

Article

Not peer-reviewed version

Antibacterial Aerogels-Based Membranes by Customized Colloidal Functionalization of TEMPO-Oxidized Cellulose Nanofibers Incorporating CuO

<u>Elena Usala</u>*, <u>Eduardo Espinosa</u>*, Wasim El Arfaoui , <u>Ramón Morcillo-Martín</u> , <u>Begoña Ferrari</u> , Zoilo Gonzalez

Posted Date: 17 October 2023

doi: 10.20944/preprints202310.1025.v1

Keywords: Colloidal heterostructures; functional aerogels; Cellulose Nanofibers; CuO nanoparticles; Antibacterial Membranes



Preprints.org is a free multidiscipline platform providing preprint service that is dedicated to making early versions of research outputs permanently available and citable. Preprints posted at Preprints.org appear in Web of Science, Crossref, Google Scholar, Scilit, Europe PMC.

Copyright: This is an open access article distributed under the Creative Commons Attribution License which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Disclaimer/Publisher's Note: The statements, opinions, and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions, or products referred to in the content.

Article

Antibacterial Aerogels-based Membranes by customized Colloidal Functionalization of TEMPO-Oxidized Cellulose Nanofibers incorporating CuO

E. Usala 1, E. Espinosa 1,3, W. El Arfaoui 1, R. Morcillo 1, B. Ferrari 2,3 and Zoilo Gonzalez 1,3*

- ¹ BioPren Group, Chemical Engineering Department, Instituto Químico para la Energía y el Medioambiente (IQUEMA), Universidad de Córdoba, 14014, Córdoba, Spain
- ² Instituto de Cerámica y Vidrio, CSIC, Campus de Cantoblanco, c/ Kelsen 5, 28049 Madrid, Spain
- ³ Unidad Asociada CSIC-UCO. Fabricación aditiva de materiales compuestos basados en celulosa funcionalizada, obtenida de residuos de biomasa
- * Correspondence: zgonzalez@uco.es

Abstract: An innovative colloidal approach is proposed here to carry out the customized functionalization of TEMPO-Oxidized Cellulose Nanofibers (CNF) incorporating non-noble inorganic nanoparticles. A heterocoagulation process is applied between the delignified CNF and as-synthetized CuO nanoparticles (CuO NPs) to formulate mixtures which are used in the preparation of aerogels with antibacterial effect, which could be used to manufacture membranes, filters, foams, etc. The involved components of formulated blending, CNF and CuO NPs, were individually obtained by using a biorefinery strategy for agricultural waste valorization together with an optimized chemical precipitation, assisted by ultrasounds. The optimization of synthesis parameters for CuO NPs have avoided the presence of undesirable species which usually requires of later thermal treatment with associated costs. The aerogels-based structure, obtained by conventional freeze-drying, acted as 3D support for CuO NPs providing a good dispersion within the cross-linked structure of the nanocellulose and facilitating direct contact of the antibacterial phase against undesirable microorganisms. All samples showed a positive response against Escherichia coli and Staphylococcus aureus. An increase of the antibacterial response of the aerogels, measured by agar disk diffusion test, has been observed with the increase of CuO NPs incorporated, obtaining width of the antimicrobial "halo" (nwhalo) from 0 to 0.6 and 0.35 for S. aureus and E. coli, respectively. Furthermore, the aerogels have been able to deactivate S. aureus and E. coli in less than 5 hours when the antibacterial assays have been analyzed by broth dilution method. From CNF-50CuO samples, an overlap nanoparticles effect produced a decrease of the antimicrobial kinetic.

Keywords: colloidal heterostructures; functional aerogels; Cellulose Nanofibers; CuO nanoparticles; antibacterial membranes

1. Introduction

The application of polymer-based porous structures (membranes, filters, foams) in wastewater treatment or in the concentration of targeted chemicals from aqueous solution is widespread. Nevertheless, the longstanding biofouling issue on structure surfaces (internal and external) has garnered enormous attention within the scientific and industrial fields. Motivated by the recent development of nanocomposite materials, the integration of antimicrobial nanophases within polymeric structures is emerging as a promising strategy to mitigate biofouling[1,2]

On the other hand, the environmental problems and the limited non-renewable resources is encouraging the search for alternative materials. The need to reduce the exploitation in fossil fuels drives the alternative use of others eco-friendly resources. Concerns about sustainability have attracted great interest in bioproducts and renewable materials, from biorefinery, as emerging solutions to satisfy a range of technological challenges[3].

In this sense, the manufacturing of composite materials, incorporating naturally derived polymers coming from valorization processes, have emerged as a promising class of engineering

materials providing new prospects for modern technology. The implementation of this type of energy-efficient is vital to develop the long-term bioeconomy concept.

Among all exploitation possibilities, the use of nanocellulose-based materials is very promising for their biocompatibility and unique properties such as low density, biodegradability, abundance, high-functionalization possibilities, and their vast applicability [4,5]. In addition to possessing the inherent properties of a polymer obtained from natural resources, cellulose nanofibers (CNF) can be combined with other functional component to gain certain added value. The combination of two or more constituents offers better properties than the individual components. In particular, the incorporation of metal oxide nanoparticles (MONPs) is considered an efficient approach to obtain potential features in the fields catalysis, water purification etc. [6]. Controlling the concentration and physicochemical characteristics (size, morphology, specific surface area, porosity, etc.) of the inorganic phases, the functionalities of composites materials can be tailored and improved. In this sense, the employment of ultrasound systems to assist the syntheses of MONPs is today considered an interesting way to regulate the nucleation and growth during their processes formation. The impact of ultrasound on crystallographic, textural and functional properties of MONPs has been intensively investigated in the last decades, revealing that final performances is influenced by the presence of cavitation bubbles that induces the ultrasonic waves[7].

In the last few years, many hybrid nanocellulose/MONPs formulations have been proposed to prepare functional composite as thin films, coated-papers, and other planar format [8,9]. But also, the manufacturing of three-dimensional networks, such as hydrogels, foams, crumpled shapes, and especially aerogels, have attracted special attention as potential framework or supporting matrix of MONPs. They really possess interesting characteristics such as immobilization ability, high specific surface area, structural integrity, mechanical robustness, high porosity, and easy separation of the material from reactions media[10,11].

