -Synephrine Induces Transcriptional Changes via the cAMP/PKA Pathway but Not Cytotoxicity or Mutagenicity in Human Gastrointestinal Cells. *J. Toxicol. Environ. Health A* **2021**, *84*, 196–212, doi:10.1080/15287394.2020.1855490.
9. Ishida, M.; Takekuni, C.; Nishi, K.; Sugahara, T. P -Synephrine Suppresses Inflammatory Responses in Lipopolysaccharide-Stimulated RAW264.7 Cells and Alleviates Systemic Inflammatory Response Syndrome in Mice. *Food Funct.* **2022**, *13*, 5229–5239, doi:10.1039/D2FO00299J.
10. Taheri, R.; Hamzkanlu, N.; Rezvani, Y.; Niroumand, S.; Samandar, F.; Amiri-Tehrani Zadeh, Z.; Saberi, M.R.; Chamani, J. Exploring the HSA/DNA/Lung Cancer Cells Binding Behavior of p-Synephrine, a Naturally Occurring Phenyl Ethanol Amine with Anti-Adipogenic Activity: Multi Spectroscopic, Molecular Dynamic and Cellular Approaches. *J. Mol. Liq.* **2022**, *368*, 120826, doi:10.1016/j.molliq.2022.120826.
11. Yemelyanov, A.; Czwornog, J.; Gera, L.; Joshi, S.; Chatterton, R.T.; Budunova, I. Novel Steroid Receptor Phyto-Modulator Compound a Inhibits Growth and Survival of Prostate Cancer Cells. *Cancer Res.* **2008**, *68*, 4763–4773, doi:10.1158/0008-5472.CAN-07-6104.
12. Adcock, I.M. Glucocorticoid-Regulated Transcription Factors. *Pulm. Pharmacol. Ther.* **2001**, *14*, 211–219, doi:10.1006/pupt.2001.0283.
13. Barnes, P.J. Glucocorticosteroids: Current and Future Directions. *Br. J. Pharmacol.* **2011**, *163*, 29–43, doi:10.1111/j.1476-5381.2010.01199.x.
14. De Bosscher, K.; Berghe, W.V.; Beck, I.M.E.; Van Molle, W.; Hennuyer, N.; Hapgood, J.; Libert, C.; Staels, B.; Louw, A.; Haegeman, G. A Fully Dissociated Compound of Plant Origin for Inflammatory Gene Repression. *Proc. Natl. Acad. Sci.* **2005**, *102*, 15827–15832, doi:10.1073/pnas.0505541102.
15. Liberman, A.C.; Antunica-Noguero, M.; Ferraz-de-Paula, V.; Palermo-Neto, J.; Castro, C.N.; Druker, J.; Holsboer, F.; Perone, M.J.; Gerlo, S.; De Bosscher, K.; et al. Compound A, a Dissociated Glucocorticoid Receptor Modulator, Inhibits T-Bet (Th1) and Induces GATA-3 (Th2) Activity in Immune Cells. *PLoS ONE* **2012**, *7*, e35155, doi:10.1371/journal.pone.0035155.
16. De Bosscher, K.; Beck, I.M.; Dejager, L.; Bougarne, N.; Gaigneaux, A.; Chateauvieux, S.; Ratman, D.; Bracke, M.; Tavernier, J.; Vanden Berghe, W.; et al. Selective Modulation of the Glucocorticoid Receptor Can Distinguish between Transrepression of NF-κB and AP-1. *Cell. Mol. Life Sci.* **2014**, *71*, 143–163, doi:10.1007/s00018-013-1367-4.
17. Ratman, D.; Vanden Berghe, W.; Dejager, L.; Libert, C.; Tavernier, J.; Beck, I.M.; De Bosscher, K. How Glucocorticoid Receptors Modulate the Activity of Other Transcription Factors: A Scope beyond Tethering. *Mol. Cell. Endocrinol.* **2013**, *380*, 41–54, doi:10.1016/j.mce.2012.12.014.
18. Zhang, T.; Liang, Y.; Zhang, J. Natural and Synthetic Compounds as Dissociated Agonists of Glucocorticoid Receptor. *Pharmacol. Res.* **2020**, *156*, 104802, doi:10.1016/j.phrs.2020.104802.
19. Robertson, S.; Rohwer, J.M.; Hapgood, J.P.; Louw, A. Impact of Glucocorticoid Receptor Density on Ligand-Independent Dimerization, Cooperative Ligand-Binding and Basal Priming of Transactivation: A Cell Culture Model. *PLoS One* **2013**, *8*, e64831, doi:10.1371/journal.pone.0064831.
20. Tsai, S.Y.; Carlstedt-Duke, J.; Weigel, N.L.; Dahlman, K.; Gustafsson, J.A.; Tsai, M.J.; O'Malley, B.W. Molecular Interactions of Steroid Hormone Receptor with Its Enhancer Element: Evidence for Receptor Dimer Formation. *Cell* **1988**, *55*, 361–369, doi:10.1016/0092-8674(88)90059-1.
21. Dembitsky, V.M.; Terent'ev, A.O.; Levitsky, D.O. Aziridine Alkaloids: Origin, Chemistry and Activity. In *Natural Products*; Ramawat, K.G., Mérillon, J.-M., Eds.; Springer Berlin Heidelberg: Berlin, Heidelberg, 2013; pp. 977–1006 ISBN 978-3-642-22143-9.
22. Wüst, S.; Tischner, D.; John, M.; Tuckermann, J.P.; Menzfeld, C.; Hanisch, U.-K.; van den Brandt, J.; Lühder, F.; Reichardt, H.M. Therapeutic and Adverse Effects of a Non-Steroidal Glucocorticoid Receptor Ligand in a Mouse Model of Multiple Sclerosis. *PLoS One* **2009**, *4*, e8202, doi:10.1371/journal.pone.0008202.

