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Article

Determination of the Copy Number of Porcine Endogenous Retroviruses (PERV) in Auckland Island Pigs Repeatedly Used for Clinical Xenotransplantation and Elimination of PERV-C

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Abstract: Auckland Island pigs represent an inbred population of feral pigs isolated on the subantarctic island for over 100 years. The animals have been maintained under pathogen-free conditions in New Zealand, they are well characterized virologically and have been used as donor sources in first clinical trials of porcine neonatal islet cell transplantation for the treatment of human diabetes patients. The animals do not carry any of the xenotransplantation-relevant viruses and in the first clinical trials no porcine viruses including porcine endogenous retroviruses (PERVs) have been transmitted to the human recipients. PERVs pose a special risk in xenotransplantation since they are part of the pig genome. When the copy number of PERVs in these animals was analyzed using droplet digital PCR, a copy number typical for Western pigs was found, confirming previous phylogenetic analyses of microsatellites and mitochondrial analyses showing a closer relationship to European pigs than to Chinese pigs. When kidney cells from very young piglets were analyzed, only around 20 PERVpol copies were detected. Using these cells as nuclear donors in a somatic cell nuclear transfer (SCNT), animals were born showing PERVpol copy numbers between 35 and 56. This data indicates that Auckland Island pigs have a lower copy number in comparison with other Western pig breeds and that the copy number is higher in adult animals compared with cells from young piglets. Most importantly, PERV-C free animals were selected and the absence of additional eight porcine viruses was demonstrated.

Keywords: porcine endogenous retroviruses; Auckland Island pigs; islet cell xenotransplantation

1. Introduction

The Auckland Islands are a group of subantarctic islands which lie some 560 kilometers south of New Zealand. Pigs were originally introduced onto these islands in 1807 by Captain Abraham Bristow and in 1840 James Ross brought more pigs onto the island [1, 2]. From the time of the last release the pig population on Auckland Island remained isolated for the next hundred years. Pigs were removed from Auckland Island in 1999 to help restore the environment to its former state and were brought to the mainland of New Zealand for conservation. 17 animals were housed, initially in quarantine, in a special, purpose-built facility in Invercargill, where they have been successfully bred [3]. The pigs are black or white to brown with black markings, have a long snout, high hair and bristle coverage for the cold conditions [1]. When the mitochondrial D-loop DNA sequences of the Auckland Island pigs were compared with sequences from domestic and wild boar, the sequence from the Auckland Island pigs clustered with domestic European breeds and they represent a single breeding population [1, 2]. Phylogenetic analyses of microsatellites also showed that they are more closely related to European pigs than Chinese pigs, which is consistent with the mitochondrial analyses [3].

These pigs have been monitored for 8 years and were found negative for all tested porcine infectious agents [4, 5]. Among the tested viruses potentially posing a risk for xenotransplantation were the porcine endogenous retro-virus (PERV), porcine cytomegalovirus (PCMV), a porcine roseolovirus (PCMV/PRV), porcine lymphotropic herpesvirus (PLHV), and porcine circovirus (PCV), for testing diagnostic PCR and serology methods were applied. In comparison with other pig breeds, the Auckland Island pigs are the “cleanest” pigs concerning virological safety. This was the reason to use pig islets from Auckland Island pigs for the treatment of humans with diabetes. In a prospective preclinical trial transplanting islet cells from Auckland Island pigs into non-human primates, no transmission of pig viruses was observed [6]. No transmission of pig viruses including PERVs was also observed in two clinical trials performed in New Zealand [7] and Argentina [8].