The incorporation of copper oxide nanoparticles (CuO NPs) in nanohybrid structures has particularly studied in the last years due to the broad spectrum of applications. One of the most important characteristics is their antimicrobial activity against bacteria, fungi, viruses, and algae, which is extremely interesting for its use as polymer nanocomposite[12]. Moreover, the CuO can be considered a cheaper alternative than the conventional use of metal noble nanoparticles. Their inherent properties also interact closely with the microorganism membranes and exhibit competitive antimicrobial effects[13]. Some authors have already worked on the preparation of nanocellulose/CuO composites. For example Heidari, H et al. carried out an in-situ synthesis of CuO on bacterial cellulose and measured the antibacterial effects against strains of Klebsiella pneumonia and Staphylococcus aureus[13]. Almasi, H. et al. compared the efficiency against E.coli, P.aeruginosa and L.monocytogenes of bacterial cellulose nanofibers and chitosan nanofibers containing precipitated On the other hand, Shankar et al. demonstrated that the characteristics of CuO, synthesized in the presence of cellulose, were additionally affected by the type of cellulose used. They found that the source of cellulose influences the shape, size, and efficiency of cellulose-CuO composite[15].

In all cases, the incorporation of inorganic NPs, including the mentioned with CuO, has been carried by using conventional wet chemical methods such as sol-gel, co-precipitation, hydrothermal synthesis[16,17], where certain relevant processing parameters have not been considered in detail.

This is the reason why the application of an alternative colloidal approach could become in a design guide to achieve major control over manufacturing processes and therefore more efficient systems. The development of high homogeneous and well-dispersed components in suspension as well as the study of their electrostatic interactions allow controlling the mixtures in liquid media obtaining optimum distributions, which are completely relevant for the subsequent shaping processes[18,19].

Therefore, the present study focused on the customized preparation of functional CNF-based aerogels incorporating antibacterial CuO by using a facile colloidal methodology. The formulation of CNF-CuO blends was carried out by a heterocoagulation route described previously with other inorganic NPs. Furthermore, an analysis of the aerogel structure and their functional properties when

2

it is functionalized with different content of CuO. All samples have been characterized by XRD, FESEM and FTIR. In addition, the antimicrobial activities of the aerogels were determined against *Staphylococcus Aureus* and *Escherichia Coli* by Broth dilution method and Agar Diffusion method.

2. Materials and Methods

2.1. Preparation of CuO NPs and CNF suspensions

The CuO nanopowder used in this work was firstly synthetized by a chemical precipitation route, assisted by a high intensity ultrasonic horn for 30 minutes (SONOPULS HD 4200 BANDELIN using the Ti plate TT213, \emptyset = 13 mm with a power of 150W, Berlin, Germany). The procedure was made by dissolving the appropriate amount of a copper salt (0.1 mol·L⁻¹ of CuSO₄·5H₂O) in 20 ml of deionized water. The reaction was carried out by adding, drop by drop, 20 ml of potassium hydroxide solution of 0.3 mol·L⁻¹. Powder obtained was filtered using polymeric filters (\emptyset = 0.22 μ m), washed with deionized water and dried at 60°C for 24 h. The parameters were adjusted to avoid the presence of undesirable species and the application of later calcination processes. The yield of the synthesis was finally determined according to stoichiometric reaction.

The crystalline structure, interlaminar distance and crystallite size of the obtained powder was studied by XRD analysis using a D8 Advance diffractometer (Bruker, monochromatic Cu K α radiation λ =1.540598Å, 40 kV; 40 mA; 2h = 10–80). Nitrogen adsorption/desorption and specific surface area on the synthetized powder were measured at 77K using an adsorption analyzer (Autosorb-iQ-2 MP/XR, Quantachrome). A thermal characterization of the synthetized powder was also made by thermogravimetric analysis (5 °C min⁻¹, T = 25–600 °C, Air atmosphere, TGA/DSC 1 STAR system (Mettler Toledo)). The size and morphology of the CuO powder was also examined by FE-SEM JEOL JSM 6300 microscope (Boston).

On the other hand, TEMPO-Oxidized Cellulose nanofibers (CNF) were obtained from wheat straw residues by following a specific procedure described previously for the research group elsewhere [20,21].

Before blending the components of composite, individually colloidal stabilization analyses of CuO and CNF were made at different pH by zeta potential measurements. The colloidal dispersion of CuO was additionally studied by adsorbing different amount of a cationic polyelectrolyte as stabilizer additive. Branched polyethylenimine (PEI, Mw 25000 Sigma-Aldrich, Spain) was used for this purpose. All Zeta potential measurements were made by laser Doppler velocimetry in a ZetasizerNano ZS (Malvern, UK). The zeta potential evolution of NP with pH was determined for suspensions with concentrations of 0.01 g·L·¹using 10-² M KCl as solvent and inert electrolyte, to maintain the ionic strength of the medium. The pH adjustment of the suspensions was carried out by addition of small quantities of 0.1 M HNO3 or KOH and controlled with a pH probe (Metrohm AG, Germany). The zeta potential saturation curve determined for the addition of PEI was measured without the inert electrolyte at pH 8. Before each measurement, NP suspensions was dispersed and homogenized by sonication, using an ultrasonic horn (Sonopuls HD4200, 100W, Bandelin). On the other hand, the homogenization of CNF was made by using high-shear homogeniser (IKA T18 digital Ultra Turrax, IKA Germany).

2.2. Preparation of Nanocellulose/CuO Aerogels

The preparation of the CNF/CuO-PEI structures was carried out by adding the CuO-PEI suspension on the CNF hydrogel inducing heterocoagulation phenomenon. Both the surface charge of CNF as well as the exactly amount of CuO-PEI to completely cover the CNF surface were also monitored in terms of zeta potential. The pH of work was around 8 to increase the protonation of amine groups of PEI favoring the adsorption and the later interaction with the carboxyl/hydroxyl groups presented in nanofibers.

For this, varying amount of the stock suspensions of CNF (0.4 wt.%) and CuO-PEI, previously prepared as reference, were combined to obtain mixtures with different percentages (5, 10, 25, 50 and 100 wt.% CuO-PEI on dry weight of CNF. The obtained samples have been named as CNF-XCuO).

3

All blends were cast in PET molds with a 25 mm diameter, frozen for 24 h and freeze-dried at -85 °C under 0.5 mBar for 72 hours in a Lyoquest -85 device.

ATR-FTIR spectra of the compounds was carried out in a Perkin Elmer spectrometer, by mounting the samples onto its diamond cell and pressing with an adjustable screw. IR transmittance was measured in the range 400- 4000 cm⁻¹ with a resolution of 4 cm⁻¹.