23. Lesovaya, E.A.; Chudakova, D.; Baida, G.; Zhidkova, E.M.; Kirsanov, K.I.; Yakubovskaya, M.G.; Budunova, I.V. The Long Winding Road to the Safer Glucocorticoid Receptor (GR) Targeting Therapies. *Oncotarget* **2022**, *13*, 408–424, doi:10.18632/oncotarget.28191.
24. Zare, F.; Solhjoo, A.; Sadeghpour, H.; Sakhteman, A.; Dehshahri, A. Structure-Based Virtual Screening, Molecular Docking, Molecular Dynamics Simulation and MM/PBSA Calculations towards Identification of Steroidal and Non-Steroidal Selective Glucocorticoid Receptor Modulators. *J. Biomol. Struct. Dyn.* **2023**, *41*, 7640–7650, doi:10.1080/07391102.2022.2123392.
25. Swart, P.; van der Merwe, K.J.; Swart, A.C.; Todres, P.C.; Hofmeyr, J.H. Inhibition of Cytochrome P-450(11)Beta by Some Naturally Occurring Acetophenones and Plant Extracts from the Shrub *Salsola Tuberculiformis*. *Planta Med.* **1993**, *59*, 139–143, doi:10.1055/s-2006-959629.
26. Stohs, S.J. Safety, Efficacy, and Mechanistic Studies Regarding Citrus Aurantium (Bitter Orange) Extract and p-Synephrine. *Phytother. Res. PTR* **2017**, *31*, 1463–1474, doi:10.1002/ptr.5879.
27. Stohs, S.J. Problems with Citrus Aurantium Information in “A Review on Botanical Species and Chemical Compounds with Appetite Suppressing Properties for Body Weight Control.” *Plant Foods Hum. Nutr.* **2013**, *68*, 329–331, doi:10.1007/s11130-013-0376-7.
28. Shara, M.; Stohs, S.J.; Mukattash, T.L. Cardiovascular Safety of Oral p -Synephrine (Bitter Orange) in Healthy Subjects: A Randomized Placebo-Controlled Cross-over Clinical Trial: Lack of Adverse Effects of p -Synephrine. *Phytother. Res.* **2016**, *30*, 842–847, doi:10.1002/ptr.5590.
29. Stohs, S.J.; Preuss, H.G.; Shara, M. A Review of the Human Clinical Studies Involving Citrus Aurantium (Bitter Orange) Extract and Its Primary Protoalkaloid p-Synephrine. *Int. J. Med. Sci.* **2012**, *9*, 527–538, doi:10.7150/ijms.4446.
30. Rossato, L.G.; Costa, V.M.; De Pinho, P.G.; Carvalho, F.; De Lourdes Bastos, M.; Remião, F. Structural Isomerization of Synephrine Influences Its Uptake and Ensuing Glutathione Depletion in Rat-Isolated Cardiomyocytes. *Arch. Toxicol.* **2011**, *85*, 929–939, doi:10.1007/s00204-010-0630-9.
31. Stohs, S.J.; Shara, M.; Ray, S.D. P -Synephrine, Ephedrine, p -octopamine and m -synephrine: Comparative Mechanistic, Physiological and Pharmacological Properties. *Phytother. Res.* **2020**, *34*, 1838–1846, doi:10.1002/ptr.6649.
32. Stohs, S.J. Physiological Functions and Pharmacological and Toxicological Effects of P-Octopamine. *Drug Chem. Toxicol.* **2015**, *38*, 106–112, doi:10.3109/01480545.2014.900069.
33. Vaysse, J.; Balayssac, S.; Gilard, V.; Desoubdizanne, D.; Malet-Martino, M.; Martino, R. Analysis of Adulterated Herbal Medicines and Dietary Supplements Marketed for Weight Loss by DOSY <sup>1</sup> H-NMR. *Food Addit. Contam. Part A* **2010**, *27*, 903–916, doi:10.1080/19440041003705821.
34. Mercader, J.; Wanecq, E.; Chen, J.; Carpené, C. Isopropylorsynephrine Is a Stronger Lipolytic Agent in Human Adipocytes than Synephrine and Other Amines Present in Citrus Aurantium. *J. Physiol. Biochem.* **2011**, *67*, 443–452, doi:10.1007/s13105-011-0078-2.
35. Koh, A.H.W.; Chess-Williams, R.; Lohning, A.E. Racemic Synephrine Found in Citrus Aurantium-Listing Pre-Workout Supplements Suggests a Non-Plant-Based Origin. *Drug Test. Anal.* **2021**, *13*, 1569–1575, doi:10.1002/dta.3042.
36. Tanaka, S.; Sekiguchi, M.; Yamamoto, A.; Aizawa, S.-I.; Sato, K.; Taga, A.; Terashima, H.; Ishihara, Y.; Kodama, S. Separation of Synephrine Enantiomers in Citrus Fruits by a Reversed Phase HPLC after Chiral Precolumn Derivatization. *Anal. Sci. Int. J. Jpn. Soc. Anal. Chem.* **2019**, *35*, 407–412, doi:10.2116/analsci.18P441.
37. Zhang, Y.; Wang, S.; Luo, J.; Lin, Y.; Xu, X.; Han, C.; Kong, L. Preparative Enantioseparation of Synephrine by Conventional and pH-Zone-Refining Counter-Current Chromatography. *J. Chromatogr. A* **2018**, *1575*, 122–127, doi:10.1016/j.chroma.2018.09.012.
38. Pellati, F.; Benvenuti, S.; Melegari, M. Enantioselective LC Analysis of Synephrine in Natural Products on a Protein-Based Chiral Stationary Phase. *J. Pharm. Biomed. Anal.* **2005**, *37*, 839–849, doi:10.1016/j.jpba.2004.09.008.
39. Brown, C.M.; McGrath, J.C.; Midgley, J.M.; Muir, A.G.; O'Brien, J.W.; Thonoor, C.M.; Williams, C.M.; Wilson, V.G. Activities of Octopamine and Synephrine Stereoisomers on Alpha-Adrenoceptors. *Br. J. Pharmacol.* **1988**, *93*, 417–429, doi:10.1111/j.1476-5381.1988.tb11449.x.
