

Review

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*Review*

# Translational Challenges and Prospective Solutions in the Implementation of Biomimetic Delivery Systems

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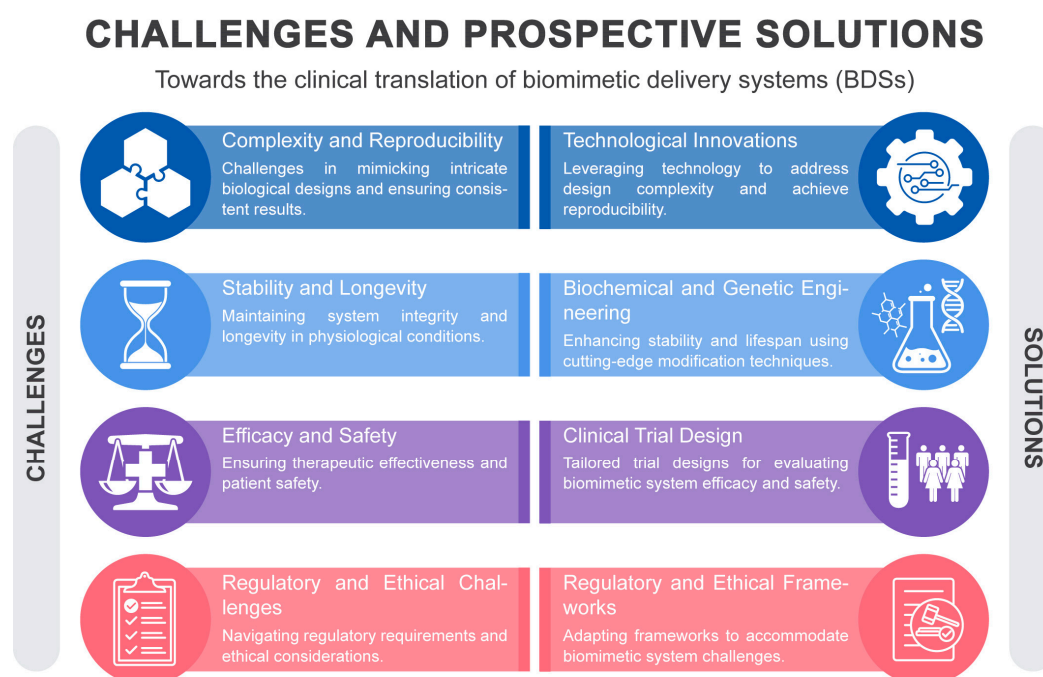
**Abstract:** Biomimetic delivery systems (BDSs), inspired by the intricate designs of biological systems, have emerged as a groundbreaking paradigm in nanomedicine, offering unparalleled advantages in therapeutic delivery. These systems, encompassing platforms such as liposomes, protein-based nanoparticles, extracellular vesicles, and polysaccharides, are lauded for their targeted delivery, minimized side effects, and enhanced therapeutic outcomes. However, the translation of BDSs from research settings to clinical applications is fraught with challenges, including reproducibility concerns, physiological stability, and rigorous efficacy and safety evaluations. Furthermore, the innovative nature of BDSs demands a reevaluation and evolution of existing regulatory and ethical frameworks. This review provides an overview of BDSs, delve into the multifaceted translational challenges and present emerging solutions, underscored by real-world case studies. Emphasizing the potential of BDSs to redefine healthcare, we advocate for sustained interdisciplinary collaboration and research. As our understanding of biological systems deepens, the future of BDSs in clinical translation appears promising, with a focus on personalized medicine and refined patient-specific delivery systems.

**Keywords:** biomimetic; bioinspired; nanodiscs; liposomes; virus-like particles; albumin; ferritin; polysaccharides; extracellular vesicles

## 1. Introduction

Biomimetic delivery systems (BDSs), defined by their ability to mimic biological systems, hold significant promise in the realm of biomedicine and nanomedicine. They leverage the principles of nature, emulating the structural or functional attributes of biological systems to enhance drug delivery capabilities [1–3]. BDSs often involve the use of naturally derived materials, the structural mimicry of biological entities, or the replication of biological processes, with the aim of improving drug delivery outcomes such as targeting Zability, controlled release, and biocompatibility [4–6]. Recent advancements in biomimicry have resulted in the creation of innovative drug delivery systems [7–9], spanning various paradigms such as liposomal carriers [10], virus-like nanoparticles (VLPs) for gene delivery [11–13], and hydrogel structures [14–16]. Additionally, new classes of delivery vehicles have emerged, including extracellular vesicles (EVs) [17,18], red blood cell (RBC)-based carriers [19,20], and nanodiscs (NDs), each presenting unique therapeutic prospects. EVs,

naturally occurring cellular delivery systems, comprised of microvesicles and exosomes [22,23], hold promise due to their bio-compatibility and targeted delivery capability [24,25], stimulating interest in their use for delivering RNA-based therapeutics [22,26]. RBCs, with their advantageous properties like long circulatory half-life and immune evasion, are under investigation as potential drug carriers, with methods involving engineering and manipulation into biomimetic nanoparticles [27][28,29]. NDs, mimicking high-density lipoproteins (HDL) [31,32], are versatile delivery platforms due to their ability to solubilize and present various drug molecules and have potential benefits for targeted cancer therapy due to preferential uptake by cancer cells [33,34].



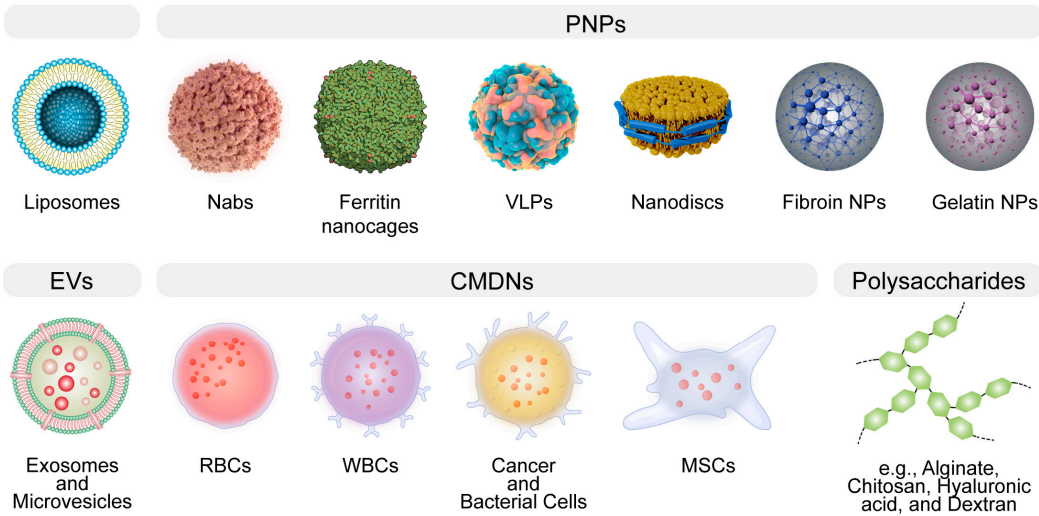
**Figure 1.** A schematic overview of the clinical translational hurdles and prospective solutions in BDS.

BDSs represent a compelling approach to drug delivery, capitalizing on the structures and mechanisms intrinsic to biological systems (Figure 2). The theoretical bedrock of biomimetic delivery systems is fundamentally rooted in the principles of self-assembly, molecular recognition, and biocompatibility [1–3]. Self-assembly refers to the process by which molecules spontaneously organize into ordered structures [35,36]. This characteristic, borrowed from nature, is widely harnessed to construct nanoscale delivery vehicles [37]. Molecular recognition refers to the ability of molecules to interact specifically with others, typically resulting in a biological function or response. This principle allows for the precise targeting of therapeutic agents to disease sites, minimizing off-target effects. Lastly, the nano-bio interface effect and biocompatibility are critical attributes of any biomimetic nanosystem intended for clinical use, ensuring that the system does not elicit adverse immune responses or toxic effects [38,39]. The paradigm of drug delivery has seen revolutionary advancement with the burgeoning interest in BDSs, which intimately mimic biological structures to enhance therapeutic efficacy. Noteworthy examples include liposomes, PNPs, extracellular vesicles, Cell membrane-derived nanocarriers, nanodiscs, and natural polymers, each boasting unique attributes for the therapeutic delivery and in situ companion diagnostics [40–42].

These advancements have catalyzed previously unattainable therapeutic opportunities, including targeted cancer therapies [43], gene editing [44], and regenerative medicine [45]. The diversity and adaptability of these BDSs underscore the significant potential of leveraging nature's design in the development of next-generation therapeutic interventions. However, the path from the bench to bedside translation is fraught with complexity. Despite the theoretical advantages of BDSs, their translation into clinical applications has been slower than expected, hindered by various

technical, biological, and regulatory challenges. For instance, issues such as scalability of production, immunogenicity, stability of the systems under physiological conditions, and navigating regulatory approvals pose significant hurdles. The urgency for such a discourse is evident. The promise of biomimicry in healthcare can only be realized when these delivery systems transition from being experimental novelties to tools readily available in the clinician's arsenal.

This review elucidates the translational challenges prevalent in the field, focusing on their intricate aspects and contemplating potential resolutions (Figure 1). Given the broad scope of this review, emphasis is placed on general themes rather than meticulous analyses of individual cases. We initially examine challenges segmented into technical, biological, and regulatory categories, before presenting emerging solutions and strategies, highlighted by instances of successful translation. Conclusively, we offer insights into the future of the BDSs field, emphasizing the revolutionary impact of these technologies on healthcare and advocating for sustained research and collaboration in this realm.



**Figure 2.** The general illustration of biomimetic delivery systems (BDSs). BDSs are designed to emulate natural structures, thereby augmenting therapeutic efficacy. Notable examples encompass liposomes, protein-based nanoparticles (PNPs), extracellular vesicles (EVs), cell membrane-derived nanocarriers (CMDNs), nanodiscs, and polysaccharides.

2. Challenges and Approaches in Clinical Translation of BDSs

2.1. Complexity and Reproducibility

In the realm of biomimetic delivery systems, different biomimetic materials and structures have been explored for their potential advantages in the delivery of therapeutic agents. Each of these systems brings unique complexities and challenges in terms of their production and ensuring their reproducibility, which is vital for their successful translation into clinical applications.

**Table 1.** A summary of the complexities, reproducibility challenges, and prospective solutions related to various BDSs.

BDS	Complexity and Reproducibility	Prospective Solutions
Liposomes	Diverse lipids induce variability.	Advanced lipid-mixing technologies.
	Sustained stability is challenging.	Freeze-thaw increases reproducibility.
	Surface alterations cause variability.	Advanced ligand conjugation methods.
	Scaling up adds variability.	Automated production control.
Protein-based NPs		
Albumin NPs	Influenced by albumin source. Uniform size & shape are hard to	High-pressure homogenization. Improved purification techniques. High



	attain. Altered surface for specific targeting. Efficient drug encapsulation control.	Throughput Screening, Microfluidics and Computational Modeling.
Protein-based nanocages	Ensuring consistent protein folding. Reproducible encapsulation. Stable surface chemistry. Efficient drug encapsulation control. Consistent drug release profiles.	Advanced bioengineering methods. Monitoring protein folding in real-time. New modification methods for stability. Innovative drug-loading for consistency. Smart release systems for specific triggers.
VLPs	Complexity in VLP assembly. Attaining purity and reproducibility. Heterogeneous surface modifications. Inconsistent therapeutic encapsulation in VLPs.	Advanced purification like SEC. Genomic engineering for optimized production. Developed specific bioconjugation techniques. High-throughput techniques for optimal encapsulation.
NDs	Component multiplicity causes variability. Consistent size and shape. Adding functional groups increases complexity. Batch-to-batch variability	Synthesis and purification for uniformity. Advanced assembly techniques. Site-specific functionalization and modular design. Standardized protocols, real-time QC, and advanced characterization.
EVs	Heterogeneity of EV populations. Differentiating EV subtypes is challenging. Possible contamination with proteins. Ensuring efficient encapsulation. Controlling release kinetics. Maintaining EV properties post-modification. Ensuring targeting specificity. EV source depends on donor cells.	Advanced centrifugation. High-resolution imaging & flow cytometry. Improved purification processes. Sonication or electroporation. Covalent and non-covalent linking. Bio-orthogonal chemistry. Molecular imprinting techniques. Standardized cell lines/biofactories.
CMDNs	Potential heterogeneity due to cell sources. Unpredictable biological interactions. Batch-to-batch differences. Enhancing nanocarrier functionality/specificity.	Improved cell culture techniques. Predictive molecular modeling & simulation. Controlled nanocarrier production via microfluidics. Surface engineering, genetic modifications, molecular tethering strategies.
<b>Polysaccharides</b>		
Alginate	Variability in alginate source/purity. Gelation process control. Encapsulation efficiency variability.	Advanced chromatography for purification. Microfluidics for consistent gel bead formation. Advanced sonication/emulsification.
Chitosan	Molecular weight influences properties. Degree of deacetylation influences properties. Replicating desired structures is challenging. Crosslinking variability affects stability. Uniform surface properties are challenging.	Advanced chromatographic techniques to standardize molecular weight. Spectroscopy for precise deacetylation. High-resolution microscopy & automated synthesis. Advanced controlled crosslinking techniques. Advanced surface characterization.
Hyaluronic acid	Variability in sources. Consistent molecular weight is crucial.	Microbial synthesis of HA for consistency. Real-time molecular weight monitoring.
Dextran	Variability in molecular weight distribution. Branching variation affects behavior. Functional group variation. Achieving consistent size/morphology is challenging.	Controlled polymerization methods. Detailed structure analysis via spectroscopy. Controlled enzymatic/chemical modifications. Microfluidics for controlled and reproducible nanosystem generation.

Liposomes, vesicular structures composed of lipid bilayers, are valuable carriers for various drugs, improving their pharmacokinetics, biodistribution, and therapeutic index, exemplified by clinically approved liposomal drugs like Doxil®/Caelyx® and AmBisome® [46]. However, challenges in clinical translation include the heterogeneous nature of liposomes affecting consistency between

batches, impacting drug delivery efficacy and therapeutic outcomes [47]. Size and lipid composition variations, stability concerns related to environmental factors, and deviations in morphology and drug release under inappropriate storage temperatures or extreme pH levels are notable issues [48–52]. To mitigate these, real-time monitoring, process analytical technologies (PAT), and techniques like nuclear magnetic resonance (NMR) spectroscopy and liquid chromatography–mass spectrometry (LC–MS) are crucial to ensure formulation consistency and rectify deviations immediately [53–55]. Challenges in liposomal drug manufacturing include the need for meticulous control over storage and handling, stringent quality control, and managing the transition from laboratory to industrial scale, all contributing to increased costs and complexity [56]. However, continuous manufacturing processes and advanced technologies, such as high-throughput screening and microfluidic systems, can enhance consistency and uniformity, ensuring precise formulation control for therapeutic outcomes [57]. In silico methods aid in designing stable liposomal systems [58,59]. While it's improbable to eradicate all challenges in liposomal drug delivery systems (LDDS), integrating advanced technologies can alleviate them, ensuring efficient and consistent production of clinically effective LDDS. The integration of these technologies into formulation and production processes is crucial in addressing the challenges comprehensively.

