

Review

Not peer-reviewed version

The Impact of Apolipoprotein E (APOE) Epigenetics on Aging and Sporadic Alzheimer's Disease

[Madia Lozupone](#)^{*}, [Vittorio Dibello](#), Rodolfo Sardone, Fabio Castellana, [Roberta Zupo](#), [Luisa Lampignano](#), [Ilaria Bortone](#), Antonio Daniele, [Antonello Bellomo](#), [Vincenzo Solfrizzi](#), [Francesco Panza](#)^{*}

Posted Date: 3 November 2023

doi: 10.20944/preprints202311.0227.v1

Keywords: apolipoprotein E; Alzheimer's disease; methylation; dementia; epigenetics; tau protein; amyloid- β ; longevity



Preprints.org is a free multidiscipline platform providing preprint service that is dedicated to making early versions of research outputs permanently available and citable. Preprints posted at Preprints.org appear in Web of Science, Crossref, Google Scholar, Scilit, Europe PMC.

Copyright: This is an open access article distributed under the Creative Commons Attribution License which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Review

The Impact of Apolipoprotein E (APOE) Epigenetics on Aging and Sporadic Alzheimer's Disease

Madia Lozupone ¹, Vittorio Dibello ², Rodolfo Sardone ³, Fabio Castellana ⁴, Roberta Zupo ⁴, Luisa Lampignano ⁵, Ilaria Bortone ¹, Antonio Daniele ^{6,7}, Antonello Bellomo ⁸, Vincenzo Solfrizzi ⁴ and Francesco Panza ⁴

¹ Department of Translational Biomedicine and Neuroscience (DiBrain), University of Bari Aldo Moro, Bari, Italy

² Department of Orofacial Pain and Dysfunction, Academic Centre for Dentistry Amsterdam (ACTA), University of Amsterdam and Vrije Universiteit Amsterdam, 1081 HV Amsterdam, The Netherlands

³ Local Healthcare Authority of Taranto, 74121 Taranto, Italy

⁴ Department of Interdisciplinary Medicine, Clinica Medica e Geriatria "Cesare Frugoni", University of Bari Aldo Moro, Bari, Italy

⁵ Local Healthcare Authority of Bari, ASL Bari, Bari, Italy

⁶ Department of Neuroscience, Catholic University of Sacred Heart, 00168 Rome, Italy

⁷ Neurology Unit, IRCCS Fondazione Policlinico Universitario A. Gemelli, 00168 Rome, Italy

⁸ Psychiatric Unit, Department of Clinical & Experimental Medicine, University of Foggia, Foggia, Italy

* Correspondence: Madia Lozupone, MD, PhD. Department of Translational Biomedicine and Neuroscience (DiBrain), University of Bari Aldo Moro, Bari, Italy. E-mail: madia.lozupone@gmail.com. Francesco Panza, MD, PhD, Department of Interdisciplinary Medicine, Clinica Medica e Geriatria "Cesare Frugoni", University of Bari Aldo Moro, Bari, Italy. E-mail: f_panza@hotmail.com

Abstract: Sporadic Alzheimer's disease (AD) derives from an interplay among environmental factors and genetic variants, while epigenetic modifications have been expected to affect the onset and progression of its complex etiopathology. Heterozygous carriers of the apolipoprotein E gene (*APOE*) ϵ 4 allele have a 4-fold increased risk of developing AD, while *APOE* ϵ 4/ ϵ 4-carriers have a 12-fold increased risk in comparison with the *APOE* ϵ 3-carriers. The main longevity factor is the homozygous *APOE* ϵ 3/ ϵ 3 genotype. In the present narrative review article, we summarized and described the role of *APOE* epigenetics in aging and AD pathophysiology. It is not fully understood how *APOE* variants may increase or decrease AD risk, but this gene is known to affect amyloid- and tau-mediated neurodegeneration directly or indirectly, also by affecting lipid metabolism and inflammation. For sporadic AD, epigenetic regulatory mechanisms may control and influence *APOE* expression in response to external insults. Diet, a major environmental factor, has been significantly associated with physical exercise, cognitive function, and the methylation level of several cytosine-phosphate-guanine (CpG) dinucleotides sites of *APOE*.

Keywords: apolipoprotein E; Alzheimer's disease; methylation; dementia; epigenetics; tau protein; amyloid- β ; longevity

1. Introduction

The study of complex diseases is based on the association among epigenetics, gene variants, and environmental factors [1,2]. The pathophysiology of Alzheimer's disease (AD) is a mixture of many pathogenic pathways and gene expression networks. The current model of AD is based on the amyloid- β ($A\beta$) hypothesis, in which a series of deterministic events leads from $A\beta$ and tau deposition to neurodegeneration and progressive decline of cognitive function. This conceptualization matches autosomal-dominant AD, defined as dominantly inherited AD with pathological confirmation, although it is less appropriate for sporadic AD. A probabilistic AD model connoted by three variants of the disease has been proposed: autosomal-dominant AD, apolipoprotein E (apolipoprotein E gene, *APOE*) ϵ 4 allele-related sporadic AD, and *APOE* ϵ 4 allele-

unrelated sporadic AD [3]. These three variants suggested a reduced weight of the A β hypothesis, giving more importance to environmental factors and lower-risk genes [3].

There were epigenetic modifications also in AD [4]. From a genetic point of view, the known risk loci showed a low penetrance in causing AD, except for A β production-related genes, and none of them have been related to different AD pathogenic pathways. On the contrary, epigenetic alterations may modify transcriptional activity globally throughout different genes and multiple biological pathways. Epigenetic mechanisms may also explain the influence of environmental stimuli such as dietary patterns, harmful exposures, and lifestyle factors on phenotypic outcomes in individuals with the same genetic variants [5]. Additionally, genetic sequence and epigenetic code were linked in a clear way. In fact, some single nucleotide polymorphisms (SNPs) are considered a common epigenetic mark because of the rearranging of cytosine-phosphate-guanine (CpG) dinucleotides with C nucleotide methylation. These CpG-altering SNPs may modulate DNA methylation levels in a *cis* or *trans* manner or they may modify gene transcription at regions enhanced of CpG known as CpG islands [6–8].

Since early 90's, many studies have showed that *APOE* could play a central role in AD neurodegeneration. For sporadic AD *APOE* allele $\epsilon 4$ is a key genetic risk factor [9–11], with a semidominant inheritance [12], and associated to the ApoE4 isoform. Conversely, in sporadic AD, the *APOE* allele $\epsilon 2$, associated to the ApoE2 isoform, could have a protective effect [13,14]. Although the increased risk for sporadic AD in *APOE* $\epsilon 4$ -carriers, the presence of the *APOE* $\epsilon 4$ allele alone is not a causal factor for AD pathology [15]. In this context, epigenetics may represent a candidate for a point of overlapping among several AD genetic risk factors, such as the *APOE* $\epsilon 4$ allele, and the AD pathophysiological processes. Human ApoE is a glycoprotein of 299-amino acids, traditionally binding phospholipids and cholesterol. ApoE is produced in 3 common isoforms (ApoE2, ApoE3, and ApoE4) differing in two amino acid residues at positions 112 and 158, and one very uncommon isoform (ApoE3r) [16].

The *APOE* variants, respectively ϵ^2 , ϵ^3 , ϵ^4 and ϵ^3r , are determined by four haplotypes, derived from the allele association of 2 common SNPs rs429358 ($C^{3,937} \rightarrow T$) and rs7412 ($C^{4,075} \rightarrow T$) at the *APOE* locus (19q13.32), coding for the different protein isoforms [16]. These *APOE* four alleles [17], are considered the most investigated variants in human Caucasian genome. Remarkably, the *APOE* exon 4 region, encompassing the $\epsilon 2/\epsilon 3/\epsilon 4$ allele variants, is a well-defined CpG islands rich area. Moreover, the two common SNPs rs429358 and rs7412 are CpG-altering and modify the CpG content of this area. This *APOE* CpG islands rich area is a transcriptional enhancer with a specificity linked to the $\epsilon 4$ allele and cell-type [18]. In the present review article, we briefly summarized and highlighted the complex epigenetic regulation of *APOE* gene in aging and sporadic AD.

2. The role of apolipoprotein e in Alzheimer's disease pathogenesis

For sporadic AD, *APOE* is the most important genetic risk factor as well as for the earlier stages of cognitive decline represented by mild cognitive impairment (MCI) [19], but its expression is poorly understood. Astrocytes and activated microglia produced the major amount of ApoE in the brain. Having one *APOE* $\epsilon 4$ allele conducts to a 4-fold increased risk of developing AD, while having two *APOE* $\epsilon 4$ alleles conducts to a 12-fold increased risk, compared to the *APOE* $\epsilon 3$ -carriers. Conversely, the uncommon heterozygous carriers of the *APOE* $\epsilon 2$ allele have a risk 40% lower for AD and the homozygous carriers have a further reduced risk [20]. It was showed that *APOE* $\epsilon 4$ -carriers with normal cognition displayed elevated A β and tau brain burden than *APOE* $\epsilon 3$ -carriers; conversely, *APOE* $\epsilon 2$ -carriers had reduced global A β burden, without differences in regional tau burden or accumulation over time [21]. The contribution in AD pathogenesis from *APOE* involves not only A β aggregation and its clearance, but also tau-mediated neurodegeneration [22], microglia impairment [23,24], astrocyte reactivity [25], and blood-brain barrier disruption [26,27].

The three ApoE isoforms bind and transport A β peptides with differential affinity during AD pathogenesis [28,29], being highest for ApoE4, intermediate for ApoE3, and lowest for ApoE2 [30,31]. Therefore, their effects are also different concerning A β aggregation and clearance, but not A β production [32,33]. ApoE also can affect tau-mediated neurodegeneration and tauopathy by

modulating microglial responses to A β plaque pathology [34–36]. Thus, different ApoE isoforms may increase or reduce the risk for AD [29,31], based on different combined effects of ApoE isoforms on both A β deposition and neurofibrillary tangles [37]. *APOE* and its $\epsilon 2/\epsilon 3/\epsilon 4$ alleles have been connected by several genetic studies to multiple physiological conditions and disorders. Epigenetic alterations could explain the association between *APOE* and its associated diseases, considering that genetic signal associated with the disease also reflects a site's sequence architecture for epigenetic code [18].