40. Kim, K.W.; Kim, H.D.; Jung, J.S.; Woo, R.S.; Kim, H.S.; Suh, H.W.; Kim, Y.H.; Song, D.K. Characterization of Antidepressant-like Effects of p-Synephrine Stereoisomers. *Naunyn. Schmiedeberg's Arch. Pharmacol.* **2001**, *364*, 21–26, doi:10.1007/s002100100416.
41. Savinkova, A.V.; Tilova, L.R.; Borisova, O.I.; Zhidkova, E.M.; Kuzin, K.A.; Kirsanov, K.I.; Belitsky, G.A.; Budunova, I.V.; Yakubovskaya, M.G.; Lesovaya, E.A. ANTI-TUMOR EFFECT OF CPDA ENANTIOMERS IN VITRO IN THE MODEL OF ACUTE LYMPHOBLASTIC LEUKEMIA. *Russ. J. Biotherapy* **2017**, *16*, 61–69, doi:10.17650/1726-9784-2017-16-1-61-69.
42. Ippolito, M.; Benovic, J.L. Biased Agonism at  $\beta$ -Adrenergic Receptors. *Cell. Signal.* **2021**, *80*, 109905, doi:10.1016/j.cellsig.2020.109905.
43. Hodavance, S.Y.; Gareri, C.; Torok, R.D.; Rockman, H.A. G Protein-Coupled Receptor Biased Agonism. *J. Cardiovasc. Pharmacol.* **2016**, *67*, 193–202, doi:10.1097/FJC.0000000000000356.

44. Ziemichod, W.; Gibula-Tarlowska, E.; Kotlinska, J.H.; Grochecki, P.; Kedzierska, E. *P* -Synephrine and Its Various Pharmacological Effects. *Curr. Issues Pharm. Med. Sci.* **2021**, *34*, 169–173, doi:10.2478/cipms-2021-0031.
45. Wu, L.; Zhang, L.; Li, B.; Jiang, H.; Duan, Y.; Xie, Z.; Shuai, L.; Li, J.; Li, J. AMP-Activated Protein Kinase (AMPK) Regulates Energy Metabolism through Modulating Thermogenesis in Adipose Tissue. *Front. Physiol.* **2018**, *9*, 122, doi:10.3389/fphys.2018.00122.
46. Mowers, J.; Uhm, M.; Reilly, S.M.; Simon, J.; Leto, D.; Chiang, S.-H.; Chang, L.; Saltiel, A.R. Inflammation Produces Catecholamine Resistance in Obesity via Activation of PDE3B by the Protein Kinases IKK $\epsilon$  and TBK1. *eLife* **2013**, *2*, e01119, doi:10.7554/eLife.01119.
47. Miyoshi, H.; Souza, S.C.; Zhang, H.-H.; Strissel, K.J.; Christoffolete, M.A.; Kovsan, J.; Rudich, A.; Kraemer, F.B.; Bianco, A.C.; Obin, M.S.; et al. Perilipin Promotes Hormone-Sensitive Lipase-Mediated Adipocyte Lipolysis via Phosphorylation-Dependent and -Independent Mechanisms. *J. Biol. Chem.* **2006**, *281*, 15837–15844, doi:10.1074/jbc.M601097200.
48. Clark, A.R.; Belvisi, M.G. Maps and Legends: The Quest for Dissociated Ligands of the Glucocorticoid Receptor. *Pharmacol. Ther.* **2012**, *134*, 54–67, doi:10.1016/j.pharmthera.2011.12.004.
49. Lee, D.A.; Gross, L.; Wittrock, D.A.; Windebank, A.J. Localization and Expression of Ciliary Neurotrophic Factor (CNTF) in Postmortem Sciatic Nerve from Patients with Motor Neuron Disease and Diabetic Neuropathy. *J. Neuropathol. Exp. Neurol.* **1996**, *55*, 915–923, doi:10.1097/00005072-199608000-00007.
50. Riccardi, C.; Bruscoli, S.; Migliorati, G. Molecular Mechanisms of Immunomodulatory Activity of Glucocorticoids. *Pharmacol. Res.* **2002**, *45*, 361–368, doi:10.1006/phrs.2002.0969.
51. Li, L.; Bonneton, F.; Chen, X.Y.; Laudet, V. Botanical Compounds and Their Regulation of Nuclear Receptor Action: The Case of Traditional Chinese Medicine. *Mol. Cell. Endocrinol.* **2015**, *401*, 221–237, doi:10.1016/j.mce.2014.10.028.
52. Petersen, R.K.; Madsen, L.; Pedersen, L.M.; Hallenborg, P.; Hagland, H.; Viste, K.; Døskeland, S.O.; Kristiansen, K. Cyclic AMP (cAMP)-Mediated Stimulation of Adipocyte Differentiation Requires the Synergistic Action of Epac- and cAMP-Dependent Protein Kinase-Dependent Processes. *Mol. Cell. Biol.* **2008**, *28*, 3804–3816, doi:10.1128/MCB.00709-07.
53. Bos, J.L. Epac Proteins: Multi-Purpose cAMP Targets. *Trends Biochem. Sci.* **2006**, *31*, 680–686, doi:10.1016/j.tibs.2006.10.002.
54. Stohs, S.J.; Preuss, H.G.; Shara, M. A Review of the Receptor-Binding Properties of p-Synephrine as Related to Its Pharmacological Effects. *Oxid. Med. Cell. Longev.* **2011**, *2011*, 482973, doi:10.1155/2011/482973.
55. Rutigliano, G.; Accorroni, A.; Zucchi, R. The Case for TAAR1 as a Modulator of Central Nervous System Function. *Front. Pharmacol.* **2017**, *8*, 987, doi:10.3389/fphar.2017.00987.