The CuO NPs distributions on cellulose-based matrix as well as the microstructural morphology of the resulting drying aerogels were additionally observed with a field emission scanning electron microscope (FESEM, Hitachi S4700, Japan)

2.3. Antimicrobial analysis of the CNF/CuO aerogels

The antimicrobial properties of the produced aerogels were tested against two different microorganisms, the Gram-positive bacteria *S.aureus* and the Gram-negative bacteria *E.coli*.

2.3.1. Agar Disk Diffusion Method

To perform the agar disk diffusion tests the CNF/CuO aerogels were cut into round shapes of approximately 10 mm diameter and exposed to UV-light for 20 minutes. Lawns of the target microorganisms ($1x 10^8$ cfu/mL) were performed in the surface of the nutrient agar (NA) plates. The resulting aerogels were placed upon the lawns and kept at 37° C for 24h. After 24h the normalized width of the antimicrobial "halo" (nw_{halo}) was determined by applying the methodology described elsewhere [22] through the following equation:

$$nw_{halo} = [(d_{iz}-d)/2]/d \tag{1}$$

where d_{iz} is the diameter of the inhibition zone and d is the disk diameter.

2.3.2. Broth dilution Method

The antimicrobial activity in liquid media of the CNF/CuO aerogels was tested. First, an aliquot of the 40% (v/v) glycerol–ether frozen stock was cultured in nutrient broth (NB) at 37° C for 24 h. To obtain cells in the stationary growth phase, bacteria were subcultured at 37° C for 24 in fresh NB. The bacterial concentration was adjusted to 10^{8} CFU/mL using the McFarland 0.5 standard (0.1-0.08 absorbance at 625 nm). Tubes containing the aerogels with different amounts of NPs in NaCl solution (0.85% w/v) were prepared. Each test tube was inoculated with 100μ L of the previously standardized solution, obtaining a final concentration of approximately 10^{6} UFC/mL in the tubes. To confirm the initial bacterial concentration, a plate count of the standard tube was carried out. Afterwards, test tubes were incubated at the optimum conditions for each microorganism (37° C) and, subsequently, samples were taken from each tube at specific time intervals: 0, 1, 2.5, 5, 7 and 24h. For each tube the appropriate serial dilutions were performed, and 0.1 mL was surface plated in NA. After the incubation period (24h at 37° C) plates were counted. As negative controls, tubes containing CNF-100% aerogels + inoculum (CNF) and inoculum + NaCl solution (Control) were tested.

3. Results and Discussion

3.1. Synthesis and characterization of the involved components

The manufacturing of the antibacterial aerogels has been carried out by functionalizing of TEMPO-Oxidized CNF, prepared from the valorization of wheat straw residues by a standard biorefinery procedure described elsewhere [20,21]. In comparison with the CNF obtained by a mechanical procedure, these CNF presented excellent physicochemical properties with a high nanofibrillation yield value (~87%), high cationic demand and carboxyl content (~1210 and ~360 $\mu eq \cdot g^{-1}$ respectively) and large specific surface area (414 $m^2 \cdot g^{-1}$)

On the other hand, previous results showed that the purity, size, and morphology of CuO nanopowders, synthesized through a conventional precipitation reaction, are affected for the type of

4

5

precursor used [23]. Moreover, a calcination process above 450 °C is required to ensure the crystallinity and purification of the CuO specie [24].

Our work demonstrates that the application of a controlled amount of ultrasonic power in a concrete volume of precursor solution (W cm⁻² mol⁻¹) allowed the instantaneous formation of a pure CuO monoclinic phase in absence of thermal treatments. The effect of the applied ultrasounds lead to higher Gibbs free energy values avoiding the presence of intermediated precipitates, like Cu(OH)₂ or Cu₂O, which are normally obtained applying lower calcination temperatures (~250-300°C). In our case, the application of 30 minutes of ultrasound was enough to finish the precipitation reaction obtaining yields close to 95%.

The XRD plot in Figure 1a confirmed that the as-synthesized powder corresponds to the pure CuO monoclinic structure. The peaks located at $2\theta = 32.57^{\circ}$; 35.56° ; 38.77° ; 48.82° ; 58.27° ; 61.59° ; 66.36° and 67.92° are assigned to the characteristic orientation planes (110), (-111), (200), (-202), (020), (202), (-113) (-311) and (220) of CuO synthetized from CuSO₄·5H₂O [25]. In our case, the influence of sono-chemistry in the powder precipitation have avoided the presence of further undesirable species, like brochantite and posnjakite, usually detected by other authors in works in which the sulfate salt was involved as chemical precursors[23]. To analyze the effect of the ultrasonic power on the physicochemical features of the synthesized CuO NPs, crystallographic parameters were also determined. The interlaminar distance were calculated from the XRD plots obtaining values of 3.88 A. The crystallite size was additionally calculated according to the Warren modification of the Scherrer formula and using both the peak position and the full width at half maximum (FWHM) of the (-111) reflection. The result was around 19.52 nm, which was close to other values reported in previous work, but in absence of ultrasound. Therefore, we could conclude that there are no significant differences in the final crystallographic ordering. TGA curves depicted in Figure 2a also confirmed that the as-prepared powder did not present relevant weight losses and phase conversions during a later calcination process in air. The slight decrease of mass (4%) was attributed to superficially physiosorbed and interlamellar water losses as well as small amounts of remaining reactants.

Figure 1c illustrates the adsorption/desorption isotherm and provides details regarding the pore size distribution of the as-synthesized CuO powder calculated using the Barrett, Joyner, and Halenda (BJH) model. The isotherm profile corresponds to Type IV isotherms. The low adsorbed volume at very low relative pressure (P/Po < 0.1) confirms the limited presence of micropores, attributed to the absence of filling without adsorbent-adsorbate interactions. The CuO powder also absorbed a minimal amount of N₂, displaying a gradual increase in the relative pressure range of 0-0.8, indicating the absence of mesopores. However, a significant amount of gas is absorbed in the relative pressure range of 0.8–1, which is characteristic of certain macroporosity. This increase after P/P0 > 0.9 represents the overall porosity of CuO contributed by the interspaces among nanoparticles, providing a higher exposed surface area to the surrounding medium. The pore size distributions, plotted as a function of the adsorbed volume of N₂, confirm the presence of interspaces/macropores over a 30-70 nm range. The SSA of nanoparticles were finally determined using the Brunauer, Emmett, and Teller (BET) method obtaining a value of 49.46 m²·g⁻¹. All this info is in concordance with the morphological details revealed in the FESEM micrograph of Figure 1d. The image revealed short nanorod-like structures with a diameter of around 30 nm. Although the primary shape of particles is 1D nanorods, the microstructural detail shows a small agglomeration degree within the mentioned interspaces among them.

