

Review

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Review

Primary Cutaneous CD30 positive Lymphoproliferative Disorders—Current Therapeutic Approach with Focus on Brentuximab Vedotin

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Abstract: One of the most common subgroups of cutaneous T-cell lymphomas is the that of primary cutaneous CD30 positive lymphoproliferative disorders. The group includes lymphomatoid papulosis (LyP) and primary cutaneous anaplastic large cell lymphoma (pcALCL), as well as some borderline cases. Recently, significant progress has been made in understanding the genetics and treatment of these disorders. This review article summarizes the clinical evidence supporting the current treatment options in these diseases. Recent years have seen the introduction of novel agents into clinical practice; most of these target CD30, such as anti-CD30 monoclonal antibodies and conjugated antibodies (brentuximab vedotin), bispecific antibodies and cellular therapies, particularly anti-CD30 CAR-T cells. This paper briefly reviews the biology of CD30 that makes it a good therapeutic target, and describes the anti-CD30 therapies that have emerged to date.

Keywords: anaplastic large cell lymphoma; Brentuximab vedotin; CD30 positive lymphomas; diagnosis; Lymphomatoid papulosis; primary cutaneous T-cell lymphoma; treatment

1. Introduction

Primary cutaneous CD30+ lymphoproliferative disorders (LPD) are the second most common group of primary cutaneous T-cell lymphomas, after mycosis fungoides (MF) [1]. The group includes primary cutaneous anaplastic large T-cell lymphoma (pcALTCL), lymphomatoid papulosis (LyP) and border-line tumours. They generally have a good prognosis, but most cases are also characterised by a long-term and recurrent course that makes everyday life difficult for patients. It is worth emphasizing that although these are indolent proliferations, they may pose a risk of internal organ involvement. In addition, although LyP is a benign disease, it is still included in the group of lymphomas, because in rare cases it can transform from LyP to MF, pcALTCL or Hodgkin lymphoma (HL). Generally LPD30+ disorders have a slow and chronic course. Currently, brentuximab vedotin (BV) is one of the newer drugs used to treat CD30+ LPD [2]. Brentuximab vedotin is a conjugate of a mouse-human chimeric IgG1 anti-CD30 antibody linked to monomethyl auristatin E (MMAE), an anti-tubulin drug. Treatment brings rapid remission of skin lesions, but also causes side effects, the most common of which is peripheral neuropathy. It is believed that the neuropathy that can develop during brentuximab vedotin therapy is caused by the inhibition of microtubule-dependent axonal transport.

CD30, also known as TNFRSF8 (TNF Receptor Superfamily Member 8) or Ki-1, is a 120 kDa glycoprotein receptor belonging to the TNF family [3]. It was first identified in 1982 as a surface marker for selected Hodgkin lymphoma (HL) cell lines [4]. The CD30 receptor is activated by the binding of its ligand: CD30L, a membrane-bound cytokine expressed on activated granulocytes and lymphocytes [5]. CD30 is most highly expressed in CD8+ and CD4+ lymphocytes [6]. Formation of the ligand complex activates the receptor and TNF-related factor recruitment (TRAF1, TRAF2,

TRAF3); it also initiates the binding of proteins to form a signalling complex inside the cell. This stimulates the nuclear factor kappa B (NFkB) pathway, and activates signalling via the mitogen-activated protein kinase (MAPK) pathways. Both pathways promote a variety of effects, including those that promote survival and prevent apoptosis of neoplastic cells. Moreover, it is postulated that CD30 activation is regulated through the expression of Jun-B, which is a transcription factor of activated proteins (AP-1) responsible for neoplastic transformation [7].

In healthy individuals, CD30+ expression is minimal, being most prominent on activated lymphocytes; even in this case, it only accounts for less than 1% of activated lymphocytes. In healthy individuals, the role of CD30 is thought to supervise the immune system by mediating information between B and T cells.

Importantly, CD30 expression is also observed on CD8+ and CD4+ lymphocytes in skin inflammation, viral infections and malignancies. Increased amounts of CD30 have been found in atopic dermatitis, psoriasis, parasitic infections (scabies) [8], as well as in viral infections with Epstein-Barr virus (EBV), human immunodeficiency virus (HIV), human T-lymphoma virus 1 (HTLV 1), HTLV 2 and molluscum contagiosum [9]. Falini et al. showed that the viral infection could increase the number of activated CD30-expressing cells from 0.1% to 95% within three days [10].

CD30 expression is characteristic of LyP and pcALCL, however, it is not unique to primary cutaneous CD30+ LD and cannot be defined as a marker of these disorders. It is also found in anaplastic large cell lymphoma (ALCL), HL, *large cell transformation of mycosis fungoides* (MF-LCT), acute myeloid leukaemia (AML), myelodysplastic syndromes (MDS), mastocytosis, CD30+ B-cell lymphomas and EBV+ hydroa vacciniforme-like-T-cell lymphoma [11]. CD30 expression varies across the six WHO-recognised LyP histological subtypes. Primary cutaneous anaplastic large T-cell lymphoma cells express CD30 at 75%.

In clinical practice, CD30 expression on the cell surface can be assessed by flow cytometry (FCM) and immunohistochemistry (IHC) [9]. The most common evaluation method is immunohistochemistry of skin specimens, which can be combined with an assessment of cell morphology. There are many different anti-CD30 antibodies that recognize epitopes on CD30, but the BerH2 antibody is used for the routine IHC assessment of CD30 [5]. Given recent molecular discoveries, therapy targeting the CD30 receptor appears attractive to researchers trying to identify targeted therapy of these lymphomas.

2. Primary Cutaneous CD30-Positive Lymphoproliferative Disorders

According to the current consensus of the fifth edition of the classification of cutaneous lymphomas of the World Health Organization (WHO) and the European Organization for Research and Treatment of Cancer (EORTC), primary cutaneous CD30+ LPD constitutes a separate group of lymphomas [1,12]. They are the second most common group of CTCLs, accounting for approximately 25% [13,14].

Primary cutaneous CD30+ LPDs include pcALCL and LyP, as well as borderline tumours with clinical and histological features lying between the two. Importantly, as the histological criteria are often not sufficient to distinguish between these diseases, researchers typically use the term CD30+ LPD during the initial clinical evaluation and especially pathological diagnosis, rather than LyP or pcALCL. A short follow-up period (8–10 weeks) may reveal spontaneous regression, which is more characteristic of LyP. This diagnosis is critical for further disease management and the initiation of appropriate therapy. Both LyP and pcALCL have different histological, clinical and immunophenotypic variants. Also, despite their histological picture, suggesting highly malignant neoplastic infiltration, they also generally have a slow course with a good prognosis [15].

2.1. Lymphomatoid Papulosis

Lymphomatoid papulosis was first described by Dr. Warren L. Macauley in 1968 as a benign, histologically malignant, self-limiting, but recurrent disorder of unknown etiology [16]. It was only after years of follow-up that biopsies from skin lesions demonstrated a histology typical of lymphoma

together with the presence of large atypical CD30 positive cells; as such, LyP was classified as a CD30+ LPD [17].

Lymphomatoid papulosis is a rare skin proliferation with an incidence of 1.2–1.9/million and an excellent prognosis, with a 10-year survival rate of approximately 100% [8]. However, patients with LyP are at risk of developing secondary malignancies, including nodal or cutaneous ALCL, HL, and MF. These lymphomas are clonally related to LyP and, according to various sources, develop in 4 to 60% of LyP patients. They can occur before, concomitantly, or after LyP [8,18,19]. The most common secondary lymphomas identified in a large retrospective cohort study of 180 patients with LyP were MF (61.4%) and ALCL (26.3%) [20]. Sauder et al. report that LyP B or C, male sex, LyP with monoclonal rearrangement of the TCR receptor, EBV infection and advanced age increase the chance of second malignancy; as such, each patient diagnosed with this disease requires increased oncological supervision [13].

The pathogenesis of LyP is unknown, but most studies suggest a genetic background based on abnormalities in the CD30 transcription [21]. Although viral infections with HTLV-1 virus and hepatitis E virus, were also suspected, these correlations were not confirmed in subsequent studies [22,23]. It has been suggested that LyP may be related to iatrogenic inflammation of the skin: Haro et al. [24] found that a patient previously treated with radiotherapy for breast cancer later developed LyP in the treated area.

Lymphomatoid papulosis manifests as polymorphic, varicella-like papules, sometimes vesicles, necrotic, ulcerated or haemorrhagic lesions [21] [Figure 1]. They are most common on the limbs and trunk; however, in some cases, they are also found on the genitals and oral cavity. The lesions may be accompanied by itching or slight tenderness, or may be asymptomatic. The main feature of LyP that distinguishes it from other types of CD30+ LPD is that the lesions spontaneously resolve within four to eight weeks, in which case, resolution may be permanent or a few years [25]. The disorder occurs in adults, with a slight predominance of men aged 20 to 40 years, but paediatric cases have also been reported [21]. The WHO-EORTS 2018 update classifies LyP into six subtypes, *viz.* LyP A, B, C, D, E, and LyP with DUSP22-IRF4 rearrangement; these differ slightly in histopathology and immunophenotype, but some features overlap [13].

