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Posted Date: 15 September 2023

doi: 10.20944/preprints202309.1088.v1

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## Article

# Fetuin-A Modulates Tumor Growth and Invasion in a Basal-Like Triple Negative Breast Cancer Cell Line, MDA-MB-468

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**Abstract:** The present studies were undertaken to address the role of fetuin-A in the growth and invasion potential in a triple negative breast cancer (TNBC) cell line, MDA-MB-468. Basal like TNBC that express high levels of ectopic fetuin-A have poorer prognosis for the patients compared to those that express low levels of the protein. We overexpressed fetuin-A in MDA-MB-468 and then determined the invasive potential of fetuin-A overexpressing cells vs controls transfected with empty vector. We also determined the adhesion and growth potential of the cells in the presence of only fetuin-A in serum free medium and also in complete medium. Our data suggest that fetuin-A overexpression significantly enhances the invasive potential of the cells and also the expression of Toll like receptor 4 (TLR4) on these cells. More importantly, the cells rely on fetuin-A-TLR4 signaling network for growth and invasion because the specific TLR4 inhibitor CLI-095 (resatorvid) abrogates fetuin-A mediated growth and invasion. Taken together, the data suggest that fetuin-A-TLR4 signaling network plays a significant role in the growth and invasion potential of TNBC.

**Keywords:** fetuin-A; triple negative; breast cancer; TLR4; invasion; basal-like

## 1. Introduction

We previously demonstrated the significance of fetuin-A in the initiation and progression of mammary tumors using a transgenic mouse model for breast cancer. In this model system, we were able to demonstrate that lack of fetuin-A in the PyMT transgenic background significantly prolonged the latency period of mammary tumor development. Furthermore, tumors that formed in mice that lacked fetuin-A were much smaller in size with low number of proliferating cells [1]. However, this model was based on liver produced fetuin-A. In the present studies we considered not only the role played by fetuin-A supplied to the tumor, but also ectopic fetuin-A produced by the tumor.

Fetuin-A is presently considered a multifunctional protein with a potential to play several roles in tumorigenesis [2]. It is a serum glycoprotein primarily synthesized and secreted by liver parenchymal cells. It has a molecular weight of approximately 50 kDa and is heavily glycosylated [3]. Whereas its main physiological function is inhibition of ectopic calcification, reports show that fetuin-A synthesized by tumor cells (ectopic fetuin-A) plays pivotal roles in the progression of head and neck squamous cell carcinomas [4], lung cancer [5], glioblastomas [6], and prostate cancer [7]. We have reported that fetuin-A added to the tumor cells plays a scaffolding role on the surfaces of a prostate cancer cell line, LNCaP, where it is concentrated on the surfaces of the cells growing as spheroids and simultaneously trap extracellular vesicles such as exosomes to effectively mediate

growth [8]. Our studies suggested that this could be a contributing pathway for growth in prostate cancer cells particularly those that do not rely entirely on androgen signaling for their growth [8]. We questioned whether the same growth strategy could be employed by breast cancer cells especially those that lack receptors for estrogens, progesterone and epidermal growth factor 2, commonly known as triple negative breast cancer (TNBC). Whereas the other subtypes of breast cancer such as luminal A and B rely on estrogens and progesterone and ... for growth, TNBC could rely on other pathways such as those propagated by fetuin-A and epidermal growth factor 1 to propel their growth [9].

On tumor cells, we have identified TLR4 as the receptor with which fetuin-A interacts to promote tumor growth [10]. Of note is the plethora of studies that have implicated TLR4 in the progression of tumor cells [11-13]. For many of these studies, it was assumed that the ligand for TLR4 activation even on tumor cells was the endotoxin, lipopolysaccharide (LPS). This signaling network was then responsible for chronic inflammatory conditions that promoted tumor progression [12, 14, 15]. Indeed LPS as ligand for TLR4 activation during tumor progression has been validated in gastric tumors [16]. However, in other tumors linking LPS in the activation of TLR4 on tumor cells was not possible suggesting there are other endogenous ligands that activate TLR4 on tumor cells [12].

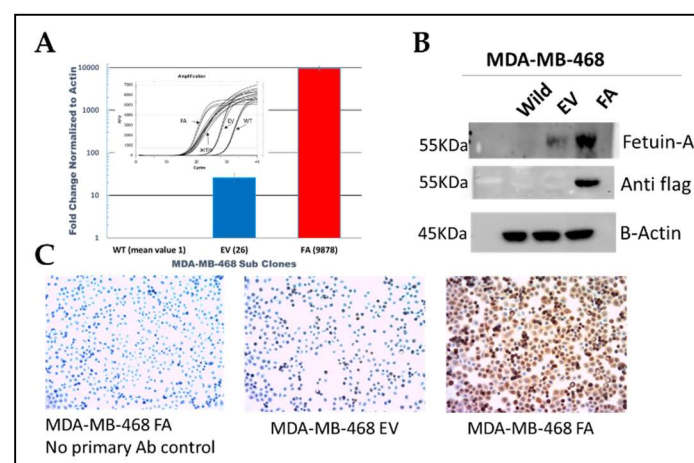
Here we have questioned the potential role of fetuin-A to modulate through TLR4, the growth and invasive capacity of a TNBC cell line, MDA-MB-468. The cell line was derived from a pleural effusion metastasis of a breast cancer patient [17]. It exhibits phenotypes that resemble basal-like breast cancer, a sub-type typically associated with TNBC. The cells are highly proliferative and invasive [18] and have been instrumental in advancing our knowledge of TNBC as well as the development of novel treatment strategies for this challenging disease [19].