The exploration of endogenous proteins like albumin in drug delivery is growing due to their biocompatibility and enhanced pharmacokinetics. However, the translation of albumin-based carriers is intricate due to challenges in modification and resultant variability [60]. Methods like covalent linkage and encapsulation are used for drug attachment to albumin [61], requiring precision to maintain albumin's integrity, and inconsistencies in these processes can lead to variations in drug loading and reproducibility [62]. While albumin is naturally benign, modifications can potentially induce immune reactions, impacting its biocompatibility, binding affinities, biodistribution, and pharmacokinetics, thereby posing a risk of undermining its inherent benefits [63,64]. Such modifications and variability in drug release kinetics can influence drug efficacy, plasma levels, and safety [65,66]. Utilizing high-resolution techniques and computational modeling can provide structural insights and predict interaction behaviors in biological settings, helping in refining drug loading and streamlining the design process [67–70]. Scaling from lab to industrial levels can impact product quality and characteristics in albumin-based systems, and the complexity of albumin modification challenges the reproducibility [60,71]. Implementing microfluidic devices [72,73], utilizing standardized albumin sources like rHSA [74], and employing automated synthesis platforms can enhance reproducibility by ensuring consistent reactions and minimizing variability and contamination [75]. The incorporation of sensors and analytical tools for real-time feedback and continuous monitoring of synthesis parameters further ensures product consistency [76–78]. The complexity and need for precise reproducibility in albumin-based delivery systems pose significant challenges, but technological advancements, from high-resolution analyses to automation, combined with strategic design, address these challenges [79,80], paving the way for broader clinical adoption.

Ferritin-based PNPs show promise for personalized medicine due to their encapsulation abilities but face translation challenges stemming from the complexity and reproducibility of assembly [81–83]. Precise control of pH and ionic strength is critical due to ferritin's conformational plasticity, and deviations can lead to irregular nanoparticles affecting drug delivery and therapeutic outcomes [84]. Standard assembly/disassembly methods and advanced spectroscopic techniques are pivotal for maintaining conditions and understanding ferritin conformational transitions [85,86]. Modifications to optimize encapsulation can disrupt self-assembly and affect size, thus impacting pharmacokinetics and pharmacodynamics [87–89]. Standardized modification protocols, including directed evolution and genetic fusion, are crucial for maintaining consistency [90]. Inherent size variability of ferritin nanoparticles poses further challenges [91], necessitating advanced separation methods and size-exclusion techniques to ensure uniform therapeutic outcomes [92]. Real-time monitoring and advanced characterization techniques like cryo-electron microscopy provide insights into structures, aiding in addressing polydispersity [93]. Integration of technology advancements such as molecular dynamics simulations offers perspectives on ferritin assembly behavior, aiding in addressing the polydispersity [94,95] for informed design, and a comprehensive approach focusing on control and

standardization can help overcome challenges and realize ferritin's clinical potential in personalized medicine.

Virus-like particles (VLPs) use the infectious properties of viruses for therapeutic delivery, relying on complex recombinant DNA technology [96], and face inherent production variability. Advanced bioinformatics tools can refine the integration of foreign DNA [97,98], reducing genetic risks and enabling exact cellular condition control, assisted by modern bioreactors and real-time monitoring [99]. These innovations, along with high-throughput screening and synthetic biology, can mitigate biological system variability and genetic instability, promoting a consistent VLP manufacturing [98,100,101]. However, purifying VLPs is complex due to their similarity to host proteins and size variation. Variations in purification methods can affect VLP yield and characteristics [102], possibly causing inconsistent therapeutic results. Nanotechnology and advanced filtration [103,104], coupled with real-time monitoring and cutting-edge spectroscopy [105–107], address these challenges by distinguishing VLPs from impurities and ensuring structural integrity. A deeper understanding of fundamental biological processes and targeted interventions, backed by advancements in technology and knowledge, are crucial for developing more efficient and reliable production strategies for VLP delivery systems.

Nanodiscs (NDs), stabilized by membrane scaffold proteins (MSPs), are discoidal structures apt for studying membrane proteins and delivering bioactive agents due to their biomimetic nature [31,32]. However, their clinical application is hindered by challenges in the complex, multi-step assembly process and reproducibility. The assembly involves the self-assembly of phospholipids and MSPs, and the correct protein-to-lipid ratio is crucial for ND integrity and function [108]. Factors like lipid type, MSP variant, and assembly conditions necessitate optimization and significantly impact the assembly complexity and reproducibility [108,109], which are essential for complying with strict pharmaceutical regulations. Minor variations could alter ND properties, affecting their *in vivo* behavior and therapeutic efficacy, leading to batch variability and translational challenges. Microfluidic automation [110], real-time monitoring [111], and design strategies, such as molecular dynamics simulations [112–114], can address assembly complexity and enhance understanding of ND behavior. The scalability of ND production is pivotal, with continuous flow synthesis being a potential solution to maintain quality and meet regulatory demands for manufacturing consistency, as traditional batch processes introduce variability and are challenging to scale [115,116]. Efficient detergent-removal strategies and the exploration of biocompatible, biodegradable detergents are vital to mitigate toxicity concerns and simplify post-assembly purification [117–119]. In conclusion, overcoming the challenges in assembly complexity, reproducibility, and scalability is crucial to harness the full potential of NDs in innovative therapeutic delivery systems.

Silk fibroin (SF) and gelatin (GA) have been extensively researched for their potential in biomimetic delivery systems, owing to their biocompatibility and adjustable degradation rates, essential for *in vivo* nanoparticle application, especially in drug delivery [120–123]. However, translational challenges arise from their inherent complexity and the associated reproducibility issues in nanoparticle fabrication. For SF, clinical application is hindered by product heterogeneity arising from variability in silk sources and fibroin properties [124]. Advanced genetic engineering tools, like CRISPR/Cas systems, and standardized fibroin extraction methods can help overcome such variability, ensuring consistent quality and properties essential for drug delivery [125–127]. Similarly, GA faces variability and reproducibility challenges due to differences in source animals and extraction methods [128–130]. High-throughput screening techniques and process standardization [131–133], including controlled crosslinking conditions and microfluidic platforms [132–135], are crucial for maintaining consistency in nanoparticle production. These enhancements, along with computational models predicting interactions between SF or GA and encapsulated drugs, contribute to achieving optimal and consistent biological performance [136–138]. Thus, standardizing sourcing, purification, and fabrication procedures, coupled with a comprehensive understanding of their impacts, are imperative for the successful clinical translation of these biomimetic systems.

Extracellular Vesicles (EVs) are notable for their potential in targeted therapeutic delivery and have gained prominence in biomedical research due to their capacity to transfer cellular information.

However, their clinical transition is impeded by challenges related to their production, heterogeneity, scalability, and stability [139,140]. EVs, originating from cell cultures, play roles in cellular communication and waste management but exhibit considerable variability in size, content, and origin, complicating manufacturing and impacting therapeutic predictability and reproducibility [141]. Controlling this variability is crucial and can be achieved using single-vesicle analysis techniques, such as nanoscale flow cytometry, and potentially through synthetic biology approaches to ensure uniform EV production [139,142–144]. Scalability remains a significant challenge, with existing methods like ultracentrifugation being inefficient and inducing structural alterations in vesicles [145]. The introduction of novel technologies like bioreactors and microfluidic platforms has revolutionized EV production by optimizing cell conditions and enhancing yield and process efficiency [146–149]. The stability of EVs is paramount, with external factors impacting their functionality and safety. Advanced lyophilization, nano-encapsulation, and cryoprotectants have been employed to enhance EV shelf life, protect vesicle integrity, and prevent aggregation [150–152]. The application of artificial intelligence and machine learning can expedite and standardize EV analysis for quality control [153]. Despite their immense therapeutic potential, the realization of EVs necessitates advancements in their biology, production optimization, and rigorous quality control to address the prevailing challenges.

Cell Membrane-Derived Nanocarriers (CMDNs), particularly from erythrocytes, present a promising frontier in targeted therapeutic delivery due to their biological stealth characteristics [20,27,28]. Nonetheless, the complexities in isolation, modification, and loading processes, coupled with the need for rigorous quality control and reproducibility, impede their clinical translation [154]. The isolation of CMDNs is intricate, involving donor cell selection, cell lysis, removal of cellular components, and each stage introduces potential variability affecting product consistency [155]. Donor cell selection, influenced by age, health, and genetics, affects nanocarrier characteristics and performance. Implementations of microfluidic technologies, automation, and the utilization of 'cell banks' with optimal donor cells can standardize processes and diminish variability [110,156]. Additionally, post-isolation engineering of CMDNs for enhanced stability, circulation, and targeted delivery introduces further complexity. Controlled conditions and precision are requisite for consistent modifications across batches, facilitated by techniques like atomic layer deposition and bio-orthogonal chemistries [157,158], with real-time monitoring ensuring uniformity [159,160]. Rigorous validation is vital for confirming drug loading and release profiles, crucial for therapeutic efficacy. The need for stringent quality control amid varied CMDN properties necessitates comprehensive quality control approaches. Techniques like nanoparticle tracking analysis and dynamic light scattering are fundamental for characterizing CMDN parameters [161]. However, inherent biological variability and multifaceted production processes exacerbate the challenges in capturing CMDN diversity. Feedback-controlled systems, like process analytical technology (PAT) [162], and computational models leveraging molecular dynamics and machine learning provide predictive insights into nanocarrier behavior and aid in optimizing production parameters [163,164]. Overcoming the production complexities, variability, and quality control challenges is pivotal for the clinical realization of CMDNs.

Polysaccharides such as alginate, chitosan, hyaluronic acid (HA), and dextran are prominent in nanoparticle synthesis due to their biocompatibility and safety [165]. However, their natural origins introduce variability in source, purification, and modification, yielding heterogeneity in nanoparticle properties which can impact the stability and reproducibility of delivery systems. The diverse sources, with variations in biological, chemical, and physical properties, influence polysaccharide properties, such as molecular weight and degree of deacetylation, thereby affecting nanoparticle attributes like size, charge, stability, and ultimately, therapeutic efficacy [166–168]. Modern extraction techniques and purification processes can mitigate batch variability, while sensor-based technologies and process adjustments aim to enhance consistency [169–172]. However, residual contaminants and modifications to polysaccharides amplify heterogeneity issues, impacting solubility, degradation, and drug loading. The employment of machine learning and artificial intelligence optimizes modification parameters, ensuring consistent processes and reduced product variability. The



inherent variability in polysaccharide-based nanoparticles alters biological interactions and poses challenges in clinical translation, affecting pharmacokinetics, biodistribution, and therapeutic efficacy [173]. Advanced characterization methods and real-time monitoring technologies, like PAT and digital twins of the production process, are crucial to control heterogeneity and enhance reproducibility [162,174]. The inherent complexity and reproducibility challenges of polysaccharides necessitate the development of standardized methods for extraction, purification, and modification, as well as advanced characterization techniques. Integrating technological advancements and innovative design strategies is pivotal for developing consistent and effective polysaccharide-based delivery systems, essential for bridging the gap from laboratory to clinical application.

2.2. Stability and Longevity

The quest for stability and longevity of BDS in physiological conditions is a complex journey marked by numerous challenges. These systems, while crafted to mimic the natural biological environment, still encounter substantial difficulties in withstanding rapid clearance or degradation within the human body. This factor reduces their therapeutic window, undermining their effectiveness in achieving the desired clinical outcomes.

**Table 2.** An overview of the stability, longevity challenges, and prospective solutions related to various biomimetic delivery systems.

BDS	Stability and Longevity Challenges	Prospective Solutions
Liposomes	Sensitivity to oxidation and hydrolysis. Fusion/aggregation in serum. Rapid clearance from circulation.	Liposome coating (e.g., PEGylation). Incorporation of cholesterol. Antioxidant inclusion.
Protein-Based NPs		
Albumin nanoparticles	Instability in harsh environments (e.g., acidic pH). Enzymatic degradation.	Cross-linking of albumin molecules. Encapsulation with protective polymers. Surface modifications.
Protein-based nanocages	Structural disintegration at non-optimal conditions. Immune recognition and clearance.	Chemical surface modifications. Incorporation of stability-enhancing ligands. Fusion with other stable proteins.
VLPs	Potential immunogenicity. Stability issues due to dynamic protein structures.	Genetic modifications. Encapsulation within protective matrices. Surface modifications to reduce immunogenicity.
NDs	Sensitivity to physiologic conditions, leading to structural alteration. Potential immune recognition.	Use of stable lipids. Protective protein inclusion. Surface modification.
Fibroin and Gelatin	Sensitivity to temperature and pH. Enzymatic degradation in vivo.	Chemical cross-linking. Incorporation into composite materials. Coating with protective polymers.
EVs	Susceptibility to clearance mechanisms. Sensitivity to physiologic	Surface modifications. PEGylation. Encapsulation within biomaterials. Cryopreservation techniques.

	conditions leading to vesicle disruption.	
CMDNs	Potential immunogenicity. Sensitivity to in vivo degradation mechanisms.	Immune camouflage techniques. Genetic modifications for enhanced stability. Surface modifications.
Polysaccharides		
Alginate	Rapid degradation in vivo. Instability in the presence of divalent cations.	Cross-linking with divalent cations. Incorporation into composite materials. Layer-by-layer assembly.
Chitosan	Solubility issues in neutral and basic pH. Rapid degradation in vivo.	Chemical modifications for solubility. Cross-linking. Layer-by-layer assembly.
Hyaluronic acid	Rapid enzymatic degradation in vivo. Instability under harsh conditions.	Derivatization and cross-linking. Hydrogel formulations. Composite materials incorporation.
Dextran	Sensitivity to oxidative conditions. Enzymatic degradation.	Cross-linking. Encapsulation within protective matrices. Blend with other stable polymers.

Liposomes are inherently unstable due to the susceptibility of phospholipids to oxidation and hydrolysis, affecting their structural integrity and function [175,176]. Oxidation leads to the formation of cytotoxic peroxidation by-products, posing substantial challenges to clinical applications [177]. Antioxidants like vitamin E and ferulic acid can neutralize oxidative damage, and their uniform distribution is facilitated by advanced techniques such as high-pressure homogenization [178–180]. The incorporation of stable phospholipids like sphingomyelin and cholesterol can further enhance membrane stability [181]. Conversely, hydrolysis disrupts liposomal structure and compromises the stability and the encapsulated agents' efficacy in physiological environments [182–185]. Encapsulation with lipid-polymer conjugates like PEG-PE and emerging techniques like electrospinning can mitigate hydrolytic degradation [186,187]. Utilizing hydrolytically stable phospholipid analogs and designing liposomes with interdigitated lipid phases or incorporating ceramides can also bolster resistance to hydrolytic degradation [188]. Therefore, a profound understanding of phospholipid oxidation and hydrolysis is essential for developing stabilization strategies, crucial for liposomes' successful clinical translation.

Albumin-based BDS, revered for their biocompatibility and molecule-binding potential, encounter numerous challenges in clinical transition due to their interactions with various bodily substances, leading to aggregation and premature therapeutic release [65,66,189]. Such interactions risk sub-optimal outcomes and affect pharmacokinetics and efficacy as they are quickly cleared by the immune system. Furthermore, enzymatic actions in the body can jeopardize their structural integrity and result in variable drug levels and adverse events [60]. Storage and transport also present challenges, including denaturation, oxidation, and aggregation [67,190–192]. Several strategies have been developed to mitigate these challenges, including nanoencapsulation and PEGylation to prevent premature interactions and extend circulation half-life [193–195]. Modifying nanoparticle size and shape, utilizing enzyme-inhibiting coatings, and employing cryopreservation and lyophilization address issues related to immune evasion, enzymatic degradation, and structural integrity [196,197]. Implementations of antioxidants, hydrogel encapsulation, and optimized buffer solutions offer protection against various stresses and maintain albumin structure [198,199]. Molecular imprinting and stimuli-responsive elements have also been utilized for improved drug loading and controlled delivery [200–203]. Hence, integrating these methodologies is pivotal in addressing the complications associated with albumin-based BDS, enabling enhanced therapeutic delivery.