56. Fleischer, L.M.; Somaiya, R.D.; Miller, G.M. Review and Meta-Analyses of TAAR1 Expression in the Immune System and Cancers. *Front. Pharmacol.* **2018**, *9*, 683, doi:10.3389/fphar.2018.00683.
57. Ramirez-Virella, J.; Leininger, G.M. The Role of Central Neurotensin in Regulating Feeding and Body Weight. *Endocrinology* **2021**, *162*, bqab038, doi:10.1210/endo/bqab038.
58. Taylor, D.R.; Hancox, R.J. Interactions between Corticosteroids and Beta Agonists. *Thorax* **2000**, *55*, 595–602, doi:10.1136/thorax.55.7.595.
59. Oakley, R.H.; Cidlowski, J.A. The Biology of the Glucocorticoid Receptor: New Signaling Mechanisms in Health and Disease. *J. Allergy Clin. Immunol.* **2013**, *132*, 1033–1044, doi:10.1016/j.jaci.2013.09.007.
60. Lesovaya, E.A.; Chudakova, D.; Baida, G.; Zhidkova, E.M.; Kirsanov, K.I.; Yakubovskaya, M.G.; Budunova, I.V. The Long Winding Road to the Safer Glucocorticoid Receptor (GR) Targeting Therapies. *Oncotarget* **2022**, *13*, 408–424, doi:10.18632/oncotarget.28191.
61. Schäcke, H.; Schottelius, A.; Döcke, W.-D.; Strehlke, P.; Jaroch, S.; Schmees, N.; Rehwinkel, H.; Hennekes, H.; Asadullah, K. Dissociation of Transactivation from Transrepression by a Selective Glucocorticoid Receptor Agonist Leads to Separation of Therapeutic Effects from Side Effects. *Proc. Natl. Acad. Sci. U. S. A.* **2004**, *101*, 227–232, doi:10.1073/pnas.0300372101.
62. Lesovaya, E.; Yemelyanov, A.; Swart, A.C.; Swart, P.; Haegeman, G.; Budunova, I. Discovery of Compound A - a Selective Activator of the Glucocorticoid Receptor with Anti-Inflammatory and Anti-Cancer Activity. *Oncotarget* **2015**, *6*, 30730–30744, doi:10.18632/oncotarget.5078.
63. Lesovaya, E.; Yemelyanov, A.; Kirsanov, K.; Popa, A.; Belitsky, G.; Yakubovskaya, M.; Gordon, L.I.; Rosen, S.T.; Budunova, I. Combination of a Selective Activator of the Glucocorticoid Receptor Compound A with a Proteasome Inhibitor as a Novel Strategy for Chemotherapy of Hematologic Malignancies. *Cell Cycle* **2013**, *12*, 133–144, doi:10.4161/cc.23048.
64. Timberlake, W.E.; Griffin, D.H. Direct Inhibition of the Uptake of Proline by Cycloheximide. *Biochem. Biophys. Res. Commun.* **1973**, *54*, 216–221, doi:10.1016/0006-291x(73)90910-8.
65. Miyai, K. [Enzyme immunoassay of hormone]. *Horumon To Rinsho* **1978**, *26*, 771–779.
66. Zhidkova, E.M.; Kuzin, K.A.; Tilova, L.R.; Savinkova, A.V.; Borisova, O.I.; Lavrova, M.D.; Maximova, V.P.; Kirsanov, K.I.; Yakubovskaya, M.G.; Lesovaya, E.A. COMPARATIVE ANALYSIS OF BIOLOGICAL EFFECTS OF SELECTIVE ACTIVATOR OF THE GLUCOCORTICOID RECEPTOR CPDA ON

- DIFFERENT SUBTYPES OF BREAST CANCER CELL LINES. *Sib. J. Oncol.* **2017**, *16*, 41–46, doi:10.21294/1814-4861-2017-16-6-41-46.
67. Louw, A. GR Dimerization and the Impact of GR Dimerization on GR Protein Stability and Half-Life. *Front. Immunol.* **2019**, *10*, 1693, doi:10.3389/fimmu.2019.01693.
  68. Chen, W.; Dang, T.; Blind, R.D.; Wang, Z.; Cavasotto, C.N.; Hittelman, A.B.; Rogatsky, I.; Logan, S.K.; Garabedian, M.J. Glucocorticoid Receptor Phosphorylation Differentially Affects Target Gene Expression. *Mol. Endocrinol.* **2008**, *22*, 1754–1766, doi:10.1210/me.2007-0219.
  69. Yemelyanov, A.; Czwornog, J.; Gera, L.; Joshi, S.; Chatterton, R.T.; Budunova, I. Novel Steroid Receptor Phyto-Modulator Compound A Inhibits Growth and Survival of Prostate Cancer Cells. *Cancer Res.* **2008**, *68*, 4763–4773, doi:10.1158/0008-5472.CAN-07-6104.
  70. Hsu, C.-L.; Chen, Y.-L.; Ting, H.-J.; Lin, W.-J.; Yang, Z.; Zhang, Y.; Wang, L.; Wu, C.-T.; Chang, H.-C.; Yeh, S.; et al. Androgen Receptor (AR) NH<sub>2</sub>- and COOH-Terminal Interactions Result in the Differential Influences on the AR-Mediated Transactivation and Cell Growth. *Mol. Endocrinol.* **2005**, *19*, 350–361, doi:10.1210/me.2004-0190.
  71. Joshi, T.; Johnson, M.; Newton, R.; Gienbycz, M.A. The Long-Acting B<sub>2</sub> -Adrenoceptor Agonist, Indacaterol, Enhances Glucocorticoid Receptor-Mediated Transcription in Human Airway Epithelial Cells in a Gene- and Agonist-Dependent Manner. *Br. J. Pharmacol.* **2015**, *172*, 2634–2653, doi:10.1111/bph.13087.
  72. Kaur, M.; Chivers, J.E.; Gienbycz, M.A.; Newton, R. Long-Acting Beta<sub>2</sub>-Adrenoceptor Agonists Synergistically Enhance Glucocorticoid-Dependent Transcription in Human Airway Epithelial and Smooth Muscle Cells. *Mol. Pharmacol.* **2008**, *73*, 203–214, doi:10.1124/mol.107.040121.