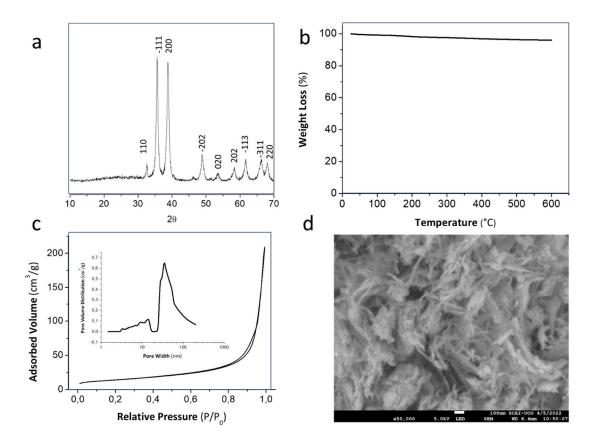


Figure 1. Characterization of as-synthetized CuONP: (A) XRD pattern (B) TGA curve, (C) N₂ adsorption/desorption isotherm and pore size distribution, and (D) FESEM micrographs.

To define the optimal conditions of colloidal processing, the evolution of zeta potential of assynthesized CuO NPs was firstly examined in a wide range of pH values (2 < pH < 11), Figure 2a. These CuO NPs exhibited an isoelectric point at pH 2.5 and a negative surface charge above this value. It is remarkable the zeta potential measurement at pH 8 was -33 mV, which is considered an idoneous condition to adsorb a cationic polyelectrolyte and improve the electrosteric effect of nanoparticles in dispersion. According to the literature[26], the branched PEI has a pKa value of 8.6, which means that their amine groups may be protonated at pH 8. The electrostatic interaction between the positive groups of PEI and the negative surface of CuO NPs was additionally followed by zeta potential analyses, Figure 2a. The negative surface of CuO NPs turned to positive with the addition of 1 wt. % of PEI and achieved a maximum of +40mV when the surfaces CuO NPs are stabilized with 6wt.% of PEI (CuO6PEI). Further colloidal characterization both CuO6PEI and asprepared CNF was carried out to check the surface charges in the liquid medium where they were then blended. The zeta potential values in function of pH have been also plotted in Figure 2b.

These results allow to select the appropriate pH conditions to ensure an effective electrostatic interaction between the negative charge of CNF and the positive surface of the CuO6PEI, Figure 2c. At pH 8, the zeta potential values were -45mV and +40mV respectively. Therefore, all mixtures formulated from both components in a unique suspension have been made at this working pH, promoting colloidal heterocoagulation phenomenon between them.

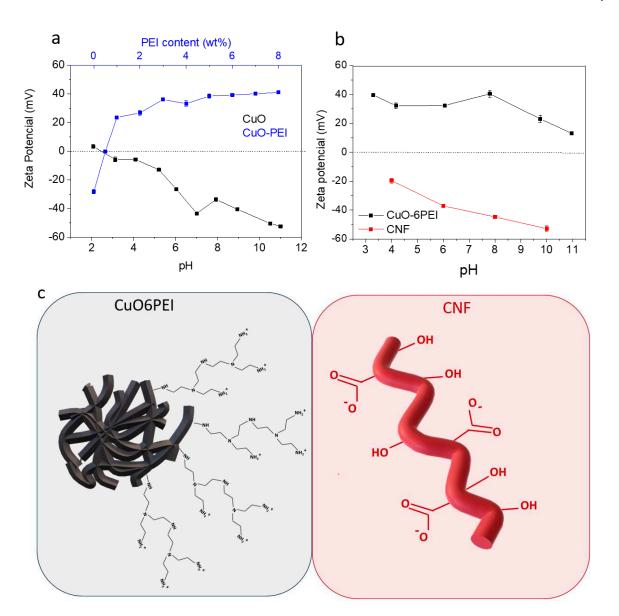


Figure 2. Evolution of Zeta potential as a function of amount of adsorbed PEI onto the particle surface and evolution of Zeta potential of CuO as a function of pH without PEI (A); Evolution of Zeta potential of CuO–6PEI and CNF as a function of pH (B); Scheme of the involved component representing their surface charge(C).

3.2. Blending Formulation and Aerogels Preparation

To define a specific ratio of the CNF/CuO6PEI formulations, different percentages of CuO6PEI were blended over the dry weight of CNF. The formation of different heterostructures was again monitored through the zeta potential evolution, Figure 3. The positive charged CuO6PEI approaches the negative sites of CNF provoking the neutralization of its functional groups. The negative zeta potential values of nude CNF turned towards a value of 22mV when the percentage of CuO6PEI added was 100wt.%. Then the values continued increasing until a maximum of 40mV when the added percentage was 1500 wt.%. No further variations were observed for higher additions of CuO6PEI.

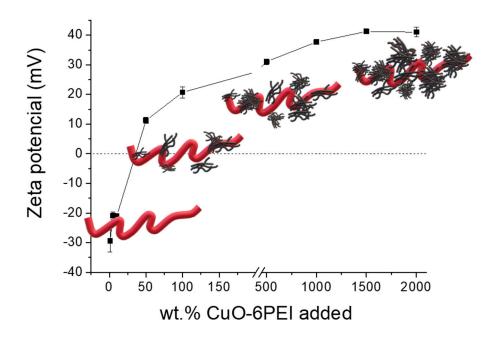


Figure 3. Evolution of Zeta potential as a function of amount of adsorbed CuO-6PEI onto the CNF.

To explore the formation of the CNF/CuO6PEI heterostructures and examine the antibacterial role of the CuO6PEI, six formulations, labelled as CNF, CNF-5CuO, CNF-10CuO, CNF-25CuO, CNF-50CuO, CNF-100CuO, were prepared by varying the percentage of added CuO-6PEI (0, 5, 10, 25, 50 and 100%) Notice that these compositions were chosen before and after turning the charge of CNF considering the antimicrobial concentrations of CuO NPs reported elsewhere[6,12,27].

After stirring, casting, and freeze-drying processes, the resulting aerogels were easily removed from the corresponding mold for further characterization, Figure 4a. FTIR spectra were acquired to obtain information about the electrostatic interaction between CuO-6PEI and CNF, Figure 4b. The characteristic cellulosic bands were observed at 3340 cm⁻¹ due to the stretching of –OH, at 2890 cm⁻¹ due to aliphatic C–H stretching, at 1606 cm⁻¹ due to C=O stretching of the carboxylate groups, at 1148 cm⁻¹ corresponds to deformation of the antisymmetric C–O–C bond, and a broad band between 1010 and 1050 cm⁻¹ was attributed to the C–OH bonds and the ether of the pyranose ring. Other peaks were additionally observed in the range of 1489–1420 cm⁻¹ due to single bond CH₂ scissoring motion in cellulose, 1368–1373 cm⁻¹ represents C-H bending, ~1317 cm⁻¹ shows CH₂ wagging, and a peak at 893 cm⁻¹ with improved sharpness associated with cellulosic β -glycosidic linkages. Similarly, peaks at 1154–1159 cm⁻¹ shows C-C ring stretching band[28].