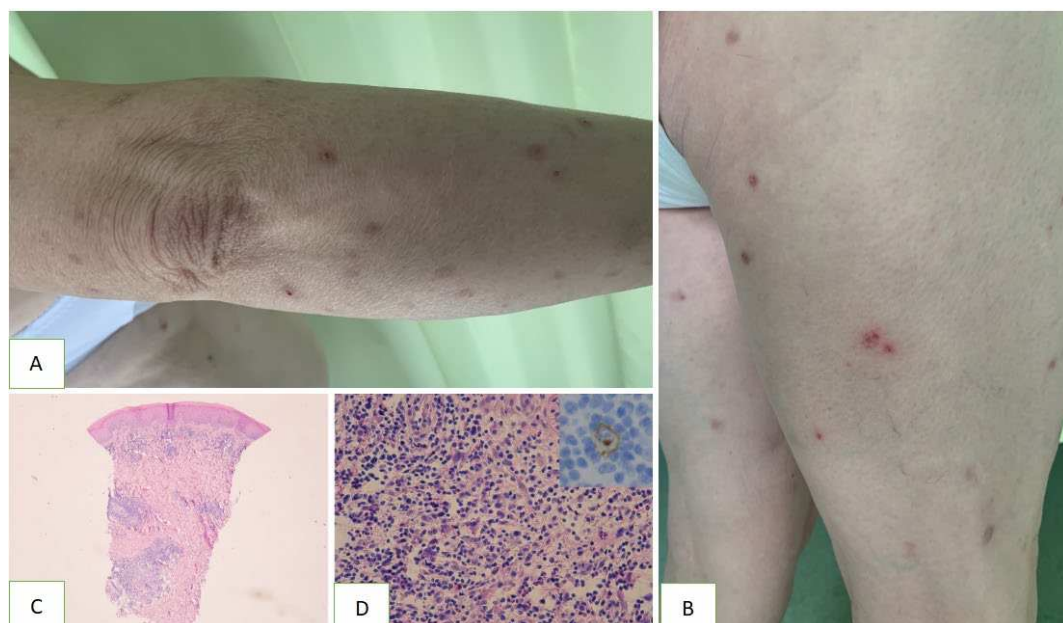


Figure 1. Clinical characteristics and histopathology of lymphoid papulosis. Scattered papules and single nodules are visible on the skin of the limbs and trunk (A). Some lesions have a hemorrhagic component, and others present disintegrations in the center (B). Typical arrangement of the cutaneous infiltrate with nodular aggregates sparing the epidermis (C). Cellular composition contains small lymphocytes and scattered eosinophils with single atypical CD30+ cells (D).

The most common subtype is LyP A >80%. All subtypes have a similar indolent clinical course, with the exception of subtype E, which manifests as vasculitis involving small and medium vessels; it may also be more aggressive and cause large ulcerative lesions [26]. The immunophenotype of the LyP cells vary by subtype, but they are represented by CD4+, CD30+, BF1+, granzyme B+, CD56+, CD8+, gamma/delta TCR (type D) cells [15].

The diagnosis of LyP is based on a combination of clinical features, histopathological and immunohistochemical findings [21]. It is necessary to perform a complete blood count, LDH determination and basic blood biochemistry. If a secondary malignancy is suspected, e.g. the presence of enlarged lymph nodes, B symptoms or skin lesions not resolving spontaneously, the diagnostics should be extended to include further laboratory and imaging tests, possibly in combination with a diagnostics with further laboratory and imaging tests, and maybe with bone marrow biopsy [27].

2.2. Primary Cutaneous Anaplastic Large Cell Lymphoma

Primary cutaneous anaplastic CD30+ cell lymphoma belongs to the subgroup of peripheral T-cell lymphomas. It was described in 1985 as a CD30-expressing malignancy of the lymphatic system, which in most cases affects only the skin [28]. It contains large cells with the pleomorphic, immunoblastic or anaplastic morphology (Figure 2). The CD30 antigen can be found in at least 75% of neoplastic cells [19,29]. The condition accounts for 9% of all CTCLs. It usually affects the elderly (median 60 years), with a greater prevalence among men, but has also been reported in children [18]. The 10-year survival rate for pcALCL is 90%, while the 5-year survival rate for ALK-positive ALCL ranges from 15 to 45% [25,30].

Primary cutaneous anaplastic large cell lymphoma is clinically manifested as solitary or multiple nodules, tumours, erythematous plaques, sometimes papular lesions, often accompanied by ulceration [Figure 2]. They tend to get bigger over weeks and usually involve the head, neck and limbs. Extensive limb disease (ELD), a variant of pcALCL, is manifested by multiple skin tumours on one limb, and is associated with disease progression and poorer prognosis [31]. Additionally, pcALCL, is less likely to demonstrate spontaneous regression than LyP, ranging from 10% to 42% of patients, and is often subject to recurrences [32]. Extracutaneous involvement is rare and most commonly involves regional lymph nodes, which are observed in 10% of patients [25]. The secondary involvement of the skin by systemic ALCL, HL and MF CD30+ should be excluded.

The classic histological picture presents limited cellular infiltration, large lymphoid cells, usually with absent or discrete epidermotropism. The immunophenotype includes CD4+, CD30+, BF1+, CD56+, granzyme B+ cells, and variable loss of CD2, CD3 and CD5. Unlike nodal ALCLs, most pcALCLs express CLA (*cutaneous lymphocyte-associated antigen*), but do not express EMA (epithelial membrane antigen) [33,34].

As in the case of LyP, the diagnosis of pcALCL is based on a combination of clinical features, histopathological and immunopathological findings. Basic laboratory tests are necessary, as in LyP; however, these should be accompanied by imaging of the whole body, preferably with PET-CT. Additionally, it is worth testing for HIV, HTLV-1 and EBV, because some T-cell lymphomas may produce secondary skin lesions and have a viral aetiology. If the lymph nodes are enlarged or metabolically active in PET, a lymph node biopsy should be performed. Bone marrow biopsy is no longer recommended in patients with pcALCL, unless the patient has systemic symptoms, cytopenia or extensive skin lesions [15,18].

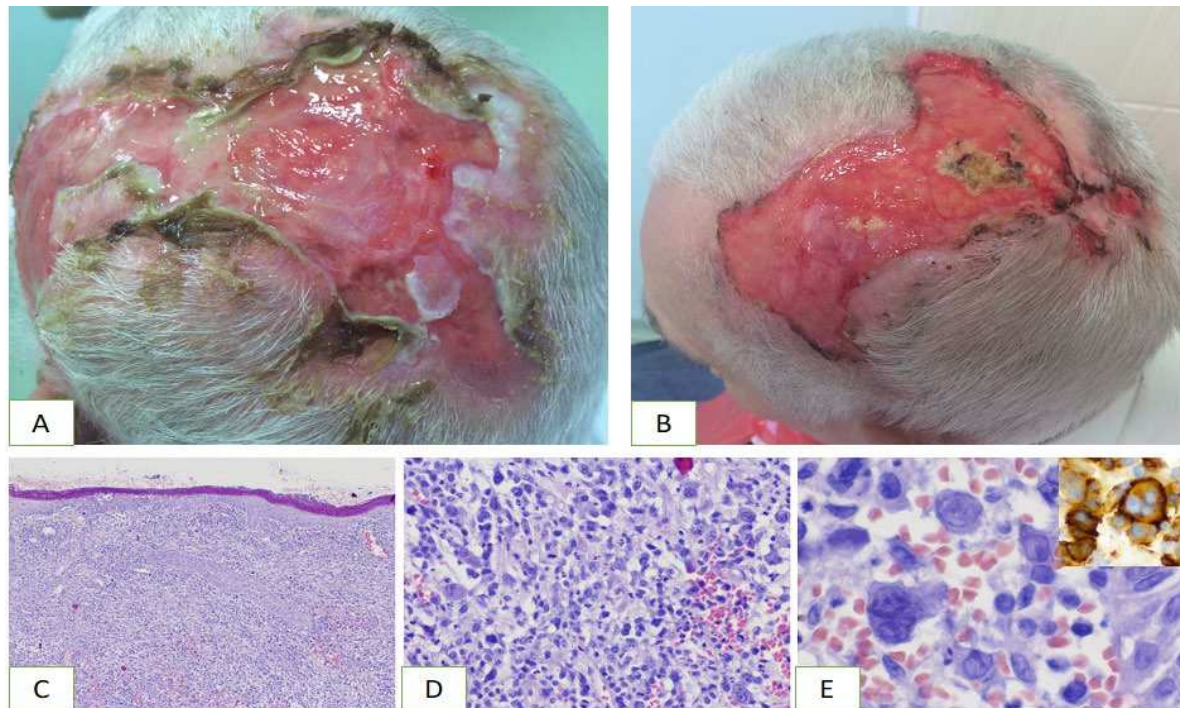


Figure 2. Clinical characteristics and histopathology of primary cutaneous anaplastic large cell lymphoma. On the top of the head there are extensive, merging ulcers accompanied by pain (A,B) Diffuse infiltrate of lymphoma cells involves the skin (C). It contains atypical, pleomorphic cells (D). Mononucleated and multinucleated cells prevails and the show strong CD30 expression (E).