  73. Rider, C.F.; Altonsy, M.O.; Mostafa, M.M.; Shah, S.V.; Sasse, S.; Manson, M.L.; Yan, D.; Kärrman-Mårdh, C.; Miller-Larsson, A.; Gerber, A.N.; et al. Long-Acting B<sub>2</sub>-Adrenoceptor Agonists Enhance Glucocorticoid Receptor (GR)-Mediated Transcription by Gene-Specific Mechanisms Rather Than Generic Effects via GR. *Mol. Pharmacol.* **2018**, *94*, 1031–1046, doi:10.1124/mol.118.112755.
  74. Tanaka, T.; Grusby, M.J.; Kaisho, T. PDLIM2-Mediated Termination of Transcription Factor NF- $\kappa$ B Activation by Intranuclear Sequestration and Degradation of the P65 Subunit. *Nat. Immunol.* **2007**, *8*, 584–591, doi:10.1038/ni1464.
  75. Wu, Q.; Li, R.; Soromou, L.W.; Chen, N.; Yuan, X.; Sun, G.; Li, B.; Feng, H. P-Syneprine Suppresses Lipopolysaccharide-Induced Acute Lung Injury by Inhibition of the NF- $\kappa$ B Signaling Pathway. *Inflamm. Res. Off. J. Eur. Histamine Res. Soc. Al* **2014**, *63*, 429–439, doi:10.1007/s00011-014-0715-7.
  76. Liu, S.; Feng, G.; Wang, G.-L.; Liu, G.-J. p38MAPK Inhibition Attenuates LPS-Induced Acute Lung Injury Involvement of NF- $\kappa$ B Pathway. *Eur. J. Pharmacol.* **2008**, *584*, 159–165, doi:10.1016/j.ejphar.2008.02.009.
  77. Cain, D.W.; Cidlowski, J.A. Immune Regulation by Glucocorticoids. *Nat. Rev. Immunol.* **2017**, *17*, 233–247, doi:10.1038/nri.2017.1.
  78. Ronchetti, S.; Migliorati, G.; Bruscoli, S.; Riccardi, C. Defining the Role of Glucocorticoids in Inflammation. *Clin. Sci.* **2018**, *132*, 1529–1543, doi:10.1042/CS20171505.
  79. Guillot, M. [Request for authorization for marketing an antibiotic for pediatric use]. *Ann. Pediatr. (Paris)* **1987**, *34*, 671–675.
  80. Cruz-Topete, D.; Cidlowski, J.A. One Hormone, Two Actions: Anti- and pro-Inflammatory Effects of Glucocorticoids. *Neuroimmunomodulation* **2015**, *22*, 20–32, doi:10.1159/000362724.
  81. Reuter, K.C.; Grunwitz, C.R.; Kaminski, B.M.; Steinhilber, D.; Radeke, H.H.; Stein, J. Selective Glucocorticoid Receptor Agonists for the Treatment of Inflammatory Bowel Disease: Studies in Mice with Acute Trinitrobenzene Sulfonic Acid Colitis. *J. Pharmacol. Exp. Ther.* **2012**, *341*, 68–80, doi:10.1124/jpet.111.183947.
  82. Cacheiro-Llaguno, C.; Hernández-Subirá, E.; Díaz-Muñoz, M.D.; Fresno, M.; Serrador, J.M.; Íñiguez, M.A. Regulation of Cyclooxygenase-2 Expression in Human T Cells by Glucocorticoid Receptor-Mediated Transrepression of Nuclear Factor of Activated T Cells. *Int. J. Mol. Sci.* **2022**, *23*, 13275, doi:10.3390/ijms232113275.
  83. Korn, S.H.; Wouters, E.F.; Wesseling, G.; Arends, J.W.; Thunnissen, F.B. Interaction between Glucocorticoids and Beta<sub>2</sub>-Agonists: Alpha and Beta Glucocorticoid-Receptor mRNA Expression in Human Bronchial Epithelial Cells. *Biochem. Pharmacol.* **1998**, *56*, 1561–1569, doi:10.1016/s0006-2952(98)00179-8.
  84. Peters, M.J.; Adcock, I.M.; Brown, C.R.; Barnes, P.J. Beta-Adrenoceptor Agonists Interfere with Glucocorticoid Receptor DNA Binding in Rat Lung. *Eur. J. Pharmacol.* **1995**, *289*, 275–281, doi:10.1016/0922-4106(95)90104-3.
  85. Zhang, Z.; Zhang, Z.-Y.; Schluesener, H.J. Compound A, a Plant Origin Ligand of Glucocorticoid Receptors, Increases Regulatory T Cells and M2 Macrophages to Attenuate Experimental Autoimmune Neuritis with Reduced Side Effects. *J. Immunol.* **2009**, *183*, 3081–3091, doi:10.4049/jimmunol.0901088.
  86. Gossye, V.; Elewaut, D.; Bougarne, N.; Bracke, D.; Van Calenbergh, S.; Haegeman, G.; De Bosscher, K. Differential Mechanism of NF- $\kappa$ B Inhibition by Two Glucocorticoid Receptor Modulators in Rheumatoid



- Arthritis Synovial Fibroblasts: NF- $\kappa$ B Inhibition by GR Modulators in RA FLS. *Arthritis Rheum.* **2009**, *60*, 3241–3250, doi:10.1002/art.24963.
87. Gossye, V.; Elewaut, D.; Van Beneden, K.; Dewint, P.; Haegeman, G.; De Bosscher, K. A Plant-Derived Glucocorticoid Receptor Modulator Attenuates Inflammation without Provoking Ligand-Induced Resistance. *Ann. Rheum. Dis.* **2010**, *69*, 291–296, doi:10.1136/ard.2008.102871.