3. Apolipoprotein E, human longevity, and Alzheimer's disease

There was a genetic association of *APOE* is with both human longevity and AD, but its mechanistic contribution in aging is largely under investigation. *APOE* pleiotropic roles may be explained by its exceptional epigenetic properties. In AD brain, these epigenetic changes could contribute to neural cell dysfunction. Additionally, several studies showed DNA methylation modifications on specific genes implicated in AD pathology such as *APOE*. In AD brain, it was showed that *APOE* CpG islands were differentially methylated in an *APOE*- and tissue-specific way [38]. In the brain of targeted replacement (TR) mice expressing human ApoE, allele variations within the major *APOE* CpG island may affect its methylation [39]. Epigenetic changes may link modified gene expression with environmental stimuli such as dietary patterns and physical exercise. In animal models, *APOE* alleles may have alterations in epigenetic regulation in response to external stimuli reported in studies on *APOE* TR mice [40].

The differences between mouse and human *APOE* gene clusters, the complexity of transcriptional control of human ApoE, and the structure of the targeting construct should be considered in the strategy for replacing mouse ApoE in the *APOE* TR models [41]. Moreover, lifestyle factors like education, alcohol consumption, smoking, and physical activity may attenuate genetic risk in the process of age-related cognitive decline and twelve modifiable risk factors might prevent or delay up to 40% of different dementias [42]. The complex interactions among age-related cognitive decline, genetics, and lifestyle may encourage behaviors maintaining cognitive health in older age [43]. At this regard, ApoE may be important for the pathophysiology of lipid metabolism [44] and central nervous system (CNS), although the role in healthy aging and longevity has seen its value growth [45–47].

Studies on longevity and healthy aging are related because subjects who live long tend to be healthy for a greater part of their lives [48]. Healthy aging can be defined as achieving older age maintaining intact cognition and/or mobility and without disabilities or multimorbidity. This last can be defined as the coexistence of two or more chronic diseases in the same subjects [49]. The detrimental effects of the *APOE* $\epsilon 4$ allele on longevity could influence the probability of a long human lifespan [48]. The *APOE* $\epsilon 2$ allele is more frequent in long-lived individuals than the $\epsilon 4$ allele [50]. Thus, the main longevity factor is the homozygous *APOE* $\epsilon 3/\epsilon 3$ genotype. The higher frequency of the $\epsilon 3$ allele in older individuals and their offspring than in controls derives from the greater amount of *APOE* $\epsilon 3/\epsilon 3$ genotype compared to the $\epsilon 2/\epsilon 3$ or $\epsilon 3/\epsilon 4$ genotypes [51].

In the pathophysiology of lipid metabolism, the role of ApoE may be related with normal/pathological aging, while its function in the pathophysiology of CNS needs further clarification [52]. In fact, in the CNS, there was about a quarter of total body cholesterol that may exert an important role in synaptic plasticity [53]. With advancing age, cholesterol metabolism may modify, and its related brain changes may be associated with the pathophysiology of AD [53]. So, in longevity and healthy aging, lipid and cholesterol maintenance are a critical factor also from an interventional point of view. The detrimental effects of *APOE* $\epsilon 4$ allele might be managed by dietary interventions [54], with a Mediterranean dietary pattern potentially including higher n-3 polyunsaturated fatty acid intakes [55,56].

4. Specific epigenetic modifications of apolipoprotein E in Alzheimer's disease

In response to environmental stimuli, epigenetic marks and signals may enable temporal combination of regulatory events through mechanisms including DNA methylation, histone

modification/chromatin conformation, and noncoding microRNAs (miRNAs). Several studies investigating DNA methylation in the *APOE* gene suggested an age-dependent flow and *APOE* DNA methylation specific for brain area. The *APOE* genomic sequence is approximately 4 kb in size (chromosome19:45408714-45412650, hg19) including its promoter. This region encompasses 172 CpG dinucleotides [57]. In the late 90s and early 2000s, polymorphic sites in the first intron and the proximal promoter the of *APOE* gene cluster (−1,019 to +407) affecting *APOE* expression have been identified [58–64] (Table 1). Notably, these polymorphisms have been related with a differential AD risk [65,66]. However, in AD, the association between these polymorphic sites and the variability of sequence in the proximal promoter with ApoE protein levels were not clearly understood. In fact, among different studies, findings on the levels of expression of *APOE* RNA and the relationship with the ApoE levels varied. In human *postmortem* brain, there was elevated methylation in AD frontal lobe of a 5'-C-phosphate-G-3' (CpG) island overlapping with exon four and downstream [67]. Interestingly, *APOE* has a well-defined CpG island not residing in the promoter region and overlapping with the *APOE* 3'-exon. In the human genome, these 3'-CpG islands are very rare representing < 1% of total CpG islands and are also conserved in other mammals [68,69]. However, the *APOE* CpG island methylation level relates to the expression level of four known *APOE* transcripts. The majority of the total *APOE* mRNA, with higher expression in the AD frontal lobe than in the frontal lobe of control subjects, is constituted by circular RNAs, miRNAs, and truncated *APOE* transcripts. The findings of several studies suggested several changes in epigenome and the regulatory role of epigenomic elements associated with the risk or clinical presentation of different neurological diseases, although the exact clinical significance of these signatures in the quantities of RNA and methylation level of CGI in the *APOE* 3'-exon was still unclear [67] (Table 1).

At the level of the individual CpG site, epigenetic regulation was showed by up/down patterns in the methylation profiles between samples and tissues. Significant differences in the global methylation levels among several brain regions were discovered across *postmortem* brain tissues. In brain regions primary affected by AD such as frontal lobe, temporal lobe, and hippocampus, methylation levels were lower. Conversely, in the cerebellum, a region apparently lacking profound pathological changes in AD but with recent important findings, the highest methylation levels were observed, suggesting that a correlation may exist between the methylation levels of the *APOE* CpG islands and the vulnerability of brain AD regions [70]. In fact, age- and AD-related alterations in several cerebellar subregions may also impact numerous functional domains, especially those affecting cognitive processing [70].

Genetic variants, which consist of CpG-altering SNP, can modify DNA methylation levels. These genetic variations may act like regulatory elements connecting genetic changes with epigenetic variability [71] (Table 1). As previously described, the *APOE* $\epsilon 2/\epsilon 3/\epsilon 4$ alleles are produced by two CpG-altering SNPs (rs429358 and rs7412) residing within the core region of the *APOE* CpG islands. The *APOE* $\epsilon 4$ allele, if compared with $\epsilon 2$ or $\epsilon 3$ alleles, adds one more CpG, further saturating a small 12 bp region with 4 CpG sites. On the contrary, the *APOE* $\epsilon 2$ allele eliminates 1 CpG and opens a 33-bp CpG-free region. Therefore, these two SNPs may alter the regional CpG burden and probably influence global DNA methylation of the CpG islands. These CpG load changes might change the binding profiles of methyl CpG-binding domain proteins, connected specifically to methylated DNA through their exclusive amino acid pattern [72].

Furthermore, within the *APOE* CpG islands, there is evidence of indirect indicators of protein binding which consist of histone marks and a DNase I hypersensitivity cluster. These findings suggested that the *APOE* CpG islands (and exon 4) may be a site for chromatin remodeling and protein binding. Considering that environmental stimuli could influence DNA methylation gradually with aging, the differences in *APOE* CpG islands methylation between healthy individuals and AD increased with age [73]. Taken together, different methylation landscapes could be represented by inheritance of different $\epsilon 2/\epsilon 3/\epsilon 4$ alleles in the *APOE* CpG islands, which could accumulate or change continuously with age, also modified by environmental factors. Recent results showed that methylation levels for most CpG sites may be in the order of *APOE* $\epsilon 4$ -carriers > *APOE* $\epsilon 3/\epsilon 3$ -carriers > *APOE* $\epsilon 2$ -carriers, considering that *APOE* $\epsilon 4$ -carriers have the greatest number of

CpG sites, while *APOE* $\epsilon 2$ carriers have the smallest number with $\epsilon 3/\epsilon 3$ in the middle [74] (Table 1). These changes could potentially alter protein binding, with some consequences on biological systems, even affecting the pathophysiological processes of multiple diseases and plasma lipids levels. *APOE* methylation could partially mediate the effects of age on plasma lipid (Figure 1).