Protein-based nanocages, led by ferritin, are a breakthrough in theranostic devices. They promise innovative drug delivery systems based on biomimetic principles. Yet, the journey to clinical use presents challenges, including structural disruption in varying in vivo environments, which might trigger unintended drug release [84]. These nanocages also risk denaturation, aggregation, or deactivation under certain conditions, necessitating specialized storage solutions. While ferritin's capability to traverse biological barriers is notable, controlling sustained drug release remains complex [204,205], with modifications for targeted delivery potentially introducing immunogenicity [206,207]. Ensuring uniformity in properties and drug potency during clinical manufacturing is imperative. Addressing these challenges demands a multidisciplinary approach, employing advancements in material science, innovative storage technologies, molecular engineering for precise drug release, advanced bioconjugation, computational simulations, high-resolution analytics, and machine learning for real-time monitoring [208,209]. This integrated methodology, combining the expertise of nanotechnologists, biologists, and pharmacologists, is crucial for unleashing the full potential of ferritin-based systems in targeted oncology.

VLPs are renowned for their precise control, defined structures, and adjustable immunogenicity, marking them as ideal candidates for targeted delivery platforms. However, their stability is compromised in demanding physiological environments due to factors such as pH fluctuations and the presence of proteases, causing potential premature therapeutic release and impacting targeting capabilities [97,210–212]. The inherent immunogenicity of VLPs, while advantageous for vaccines, poses a significant challenge for drug delivery, as it can provoke immune responses leading to rapid clearance and possible side effects [12]. Addressing these issues involves incorporating pH-responsive modifications and protease-resistant motifs to enhance stability [213–216], leveraging nanotechnology and surface modifications to augment targeting precision [217–219], and developing innovative strategies like "stealth" VLPs and biomimetic coatings to balance immunogenicity [220,221]. Such developments are pivotal in evolving VLPs into efficient, stable therapeutic delivery systems, poised to yield enhanced clinical outcomes.

NDs serve as versatile drug delivery platforms but are hampered by challenges stemming from their amphiphilic lipid nature, causing instability in size, shape, and functional efficacy. Factors including temperature, pH, and ionic strength can induce lipid phase transitions and nanodisc aggregation, potentially causing premature drug release and reducing therapeutic efficacy [222,223]. The vulnerability of NDs to oxidation and enzymatic degradation poses significant concerns regarding their longevity and interaction with serum proteins can further induce instability [224]. Additionally, the formation of a protein corona can lead to swift immune clearance and can elicit immune responses, thereby raising safety concerns [225]. Strategies to enhance ND stability include reinforcing the lipid layer, incorporating antioxidants, and PEGylation, and developing stimuli-responsive NDs, all of which are crucial to maintaining ND biocompatibility and therapeutic potency [31,42,226,227]. The advancement in these strategies holds the potential to revolutionize ND-based drug delivery systems.

Fibroin and gelatin, due to their biocompatibility and biodegradability, are widely used in biomimetic delivery systems but face challenges related to stability and longevity under physiological conditions [228,229]. These proteins are susceptible to enzymatic degradation and pH variations, which affect their structural integrity and could lead to premature therapeutic release. Additionally, traditional sterilization methods can compromise their structural effectiveness for drug delivery. Several strategies are being developed to overcome these challenges. Chemical crosslinking and blending with synthetic polymers enhance resistance to degradation and improve mechanical properties [230–233]. Integration of bioinert nanoparticles and lyophilization offers stability and controlled drug release [234,235]. Innovations like pH-responsive coatings [236,237], coacervation and electrospinning optimized encapsulation technologies [238–241], and novel fabrication and sterilization methods, like supercritical carbon dioxide-based NP formation methodologies and cold plasma sterilization [242,243], are being explored to maintain material integrity and safety. These advancements reinforce the significance of fibroin and gelatin in evolving biomedical applications.

EVs exhibit promising capabilities for targeted therapies due to their unique biological functionality but face substantial challenges in maintaining stability and longevity [151,244–246]. Physiological factors, along with difficulties in isolation, purification, and modification, can alter EV structure and hinder therapeutic delivery capabilities [246–250]. The unstable nature of EVs necessitates advancements in methodology to preserve functionality during storage, transport, and therapeutic loading, with issues such as sensitivity to freeze-thaw cycles and long-term storage further complicating their utilization [152,251]. Strategies such as encapsulation technologies [252,253], surface modifications [254], and advanced isolation methods are being developed to address these challenges [148,149,255]. Additionally, innovations in cryoprotectants, packaging, and transport solutions are being explored to enhance EV stability and integrity [197,256]. The advancement of these strategies, coupled with interdisciplinary collaboration, is pivotal for harnessing the therapeutic potential of EVs in modern medicine.

In biomimetic delivery, CMDNs, particularly those derived from red blood cells (RBCs), display significant stability and longevity challenges and can trigger immune responses leading to premature clearance due to alterations during the extraction and modification processes [257]. The mononuclear phagocyte system (MPS) recognizes altered RBC-derived nanocarriers, reducing their bloodstream longevity [258]. Solutions like surface camouflage, synthetic RBC mimetics, and the controlled release (like pH, temperature, or particular biomolecules) of immunosuppressive agents are being explored to mitigate these challenges and prolong circulation [259,260]. Furthermore, preserving structural integrity and maintaining optimal stability and efficacy during storage and transport is crucial, with enhancements via nanoengineering, refined cryopreservation, lyophilization methods, and innovative preservatives being pivotal [261–263]. The development of CMDNs necessitates a multidisciplinary approach, combining biotechnology, materials science, and pharmacology, to optimize the stability, longevity, and controlled release kinetics of RBC-derived nanocarriers, heralding advancements in therapeutic delivery systems.

Polysaccharide-based carriers such as alginate, chitosan, hyaluronic acid, and dextran exhibit unique stability issues. Alginate and chitosan are notable for their biocompatibility and biodegradability but are susceptible to instability due to their hydrophilic nature, resulting in vulnerability to environmental factors like pH and ionic strength [264,265]. Chemical modifications and protective coatings can address these vulnerabilities, improving their resilience. Hyaluronic acid faces stability issues due to susceptibility to enzymatic degradation by hyaluronidases, affecting its longevity and therapeutic effect [266]. The introduction of enzyme inhibitors or structural modifications can improve its resistance. Dextran, while soluble and biocompatible, is sensitive to microbial contamination affecting its long-term stability [266]. Enhanced sterilization, incorporation of antimicrobial agents, and encapsulation techniques can mitigate this susceptibility. The formulation of these polysaccharides into nanoparticles or microspheres offers improved stability and controlled therapeutic release, symbolizing a promising development in creating robust delivery platforms [165,172,267]. Furthermore, the profound potential of polysaccharide-based BDSs is notably challenged by inherent stability issues. The integration of technological advancements, innovative design, chemical modifications, and protective strategies is crucial for realizing their full therapeutic capabilities, promoting the development of more resilient and efficient delivery platforms. Despite the revolutionary prospects of these delivery systems in drug delivery, stability and longevity challenges in physiological conditions, storage, and transport require continuous research, development, and optimization of fabrication and handling processes. This emphasizes the need for stabilizing agents and optimized procedures to enhance the clinical translatability of these promising systems.

### 2.3. Efficacy and Safety

The efficacy and safety of therapeutic agents, especially BDSs that emulate natural biological entities, are fundamental to their clinical utility. These BDSs are anticipated to provide efficacy comparable or superior to existing treatments with a satisfactory safety profile, but their clinical translation encounters substantial challenges such as unpredictable *in vivo* behavior, potential off-



target effects, and unexpected immune responses [268]. Comprehensive evaluation, including preclinical and clinical studies of pharmacokinetics, pharmacodynamics, and immunogenicity, is pivotal to establish therapeutic validity.

Liposome-based BDSs, noted for their ability to encapsulate diverse agents, promise enhanced drug solubility and targeted delivery [46]. However, intrinsic challenges exist, impacting therapeutic efficacy and safety [269]. Variations in entrapment efficiency can result in sub-optimal drug concentrations, affecting therapeutic outcomes. Challenges with drug release kinetics, premature or delayed, can compromise drug effectiveness [270]. Rapid clearance and degradation in biological fluids, and interactions with serum proteins, enzymes, or immune cells, diminish drug bioavailability [268]. Inaccurate targeting and off-target interactions can necessitate higher doses, inducing potential side effects. Liposomal formulations, especially those modified with targeting ligands, may elicit immune responses, ranging from allergies to severe anaphylaxis [271,272], and certain liposomal components can exhibit toxicity. The variability in the Enhanced Permeability and Retention (EPR) effect introduces an additional complexity [273]. Rapid drug release due to destabilization presents overdose risks [274]. Despite the potential of liposomal systems, these multifaceted concerns necessitate meticulous consideration and ongoing refinement.

The clinical translation of liposomal technologies, exemplified by pioneering formulations like Doxil® and AmBisome®, highlights the innovation in therapeutic delivery. Doxil®, a paradigmatic FDA-approved nanodrug, utilized adaptive trial designs for dynamic dose adjustments, balancing efficacy with safety and showcasing the importance of real-time data-based refinements [275]. AmBisome® distinguished itself with a meticulous comparative approach in clinical trials, revealing its superior therapeutic index in antifungal treatments [276,277]. The imperative theme is the necessity for adaptable and flexible trial designs, with multi-arm multi-stage (MAMS) designs being efficient by allowing simultaneous evaluations of various formulations, accelerating development, and optimizing resource allocation [278,279]. In the post-approval phase, the integration of real-world evidence (RWE) and stringent post-marketing surveillance are crucial, providing insights into long-term safety, rare side effects, adherence patterns, and therapeutic outcomes in diverse populations [280,281]. This approach, drawing from the foundational successes of Doxil® and AmBisome®, informs and refines subsequent clinical trials and therapeutic guidelines. The clinical success of liposomal technologies underscores the essential role of innovative trial designs, adaptability, and ongoing evaluation in advancing liposomal therapeutics from experimental to established clinical treatments.

PNPs, encompassing a diverse set of biomaterials such as Albumin nanoparticles, Protein-based nanocages exemplified by ferritin, VLPs, NDs, Fibroin, and Gelatin, are advancing to the forefront of drug delivery research due to their inherent biocompatibility, biodegradability, and potential for precision-targeted therapeutic delivery. With Albumin nanoparticles, despite being synthesized from endogenous proteins, the inherent risk lies in the potential elicitation of immunogenic reactions, stemming from slight alterations or impurities during the nanoparticle formation process [282]; moreover, their inherent stability is also a concern as degradation can substantially affect drug release kinetics, leading to suboptimal therapeutic effects [60,192]. Turning to Protein-based nanocages, specifically ferritin, they display the dual challenges of potentially inconsistent drug loading efficiencies, which directly impact the therapeutic dosing [205,283], and a heightened sensitivity to environmental factors like pH or temperature; this sensitivity might result in unintended, premature drug release [84,205]. Additionally, their natural role in iron storage poses concerns over inadvertently disrupting iron homeostasis in the body [284]. VLPs, while ingeniously designed to lack viral genetic material, are not without concerns, primarily rooted in the potential of evoking systemic immune reactions. Their complex synthesis pathway also introduces the risk of production inconsistencies and, albeit rarely, a shadow of concern regarding potential mutations, raising the specter of inadvertently reintroducing pathogenic properties [12,285]. NDs, in their design, carry lipid-based structures, which render them susceptible to oxidation or hydrolysis, challenges further exacerbated by potential size inconsistencies that can lead to variable biodistribution, affecting their therapeutic reach and efficacy [108]. Finally, the naturally-derived PNPs, Fibroin and Gelatin,

introduce their own set of challenges: their natural sourcing can lead to variability in nanoparticle properties between batches, potential toxicity stemming from the use of chemical crosslinkers, and the concern of rapid degradation in physiological settings, which can obstruct the controlled, sustained release of therapeutic agents [120,128,134,286]. In summation, while the promise of PNPs in revolutionizing therapeutic delivery is undeniable, their path is fraught with multifaceted scientific challenges that mandate rigorous research and optimization before clinical fruition.

EVs and CMDNs, including exosomes, microvesicles, and apoptotic bodies, are prominent for their therapeutic delivery potential due to their biocompatibility and capability for targeted delivery, offering advantages over synthetic carriers. Nevertheless, integrating them into clinical paradigms requires rigorous evaluation of therapeutic efficacy and safety [287,288]. Achieving site-specific delivery is challenging, potentially leading to off-target effects [289]. Stability during storage is vital, with factors like temperature fluctuations compromising therapeutic potential [151]. Immunogenicity is a significant concern; while autologous sources mitigate risks, large-scale production from allogenic or xenogenic sources amplifies associated risks [290]. Batch-to-batch variability and contamination risks during isolation compound safety concerns [291]. The potential for horizontal gene transfer by exosomes could inadvertently transfer detrimental genes. As the biomedical field progresses with the rise of EVs and CMDNs, there's an escalating need for reconfigured clinical trial frameworks to address the unique challenges associated with these therapies, particularly due to variable cargo loading efficiencies influenced by variations in vesicular dimensions, intricate lipidomic architectures, and membrane biomechanics [292,293]. Adaptive clinical trial designs become indispensable, allowing for modifiable responses based on interim findings and leveraging real-time pharmacokinetic feedback to optimize dosages [294,295]. The MAMS designs are noteworthy, enabling concurrent evaluations to optimize therapeutic precision [296]. Integration of real-world data is crucial to understand the longitudinal stability and efficacy in real clinical settings, balancing trial controls with patient variability. Safety evaluations should consider the diverse origins of EVs, employing basket and umbrella trial structures to assess immunogenicity risks across different patient cohorts [297]. Sequential Multiple Assignment Randomized Trials (SMART) designs, renowned for flexibility, are pivotal to counter variability and contamination threats, allowing treatment recalibrations based on evolving responses or risks [298,299]. The latent risk in exosomes mediating detrimental horizontal gene transfers demands meticulous dynamic surveillance mechanisms, supported by Bayesian analytical paradigms. In conclusion, to realize the potential of EVs and cell membrane-based nanocarriers without compromising safety, clinical trial methodologies must evolve, incorporating innovative, adaptive, and rigorous designs.

Polysaccharide-based BDSs are renowned for their biocompatibility, biodegradability, and functional modification capacities, making them prominent in drug delivery research [165,172,267,300]. However, alginate exhibits challenges like burst release patterns and syneresis, impacting optimal drug concentrations and release kinetics, and posing potential overdose concerns [301,302]. Contaminants in alginate can also provoke inflammatory responses. Chitosan's solubility is pH-dependent, affecting its efficacy in diverse bodily microenvironments, and variations in its molecular weight distribution can lead to discrepancies in drug loading and release profiles [265]. Its biodegradation kinetics can leave residual fragments in vivo, raising safety concerns including rare allergic reactions. HA's propensity for rapid enzymatic degradation limits its suitability for sustained drug delivery, and its purity is crucial if derived from animal sources to avoid immune responses or pathogen transmission [266]. Dextran, though versatile, presents challenges with variable molecular weights affecting delivery profiles and has rare instances of induced anaphylactic reactions associated with higher molecular weights [166,303].