3. Genetics of Primary Cutaneous CD30 Positive Lymphoproliferative Disorders

Molecular tests ease the diagnosis of cutaneous CD30+ lymphoproliferative disorders and help differentiate between LyP and pcALCL. It is also important to differentiate between pcALCL and secondary skin involvement by anaplastic lymphoma kinase (ALK)-positive and ALK-negative ALCL: the latter require different treatment methods and entail worse prognosis. ALK is a tyrosine kinase receptor. Its physiological expression is limited to a few cell types, such as endothelial cells or glial cells. ALK fusion has been noted in pcALCL but not LyP. Until recently, pcALCL has been considered an ALK-negative lymphoma, although pcALCL may also be ALK-positive [35]; this is a rare phenomenon which occurs practically only in the paediatric population and is associated with progression to the secondary systemic involvement [36].

A new LyP subtype was recently identified, characterised by a rearrangement of DUSP22 and IRF4 (Interferon Regulatory Factor 4), a tumour suppressor gene regulating T-cell proliferation, at the 6p25.3 locus; the same rearrangements have been previously reported in pcALCL, where they are observed in 20–57% of cases [37]. In the future, this may allow differential diagnosis between pcALCL and *large cell transformation* of MF (MF-LCT): MF-LCT is negative for IRF4 translocations, while pcALCL is often positive [38].

Another study identified chromosomal translocations targeting CD30+ LPD tyrosine kinases. Among the 47 patients with LPD, 4% carried the NPM1 (5q35) and TYK2 (19p13) fusion, which encodes the NPM1-TYK2 protein. This protein promotes cell proliferation by activating the STAT1/3/5 pathway; this information may be valuable when using tyrosine kinase inhibitors in patients carrying this fusion [35].

Sun et al. found that in most LPDs, the SATB1 protein (binding AT1-rich sequences), i.e. a nuclear protein of thymocytes that plays a role in T cell development, is overexpressed in CD30+ lymphocytes. Overexpression has been noted in 91.7% of LyP and 38.1% of pcALCL cases, with the prevalence increasing as the disease progresses. Interestingly, these cases have a better response to methotrexate (MTX) [39].

Epigenetics can also be used to differentiate between CD30 neoplastic proliferation and CD30+ inflammatory infiltrates. De Souza et al. evaluated the expression of intracellular 5-hydroxymethylcytosine (5-hmC) caused by DNA cytosine methylation at position 5 [40]. This phenomenon occurs in several malignancies. The authors report that 5-hmC expression occurred in 27 LyP and 14 pcALCL cases and in 19 CD30+ inflammatory infiltrates. In contrast, a complete loss of 5-hmC expression was observed in 63% of LyP and 53% of pcALCL cases; such a lack of expression was a hallmark of CD30+ LPD, and this may help to distinguish neoplastic diseases from CD30+ inflammatory infiltrates [40].

Finally, Kamstrup et al. demonstrated Notch expression in 12 LyP and in 11 pcALCL cases; the transmembrane Notch receptor influences T lymphocyte proliferation, and demonstrates strong expression in pcALCL lymphocytes but lower in LyP cells. This discovery may determine the future use of targeted therapy with Notch antagonists and may help to differentiate LyP from pcALCL [41].

4. Pharmacology of Brentuximab Vedotin

Brentuximab vedotin is a conjugate of a mouse-human chimeric IgG1 anti-CD30 antibody linked to monomethyl auristatin E (MMAE), which is an anti-tubulin drug. MMAE is a synthetic drug based on the auristatin structure. It inhibits microtubules by promoting the breakdown of these structures and inhibiting tubulin polymerization. After entering the body, the conjugate binds selectively to CD30 on the surface of LPD30+ cells. It then enters the cell by endocytosis and joins the lysosomes as a vesicle, where free MMAE is released by proteolytic enzymes. This inhibits the polymerization of tubulin in the cellular cytoskeleton, arrests the G2/M cell cycle and leads to apoptosis of LPD30+ cells (Figure 3) [42]. MMAE can induce the death of neighbouring cells by diffusion across the cell membrane [43]. Brentuximab vedotin was first approved for the treatment of classic HL and ALCL in the US in 2011 and in Europe in 2012. According to the NCCN 2021 recommendations, Brentuximab vedotin is a basic drug for the treatment of pcALCL and a first- or second-line drug in the case of CD30+ MF [44]. As part of the drug program (B66) in Poland, BV is indicated in patients with histopathologically-confirmed cutaneous T-cell lymphoma, *viz.* pcALCL and MF. However, this program requires the CD30 antigen to be confirmed immunohistochemically in at least one of biopsies.

Jagadeesh et al.[45] report the efficacy of BV to be independent of the percentage of CD30 cells. The authors analyzed five studies encompassing a total of 275 patients with PTCL, CTCL and B-cell non-Hodgkin lymphoma (NHL). In 140 patients, tumours presented CD30 expression <10%; among these, 60 patients demonstrated CD30 expression undetectable by immunohistochemistry (IHC). No significant differences in overall response rates (ORR) were observed between patients with CD30 expression >10% and those with <10% in any of the studies. The median duration of the response was also similar. Therefore, CD30 expression, as measured by standard IHC, does not appear to influence the clinical benefits of using BV.

As BV treatment may result in tumour lysis syndrome (TLS), appropriate prophylactic treatment should be applied, including a pre-treatment determination of uric acid level, and the application of prednisone, allopurinol, anticoagulant therapy and hydration. In patients with advanced disease, rasburicase may be considered, as well as hospitalization for the first administration of the drug, with subsequent observation over several days. In addition, blood tests should be performed one week after administration in order to monitor possible bone marrow damage. Premedication of an allergic reaction should also be considered, especially before the first administration of the drug. Contraception is required during BV treatment. Additionally, a man should consider semen analysis prior to treatment [46].

BV is generally well tolerated, however, side effects are frequent and present a challenge for haematologists and dermatologists. In order of frequency, they most often include infections, followed by peripheral sensory neuropathy, nausea, fatigue, diarrhoea, pyrexia, upper respiratory tract infection and neutropenia. One of the most serious, but also the rarest, side effects is reactivation of the John Cunningham (JC) virus, which causes progressive multifocal leukoencephalopathy

(PML). PML is a demyelinating disease of the CNS that is often fatal [46]. Most data related to the side effects of BV have been described in patients suffering from Hodgkin lymphoma (HL).

One of the biggest concerns associated with BV treatment is peripheral neurotoxicity, which is observed to some degree in most patients receiving BV; this is the main reason for dose modifications, delays in administration or discontinuation of treatment, and may affect the efficiency of the therapy. Peripheral neuropathy can cause sensory, motor and autonomic dysfunctions of the peripheral nerves. Importantly, peripheral neuropathy is largely reversible, but may persist for months or years after treatment. Therefore, it may become a major survival problem for patients treated with the drug [47]. The mechanism of developing neuropathy during BV therapy is related to the inhibition of microtubule-dependent axonal transport. Microtubules maintain axonal transport between nerve cell bodies and distal nerve endings [48]. Axonal degeneration typically results in neuropathy of the most peripheral nerve endings, starting at the most distal parts of the extremities, such as the fingertips and pulp of the toes; however, this condition later progresses proximally towards the trunk. Of note, peripheral neuropathy has been associated with other MMAE (Monomethyl auristatin E) antibody conjugates, such as enfortumab-vedotin (Padcev) used in the treatment of metastatic urothelial carcinoma and or polatuzumab-vedotin (Polivy) used in diffuse large B-cell lymphoma [49,50]. Interestingly, neurons do not express CD30, as confirmed by the pathological data from a sural nerve biopsy of a patient with BV-induced peripheral neuropathy [51]. Although the route by which MMAE reaches the peripheral nerves remains unclear, several potential mechanisms exist. Firstly, monomethylauristatin E can diffuse from CD30-positive lymphoma cells into the extracellular matrix and kill the surrounding CD30-negative cells, although the extent of MMAE action outside the tumour environment is unknown [52]. Also, the drug can be released before being internalized with CD30+ cells, thus damaging other cells [53]. Although small molecules, such as MMAE, can enter cells by passive diffusion and be transported within peripheral nerve cells, systemic levels of unconjugated MMAE are typically very low, and are hence unlikely to damage the peripheral nerves [54].

Brentuximab vedotin may cause motor, sensory and autonomic nerve dysfunction. Patients most often complain of sensory symptoms, including abnormal vibratory sensation (80%), abnormal tactile perception (80%), paresthesia (70%), numbness (70%), tingling (60%), burning (40%). They may also report allodynia, i.e. pain in response to usually painless stimuli, or hyperalgesia, i.e. excessive pain in response to painful stimuli. Distal loss of vibratory sensation is most readily detected on physical examination [55].

The level of exposure to MMAE and its duration in the peripheral nerve tissue is believed to most strongly influence peripheral neuropathy. More severe neuropathy was associated with more cycles of BV and more frequent dosing [54]. It was found that 22% of patients experienced peripheral neuropathy at a dose of 1.8 mg/kg, given every three weeks [56] compared to 66% at a weekly dose of 0.4-1.4 mg/kg [57]. This may be due to the repair mechanisms having insufficient time to act.