Here we analyze the number of PERV proviruses in the genome of adult Auckland Island pigs, in cells from young piglets as well as in animals obtained by somatic cell nuclear transfer (SCNT) using these piglet cells. Furthermore, we screened these animals for an additional 8 xenotransplantation-relevant viruses, among them PCMV/PRV. PCMV/PRV is of special interest, it is known to significantly reduce the survival time of non-human primates after transplantation of PCMV/PRV-positive kidneys or hearts [9-12]. PCMV/PRV was also transmitted in the first clinical trial transplanting a pig heart into a patient in Baltimore and contributed to the death of the patient [13, 14]. We confirmed the absence of PCMV/PRV and other potentially zoonotic viruses in the Auckland Island pigs.

It is known that PERV-C can recombine with PERV-A in living pigs and the resulting PERV-A/C recombinant viruses are characterized by higher virus titers compared with the paternal PERV-A [15-17]. Release of human tropic PERV-A/C from mitogen-triggered peripheral mononuclear cells (PBMCs) was observed in the case of different minipigs as well as ill pigs [18-22]. Since PERV-C is in contrast to PERV-A and PERV-B not present in all pigs, we screened the Auckland Island pigs for PERV-C and selected PERV-C animals for further breeding. The absence of PERV-C will prevent a recombination with PERV-A.

2. Materials and Methods

2.1. Animals and Cells

Auckland Island pigs were bred as described by the NZ Xeno Ltd, Auckland, New Zealand [4, 5]. DNA and blood from two groups of animals were sent to Berlin and the PERV copy numbers were analyzed. Later, kidney cells from four animals (two male, two female) at an age of about 4 weeks were sent to Germany and the PERV copy numbers were analyzed. Nuclei from these cells were used for an in vitro SCNT into matured oocytes which were collected from abattoir derived ovaries from German Landrace/Piétrain hybrid gilts. The SCNT was performed at the Chair for Molecular Animal Breeding and Biotechnology, and Center for Innovative Medical Models (CiMM) of the Ludwig Maximilian University Munich. DNA was obtained from animals of the first and second generation and analyzed for the PERV copy number. As an internal standard the porcine embryonic kidney cell line PK15 (ACC 640, from the Leibniz Institute DSMZ German Collection of Microorganisms and Cell lines, Braunschweig, Germany) was used.

2.2. DNA Isolation

DNA was extracted at XenoNZ and CiMM, DNA was isolated from blood using two DNA extraction methods: DNeasy Blood and Tissue kit (Qiagen GmbH, Hilden, Germany) or phenol chloroform extraction using TRIzol Reagent (Invitrogen, Carlsbad, CA, USA) following the manufacturer's instructions. DNA was quantified using a Qubit 3.0 Fluorometer (Thermo Fischer), and the 260 nm/280 nm ratio was determined using a NanoDrop ND-1000 (Thermo Fisher Scientific Inc., Worcester, MA, USA).

2.3. Droplet Digital PCR

Droplet digital PCR (ddPCR) was performed according to the manufacturer's instructions (Bio-Rad, Hercules, CA, USA, [\[http://www.bio-rad.com/de-de/applications-technologies/droplet-digital-pcr-ddpcr-technology?ID=MDV31M4VY\]](http://www.bio-rad.com/de-de/applications-technologies/droplet-digital-pcr-ddpcr-technology?ID=MDV31M4VY)) using a QX200 droplet generator and a QX100 droplet reader (Bio-Rad). Purified genomic DNA (100 ng genomic DNA) was digested with MseI (New England Biolabs, Ipswich, MA, USA) (20U) at 37°C for 1 hour, and the restriction enzyme was heat-inactivated. The DNA digest was diluted to 5-10 ng/μL for the ddPCR. The ddPCR mix consisted of 10 μL 2X ddPCR Master mix, 1.8 μL of each 10 μmol/L target primers (Table 1), 0.5 μL of each 10 μmol/L probes (FAM/HEX) (Table 1), 2.5-10 ng digested DNA, and water to a total volume of 20 μL. The following cycling conditions were used: 10-minute initial enzyme activation at 95°C, 30-second denaturation at 94°C, 1-min annealing.

Table 1. Primers and probes used for the estimation of the PERV copy number and the detection of PERV-C.