If the comparative analysis is concretely focused on peaks corresponding to hydroxyl and carboxyl groups (3340 and 1600cm⁻¹ respectively) a notorious decrease in their intensity was appreciated when the amount of CuO6PEI was increased, whereas the prominence of peaks corresponding to the Cu(II)-O bond (at 604 and 499 cm⁻¹) presented an oppositely tendency and was higher with the increase of CuO concentration.

Recent studies have demonstrated that the increase and narrowing of the band corresponding to carboxyl groups (around 1600cm⁻¹) is associated to the amount of oxidative reactant during TEMPO pre-treatment [29]. However, the decrease and widening of this peak for the same TEMPO pre-treatment conditions (similarly to this study) are associated to the electrostatic interactions between CNF and CuO6PEI. These electrostatic interactions are produced between the negative charge of the hydroxyl/carboxyl groups belong to CNF and the positive charge of the PEI groups which are surface modifying the CuONPs. As expected, the more the CuO6PEI amount increases, the higher the electrostatic interaction is, and the higher intensity variations are.

Therefore, we can confirm the effectiveness of the colloidal heterocoagulation treatment for functionalizing the CNF, using the same oxidative capacity of the selective oxidation of cellulose.

a



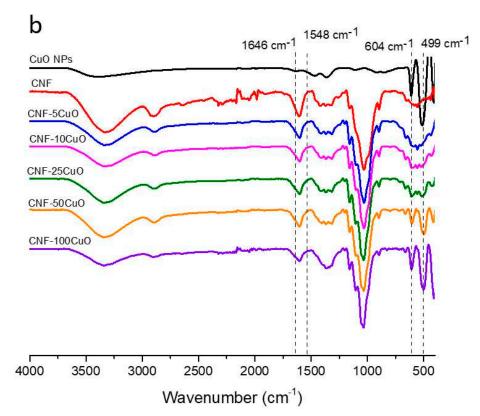


Figure 4. a) Picture and b) FTIR spectra of CNF-XCuO6PEI aerogels.

To analyze the internal structure and microstructural details of different aerogels formulated, FESEM images were taken (Figure 5).

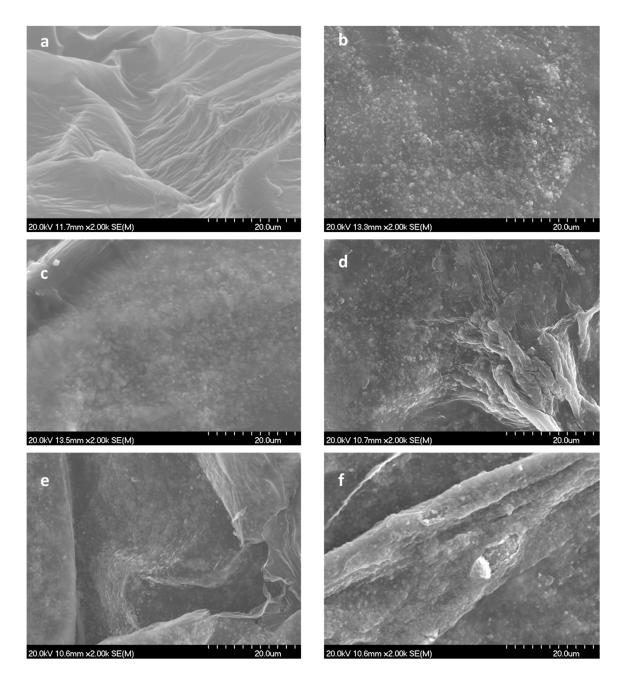


Figure 5. Scanning electron micrographs of a fractured (top view) aerogel. Letters a, b, c, d, e and f correspond to CNF, CNF-5CuO, CNF-10CuO, CNF-25CuO, CNF-50CuO and CNF-100CuO, respectively.

The typical wrinkled structure usually obtained for aerogels was herein analyzed at high magnification to appreciate the details of CuO6PEI nanoparticles over the reticulated CNF.

FESEM images evidence that an increase in the ratio of the inorganic nanophase has a significant effect on the aerogels microstructures. CuO6PEI NPs are visualized covering the surfaces of all aerogels except the reference one. The more the CuO6PEI content increases, the lower the exposed surface of CNF.

The CNF-5CuO, CNF-10CuO, CNF-25CuO samples with a lower inorganic content showed a high dispersion of nanoparticles, avoiding the presence of large agglomerates. The effect of the previous CuO NP dispersion/stabilization confirms the effectiveness of colloidal method when we are dealing with blending of components for these composite materials. On the other hand, the CNF-50CuO and CNF100CuO samples with higher inorganic content showed nanoparticles overlap. They

are settled into cellulose surfaces forming interconnected layers, which are actively exposed towards the external medium, which could potentially affect their functional behavior.

3.3. Antimicrobial activity of Aerogels

Results of the agar disk diffusion test are presented in Figure 6. Overall, the antibacterial activity of the aerogels was fruitful, obtaining width of the antimicrobial "halo" (nwhalo) from 0 to 0.6 and 0.35 for *S. aureus* and *E.coli*, respectively. For both bacteria, the inhibition zone was directly correlated with the amount of CuO NPs in the aerogels reaching the maximum inhibition zone with 100%-CuO NPs samples. Since copper ions must diffuse into the agar to inhibit germination and bacterial growth, the higher the amount of CuO NPs in the aerogels the more ions available to diffuse into agar, leading to larger diameters. Comparing the effect of the CuO NPs on both microorganisms we observe a higher effectiveness of the aerogels against the gram-positive *S. aureus* compared to the gram-negative *E. coli*. This observation is also in agreement with [30] and [31], who reported Gram-negative strains present more resistance/tolerance against ZnO and CuO nanoparticles over Gram-positive bacterial strains. The outer membrane (OM) present in Gram-negative bacteria acts as a protective barrier against antimicrobial agents since they must pass the outer membrane to access their targets, conferring greater resistance compared to Gram-positive bacteria [32].