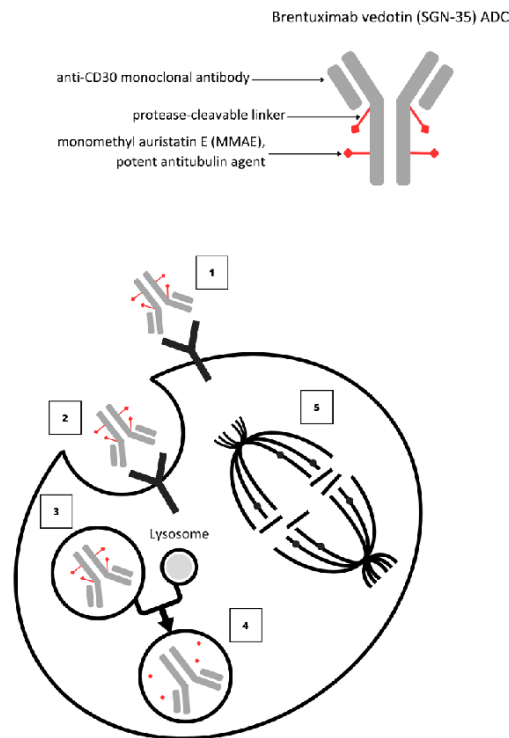


Figure 3. Brentuximab vedotin—mechanism of action. ADC binds to CD30-ADC-CD30-complex (1). Endocytosis (2). ADC-CD30 complex traffics to lysosome (3). MMAE released by lysosomal proteases (4). MMAE binds to tubulin and disrupts microtubule network leading to G2/M cell-cycle arrest and apoptosis (5).

In the ALCANZA study, 44 of the 66 (67%) patients from the BV group experienced peripheral neuropathy: eighteen patients demonstrated Grade 1 neuropathy, 20 had Grade 2 and six had Grade 3; no cases of Grade 4 were noted. Of the 44 patients, 23 (52%) required treatment modifications, e.g. dose reduction, dose interruption or infusion delay, while nine (14%) permanently discontinued BV treatment. In the final data, 26 of the 44 patients experienced complete resolution of peripheral neuropathy while 12 exhibited an improvement to Grade 1 or 2 [58]. Patients treated with BV should receive multidisciplinary care. In such cases, a neurologist can quickly detect even discreet symptoms of peripheral neuropathy by physical examination. The first symptom to appear seems to be loss of vibratory sensation, which can be assessed with a tuning fork [51,55]. Pharmacological treatment of peripheral neuropathy focuses mainly on symptomatic treatment, mainly using duloxetine; the drug is an inhibitor of serotonin and norepinephrine reuptake, recommended for brentuximab vedotin-induced peripheral neuropathy by the American Society of Clinical Oncology clinical practice guidelines [59]. This drug relieves numbness and tingling and is effective in reducing pain [60].

5. Standard Treatment

5.1. Lymphomatoid Papulosis

The best-documented therapeutic approach to LyP involves the use of topical glucocorticoids, methotrexate and phototherapy (UVA, UVB). Nevertheless, the initial watch-and-see strategy may be the first-line treatment due to the mild and self-limiting nature of the disease. Importantly, there is no evidence that early treatment prevents the development of a second malignancy or affects the course of the disease [61]. A large retrospective, multicenter study showed no difference in the final CR (Complete response) in patients treated with one of the options: topical corticosteroids, methotrexate or phototherapy [62]. Both UVB (311) and PUVA therapy have been successfully used

in the treatment of LyP. Generally, PUVA is superior to UVB nb311, however, due to the recurrent nature of LyP and the limited lifetime dose of PUVA, UVB nb311 is usually the first-line treatment [63]. Kempf et al. [27] report 27% CR and 68% PR among 19 patients treated with PUVA. In most cases, phototherapy treatment needs to be restarted when the disease recurs.

One effective treatment option in LyP is methotrexate; however, discontinuation is associated with a relapse rate of approximately 40% and long-term therapy is needed. Fernandez-De Misa et al. studied 48 patients treated with <20 mg methotrexate per week, resulting in a CR in 25 patients, a PR in 21 patients, and non-response in two [62]. Among the patients with a CR, 92% experienced a LyP relapse after discontinuation of methotrexate therapy. Vonderheid et al. [64] studied 45 patients treated with methotrexate at a median dose of 20 mg/week for 39 months. In most cases, response to treatment was observed within four weeks, and long-term disease control was reported in 39 patients. After achieving a response to methotrexate, the frequency of methotrexate dosing was reduced to as low as 1×/month. After methotrexate discontinuation, 10 patients remained LyP-free for a median follow-up of 127 months.

Although methotrexate therapy is a long-term option, it has been found that reducing the dosing to a weekly dose after remission of skin lesions is effective. Other ways of treating LyP have been described, such as local radiotherapy, interferon-alpha, bexarotene, multi-drug chemotherapy and mycophenolic acid derivatives. However, these methods are less common or have not been adopted at all in everyday medical practice.

One of the newer treatments of refractory, extensive cases is CD30-targeted therapy with BV. Lewis et al. used BV in 12 patients with LyP, all of whom received 1.8 mg/kg brentuximab vedotin infused over 30 minutes, every 21 days [65]. All the patients responded to BV, and seven showed a complete response. For all patients, time to response was three weeks. The median duration of response was 20 weeks (between six and 103 weeks). In five patients, a relapse was reported with a median time to relapse of 12 weeks (between six and 41 weeks). One patient with relapse was re-treated and remained in a partial response for more than 23 months.

5.2. Primary Cutaneous Anaplastic Large T-Cell Lymphoma

Due to its similarity to LyP, the treatment of pcALCL is based on similar therapeutic methods. In the case of single lesions, the preferred approach consists of local radiotherapy and surgical excision of the lesions. In multifocal pcALCL, systemic therapy is applied. The literature also describes new possibilities for targeted therapies based on molecular data. Single lesions can be surgically removed or subjected to local radiotherapy at 30–40 Gy [66,67]. In most cases, these are first-line therapies. Importantly, the two treatment methods are not used in combination, as this has not been found to yield any further benefits. While such management leads to remission, it typically results in relapse that requires additional therapy.

In the case of multifocal lesions, methotrexate is the treatment of choice. Due to the long-term nature of the therapy and the need to reduce toxicity, it is used in the lowest possible weekly dose allowing for remission. Once the disease is in remission, the frequency of administration can be reduced. A large retrospective study showed a 77% response rate within four weeks, although ongoing therapy is required to prevent the recurrence of skin lesions [64].

In addition, the NCCN 2021 guidelines recommend bexarotene for multifocal lesions in pcALCL. It is an oral drug, a vitamin A derivative that binds to the X receptor. However, as noted in the guidelines, the data on its use in pcALCL has been obtained from limited case reports, and it has yet to be accepted into everyday medical practice [44].

Currently, the favoured treatment for cases of extracutaneous involvement in pcALCL, characterised by the N1 peripheral lymph nodes, is multidrug chemotherapy based on the CHP (cyclophosphamide, doxorubicin, prednisone) or CHOP (CHP + vincristine) regimen. However, the preferred regimen for N1 cALCL is brentuximab vedotin with or without local radiotherapy [44,68–74]. Duvic et al. presented the results of the treatment with brentuximab vedotin conducted in 48 patients, of which 11 were diagnosed with pcALCL or LyP (2 with pcALCL, 9 with LyP), the remaining patients suffered from MF CD30+, LyP/MF, pcALCL/LyP or pcALCL/MF [69]. All patients

received intravenous BV 1.8 mg/kg every three weeks. Interestingly, skin lesions in LyP and pcALCL responded faster to the treatment than the lesions in MF, but demonstrated a shorter response. Time to response in LyP/pcALCL was three weeks (range: three to nine weeks) compared to 12 weeks (range: three to 39 weeks) in MF patients. The median duration of response was 26 weeks (range: six to 44 weeks) in LyP/pcALCL patients compared to 32 weeks (range: three to 93 weeks) in MF patients. The ORR (Objective Response Rate) for all patients was 100%.

Muniesa et al. analyzed 67 patients with CTCL who were treated with BV from the Spanish Registry of Cutaneous Lymphomas (RELCP) [70]. The study included 12 patients with CD30 lymphoproliferative disorders, two with LyP and 10 with pcALCL. Eight patients with pcALCL presented with extracutaneous involvement. The patients received a median of seven BV infusions, and the mean follow-up period was 18 months. Most patients this group showed a rapid and durable response to BV with a median PFS (Progression free survival) of 23,2 months. In patients with LPD CD30+, ORR was 84% and CR rate was 42%. These results confirm the effectiveness of brentuximab vedotin treatment.

The largest international, randomized, multicenter phase III study (ALCANZA) compared the effects of the treatment with BV 1.8 mg/kg, administered every three weeks, for up to 16 cycles with bexarotene 300 mg/m² given daily for up to 48 weeks and oral methotrexate 5–50 mg administered weekly for up to 48 weeks. The study groups comprised 128 previously-treated patients with CD30+ MF (97 patients) and pcALCL (31 patients). The patients were randomly assigned to each group: 16 patients with pcALCL received brentuximab vedotin and 15 patients were given bexarotene or methotrexate. The primary endpoint was the proportion of patients who achieved an *objective global* response lasting at least four months (ORR4). The ORR4 values were 69% (n = 11) in the BV group compared 20% (n = 3) in the bexarotene/methotrexate group. Importantly, progression-free survival (PFS) was 22.2 months longer for BV compared to bexarotene/methotrexate (i.e. 27.5 vs. 5.3 months) [59,71]. Based on the findings, brentuximab vedotin was approved at a dose of 1.8 mg/kg for the treatment of pcALCL and MF CD30+.