A holistic assessment of polysaccharide-based BDSs necessitates a transition from traditional to more innovative, flexible clinical trial designs, with adaptive designs becoming pivotal for modifications like dose titrations based on interim analyses, addressing biomimetics' unpredictability [275,294,295]. The efficient MAMS design allows simultaneous evaluations of diverse formulations, swiftly sidelining suboptimal candidates, while platform trials provide a

dynamic scaffold for continuous comparison of polysaccharide derivatives [279,295,296]. The specificity of Umbrella and Basket trials is invaluable for discerning patient subpopulations benefiting from particular formulations, enhancing the precision medicine [297]. Integration of RWE is crucial, offering insights into broader clinical scenarios and assessing the real-world effectiveness of polysaccharide-based BDSs [281]. Incorporating patient feedback in patient-centric trials facilitates comprehensive assessments of biocompatibility and efficacy [304]. However, the employment of such innovative designs involves complexities; they require sophisticated statistical methodologies, continuous monitoring, and transparent, ethical decision-making. In summary, the clinical validation of polysaccharide-based BDS systems is intrinsically linked to the strategic employment of these innovative, nuanced trial designs in therapeutic applications.

In conclusion, while each biomimetic delivery system carries unique opportunities, they all share common challenges in terms of their in vivo behavior, safety, and efficacy profiles. To enable their clinical translation, a comprehensive understanding of these challenges and the development of strategies to address them is crucial. This should include extensive preclinical and clinical evaluation of their pharmacokinetics, pharmacodynamics, and potential for inducing immunogenicity. The successful resolution of these challenges will unlock the therapeutic potential of these biomimetic delivery systems, improving patient outcomes across a range of diseases and conditions.

#### *2.4. Regulatory and Ethical Challenges*

The clinical implementation journey of BDSs encompasses intricate regulatory necessities and significant ethical considerations, often aligning with advanced therapy medicinal products (ATMPs) or nanomedicines, requiring specialized regulatory pathways [305,306]. The diverse forms of BDSs like Liposomes, Albumin, CMDNs, and various polysaccharides, necessitate the formulation of innovative regulatory guidelines and consistent dialogue between researchers and regulatory entities to navigate the clinical translation pathway. Ethical considerations become paramount especially with human-derived biomimetic materials like EVs or RBCs [307], necessitating thorough informed consent processes, strict privacy protection measures, and equitable access considerations encompassing production cost, pricing, and healthcare infrastructure disparities. While addressing technical challenges is crucial, ethical concerns require equal emphasis, requiring a multidimensional approach to harmonize scientific innovation, regulatory compliance, and ethical responsibility in the clinical translation of biomimetic delivery systems.

Liposomes require intricate characterization due to their diverse properties, raising regulatory and informed consent complexities [308], and their predisposition to degradation necessitates stabilization efforts. PNPs like albumin and ferritin pose risks of adverse immune reactions, batch variability, and contamination, particularly from animal-derived proteins, which elevate ethical concerns and can limit acceptability among certain demographics [309]. VLPs, although non-pathogenic, invoke apprehension about potential immunogenic responses and necessitate elevated consent standards due to uncertainties surrounding their long-term effects [310]. NDs, being relatively novel, face challenges in standardization and harbor unresolved ethical considerations. EVs present hurdles in achieving reproducible isolation and purification protocols and pose potential risks in transmitting undesired biomolecules, emphasizing the need for transparency [248–250,290]. CMDNs face challenges in preserving native membrane characteristics while balancing potential immunogenic reactions, especially when sourced from human tissues. Polysaccharides bring forth challenges related to consistency and contamination [172], with their derivation methods potentially conflicting with the preferences or beliefs of certain patient groups, thus intensifying ethical dilemmas.

The emerging BDSs epitomize the integration of nature's complex designs with human technological developments and have brought to the forefront an urgent necessity for advanced regulatory and ethical frameworks tailored to their nuances (Table 3). Historically, the edifice of regulatory standards has been anchored on principles of safety, efficacy, and quality, further buttressed by ethical cornerstones such as informed consent, equity, and transparency. These tried-and-true paradigms, though effective for conventional therapeutics, grapple with the

multifaceted challenges inherent to BDS. A hallmark feature of these systems is their biological variability and complexity, which, while promising targeted precision, complicates the path to achieving consistent reproducibility, i.e., a gold standard in therapeutic evaluations. This variability is compounded by BDS's novel and potentially multifactorial mechanisms of action, which can diverge significantly from traditional therapeutics and demand a deeper level of scrutiny. Further, owing to their intimate mimicry of biological systems, BDSs introduce the possibility of unprecedented interactions with native biological entities, necessitating rigorous preemptive assessment and monitoring. On the ethical front, the material source variability of BDSs introduces intricate layers of concerns, spanning from informed consent and potential exploitation to uncharted territories of long-term biocompatibility and unforeseen systemic effects. The sophistication and innovation underlying BDSs, while promising groundbreaking therapeutic solutions, might also inadvertently escalate production and distribution costs, thus catalyzing debates on equitable accessibility, especially in socioeconomically diverse settings. As the biomedical community stands at this crossroads, a forward-thinking regulatory strategy is of paramount importance. This strategy should champion adaptive oversight mechanisms, foster interdisciplinary dialogues, and advocate for harmonized global standards, ensuring that BDS innovations are not siloed but shared collaboratively. Concurrently, ethical protocols require a renaissance, one that broadens the boundaries of informed consent, deepens stakeholder participation, and relentlessly pursues transparency, ensuring that the transformative potential of BDSs is harmoniously balanced with societal, moral, and patient-centric imperatives. The dawn of biomimetic delivery systems demands a rethinking of our regulatory and ethical scaffolds. While the challenges are intricate, they present an opportunity: to shape a future where innovation flourishes within robust societal safeguards, ensuring that advancements in drug delivery truly serve humanity's best interests.

**Table 3.** Overall insights into regulatory and ethical challenges for BDS.

Categories	Insights
Regulatory Challenges	<ul style="list-style-type: none"><li>• Biological variability &amp; complexity</li><li>• Achieving consistent reproducibility</li><li>• Potential unprecedented interactions with biological entities</li></ul>
Regulatory Frameworks	<ul style="list-style-type: none"><li>• Adaptive oversight mechanisms</li><li>• Interdisciplinary dialogues</li><li>• Harmonized global standards</li></ul>
Ethical Challenges	<ul style="list-style-type: none"><li>• Sourcing material from sentient entities (humans/animals)</li><li>• Informed consent and potential exploitation</li><li>• Equitable accessibility in diverse settings</li><li>• Broadened boundaries of informed consent</li></ul>
Ethical Frameworks	<ul style="list-style-type: none"><li>• Stakeholder participation</li><li>• Pursuit of transparency</li></ul>

3. Conclusions

BDSs have emerged as a transformative frontier in nanomedicine, promising unparalleled advantages in drug delivery and therapeutic modalities. These systems, rooted in the principles of self-assembly, molecular recognition, and biocompatibility, encompass a variety of platforms such as liposomes [46], PNPs [32,120,122,204,311,312], extracellular vesicles [17], and polysaccharides [267]. Their clinical applications have been praised for achievements in targeted delivery, reduced side effects, and improved therapeutic outcomes. However, the journey of these innovative delivery systems from the lab bench to bedside is not without its hurdles. The inherent complexity of biomimetic designs poses challenges in ensuring reproducibility, a crucial factor in clinical applications. The physiological environment presents issues related to stability and longevity of these delivery systems. Moreover, the efficacy and safety of these novel therapies, although promising, need rigorous evaluation. Beyond the technical challenges lie intricate regulatory mazes and ethical considerations that must be navigated to achieve successful clinical translation. To overcome these challenges, the scientific community has turned to various strategies. Technological innovations have



been at the forefront, addressing issues of complexity and reproducibility. The exploration and integration of advanced biomaterials aim to bolster the stability and lifespan of biomimetic systems in physiological settings. Recognizing the unique properties and challenges of biomimetic delivery, there has been a push for innovative clinical trial designs that can more aptly evaluate their efficacy and safety. Furthermore, it's evident that the traditional regulatory and ethical frameworks might fall short, necessitating the evolution of these frameworks in alignment with the innovative nature of biomimetic delivery systems. Real-world case studies provide tangible evidence of these challenges and, more importantly, shed light on successful strategies and interventions that have paved the way for clinical translation. These instances not only offer insights but also inspire confidence in the potential of biomimetic delivery systems to revolutionize healthcare.

The future holds substantial promise for the clinical translation of BDSs as advancements in understanding biological systems continue to refine the design and capabilities of BDSs. Anticipated innovations, emerging from interdisciplinary collaborations among biologists, chemists, engineers, and clinicians, will likely be more refined, efficient, and personalized, aligning with individual patient profiles for optimized outcomes. The evolving familiarity of global regulatory bodies with BDSs anticipates the establishment of more streamlined guidelines, expediting clinical translation. Initial challenges and learnings in clinical translation will be instrumental in refining subsequent iterations of BDSs for enhanced clinical application. In essence, BDSs, merging nature's design with human ingenuity, have immense potential in revolutionizing drug delivery, and despite existing challenges, the commitment of the scientific community and ongoing technological and regulatory advancements underline a future replete with potential.

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## References

1. Sheikhpour, M.; Barani, L.; Kasaeian, A. Biomimetics in drug delivery systems: A critical review. *J Control Release* 2017; 253: 97-109
2. Vincent, J.F.V. Biomimetics — a review. *Proceedings of the Institution of Mechanical Engineers, Part H: Journal of Engineering in Medicine* 2009; 223: 919-939
3. Vincent, J.F.V.; Bogatyreva, O.A.; Bogatyrev, N.R.; et al. Biomimetics: its practice and theory. *Journal of The Royal Society Interface* 2006; 3: 471-482
4. Venkatesh, S.; Byrne, M.E.; Peppas, N.A.; et al. Applications of biomimetic systems in drug delivery. *Expert Opin Drug Deliv* 2005; 2: 1085-1096
5. Fukuta, T.; Kogure, K. Biomimetic Nanoparticle Drug Delivery Systems to Overcome Biological Barriers for Therapeutic Applications. *Chemical and Pharmaceutical Bulletin* 2022; 70: 334-340
6. Chen, Y.-x.; Wei, C.-x.; Lyu, Y.-q.; et al. Biomimetic drug-delivery systems for the management of brain diseases. *Biomaterials Science* 2020; 8: 1073-1088
7. Chen, L.; Hong, W.; Ren, W.; et al. Recent progress in targeted delivery vectors based on biomimetic nanoparticles. *Signal Transduction and Targeted Therapy* 2021; 6: 225
8. Rasheed, T.; Nabeel, F.; Raza, A.; et al. Biomimetic nanostructures/cues as drug delivery systems: a review. *Materials Today Chemistry* 2019; 13: 147-157
9. Zhang, M.; Du, Y.; Wang, S.; et al. A Review of Biomimetic Nanoparticle Drug Delivery Systems Based on Cell Membranes. *Drug Design, Development and Therapy* 2020; 14: 5495-5503

10. Chandrawati, R.; Caruso, F. Biomimetic Liposome- and Polymersome-Based Multicompartmentalized Assemblies. *Langmuir* 2012; 28: 13798-13807
11. Tariq, H.; Batool, S.; Asif, S.; et al. Virus-Like Particles: Revolutionary Platforms for Developing Vaccines Against Emerging Infectious Diseases. *Frontiers in Microbiology* 2022; 12:
12. Nooraei, S.; Bahrulolum, H.; Hoseini, Z.S.; et al. Virus-like particles: preparation, immunogenicity and their roles as nanovaccines and drug nanocarriers. *Journal of Nanobiotechnology* 2021; 19: 59
13. Banskota, S.; Raguram, A.; Suh, S.; et al. Engineered virus-like particles for efficient delivery of therapeutic proteins. *Cell* 2022; 185: 250-265.e216
14. Geckil, H.; Xu, F.; Zhang, X.; et al. Engineering hydrogels as extracellular matrix mimics. *Nanomedicine (Lond)* 2010; 5: 469-484
15. Zhang, Y.; Xu, Y.; Gao, J. The engineering and application of extracellular matrix hydrogels: a review. *Biomaterials Science* 2023; 11: 3784-3799
16. González-Díaz, E.C.; Varghese, S. Hydrogels as Extracellular Matrix Analogs. *Gels* 2016; 2: 20
17. Vader, P.; Mol, E.A.; Pasterkamp, G.; et al. Extracellular vesicles for drug delivery. *Adv Drug Deliv Rev* 2016; 106: 148-156
18. Herrmann, I.K.; Wood, M.J.A.; Fuhrmann, G. Extracellular vesicles as a next-generation drug delivery platform. *Nature Nanotechnology* 2021; 16: 748-759
19. Muzykantov, V.R. Drug delivery by red blood cells: vascular carriers designed by mother nature. *Expert Opin Drug Deliv* 2010; 7: 403-427
20. Villa, C.H.; Anselmo, A.C.; Mitragotri, S.; et al. Red blood cells: Supercarriers for drugs, biologicals, and nanoparticles and inspiration for advanced delivery systems. *Adv Drug Deliv Rev* 2016; 106: 88-103
21. Pol, E.v.d.; Böing, A.N.; Harrison, P.; et al. Classification, Functions, and Clinical Relevance of Extracellular Vesicles. *Pharmacological Reviews* 2012; 64: 676-705
22. Kalluri, R.; LeBleu, V.S. The biology, function, and biomedical applications of exosomes. *Science* 2020; 367: eaau6977
23. Raposo, G.; Stoorvogel, W. Extracellular vesicles: Exosomes, microvesicles, and friends. *Journal of Cell Biology* 2013; 200: 373-383
24. Muralidharan-Chari, V.; Clancy, J.W.; Sedgwick, A.; et al. Microvesicles: mediators of extracellular communication during cancer progression. *Journal of Cell Science* 2010; 123: 1603-1611
25. Théry, C.; Zitvogel, L.; Amigorena, S. Exosomes: composition, biogenesis and function. *Nature Reviews Immunology* 2002; 2: 569-579
26. O'Brien, K.; Breyne, K.; Ughetto, S.; et al. RNA delivery by extracellular vesicles in mammalian cells and its applications. *Nature Reviews Molecular Cell Biology* 2020; 21: 585-606
27. Villa, C.H.; Cines, D.B.; Siegel, D.L.; et al. Erythrocytes as Carriers for Drug Delivery in Blood Transfusion and Beyond. *Transfusion Medicine Reviews* 2017; 31: 26-35
28. Xia, Q.; Zhang, Y.; Li, Z.; et al. Red blood cell membrane-camouflaged nanoparticles: a novel drug delivery system for antitumor application. *Acta Pharm Sin B* 2019; 9: 675-689
29. Glassman, P.M.; Villa, C.H.; Ukidve, A.; et al. Vascular Drug Delivery Using Carrier Red Blood Cells: Focus on RBC Surface Loading and Pharmacokinetics. *Pharmaceutics* 2020, 12, 440, doi:10.3390/pharmaceutics12050440.
30. Tsujita, M.; Wolska, A.; Gutmann, D.A.P.; et al. Reconstituted Discoidal High-Density Lipoproteins: Bioinspired Nanodiscs with Many Unexpected Applications. *Current Atherosclerosis Reports* 2018; 20: 59
31. Murakami, T. Phospholipid nanodisc engineering for drug delivery systems. *Biotechnology Journal* 2012; 7: 762-767
32. Bariwal, J.; Ma, H.; Altenberg, G.A.; et al. Nanodiscs: a versatile nanocarrier platform for cancer diagnosis and treatment. *Chemical Society Reviews* 2022; 51: 1702-1728
33. Traughber, C.A.; Opoku, E.; Brubaker, G.; et al. Uptake of high-density lipoprotein by scavenger receptor class B type 1 is associated with prostate cancer proliferation and tumor progression in mice. *J Biol Chem* 2020; 295: 8252-8261
34. Baranova, I.N.; Kurlander, R.; Bocharov, A.V.; et al. Role of human CD36 in bacterial recognition, phagocytosis, and pathogen-induced JNK-mediated signaling. *J Immunol* 2008; 181: 7147-7156
35. Lei, Z.; Wang, J.; Lv, P.; et al. Biomimetic synthesis of nanovesicles for targeted drug delivery. *Science Bulletin* 2018; 63: 663-665
36. Tu, R.S.; Tirrell, M. Bottom-up design of biomimetic assemblies. *Adv Drug Deliv Rev* 2004; 56: 1537-1563
37. Chen, Z.; Chen, X.; Huang, J.; et al. Harnessing Protein Corona for Biomimetic Nanomedicine Design. *Biomimetics* 2022; 7: 126
38. Tang, Z.; Xiao, Y.; Kong, N.; et al. Nano-bio interfaces effect of two-dimensional nanomaterials and their applications in cancer immunotherapy. *Acta Pharm Sin B* 2021; 11: 3447-3464
39. Liu, Y.; Wang, J.; Xiong, Q.; et al. Nano-Bio Interactions in Cancer: From Therapeutics Delivery to Early Detection. *Acc Chem Res* 2021; 54: 291-301