The impetus for the present studies were analysis of breast cancer survival data (Cancer Genome Atlas, TCGA [20]. In these studies, basal-like as well as basal-like triple negative breast cancer cells that expressed high levels of fetuin-A had significantly poor overall survival compared to those that expressed low levels of the protein. This was not replicated in both luminal A and luminal B subtypes which are the most common breast cancer sub-types [20]. The studies underscored the significance of ectopic fetuin-A in the progression of breast cancer.

## 2. Results

### 2.1. Overexpression of fetuin-A in MDA-MB-468.

We utilized the lentiviral system to stably overexpress fetuin-A in the basal like triple negative breast cancer cell line MDA-MB-468, at the mRNA (Figure 1A, red bar) and protein (Figure 1B and 1C) levels.

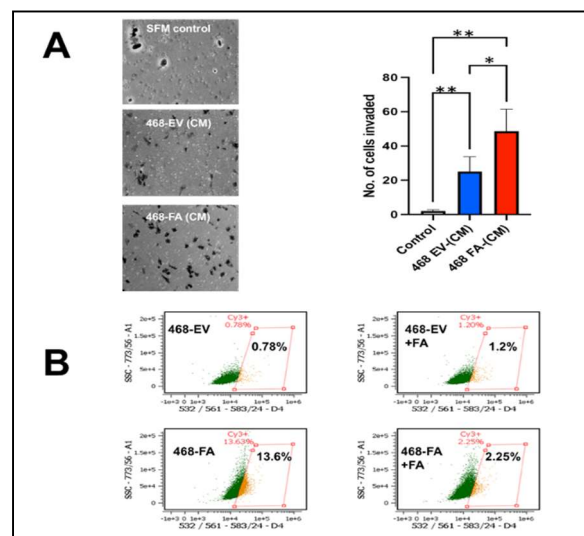


**Figure 1. Overexpression of fetuin-A (Fet-A) in MDA-MB-468 TNBC cell line.** MDA-MB-468 cells were transfected with either empty vector (EV) or fetuin-A expression vector (FA). Expression level was then determined by QT-RTPCR (panel A), Western blot (panel B) and IHC (panel C).

Both WT and wild-type transfected with empty vector (EV) expressed baseline levels of fetuin-A as shown in **Figure 1A & B**. Because previous data from our group and others had shown that ectopically produced fetuin-A promotes adhesion and motility of other tumor cell types, we next considered assaying for invasion capacity in MDA-MB-468 cells over-expressing fetuin-A.

## 2.2. Forced expression of fetuin-A in MDA-MB-468-FA increased invasion capacity of the cells relative to transfection controls and also upregulated TLR4 expression.

In 4 separate experiments, MDA-MB-468-FA cells were significantly more invasive as compared to their transfection controls, MDA-MB-468-EV (**Figure 2A**). We previously demonstrated that uptake of fetuin-A culminating in adhesion in tumor cells was mediated by TLR4 [10]. Expression of TLR4 in tumor cells however varies from one cell type to the other and appears to be under tight regulatory control. For example, report by Mehmeti et al. [21] demonstrated very low almost negligible levels of TLR4 in MDA-MB-468. We also recorded very low levels of TLR4 in these cells. We questioned whether the expression levels of TLR4 in MDA-MB-468 could be influenced by either added fetuin-A or forced expression of fetuin-A in these cells. As expected, our data show that MDA-MB-468-EV in serum free medium, expressed very low levels of TLR4 (0.78%) as measured by flow cytometry (**Figure 2B**). However, in the presence of fetuin-A (Fet-A) (2 mg/ml) in serum free medium, there was a slight increase in the surface levels of TLR4 (1.2%). The overexpression of fetuin-A mRNA in MDA-MB-468-FA in serum free medium on the other hand, dramatically increased TLR4 expression by almost 20-fold (13.4%). This high level was however attenuated in the presence of fetuin-A (2 mg/ml) to 2.25% (**Figure 2B**).