In a subsequent study, Ribereau-Gayon et al. evaluated the effectiveness of treatment for aggressive CD30+ CTCL. From April 2015 to January 2022, seven patients were included in the study: four with mycosis fungoides with large cell transformation, one with primary cutaneous aggressive epidermotropic CD8-positive T-cell lymphoma and two with pcALCL. The patients with pcALCL outside the skin had lymph node involvement [72]. They had no B symptoms or elevated LDH levels. In all biopsies, anaplastic cells showed strong CD 30 positivity. The aim of the study was to determine the effectiveness of treatment with a combination of anthracycline-based polychemotherapy (cyclophosphamide, doxorubicin, prednisolone) with brentuximab vedotin. Patients received six cycles of BV+CHP administered at three-week intervals: BV was given at 1.8 mg/kg, cyclophosphamide at 750 mg/m², doxorubicin at 50 mg/m² intravenously on day 1 of each cycle; prednisone was given at a dose of 100 mg, once daily, orally, on days 1 to 5. Additionally, all patients received G-CSF and had regular skin and lymph node examinations. The first PET-CT evaluation was performed after six cycles [72]. The follow-up period was 16.4 months in one patient and 9.6 months in the other. Both subjects achieved ORR4. PFS was 16.4 months in the first patient and 9.6 months in the second. Both patients achieved CR, with no progression or recurrence observed at the end of the study. The pcALCL patients responded more quickly to treatment, which is consistent with previous studies of BV monotherapy; however, this was the first real-world cohort study to be performed; the findings indicate BV to have a favorable safety profile and good effect. It also appears to be an alternative instead of CHOP for aggressive treatment of CD30+ CTCL in cases of extracutaneous involvement, but requires further study.

Milan et al. report the case of a patient with pcALCL who was treated with a short course of BV. The patient achieved a complete remission of the disease after two cycles of treatment. In cases of pcALCL, CD30 is expressed on >75% of cells; therefore, approximately 75% of cells should die after each cycle of BV [73]. The patient showed a significant reduction in lesion size, consistent with the MMAE pharmacokinetics, which peaks at one to three days after infusion.

Guarnera et al. describe the case of a 45-year-old patient with extracutaneous, recurrent and aggressive pcALCL. The patient was treated with autologous stem cell transplantation (ASCT) with consolidation with BV [74]; BV was administered at a dose of 1.8 mg/kg every 21 days for six cycles, resulting in complete remission of the disease. The patient subsequently underwent ASCT and a further 10 cycles of BV as part of consolidation therapy, for a total of 16 cycles. Thirty months after the last dose of BV, the patient remained in complete remission. Larger clinical studies are summarized in Table 1.

Table 1. Larger clinical trials of brentuximab vedotin in primary cutaneous CD30 positive lymphoproliferative disorders.

Authors/ Reference	Disease and Number of Patients	Treatment	Prior Systemic Therapies	Response	Median Response Duration (Range)	OS	Adverse Events
Duvic et al. 2015 [69]	Total 48, MF 28, pc-ALCL 2, LyP 9, LyP and MF 7, ALCL/LyP/MF 2	BV at 1.8mg/kg/iv every 21 days for a maximum of eight doses.	Median 2 (1– 10) for MF, and 1 (0–5) for LyP/pc- ALCL	Total OR - 73%, CR— 35%, MF OR 54%, ALCL/LyP/MF OR 100%	LyP/pc-ALCL 26 weeks (6 - 44), MF -32 weeks (3 to 93)	14.7 years (10.2—not reached) from 1st diagnosis	Peripheral neuropathy (67%), nausea (19%), fatigue (35%), rash (24%), diarrhea (15%), myalgia (17%), localized skin infection (15%), neutropenia (15%), alopecia (11%)
Lewis et al. 2017 [65]	LyP -12	BV 1.8 mg/kg every 21 days for maximum of 8 doses	GKS—10 MTX—4 UV-B—3	OR—100%, CR 58%	20 weeks (6- 103 weeks)	NR	Peripheral neuropathy 83%, fatigue 58%, nausea 50%, diarrhea 42%, hypersensitivity drug rash 42%, headache 33%, neutropenia 17%, abdominal pain 17%, vomiting 17%
Horwitz et al. 2021 [59]	pcALCL -64	BV 1.8 mg/kg every 3 weeks, for up to 16 3- week cycles	At least 1 prior systemic therapy or radiotherapy	OR 54.7%, CR 17.2%	16.7 m	3-yrs OS - 64.4%	Peripheral neuropathy -67%, nausea -36%, diarrhea - 29%, fatigue -29%, vomiting—17%, alopecia— 15%, pruritus—17%, pyrexia—17%
Muniesa et al. 2023 [70]	Total 67, MF— 48, SS—7, pcALCL—10, LyP -2	BV 1.8mg/kg iv every 3 weeks	Mean previous systemic treatments - of 4 (1-11)	Total OR 67%, CR 37%; pcALCL and LyP OR 84%, CR 42%	Total 10.3 m, pcALCL and LyP 23.2 m	1 death due to disease progression	Peripheral neuropathy 57%. Serious AE : Skin rash -2, local skin infection—2, sepsis -1, inflammatory adenopathy 1, pyoderma gangrenosum 1, palmoplantar pompholyx 1, anemia +/- thrombopenia 1, hepatitis -1, pancreatitis -1, acute renal failure -1, asthenia -1.
Ribereau— Gayon et. al 2023 [72]	Total 7, MF—4, pcALCL—2, aggressive epidermotropic T-cell lymphoma -1	BV + CHP adminis- tered at 3- week intervals*	0–2 pts, 1–3 pts, 2–1 pt, 3–1pt.	OR 6/7 (86%) CR -3 PR -3	14.9 m (11.6- 16.4)	No data	Anemia -6, diarrhea -5, fatigue -5, pruritus -4, nausea -3, bone pain -3, thrombopenia -3, and neutropenia -3.

* BV 1.8 mg/kg, cyclophosphamide 750 mg/m², doxorubicin 50 mg/m² iv on day 1 of each cycle, and prednisone at 100 mg once daily orally days 1 to 5 of each cycle.

6. Perspectives

Currently, the recommended first-line therapies for treating localized primary cutaneous CD30 positive lymphoproliferative disorders are complete surgical excision and local radiotherapy.

However, the most appropriate treatment for generalized and relapsed/ refractory patients remains more controversial. In recent years, BV has been indicated as one of the best options for achieving high response rates with low toxicity; however, new molecular-based therapies are still being sought for these patients [75]. One option is based on the use of Janus JAKs, a family of non-receptor tyrosine kinases involved in the intracellular signal transduction of the JAK-STAT pathway [76]. The signal cascade influences the life processes in the cell. STAT3 or JAK1 mutations are associated with pSTAT3 in pcALCL, with NPM1-TYK2 gene fusion and oncogenic activation of STAT3. Upon activation by cytokines, it is phosphorylated and translocated to the nucleus, where it acts as a transcription activator, affecting many cellular processes, including proliferative and anti-apoptotic effects. *In vitro* studies have found inhibitors of these kinases to potentially be effective in controlling pcALCL cells [76].

NOTCH-1 is a transmembrane protein that controls the further fate of the cell by playing a role in developmental processes [77,78]. It is a signaling network that regulates interactions between neighboring cells. The release of NOTCH1 (ICN1) from the membrane-associated protein NOTCH1 is prevented by gamma-secretase (γ -secretase) inhibitors. The resulting cascade downregulates the nuclear factor kB (NFkB) pathway of tumor cells, leading to the inactivation of survival genes.

Some types of hematological malignancies, including ALCL, are driven by the anaplastic lymphoma kinase (ALK) gene [79]. A variety of ALK inhibitors are available, including crizotinib, ceritinib, lorlatinib, alectinib, entrectinib and brigatinib, that can downregulate the STAT3 pathway in patients with ALK positive pcALCL, thereby causing apoptosis among tumor cells [80].

Other agents with possible activity in pcALCL LyP include Denileukin Diftox (DD) and adalimumab [81–84]. Denileukin Diftox is a chimeric immunotoxin consisting of interleukin 2 (IL-2) and the cytotoxic domain of diphtheria toxin. It acts on cells that express the interleukin-2 receptor, such as activated T cells in cutaneous T-cell lymphomas. After binding to the receptor, it undergoes endocytosis, and releases a diphtheria toxin that can inhibit protein synthesis, resulting in cell apoptosis. [81]. Denileukin Diftox was approved by the U.S. Food and Drug Administration in 1999 for the treatment of relapsing CTCL. Adalimumab is a fully human anti-tumor necrosis factor α (anti-TNF α) monoclonal antibody that specifically blocks the interaction of TNF α with its receptors [83]. There are some suggestions that adalimumab can induce a response in LyP and pcALCL [83]. CD30 and TNF- α receptors are surface receptors; they are known to indirectly activate NF-kB, which in the case of lymphoma, leads to uncontrolled proliferation. Ligation of CD30 by CD30 inhibitors blocks the activation of the NF-kB pathway, resulting in cell apoptosis. In addition, inhibition of TNF- α by adalimumab may block NF-kB pathway activation, leading to apoptosis of CD30+ cells [83]. However, some authors indicate that adalimumab can trigger lymphoma in patients treated with anti-TNF α drugs [85,86].