  88. Reber, L.L.; Daubeuf, F.; Plantinga, M.; De Cauwer, L.; Gerlo, S.; Waelput, W.; Van Calenbergh, S.; Tavernier, J.; Haegeman, G.; Lambrecht, B.N.; et al. A Dissociated Glucocorticoid Receptor Modulator Reduces Airway Hyperresponsiveness and Inflammation in a Mouse Model of Asthma. *J. Immunol.* **2012**, *188*, 3478–3487, doi:10.4049/jimmunol.1004227.
  89. Saksida, T.; Vujicic, M.; Nikolic, I.; Stojanovic, I.; Haegeman, G.; Stosic-Grujicic, S. Compound A, a Selective Glucocorticoid Receptor Agonist, Inhibits Immunoinflammatory Diabetes, Induced by Multiple Low Doses of Streptozotocin in Mice. *Br. J. Pharmacol.* **2014**, *171*, 5898–5909, doi:10.1111/bph.12892.
  90. Xu, W.W.; Zheng, C.-C.; Huang, Y.-N.; Chen, W.-Y.; Yang, Q.-S.; Ren, J.-Y.; Wang, Y.-M.; He, Q.-Y.; Liao, H.-X.; Li, B. Synephrine Hydrochloride Suppresses Esophageal Cancer Tumor Growth and Metastatic Potential through Inhibition of Galectin-3-AKT/ERK Signaling. *J. Agric. Food Chem.* **2018**, *66*, 9248–9258, doi:10.1021/acs.jafc.8b04020.
  91. Leão, T.K.; Ribeiro, D.L.; Machado, A.R.T.; Costa, T.R.; Sampaio, S.V.; Antunes, L.M.G. Synephrine and Caffeine Combination Promotes Cytotoxicity, DNA Damage and Transcriptional Modulation of Apoptosis-Related Genes in Human HepG2 Cells. *Mutat. Res. Genet. Toxicol. Environ. Mutagen.* **2021**, *868–869*, 503375, doi:10.1016/j.mrgentox.2021.503375.
  92. Ribeiro, D.L.; Machado, A.R.T.; da Silva Machado, C.; Santos, P.W. da S.; Aissa, A.F.; Barcelos, G.R.M.; Antunes, L.M.G. Analysis of the Cytotoxic, Genotoxic, Mutagenic, and pro-Oxidant Effect of Synephrine, a Component of Thermogenic Supplements, in Human Hepatic Cells in Vitro. *Toxicology* **2019**, *422*, 25–34, doi:10.1016/j.tox.2019.04.010.
  93. Lanshakov, D.A.; Sukhareva, E.V.; Bulygina, V.V.; Lagunov, T.A.; Kalinina, T.S. Effects of neonatal dexamethasone and CpdA on the expression of genes for apoptosis regulator proteins in the neonatal hippocampus. *Integr. Physiol.* **2021**, *2*, 41–48, doi:10.33910/2687-1270-2021-2-1-41-48.
  94. Cui, Z.; Lee, Y.; Lee, Y.; Park, D. P-Synephrine Suppresses Glucose Production but Not Lipid Accumulation in H4IIE Liver Cells. *J. Med. Food* **2015**, *18*, 76–82, doi:10.1089/jmf.2013.3133.
  95. Klotz, A.; Baida, G.; Bhalla, P.; Haegeman, G.; Budunova, I. Selective Activator of the Glucocorticoid Receptor Compound A Dissociates Therapeutic and Atrophogenic Effects of Glucocorticoid Receptor Signaling in Skin. *J. Cancer Prev.* **2015**, *20*, 250–259, doi:10.15430/JCP.2015.20.4.250.
  96. Vatsavai, L.K.; Kilari, E.K. Interaction of P-Synephrine on the Pharmacodynamics and Pharmacokinetics of Gliclazide in Animal Models. *J. Ayurveda Integr. Med.* **2018**, *9*, 183–189, doi:10.1016/j.jaim.2017.04.010.
  97. Guo, Y.-J.; Pan, W.-W.; Liu, S.-B.; Shen, Z.-F.; Xu, Y.; Hu, L.-L. ERK/MAPK Signalling Pathway and Tumorigenesis. *Exp. Ther. Med.* **2020**, *19*, 1997–2007, doi:10.3892/etm.2020.8454.
  98. Jain, R.; Watson, U.; Vasudevan, L.; Saini, D.K. ERK Activation Pathways Downstream of GPCRs. *Int. Rev. Cell Mol. Biol.* **2018**, *338*, 79–109, doi:10.1016/bs.ircmb.2018.02.003.
  99. Segalés, J.; Perdiguero, E.; Muñoz-Cánoves, P. Regulation of Muscle Stem Cell Functions: A Focus on the P38 MAPK Signaling Pathway. *Front. Cell Dev. Biol.* **2016**, *4*, 91, doi:10.3389/fcell.2016.00091.
  100. Takagi, M.; Kimura, K.; Nakashima, K.-I.; Hirai, T.; Inoue, M. Induction of Beige Adipocytes by Naturally Occurring B3-Adrenoceptor Agonist p-Synephrine. *Eur. J. Pharmacol.* **2018**, *836*, 67–74, doi:10.1016/j.ejphar.2018.08.011.
  101. da Silva-Pereira, J.F.; Bubna, G.A.; Gonçalves, G. de A.; Bracht, F.; Peralta, R.M.; Bracht, A. Fast Hepatic Biotransformation of P-Synephrine and p-Octopamine and Implications for Their Oral Intake. *Food Funct.* **2016**, *7*, 1483–1491, doi:10.1039/c6fo00014b.
  102. Maggiorani, D.; Manzella, N.; Edmondson, D.E.; Mattevi, A.; Parini, A.; Binda, C.; Miale-Perez, J. Monoamine Oxidases, Oxidative Stress, and Altered Mitochondrial Dynamics in Cardiac Ageing. *Oxid. Med. Cell. Longev.* **2017**, *2017*, 3017947, doi:10.1155/2017/3017947.