Name	Sequence	Location (nucleotide number)	Accession number	Reference
PERV pol1-forward	CGACTGCCCAAGGGTCAA	3568-3587	HM159246	Yang et al., 2015 [23]
PERV pol2-reverse	TCTCTCCTGCAAATCTGGGCC	3803-3783		
PERV pol probe	/56FAM/CACGTACTGGAGGAGGGTCACCTG	3678-3655		
Pig actin forward	TAACCGATCCTTCAGCATT			Krüger et al., 2020 [24]
Pig actin reverse	TGGTTCAAAGCTTGCATCATA			
Pig actin probe	/5HEX/CGTGGGGATGCTCCTGAGAAAG			
Pig GAPDH forward	TTCACTCCGACCTTCACCAT	3951-3970	NC_010447.5 (396823)	Krüger et al., 2020 [24]
Pig GAPDH reverse	CCCGCATCTAATGTTCTCTTC	4022-4001		
Pig GAPDH probe	/5HEX/CAGCCGCGTCCCTGAGACAC	3991-3972		
PERV-C forward	CTGACCTGGATTAGAACTGG	6606-6625	AM229312	Takeuchi et al. [25, 26]
PERV-C reverse	ATGTTAGAGGATGGTCTGG	6867-6886		



2.3. Spleen and Liver Tissues

Frozen samples of spleen and liver tissues from dead born (day 0), 9-30 days old and older than 12 weeks piglets from the CiMM were used. DNA was isolated as described above and the PERV copy number was measured in both organs. and extension at 59°C (45 cycles), and final 10-min enzyme deactivation at 98°C by using a Master cycler ProS (Eppendorf, Hamburg, Germany). The temperature ramp rate was 2°C per second. To run a gradient, temperatures between 57 and 62°C were used.

2.4. Tests for PERV-C

DNA from the Auckland Island pigs were used to analyze the presence of PERV-C. Primers and conditions used for were described [25, 26] (Table 2).

2.5. Tests for Additional Porcine Viruses

DNA, RNA and blood from the Auckland Island pigs were used to analyze the presence of different DNA viruses and the RNA virus HEV. DNA was isolated as described above, and RNA was isolated using the RNeasy Mini kit (Qiagen, Hilden, Germany). Primers and condition used for the PCR or real-time PCR assays have been described for PCMV/PRV [27, 28], PCV2 [29], PCV3 [30], PLHV-1, PLHV-2, and PLHV-3 [31] as well as for HEV [31]. The Western blot and ELISA to detect antibodies against HEV using recombinant proteins were described as well [31].

3. Results

3.1. PERV Copy Number in the Genome of Adult Auckland Island Pigs

To determine the number of integrated PERVs, a ddPCR was developed which uses primers binding to a highly conserved region of the polymerase gene (pol) of PERV [24, 32]. DNA from 30 Auckland Island pigs, bred in New Zealand, were analyzed using the ddPCR and a PERVpol copy number of around 60 was measured (Figure 1). This copy number was similar compared with the copy number of Göttingen minipigs and lower compared with Aachen minipigs, investigated in parallel. DNA from kidneys of Göttingen minipigs revealed between 45 and 78 PERVpol copies using GAPDH as a reference and in a second study analyzing 16 Göttingen minipigs a median number of 64 (45-93) was found [32]. When DNA samples from Aachen minipigs were analyzed, a median number of 69 (34-97) was determined [32].