Both pcALCL and LyP are characterized by high expression of CD30, and as such, may be a suitable targets for chimeric antigen receptor T-cell (CAR-T) therapy [87]. Preclinical studies demonstrated that CD30 CAR-T cells lyse MyLa CTCL cells and inhibit tumor growth [88]. Also, a study of nine patients with CD30+ lymphoma found treatment based on a CD30 CAR-T infusion to be well tolerated with no immunodeficiency noted [89]. Although many studies have evaluated the effectiveness of CD30 CAR-T, especially for the treatment of HL, no studies have yet been performed on CTCL. Importantly, both pcALCL and LyP demonstrate higher CD30 expression than MF and SS, and CAR-T therapy would hypothetically be more effective in tumors with high CD30 expression.

7. Conclusion

Long-term observation has shown that BV targeted therapy is effective and safe in LPD30+, especially in pcALCL. Future research should focus on developing alternative regimens, including longer intervals between infusions, fewer cycles, or administering BV directly to single lesions. In addition, there is a need to develop novel treatment regimens that can reduce the incidence of side effects without affecting the final treatment responses; among these, bispecific antibodies and cellular therapies such as anti-CD30 CAR-T cells appear particularly most promising.

Abbreviations: ALCL—anaplastic large-cell lymphoma; LyP—lymphomatoid papulosis; pc—primary cutaneous, BV—brentuximab vedotin, CHP—cyclophosphamide, doxorubicin, prednisone; CR—complete response, GKS—steroids, GKS—steroids, LyP—lymphoid papulosis, m—months, pcALCL—primary cutaneous anaplastic large T-cell lymphoma, MF—mycosis fungoides, MTX—methotrexate, NR—not reported, mGKS—topical steroids, OR—overall response, PR—partial response.

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References

1. Willemze R, Cerroni L, Kempf W, Berti E, Facchetti F, Swerdlow SH, Jaffe ES. The 2018 update of the WHO-EORTC classification for primary cutaneous lymphomas. *Blood*. 2019;133:1703-1714.
2. Yi JH, Kim SJ, Kim WS. Brentuximab vedotin: clinical updates and practical guidance. *Blood Res*. 2017 ;52:243-253.
3. Stein H, Foss HD, Dürkop H, Marafioti T, Delsol G, Pulford K, Pileri S, Falini B. CD30(+) anaplastic large cell lymphoma: a review of its histopathologic, genetic, and clinical features. *Blood*. 2000;96:3681-3695.
4. Anderson JR, Armitage JO, Weisenburger DD. Epidemiology of the non-Hodgkin's lymphomas: Distributions of the major subtypes differ by geographic locations. *Annals of Oncology*. 1998;9:717-720.
5. van der Weyden CA, Pileri SA, Feldman AL, Whisstock J, Prince HM. Understanding CD30 biology and therapeutic targeting: a historical perspective providing insight into future directions. *Blood Cancer J*. 2017 ;7:e603
6. Zheng B, Fiumara P, Li YV, Georgakis G, Snell V, Younes M, Vauthey JN, Carbone A, Younes A. MEK/ERK pathway is aberrantly active in Hodgkin disease: a signaling pathway shared by CD30, CD40, and RANK that regulates cell proliferation and survival. *Blood*. 2003;102:1019-1027.
7. Garces de Los Fayos Alonso I, Liang HC, Turner SD, Lagger S, Merkel O, Kenner L. The Role of Activator Protein-1 (AP-1) Family Members in CD30-Positive Lymphomas. *Cancers (Basel)*. 2018;10:93
8. Di Raimondo C, Parekh V, Song JY, Rosen ST, Querfeld C, Zain J, Martinez XU, Abdulla FR. Primary Cutaneous CD30+ Lymphoproliferative Disorders: a Comprehensive Review. *Curr Hematol Malig Rep*. 2020;15:333-342
9. Karube K, Kakimoto Y, Tonozuka Y, Ohshima K. The expression of CD30 and its clinico-pathologic significance in peripheral T-cell lymphomas. *Expert Rev Hematol*. 2021;14:777-787.
10. Falini B, Pileri S, Pizzolo G, Dürkop H, Flenghi L, Stirpe F, Martelli MF, Stein H. CD30 (Ki-1) molecule: a new cytokine receptor of the tumor necrosis factor receptor superfamily as a tool for diagnosis and immunotherapy. *Blood*. 1995 ;85:1-14
11. Zheng W, Medeiros LJ, Hu Y, Powers L, Cortes JE, Ravandi-Kashani F, Kantarjian HH, Wang SA. CD30 expression in high-risk acute myeloid leukemia and myelodysplastic syndromes. *Clin Lymphoma Myeloma Leuk*. 2013;13:307-314.
12. Alaggio R, Amador C, Anagnostopoulos I, Attygalle AD, Araujo IBO, Berti E, Bhagat G, Borges AM, Boyer D, Calaminici M, Chadburn A, et al. The 5th edition of the World Health Organization Classification of Haematolymphoid Tumours: Lymphoid Neoplasms. *Leukemia*. 2022;36:1720-1748.
13. Sauder MB, O'Malley JT, LeBoeuf NR. CD30+ Lymphoproliferative Disorders of the Skin. *Hematol Oncol Clin North Am*. 2017;31:317-334.
14. Kempf W, Zimmermann AK, Mitteldorf C. Cutaneous lymphomas—An update 2019. *Hematol Oncol*. 2019 ;37(S1):43-47.
15. Nikolaenko L, Zain J, Rosen ST, Querfeld C. CD30-Positive Lymphoproliferative Disorders. *Cancer Treat Res*. 2019;176:249-268.
16. Macaulay WL. Lymphomatoid papulosis. A continuing self-healing eruption, clinically benign--histologically malignant. *Arch Dermatol*. 1968;97:23-30.