  103. Jakopin, Ž. Risks Associated with Fat Burners: A Toxicological Perspective. *Food Chem. Toxicol. Int. J. Publ. Br. Ind. Biol. Res. Assoc.* **2019**, *123*, 205–224, doi:10.1016/j.fct.2018.10.051.
  104. Hardie, D.G. AMPK: A Target for Drugs and Natural Products with Effects on Both Diabetes and Cancer. *Diabetes* **2013**, *62*, 2164–2172, doi:10.2337/db13-0368.
  105. Chen, P.-I.; Cao, A.; Miyagawa, K.; Tojais, N.F.; Hennigs, J.K.; Li, C.G.; Sweeney, N.M.; Inglis, A.S.; Wang, L.; Li, D.; et al. Amphetamines Promote Mitochondrial Dysfunction and DNA Damage in Pulmonary Hypertension. *JCI Insight* **2017**, *2*, e90427, doi:10.1172/jci.insight.90427.
  106. Opferman, J.T.; Kothari, A. Anti-Apoptotic BCL-2 Family Members in Development. *Cell Death Differ.* **2018**, *25*, 37–45, doi:10.1038/cdd.2017.170.
  107. Kalfeist, L.; Galland, L.; Ledys, F.; Ghiringhelli, F.; Limagne, E.; Ladoire, S. Impact of Glucocorticoid Use in Oncology in the Immunotherapy Era. *Cells* **2022**, *11*, 770, doi:10.3390/cells11050770.



108. Greenstein, S.; Ghias, K.; Krett, N.L.; Rosen, S.T. Mechanisms of Glucocorticoid-Mediated Apoptosis in Hematological Malignancies. *Clin. Cancer Res. Off. J. Am. Assoc. Cancer Res.* **2002**, *8*, 1681–1694.
109. Flentje, M.; Kimmig, B.; Kuttig, H.; zum Winkel, K. [Electron therapy of malignant parotid tumors]. *Strahlenther. Onkol. Organ Dtsch. Rontgengesellschaft Al* **1988**, *164*, 136–140.
110. Pufall, M.A. Glucocorticoids and Cancer. *Adv. Exp. Med. Biol.* **2015**, *872*, 315–333, doi:10.1007/978-1-4939-2895-8\_14.
111. Rubenstein, N.M.; Guan, Y.; Woo, P.L.; Firestone, G.L. Glucocorticoid Down-Regulation of RhoA Is Required for the Steroid-Induced Organization of the Junctional Complex and Tight Junction Formation in Rat Mammary Epithelial Tumor Cells. *J. Biol. Chem.* **2003**, *278*, 10353–10360, doi:10.1074/jbc.M213121200.
112. Zheng, Y.; Izumi, K.; Li, Y.; Ishiguro, H.; Miyamoto, H. Contrary Regulation of Bladder Cancer Cell Proliferation and Invasion by Dexamethasone-Mediated Glucocorticoid Receptor Signals. *Mol. Cancer Ther.* **2012**, *11*, 2621–2632, doi:10.1158/1535-7163.MCT-12-0621.
113. Law, M.E.; Corsino, P.E.; Jahn, S.C.; Davis, B.J.; Chen, S.; Patel, B.; Pham, K.; Lu, J.; Sheppard, B.; Nørgaard, P.; et al. Glucocorticoids and Histone Deacetylase Inhibitors Cooperate to Block the Invasiveness of Basal-like Breast Cancer Cells through Novel Mechanisms. *Oncogene* **2013**, *32*, 1316–1329, doi:10.1038/onc.2012.138.
114. Lin, K.-T.; Wang, L.-H. New Dimension of Glucocorticoids in Cancer Treatment. *Steroids* **2016**, *111*, 84–88, doi:10.1016/j.steroids.2016.02.019.
115. Taslimi, P.; Akıncioğlu, H.; Gülçin, İ. Synephrine and Phenylephrine Act as A-amylase, A-glycosidase, Acetylcholinesterase, Butyrylcholinesterase, and Carbonic Anhydrase Enzymes Inhibitors. *J. Biochem. Mol. Toxicol.* **2017**, *31*, e21973, doi:10.1002/jbt.21973.
116. Hong, N.-Y.; Cui, Z.-G.; Kang, H.-K.; Lee, D.-H.; Lee, Y.-K.; Park, D.-B. P-Synephrine Stimulates Glucose Consumption via AMPK in L6 Skeletal Muscle Cells. *Biochem. Biophys. Res. Commun.* **2012**, *418*, 720–724, doi:10.1016/j.bbrc.2012.01.085.
117. Ratamess, N.A.; Bush, J.A.; Kang, J.; Kraemer, W.J.; Stohs, S.J.; Nocera, V.G.; Leise, M.D.; Diamond, K.B.; Campbell, S.C.; Miller, H.B.; et al. The Effects of Supplementation with P-Synephrine Alone and in Combination with Caffeine on Metabolic, Lipolytic, and Cardiovascular Responses during Resistance Exercise. *J. Am. Coll. Nutr.* **2016**, *35*, 657–669, doi:10.1080/07315724.2016.1150223.
118. Stohs, S.J.; Badmaev, V. A Review of Natural Stimulant and Non-Stimulant Thermogenic Agents. *Phytother. Res. PTR* **2016**, *30*, 732–740, doi:10.1002/ptr.5583.
119. Maldonado, M.R.; Bracht, L.; de Sá-Nakanishi, A.B.; Corrêa, R.C.G.; Comar, J.F.; Peralta, R.M.; Bracht, A. Actions of P-Synephrine on Hepatic Enzyme Activities Linked to Carbohydrate Metabolism and ATP Levels in Vivo and in the Perfused Rat Liver. *Cell Biochem. Funct.* **2018**, *36*, 4–12, doi:10.1002/cbf.3311.