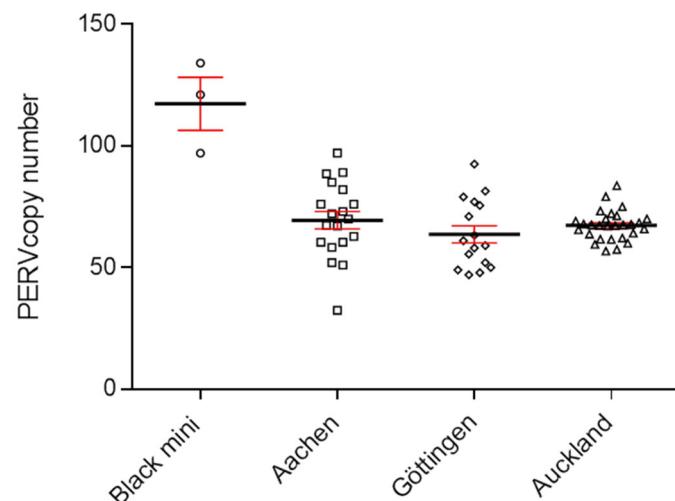


Figure 1. Determination of the PERVpol copy number of 30 Auckland Island pigs bred in New Zealand by ddPCR. For comparison the PERVpol copy number of Göttingen minipigs and Aachen minipigs is shown [32]. Porcine GAPDH was chosen as reference gene.

At a later point in time, DNA from another 14 Auckland Island pigs, also bred in New Zealand were analyzed and copy numbers between 45 and 63 were estimated (Figure 2). This experiment confirmed the previous results shown in Figure 1, but showed a broader range of copy numbers.

3.2. Selection of PERV-C Negative Animals

When the 14 animals tested for the PERV copy number above (Figure 2) and another pig (494) were tested using a PCR, eight out of 15 animals (53%) were PERV-C negative (Table 2). These investigations had the goal to select four PERV-C-free animals from which kidney cells were obtained, which could later be used for the SCNT.

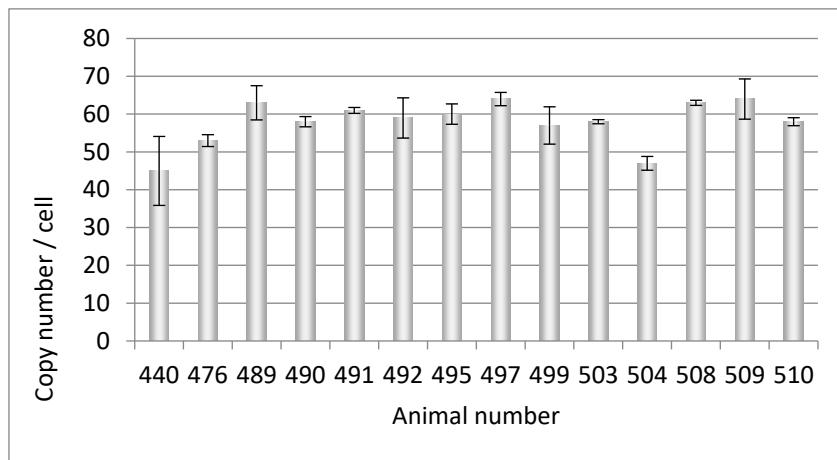


Figure 2. Determination of the PERVpol copy number of 14 further Auckland Island pigs bred in New Zealand by ddPCR at a later time point.

Table 2. Testing of Auckland Island pigs (animals 440-510) and embryonic cells (w, female, m, male) for different viruses using PCR methods and a Western blot assay and ELISA (HEV). PERV, median copy number (Figure 1); n.t., not tested, n.a., not applicable.

49	62	+	-	-	-	n.t.	-	-	-
2									
49	73	+	-	-	-	-	-	-	-
4									
49	66	-	-	-	-	-	-	-	-
5									
49	69	-	-	-	-	-	-	-	-
7									
49	62	-	-	-	-	-	-	-	-
9									
50	62	-	-	-	-	-	-	-	-
3									
50	57	-	-	-	-	n.t.	-	-	-
4									
50	64	+	-	-	-	n.t.	-	-	-
8									
50	67	+	-	-	-	n.t.	-	-	-
9									
51	60	+	-	-	-	n.t.	-	-	-
0									
f1	21	-	-	n.t.	n.a.	n.a.	n.t.	-	n.t.
f2	20	-	-	n.t.	n.a.	n.a.	n.t.	-	n.t.
m	20	-	-	n.t.	n.a.	n.a.	n.t.	-	n.t.
1									
m	22	-	-	n.t.	n.a.	n.a.	n.t.	-	n.t.
2									