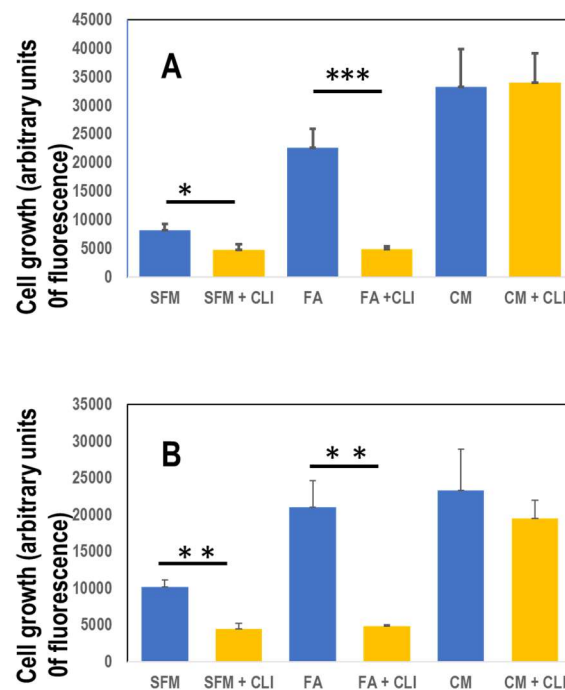


**Figure 2. Enhanced invasion capacity in fetuin-A overexpressing MDA—MB-468 cells.** In panel A, fetuin-A overexpressing cells, 468 FA invaded through a bed of Matrigel more readily compared to 468 EV controls (\*P < 0.05; N= 6). In panel B, 468-EV express the least (0.78%) TLR4 on their surfaces. This increases slightly (1.2%) upon addition of FetA. However, 468-FA express the highest level of surface TLR4 (13.6%) which decreases upon addition of external FA (2.25%).

## 2.3. Growth of MDA-MB-468-FA in 2-dimensional (2-D) as well as 3-dimensional (3-D) is influenced by fetuin-A.

In order to grow, the TNBC cells still need either the presence of fetuin-A (FA) in serum free medium or complete medium containing 10% fetal bovine serum (**Figure 3A&B**). In order for fetuin-A to mediate cellular adhesion and growth in 2-D, its uptake (which can be inhibited by a specific TLR4 inhibitor) is a pre-requisite [10]. Inhibition of fetuin-A uptake attenuated cellular adhesion and growth only in those wells containing fetuin-A in serum free medium (**Figure 3A**). Adhesion and growth of cells growing in complete medium which contains fetuin-A (~2 mg/ml) in addition to other

adhesion molecules was not affected significantly by the TLR4 inhibitor, CLI-095 (28  $\mu$ M) in 2-D cultures (**Figure 3A**). There was moderate growth of cells in wells containing fetuin-A (2 mg/ml) in serum free medium but the growth was significantly inhibited by CLI-095 (**Figure 3A**). As previously reported for prostate cancer cell line (8), the MDA-MB-468 cells grew well in 3-D culture wells containing fetuin-A (2 mg/ml) in serum free medium as well as complete medium (forming grapelike clusters of cells) but grew poorly in SFM (**Figure 3B**). Interestingly, the 3-D growth of cells in wells containing fetuin-A in SFM was significantly attenuated by CLI-095 (28  $\mu$ M) but not the 3-D growth of the cells in complete medium (**Figure 3B**).

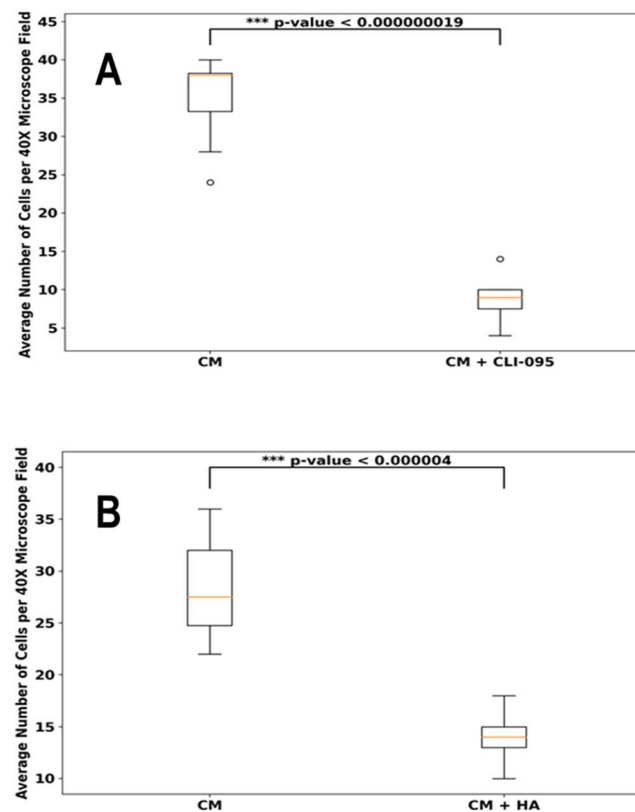


**Figure 3. Fetuin-A mediates 2-D and 3-D growth of MDA-MB-468 via TLR4 signaling.** The cells were seeded ( $2 \times 10^4$  cells/well) in attachment (2-D) (**panel A**) or ultra-low attachment (3-D) (**panel B**) 96-well plates in SFM; SFM + CLI-095 (CLI); Fetuin-A (FA); FA + CLI; complete medium (CM); and in CM + CLI. After 6 days of growth, Alamar blue was added to each well, incubated for 2 h and fluorescence measured as described. \* $P < 0.05$ ; \*\* $P < 0.001$ ; \*\*\* $P < 0.0001$ ;  $N = 4$ .

#### 2.4. Invasion of MDA-MB-468 cells is attenuated by the specific TLR4 inhibitor, CLI-095 cells and in the presence of complete medium depleted of fetuin-A.