40. Chen, Z.; Chen, X.; Liu, G.; et al. Editorial: The Application of Nanoengineering in Advanced Drug Delivery and Translational Research. *Front Bioeng Biotechnol* 2022; 10: 886109
41. Li, L.; Wang, J.; Kong, H.; et al. Functional biomimetic nanoparticles for drug delivery and theranostic applications in cancer treatment. *Science and Technology of Advanced Materials* 2018; 19: 771-790
42. Wang, J.; Wang, A.Z.; Lv, P.; et al. Advancing the Pharmaceutical Potential of Bioinorganic Hybrid Lipid-Based Assemblies. *Adv Sci (Weinh)* 2018; 5: 1800564
43. Guido, C.; Maiorano, G.; Cortese, B.; et al. Biomimetic Nanocarriers for Cancer Target Therapy. *Bioengineering* 2020; 7: 111
44. Sabu, C.; Rejo, C.; Kotta, S.; et al. Bioinspired and biomimetic systems for advanced drug and gene delivery. *J Control Release* 2018; 287: 142-155
45. Liu, S.; Yu, J.-M.; Gan, Y.-C.; et al. Biomimetic natural biomaterials for tissue engineering and regenerative medicine: new biosynthesis methods, recent advances, and emerging applications. *Military Medical Research* 2023; 10: 16
46. Liu, P.; Chen, G.; Zhang, J. A Review of Liposomes as a Drug Delivery System: Current Status of Approved Products, Regulatory Environments, and Future Perspectives. *Molecules* 2022; 27: 1372
47. Agmo Hernández, V.; Karlsson, G.; Edwards, K. Intrinsic Heterogeneity in Liposome Suspensions Caused by the Dynamic Spontaneous Formation of Hydrophobic Active Sites in Lipid Membranes. *Langmuir* 2011; 27: 4873-4883
48. Maritim, S.; Boulas, P.; Lin, Y. Comprehensive analysis of liposome formulation parameters and their influence on encapsulation, stability and drug release in glibenclamide liposomes. *Int J Pharm* 2021; 592: 120051
49. Mayer, L.D.; Tai, L.C.L.; Ko, D.S.C.; et al. Influence of Vesicle Size, Lipid Composition, and Drug-to-Lipid Ratio on the Biological Activity of Liposomal Doxorubicin in Mice. *Cancer Research* 1989; 49: 5922-5930
50. Crowe, J.H.; Crowe, L.M. Factors affecting the stability of dry liposomes. *Biochimica et Biophysica Acta (BBA) - Biomembranes* 1988; 939: 327-334
51. Senior, J.H. Fate and behavior of liposomes in vivo: a review of controlling factors. *Crit Rev Ther Drug Carrier Syst* 1987; 3: 123-193
52. Sułkowski, W.W.; Pentak, D.; Nowak, K.; et al. The influence of temperature, cholesterol content and pH on liposome stability. *Journal of Molecular Structure* 2005; 744-747: 737-747
53. Hupfeld, S.; Holsæter, A.M.; Skar, M.; et al. Liposome Size Analysis by Dynamic/Static Light Scattering upon Size Exclusion-/Field Flow-Fractionation. *Journal of Nanoscience and Nanotechnology* 2006; 6: 3025-3031
54. Doyen, C.; Larquet, E.; Coureux, P.-D.; et al. Nuclear Magnetic Resonance Spectroscopy: A Multifaceted Toolbox to Probe Structure, Dynamics, Interactions, and Real-Time In Situ Release Kinetics in Peptide-Liposome Formulations. *Mol Pharm* 2021; 18: 2521-2539
55. Siriwardane, D.A.; Wang, C.; Jiang, W.; et al. Quantification of phospholipid degradation products in liposomal pharmaceutical formulations by ultra performance liquid chromatography-mass spectrometry (UPLC-MS). *Int J Pharm* 2020; 578: 119077
56. Wagner, A.; Vorauer-Uhl, K. Liposome Technology for Industrial Purposes. *Journal of Drug Delivery* 2011; 2011: 591325
57. Carugo, D.; Bottaro, E.; Owen, J.; et al. Liposome production by microfluidics: potential and limiting factors. *Sci Rep* 2016; 6: 25876
58. Parchekani, J.; Allahverdi, A.; Taghdir, M.; et al. Design and simulation of the liposomal model by using a coarse-grained molecular dynamics approach towards drug delivery goals. *Sci Rep* 2022; 12: 2371
59. Jämbeck, J.P.M.; Eriksson, E.S.E.; Laaksonen, A.; et al. Molecular Dynamics Studies of Liposomes as Carriers for Photosensitizing Drugs: Development, Validation, and Simulations with a Coarse-Grained Model. *Journal of Chemical Theory and Computation* 2014; 10: 5-13
60. Langer, K.; Anhorn, M.G.; Steinhäuser, I.; et al. Human serum albumin (HSA) nanoparticles: Reproducibility of preparation process and kinetics of enzymatic degradation. *Int J Pharm* 2008; 347: 109-117
61. Bertucci, C.; Domenici, E. Reversible and covalent binding of drugs to human serum albumin: methodological approaches and physiological relevance. *Current medicinal chemistry* 2002; 9: 1463-1481
62. Galisteo-González, F.; Molina-Bolívar, J.A. Systematic study on the preparation of BSA nanoparticles. *Colloids and Surfaces B: Biointerfaces* 2014; 123: 286-292
63. Maciążek-Jurczyk, M.; Szkudlarek, A.; Chudzik, M.; et al. Alteration of human serum albumin binding properties induced by modifications: A review. *Spectrochim Acta A Mol Biomol Spectrosc* 2018; 188: 675-683
64. Hornok, V. Serum Albumin Nanoparticles: Problems and Prospects. *Polymers* 2021; 13: 3759
65. Das, S.; Jagan, L.; Isiah, R.; et al. Nanotechnology in oncology: Characterization and in vitro release kinetics of cisplatin-loaded albumin nanoparticles: Implications in anticancer drug delivery. *Indian Journal of Pharmacology* 2011; 43: 409
66. Kulig, K.; Ziabka, M.; Pilarczyk, K.; et al. Physicochemical Study of Albumin Nanoparticles with Chlorambucil. *Processes* 2022; 10: 1170

67. Baler, K.; Martin, O.A.; Carignano, M.A.; et al. Electrostatic Unfolding and Interactions of Albumin Driven by pH Changes: A Molecular Dynamics Study. *The Journal of Physical Chemistry B* 2014; 118: 921-930
68. Narwal, M.; Kumar, D.; Mukherjee, T.K.; et al. Molecular dynamics simulation as a tool for assessment of drug binding property of human serum albumin. *Molecular Biology Reports* 2018; 45: 1647-1652
69. Kaboli, S.F.; Mehrnejad, F.; Nematollahzadeh, A. Molecular modeling prediction of albumin-based nanoparticles and experimental preparation, characterization, and in-vitro release kinetics of prednisolone from the nanoparticles. *Journal of Drug Delivery Science and Technology* 2021; 64: 102588
70. Amirinasab, M.; Dehestani, M. Theoretical aspects of interaction of the anticancer drug cytarabine with human serum albumin. *Structural Chemistry* 2023:
71. Muthu, M.S.; Wilson, B. Challenges posed by the scale-up of nanomedicines. *Nanomedicine (Lond)* 2012; 7: 307-309
72. Shrimal, P.; Jadeja, G.; Patel, S. A review on novel methodologies for drug nanoparticle preparation: Microfluidic approach. *Chemical Engineering Research and Design* 2020; 153: 728-756
73. Hakala, T.A.; Davies, S.; Toprakcioglu, Z.; et al. A Microfluidic Co-Flow Route for Human Serum Albumin-Drug-Nanoparticle Assembly. *Chemistry – A European Journal* 2020; 26: 5965-5969
74. Tuan Giam Chuang, V.; Kragh-Hansen, U.; Otagiri, M. Pharmaceutical Strategies Utilizing Recombinant Human Serum Albumin. *Pharm Res* 2002; 19: 569-577
75. Liu, X.; Mohanty, R.P.; Maier, E.Y.; et al. Controlled loading of albumin-drug conjugates ex vivo for enhanced drug delivery and antitumor efficacy. *J Control Release* 2020; 328: 1-12
76. Ghadami, S.A.; Ahmadi, Z.; Moosavi-Nejad, Z. The albumin-based nanoparticle formation in relation to protein aggregation. *Spectrochim Acta A Mol Biomol Spectrosc* 2021; 252: 119489
77. Dawoud, M.H.S.; Abdel-Daim, A.; Nour, M.S.; et al. A Quality by Design Paradigm for Albumin-Based Nanoparticles: Formulation Optimization and Enhancement of the Antitumor Activity. *Journal of Pharmaceutical Innovation* 2023:
78. Sønderby, P.; Bukrinski, J.T.; Hebditch, M.; et al. Self-Interaction of Human Serum Albumin: A Formulation Perspective. *ACS Omega* 2018; 3: 16105-16117
79. Vogel, V.; Langer, K.; Balthasar, S.; et al. Characterization of serum albumin nanoparticles by sedimentation velocity analysis and electron microscopy. Berlin, Heidelberg, 2002; pp. 31-36.
80. Spada, A.; Emami, J.; Tuszynski, J.A.; et al. The Uniqueness of Albumin as a Carrier in Nanodrug Delivery. *Mol Pharm* 2021; 18: 1862-1894
81. Tesarova, B.; Musilek, K.; Rex, S.; et al. Taking advantage of cellular uptake of ferritin nanocages for targeted drug delivery. *J Control Release* 2020; 325: 176-190
82. Zhang, Y.; Orner, B.P. Self-Assembly in the Ferritin Nano-Cage Protein Superfamily. *Int J Mol Sci* 2011; 12: 5406-5421
83. Honarmand Ebrahimi, K.; Hagedoorn, P.-L.; Hagen, W.R. Unity in the Biochemistry of the Iron-Storage Proteins Ferritin and Bacterioferritin. *Chemical Reviews* 2015; 115: 295-326
84. Stühn, L.; Auernhammer, J.; Dietz, C. pH-depended protein shell dis- and reassembly of ferritin nanoparticles revealed by atomic force microscopy. *Sci Rep* 2019; 9: 17755
85. Nakahara, Y.; Endo, Y.; Inoue, I. Construction Protocol of Drug-Protein Cage Complexes for Drug Delivery System. In *Protein Cages: Design, Structure, and Applications*, Ueno, T., Lim, S., Xia, K., Eds.; Springer US: New York, NY, 2023; pp. 335-347.
86. Liu, Y.; Zhao, G. Reassembly Design of Ferritin Cages. In *Protein Cages: Design, Structure, and Applications*, Ueno, T., Lim, S., Xia, K., Eds.; Springer US: New York, NY, 2023; pp. 69-78.
87. Wade, V.J.; Levi, S.; Arosio, P.; et al. Influence of site-directed modifications on the formation of iron cores in ferritin. *Journal of Molecular Biology* 1991; 221: 1443-1452
88. Zhang, B.; Tang, G.; He, J.; et al. Ferritin nanocage: A promising and designable multi-module platform for constructing dynamic nanoassembly-based drug nanocarrier. *Adv Drug Deliv Rev* 2021; 176: 113892
89. Yang Caiyun, C.C., Cai Yao, Zhang Tongwei, Pan Yongxin. The Surface Modification of Ferritin and Its Applications. *Progress in Chemistry* 2016; 28: 91-102
90. Wang, C.; Zhang, C.; Li, Z.; et al. Extending Half Life of H-Ferritin Nanoparticle by Fusing Albumin Binding Domain for Doxorubicin Encapsulation. *Biomacromolecules* 2018; 19: 773-781
91. Zhang, S.; Zang, J.; Chen, H.; et al. The Size Flexibility of Ferritin Nanocage Opens a New Way to Prepare Nanomaterials. *Small* 2017; 13: 1701045
92. Giddings, J.C.; Yang, F.J.; Myers, M.N. Flow field-flow fractionation as a methodology for protein separation and characterization. *Analytical Biochemistry* 1977; 81: 395-407
93. He, D.; Hughes, S.; Vanden-Hehir, S.; et al. Structural characterization of encapsulated ferritin provides insight into iron storage in bacterial nanocompartments. *eLife* 2016; 5: e18972
94. Li, Z.; Maity, B.; Hishikawa, Y.; et al. Importance of the Subunit-Subunit Interface in Ferritin Disassembly: A Molecular Dynamics Study. *Langmuir* 2022; 38: 1106-1113