3.3. PERV Copy Number in the Genome of Cell Lines from Very Young Piglets

DNA from four kidney cell lines from very young piglets were analyzed which had been selected as PERV-C negative. A very low copy number of 20-22 copies was found (Figure 3).

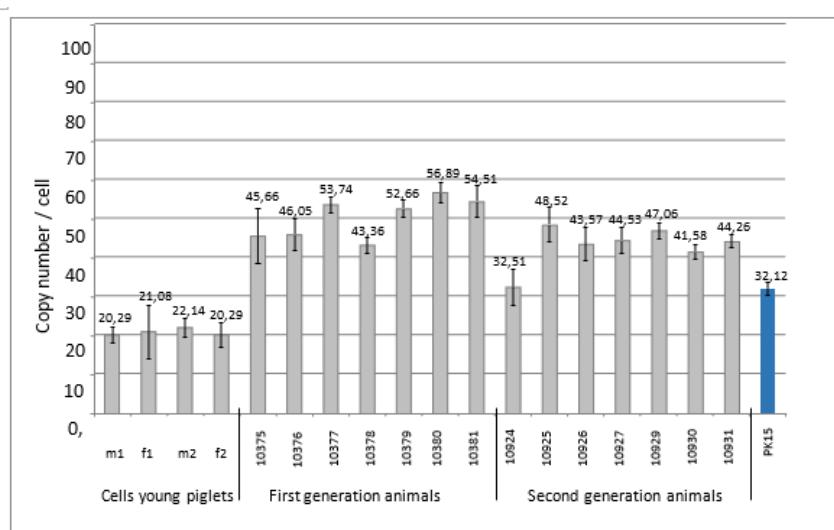


Figure 3. Determination of the PERVpol copy number of four kidney cell lines from very young Auckland Island pigs, and from the first and second generation of pigs obtained by nuclear transfer from the embryonic kidney cells. The copy number of PK15 cells was determined as control. m, male, f, female.

3.4. PERV Copy Number in the Genome of Auckland Island Pigs Obtained by SCNT

The nuclei of these PERV-C-negative embryonic kidney cells were used for an *in vitro* SCNT into matured oocytes collected from abattoir derived ovaries from German Landrace/Piétrain hybrid gilts. The animals of the first and second generation were analyzed for the PERVpol copy number. The copy numbers of the individual pigs ranged from 32 to 57, but were lower compared with the numbers of the Auckland Island pigs analyzed in the past (Figure 1, Figure 2), and lower than other pig breeds such as the Göttingen minipigs, and the Aachen minipigs [32]. The copy number found in PK15 cells correlated well with the copy number detected previously. Since different copy numbers of integrated PERV were found in different PK15 cell lines [32], here the PK15 cells ordered from the Leibniz Institute DSMZ German Collection of Microorganisms and Cell lines, Braunschweig, Germany (ACC 640), were used and the measured copy number was 32.12 +/- 1.7, which was in agreement with our previous findings [24, 32].

3.5. Increase of the PERV Copy Number with Age

In order to obtain additional evidence for the increase of the PERV copy number with age, the copy number was determined in two different organs of dead born (day 0), 9-30 days old and older than 12 weeks piglets from the CiMM. The copy number was determined in the spleen and liver and the difference between the copy number was depicted dependend on the age of the animals (Figure 4). It was obvious that there were differences in the copy number between spleen and liver, indicating that in addition to the copy number in the germ line de novo integrations took place in one or the other organ. The de novo integrations was not tissue-specific, in some animals the higher copy number was in the spleen, in others in the liver. The difference was larger in older piglets, indicating that the copy number increased in dependence on the age of the animals (Figure 4).