Invasion of the TNBC cell line was attenuated in the presence of a specific inhibitor of TLR4, CLI-095 also known as resatorvid at a concentration of 28  $\mu$ M that we previously showed was effective in inhibiting motility and invasion in other tumor cells [10] (**Figure 4A**)





**Figure 4. Attenuation of fetuin-A mediated invasion by inhibition of TLR4 signaling network.** Invasion of MDA-MB-468 cells through Matrigel is attenuated in the presence of CLI-095 in the upper chamber (A) and complete medium depleted of fetuin-A (CM + HA) in the lower chambers (B) of transwell plates. .

We previously proposed that the major chemoattractant in complete medium is fetuin-A. This is because we could replace complete medium in the bottom wells of trans-well invasion and motility assays with fetuin-A at approximately 2 mg/ml and still maintain chemoattraction [22]. We therefore depleted L15 medium containing 10% fetal bovine serum and repeated trans-well assays. We demonstrated in three separate experiments that invasion was significantly attenuated in the presence of complete medium depleted of fetuin-A (CM + HA) relative to complete medium (CM) control (Figure 4B).

### 3. Discussion

The role that fetuin-A synthesized by cancer cells plays in tumor cell growth has been rather difficult to explain from a mechanistic viewpoint, given that the concentration of fetuin-A in the blood is relatively high and likely to supply all the needs of the growing tumor mass. Indeed, in the present studies, the ectopically produced fetuin-A was not sufficient to drive the growth of the cells in serum free medium as had been shown for other tumor cells such as head and neck squamous cell carcinomas [4] and glioblastoma [6]. The report by Mintz et al suggested that the ectopically produced fetuin-A drove the progression of prostate cancer culminating in the metastatic spread to the bones [7]. We hereby demonstrate that overexpression of fetuin-A by the basal like triple negative breast cancer cell line, MDA-MB-468, enhances its capacity for invasion but not growth in serum free medium.

The basal like triple negative breast cancer cell line MDA-MB-468 is an excellent prototype of the cancer it represents in that it lacks estrogen, progesterone and epidermal growth factor 2 receptors [17, 21]. Triple negative breast cancer, a type of basal breast cancer that disproportionately affect women of color has poor prognosis with fewer treatment options [23]. A recent survey of TCGA

survival data (Kaplan Meir Plotter) showed that basal type breast cancers including triple negative that express high levels of fetuin-A, have worse overall survival compared to those that express low levels of the protein [20], suggesting that fetuin-A upregulates metastatic processes in these cells. Based on these studies, we overexpressed fetuin-A in MDA-MB-468 to determine if this was sufficient to confer rapid growth, adherence and spreading in these cells, cellular phenotypes that we had previously attributed to fetuin-A [24]. Whereas overexpression of fetuin-A in MDA-MB-468 did not confer growth advantage (2-D or 3-D) to these cells, the increased invasive capacity as well as TLR4 expression in the TNBC cells resulting from overexpression of ectopic fetuin-A, suggested that other signaling pathways that promote invasion and metastasis are also upregulated.

The basal like triple negative breast cancer cell line, MDA-MB-468 express very low levels of TLR4 [21]. We believe this is the first report showing that fetuin-A overexpression can upregulate TLR4 expression in tumor cells. Even more interesting was the fact that in the transfection controls where the expression of TLR4 was almost negligible, the presence of fetuin-A at approximately 2 mg/ml was sufficient to increase surface expression of TLR4 underscoring the role of fetuin-A in the tight regulation of surface expression of TLR4 on tumor cells. A number of studies have reported the role of TLR4 in tumor motility, invasion and metastasis [12]. However, the ligand that initiates TLR4 signaling in tumor cells in particular has been a subject of intense debate. Whether it is the natural ligand for TLR4 lipopolysaccharide (LPS) or some other cellular protein such as high-mobility group box-1 protein (HMGB1)[12]. Fetuin-A was originally proposed to be a ligand for TLR4 by Pal et al [25]. The publicly available protein/protein binding data (STRING) shows that other than serum albumin, TLR4 has the strongest binding interaction with fetuin-A. The present studies suggest that cell surface TLR4 is the receptor through which fetuin-A interacts with MDA-MB-468 cells. It is not unusual for a ligand to modulate the expression of its receptor and vice-versa [26, 27].

Fetuin-A supplied to the tumor cells either from the medium or blood (*in vivo*) modulates cellular adhesion, motility, and growth in a very elaborate mechanism. To begin with, fetuin-A has to enter the cell as a prelude to cell adhesion and cell spreading. We have reported the ability of fetuin-A to prepare a unique population of extracellular vesicles/exosomes that then promote the cellular adhesion and spreading. We have demonstrated that only those exosomes that are isolated and secreted in the presence of fetuin-A have the ability to promote adhesion and cell spreading. Those prepared in the absence of fetuin-A, lack this ability [24]. In the present studies we show that fetuin-A alone is sufficient to drive the adhesion and spreading and growth of MDA-MB-468 cells both in 2-D as well as 3-D, processes that were abrogated by the specific TLR4 inhibitor. These we believe are the cells that also have the propensity for invasion.

The present data suggest that even though fetuin-A is capable of signaling growth and adhesion in MDA-MB-468 cells, other adhesion and growth signals are at play in the presence of complete medium, so the cells will use these default pathways and bypass the fetuin-A/TLR4 signaling network when it is specifically targeted by CLI-095. The fact that CLI-095 only reduced the 2-D and 3-D growth in wells containing fetuin-A but not complete medium shows that the action of CLI-095 was specific for the fetuin-A mediated adhesion and growth.