95. Anjum, F.; Shahwan, M.; Alhumaydhi, F.A.; et al. Mechanistic insight into the binding between Ferritin and Serotonin: Possible implications in neurodegenerative diseases. *Journal of Molecular Liquids* 2022; 351: 118618
96. Banskota, S.; Raguram, A.; Suh, S.; et al. Engineered virus-like particles for efficient in vivo delivery of therapeutic proteins. *Cell* 2022; 185: 250-265.e216
97. Zhang, L.; Lua, L.H.L.; Middelberg, A.P.J.; et al. Biomolecular engineering of virus-like particles aided by computational chemistry methods. *Chemical Society Reviews* 2015; 44: 8608-8618
98. Charlton Hume, H.K.; Vidigal, J.; Carrondo, M.J.T.; et al. Synthetic biology for bioengineering virus-like particle vaccines. *Biotechnology and Bioengineering* 2019; 116: 919-935
99. Fuenmayor, J.; Gòdia, F.; Cervera, L. Production of virus-like particles for vaccines. *New Biotechnology* 2017; 39: 174-180
100. Santi, L.; Huang, Z.; Mason, H. Virus-like particles production in green plants. *Methods* 2006; 40: 66-76
101. Mohr, J.; Chuan, Y.P.; Wu, Y.; et al. Virus-like particle formulation optimization by miniaturized high-throughput screening. *Methods* 2013; 60: 248-256
102. Huhti, L.; Blazevic, V.; Nurminen, K.; et al. A comparison of methods for purification and concentration of norovirus GII-4 capsid virus-like particles. *Archives of Virology* 2010; 155: 1855-1858
103. Rocha, J.M. Aqueous two-phase systems and monolithic chromatography as alternative technological platforms for virus and virus-like particle purification. *Journal of Chemical Technology & Biotechnology* 2021; 96: 309-317
104. Marchel, M.; Niewisiewicz, J.; Coroadinha, A.S.; et al. Purification of virus-like particles using aqueous biphasic systems composed of natural deep eutectic solvents. *Separation and Purification Technology* 2020; 252: 117480
105. Ladd Effio, C.; Oelmeier, S.A.; Hubbuch, J. High-throughput characterization of virus-like particles by interlaced size-exclusion chromatography. *Vaccine* 2016; 34: 1259-1267
106. Pereira Aguilar, P.; González-Domínguez, I.; Schneider, T.A.; et al. At-line multi-angle light scattering detector for faster process development in enveloped virus-like particle purification. *Journal of Separation Science* 2019; 42: 2640-2649
107. Dourous, L.; Rosa-Calatrava, M.; Petiot, E. Advances in influenza virus-like particles bioprocesses. *Expert Review of Vaccines* 2019; 18: 1285-1300
108. Denisov, I.G.; Grinkova, Y.V.; Lazarides, A.A.; et al. Directed Self-Assembly of Monodisperse Phospholipid Bilayer Nanodiscs with Controlled Size. *Journal of the American Chemical Society* 2004; 126: 3477-3487
109. Xu, D.; Chen, X.; Li, Y.; et al. Reconfigurable Peptide Analogs of Apolipoprotein A-I Reveal Tunable Features of Nanodisc Assembly. *Langmuir* 2023; 39: 1262-1276
110. Wade, J.H.; Jones, J.D.; Lenov, I.L.; et al. Microfluidic platform for efficient Nanodisc assembly, membrane protein incorporation, and purification. *Lab on a Chip* 2017; 17: 2951-2959
111. Goluch, E.D.; Shaw, A.W.; Sligar, S.G.; et al. Microfluidic patterning of nanodisc lipid bilayers and multiplexed analysis of protein interaction. *Lab on a Chip* 2008; 8: 1723-1728
112. Xu, D.; Chen, X.; Chen, Z.; et al. An in Silico Approach to Reveal the Nanodisc Formulation of Doxorubicin. *Front Bioeng Biotechnol* 2022; 10: 859255
113. Bengtsen, T.; Holm, V.L.; Kjølbye, L.R.; et al. Structure and dynamics of a nanodisc by integrating NMR, SAXS and SANS experiments with molecular dynamics simulations. *eLife* 2020; 9: e56518
114. Pourmousa, M.; Pastor, R.W. Molecular dynamics simulations of lipid nanodiscs. *Biochimica et Biophysica Acta (BBA) - Biomembranes* 2018; 1860: 2094-2107
115. Yoshida, J.-i.; Nagaki, A.; Yamada, D. Continuous flow synthesis. *Drug Discovery Today: Technologies* 2013; 10: e53-e59
116. Julien, J.A.; Fernandez, M.G.; Brandmier, K.M.; et al. Rapid preparation of nanodiscs for biophysical studies. *Archives of Biochemistry and Biophysics* 2021; 712: 109051
117. Pedrazzani, R.; Ceretti, E.; Zerbini, I.; et al. Biodegradability, toxicity and mutagenicity of detergents: Integrated experimental evaluations. *Ecotoxicology and Environmental Safety* 2012; 84: 274-281
118. Sobrino-Figueroa, A.S. Evaluation of oxidative stress and genetic damage caused by detergents in the zebrafish *Danio rerio* (Cyprinidae). *Comparative Biochemistry and Physiology Part A: Molecular & Integrative Physiology* 2013; 165: 528-532
119. Justesen, B.H.; Günther-Pomorski, T. Chromatographic and electrophoretic methods for nanodisc purification and analysis. *Reviews in Analytical Chemistry* 2014; 33: 165-172
120. Howard, F.H.; Gao, Z.; Mansor, H.B.; et al. Silk Fibroin Nanoparticles: A Biocompatible Multi-Functional Polymer for Drug Delivery. 2023:
121. Foox, M.; Zilberman, M. Drug delivery from gelatin-based systems. *Expert Opin Drug Deliv* 2015; 12: 1547-1563
122. Elzoghby, A.O. Gelatin-based nanoparticles as drug and gene delivery systems: Reviewing three decades of research. *J Control Release* 2013; 172: 1075-1091

123. Yasmin, R.; Shah, M.; Khan, S.A.; et al. Gelatin nanoparticles: a potential candidate for medical applications. *Nanotechnology Reviews* 2017; 6: 191-207
124. Nguyen, T.P.; Nguyen, Q.V.; Nguyen, V.-H.; et al. Silk Fibroin-Based Biomaterials for Biomedical Applications: A Review. *Polymers* 2019; 11: 1933
125. Baci, G.-M.; Cucu, A.-A.; Giurgiu, A.-I.; et al. Advances in Editing Silkworms (*Bombyx mori*) Genome by Using the CRISPR-Cas System. *Insects* 2022; 13: 28
126. Aznar-Cervantes, S.D.; Vicente-Cervantes, D.; Meseguer-Olmo, L.; et al. Influence of the protocol used for fibroin extraction on the mechanical properties and fiber sizes of electrospun silk mats. *Mater Sci Eng C Mater Biol Appl* 2013; 33: 1945-1950
127. DeBari, M.K.; King, C.I., III; Altgold, T.A.; et al. Silk Fibroin as a Green Material. *ACS Biomater Sci Eng* 2021; 7: 3530-3544
128. Khan, S.A. Mini-Review: Opportunities and challenges in the techniques used for preparation of gelatin nanoparticles. *Pak. J. Pharm. Sci* 2020; 33: 221-228
129. Ahmed, M.A.; Al-Kahtani, H.A.; Jaswir, I.; et al. Extraction and characterization of gelatin from camel skin (potential halal gelatin) and production of gelatin nanoparticles. *Saudi Journal of Biological Sciences* 2020; 27: 1596-1601
130. Zhou, P.; Regenstein, J.M. Effects of Alkaline and Acid Pretreatments on Alaska Pollock Skin Gelatin Extraction. *Journal of Food Science* 2005; 70: c392-c396
131. Eggert, S.; Kahl, M.; Bock, N.; et al. An open-source technology platform to increase reproducibility and enable high-throughput production of tailorable gelatin methacryloyl (GelMA) - based hydrogels. *Materials & Design* 2021; 204: 109619
132. Xia, Y.; Xu, R.; Ye, S.; et al. Microfluidic Formulation of Curcumin-Loaded Multiresponsive Gelatin Nanoparticles for Anticancer Therapy. *ACS Biomater Sci Eng* 2023; 9: 3402-3413
133. Solomun, J.I.; Totten, J.D.; Wongpinyochit, T.; et al. Manual Versus Microfluidic-Assisted Nanoparticle Manufacture: Impact of Silk Fibroin Stock on Nanoparticle Characteristics. *ACS Biomater Sci Eng* 2020; 6: 2796-2804
134. Jahanshahi, M.; Sanati, M.H.; Hajizadeh, S.; et al. Gelatin nanoparticle fabrication and optimization of the particle size. *physica status solidi (a)* 2008; 205: 2898-2902
135. Madkhali, O.; Mekhail, G.; Wettig, S.D. Modified gelatin nanoparticles for gene delivery. *Int J Pharm* 2019; 554: 224-234
136. Ghareh nazifam, Z.; Dolatabadi, R.; Baniassadi, M.; et al. Computational analysis of vincristine loaded silk fibroin hydrogel for sustained drug delivery applications: Multiphysics modeling and experiments. *Int J Pharm* 2021; 609: 121184
137. Hathout, R.M.; Metwally, A.A.; Woodman, T.J.; et al. Prediction of Drug Loading in the Gelatin Matrix Using Computational Methods. *ACS Omega* 2020; 5: 1549-1556
138. Carmelo-Luna, F.J.; Mendoza-Wilson, A.M.; Ramos-Clamont Montfort, G.; et al. Synthesis and experimental/computational characterization of sorghum procyanidins-gelatin nanoparticles. *Bioorganic & Medicinal Chemistry* 2021; 42: 116240
139. Bordanaba-Florit, G.; Royo, F.; Kruglik, S.G.; et al. Using single-vesicle technologies to unravel the heterogeneity of extracellular vesicles. *Nature Protocols* 2021; 16: 3163-3185
140. Ingato, D.; Lee, J.U.; Sim, S.J.; et al. Good things come in small packages: Overcoming challenges to harness extracellular vesicles for therapeutic delivery. *J Control Release* 2016; 241: 174-185
141. Tkach, M.; Théry, C. Communication by Extracellular Vesicles: Where We Are and Where We Need to Go. *Cell* 2016; 164: 1226-1232
142. Vogel, R.; Coumans, F.A.W.; Maltesen, R.G.; et al. A standardized method to determine the concentration of extracellular vesicles using tunable resistive pulse sensing. *J Extracell Vesicles* 2016; 5: 31242
143. Morales-Kastresana, A.; Telford, B.; Musich, T.A.; et al. Labeling Extracellular Vesicles for Nanoscale Flow Cytometry. *Sci Rep* 2017; 7: 1878
144. Srivastava, A.; Amreddy, N.; Pareek, V.; et al. Progress in extracellular vesicle biology and their application in cancer medicine. *Wiley Interdiscip Rev Nanomed Nanobiotechnol* 2020; 12: e1621
145. Allelein, S.; Medina-Perez, P.; Lopes, A.L.H.; et al. Potential and challenges of specifically isolating extracellular vesicles from heterogeneous populations. *Sci Rep* 2021; 11: 11585
146. Yan, I.K.; Shukla, N.; Borrelli, D.A.; et al. Use of a Hollow Fiber Bioreactor to Collect Extracellular Vesicles from Cells in Culture. In *Extracellular RNA: Methods and Protocols*, Patel, T., Ed.; Springer New York: New York, NY, 2018; pp. 35-41.
147. Kang, H.; Bae, Y.-h.; Kwon, Y.; et al. Extracellular Vesicles Generated Using Bioreactors and their Therapeutic Effect on the Acute Kidney Injury Model. *Adv Healthc Mater* 2022; 11: 2101606
148. Davies, R.T.; Kim, J.; Jang, S.C.; et al. Microfluidic filtration system to isolate extracellular vesicles from blood. *Lab on a Chip* 2012; 12: 5202-5210

149. Gholizadeh, S.; Shehata Draz, M.; Zarghooni, M.; et al. Microfluidic approaches for isolation, detection, and characterization of extracellular vesicles: Current status and future directions. *Biosensors and Bioelectronics* 2017; 91: 588-605
150. Görgens, A.; Corso, G.; Hagey, D.W.; et al. Identification of storage conditions stabilizing extracellular vesicles preparations. *J Extracell Vesicles* 2022; 11: e12238
151. Zeng, Y.; Qiu, Y.; Jiang, W.; et al. Biological Features of Extracellular Vesicles and Challenges. *Frontiers in Cell and Developmental Biology* 2022; 10: 816698
152. Trenkenschuh, E.; Richter, M.; Heinrich, E.; et al. Enhancing the Stabilization Potential of Lyophilization for Extracellular Vesicles. *Adv Healthc Mater* 2022; 11: 2100538
153. Gómez-de-Mariscal, E.; Maška, M.; Kotrbová, A.; et al. Deep-Learning-Based Segmentation of Small Extracellular Vesicles in Transmission Electron Microscopy Images. *Sci Rep* 2019; 9: 13211
154. Bourgeaux, V.; Lanao, J.M.; Bax, B.E.; et al. Drug-loaded erythrocytes: on the road toward marketing approval. *Drug Design, Development and Therapy* 2016; 10: 665-676
155. Chugh, V.; Vijaya Krishna, K.; Pandit, A. Cell Membrane-Coated Mimics: A Methodological Approach for Fabrication, Characterization for Therapeutic Applications, and Challenges for Clinical Translation. *ACS Nano* 2021; 15: 17080-17123
156. Rao, L.; Cai, B.; Bu, L.-L.; et al. Microfluidic Electroporation-Facilitated Synthesis of Erythrocyte Membrane-Coated Magnetic Nanoparticles for Enhanced Imaging-Guided Cancer Therapy. *ACS Nano* 2017; 11: 3496-3505
157. Yang, H.-C.; Waldman, R.Z.; Chen, Z.; et al. Atomic layer deposition for membrane interface engineering. *Nanoscale* 2018; 10: 20505-20513
158. Huang, L.-L.; Nie, W.; Zhang, J.; et al. Cell-Membrane-Based Biomimetic Systems with Bioorthogonal Functionalities. *Acc Chem Res* 2020; 53: 276-287
159. Yousefi, N.; Tufenkji, N. Probing the Interaction between Nanoparticles and Lipid Membranes by Quartz Crystal Microbalance with Dissipation Monitoring. *Frontiers in Chemistry* 2016; 4: 46
160. McDonnell, J.M. Surface plasmon resonance: towards an understanding of the mechanisms of biological molecular recognition. *Current Opinion in Chemical Biology* 2001; 5: 572-577
161. Fang, R.H.; Kroll, A.V.; Gao, W.; et al. Cell Membrane Coating Nanotechnology. *Adv Mater* 2018; 30: 1706759
162. Read, E.K.; Park, J.T.; Shah, R.B.; et al. Process analytical technology (PAT) for biopharmaceutical products: Part I. concepts and applications. *Biotechnology and Bioengineering* 2010; 105: 276-284
163. Zhang, X.; Ma, G.; Wei, W. Simulation of nanoparticles interacting with a cell membrane: probing the structural basis and potential biomedical application. *NPG Asia Materials* 2021; 13: 52
164. Singh, A.V.; Maharjan, R.-S.; Kanase, A.; et al. Machine-Learning-Based Approach to Decode the Influence of Nanomaterial Properties on Their Interaction with Cells. *ACS Appl Mater Interfaces* 2021; 13: 1943-1955
165. Barclay, T.G.; Day, C.M.; Petrovsky, N.; et al. Review of polysaccharide particle-based functional drug delivery. *Carbohydr Polym* 2019; 221: 94-112
166. Díaz-Montes, E. Dextran: Sources, Structures, and Properties. *Polysaccharides* 2021; 2: 554-565
167. Huang, M.; Khor, E.; Lim, L.-Y. Uptake and Cytotoxicity of Chitosan Molecules and Nanoparticles: Effects of Molecular Weight and Degree of Deacetylation. *Pharm Res* 2004; 21: 344-353
168. Bhattacharya, D.S.; Svehkarev, D.; Bapat, A.; et al. Sulfation Modulates the Targeting Properties of Hyaluronic Acid to P-Selectin and CD44. *ACS Biomater Sci Eng* 2020; 6: 3585-3598
169. Mena-García, A.; Ruiz-Matute, A.I.; Soria, A.C.; et al. Green techniques for extraction of bioactive carbohydrates. *TrAC Trends in Analytical Chemistry* 2019; 119: 115612
170. Huang, G.; Chen, F.; Yang, W.; et al. Preparation, deproteinization and comparison of bioactive polysaccharides. *Trends in Food Science & Technology* 2021; 109: 564-568
171. Zheng, D.; Yang, K.; Nie, Z. Engineering heterogeneity of precision nanoparticles for biomedical delivery and therapy. *VIEW* 2021; 2: 20200067
172. Plucinski, A.; Lyu, Z.; Schmidt, B.V. Polysaccharide nanoparticles: From fabrication to applications. *J Mater Chem B* 2021; 9: 7030-7062
173. Galmarini, S.; Hanusch, U.; Giraud, M.; et al. Beyond Unpredictability: The Importance of Reproducibility in Understanding the Protein Corona of Nanoparticles. *Bioconjugate Chemistry* 2018; 29: 3385-3393
174. Bastogne, T. Quality-by-design of nanopharmaceuticals – a state of the art. *Nanomedicine* 2017; 13: 2151-2157
175. Schnitzer, E.; Pinchuk, I.; Bor, A.; et al. Oxidation of liposomal cholesterol and its effect on phospholipid peroxidation. *Chemistry and Physics of Lipids* 2007; 146: 43-53
176. Schnitzer, E.; Pinchuk, I.; Lichtenberg, D. Peroxidation of liposomal lipids. *European Biophysics Journal* 2007; 36: 499-515
177. Inglut, C.T.; Sorrin, A.J.; Kuruppu, T.; et al. Immunological and Toxicological Considerations for the Design of Liposomes. *Nanomaterials (Basel)* 2020; 10: 190