Table 1. Overview of studies illustrating epigenetic signatures of apolipoprotein E gene (*APOE*) in aging and Alzheimer's disease (AD).

| <i>APOE</i> exons, promoter, and CGI | | | | |
|--------------------------------------|-----------------|--|---|--|
| Study | Study design | Sample size | Age or mean age at death (years) | Principal findings |
| Lambert et al., 1998 [59] | Cross-sectional | AD: 573 Controls: 509 | AD: 73.8±8.1 Controls: 70.4±7.9 | Among three <i>APOE</i> promoter mutations (−491 AT, −427 CT and Th1/E47cs), the Th1/E47cs T allele was associated with an increased AD risk, while the −491 T allele was associated with a decreased risk, independently of the <i>APOE</i> $\epsilon 2/\epsilon 3/\epsilon 4$ polymorphism effect. The −427 CT polymorphism was not associated with AD. In addition to the qualitative effect of the <i>APOE</i> $\epsilon 2/\epsilon 3/\epsilon 4$ polymorphisms on the AD occurrence, the quantitative variation of expression of these alleles due to functional <i>APOE</i> promoter mutations, may be a key determinant of AD development |
| Lambert et al., 1998 [61] | Cross-sectional | AD: 310 Controls: 293 | AD: 72-91 Controls: 75-102 | The Th1/E47cs T allele was associated with an increased risk of developing AD (odds ratio, OR = 1.29) and the OR was 1.79 for individuals bearing at least one T allele |
| Yu et al., 2013 [71] | Cross-sectional | Frontal lobe AD: 9 Controls: 6 | Frontal lobe AD: 86.8±6.9 Controls: 87.9±8.6 | <i>APOE</i> CGI exhibited transcriptional enhancer/silencer activity and differentially modulates expression of genes at the <i>APOE</i> locus in a cell type-, DNA methylation- and $\epsilon 2/\epsilon 3/\epsilon 4$ allele-specific manner. These findings implicated a novel functional role for a 3'-exon CGI and supported a modified mechanism of action for <i>APOE</i> in disease risk, involving also an epigenetically regulated transcriptional program at the <i>APOE</i> locus driven by the <i>APOE</i> CGI |
| Lee et al., 2020 [67] | Cross-sectional | Frontal lobe AD: 44 Controls: 21 Cerebellum AD: 51 Controls: 25 | Frontal lobe AD: 86.8±6.9 Controls: 87.9±8.6 Cerebellum AD: 74.6±9.3 Controls: 73.5±10.9 | <i>APOE</i> has a single CpG island (CGI) that overlaps with its 3'-exon. In this study, the presence of <i>APOE</i> circular RNA (circRNA) was discovered and found that circRNA and full-length mRNA each constitute approximately one third of the total <i>APOE</i> RNA, with truncated mRNAs likely constituting some of the missing fraction. All <i>APOE</i> RNA species demonstrated significantly higher expression in AD frontal lobe than in control frontal lobe, suggesting a possible modified mechanism of gene |

| | | | | |
|--|-----------------|-----------------------|-------|--|
| action for <i>APOE</i> in AD involving also an epigenetically regulated transcriptional program driven by DNA methylation in the <i>APOE</i> CGI | | | | |
| Ma et al., 2015 [74] | Cross-sectional | 475 men and 518 women | 18-87 | The 13 <i>APOE</i> CpG sites were categorized into three groups: Group 1 showed hypermethylation (> 50%, in the promoter region), Group 2 exhibited hypomethylation (< 50%, in the first two exons and introns), and Group 3 showed hypermethylation (> 50%, in the exon 4. <i>APOE</i> methylation was significantly associated with age and plasma total cholesterol and <i>APOE</i> methylation patterns differed across <i>APOE</i> ϵ variants and the promoter variant rs405509, which further showed a significant interaction with age |

CGI: 5'-C-phosphate-G-3' (CpG) island; CpG: cytosine-phosphate-guanine.

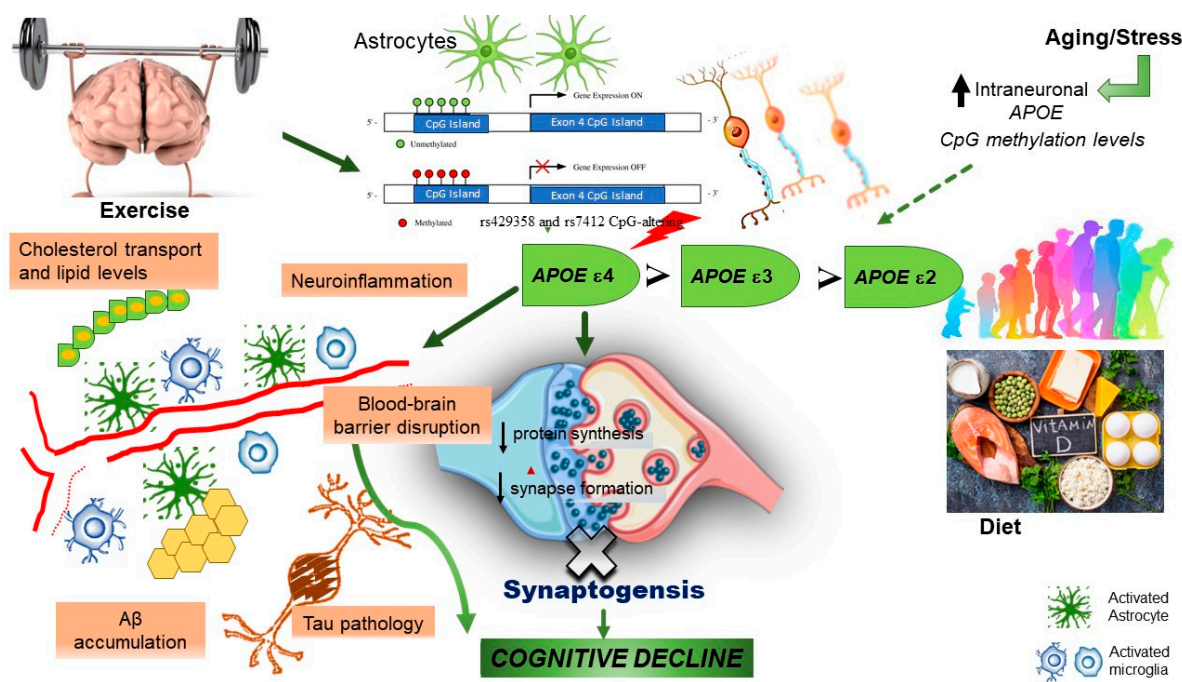


Figure 1. Despite the high lifetime risk linked to the presence of the apolipoprotein E (*APOE*) $\epsilon 3/\epsilon 4$ and *APOE* $\epsilon 4/\epsilon 4$ genotypes (the greatest risk factor for developing Alzheimer’s Disease, AD), stochastic factors (such as environment, diet, physical exercise, and ageing), may play a significant role. *APOE* pleiotropic roles may be explained by its exceptional epigenetic properties. The *APOE* $\epsilon 2/\epsilon 3/\epsilon 4$ alleles are produced by two cytosine-phosphate-guanine (CpG)-altering SNPs (rs429358 and rs7412) residing within the core region of the *APOE* CpG islands. *APOE* $\epsilon 4$ carriers have the greatest number of CpG dinucleotides sites, while *APOE* $\epsilon 2$ carriers have the smallest number, so methylation levels for most CpG sites are in the order of *APOE* $\epsilon 4$ carriers > *APOE* $\epsilon 3/\epsilon 3$ > *APOE* $\epsilon 2$ carriers. The role of *APOE* in AD pathogenesis involves not only amyloid- β (ab) aggregation and clearance, but also tau-mediated neurodegeneration, microglia dysfunction, astrocyte reactivity, and blood-brain barrier disruption.

In the epigenetic landscape, miRNAs are known to be small non-coding RNAs with a length of ~ 22 nucleotides They are also implicated in AD, as showed by the altered expression of miRNA 650

(miR-650) in AD brains [75]. Bioinformatic analysis showed that miR-650 may target the expression of three components associated to AD: *APOE*, presenilin 1 (PSEN1), and cyclin-dependent kinase 5 (CDK5), with recent findings confirming that miR-650 may reduce *in vitro* the expression of *APOE*, PSEN1, and CDK5 [75].

5. Epigenetics of apolipoprotein E and cognitive function: contrasting evidence in Alzheimer's disease

Several lifestyle and environmental stimuli could explain the effects of *APOE* genotype on AD and cognitive functioning, such as exercise [76], education [77], and vitamin D status [78]. Among implications for the development and progression of AD, vitamin D supplementation may be another potential strategy to consider for the *APOE* ϵ 4 allele-carriers. Some reports showed that higher vitamin D concentrations in *APOE* ϵ 4 homozygous carriers allow to perform better at memory scores [79]. Then, compared to the *APOE* ϵ 3/ ϵ 3-carriers, the *APOE* ϵ 4-carriers showed earlier onset of cognitive impairment in AD. Although, after the disease onset, the effect of *APOE* genotype on the progression of cognitive impairment remained debated [80].

For this reason, epigenetic modifications of *APOE* such as DNA methylation have a central role in maintaining cognitive function in older age. Growing DNA methylation levels at the *APOE* promoter region were found on *postmortem* prefrontal cortex samples of sporadic AD individuals by mass spectrometry [81]. Numerous previous studies have investigated the association between *APOE* DNA methylation and AD or MCI [82–84]. Instead, the association between *APOE* DNA methylation and cognitive function in healthy subjects without cognitive impairment was evaluated by two studies with controversial findings [85,86]. Liu and colleagues found an inverse association between DNA methylation in the *APOE* gene region and delayed recall capacity among 289 older African Americans people with a mean age of 67 years during normal cognitive aging [85]. Conversely, the other study conducted in a large European cohort, observed no association between general cognitive functioning and *APOE* DNA methylation [86].

Many reports have suggested that neuroinflammation may have a key role in AD pathogenesis [87]. Dietary habits are known to influence systemic inflammation, neuroinflammation, and inflammaging [88]. A recent study conducted in a cohort of racially diverse middle-aged people ($n = 411$), pursued to identify DNA methylation sites associated with cognitive function in the genomic region of *APOE*. About inflammatory potential of the diet, among the dietary inflammatory index, cognitive performance, and the methylation level of several CpG sites have been detected significant relationships [89].

However, studies are contrasting at this regard, and if epigenetic biomarkers could be used for predicting AD is still unclear. In the *APOE* gene, DNA methylation at two CpG sites (3/13) that are known to show age-dependent changes, was related with the total cholesterol and high-density lipoprotein cholesterol ratio, but not with cognitive status, family history of AD, or the risk of cardiovascular disease in a blood-based DNA methylation study of 5828 people from the Generation Scotland cohort [90]. These findings supported that there is no evidence yet for considering *APOE* methylation as a biomarker for predicting AD or cardiovascular disease, although *APOE* methylation was associated with the blood levels of cholesterol [90].