Lastly the data suggest that in addition to other known chemo-attractants such as EGF, fetuin-A is the dominant chemo-attractant in complete medium (CM) that is usually added to the lower chambers in trans-well invasion and motility assays [22]. The depletion of fetuin-A in complete medium significantly attenuated the ability of CM to attract the cells as they invaded through a bed of Matrigel. Likewise, in blood vessels next to a growing tumor mass and where fetuin-A is maintained at a concentration of approximately 0.3 mg/ml, it is likely to play a role in the initial attraction of invading tumor cells during the process of intravasation.

In summary we have demonstrated that elevated expression of fetuin-A in a triple negative breast cancer cell line promotes invasion capacity of the cells and upregulates TLR4 expression in these cells. This could explain why TNBC which express high levels of ectopic fetuin-A have poorer prognosis. The data also suggest that fetuin-A in the medium can modulate the surface expression of TLR4 on tumor cells (either upregulation or downregulation) to promote adhesion, invasion and

growth. More importantly our data show that fetuin-A is the putative ligand that signals through TLR4 to promote the growth and invasive capacity of MDA-MB-468.

## 4. Materials and Methods

### 4.1. Cells

MDA-MB-468 was purchased from ATCC, Manassas, Va. The cells were cultured in L15 (Leibovitz's) medium supplemented with 10% fetal bovine serum, 1% NaHCO<sub>3</sub> and 1x antibiotic/antimycotic (Life Technologies, Grand Island, NY, U.S.A). The cells were maintained in humidified CO<sub>2</sub> incubator at 37°C. The serum-free medium (SFM) used in the studies contained 1% (w/v) bovine serum albumin.

### 4.2. Over expression of Fetuin-A (FA) by Lentiviral method

In order to confer stable overexpression, a lentiviral vector was used for gene delivery. In brief, HEK 293FT cells with a low passage number were seeded in 10-cm plates at a density of  $4.0 \times 10^6$  cells and allowed to achieve 80% confluency overnight. On the next day, 2 h before transduction, the medium was replaced with 10 ml fresh, pre-warmed DMEM containing 10% FBS. A total of 20 µg of DNA plasmids (10 µg of flag-tagged Fetuin-A-expression vector and 10 µg of psPAX2 packaging and pMD2. G envelope plasmid DNA) were combined in 1 ml serum-free, antibiotic-free DMEM in a 10-cm plate. The control plates comprised GFP containing vectors. Then, the cells were transduced using OriGene TurboFectin 8.0 Transfection Reagent (Cat # TF81005). The medium was removed around 14–16 h after transduction, and 8 ml/dish fresh preheated DMEM containing 10% FBS was added. After 12 h, the supernatant was collected, and this procedure was continued for 72 h at 8–12 h intervals. When all the supernatants were collected, centrifugation was performed at 1,500 rpm for 5 min; the supernatant was filtered through a 0.45-µm filter. The viral titer was detected using qPCR Lentiviral titer kit (LV900) from abm. The filtered supernatant was aliquoted and stored at -80°C.

MDA-MB-468 cells were plated in 6 well plates at a density of  $0.5 \times 10^5$  cells per well 24 h prior to viral infection and grown at 37°C with 5% CO<sub>2</sub> overnight. On the day of infection, lentiviral particles were added to cells with the mixture of complete media and 8 µg/mL polybrene. The cells were incubated with virus for 8 hours, and then medium containing FBS was added. Second round of infection was done for next 8 hours and media was changed. Forty-eight hours later, the infected cells were subjected to selection with 2 mg/mL puromycin for 1 week.

### 4.3. Reverse Transcriptase and Real-Time PCR.

MDA-MB-468 cells were grown in 10 cm dishes and cultured in complete medium containing 10% FBS. Total RNA was isolated from the cells using RNeasy MiniKit from QIAGEN (Valencia, CA). For the single-strand cDNA synthesis, 1 µg of total RNA was reverse-transcribed using High-Capacity cDNA Reverse Transcription Kit according to manufacturer's protocol from ThermoFisher Scientific (Catalog number: 4368814). Real time PCR was performed using Fetuin-A Taqman gene specific assays (Catalog number: Hs00155659\_m1) and TaqMan™ Fast Advanced Master Mix (Catalog number: 4444556). Beta Actin was used as endogenous controls. The expression levels of the genes were normalized against those of the endogenous control using the 2-ΔΔCt method, and p < 0.05 considered statistically significant (\*p < 0.05; \*\*p < 0.01; \*\*\*p < 0.001).

### 4.4. Western Blot Assays:

Western blotting was done following standard protocols. Proteins were separated on SDS-polyacrylamide gels and transferred onto nitrocellulose membranes. Membranes were blocked with 5% non-fat milk and incubated with the primary antibodies AHSG (R&D, Cat # MAB 1184), anti-flag (Sigma, Cat # F3165), Actin (Cell Signaling, Cat # 4967) overnight at 4°C. Membranes were washed and incubated with horseradish peroxidase-conjugated secondary antibodies at room temperature.



The antigen-antibody complexes were detected using enhanced chemiluminescence detection reagents. Protein bands were imaged using a ChemiDoc gel imaging system.