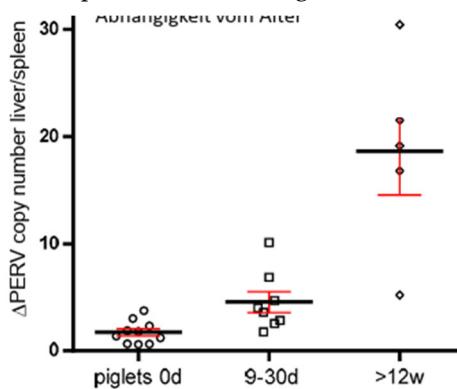


Figure 4. Presentation of the differences of the PERV copy number in spleen and liver in dependence on the age of the animals. The PERV copy number was determined in both organs of dead born piglets (day 0), 9-30 days old and 12 weeks old animals using ddPCR and the difference was indicated in dependence on the age.

3.6. Further Characterization of the Auckland Island Pigs

In order to characterize the Auckland Island pigs further, PCR-based methods for the detection of PERV-C, PCMV/PRV, PCV1, PCV2, PCV3, PLHV-1, PLHV-2, and PLHV-3 as well as well as a reverse transcriptase (RT) RT-PCR for HEV were performed (Table 2). The tests for all these viruses were negative. Negative was also a Western blot assay and an ELISA screening for antibodies against HEV, indicating that these animals are free of HEV. In addition to the Auckland Island pigs, the four

kidney cell lines from very young piglets were tested for PERV-C, PCMV/PRV, PCV3 and were found negative (Table 2).

4. Discussion

For the first time, a determination of the number of integrated PERVs in Auckland Island pigs was performed using the ddPCR method. The Auckland Island pigs had been used in the first xenotransplantation trials in New Zealand and Argentina [7, 8], they are virologically well characterized [4-6] and are the “cleanest” pigs compared with other pig breeds.

In the past it had been shown that the PERV copy number differs in dependence on the pig strain, on the individual animal tested, and on the method used (for review see [33]). Auckland Island pigs had been reported previously to contain three to 37 copies using a real-time PCR assay and a light cycler [5] and four to 40 copies were found using a PCR-based limited dilution assay (PLDA) [7]. In another study, relative gene dosages of PERV in Auckland Island pigs ranged from 2.8 ± 0.1 to 68 ± 1.6 at an average of 17.6 ± 10.4 using a real-time PCR [34]. This indicates a generally lower copy number compared to the numbers found here, but some animals had even a higher copy number (68). In our study we found that the PERV copy number is much lower in cells from very young piglets used for the SCNT (20-22) compared with the copy number in adult animals (ranging from 32 to 59 in the animals obtained from these cells by SCNT) (Figure 3).

This data confirms that the copy number depends on the method used and probably on the age of the animals. It is well known that PERVs are active and in addition to the endogenous proviruses there are *de novo* integrated proviruses in somatic cells. In the result, the number of integrated PERV proviruses differs from organ to organ (for review of numerous publications see [33]) and even in different parts of a single organ [24] in living pigs. The finding that the copy number is lower in cells from very young animals correlates with findings in another study: When analyzing 14-21-day-old neonatal and 1-2-year-old adult Belgian landrace pigs a significant increase in the copy number over time was also observed [35]. In these animals, an difference between least five copies and 80 copies was reported, indicating that the viruses are active and replicating in the neonatal pig. In Figure 5 a schematic presentation of these events is given. Indicated are the copy numbers found in the somatic cells of the animals and in the cells from young animals used for SCNT measured by ddPCR. Considering that copy numbers in adult animals are the result of *de novo* integrations in somatic cells, it logically follows that the number of proviruses in the germline can be at most 20 (Figure 5).