6. Conclusions

In the panorama of current available evidence, the investigation of healthy aging and longevity is currently of remarkable interest. *APOE* could be considered an epigenetic mediator of senescence considering that different ApoE biochemical pathways in lipid metabolism, neuroinflammation and neurodegeneration may contribute to longevity and healthy aging. Nonetheless, such areas of investigation are still increasing, since ApoE function in neurodegenerative diseases such as AD cannot be uniquely explained by ApoE effects in lipid metabolism. Furthermore, the imbalance in the ApoE isoforms could explain the pathophysiological process of cognitive impairment linked to sporadic AD [91].

Stochastic factors (such environmental, diet, and pollution) may play a significant role in sporadic AD, despite the elevated lifetime risk linked to *APOE* $\epsilon 3/\epsilon 4$ and *APOE* $\epsilon 4/\epsilon 4$ genotypes. Indeed, according to the notion of stochastic risk or protective factors and although it is known that *APOE* $\epsilon 4/\epsilon 4$ -carriers developed dementia about 10 years earlier than *APOE* $\epsilon 2$ carriers [92], there was still significant discrepancy in the age of onset for *APOE* $\epsilon 4/\epsilon 4$ -carriers (standard deviation of 6 years) [93]. During the process of aging, the accumulation of molecular alterations driven by genetic and epigenetic events in the organism lead to a loss of phenotypic plasticity over time. Also, epigenetics may be altered during the process of aging, and it is particularly important as age is the greatest risk factor for developing AD [94].

Ethical Approval: This article does not contain any studies with human participants or animals performed by any of the authors.

Informed Consent: For this type of study, formal consent is not required.

Conflicts of Interest: The authors have no relevant affiliations or financial involvement with any organization or entity with a financial interest in or financial conflict with the subject matter or materials discussed in the manuscript. This includes employment, consultancies, honoraria, stock ownership or options, expert testimony, grants or patents received or pending, or royalties.

References

- Heyn, H. A symbiotic liaison between the genetic and epigenetic code. *Front. Genet.* **2014**, *5*, 113.
- Mill, J.; Heijmans, B.T. From promises to practical strategies in epigenetic epidemiology. *Nat. Rev. Genet.* **2013**, *14*, 585–594.
- Frisoni, G.B.; Altomare, D.; Thal, D.R.; Ribaldi, F.; van der Kant, R.; Ossenkoppele, R.; Blennow, K.; Cummings, J.; van Duijn, C.; Nilsson, P.M.; Dietrich, P.Y.; Scheltens, P.; Dubois, B. The probabilistic model of Alzheimer disease: the amyloid hypothesis revised. *Nat. Rev. Neurosci.* **2022**, *23*, 53–66.
- Coppieters, N.; Dragunow, M. Epigenetics in Alzheimer's disease: A focus on DNA modifications. *Curr. Pharm. Des.* **2011**, *17*, 3398–3412.
- Cavalli, G.; Heard, E. Advances in epigenetics link genetics to the environment and disease. *Nature* **2019**, *571*, 489–499.
- Zhang, D.; Cheng, L.; Badner, J.A.; Chen, C.; Chen, Q.; Luo, W.; Craig, D.W.; Redman, M.; Gershon, E.S.; Liu, C. Genetic control of individual differences in gene-specific methylation in human brain. *Am. J. Hum. Genet.* **2010**, *86*, 411–419.
- Gibbs, J.R.; van der Brug, M.P.; Hernandez, D.G.; Traynor, B.J.; Nalls, M.A.; Lai, S.L.; Arepalli, S.; Dillman, A.; Rafferty, I.P.; Troncoso, J.; Johnson, R.; Zielke, H.R.; Ferrucci, L.; Longo, D.L.; Cookson, M.R.; Singleton, A.B. Abundant quantitative trait loci exist for DNA methylation and gene expression in human brain. *PLoS Genet.* **2010**, *6*, e1000952.
- Shoemaker, R.; Deng, J.; Wang, W.; Zhang, K. Allele-specific methylation is prevalent and is contributed by CpG-SNPs in the human genome. *Genome Res.* **2010**, *20*, 883–889.
- Corder, E.H.; Saunders, A.M.; Strittmatter, W.J.; Schmechel, D.E.; Gaskell, P.C.; Small, G.W.; Roses, A.D.; Haines, J.L.; Pericak-Vance, M.A. Gene dose of apolipoprotein e type 4 allele and the risk of Alzheimer's disease in late onset families. *Science* **1993**, *261*, 921–923.
- Chartier-Harlin, M.C.; Parfitt, M.; Legrain, S.; Pérez-Tur, J.; Brousseau, T.; Evans, A.; Berr, C.; Odile V.; Roques, P.; Gourlet, V.; Fruchart, J.C.; Delacourte, A.; Rossor, M.; Amouyel, P. Apolipoprotein E, epsilon 4 allele as a major risk factor for sporadic early and late-onset forms of Alzheimer's disease: analysis of the 19q13,2 chromosomal region. *Hum. Mol. Genet.* **1994**, *3*, 569–574.
- Farrer, L.A.; Cupples, L.A.; Haines, J.L.; Hyman, B.; Kukull, W.A.; Mayeux, R.; Myers, R.H.; Pericak-Vance, M.A.; Risch, N.; van Duijn, C.M. Effects of age, sex, and ethnicity on the association between apolipoprotein e genotype and Alzheimer disease. a meta-analysis. APOE and Alzheimer disease meta-analysis consortium. *JAMA* **1997**, *278*, 1349–1356.
- Genin, E.; Hannequin, D.; Wallon, D.; Sleegers, K.; Hiltunen, M.; Combarros, O.; Bullido, M.J.; Engelborghs, S.; De Deyn, P.; Berr, C.; Pasquier, F.; Dubois, B.; Tognoni, G.; Fiévet, N.; Brouwers, N.; Bettens, K.; Arosio, B.; Coto, E.; Del Zompo, M.; Mateo, I.; Epelbaum, J.; Frank-Garcia, A.; Helisalmi, S.; Porcellini, E.; Pilotto, A.; Forti, P.; Ferri, R.; Scarpini, E.; Siciliano, G.; Solfrizzi, V.; Sorbi, S.; Spalletta, G.; Valdivieso, F.; Vepsäläinen, S.; Alvarez, V.; Bosco, P.; Mancuso, M.; Panza, F.; Nacmias, B.; Bossù, P.; Hanon, O.; Piccardi, P.; Annoni, G.; Seripa, D.; Galimberti, D.; Licastro, F.; Soininen, H.; Dartigues, J.F.; Kamboh, M.I.; Van Broeckhoven, C.; Lambert, J.C.; Amouyel, P.; Campion, D. APOE and Alzheimer disease: a major gene with semi-dominant inheritance. *Mol. Psychiatry* **2011**, *16*, 903–907.