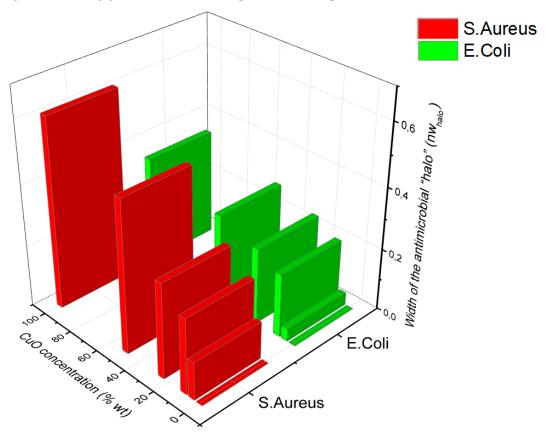


Figure 6. Antibacterial activity against *E. coli* and *S. aureus* by Agar Diffusion method.

Since disk diffusion method is not capable to distinguish between bacterial growth inhibition and bacterial death, the antimicrobial kinetics of the CuO NPs aerogels were additionally studied by the broth dilution method. Figure 7 shows the antimicrobial behavior of the samples in liquid media. Overall, the composites were able to eliminate both pathogenic bacteria in less than 5 hours when CuO NPs were present in aerogels at concentrations below 50% wt.

It was noted that CNF samples caused a slight reduction in the first few hours of both studies, remaining constant thereafter. This phenomenon can be justified through different theories. On one side, an antibacterial activity of Oxidized Nanocellulose with different Carboxyl content has been reported [33]. But also, since the bacteria are in liquid media, the porous structure of CNF aerogels

could function as a physical impairment (network) where bacterial cells are retained leading to their accumulation[34]. In our analyses, the cell retention was different for both bacteria, reaching reductions of around 1 log for S. aureus compared to 0.7 log for E. coli. A reasonable answer to this phenomenon could be the different features of both bacteria and the way they are arranged. First, Staphylococcus aureus is a coccus, non-flagellated bacteria of around 1µm diameter, that is grouped in clusters[35]. In addition, Escherichia coli is a rod shape, flagellated bacteria with dimensions of approximately 1 x 0.5 µm and normally is found arranged singly or in pairs[36]. The smaller size of E.coli along with its active motility in liquid media, causes a lower cell retention in the aerogels compared to S. aureus. A clear reduction in bacterial counts was evidenced when CuO NPs were added into the samples. As shown in Figure 7a and 7b, the aerogels were able to deactivate S. aureus and E. coli in less than 5 hours, except for CNF-50CuO samples. The size, shape, and charge of CuO NPs are crucial factors that define the ability to kill bacteria[37]. Usually, the smaller the nanoparticle the more effective the cell damage, since the surface area to volume ratio is larger as compared to bigger particles. However, the high amount of CuO NPs in CNF-50CuO samples leads to their agglomeration[13], reducing the specific surface area and therefore delaying cell death to 7h. Notice that CNF-100CuO samples were not included due they presented worse behaviors

Comparing the bactericidal effect for both bacteria, a greater reduction of *S. aureus* was evidenced in the first hour reaching log reductions of 1.4, 0.99, 0.82 and 2.27 for CNF-5CuO, CNF-10CuO, CNF-25CuO, and CNF-50CuO samples, respectively. Despite the protection of *E.coli* against antimicrobial agents, a sharp decrease was observed between 1h and 2.5h. During the first hour of study, *E. coli* is able to place in favorable areas where the concentrations of Cu²⁺ ions in the solution are low. However, the higher release of ions from the materials to the medium at 2.5h, together with the high cationic affinity conferred by the negatively charged lipopolysaccharide molecules present in the outer membrane of *E.coli* [37], causes concentrations to decay to those obtained for *S. aureus*.

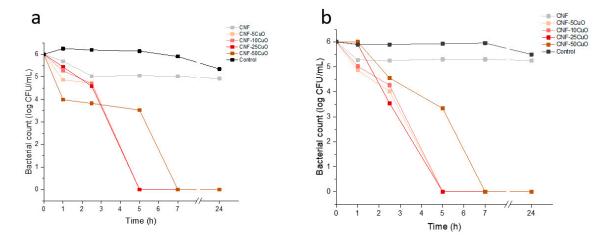


Figure 7. Antibacterial activity of CNF/CuO aerogels against S. aureus (a) and E. coli (b).

4. Conclusions

It has been demonstrated that the application of a controlled amount of ultrasonic power in a concrete volume of sulfate precursor solution (W cm⁻² mol⁻¹) allows the instantaneous formation of a pure CuO monoclinic nanophase. The direct obtaining of this specie avoids the application of further thermal treatments that are conventionally required in absence of sonochemistry.

In terms of processing of nanocomposite materials, it can be concluded that the design of CNF–CuO heterostructures by means of a colloidal route provides numerous possibilities that are unattainable using traditional methods, confirming the fundamental hypothesis of the present work. It was shown that TEMPO-Oxidized CNF showed idoneous surface charge to promote their electrostatic interaction with highly dispersed CuO nanostructures. It is observed that with increasing CuO concentration in the formulation, the CNF surface is completely covered, increasing the

13

All samples showed a positive response against Escherichia coli and Staphylococcus aureus. An in-crease of the antibacterial response of the aerogels, measured by agar disk diffusion test, has been observed with the increase of CuO NPs incorporated, obtaining width of the antimicrobial "halo" (nwhalo) from 0 to 0,6 and 0,35 for S. aureus and E. coli, respectively. Furthermore, the aerogels have been able to deactivate S. aureus and E. coli in less than 5 hours when the antibacterial assays have been analyzed by broth dilution method.

It is therefore concluded that the use of CNF as a support for antimicrobial materials favors the adequate dispersion and support of the nanoparticles, and its aerogel conformation and specific cross-linked structure facilitate good contact and transfer towards different media, increasing the efficiency of the antimicrobial activity.

Although this work involved the incorporation of CuO NPs into a specific nanocellulose-based support through cross-linking phenomena, the colloidal procedure, herein described, opens a door to ad hoc design of multifunctional membranes, filters, or flexible substrates, which require the inclusion of other alternative compositions.

Author Contributions: "Conceptualization, E.U, E,E and Z.G.; methodology, E.U, W.EA and R.M.; validation, E.E., B.F and Z.G.; formal analysis, E.U, W.EA and R.M.; data curation, E.E., B.F and Z.G.; writing—original draft preparation, E.U and R.M.; writing—review and editing, E.E., B.F and Z.G.; visualization, E.E., B.F and Z.G.; supervision, E.E and Z.G.; project administration, Z.G.; funding acquisition, Z.G. All authors have read and agreed to the published version of the manuscript."