178. Sainaga Jyothi, V.G.S.; Bulusu, R.; Venkata Krishna Rao, B.; et al. Stability characterization for pharmaceutical liposome product development with focus on regulatory considerations: An update. *Int J Pharm* 2022; 624: 122022
179. Barnadas-Rodríguez, R.; Sabés, M. Factors involved in the production of liposomes with a high-pressure homogenizer. *Int J Pharm* 2001; 213: 175-186
180. Karn, P.R.; Cho, W.; Park, H.-J.; et al. Characterization and stability studies of a novel liposomal cyclosporin A prepared using the supercritical fluid method: comparison with the modified conventional Bangham method. *Int J Nanomedicine* 2013; 8: 365-377
181. Webb, M.S.; Harasym, T.O.; Masin, D.; et al. Sphingomyelin-cholesterol liposomes significantly enhance the pharmacokinetic and therapeutic properties of vincristine in murine and human tumour models. *British Journal of Cancer* 1995; 72: 896-904
182. Roy, B.; Guha, P.; Bhattarai, R.; et al. Influence of Lipid Composition, pH, and Temperature on Physicochemical Properties of Liposomes with Curcumin as Model Drug. *Journal of Oleo Science* 2016; 65: 399-411
183. Grit, M.; Crommelin, D.J.A. Chemical stability of liposomes: implications for their physical stability. *Chemistry and Physics of Lipids* 1993; 64: 3-18
184. Arouri, A.; Hansen, A.H.; Rasmussen, T.E.; et al. Lipases, liposomes and lipid-prodrugs. *Current Opinion in Colloid & Interface Science* 2013; 18: 419-431
185. Flaten, G.E.; Chang, T.-T.; Phillips, W.T.; et al. Liposomal formulations of poorly soluble camptothecin: drug retention and biodistribution. *Journal of Liposome Research* 2013; 23: 70-81
186. Basáñez, G.; Goñi, F.M.; Alonso, A. Poly(ethylene glycol)-lipid conjugates inhibit phospholipase C-induced lipid hydrolysis, liposome aggregation and fusion through independent mechanisms. *FEBS Letters* 1997; 411: 281-286
187. Mickova, A.; Buzgo, M.; Benada, O.; et al. Core/Shell Nanofibers with Embedded Liposomes as a Drug Delivery System. *Biomacromolecules* 2012; 13: 952-962
188. Maurer, N.; Fenske, D.B.; Cullis, P.R. Developments in liposomal drug delivery systems. *Expert Opinion on Biological Therapy* 2001; 1: 923-947
189. Mishra, V.; Heath, R.J. Structural and Biochemical Features of Human Serum Albumin Essential for Eukaryotic Cell Culture. *Int J Mol Sci* 2021; 22: 8411
190. Lebedeva, N.S.; Yurina, E.S.; Gubarev, Y.A.; et al. Molecular mechanisms causing albumin aggregation. The main role of the porphyrins of the blood group. *Spectrochim Acta A Mol Biomol Spectrosc* 2021; 246: 118975
191. Wang, S.-L.; Lin, S.-Y.; Li, M.-J.; et al. Temperature effect on the structural stability, similarity, and reversibility of human serum albumin in different states. *Biophysical Chemistry* 2005; 114: 205-212
192. Oliva, A.; Santoveña, A.; Llabres, M.; et al. Stability Study of Human Serum Albumin Pharmaceutical Preparations. *Journal of Pharmacy and Pharmacology* 2010; 51: 385-392
193. A. A. Aljabali, A.; A. Bakshi, H.; L. Hakkim, F.; et al. Albumin Nano-Encapsulation of Piceatannol Enhances Its Anticancer Potential in Colon Cancer Via Downregulation of Nuclear p65 and HIF-1 $\alpha$ . *Cancers* 2020; 12: 113
194. Narayana, S.; Gulzar Ahmed, M.; Nasrine, A. Effect of nano-encapsulation using human serum albumin on anti-angiogenesis activity of bevacizumab to target corneal neovascularization: Development, optimization and in vitro assessment. *Materials Today: Proceedings* 2022; 68: 93-104
195. Fahrlander, E.; Schelhaas, S.; Jacobs, A.H.; et al. PEGylated human serum albumin (HSA) nanoparticles: preparation, characterization and quantification of the PEGylation extent. *Nanotechnology* 2015; 26: 145103
196. Niknejad, H.; Mahmoudzadeh, R. Comparison of different crosslinking methods for preparation of docetaxel-loaded albumin nanoparticles. *Iranian journal of pharmaceutical research: IJPR* 2015; 14: 385
197. Anhorn, M.G.; Mahler, H.-C.; Langer, K. Freeze drying of human serum albumin (HSA) nanoparticles with different excipients. *Int J Pharm* 2008; 363: 162-169
198. Anraku, M.; Kouno, Y.; Kai, T.; et al. The role of N-acetyl-methionine as a new stabilizer for albumin products. *Int J Pharm* 2007; 329: 19-24
199. Meng, R.; Zhu, H.; Deng, P.; et al. Research progress on albumin-based hydrogels: Properties, preparation methods, types and its application for antitumor-drug delivery and tissue engineering. *Front Bioeng Biotechnol* 2023; 11: 1137145
200. Zhao, M.; Li, Z.; Li, X.; et al. Molecular imprinting of doxorubicin by refolding thermally denatured bovine serum albumin and cross-linking with hydrogel network. *Reactive and Functional Polymers* 2021; 168: 105036
201. Zhang, J.; Hao, Y.; Tian, X.; et al. Multi-stimuli responsive molecularly imprinted nanoparticles with tailorable affinity for modulated specific recognition of human serum albumin. *J Mater Chem B* 2022; 10: 6634-6643
202. Raja, S.T.K.; Thiruselvi, T.; Mandal, A.B.; et al. pH and redox sensitive albumin hydrogel: A self-derived biomaterial. *Sci Rep* 2015; 5: 15977



203. Zheng, C.; Wang, L.; Gao, C. pH-sensitive bovine serum albumin nanoparticles for paclitaxel delivery and controlled release to cervical cancer. *Applied Nanoscience* 2022; 12: 4047-4057
204. Khoshnejad, M.; Parhiz, H.; Shuvaev, V.V.; et al. Ferritin-based drug delivery systems: Hybrid nanocarriers for vascular immunotargeting. *J Control Release* 2018; 282: 13-24
205. Yin, S.; Davey, K.; Dai, S.; et al. A critical review of ferritin as a drug nanocarrier: Structure, properties, comparative advantages and challenges. *Particuology* 2022; 64: 65-84
206. Houser, K.V.; Chen, G.L.; Carter, C.; et al. Safety and immunogenicity of a ferritin nanoparticle H2 influenza vaccine in healthy adults: a phase 1 trial. *Nature Medicine* 2022; 28: 383-391
207. Morikawa, K.; Oseko, F.; Morikawa, S. A Role for Ferritin in Hematopoiesis and the Immune System. *Leukemia & Lymphoma* 1995; 18: 429-433
208. Sun, X.; Hong, Y.; Gong, Y.; et al. Bioengineered Ferritin Nanocarriers for Cancer Therapy. *Int J Mol Sci* 2021; 22: 7023
209. Singh, A.; Singhal, B. Role of Machine Learning in Bioprocess Engineering: Current Perspectives and Future Directions. In *Design and Applications of Nature Inspired Optimization: Contribution of Women Leaders in the Field*, Singh, D., Garg, V., Deep, K., Eds.; Springer International Publishing: Cham, 2022; pp. 39-54.
210. Gupta, R.; Arora, K.; Roy, S.S.; et al. Platforms, advances, and technical challenges in virus-like particles-based vaccines. *Frontiers in Immunology* 2023; 14: 1123805
211. Lv, P.; Liu, X.; Chen, X.; et al. Genetically Engineered Cell Membrane Nanovesicles for Oncolytic Adenovirus Delivery: A Versatile Platform for Cancer Virotherapy. *Nano Letters* 2019; 19: 2993-3001
212. Suffian, I.F.B.M.; Al-Jamal, K.T. Bioengineering of virus-like particles as dynamic nanocarriers for in vivo delivery and targeting to solid tumours. *Adv Drug Deliv Rev* 2022; 180: 114030
213. Biabanikhankahdani, R.; Alitheen, N.B.M.; Ho, K.L.; et al. pH-responsive Virus-like Nanoparticles with Enhanced Tumour-targeting Ligands for Cancer Drug Delivery. *Sci Rep* 2016; 6: 37891
214. Hu, H.; Steinmetz, N.F. Doxorubicin-Loaded Physalis Mottle Virus Particles Function as a pH-Responsive Prodrug Enabling Cancer Therapy. *Biotechnology Journal* 2020; 15: 2000077
215. Serradell, M.C.; Rupil, L.L.; Martino, R.A.; et al. Efficient oral vaccination by bioengineering virus-like particles with protozoan surface proteins. *Nature Communications* 2019; 10: 361
216. Ali, A.; Ganguillet, S.; Turgay, Y.; et al. Surface crosslinking of virus-like particles increases resistance to proteases, low pH and mechanical stress for mucosal applications. *bioRxiv* 2023; 2023.2007.2029.550271
217. Shi, L.; Sanyal, G.; Ni, A.; et al. Stabilization of human papillomavirus virus-like particles by non-ionic surfactants. *Journal of Pharmaceutical Sciences* 2005; 94: 1538-1551
218. Schumacher, J.; Bacic, T.; Staritzbichler, R.; et al. Enhanced stability of a chimeric hepatitis B core antigen virus-like-particle (HBcAg-VLP) by a C-terminal linker-hexahistidine-peptide. *Journal of Nanobiotechnology* 2018; 16: 39
219. Gleiter, S.; Lilie, H. Coupling of antibodies via protein Z on modified polyoma virus-like particles. *Protein Science* 2001; 10: 434-444
220. Segel, M.; Lash, B.; Song, J.; et al. Mammalian retrovirus-like protein PEG10 packages its own mRNA and can be pseudotyped for mRNA delivery. *Science* 2021; 373: 882-889
221. Himbert, S.; Rheinstädter, M. Erythro-VLP: Erythrocyte Virus-Like-Particles. *Biophysical Journal* 2021; 120: 196a
222. Grushin, K.; White, M.A.; Stoilova-McPhie, S. Reversible stacking of lipid nanodiscs for structural studies of clotting factors. *Nanotechnology Reviews* 2017; 6: 139-148
223. Hoi, K.K.; Robinson, C.V.; Marty, M.T. Unraveling the Composition and Behavior of Heterogeneous Lipid Nanodiscs by Mass Spectrometry. *Analytical Chemistry* 2016; 88: 6199-6204
224. Damiani, S.; Scheberl, A.; Zayni, S.; et al. Albumin-bound nanodiscs as delivery vehicle candidates: Development and characterization. *Biophysical Chemistry* 2019; 251: 106178
225. Chen, T.; Pan, F.; Luo, G.; et al. Morphology-driven protein corona manipulation for preferential delivery of lipid nanodiscs. *Nano Today* 2022; 46: 101609
226. Dane, E.L.; Belessiotis-Richards, A.; Backlund, C.; et al. STING agonist delivery by tumour-penetrating PEG-lipid nanodiscs primes robust anticancer immunity. *Nature Materials* 2022; 21: 710-720
227. Zhang, W.; Sun, J.; Liu, Y.; et al. PEG-Stabilized Bilayer Nanodisks As Carriers for Doxorubicin Delivery. *Mol Pharm* 2014; 11: 3279-3290
228. Li, Z.; Gu, L. Effects of Mass Ratio, pH, Temperature, and Reaction Time on Fabrication of Partially Purified Pomegranate Ellagitannin-Gelatin Nanoparticles. *Journal of Agricultural and Food Chemistry* 2011; 59: 4225-4231
229. Lammel, A.S.; Hu, X.; Park, S.-H.; et al. Controlling silk fibroin particle features for drug delivery. *Biomaterials* 2010; 31: 4583-4591
230. Yan, H.-B.; Zhang, Y.-Q.; Ma, Y.-L.; et al. Biosynthesis of insulin-silk fibroin nanoparticles conjugates and in vitro evaluation of a drug delivery system. *Journal of Nanoparticle Research* 2009; 11: 1937-1946
231. Leo, E.; Angela Vandelli, M.; Cameroni, R.; et al. Doxorubicin-loaded gelatin nanoparticles stabilized by glutaraldehyde: Involvement of the drug in the cross-linking process. *Int J Pharm* 1997; 155: 75-82