13. Corder, E.H.; Saunders, A.M.; Risch, N.J.; Strittmatter, W.J.; Schmechel, D.E.; Gaskell, P.C. Jr; Rimmer, J. B.; Locke, P. A.; Conneally, P. M.; Schmechel, K. E.; Small, G. W.; Roses, A. D.; Haines, J. L.; Pericak-Vance, M. A. Protective effect of apolipoprotein e type 2 allele for late onset Alzheimer disease. *Nat. Genet.* **1994**, *7*, 180–184.
14. Panza, F.; Solfrizzi, V.; Torres, F.; Mastroianni, F.; Colacicco, A.M.; Basile, A.M.; Capurso, C.; D'Introno, A.; Del Parigi, A.; Capurso, A. Apolipoprotein E in Southern Italy: protective effect of epsilon 2 allele in early- and late-onset sporadic Alzheimer's disease. *Neurosci. Lett.* **2000**, *292*, 79–82.
15. Mayeux, R.; Saunders, A.M.; Shea, S.; Mirra, S.; Evans, D.; Roses, A.D.; Hyman, B.T.; Crain, B.; Tang, M.X.; Phelps, C.H. Utility of the apolipoprotein E genotype in the diagnosis of Alzheimer's disease. Alzheimer's disease centers consortium on apolipoprotein E and Alzheimer's disease. *N. Engl. J. Med.* **1998**, *338*, 506–511.
16. Seripa, D.; D'Onofrio, G.; Panza, F.; Cascavilla, L.; Masullo, C.; Pilotto, A. The genetics of the human APOE polymorphism. *Rejuvenation Res.* **2011**, *14*, 491–500.
17. Nickerson, D.A.; Taylor, S.L.; Fullerton, S.M.; Weiss, K.M.; Clark, A.G.; Stengård, J.H.; Salomaa, V.; Boerwinkle, E.; Sing, C.F. Sequence diversity and large-scale typing of SNPs in the human apolipoprotein E gene. *Genome Res.* **2000**, *10*, 1532–1545.
18. Yu, C.E.; Foraker, J. Epigenetic considerations of the APOE gene. *Biomol. Concepts* **2015**, *6*, 77–84.
19. Serrano-Pozo, A.; Das, S.; Hyman, B.T. APOE and Alzheimer's disease: advances in genetics, pathophysiology, and therapeutic approaches. *Lancet Neurol.* **2021**, *20*, 68–80.
20. Reiman, E.M.; Arboleda-Velasquez, J.F.; Quiroz, Y.T.; Huentelman, M.J.; Beach, T.G.; Caselli, R.J.; Chen, Y.; Su, Y.; Myers, A.J.; Hardy, J.; Paul Vonsattel, J.; Younkin, S.G.; Bennett, D.A.; De Jager, P.L.; Larson, E.B.; Crane, P.K.; Keene, C.D.; Kamboh, M.I.; Kofler, J.K.; Duque, L.; Gilbert, J.R.; Gwirtsman, H.E.; Buxbaum, J.D.; Dickson, D.W.; Frosch, M.P.; Ghetti, B.F.; Lunetta, K.L.; Wang, L.S.; Hyman, B.T.; Kukull, W.A.; Foroud, T.; Haines, J.L.; Mayeux, R.P.; Pericak-Vance, M.A.; Schneider, J.A.; Trojanowski, J.Q.; Farrer, L.A.; Schellenberg, G.D.; Beecham, G.W.; Montine, T.J.; Jun, G.R.; Alzheimer's Disease Genetics Consortium. Exceptionally low likelihood of Alzheimer's dementia in APOE2 homozygotes from a 5,000-person neuropathological study. *Nat. Commun.* **2020**, *11*, 667.
21. Salvadó, G.; Grothe, M.J.; Groot, C.; Moscoso, A.; Schöll, M.; Gispert, J.D.; Ossenkoppele, R.; Alzheimer's Disease Neuroimaging Initiative. Differential associations of APOE-ε2 and APOE-ε4 alleles with PET-measured amyloid-β and tau deposition in older individuals without dementia. *Eur. J. Nucl. Med. Mol. Imaging* **2021**, *48*, 2212–2224.
22. Theriault, J.; Benedet, A.L.; Pascoal, T.A.; Mathotaarachchi, S.; Chamoun, M.; Savard, M.; Thomas, E.; Kang, M.S.; Lussier, F.; Tissot, C.; Parsons, M.; Qureshi, M.N.I.; Vitali, P.; Massarweh, G.; Soucy, J.P.; Rej, S.; Saha-Chaudhuri, P.; Gauthier, S.; Rosa-Neto, P. Association of Apolipoprotein E 4 With Medial Temporal Tau Independent of Amyloid-β. *JAMA Neurol.* **2020**, *77*, 470–479.
23. Krasemann, S.; Madore, C.; Cialic, R.; Baufeld, C.; Calcagno, N.; El Fatimy, R.; Beckers, L.; O'Loughlin, E.; Xu, Y.; Fanek, Z.; Greco, D.J.; Smith, S.T.; Tweet, G.; Humulock, Z.; Zrzavy, T.; Conde-Sanroman, P.; Gacias, M.; Weng, Z.; Chen, H.; Tjon, E.; Mazaheri, F.; Hartmann, K.; Madi, A.; Ulrich, J.D.; Glatzel, M.; Worthmann, A.; Heeren, J.; Budnik, B.; Lemere, C.; Ikezu, T.; Heppner, F.L.; Litvak, V.; Holtzman, D.M.; Lassmann, H.; Weiner, H.L.; Ochando, J.; Haass, C.; Butovsky, O. The TREM2-APOE Pathway Drives the Transcriptional Phenotype of Dysfunctional Microglia in Neurodegenerative Diseases. *Immunity* **2017**, *47*, 566–81.e9.
24. Chen, Y.; Hong, T.; Chen, F.; Sun, Y.; Wang, Y.; Cui, L. Interplay between microglia and Alzheimer's disease-focus on the most relevant risks: APOE genotype, sex and age. *Front. Aging Neurosci.* **2021**, *13*, 631827.
25. Chung, W.S.; Verghese, P.B.; Chakraborty, C.; Joung, J.; Hyman, B.T.; Ulrich, J.D.; Holtzman, D.M.; Barres, B.A. Novel allele-dependent role for APOE in controlling the rate of synapse pruning by astrocytes. *Proc. Natl. Acad. Sci. U. S. A.* **2016**, *113*, 10186–10191.
26. Blanchard, J.W.; Bula, M.; Davila-Velderrain, J.; Akay, L.A.; Zhu, L.; Frank, A.; Victor, M.B.; Bonner, J.M.; Mathys, H.; Lin, Y.T.; Ko, T.; Bennett, D.A.; Cam, H.P.; Kellis, M.; Tsai, L.H. Reconstruction of the human blood-brain barrier in vitro reveals a pathogenic mechanism of APOE4 in pericytes. *Nat. Med.* **2020**, *26*, 952–963.
27. Riphagen, J.M.; Ramakers, I.H.G.M.; Freeze, W.M.; Pagen, L.H.G.; Hanseeuw, B.J.; Verbeek, M.M.; Verhey, F.R.J.; Jacobs, H.I.L. Linking APOE-ε4, blood-brain barrier dysfunction, and inflammation to Alzheimer's pathology. *Neurobiol. Aging* **2020**, *85*, 96–103.
28. Strittmatter, W.J.; Saunders, A.M.; Schmechel, D.; Pericak-Vance, M.; Enghild, J.; Salvesen, G.S.; Roses, A.D. Apolipoprotein e: high-avidity binding to beta-amyloid and increased frequency of type 4 allele in late-onset familial Alzheimer disease. *Proc. Natl. Acad. Sci. U.S.A.* **1993**, *90*, 1977–1981.
29. Fagan, A.M.; Holtzman, D.M. Astrocyte lipoproteins, effects of ApoE on neuronal function, and role of ApoE in amyloid-beta deposition in vivo. *Microsc. Res. Tech.* **2000**, *50*, 297–304.

30. Tokuda, T.; Calero, M.; Matsubara, E.; Vidal, R.; Kumar, A.; Permanne, B.; Zlokovic, B.; Smith, J.D.; Ladu, M.J.; Rostagno, A.; Frangione, B.; Ghiso, J. Lipidation of apolipoprotein e influences its isoform-specific interaction with Alzheimer's amyloid beta peptides. *Biochem. J.* **2002**, *348*, 359–365.
31. Deane, R.; Sagare, A.; Hamm, K.; Parisi, M.; Lane, S.; Finn, M.B.; Holtzman, D.M.; Zlokovic, B.V. ApoE isoform-specific disruption of amyloid beta peptide clearance from mouse brain. *J. Clin. Invest.* **2008**, *118*, 4002–40013.
32. Castellano, J.M.; Kim, J.; Stewart, F.R.; Jiang, H.; DeMattos, R.B.; Patterson, B.W.; Fagan, A.M.; Morris, J.C.; Mawuenyega, K.G.; Cruchaga, C.; Goate, A.M.; Bales, K.R.; Paul, S.M.; Bateman, R.J.; Holtzman, D.M. Human ApoE isoforms differentially regulate brain amyloid-beta peptide clearance. *Sci. Transl. Med.* **2011**, *3*, 89ra57.
33. Huang, Y.A.; Zhou, B.; Wernig, M.; Sudhof, T.C. ApoE2, ApoE3, and ApoE4 differentially stimulate APP transcription and A-beta secretion. *Cell* **2017**, *168*, 427.e21–41.e21.
34. Sala Frigerio, C.; Wolfs, L.; Fattorelli, N.; Thrupp, N.; Voytyuk, I.; Schmidt, I.; Mancuso, R.; Chen, W.T.; Woodbury, M.E.; Srivastava, G.; Möller, T.; Hudry, E.; Das, S.; Saido, T.; Karran, E.; Hyman, B.; Perry, V.H.; Fiers, M.; De Strooper, B. The Major Risk Factors for Alzheimer's Disease: Age, Sex, and Genes Modulate the Microglia Response to Ab Plaques. *Cell Rep.* **2019**, *27*, 1293–306.e6.
35. Shi, Y.; Holtzman, D.M. Interplay between innate immunity and Alzheimer disease: APOE and TREM2 in the spotlight. *Nat. Rev. Immunol.* **2018**, *18*, 759–772.
36. Zhao, N.; Liu, C.C.; Van Ingelgom, A.J.; Linares, C.; Kurti, A.; Knight, J.A.; Heckman, M.G.; Diehl, N.N.; Shinohara, M.; Martens, Y.A.; Attrebi, O.N.; Petrucelli, L.; Fryer, J.D.; Wszolek, Z.K.; Graff-Radford, N.R.; Caselli, R.J.; Sanchez-Contreras, M.Y.; Rademakers, R.; Murray, M.E.; Koga, S.; Dickson, D.W.; Ross, O.A.; Bu, G. APOE ϵ 2 is associated with increased tau pathology in primary tauopathy. *Nat. Commun.* **2018**, *9*, 4388.
37. Lozupone, M.; Panza F. Impact of apolipoprotein E isoforms on sporadic Alzheimer's disease: beyond the role of amyloid beta. *Neural Regen Res.* **2024**, *19*(1), 80–83.
38. Foraker, J.; Millard, S.P.; Leong, L.; Thomson, Z.; Chen, S.; Keene, C.D.; Bekris, L.M.; Yu, C.E. The APOE Gene is Differentially Methylated in Alzheimer's Disease. *J. Alzheimers Dis.* **2015**, *48*, 745–55.
39. Rueter, J.; Rimbach, G.; Huebbe, P. Allelic variation within the major APOE CpG island affects its methylation in the brain of targeted replacement mice expressing human APOE. *Biochim. Biophys. Acta Gene Regul. Mech.* **2023**, *1866*, 194942.
40. Lefterov, I.; Fitz, N.F.; Lu, Y.; Koldamova, R. APOE ϵ 4 and risk of Alzheimer's disease - time to move forward. *Front. Neurosci.* **2023**, *17*, 1195724.
41. Sullivan, P. M.; Mezdour, H.; Aratani, Y.; Knouff, C.; Najib, J.; Reddick, R. L.; Quarfordt, S.H.; Maeda, N. Targeted replacement of the mouse apolipoprotein E gene with the common human APOE3 allele enhances diet-induced hypercholesterolemia and atherosclerosis. *J. Biol. Chem.* **1997**, *272*, 17972–17980.
42. Livingston, G.; Huntley, J.; Sommerlad, A.; Ames, D.; Ballard, C.; Banerjee, S.; Brayne, C.; Burns, A.; Cohen-Mansfield, J.; Cooper, C.; Costafreda, S.G.; Dias, A.; Fox, N.; Gitlin, L.N.; Howard, R.; Kales, H.C.; Kivimäki, M.; Larson, E.B.; Ogunniyi, A.; Orgeta, V.; Ritchie, K.; Rockwood, K.; Sampson, E.L.; Samus, Q.; Schneider, L.S.; Selbæk, G.; Teri, L.; Mukadam, N. Dementia prevention, intervention, and care: 2020 report of the Lancet Commission. *Lancet* **2020**, *396*, 413–446.
43. Reas, E.T.; Laughlin, G.A.; Bergstrom, J.; Kritz-Silverstein, D.; Barrett-Connor, E.; McEvoy, L.K. Effects of APOE on cognitive aging in community-dwelling older adults. *Neuropsychology* **2019**, *33*, 406–416.
44. Mahley, R.W. Apolipoprotein E: cholesterol transport protein with expanding role in cell biology. *Science* **1988**, *240*, 622–630.
45. Siest, G.; Pillot, T.; Regis-Bailly, A.; Leininger-Muller, B.; Steinmetz, J.; Galteau, M.M.; Visvikis, S. Apolipoprotein E: An important gene and protein to follow in laboratory medicine. *Clin. Chem.* **1995**, *41*, 1068–1086.
46. Seripa, D.; Franceschi, M.; Matera, M.G.; Panza, F.; Kehoe, P.G.; Gravina, C.; Orsitto, G.; Solfrizzi, V.; Di Minno, G.; Dallapiccola, B.; Pilotto, A. Sex differences in the association of apolipoprotein e and angiotensin-converting enzyme gene polymorphisms with healthy aging and longevity: a population-based study from Southern Italy. *J. Gerontol. A Biol. Sci. Med. Sci.* **2006**, *61*, 918–923.
47. Seripa, D.; Panza, F.; Franceschi, M.; D'Onofrio, G.; Solfrizzi, V.; Dallapiccola, B.; Pilotto, A. Non-apolipoprotein E and apolipoprotein E genetics of sporadic Alzheimer's disease. *Ageing Res. Rev.* **2009**, *8*, 214–236.
48. Brooks-Wilson, A.R. Genetics of healthy aging and longevity. *Hum. Genet.* **2013**, *132*, 1323–1338.
49. Sebastiani, P.; Gurinovich, A.; Nygaard, M.; Sasaki, T.; Sweigart, B.; Bae, H.; Andersen, S.L.; Villa, F.; Atzmon, G.; Christensen, K.; Arai, Y.; Barzilai, N.; Puca, A.; Christiansen, L.; Hirose, N.; Perls, T.T. APOE alleles and extreme human longevity. *J. Gerontol. A Biol. Sci. Med. Sci.* **2018**, *74*, 44–51.
50. Smith, J.D. Apolipoproteins and aging: emerging mechanisms. *Ageing Res. Rev.* **2002**, *1*, 345–365.
51. Feng, J.; Xiang, L.; Wan, G.; Qi, K.; Sun, L.; Huang, Z.; Zheng, C.; Lv, Z.; Hu, C.; Yang, Z. Is APOE ϵ 3 a favorable factor for the longevity: an association study in Chinese population. *J. Genet.* **2011**, *90*, 343–347.