17. Kempf W. CD30+ lymphoproliferative disorders: histopathology, differential diagnosis, new variants, and simulators. *J Cutan Pathol*. 2006 ;33(S1):58–70.
18. Chen C, Gu YD, Geskin LJ. A Review of Primary Cutaneous CD30+ Lymphoproliferative Disorders. *Hematol Oncol Clin North Am*. 2019 ;33:121-134.
19. Prieto-Torres L, Rodriguez-Pinilla SM, Onaindia A, Ara M, Requena L, Piris M. CD30-positive primary cutaneous lymphoproliferative disorders: Molecular alterations and targeted therapies. *Haematologica*. 2019 ;104:226–235.
20. Wieser I, Oh CW, Talpur R, Duvic M. Lymphomatoid papulosis: Treatment response and associated lymphomas in a study of 180 patients. *J Am Acad Dermatol*. 2016;74:59–67.
21. Nowicka D, Mertowska P, Mertowski S, Hymos A, Forma A, Michalski A, Morawska I, Hryniewicz R, Niedźwiedzka-Rystwej P, Grywalska E. Etiopathogenesis, Diagnosis, and Treatment Strategies for Lymphomatoid Papulosis with Particular Emphasis on the Role of the Immune System. *Cells*. 2022;11:3697.
22. Namba H, Hamada T, Iwatsuki K. Human T-cell leukemia virus type 1-positive lymphomatoid papulosis. *European Journal of Dermatology*. 2016;26:194–195.
23. Mallet V, Bruneau J, Zuber J, Alanio C, Leclerc-Mercier S, Roque-Afonso AM, Kraft ARM, Couronné L, Roulot D, Wedemeyer H, Albert ML, Hillon P, Laroche L, Pol S, Hermine O. Hepatitis E virus-induced primary cutaneous CD30(+) T cell lymphoproliferative disorder. *J Hepatol*. 2017;67:1334-1339.
24. Haro R, Juarez A, Díaz JL, Santonja C, Manzarbeitia F, Requena L. Regional lymphomatoid papulosis of the breast restricted to an area of prior radiotherapy. *Cutis*. 2016;97:E15-E19.
25. Bekkenk MW, Geelen FA, van Voorst Vader PC, Heule F, Geerts ML, van Vloten WA, Meijer CJ, Willemze R. Primary and secondary cutaneous CD30(+) lymphoproliferative disorders: a report from the Dutch Cutaneous Lymphoma Group on the long-term follow-up data of 219 patients and guidelines for diagnosis and treatment. *Blood*. 2000 5;95:3653-3661
26. Kempf W, Kazakov DV, Schäfer L, Rütten A, Mentzel T, Paredes BE, Palmedo G, Panizzon RG, Kutzner H. Angioinvasive lymphomatoid papulosis: a new variant simulating aggressive lymphomas. *Am J Surg Pathol*. 2013 ;37:1-13
27. Kempf W, Pfaltz K, Vermeer MH, Cozzio A, Ortiz-Romero PL, Bagot M, Olsen E, Kim YH, Dummer R, Pimpinelli N, et al. EORTC, ISCL, and USCLC consensus recommendations for the treatment of primary cutaneous CD30-positive lymphoproliferative disorders: lymphomatoid papulosis and primary cutaneous anaplastic large-cell lymphoma. *Blood*. 2011;118:4024-4035
28. Stein H, Mason DY, Gerdes J, O'Connor N, Wainscoat J, Pallesen G, Gatter K, Falini B, Delsol G, Lemke H, et al. The expression of the Hodgkin's disease associated antigen Ki-1 in reactive and neoplastic lymphoid tissue: evidence that Reed-Sternberg cells and histiocytic malignancies are derived from activated lymphoid cells. *Blood*. 1985;66:848-858.
29. Saleh JS, Subtil A, Hristov AC. Primary cutaneous T-cell lymphoma: a review of the most common entities with focus on recent updates. *Hum Pathol*. 2023;138:76–102.
30. Swerdlow SH, Campo E, Pileri SA, Harris NL, Stein H, Siebert R, Advani R, Ghielmini M, Salles GA, Zelenetz AD, Jaffe ES. The 2016 revision of the World Health Organization classification of lymphoid neoplasms. *Blood*. 2016;127:2375-2390.
31. Woo DK, Jones CR, Vanoli-Storz MN, Kohler S, Reddy S, Advani R, Hoppe RT, Kim YH. Prognostic factors in primary cutaneous anaplastic large cell lymphoma: characterization of clinical subset with worse outcome. *Arch Dermatol*. 2009;145:667-674
32. Vu K, Ai W. Update on the Treatment of Anaplastic Large Cell Lymphoma. *Curr Hematol Malig Rep*. 2018;13:135–141.
33. Benner MF, Willemze R. Applicability and prognostic value of the new TNM classification system in 135 patients with primary cutaneous anaplastic large cell lymphoma. *Arch Dermatol*. 2009;145:1399-1404
34. Kempf W. A new era for cutaneous CD30-positive T-cell lymphoproliferative disorders. *Semin Diagn Pathol*. 2017;34:22–35.
35. Querfeld C, Khan I, Mahon B, Nelson BP, Rosen ST, Evens AM. Primary cutaneous and systemic anaplastic large cell lymphoma: clinicopathologic aspects and therapeutic options. *Oncology (Williston Park)*. 2010;24:574–87.
36. Pulitzer M, Ogunrinade O, Lin O, Steinherz P. ALK-positive (2p23 rearranged) anaplastic large cell lymphoma with localization to the skin in a pediatric patient. *J Cutan Pathol*. 2015;42:182–187.
37. Karai LJ, Kadin ME, Hsi ED, Sluzevich JC, Ketterling RP, Knudson RA, Feldman AL. Chromosomal rearrangements of 6p25.3 define a new subtype of lymphomatoid papulosis. *Am J Surg Pathol*. 2013;37:1173-1181.
38. Fauconneau A, Pham-Ledard A, Cappellen D, Frison E, Prochazkova-Carlotti M, Parrens M, Dalle S, Joly P, Viraben R, Franck F, et al. Assessment of diagnostic criteria between primary cutaneous anaplastic large-

- cell lymphoma and CD30-rich transformed mycosis fungoides; a study of 66 cases. *Br J Dermatol*. 2015;172:1547-1554
39. Sun J, Yi S, Qiu L, Fu W, Wang A, Liu F, Wang L, Wang T, Chen H, Wang L,, et al. SATB1 Defines a Subtype of Cutaneous CD30+ Lymphoproliferative Disorders Associated with a T-Helper 17 Cytokine Profile. *J Invest Dermatol*. 2018 Aug;138(8):1795–804.
 40. De Souza A, Tinguely M, Pfaltz M, Burghart DR, Kempf W. Loss of expression of 5-hydroxymethylcytosine in CD30-positive cutaneous lymphoproliferative disorders. *J Cutan Pathol*. 2014;41:901–906.
 41. Kamstrup MR, Ralfkiaer E, Skovgaard GL, Gniadecki R. Potential involvement of Notch1 signalling in the pathogenesis of primary cutaneous CD30-positive lymphoproliferative disorders. *Br J Dermatol*. 2008;158:747–753.
 42. Francisco JA, Cervený CG, Meyer DL, Mixan BJ, Klussman K, Chace DF, Rejniak SX, Gordon KA, DeBlanc R, Toki BE, et al. cAC10-vcMMAE, an anti-CD30-monomethyl auristatin E conjugate with potent and selective antitumor activity. *Blood*. 2003;102:1458–1465.
 43. van de Donk NWCJ, Dhimolea E. Brentuximab vedotin. *MAbs*. 2012;4:458–465.
 44. Horwitz SM, Ansell S, Ai WZ, Barnes J, Barta SK, Brammer J, Clemens MW, Dogan A, Foss F, Ghione P, et al. T-Cell Lymphomas, Version 2.2022, NCCN Clinical Practice Guidelines in Oncology. *J Natl Compr Canc Netw*. 2022;20:285-308.
 45. Jagadeesh D, Horwitz S, Bartlett NL, Kim Y, Jacobsen E, Duvic M, Little M, Trepicchio W, Fenton K, Onsum Met al. Response to Brentuximab Vedotin by CD30 Expression in Non-Hodgkin Lymphoma. *Oncologist*. 2022;27:864-873.
 46. https://www.ema.europa.eu/en/documents/product-information/adcetris-epar-product-information_en.pdf.
 47. Velasco R, Domingo-Domenech E, Sureda A. Brentuximab-Induced Peripheral Neurotoxicity: A Multidisciplinary Approach to Manage an Emerging Challenge in Hodgkin Lymphoma Therapy. *Cancers (Basel)*. 2021;13:6125
 48. Mariotto S, Ferrari S, Sorio M, Benedetti F, Tridente G, Cavallaro T, Gajofatto A, Monaco S. Brentuximab vedotin: axonal microtubule's Apolloyon. *Blood Cancer J*. 2015;5:e343.
 49. Donaghy H. Effects of antibody, drug and linker on the preclinical and clinical toxicities of antibody-drug conjugates. *MAbs*. 2016;8:659–671.
 50. Lu D, Gillespie WR, Girish S, Agarwal P, Li C, Hirata J, Chu YW, Kagedal M, Leon L, Maiya V, Jin JY. Time-to-Event Analysis of Polatuzumab Vedotin-Induced Peripheral Neuropathy to Assist in the Comparison of Clinical Dosing Regimens. *CPT Pharmacometrics Syst Pharmacol*. 2017 ;6:401-408.
 51. Corbin ZA, Nguyen-Lin A, Li S, Rahbar Z, Tavallaee M, Vogel H, Salva KA, Wood GS, Kim YH, Nagpal S. Characterization of the peripheral neuropathy associated with brentuximab vedotin treatment of Mycosis Fungoides and Sézary Syndrome. *J Neurooncol*. 2017;132:439-446
 52. Deng C, Pan B, O'Connor OA. Brentuximab vedotin. *Clin Cancer Res*. 2013;19:22–27.
 53. Brown MP, Staudacher AH. Could bystander killing contribute significantly to the antitumor activity of brentuximab vedotin given with standard first-line chemotherapy for Hodgkin lymphoma? *Immunotherapy*. 2014;6:371–375.
 54. Stagg NJ, Shen BQ, Brunstein F, Li C, Kamath AV, Zhong F, Schutten M, Fine BM. Peripheral neuropathy with microtubule inhibitor containing antibody drug conjugates: Challenges and perspectives in translatability from nonclinical toxicology studies to the clinic. *Regul Toxicol Pharmacol*. 2016;82:1-13
 55. Mariotto S, Tecchio C, Sorio M, Bertolasi L, Turatti M, Tozzi MC, Benedetti F, Cavaletti G, Monaco S, Ferrari S. Clinical and neurophysiological serial assessments of brentuximab vedotin-associated peripheral neuropathy. *Leuk Lymphoma*. 2019;60:2806-2809.
 56. Younes A, Bartlett NL, Leonard JP, Kennedy DA, Lynch CM, Sievers EL, Forero-Torres A. Brentuximab vedotin (SGN-35) for relapsed CD30-positive lymphomas. *N Engl J Med*. 2010 ;363:1812-1821
 57. Fanale MA, Forero-Torres A, Rosenblatt JD, Advani RH, Franklin AR, Kennedy DA, Han TH, Sievers EL, Bartlett NL. A phase I weekly dosing study of brentuximab vedotin in patients with relapsed/refractory CD30-positive hematologic malignancies. *Clin Cancer Res*. 2012;18:248-255.
 58. Alberti P, Bernasconi DP, Cornblath DR, Merkies ISJ, Park SB, Velasco R, et al. Prospective Evaluation of Health Care Provider and Patient Assessments in Chemotherapy-Induced Peripheral Neurotoxicity. *Neurology*. 2021 Aug 17;97(7):e660–72.
 59. Horwitz SM, Scarisbrick JJ, Dummer R, Whittaker S, Duvic M, Kim YH, Quaglini P, Zinzani PL, Bechter O, Eradat H, et al. Randomized phase 3 ALCANZA study of brentuximab vedotin vs physician's choice in cutaneous T-cell lymphoma: final data. *Blood Adv*. 2021;5:5098-5106.
 60. Prince HM, Kim YH, Horwitz SM, Dummer R, Scarisbrick J, Quaglini P, Zinzani PL, Wolter P, Sanches JA, Ortiz-Romero PL et al. Brentuximab vedotin or physician's choice in CD30-positive cutaneous T-cell