Funding: This research was funded by MCIN/AEI/10.13039/501100011033/, through the projects PID2020-113365RA-I00 and TED2021-129920A-C42.

Data Availability Statement: Data available on request

Acknowledgments: The authors are grateful to MCIN/AEI/10.13039/501100011033 projects PID2020-113365RA-I00 and TED2021-129920A-C42 for the transfer of knowledge and results that have allowed their application in this work. The authors also thank the staff of the Microscopy Unit of the Central Research Support Service (SCAI) and the staff of Instituto Químico para la Energía y el Medioambiente (IQUEMA) of the Universidad de Córdoba

Conflicts of Interest: The authors declare no conflict of interest.

References

- Zhu, J.; Hou, J.; Zhang, Y.; Tian, M.; He, T.; Liu, J.; Chen, V. Polymeric Antimicrobial Membranes Enabled by Nanomaterials for Water Treatment. J. Memb. Sci. 2018, 550, 173–197, doi:https://doi.org/10.1016/j.memsci.2017.12.071.
- 2. Ren, G.; Wan, K.; Kong, H.; Guo, L.; Wang, Y.; Liu, X.; Wei, G. Recent Advance in Biomass Membranes: Fabrication, Functional Regulation, and Antimicrobial Applications. *Carbohydr. Polym.* **2023**, 305, 120537, doi:10.1016/J.CARBPOL.2023.120537.
- 3. Reshmy, R.; Paulose, T.A.P.; Philip, E.; Thomas, D.; Madhavan, A.; Sirohi, R.; Binod, P.; Kumar Awasthi, M.; Pandey, A.; Sindhu, R. Updates on High Value Products from Cellulosic Biorefinery. *Fuel* **2022**, *308*, 122056, doi:https://doi.org/10.1016/j.fuel.2021.122056.
- 4. Doh, H.; Lee, M.H.; Whiteside, W.S. Physicochemical Characteristics of Cellulose Nanocrystals Isolated from Seaweed Biomass. *Food Hydrocoll.* **2020**, *102*, 105542, doi:10.1016/J.FOODHYD.2019.105542.
- 5. Sharma, A.; Thakur, M.; Bhattacharya, M.; Mandal, T.; Goswami, S. Commercial Application of Cellulose Nano-Composites A Review. *Biotechnol. Reports* **2019**, 21, e00316, doi:https://doi.org/10.1016/j.btre.2019.e00316.
- 6. Ahmed A. Oun, S.S.; Rhim, J.-W. Multifunctional Nanocellulose/Metal and Metal Oxide Nanoparticle Hybrid Nanomaterials. *Crit. Rev. Food Sci. Nutr.* **2020**, *60*, 435–460, doi:10.1080/10408398.2018.1536966.
- Cabanas-Polo, S.; Gonzalez, Z.; Sanchez-Herencia, A.J.; Ferrari, B. Influence of Ultrasound on the Instantaneous Synthesis of Tridimensional α-Ni(OH)2 Nanostructures and Derived NiO Nanoparticles. *CrystEngComm* 2015, 17, 6193 – 6206, doi:10.1039/c5ce00876j.
- 8. Wu, J.; Chen, Y.; Shen, W.; Wu, Y.; Corriou, J.-P. Highly Sensitive, Flexible and Transparent TiO2/Nanocellulose Humidity Sensor for Respiration and Skin Monitoring. *Ceram. Int.* **2023**, 49, 2204–2214, doi:https://doi.org/10.1016/j.ceramint.2022.09.187.
- 9. Xue, Y.; Wang, Z.; Dutta, A.; Chen, X.; Gao, P.; Li, R.; Yan, J.; Niu, G.; Wang, Y.; Du, S.; et al.