52. Mahley, R.W.; Rall, S.C. Jr. Apolipoprotein E: far more than a lipid transport protein. *Annu. Rev. Genomics Hum. Genet.* **2000**, *1*, 507–537.
53. Yanagisawa, K. Cholesterol and pathological processes in Alzheimer's disease. *J. Neurosci. Res.* **2002**, *70*, 361–366.
54. Ordovas, J.M.; Lopez-Miranda, J.; Mata, P.; Perez-Jimenez, F.; Lichtenstein, A.H.; Schaefer, E.J. Gene-diet interaction in determining plasma lipid response to dietary intervention. *Atherosclerosis* **1995**, *118* (Suppl), S11–S27.
55. Grimm, M.O.W.; Michaelson, D.M.; Hartmann, T. Omega-3 fatty acids, lipids, and ApoE lipidation in Alzheimer's disease: a rationale for multi-nutrient dementia prevention. *J. Lipid Res.* **2017**, *58*, 2083–2101.
56. Bos, M.M.; Noordam, R.; Blauw, G.J.; Slagboom, P.E.; Rensen, P.C.N.; van Heemst, D. The ApoE ϵ 4 Isoform: Can the Risk of Diseases be Reduced by Environmental Factors? *J. Gerontol. A Biol. Sci. Med. Sci.* **2019**, *74*, 99–107.
57. Paik, Y.K.; Chang, D.J.; Reardon, C.A.; Walker, M.D.; Taxman, E.; Taylor, J.M. Identification and characterization of transcriptional regulatory regions associated with expression of the human apolipoprotein E gene. *J. Biol. Chem.* **1988**, *263*, 13340–13349.
58. Mui, S.; Briggs, M.; Chung, H.; Wallace, R. B.; Gomez-Isla, T.; Rebeck, G. W.; Hyman, B.T. A newly identified polymorphism in the apolipoprotein E enhancer gene region is associated with Alzheimer's disease and strongly with the epsilon 4 allele. *Neurology* **1996**, *47*, 196–201.
59. Lambert, J. C.; Berr, C.; Pasquier, F.; Delacourte, A.; Frigard, B.; Cottel, D.; Pérez-Tur, J.; Mouroux, V.; Mohr, M.; Cécyre, D.; Galasko, D.; Lendon, C.; Poirier, J.; Hardy, J.; Mann, D.; Amouyel, P.; Chartier-Harlin, M.C. Pronounced impact of Th1/E47cs mutation compared with-491 AT mutation on neural APOE gene expression and risk of developing Alzheimer's disease. *Hum. Mol. Genet.* **1998**, *7*, 1511–1516.
60. Lambert, J. C.; Brousseau, T.; Defosse, V.; Evans, A.; Arveiler, D.; Ruidavets, J. B.; Haas, B.; Cambou, J.P.; Luc, G.; Ducimetière, P.; Cambien, F.; Chartier-Harlin, M.C.; Amouyel, P. Independent association of an APOE gene promoter polymorphism with increased risk of myocardial infarction and decreased APOE plasma concentrations-the ECTIM study. *Hum. Mol. Genet.* **2000**, *9*, 57–61.
61. Lambert, J. C.; Pasquier, F.; Cottel, D.; Frigard, B.; Amouyel, P.; Chartier-Harlin, M. C. A new polymorphism in the APOE promoter associated with risk of developing Alzheimer's disease. *Hum. Mol. Genet.* **1998**, *7*, 533–540.
62. Lambert, J. C.; Perez-Tur, J.; Dupire, M. J.; Galasko, D.; Mann, D.; Amouyel, P.; Hardy, J.; Delacourte, A.; Chartier-Harlin, M.C. Distortion of allelic expression of apolipoprotein E in Alzheimer's disease. *Hum. Mol. Genet.* **1997**, *6*, 2151–2154.
63. Bullido, M. A. J.; and Valdivieso, F. Apolipoprotein E gene promoter polymorphisms in Alzheimer's disease. *Microsc. Res. Tech.* **2000**, *50*, 261–267.
64. Lumsden, A. L.; Mulugeta, A.; Zhou, A.; Hypponen, E. Apolipoprotein E (APOE) genotype-associated disease risks: a phenome-wide, registry-based, case-control study utilising the UK biobank. *EBioMedicine* **2020**, *59*, 102954.
65. Artiga, M. J.; Bullido, M. J.; Frank, A.; Sastre, I.; Recuero, M.; García, M. A.; Lendon, C.L.; Han, S.W.; Morris, J.C.; Vázquez, J.; Goate, A.; Valdivieso, F. Risk for Alzheimer's disease correlates with transcriptional activity of the APOE gene. *Hum. Mol. Genet.* **1998**, *7*, 1887–1892.
66. Sims, R.; van der Lee, S. J.; Naj, A. C.; Bellenguez, C.; Badarinarayan, N.; Jakobsdottir, J.; Kunkle, B.W.; Boland, A.; Raybould, R.; Bis, J.C.; Martin, E.R.; Grenier-Boley, B.; Heilmann-Heimbach, S.; Chouraki, V.; Kuzma, A.B.; Sleegers, K.; Vronskaya, M.; Ruiz, A.; Graham, R.R.; Olsos, R.; Hoffmann, P.; Grove, M.L.; Vardarajan, B.N.; Hiltunen, M.; Nöthen, M.M.; White, C.C.; Hamilton-Nelson, K.L.; Epelbaum, J.; Maier, W.; Choi, S.H.; Beecham, G.W.; Dulary, C.; Herms, S.; Smith, A.V.; Funk, C.C.; Derbois, C.; Forstner, A.J.; Ahmad, S.; Li, H.; Bacq, D.; Harold, D.; Satizabal, C.L.; Valladares, O.; Squassina, A.; Thomas, R.; Brody, J.A.; Qu, L.; Sánchez-Juan, P.; Morgan, T.; Wolters, F.J.; Zhao, Y.; García, F.S.; Denning, N.; Fornage, M.; Malamon, J.; Naranjo, M.C.D.; Majounie, E.; Mosley, T.H.; Dombroski, B.; Wallon, D.; Lupton, M.K.; Dupuis, J.; Whitehead, P.; Fratiglioni, L.; Medway, C.; Jian, X.; Mukherjee, S.; Keller, L.; Brown, K.; Lin, H.; Cantwell, L.B.; Panza, F.; McGuinness, B.; Moreno-Grau, S.; Burgess, J.D.; Solfrizzi, V.; Proitsi, P.; Adams, H.H.; Allen, M.; Seripa, D.; Pastor, P.; Cupples, L.A.; Price, N.D.; Hannequin, D.; Frank-García, A.; Levy, D.; Chakrabarty, P.; Caffarra, P.; Giegling, I.; Beiser, A.S.; Giedraitis, V.; Hampel, H.; Garcia, M.E.; Wang, X.; Lannfelt, L.; Mecocci, P.; Eiriksdottir, G.; Crane, P.K.; Pasquier, F.; Boccardi, V.; Henández, I.; Barber, R.C.; Scherer, M.; Tarraga, L.; Adams, P.M.; Leber, M.; Chen, Y.; Albert, M.S.; Riedel-Heller, S.; Emilsson, V.; Beekly, D.; Braae, A.; Schmidt, R.; Blacker, D.; Masullo, C.; Schmidt, H.; Doody, R.S.; Spalletta, G.; Longstreth, W.T. Jr.; Fairchild, T.J.; Bossù, P.; Lopez, O.L.; Frosch, M.P.; Sacchinelli, E.; Ghetti, B.; Yang, Q.; Huebinger, R.M.; Jessen, F.; Li, S.; Kamboh, M.I.; Morris, J.; Sotolongo-Grau, O.; Katz, M.J.; Corcoran, C.; Dunstan, M.; Braddel, A.; Thomas, C.; Meggy, A.; Marshall, R.; Gerrish, A.; Chapman, J.; Aguilar, M.; Taylor, S.; Hill, M.; Fairén, M.D.; Hodges, A.; Vellas, B.; Soininen, H.; Kloszewska, I.; Daniilidou, M.; Uphill, J.; Patel, Y.; Hughes, J.T.; Lord, J.; Turton, J.; Hartmann, A.M.; Cecchetti, R.; Fenoglio, C.; Serpente, M.; Arcaro, M.; Caltagirone, C.; Orfei, M.D.; Ciarrella, A.; Pichler, S.; Mayhaus, M.; Gu, W.; Lleó, A.; Fortea,