232. Kaul, G.; Amiji, M. Long-Circulating Poly(Ethylene Glycol)-Modified Gelatin Nanoparticles for Intracellular Delivery. *Pharm Res* 2002; 19: 1061-1067
233. Totten, J.D.; Wongpinyochit, T.; Carrola, J.; et al. PEGylation-Dependent Metabolic Rewiring of Macrophages with Silk Fibroin Nanoparticles. *ACS Appl Mater Interfaces* 2019; 11: 14515-14525
234. Gonçalves, A.S.C.; Rodrigues, C.F.; Fernandes, N.; et al. IR780 loaded gelatin-PEG coated gold core silica shell nanorods for cancer-targeted photothermal/photodynamic therapy. *Biotechnology and Bioengineering* 2022; 119: 644-656
235. Jia, L.; Guo, L.; Zhu, J.; et al. Stability and cytocompatibility of silk fibroin-capped gold nanoparticles. *Mater Sci Eng C Mater Biol Appl* 2014; 43: 231-236
236. Curcio, M.; Altimari, I.; Spizzirri, U.G.; et al. Biodegradable gelatin-based nanospheres as pH-responsive drug delivery systems. *Journal of Nanoparticle Research* 2013; 15: 1581
237. Sun, N.; Lei, R.; Xu, J.; et al. Fabricated porous silk fibroin particles for pH-responsive drug delivery and targeting of tumor cells. *Journal of Materials Science* 2019; 54: 3319-3330
238. Deveci, S.S.; Basal, G. Preparation of PCM microcapsules by complex coacervation of silk fibroin and chitosan. *Colloid and Polymer Science* 2009; 287: 1455-1467
239. Zhou, Y.; Yang, H.; Liu, X.; et al. Electrospinning of carboxyethyl chitosan/poly(vinyl alcohol)/silk fibroin nanoparticles for wound dressings. *Int J Biol Macromol* 2013; 53: 88-92
240. Fathollahipour, S.; Abouei Mehrizi, A.; Ghaee, A.; et al. Electrospinning of PVA/chitosan nanocomposite nanofibers containing gelatin nanoparticles as a dual drug delivery system. *Journal of Biomedical Materials Research Part A* 2015; 103: 3852-3862
241. Patra, S.; Basak, P.; Tibarewala, D.N. Synthesis of gelatin nano/submicron particles by binary nonsolvent aided coacervation (BNAC) method. *Mater Sci Eng C Mater Biol Appl* 2016; 59: 310-318
242. Xie, M.; Fan, D.; Li, Y.; et al. Supercritical carbon dioxide-developed silk fibroin nanopatform for smart colon cancer therapy. *Int J Nanomedicine* 2017; 12: 7751-7761
243. Rnjak-Kovacina, J.; DesRochers, T.M.; Burke, K.A.; et al. The Effect of Sterilization on Silk Fibroin Biomaterial Properties. *Macromolecular Bioscience* 2015; 15: 861-874
244. Jeyaram, A.; Jay, S.M. Preservation and Storage Stability of Extracellular Vesicles for Therapeutic Applications. *The AAPS Journal* 2017; 20: 1
245. Yuan, F.; Li, Y.-M.; Wang, Z. Preserving extracellular vesicles for biomedical applications: consideration of storage stability before and after isolation. *Drug Delivery* 2021; 28: 1501-1509
246. Midekessa, G.; Godakumara, K.; Ord, J.; et al. Zeta Potential of Extracellular Vesicles: Toward Understanding the Attributes that Determine Colloidal Stability. *ACS Omega* 2020; 5: 16701-16710
247. Schulz, E.; Karagianni, A.; Koch, M.; et al. Hot EVs – How temperature affects extracellular vesicles. *Eur J Pharm Biopharm* 2020; 146: 55-63
248. Ramirez, M.I.; Amorim, M.G.; Gadelha, C.; et al. Technical challenges of working with extracellular vesicles. *Nanoscale* 2018; 10: 881-906
249. Clemmens, H.; Lambert, D.W. Extracellular vesicles: translational challenges and opportunities. *Biochemical Society Transactions* 2018; 46: 1073-1082
250. Melling, G.E.; Carollo, E.; Conlon, R.; et al. The Challenges and Possibilities of Extracellular Vesicles as Therapeutic Vehicles. *Eur J Pharm Biopharm* 2019; 144: 50-56
251. Bahr, M.M.; Amer, M.S.; Abo-El-Sooud, K.; et al. Preservation techniques of stem cells extracellular vesicles: a gate for manufacturing of clinical grade therapeutic extracellular vesicles and long-term clinical trials. *International Journal of Veterinary Science and Medicine* 2020; 8: 1-8
252. Lv, K.; Li, Q.; Zhang, L.; et al. Incorporation of small extracellular vesicles in sodium alginate hydrogel as a novel therapeutic strategy for myocardial infarction. *Theranostics* 2019; 9: 7403-7416
253. Piffoux, M.; Silva, A.K.A.; Wilhelm, C.; et al. Modification of Extracellular Vesicles by Fusion with Liposomes for the Design of Personalized Biogenic Drug Delivery Systems. *ACS Nano* 2018; 12: 6830-6842
254. Richter, M.; Vader, P.; Fuhrmann, G. Approaches to surface engineering of extracellular vesicles. *Adv Drug Deliv Rev* 2021; 173: 416-426
255. Gardiner, C.; Vizio, D.D.; Sahoo, S.; et al. Techniques used for the isolation and characterization of extracellular vesicles: results of a worldwide survey. *J Extracell Vesicles* 2016; 5: 32945
256. de Jong, O.G.; Kooijmans, S.A.A.; Murphy, D.E.; et al. Drug Delivery with Extracellular Vesicles: From Imagination to Innovation. *Acc Chem Res* 2019; 52: 1761-1770
257. Malhotra, S.; Dumoga, S.; Singh, N. Red blood cells membrane-derived nanoparticles: Applications and key challenges in their clinical translation. *Wiley Interdiscip Rev Nanomed Nanobiotechnol* 2022; 14: e1776
258. Abbina, S.; Parambath, A. 14 - PEGylation and its alternatives: A summary. In *Engineering of Biomaterials for Drug Delivery Systems*, Parambath, A., Ed.; Woodhead Publishing: 2018; pp. 363-376.
259. Doshi, N.; Zahr, A.S.; Bhaskar, S.; et al. Red blood cell-mimicking synthetic biomaterial particles. *Proc Natl Acad Sci U S A* 2009; 106: 21495-21499
260. Krishnan, N.; Fang, R.H.; Zhang, L. Engineering of stimuli-responsive self-assembled biomimetic nanoparticles. *Adv Drug Deliv Rev* 2021; 179: 114006

261. Hu, C.-M.J.; Fang, R.H.; Zhang, L. Erythrocyte-Inspired Delivery Systems. *Adv Healthc Mater* 2012; 1: 537-547
262. Scott, K.L.; Lecak, J.; Acker, J.P. Biopreservation of Red Blood Cells: Past, Present, and Future. *Transfusion Medicine Reviews* 2005; 19: 127-142
263. Han, Y.; Quan, G.B.; Liu, X.Z.; et al. Improved preservation of human red blood cells by lyophilization. *Cryobiology* 2005; 51: 152-164
264. Lee, K.Y.; Mooney, D.J. Alginate: Properties and biomedical applications. *Progress in Polymer Science* 2012; 37: 106-126
265. Kumar, M.N.V.R.; Muzzarelli, R.A.A.; Muzzarelli, C.; et al. Chitosan Chemistry and Pharmaceutical Perspectives. *Chemical Reviews* 2004; 104: 6017-6084
266. Meyer, K. THE BIOLOGICAL SIGNIFICANCE OF HYALURONIC ACID AND HYALURONIDASE. *Physiological Reviews* 1947; 27: 335-359
267. Sirisha, V.; D'Souza, J.S. Polysaccharide-based nanoparticles as drug delivery systems. *Marine OMICS* 2016; 18: 663-702
268. Moosavian, S.A.; Bianconi, V.; Pirro, M.; et al. Challenges and pitfalls in the development of liposomal delivery systems for cancer therapy. *Seminars in Cancer Biology* 2021; 69: 337-348
269. Sercombe, L.; Veerati, T.; Moheimani, F.; et al. Advances and Challenges of Liposome Assisted Drug Delivery. *Frontiers in Pharmacology* 2015; 6: 286
270. Sawant, R.R.; Torchilin, V.P. Challenges in Development of Targeted Liposomal Therapeutics. *The AAPS Journal* 2012; 14: 303-315
271. Guan, J.; Shen, Q.; Zhang, Z.; et al. Enhanced immunocompatibility of ligand-targeted liposomes by attenuating natural IgM absorption. *Nature Communications* 2018; 9: 2982
272. Muro, S. Challenges in design and characterization of ligand-targeted drug delivery systems. *J Control Release* 2012; 164: 125-137
273. Maeda, H. Toward a full understanding of the EPR effect in primary and metastatic tumors as well as issues related to its heterogeneity. *Adv Drug Deliv Rev* 2015; 91: 3-6
274. Deshpande, P.P.; Biswas, S.; Torchilin, V.P. Current trends in the use of liposomes for tumor targeting. *Nanomedicine (Lond)* 2013; 8: 1509-1528
275. Pallmann, P.; Bedding, A.W.; Choodari-Oskoei, B.; et al. Adaptive designs in clinical trials: why use them, and how to run and report them. *BMC Medicine* 2018; 16: 29
276. Meunier, F.; Prentice, H.G.; Ringdén, O. Liposomal amphotericin B (AmBisome): safety data from a phase II/III clinical trial. *Journal of Antimicrobial Chemotherapy* 1991; 28: 83-91
277. Stone, N.R.H.; Bicanic, T.; Salim, R.; et al. Liposomal Amphotericin B (AmBisome®): A Review of the Pharmacokinetics, Pharmacodynamics, Clinical Experience and Future Directions. *Drugs* 2016; 76: 485-500
278. Parmar, M.K.B.; Barthel, F.M.-S.; Sydes, M.; et al. Speeding up the Evaluation of New Agents in Cancer. *J Natl Cancer Inst* 2008; 100: 1204-1214
279. Wason, J.M.S.; Jaki, T. Optimal design of multi-arm multi-stage trials. *Statistics in Medicine* 2012; 31: 4269-4279
280. Burcu, M.; Manzano-Salgado, C.B.; Butler, A.M.; et al. A Framework for Extension Studies Using Real-World Data to Examine Long-Term Safety and Effectiveness. *Therapeutic Innovation & Regulatory Science* 2022; 56: 15-22
281. Sherman, R.E.; Anderson, S.A.; Dal Pan, G.J.; et al. Real-World Evidence — What Is It and What Can It Tell Us? *New England Journal of Medicine* 2016; 375: 2293-2297
282. Kianfar, E. Protein nanoparticles in drug delivery: animal protein, plant proteins and protein cages, albumin nanoparticles. *Journal of Nanobiotechnology* 2021; 19: 159
283. Mollazadeh, S.; Yazdimamaghani, M.; Yazdian-Robati, R.; et al. New insight into the structural changes of apoferritin pores in the process of doxorubicin loading at an acidic pH: Molecular dynamics simulations. *Computers in Biology and Medicine* 2022; 141: 105158
284. Arosio, P.; Levi, S. Ferritin, iron homeostasis, and oxidative damage<sup>1, 2</sup> Guest Editor: Mario Comporti  
<sup>2</sup>This article is part of a series of reviews on "Iron and Cellular Redox Status." The full list of papers may be found on the homepage of the journal. *Free Radical Biology and Medicine* 2002; 33: 457-463
285. Harro, C.D.; Pang, Y.-Y.S.; Roden, R.B.S.; et al. Safety and Immunogenicity Trial in Adult Volunteers of a Human Papillomavirus 16 L1 Virus-Like Particle Vaccine. *J Natl Cancer Inst* 2001; 93: 284-292
286. Carissimi, G.; Montalbán, M.G.; Fuster, M.G.; et al. Silk Fibroin Nanoparticles: Synthesis and Applications as Drug Nanocarriers. *21st Century Nanostructured Materials: Physics, Chemistry, Classification, and Emerging Applications in Industry, Biomedicine, and Agriculture* 2022: 205
287. Meng, W.; He, C.; Hao, Y.; et al. Prospects and challenges of extracellular vesicle-based drug delivery system: considering cell source. *Drug Delivery* 2020; 27: 585-598
288. Le, Q.-V.; Lee, J.; Lee, H.; et al. Cell membrane-derived vesicles for delivery of therapeutic agents. *Acta Pharm Sin B* 2021; 11: 2096-2113

289. Noren Hooten, N.; Yáñez-Mó, M.; DeRita, R.; et al. Hitting the Bullseye: Are extracellular vesicles on target? *J Extracell Vesicles* 2020; 10: e12032
290. Ortiz, A. Not all extracellular vesicles were created equal: clinical implications. *Annals of Translational Medicine* 2017; 5: 111
291. Ghodsi, M.; Cloos, A.-S.; Mozaheb, N.; et al. Variability of extracellular vesicle release during storage of red blood cell concentrates is associated with differential membrane alterations, including loss of cholesterol-enriched domains. *Frontiers in Physiology* 2023; 14: 1205493
292. Loch-Neckel, G.; Matos, A.T.; Vaz, A.R.; et al. Challenges in the Development of Drug Delivery Systems Based on Small Extracellular Vesicles for Therapy of Brain Diseases. *Frontiers in Pharmacology* 2022; 13: 839790
293. Ravasco, J.M.J.M.; Paiva-Santos, A.C.; Conde, J. Technological challenges of biomembrane-coated top-down cancer nanotherapy. *Nature Reviews Bioengineering* 2023; 1: 156-158
294. Saxena, A.; Rubens, M.; Ramamoorthy, V.; et al. A Brief Overview of Adaptive Designs for Phase I Cancer Trials. *Cancers* 2022; 14: 1566
295. Angus, D.C.; Alexander, B.M.; Berry, S.; et al. Adaptive platform trials: definition, design, conduct and reporting considerations. *Nature Reviews Drug Discovery* 2019; 18: 797-807
296. Ghosh, P.; Liu, L.; Senchaudhuri, P.; et al. Design and monitoring of multi-arm multi-stage clinical trials. *Biometrics* 2017; 73: 1289-1299
297. Park, J.J.H.; Hsu, G.; Siden, E.G.; et al. An overview of precision oncology basket and umbrella trials for clinicians. *CA: A Cancer Journal for Clinicians* 2020; 70: 125-137
298. Kidwell, K.M.; Almirall, D. Sequential, Multiple Assignment, Randomized Trial Designs. *JAMA* 2023; 329: 336-337
299. Kim, H.M. Get SMART — Understanding Sequential Multiple Assignment Randomized Trials. *NEJM Evidence* 2023; 2: EVIDe2300031
300. Miao, T.; Wang, J.; Zeng, Y.; et al. Polysaccharide-Based Controlled Release Systems for Therapeutics Delivery and Tissue Engineering: From Bench to Bedside. *Adv Sci (Weinh)* 2018; 5: 1700513
301. Yasmin, F.; Chen, X.; Eames, B.F. Effect of Process Parameters on the Initial Burst Release of Protein-Loaded Alginate Nanospheres. *Journal of Functional Biomaterials* 2019; 10: 42
302. Hannon, G.; Prina-Mello, A. Endotoxin contamination of engineered nanomaterials: Overcoming the hurdles associated with endotoxin testing. *Wiley Interdiscip Rev Nanomed Nanobiotechnol* 2021; 13: e1738
303. Paull, J. A Prospective Study of Dextran-induced Anaphylactoid Reactions in 5745 Patients. *Anaesthesia and Intensive Care* 1987; 15: 163-167
304. Stegemann, S.; Klingmann, V.; Reidemeister, S.; et al. Patient-centric drug product development: Acceptability across patient populations – Science and evidence. *Eur J Pharm Biopharm* 2023; 188: 1-5
305. Wasti, S.; Lee, I.H.; Kim, S.; et al. Ethical and legal challenges in nanomedical innovations: a scoping review. *Frontiers in Genetics* 2023; 14: 1163392
306. Paradise, J. Regulating nanomedicine at the food and drug administration. *AMA journal of ethics* 2019; 21: 347-355
307. Allon, I.; Ben-Yehudah, A.; Dekel, R.; et al. Ethical issues in nanomedicine: Tempest in a teapot? *Medicine, Health Care and Philosophy* 2017; 20: 3-11
308. Belfiore, L.; Saunders, D.N.; Ranson, M.; et al. Towards clinical translation of ligand-functionalized liposomes in targeted cancer therapy: Challenges and opportunities. *J Control Release* 2018; 277: 1-13
309. Clark, D.P.; Pazdernik, N.J. Chapter 21 - Viral and Prion Infections. In *Biotechnology (Second Edition)*, Clark, D.P., Pazdernik, N.J., Eds.; Academic Cell: Boston, 2016; pp. 663-685.
310. Mittal, M.; Banerjee, M.; Lua, L.H.; et al. Current status and future challenges in transitioning to continuous bioprocessing of virus-like particles. *Journal of Chemical Technology & Biotechnology* 2022; 97: 2376-2385
311. Larsen, M.T.; Kuhlmann, M.; Hvam, M.L.; et al. Albumin-based drug delivery: harnessing nature to cure disease. *Molecular and Cellular Therapies* 2016; 4: 3
312. Rohovie, M.J.; Nagasawa, M.; Swartz, J.R. Virus-like particles: Next-generation nanoparticles for targeted therapeutic delivery. *Bioengineering & Translational Medicine* 2017; 2: 43-57

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