The fact that the number of integrated proviruses increased by age was also supported by a comparison of the copy number in two different tissues, spleen and liver, of other than Auckland Island pigs (Figure 4). Differences in the number of integrated copies means that in one or the other organs additional copies appeared in addition to the copy number in the germ line. The fact that the difference increases with age indicates that more new copies were integrated with age.

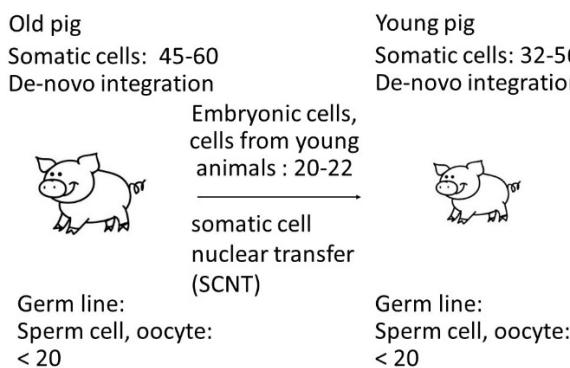


Figure 5. Copy number of PERV in somatic cells and in cells used for SCNT indicating increase of the copy number in adult animals.

The differences in the copy number of PERV in Auckland Island pigs when measuring by different methods [5, 7, 34 and this paper] do not only depend on the method. Based on our findings that the copy number increases due to *de novo* integrations, it also depends on the age of the animal which was not known to us and therefore cannot be taken into account. It would be highly interesting to test the PERV copy number in organs of a single animal over time. Furthermore, it may also depend on the tissue and the amount of PBMCs present in this tissue. PBMCs are prone for release of different PERVs [36] and *de novo* integration evidenced by the presence of integrated and release of infectious recombinant PERV-A/C from PBMCs [18, 20, 21, 36]. PERV-A/C have never been found in the germ line of pigs [37].

Auckland Island pigs are descendants of Western pigs brought to the island [1, 2]. This was confirmed by phylogenetic analyses of microsatellites and mitochondrial analyses showing that they are more closely related to European pigs than Chinese pigs [3]. The PERV copy number in Western pigs is higher compared with Chinese pigs. By analyzing 63 high-quality porcine whole-genome resequencing data, it was found that the PERV copy numbers in Chinese pigs were lower (32.0 ± 4.0) than in Western pigs (49.1 ± 6.5) [38].

From all methods ever used to count PERV copies in the DNA of pigs, such as Southern blot [39, 40], PCR titration [41], real-time PCR [42-47], fluorescence in situ hybridization (FISH) [30], PCR-based limited dilution assay [5], and genome wide sequencing [31], the ddPCR seems to be the most accurate method [24, 32, 33]. For example, using Southern blot analysis about 50 PERV copies were found in certain pig breeds [39, 40], but PCR titration [41] or real-time PCR [42-47] detected from one to 98 copies. By using FISH, 19 copies of PERV-A and 13 copies of PERV-B were found in Westran pigs [48]. Using genome-wide sequencing, 20 gammaretrovirus copies and 4 betaretrovirus copies were detected in Duroc pigs [49], indicating that sequencing does not detect all integrated viruses.

DdPCR was found to have numerous favorable features in comparison with all other methods including real-time PCR (for review see [23, 24, 32, 33]). One major advantage is that reference standard curves are not required, and bias effects arising from amplification efficiency and PCR inhibitors were reduced. The ddPCR technique provides absolute quantification based on the principles of sample partitioning and Poisson statistics, thus overcoming the normalization and calibrator issues; it has shown increased precision and sensitivity; it is relatively insensitive to PCR inhibitors and directly provides the result of the analysis expressed as number of copies of target per microliter of reaction [50].

Using ddPCR, fifty-nine copies were found when analyzing kidney fibroblasts from multi-transgenic pigs expressing the human transgenes CD46, CD55, CD59, HO1, and A20 [32]. When wild boars in and around Berlin were analyzed using droplet digital PCR, three to 69 PERV copies were found. A lower copy number, but a higher variability of this number was found in wild boars compared to domestic pigs, including minipigs [24].