- J.; Blesa, R.; Barber, I.S.; Brookes, K.; Cupidi, C.; Maletta, R.G.; Carrell, D.; Sorbi, S.; Moebus, S.; Urbano, M.; Pilotto, A.; Kornhuber, J.; Bosco, P.; Todd, S.; Craig, D.; Johnston, J.; Gill, M.; Lawlor, B.; Lynch, A.; Fox, N.C.; Hardy, J.; ARUK Consortium; Albin, R.L.; Apostolova, L.G.; Arnold, S.E.; Asthana, S.; Atwood, C.S.; Baldwin, C.T.; Barnes, L.L.; Barral, S.; Beach, T.G.; Becker, J.T.; Bigio, E.H.; Bird, T.D.; Boeve, B.F.; Bowen, J.D.; Boxer, A.; Burke, J.R.; Burns, J.M.; Buxbaum, J.D.; Cairns, N.J.; Cao, C.; Carlson, C.S.; Carlsson, C.M.; Carney, R.M.; Carrasquillo, M.M.; Carroll, S.L.; Diaz, C.C.; Chui, H.C.; Clark, D.G.; Cribbs, D.H.; Crocco, E.A.; DeCarli, C.; Dick, M.; Duara, R.; Evans, D.A.; Faber, K.M.; Fallon, K.B.; Fardo, D.W.; Farlow, M.R.; Ferris, S.; Foroud, T.M.; Galasko, D.R.; Gearing, M.; Geschwind, D.H.; Gilbert, J.R.; Graff-Radford, N.R.; Green, R.C.; Growdon, J.H.; Hamilton, R.L.; Harrell, L.E.; Honig, L.S.; Huentelman, M.J.; Hulette, C.M.; Hyman, B.T.; Jarvik, G.P.; Abner, E.; Jin, L.W.; Jun, G.; Karydas, A.; Kaye, J.A.; Kim, R.; Kowall, N.W.; Kramer, J.H.; LaFerla, F.M.; Lah, J.J.; Leverenz, J.B.; Levey, A.I.; Li, G.; Lieberman, A.P.; Lunetta, K.L.; Lyketsos, C.G.; Marson, D.C.; Martiniuk, F.; Mash, D.C.; Masliah, E.; McCormick, W.C.; McCurry, S.M.; McDavid, A.N.; McKee, A.C.; Mesulam, M.; Miller, B.L.; Miller, C.A.; Miller, J.W.; Morris, J.C.; Murrell, J.R.; Myers, A.J.; O'Bryant, S.; Olichney, J.M.; Pankratz, V.S.; Parisi, J.E.; Paulson, H.L.; Perry, W.; Peskind, E.; Pierce, A.; Poon, W.W.; Potter, H.; Quinn, J.F.; Raj, A.; Raskind, M.; Reisberg, B.; Reitz, C.; Ringman, J.M.; Roberson, E.D.; Rogaeva, E.; Rosen, H.J.; Rosenberg, R.N.; Sager, M.A.; Saykin, A.J.; Schneider, J.A.; Schneider, L.S.; Seeley, W.W.; Smith, A.G.; Sonnen, J.A.; Spina, S.; Stern, R.A.; Swerdlow, R.H.; Tanzi, R.E.; Thornton-Wells, T.A.; Trojanowski, J.Q.; Troncoso, J.C.; Van Deerlin, V.M.; Van Eldik, L.J.; Vinters, H.V.; Vonsattel, J.P.; Weintraub, S.; Welsh-Bohmer, K.A.; Wilhelmsen, K.C.; Williamson, J.; Wingo, T.S.; Woltjer, R.L.; Wright, C.B.; Yu, C.E.; Yu, L.; Garzia, F.; Golamaully, F.; Septier, G.; Engelborghs, S.; Vandenbergh, R.; De Deyn, P.P.; Fernandez, C.M.; Benito, Y.A.; Thonberg, H.; Forsell, C.; Lilius, L.; Kinhult-Ståhlbom, A.; Kilander, L.; Brundin, R.; Concar, L.; Helisalmi, S.; Koivisto, A.M.; Haapasalo, A.; Dermecourt, V.; Fievet, N.; Hanon, O.; Dufouil, C.; Brice, A.; Ritchie, K.; Dubois, B.; Himali, J.J.; Keene, C.D.; Tschanz, J.; Fitzpatrick, A.L.; Kukull, W.A.; Norton, M.; Aspelund, T.; Larson, E.B.; Munger, R.; Rotter, J.I.; Lipton, R.B.; Bullido, M.J.; Hofman, A.; Montine, T.J.; Coto, E.; Boerwinkle, E.; Petersen, R.C.; Alvarez, V.; Rivadeneira, F.; Reiman, E.M.; Gallo, M.; O'Donnell, C.J.; Reisch, J.S.; Bruni, A.C.; Royall, D.R.; Dichgans, M.; Sano, M.; Galimberti, D.; St George-Hyslop, P.; Scarpini, E.; Tsuang, D.W.; Mancuso, M.; Bonuccelli, U.; Winslow, A.R.; Daniele, A.; Wu, C.K.; GERAD/PERADES; CHARGE; ADGC; EADI; Peters, O.; Nacmias, B.; Riemenschneider, M.; Heun, R.; Brayne, C.; Rubinsztein, D.C.; Bras, J.; Guerreiro, R.; Al-Chalabi, A.; Shaw, C.E.; Collinge, J.; Mann, D.; Tsolaki, M.; Clarimón, J.; Sussams, R.; Lovestone, S.; O'Donovan, M.C.; Owen, M.J.; Behrens, T.W.; Mead, S.; Goate, A.M.; Uitterlinden, A.G.; Holmes, C.; Cruchaga, C.; Ingelsson, M.; Bennett, D.A.; Powell, J.; Golde, T.E.; Graff, C.; De Jager, P.L.; Morgan, K.; Ertekin-Taner, N.; Combarros, O.; Psaty, B.M.; Passmore, P.; Younkin, S.G.; Berr, C.; Gudnason, V.; Rujescu, D.; Dickson, D.W.; Dartigues, J.F.; DeStefano, A.L.; Ortega-Cubero, S.; Hakonarson, H.; Campion, D.; Boada, M.; Kauwe, J.K.; Farrer, L.A.; Van Broeckhoven, C.; Ikram, M.A.; Jones, L.; Haines, J.L.; Tzourio, C.; Launer, L.J.; Escott-Price, V.; Mayeux, R.; Deleuze, J.F.; Amin, N.; Holmans, P.A.; Pericak-Vance, M.A.; Amouyel, P.; van Duijn, C.M.; Ramirez, A.; Wang, L.S.; Lambert, J.C.; Seshadri, S.; Williams, J.; Schellenberg, G.D. Rare coding variants in PLCG2, ABI3, and TREM2 implicate microglial-mediated innate immunity in Alzheimer's disease. *Nat. Genet.* **2017**, *49*, 1373–1384.
67. Lee, E.G.; Tulloch, J.; Chen, S.; Leong, L.; Saxton, A. D.; Kraemer, B.; Darvas, M.; Keene, C.D.; Shutes-David, A.; Todd, K.; Millard, S.; Yu, C.E. Redefining transcriptional regulation of the APOE gene and its association with Alzheimer's disease. *PLoS One* **2020**, *15*, e0227667.
 68. Medvedeva, Y.A.; Fridman, M.V.; Oparina, N.J.; Malko, D.B.; Ermakova, E.O.; Kulakovskiy, I.V.; Heinzl, A.; Makeev, V.J. Intergenic, gene terminal, and intragenic CpG islands in the human genome. *BMC Genomics* **2010**, *11*, 48.
 69. Maunakea, A.K.; Nagarajan, R.P.; Bilenky, M.; Ballinger, T.J.; D'Souza, C.; Fouse, S.D.; Johnson, B.E.; Hong, C.; Nielsen, C.; Zhao, Y.; Turecki, G.; Delaney, A.; Varhol, R.; Thiessen, N.; Shchors, K.; Heine, V.M.; Rowitch, D.H.; Xing, X.; Fiore, C.; Schillebeeckx, M.; Jones, S.J.; Haussler, D.; Marra, M.A.; Hirst, M.; Wang, T.; Costello, J.F. Conserved role of intragenic DNA methylation in regulating alternative promoters. *Nature* **2010**, *466*, 253–257.
 70. Gellersen, H.M.; Guell, X.; Sami, S. Differential vulnerability of the cerebellum in healthy ageing and Alzheimer's disease. *Neuroimage Clin.* **2021**, *30*, 102605.
 71. Yu, C.E.; Cudaback, E.; Foraker, J.; Thomson, Z.; Leong, L.; Lutz, F.; Gill, J.A.; Saxton, A.; Kraemer, B.; Navas, P.; Keene, C.D.; Montine, T.; Bekris, L.M. Epigenetic signature and enhancer activity of the human APOE gene. *Hum. Mol. Genet.* **2013**, *22*, 5036–5047.
 72. Hendrich, B.; Bird, A. Identification and characterization of a family of mammalian methyl-CpG binding proteins. *Mol. Cell. Biol.* **1998**, *18*, 6538–47.
 73. Cedar, H.; Bergman, Y. Programming of DNA methylation patterns. *Annu. Rev. Biochem.* **2012**, *81*, 97–117.
 74. Ma, Y.; Smith, C.E.; Lai, C.Q.; Irvin, M.R.; Parnell, L.D.; Lee, Y.C.; Pham, L.; Aslibekyan, S.; Claas, S.A.; Tsai, M.Y.; Borecki, I.B.; Kabagambe, E.K.; Berciano, S.; Ordovás, J.M.; Absher, D.M.; Arnett, D.K. Genetic