120. Nagai, H.; Kaisho, T.; Yokoyama, K.; Asakawa, T.; Fujita, H.; Matsumiya, K.; Noguchi, J.; Tsuchimori, K.; Nishizawa, N.; Kanematsu-Yamaki, Y.; et al. Differential Effects of Selective Agonists of Neuromedin U1 and U2 Receptors in Obese and Diabetic Mice. *Br. J. Pharmacol.* **2018**, *175*, 359–373, doi:10.1111/bph.14077.
121. Zheng, X.; Guo, L.; Wang, D.; Deng, X. P-Synephrine: A Novel Agonist for Neuromedin U2 Receptor. *Biol. Pharm. Bull.* **2014**, *37*, 764–770, doi:10.1248/bpb.b13-00788.
122. Beaupere, C.; Liboz, A.; Fève, B.; Blondeau, B.; Guillemain, G. Molecular Mechanisms of Glucocorticoid-Induced Insulin Resistance. *Int. J. Mol. Sci.* **2021**, *22*, 623, doi:10.3390/ijms22020623.
123. Baida, G.; Bhalla, P.; Kirsanov, K.; Lesovaya, E.; Yakubovskaya, M.; Yuen, K.; Guo, S.; Lavker, R.M.; Readhead, B.; Dudley, J.T.; et al. REDD1 Functions at the Crossroads between the Therapeutic and Adverse Effects of Topical Glucocorticoids. *EMBO Mol. Med.* **2015**, *7*, 42–58, doi:10.15252/emmm.201404601.
124. Dewint, P.; Gossye, V.; De Bosscher, K.; Vanden Berghe, W.; Van Beneden, K.; Deforce, D.; Van Calenbergh, S.; Müller-Ladner, U.; Vander Cruyssen, B.; Verbruggen, G.; et al. A Plant-Derived Ligand Favoring Monomeric Glucocorticoid Receptor Conformation with Impaired Transactivation Potential Attenuates Collagen-Induced Arthritis. *J. Immunol.* **2008**, *180*, 2608–2615, doi:10.4049/jimmunol.180.4.2608.
125. Visser, K.; Smith, C.; Louw, A. Interplay of the Inflammatory and Stress Systems in a Hepatic Cell Line: Interactions between Glucocorticoid Receptor Agonists and Interleukin-6. *Endocrinology* **2010**, *151*, 5279–5293, doi:10.1210/en.2010-0368.
126. Delaunay, F.; Khan, A.; Cintra, A.; Davani, B.; Ling, Z.C.; Andersson, A.; Ostenson, C.G.; Gustafsson, J.; Efendic, S.; Okret, S. Pancreatic Beta Cells Are Important Targets for the Diabetogenic Effects of Glucocorticoids. *J. Clin. Invest.* **1997**, *100*, 2094–2098, doi:10.1172/JCI119743.
127. Mazziotti, G.; Gazzaruso, C.; Giustina, A. Diabetes in Cushing Syndrome: Basic and Clinical Aspects. *Trends Endocrinol. Metab. TEM* **2011**, *22*, 499–506, doi:10.1016/j.tem.2011.09.001.
128. Zhidkova, E.M.; Lylova, E.S.; Grigoreva, D.D.; Kirsanov, K.I.; Osipova, A.V.; Kulikov, E.P.; Mertsalov, S.A.; Belitsky, G.A.; Budunova, I.; Yakubovskaya, M.G.; et al. Nutritional Sensor REDD1 in Cancer and Inflammation: Friend or Foe? *Int. J. Mol. Sci.* **2022**, *23*, 9686, doi:10.3390/ijms23179686.
129. Rivera-Gonzalez, G.C.; Klopot, A.; Sabin, K.; Baida, G.; Horsley, V.; Budunova, I. Regulated in Development and DNA Damage Responses 1 Prevents Dermal Adipocyte Differentiation and Is Required

- for Hair Cycle-Dependent Dermal Adipose Expansion. *J. Invest. Dermatol.* **2020**, *140*, 1698-1705.e1, doi:10.1016/j.jid.2019.12.033.
130. Schäcke, H.; Zollner, T.M.; Döcke, W.D.; Rehwinkel, H.; Jaroach, S.; Skuballa, W.; Neuhaus, R.; May, E.; Zügel, U.; Asadullah, K. Characterization of ZK 245186, a Novel, Selective Glucocorticoid Receptor Agonist for the Topical Treatment of Inflammatory Skin Diseases. *Br. J. Pharmacol.* **2009**, *158*, 1088–1103, doi:10.1111/j.1476-5381.2009.00238.x.
  131. Watson, A.; Lipina, C.; McArdle, H.J.; Taylor, P.M.; Hundal, H.S. Iron Depletion Suppresses mTORC1-Directed Signalling in Intestinal Caco-2 Cells via Induction of REDD1. *Cell. Signal.* **2016**, *28*, 412–424, doi:10.1016/j.cellsig.2016.01.014.
  132. Ratamess, N.A.; Bush, J.A.; Stohs, S.J.; Ellis, N.L.; Vought, I.T.; O'Grady, E.A.; Kuper, J.D.; Hasan, S.B.; Kang, J.; Faigenbaum, A.D. Acute Cardiovascular Effects of Bitter Orange Extract (p-Synephrine) Consumed Alone and in Combination with Caffeine in Human Subjects: A Placebo-Controlled, Double-Blind Study. *Phytother. Res. PTR* **2018**, *32*, 94–102, doi:10.1002/ptr.5953.
  133. Shara, M.; Stohs, S.J.; Smadi, M.M. Safety Evaluation of P-Synephrine Following 15 Days of Oral Administration to Healthy Subjects: A Clinical Study. *Phytother. Res. PTR* **2018**, *32*, 125–131, doi:10.1002/ptr.5956.
  134. Mak, J.C.; Nishikawa, M.; Barnes, P.J. Glucocorticosteroids Increase Beta 2-Adrenergic Receptor Transcription in Human Lung. *Am. J. Physiol.-Lung Cell. Mol. Physiol.* **1995**, *268*, L41–L46, doi:10.1152/ajplung.1995.268.1.L41.

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