#### 4.5. Immuno-histochemistry Assays (IHC).

Control and over-expressed cells were seeded at an appropriate density and allowed to attach to the slide (30,000 cells/chamber in an 8-chamber slide). Cells were plated one day before staining to achieve 60-80% confluency. The following day cells were fixed with 3% formaldehyde for 15 min at room temperature and gently washed in PBS 3 times (5 min/wash). They were stained, as reported previously (Nangami et al., 2016).

#### 4.6. Invasion Assays:

The assay was done essentially as previously described [10]. The same number of cells (125,000 cells/chamber) in SFM were added to the upper chambers (500  $\mu$ l) of Boyden Chamber trans-well. The bottom chambers contained 500  $\mu$ l of L15 medium containing 10% fetal bovine serum (FBS)-complete medium (CM). As controls, some of the bottom wells contained serum free medium containing 1% (w/v) BSA. After 24 h, the invading cells migrated to the underside of the polycarbonate filters with Matrigel coating. After removing the cells remaining in the upper chambers with cotton swabs, the invading cells were fixed in 4% formalin, washed in PBS and then stained with crystal violet as described [10], and viewed under a microscope. The number of cells in 20X field of view were counted and expressed as mean  $\pm$  S.D. (N = 6)

#### 4.7. Regulation of TLR4 surface expression by Fetuin-A.

MDA-MB-468 cells are reported to express very low levels of TLR4, almost negligible [21]. We questioned whether fetuin-A could modulate cell surface expression of TLR4 in these cells, either upregulation or downregulation. The cells (MDA-MB-468, MDA-MB-468-EV and MDA-MB-468FA) were incubated at 37°C in humidified CO<sub>2</sub> incubator until approximately 80% confluent. They were then washed in warm PBS and detached with 2 mM EDTA in PBS. The cells were pelleted and re-suspended as single cells in cold FACS buffer (PBS containing 1% BSA) in Eppendorf tubes (1  $\times$  10<sup>6</sup> cells per tube). To determine the influence of added fetuin-A on cell surface TLR4 expression, the cells were suspended in FACS buffer containing fetuin-A (2 mg/ml). The tubes were incubated without (unlabeled controls) or with Cy3-labeled anti-human CD284 (TLR4 antibody; Bio-Legend, San Diego, CA USA; 10  $\mu$ l per tube) for 30 min at 4°C with end on end rotation. The cells were then washed 3x with cold FACS buffer and analyzed on a Cell-Stream flow cytometer (Luminex) using channel D4 with excitation wavelength 532 nm, and emission 583/24 nm.

#### 4.8. Invasion of MDA-MB-468 is modulated by the Fetuin-A-TLR4 signaling network.

Even though advances in cell and molecular biology approaches has shed light on the mechanisms of invasion and motility, there are still a number of gaps in our knowledge of the process. In most motility and invasion assays based on Boyden trans-well set up, complete medium containing 10% v/v fetal bovine serum is usually included in the bottom wells or chambers. It is then assumed that the growth factors in the fetal bovine serum such as EGF would be the chemo-attractants. We have proposed that uptake of fetuin-A (inhibited by CLI-095, a specific inhibitor of TLR4 signaling) along its concentration gradient from the bottom wells initiates the process of invasion [10]. We therefore repeated the invasion assays using parental MDA-MB-468 to assess the ability of TLR4 inhibitor to influence invasion of the cells through a bed of Matrigel. Cells (125,000 cells/chamber) in SFM without (control chambers) or containing CLI-095 dissolved in DMSO at a final concentration of (10  $\mu$ g/mL; 28  $\mu$ M), were added to the upper chambers (coated with Matrigel) of Boyden Chamber trans-well set-up. The control chambers contained the same amount of DMSO as experimental chambers. The bottom chambers contained 500  $\mu$ l of complete medium. In order to demonstrate that it is fetuin-A in complete medium that initiated the invasion process, we repeated the experiment with complete medium depleted of fetuin-A. The depletion was done by incubating

the medium with hydroxyapatite overnight at 4°C followed by centrifugation (5,000 × g) to remove the hydroxyapatite nanoparticles essentially as described [8].

#### 4.9. 2-Dimensional (2-D) and 3-Dimensional (3-D) growth of MDA-MB-468 cells in the presence of medium containing none (SFM) or either fetuin-A (2 mg/ml) or fetal bovine serum (CM) (10% v/v).

The MDA-MB-468 parental cell lines were plated quadruplicates either in treated 96-well culture plates (2D) or ultra-low attachment plates (3-D) at  $2 \times 10^4$  cells/well in 200 µl of medium. Cell growth was evaluated in; 1) SFM, 2) SFM + CLI-095, 3) Fetuin-A (FA), 4) FA + CLI-095, 5) CM and 6) CM + CLI-095. The final CLI-095 concentration used in the studies (10 µg/mL; ~28 µM) effectively inhibited fetuin-A uptake in these cells (data not shown). After 5 days of growth at 37°C in a humidified CO<sub>2</sub> incubator, Alamar Blue (20 µl) was added to each well and incubated further at 37°C for 2 h. The plates were then read using a BioTek Neo2 multiplate reader (Agilent) with excitation wavelength at 560 nm and emission wavelength at 590 nm, giving as an accurate estimation of cell numbers in each well expressed as arbitrary units of fluorescence.