- Superhydrophobic, Stretchable Kirigami Pencil-on-Paper Multifunctional Device Platform. *Chem. Eng. J.* **2023**, *465*, 142774, doi:https://doi.org/10.1016/j.cej.2023.142774.
- 10. Chen, Y.; Zhang, L.; Yang, Y.; Pang, B.; Xu, W.; Duan, G.; Jiang, S.; Zhang, K.; Chen, Y.; Duan, G.; et al. Recent Progress on Nanocellulose Aerogels: Preparation, Modification, Composite Fabrication, Applications. *Adv. Mater.* **2021**, *33*, 2005569, doi:10.1002/ADMA.202005569.
- 11. Paul, J.; Ahankari, S.S. Nanocellulose-Based Aerogels for Water Purification: A Review. *Carbohydr. Polym.* **2023**, 309, 120677, doi:10.1016/J.CARBPOL.2023.120677.
- 12. Shankar, S.; Wang, L.F.; Rhim, J.W. Preparation and Properties of Carbohydrate-Based Composite Films Incorporated with CuO Nanoparticles. *Carbohydr. Polym.* **2017**, 169, 264–271, doi:10.1016/J.CARBPOL.2017.04.025.
- Heidari, H.; Teimuri, F.; Ahmadi, A.R. Nanocellulose-Based Aerogels Decorated with Ag, CuO and ZnO Nanoparticles: Synthesis, Characterization and the Antibacterial Activity. *Polyhedron* 2022, 213, 115629, doi:10.1016/J.POLY.2021.115629.
- 14. Almasi, H.; Jafarzadeh, P.; Mehryar, L. Fabrication of Novel Nanohybrids by Impregnation of CuO Nanoparticles into Bacterial Cellulose and Chitosan Nanofibers: Characterization, Antimicrobial and Release Properties. *Carbohydr. Polym.* **2018**, *186*, 273–281, doi:10.1016/J.CARBPOL.2018.01.067.
- 15. Shankar, S.; Oun, A.A.; Rhim, J.-W. Preparation of Antimicrobial Hybrid Nano-Materials Using Regenerated Cellulose and Metallic Nanoparticles. *Int. J. Biol. Macromol.* **2018**, 107, 17–27, doi:https://doi.org/10.1016/j.ijbiomac.2017.08.129.
- 16. Parashar, M.; Vivek, ·; Shukla, K.; Singh, R. Metal Oxides Nanoparticles via Sol-Gel Method: A Review on Synthesis, Characterization and Applications. **2020**, *31*, 3729–3749, doi:10.1007/s10854-020-02994-8.
- 17. Postnova, I.; Khlebnikov, O.; Silant'ev, V.; Shchipunov, Y. Dimensionally Stable Cellulosic Aerogels Functionalized by Titania. *Pure Appl. Chem.* **2018**, *90*, 1755–1771, doi:doi:10.1515/pac-2018-0706.
- 18. Antoniw, J.M.; Hallman, M.T.; Kiriakou, M. V; Morse, T.; Cranston, E.D. Colloidal Stability Window for Carboxylated Cellulose Nanocrystals: Considerations for Handling, Characterization, and Formulation. *Langmuir* **2023**, *39*, 10321 10334, doi:10.1021/acs.langmuir.3c00319.
- 19. Franks, G. V; Tallon, C.; Studart, A.R.; Sesso, M.L.; Leo, S. Colloidal Processing: Enabling Complex Shaped Ceramics with Unique Multiscale Structures. *J. Am. Ceram. Soc.* **2017**, 100, 458–490, doi:https://doi.org/10.1111/jace.14705.
- 20. Carrasco, S.; Espinosa, E.; González, Z.; Cruz-Yusta, M.; Sánchez, L.; Rodríguez, A. Simple Route to Prepare Composite Nanocellulose Aerogels: A Case of Photocatalytic De-NOx Materials Application. *ACS Sustain. Chem.* \& Eng. 2023, 11, 2354–2363, doi:10.1021/acssuschemeng.2c06170.
- 21. De Haro-Niza, J.; Rincón, E.; Gonzalez, Z.; Espinosa, E.; Rodríguez, A. Nanocellulose from Spanish Harvesting Residues to Improve the Sustainability and Functionality of Linerboard Recycling Processes. *Nanomaterials* **2022**, *12*, doi:10.3390/nano12244447.
- 22. AU Martí, M.; AU Frígols, B.; AU Serrano-Aroca, A. Antimicrobial Characterization of Advanced Materials for Bioengineering Applications. *JoVE* **2018**, e57710, doi:doi:10.3791/57710.
- 23. Rangel, W.M.; Boca Santa, R.A.A.; Riella, H.G. A Facile Method for Synthesis of Nanostructured Copper (II) Oxide by Coprecipitation. *J. Mater. Res. Technol.* **2020**, *9*, 994–1004, doi:https://doi.org/10.1016/j.jmrt.2019.11.039.
- 24. Phiwdang, K.; Suphankij, S.; Mekprasart, W.; Pecharapa, W. Synthesis of CuO Nanoparticles by Precipitation Method Using Different Precursors. *Energy Procedia* **2013**, *34*, 740–745, doi:https://doi.org/10.1016/j.egypro.2013.06.808.
- 25. Jagadeesan, M.S.; Movlaee, K.; Krishnakumar, T.; Leonardi, S.G.; Neri, G. One-Step Microwave-Assisted Synthesis and Characterization of Novel CuO Nanodisks for Non-Enzymatic Glucose Sensing. *J. Electroanal. Chem.* **2019**, *835*, 161–168, doi:https://doi.org/10.1016/j.jelechem.2019.01.024.
- 26. Gonzalez, Z.; Yus, J.; Sanchez-Herencia, A.J.; Dewalque, J.; Manceriu, L.; Henrist, C.; Ferrari, B. A Colloidal Approach to Prepare Binder and Crack-Free TiO2 Multilayer Coatings from Particulate Suspensions: Application in DSSCs. *J. Eur. Ceram. Soc.* **2019**, *39*, 366–375, doi:10.1016/j.jeurceramsoc.2018.09.018.
- 27. Ferreyra Maillard, A.P.V.; Espeche, J.C.; Maturana, P.; Cutro, A.C.; Hollmann, A. Zeta Potential beyond Materials Science: Applications to Bacterial Systems and to the Development of Novel Antimicrobials. *Biochim. Biophys. Acta Biomembr.* **2021**, *1863*, 183597, doi:10.1016/j.bbamem.2021.183597.
- 28. Kaffashsaie, E.; Yousefi, H.; Nishino, T.; Matsumoto, T.; Mashkour, M.; Madhoushi, M.; Kawaguchi, H. Direct Conversion of Raw Wood to TEMPO-Oxidized Cellulose Nanofibers. *Carbohydr. Polym.* **2021**, 262, doi:10.1016/J.CARBPOL.2021.117938.
- 29. Morcillo-Martín, R.; Espinosa, E.; Rabasco-Vílchez, L.; Sanchez, L.M.; de Haro, J.; Rodríguez, A. Cellulose Nanofiber-Based Aerogels from Wheat Straw: Influence of Surface Load and Lignin Content on Their Properties and Dye Removal Capacity. *Biomolecules* **2022**, *12*, doi:10.3390/biom12020232.

15

- 31. Premanathan, M.; Karthikeyan, K.; Jeyasubramanian, K.; Manivannan, G. Selective Toxicity of ZnO Nanoparticles toward Gram-Positive Bacteria and Cancer Cells by Apoptosis through Lipid Peroxidation. *Nanomedicine Nanotechnology, Biol. Med.* **2011**, *7*, 184–192, doi:https://doi.org/10.1016/j.nano.2010.10.001.
- 32. Breijyeh, Z.; Jubeh, B.; Karaman, R. Resistance of Gram-Positive Bacteria to Current Antibacterial Agents and Overcoming Approaches. *Molecules* **2020**, 25, doi:10.3390/molecules25122888.
- 33. Tian, H.; Li, W.; Chen, C.; Yu, H.; Yuan, H. Antibacterial Activity and Mechanism of Oxidized Bacterial Nanocellulose with Different Carboxyl Content. *Macromol. Biosci.* **2023**, 23, 2200459, doi:https://doi.org/10.1002/mabi.202200459.
- 34. Wang, F.; Dai, J.; Huang, L.; Si, Y.; Yu, J.; Ding, B. Biomimetic and Superelastic Silica Nanofibrous Aerogels with Rechargeable Bactericidal Function for Antifouling Water Disinfection. *ACS Nano* **2020**, *14*, 8975–8984, doi:10.1021/acsnano.0c03793.
- 35. Gnanamani, A.; Hariharan, P.; Paul-Satyaseela, M. Staphylococcus Aureus: Overview of Bacteriology, Clinical Diseases, Epidemiology, Antibiotic Resistance and Therapeutic Approach.; 2017.
- 36. Ananthanarayan, R.; Jayaram Paniker, C.K. Textbook of Microbiology; Universiti.; 2017;
- 37. Slavin, Y.N.; Asnis, J.; Häfeli, U.O.; Bach, H. Metal Nanoparticles: Understanding the Mechanisms behind Antibacterial Activity. *J. Nanobiotechnology* **2017**, *15*, 1–20, doi:10.1186/s12951-017-0308-z.

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.