- lymphoma (ALCANZA): an international, open-label, randomised, phase 3, multicentre trial. *Lancet*. 2017 ;390:555-566.
61. AbuHilal M, Walsh S, Shear N. Associated Hematolymphoid Malignancies in Patients With Lymphomatoid Papulosis: A Canadian Retrospective Study. *J Cutan Med Surg*. 2017;21:507-512.
 62. Fernández-de-Misa R, Hernández-Machín B, Servitje O, Valentí-Medina F, Maroñas-Jiménez L, Ortiz-Romero PL, Sánchez Schmidt J, Pujol RM, Gallardo F, Pau-Charles I, et al. First-line treatment in lymphomatoid papulosis: a retrospective multicentre study. *Clin Exp Dermatol*. 2018;43:137-143.
 63. Stern RS, Liebman EJ, Våkevå L. Oral psoralen and ultraviolet-A light (PUVA) treatment of psoriasis and persistent risk of nonmelanoma skin cancer. PUVA Follow-up Study. *J Natl Cancer Inst*. 1998;90:1278-1284.
 64. Vonderheid EC, Sajjadian A, Kadin ME. Methotrexate is effective therapy for lymphomatoid papulosis and other primary cutaneous CD30-positive lymphoproliferative disorders. *J Am Acad Dermatol*. 1996 ;34:470-481.
 65. Lewis DJ, Talpur R, Huen AO, Tetzlaff MT, Duvic M. Brentuximab Vedotin for Patients With Refractory Lymphomatoid Papulosis: An Analysis of Phase 2 Results. *JAMA Dermatol*. 2017;153:1302-1306.
 66. Beljaards RC, Kaudewitz P, Berti E, Gianotti R, Neumann C, Rosso R, Paulli M, Meijer CJ, Willemze R. Primary cutaneous CD30-positive large cell lymphoma: definition of a new type of cutaneous lymphoma with a favorable prognosis. A European Multicenter Study of 47 patients. *Cancer*. 1993 ;71:2097-2104.
 67. Yu JB, McNiff JM, Lund MW, Wilson LD. Treatment of Primary Cutaneous CD30+ Anaplastic Large-Cell Lymphoma With Radiation Therapy. *International Journal of Radiation Oncology*Biophysics*. 2008;70:1542-1545.
 68. Enos TH, Feigenbaum LS, Wickless HW. Brentuximab vedotin in CD30+ primary cutaneous T-cell lymphomas: a review and analysis of existing data. *Int J Dermatol*. 2017;56:1400-1405.
 69. Duvic M, Tetzlaff MT, Gangar P, Clos AL, Sui D, Talpur R. Results of a Phase II Trial of Brentuximab Vedotin for CD30+ Cutaneous T-Cell Lymphoma and Lymphomatoid Papulosis. *J Clin Oncol*. 2015 ;33:3759-3765.
 70. Muniesa C, Gallardo F, García-Doval I, Estrach MT, Combalia A, Morillo-Andújar M, De la Cruz-Vicente F, Machan S, Moya-Martínez C, Rovira R, et al. Brentuximab vedotin in the treatment of cutaneous T-cell lymphomas: Data from the Spanish Primary Cutaneous Lymphoma Registry. *J Eur Acad Dermatol Venereol*. 2023;37:57-64.
 71. Prince HM, Kim YH, Horwitz SM, Dummer R, Scarisbrick J, Quaglino P, Zinzani PL, Wolter P, Sanches JA, Ortiz-Romero PL, et al. Brentuximab vedotin or physician's choice in CD30-positive cutaneous T-cell lymphoma (ALCANZA): an international, open-label, randomised, phase 3, multicentre trial. *Lancet*. 2017 ;390:555-66.
 72. Ribereau-Gayon E, Donzel M, Pham F, Romain-Scelle N, Perier-Muzet M, Balme B, Traverse-Glehen A, Ghesquière H, Dalle S. Brentuximab-vedotin in combination with cyclophosphamide, doxorubicin, prednisolone for the treatment of aggressive CD30-positive cutaneous T-cell lymphomas. *Leuk Lymphoma*. 2023;64:1424-1432.
 73. Milan E, Miceli P, Sernicola A, Finotto S, Marino D, Alaibac M. Complete remission of primary cutaneous anaplastic large cell lymphoma after a short course of brentuximab vedotin. *Mol Clin Oncol*. 2021;14:121.
 74. Guarnera L, Meconi F, Poggi M, Esposito F, Rizzo M, Rapisarda VM, Zizzari A, Di Raimondo C, Pupo L, Anemona L, Cantonetti M. Aggressive Primary Cutaneous Anaplastic T-Cell Lymphoma Successfully Treated with Autologous Stem Cell Transplant and Brentuximab Vedotin Consolidation: Case Report and Review of the Literature. *Hematol Rep*. 2022;14:61-66.
 75. Prieto-Torres L, Rodríguez-Pinilla SM, Onaindia A, Ara M, Requena L, Piris MÁ. CD30-positive primary cutaneous lymphoproliferative disorders: molecular alterations and targeted therapies. *Haematologica*. 2019;104:226-235.
 76. Chen E, Staudt LM, Green AR. Janus kinase deregulation in leukemia and lymphoma. *Immunity*. 2012 ;36:529-541.
 77. Kamstrup MRR, Biskup E, Gjerdrum LMR. The importance of Notch signaling in peripheral T-cell lymphomas. *Leuk Lymphoma*. 2014; 55:639-644.
 78. Larose H, Prokoph N, Matthews JD, Schleder M, Högl S, Alsulami AF, Ducray SP, Nuglozeh E, Fazaludeen FMS, Elmouna A, et al. Whole Exome Sequencing reveals NOTCH1 mutations in anaplastic large cell lymphoma and points to Notch both as a key pathway and a potential therapeutic target. *Haematologica*. 2021 ;106:1693-1704
 79. Brivio E, Baruchel A, Beishuizen A, Bourquin JP, Brown PA, Cooper T, Gore L, Kolb EA, Locatelli F, Maude SL, et al.. Targeted inhibitors and antibody immunotherapies: Novel therapies for paediatric leukaemia and lymphoma. *Eur J Cancer*. 2022;164:1-17.

80. Zhang XR, Chien PN, Nam SY, Heo CY. Anaplastic Large Cell Lymphoma: Molecular Pathogenesis and Treatment. *Cancers (Basel)*. 2022;14:1650.
81. Amagai M, Furudate S, Ohuchi K, Takahashi T, Yamazaki E, Chiba H, Asano Y, Fujimura T. Successful Treatment of Primary Cutaneous Anaplastic Large Cell Lymphoma with Denileukin Difitox. *Case Rep Oncol*. 2022 ;15:726–731.
82. Lewis DJ, Dao H, Nagarajan P, Duvic M. Primary cutaneous anaplastic large-cell lymphoma: Complete remission for 13 years after denileukin difitox. *JAAD Case Rep*. 2017;3:501–504.
83. Ottevanger R, Melchers RC, Quint KD. Remittance of primary cutaneous CD30+ lymphoproliferative disorder in a patient on adalimumab. *JAAD Case Rep*. 2022;22:34–37.
84. 84 Wolska-Washer A, Smolewski P, Robak T. Advances in the pharmacotherapeutic options for primary nodal peripheral T-cell lymphoma. *Expert Opin Pharmacother*. 2021;22:1203-1215
85. Yang C, Huang J, Huang X, Huang S, Cheng J, Liao W, Chen X, Wang X, Dai S. Risk of Lymphoma in Patients With Inflammatory Bowel Disease Treated With Anti-tumour Necrosis Factor Alpha Agents: A Systematic Review and Meta-analysis. *J Crohns Colitis*. 2018 ;12:1042–1052.
86. Bittencourt AL, Oliveira PD, Bittencourt VG, Carvalho EM, Farre L. Adult T-cell leukemia/lymphoma triggered by adalimumab. *J Clin Virol*. 2013;58:494-496.
87. 87 To V, Evtimov VJ, Jenkin G, Pupovac A, Trounson AO, Boyd RL. CAR-T cell development for Cutaneous T cell Lymphoma: current limitations and potential treatment strategies. *Front Immunol*. 2022;13:968395
88. 88 Hombach AA, Görgens A, Chmielewski M, Murke F, Kimpel J, Giebel B, Abken H. Superior Therapeutic Index in Lymphoma Therapy: CD30(+) CD34(+) Hematopoietic Stem Cells Resist a Chimeric Antigen Receptor T-cell Attack. *Mol Ther*. 2016;24:1423-1434
89. 89. Ramos CA, Ballard B, Zhang H, Dakhova O, Gee AP, Mei Z, Bilgi M, Wu MF, Liu H, Grilley B, et al. Clinical and immunological responses after CD30-specific chimeric antigen receptor-redirectioned lymphocytes. *J Clin Invest*. 2017;127:3462-3471

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