#### 4.10. Statistics

Each Figure is representative of at least 3 separate experiments. Graphs and statistical analyses were generated using either Microsoft Excel or Graph Pad Prism packages. P values less than 0.05 were considered significant.

**Author Contributions:** Conceptualization, D.K. and J.O.; methodology, D.K., A.S., O.K., M.G.I., and J.O.; software, P.O.; validation, J.O., D.K., and P.O.; formal analysis, D.K.; investigation, J.O.; resources, J.O.; data curation, D.K., and P.O.; writing—original draft preparation, D.K. and J.O.; writing—review and editing, J.O.; visualization, J.O.; supervision, J.O.; project administration, J.O.; funding acquisition, J.O., and A.S. All authors have read and agreed to the published version of the manuscript.”.

**Funding:** This research was funded by NIH/NIMHD-U54 MD007586-03 (MRRCC)(J.O.); NIH-NCI-U54 CA 163069 (J.O.) and NIGMS-9SC1 GM139814-05 (AS).

**Data Availability Statement:** Data is contained within the article.

**Acknowledgments:** We thank Dr. Ajay Kumar Singh for data analysis.

**Conflicts of Interest:** The authors declare no conflict of interest and the funders had no role in the design of the study; in the collection, analyses, or interpretation of data; in the writing of the manuscript; or in the decision to publish the results.

## References

- Guillory, B., Sakwe, A. M., Saria, M., Thompson, P., Adhiambo, C., Koumangoye, R., Ballard, B., Binhazim, A., Cone, C., Jahanen-Dechent, W. & Ochieng, J. (2010) Lack of fetuin-A (alpha2-HS-glycoprotein) reduces mammary tumor incidence and prolongs tumor latency via the transforming growth factor-beta signaling pathway in a mouse model of breast cancer, *Am J Pathol.* **177**, 2635-44.
- Mori, K., Emoto, M. & Inaba, M. (2011) Fetuin-A: a multifunctional protein, *Recent patents on endocrine, metabolic & immune drug discovery.* **5**, 124-46.
- Kovarova, M., Kalbacher, H., Peter, A., Haring, H. U., Didangelos, T., Stefan, N., Birkenfeld, A., Schleicher, E. & Kantartzis, K. (2021) Detection and Characterization of Phosphorylation, Glycosylation, and Fatty Acid Bound to Fetuin A in Human Blood, *Journal of clinical medicine.* **10**.
- Thompson, P. D., Sakwe, A., Koumangoye, R., Yarbrough, W. G., Ochieng, J. & Marshall, D. R. (2014) Alpha-2 Heremans Schmid Glycoprotein (AHSG) modulates signaling pathways in head and neck squamous cell carcinoma cell line SQ20B, *Exp Cell Res.* **321**, 123-32.
- Yu, C. J., Wang, C. L., Wang, C. I., Chen, C. D., Dan, Y. M., Wu, C. C., Wu, Y. C., Lee, I. N., Tsai, Y. H., Chang, Y. S. & Yu, J. S. (2011) Comprehensive proteome analysis of malignant pleural effusion for lung cancer biomarker discovery by using multidimensional protein identification technology, *Journal of proteome research.* **10**, 4671-82.
- Nangami, G. N., Sakwe, A. M., Izban, M. G., Rana, T., Lammers, P. E., Thomas, P., Chen, Z. & Ochieng, J. (2016) Fetuin-A (alpha 2HS glycoprotein) modulates growth, motility, invasion, and senescence in high-grade astrocytomas, *Cancer medicine.* **5**, 3532-3543.