- variants modify the effect of age on APOE methylation in the Genetics of Lipid Lowering Drugs and Diet Network study. *Aging Cell* **2015**, *14*, 49-59.
75. Lin, L.; Liu, X.; Cheng, X.; Li, Y.; Gearing, M.; Levey, A.; Huang, X.; Li, Y.; Jin, P.; Li, X. MicroRNA-650 Regulates the Pathogenesis of Alzheimer's Disease Through Targeting Cyclin-Dependent Kinase 5. *Mol. Neurobiol.* **2023**, *60*, 2426-2441.
 76. De Marco, M.; Clough, P.J.; Dyer, C.E.; Vince, R.V.; Waby, J.S.; Midgley, A.W.; Venneri, A. Apolipoprotein E epsilon allele modulates the immediate impact of acute exercise on prefrontal function. *Behav. Genet.* **2014**, *45*, 106-16.
 77. Cook, C.J.; Fletcher, J.M. Can education rescue genetic liability for cognitive decline? *Social Sci. Med.* **2015**, *127*, 159-70.
 78. Maddock, J.; Cavadino, A.; Power, C.; Hyppönen, E. 25-hydroxyvitamin D, APOE ε4 genotype and cognitive function: findings from the 1958 British birth cohort. *Eur. J. Clin. Nutr.* **2015**, *69*, 505-8.
 79. Panza, F.; La Montagna, M.; Lampignano, L.; Zupo, R.; Bortone, I.; Castellana, F.; Sardone, R.; Borracono, L.; Dibello, V.; Resta, E.; Altamura, M.; Daniele, A.; Lozupone, M. Vitamin D in the development and progression of alzheimer's disease: implications for clinical management. *Expert Rev. Neurother.* **2021**, *2*, 287-301.
 80. Suzuki, K.; Hirakawa, A.; Ihara, R.; Iwata, A.; Ishii, K.; Ikeuchi, T.; Sun, C.K.; Donohue, M.; Iwatsubo, T.; Alzheimer's Disease Neuroimaging Initiative, Japanese Alzheimer's Disease Neuroimaging Initiative. Effect of apolipoprotein E ε4 allele on the progression of cognitive decline in the early stage of Alzheimer's disease. *Alzheimers Dement. (NY)* **2020**, *6*, e12007.
 81. Wang, S.C.; Oelze, B.; Schumacher, A. Age-specific epigenetic drift in late-onset Alzheimer's disease. *PLoS One* **2008**, *3*, e2698.
 82. Karlsson, I.K.; Ploner, A.; Wang, Y.; Gatz, M.; Pedersen, N.L.; Hagg, S. Apolipoprotein E DNA methylation and late-life disease. *Int. J. Epidemiol.* **2018**, *47*, 899-907.
 83. Shao, Y.; Shaw, M.; Todd, K.; Khrestian, M.; D'Aleo, G.; Barnard, P.J.; Zahratka, J.; Pillai, J.; Yu, C.E.; Keene, C.D.; Leverenz, J.B.; Bekris, L.M. DNA methylation of TOMM40-APOE-APOC2 in Alzheimer's disease. *J. Hum. Genet.* **2018**, *63*, 459-471.
 84. Mancera-Paez, O.; Estrada-Orozco, K.; Mahecha, M.F.; Cruz, F.; Bonilla-Vargas, K.; Sandoval, N.; Guerrero, E.; Salcedo-Tacuma, D.; Melgarejo, J.D.; Vega, E.; Ortega-Rojas, J.; Roman, G.C.; Pardo-Turriago, R.; Arboleda, H. Differential methylation in APOE (Chr19; exon four; from 44,909,188 to 44,909,373/hg38) and increased apolipoprotein E plasma levels in subjects with mild cognitive impairment. *Int. J. Mol. Sci.* **2019**, *20* (6).
 85. Liu, J.; Zhao, W.; Ware, E.B.; Turner, S.T.; Mosley, T.H.; Smith, J.A. DNA methylation in the APOE genomic region is associated with cognitive function in African Americans. *BMC Med. Genomics* **2018**, *11*, 43.
 86. Mur, J.; McCartney, D.L.; Walker, R.M.; Campbell, A.; Bermingham, M.L.; Morris, S.W.; Porteous, D.J.; McIntosh, A.M.; Deary, I.J.; Evans, K.L.; Marioni, R.E. DNA methylation in APOE: The relationship with Alzheimer's and with cardiovascular health. *Alzheimers Dement. (NY)* **2020**, *6*, e12026.
 87. Heneka, M.T.; Carson, M.J.; El Khoury, J.; Landreth, G.E.; Brosseron, F.; Feinstein, D.L.; Jacobs, A.H.; Wyss-Coray, T.; Vitorica, J.; Ransohoff, R.M.; Herrup, K.; Frautschy, S.A.; Finsen, B.; Brown, G.C.; Verkhratsky, A.; Yamanaka, K.; Koistinaho, J.; Latz, E.; Halle, A.; Petzold, G.C.; Town, T.; Morgan, D.; Shinohara, M.L.; Perry, V.H.; Holmes, C.; Bazan, N.G.; Brooks, D.J.; Hunot, S.; Joseph, B.; Deigenesch, N.; Garaschuk, O.; Boddeke, E.; Dinarello, C.A.; Breitner, J.C.; Cole, G.M.; Golenbock, D.T.; Kummer, M.P. Neuroinflammation in Alzheimer's disease. *Lancet Neurol.* **2015**, *14*, 388-405.
 88. Griseta, C.; Battista, P.; Castellana, F.; Colonna, I.; Sciarra, S.; Zupo, R.; Bortone, I.; Lampignano, L.; Tirelli, S.; Bernardino, G.; Mollica, A.; Lozupone, M.; Panza, F.; Fiore, P.; Minafra, B.; Sardone, R. Serum levels of IL-6 are associated with cognitive impairment in the salus in apulia population-based study. *Heliyon* **2023**, *9*, e13972.
 89. Shen, B.; Hernandez, D.G.; Chitralla, K.N.; Fanelli-Kuczmarski, M.T.; Noren Hooten, N.; Pacheco, N.L.; Mode, N.A.; Zonderman, A.B.; Ezike, N.; Evans, M.K. APOE gene region methylation is associated with cognitive performance in middle-aged urban adults. *Neurobiol. Aging* **2022**, *116*, 41-48.
 90. Mur, J.; McCartney, D.L.; Walker, R.M.; Campbell, A.; Bermingham, M.L.; Morris, S.W.; Porteous, D.J.; McIntosh, A.M.; Deary, I.J.; Evans, K.L.; Marioni, R.E. DNA methylation in APOE: The relationship with Alzheimer's and with cardiovascular health. *Alzheimers Dement. (NY)* **2020**, *6*, e12026.
 91. Lozupone, M.; Imbimbo, B.P.; Balducci, C.; Lo Vecchio, F.; Bisceglia, P.; Latino, R.R.; Leone, M.; Dibello, V.; Solfrizzi, V.; Greco, A.; Daniele, A.; Watling, M.; Seripa, D.; Panza, F. Does the imbalance in the apolipoprotein E isoforms underlie the pathophysiological process of sporadic Alzheimer's disease? *Alzheimers Dement. (NY)* **2023**, *19*, 353-368.
 92. Slooter, A. J. C.; Cruts, M.; Kalmijn, S.; Hofman, A.; Breteler, M.M.; Van Broeckhoven, C.; van Duijn, C.M. Risk estimates of dementia by apolipoprotein E genotypes from a population-based incidence study: The Rotterdam Study. *Arch. Neurol.* **1998**, *55*, 964-968.

93. Reiman, E. M.; Arboleda-Velasquez, J.F.; Quiroz, Y.T.; Huentelman, M.J.; Beach, T.G.; Caselli, R.J.; Chen, Y.; Su, Y.; Myers, A.J.; Hardy, J.; Paul Vonsattel, J.; Younkin, S.G.; Bennett, D.A.; De Jager, P.L.; Larson, E.B.; Crane, P.K.; Keene, C.D.; Kamboh, M.I.; Kofler, J.K.; Duque, L.; Gilbert, J.R.; Gwirtsman, H.E.; Buxbaum, J.D.; Dickson, D.W.; Frosch, M.P.; Ghetti, B.F.; Lunetta, K.L.; Wang, L.S.; Hyman, B.T.; Kukull, W.A.; Foroud, T.; Haines, J.L.; Mayeux, R.P.; Pericak-Vance, M.A.; Schneider, J.A.; Trojanowski, J.Q.; Farrer, L.A.; Schellenberg, G.D.; Beecham, G.W.; Montine, T.J.; Jun, G.R.; Alzheimer's Disease Genetics Consortium. Exceptionally low likelihood of Alzheimer's dementia in APOE2 homozygotes from a 5,000-person neuropathological study. *Nat. Commun.* **2020**, *11*, 667
94. Pogribny, I.P.; Vanyushin, B.F. Age-related Genomic Hypomethylation. In: Tollefsbol TO, ed. *Epigenetics of Aging*; Springer 2009, 11-27.

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.