In contrast to human endogenous retroviruses (HERVs), which are mostly defective and not replication-competent, PERVs can be released from normal pig cells and at least PERV-A and -B are infectious for certain human cells. Human embryonic kidney 293 cells are the most susceptible cells, mainly because these cells lost numerous intracellular restriction factors [51]. Other human cell lines such as THP-1 and C8166 cells could be infected, but only with a very low virus production compared with the 293 cells [52]. Important is the fact that primary human cells such as endothelial cells, vascular fibroblast, and mesangial cells could be infected with PERV [52, 53], but also with a low virus production and primary human PBMCs could only be infected with a human-cell-adapted PERV, characterized by a higher number of enhancer repeats and a high titer [54].

All PCR-based methods used to estimate the PERV copy number have a common limitation, in that they used primers mostly for the highly conserved pol gene. This allows to detect all PERVs, i.e. PERV-A, PERV-B and PERV-C. However, the detection of this sequence does not mean that there is an intact provirus able to produce infectious virus particles. Therefore, the estimated number of integrated proviruses measured with PCR-based methods tells us nothing concerning the safety of xenotransplantation, e.g., the risk posed by PERVs [55]. Only sequencing the pig genome could give more reliable information. However, sequencing of repetitive, nearly identical sequences is always

associated with problems as has been seen with hidden sequences of the human endogenous retrovirus HERV-K [56, 57]. In these investigations, in addition to 91 full-length HERV-K proviruses previously found in the human genome draft sequence hundreds more proviruses were detected. Sequencing the genome of a domestic Duroc pig, 212 endogenous retrovirus sequences were detected, among them 9 PERV-A, 11 PERV-B and 4 betaretrovirus copies [48], a lower number of PERVs compared with other methods [33].

One major result of the work was the elimination of PERV-C. Among the Auckland Island pigs analyzed in the first experiment approximately 53% of the animals were PERV-C-negative (Table 2). PERV-C is an ecotropic virus, infecting only pig cells and therefore does not pose a direct risk for xenotransplantation. However, recombinants between the human-tropic PERV-A and PERV-C have been observed, which are able to infect human cells and which are characterized by higher replication rates [58-60]. At present PERV-A/C were found mainly in minipigs, either released from their PBMCs [37, 60-62] or present in some organs of minipigs with melanomas [63] or even apparently healthy pigs [64]. One exception was the finding of PERV-A/C in farm animals with infectious diseases [22]. However, PERV-A/C was only found in somatic cells, never in the germ line [21, 65-67].

Compared with PERV-A and -B, PERV-C appears to be even younger in the pig lineage as not all domestic pigs have PERV-C sequence [39]. Miniature swine may be a unique pig breed as they possess more copies of PERV-C sequence (typically five to nine copies) than many other pig herds (one to three copies [40]), suggesting that PERV-C is active in miniature swine [19]. Although only limited studies have been performed on PERV transmission to human cells using primary cultures from other pig herds, it is intriguing that A/C recombinant PERV have been reported mainly from miniature swine but not from other herds [19]. However, there is no risk of PERV-A/C release from Auckland Island pigs generated by SCNT using nuclei from PERV-C-free Auckland Island pigs.

5. Conclusions

Auckland Island pigs had been used repeatedly as donors for pig islet cell transplantation into non-human primates and diabetic patients. In all preclinical and clinical trials no porcine viruses including PERV had been transmitted.

Auckland Island pigs are well characterized, they are free of many potentially pathogenic viruses and have a low PERV copy number in comparison with other pig breeds. The copy number is much lower in cells from very young piglets and in animals obtained by SCNT using nuclei from Auckland Island pig cells. To enhance the virus safety, PERV-C negative animals were selected.

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