7. Mintz, P. J., Rietz, A. C., Cardo-Vila, M., Ozawa, M. G., Dondossola, E., Do, K. A., Kim, J., Troncoso, P., Logothetis, C. J., Sidman, R. L., Pasqualini, R. & Arap, W. (2015) Discovery and horizontal follow-up of an autoantibody signature in human prostate cancer, *Proc Natl Acad Sci U S A*. **112**, 2515-20.
8. Ochieng, J., Korolkova, O. Y., Li, G., Jin, R., Chen, Z., Matusik, R. J., Adunyah, S., Sakwe, A. M. & Ogunkua, O. (2022) Fetuin-A Promotes 3-Dimensional Growth in LNCaP Prostate Cancer Cells by Sequestering Extracellular Vesicles to Their Surfaces to Act as Signaling Platforms, *International journal of molecular sciences*. **23**.
9. Ferrari, P., Scatena, C., Ghilli, M., Bargagna, I., Lorenzini, G. & Nicolini, A. (2022) Molecular Mechanisms, Biomarkers and Emerging Therapies for Chemotherapy Resistant TNBC, *International journal of molecular sciences*. **23**.
10. Thomas, P. L., Nangami, G., Rana, T., Evans, A., Williams, S. D., Crowell, D., Shanker, A., Sakwe, A. M. & Ochieng, J. (2020) The rapid endocytic uptake of fetuin-A by adherent tumor cells is mediated by Toll-like receptor 4 (TLR4), *FEBS open bio*. **10**, 2722-2732.
11. Thuringer, D., Hammann, A., Benikhlef, N., Fourmaux, E., Bouchot, A., Wettstein, G., Solary, E. & Garrido, C. (2011) Transactivation of the epidermal growth factor receptor by heat shock protein 90 via Toll-like receptor 4 contributes to the migration of glioblastoma cells, *J Biol Chem*. **286**, 3418-28.
12. Oblak, A. & Jerala, R. (2011) Toll-like receptor 4 activation in cancer progression and therapy, *Clinical & developmental immunology*. **2011**, 609579.
13. Szajnik, M., Szczepanski, M. J., Czystowska, M., Elishaev, E., Mandapathil, M., Nowak-Markwitz, E., Spaczynski, M. & Whiteside, T. L. (2009) TLR4 signaling induced by lipopolysaccharide or paclitaxel regulates tumor survival and chemoresistance in ovarian cancer, *Oncogene*. **28**, 4353-63.
14. Yokota, S., Okabayashi, T., Rehli, M., Fujii, N. & Amano, K. (2010) Helicobacter pylori lipopolysaccharides upregulate toll-like receptor 4 expression and proliferation of gastric epithelial cells via the MEK1/2-ERK1/2 mitogen-activated protein kinase pathway, *Infection and immunity*. **78**, 468-76.
15. Su, B., Ceponis, P. J., Lebel, S., Huynh, H. & Sherman, P. M. (2003) Helicobacter pylori activates Toll-like receptor 4 expression in gastrointestinal epithelial cells, *Infection and immunity*. **71**, 3496-502.
16. Ito, N., Tsujimoto, H., Ueno, H., Xie, Q. & Shinomiya, N. (2020) Helicobacter pylori-Mediated Immunity and Signaling Transduction in Gastric Cancer, *Journal of clinical medicine*. **9**.
17. Zhang, R. D., Fidler, I. J. & Price, J. E. (1991) Relative malignant potential of human breast carcinoma cell lines established from pleural effusions and a brain metastasis, *Invasion Metastasis*. **11**, 204-15.
18. Hugo, H. J., Gunasinghe, N., Hollier, B. G., Tanaka, T., Blick, T., Toh, A., Hill, P., Gilles, C., Waltham, M. & Thompson, E. W. (2017) Epithelial requirement for in vitro proliferation and xenograft growth and metastasis of MDA-MB-468 human breast cancer cells: oncogenic rather than tumor-suppressive role of E-cadherin, *Breast cancer research : BCR*. **19**, 86.
19. Cingir Koker, S., Yalcin, B. & Dogan Turacli, I. (2022) Metformin resistant MDA-MB-468 cells exhibit EMT-like phenotype and increased migration capacity, *Molecular biology reports*. **49**, 5973-5984.
20. Gyorffy, B. (2021) Survival analysis across the entire transcriptome identifies biomarkers with the highest prognostic power in breast cancer, *Computational and structural biotechnology journal*. **19**, 4101-4109.
21. Mehmeti, M., Allaoui, R., Bergenfelz, C., Saal, L. H., Ethier, S. P., Johansson, M. E., Jirstrom, K. & Leandersson, K. (2015) Expression of functional toll like receptor 4 in estrogen receptor/progesterone receptor-negative breast cancer, *Breast cancer research : BCR*. **17**, 130.
22. Nangami, G. N., Watson, K., Parker-Johnson, K., Okereke, K. O., Sakwe, A., Thompson, P., Frimpong, N. & Ochieng, J. (2013) Fetuin-A (alpha2HS-glycoprotein) is a serum chemo-attractant that also promotes invasion of tumor cells through Matrigel, *Biochem Biophys Res Commun*. **438**, 660-5.
23. Ogden, A., Garlapati, C., Li, X. B., Turaga, R. C., Oprea-Ilie, G., Wright, N., Bhattarai, S., Mittal, K., Wetherilt, C. S., Krishnamurti, U., Reid, M. D., Jones, M., Gupta, M., Osan, R., Pattni, S., Riaz, A., Klimov, S., Rao, A., Cantuaria, G., Rida, P. C. & Aneja, R. (2017) Multi-institutional study of nuclear KIFC1 as a biomarker of poor prognosis in African American women with triple-negative breast cancer, *Scientific reports*. **7**, 42289.
24. Watson, K., Koumangoye, R., Thompson, P., Sakwe, A. M., Patel, T., Pratap, S. & Ochieng, J. (2012) Fetuin-A triggers the secretion of a novel set of exosomes in detached tumor cells that mediate their adhesion and spreading, *FEBS letters*. **586**, 3458-63.
25. Pal, D., Dasgupta, S., Kundu, R., Maitra, S., Das, G., Mukhopadhyay, S., Ray, S., Majumdar, S. S. & Bhattacharya, S. (2012) Fetuin-A acts as an endogenous ligand of TLR4 to promote lipid-induced insulin resistance, *Nature medicine*. **18**, 1279-85.
26. Hadcock, J. R. & Malbon, C. C. (1991) Regulation of receptor expression by agonists: transcriptional and post-transcriptional controls, *Trends in neurosciences*. **14**, 242-7.
27. Infantino, S., Moepps, B. & Thelen, M. (2006) Expression and regulation of the orphan receptor RDC1 and its putative ligand in human dendritic and B cells, *J Immunol*. **176**, 2197-207.