

# Prevalence and New Host Records of Avian Blood Parasites in Songbirds from the West of Iran: Insights into Haemosporidian Infections and the Impact of Humidity

[Mohammad Aljehni](#) , Omid Mirshamsi , [Mansor Aliabadian](#) \*

Posted Date: 13 September 2023

doi: 10.20944/preprints202309.0822.v1

Keywords: Blood parasites, Iran, Leucocytozoon spp, Songbirds.



Preprints.org is a free multidiscipline platform providing preprint service that is dedicated to making early versions of research outputs permanently available and citable. Preprints posted at Preprints.org appear in Web of Science, Crossref, Google Scholar, Scilit, Europe PMC.

Copyright: This is an open access article distributed under the Creative Commons Attribution License which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

## Article

# Prevalence and New Host Records of Avian Blood Parasites in Songbirds from the West of Iran: Insights into Haemosporidian Infections and the Impact of Humidity

Mohammad Aljehni <sup>1</sup>, Omid Mirshamsi <sup>1,2</sup> and Mansour Aliabadian <sup>1,2,\*</sup>

<sup>1</sup> Department of Biology, Faculty of Science, Ferdowsi University of Mashhad, Mashhad 9177948974, Iran; mohamad.aljahny@gmail.com (M.A.); mirshams@um.ac.ir (O.M.); Tel.: +98-3880-5517.

<sup>2</sup> Research Department of Zoological Innovations (RDZI), Institute of Applied Zoology, Faculty of Science, Ferdowsi University of Mashhad, Mashhad 9177948974, Iran.

\* Correspondence: author: Mansour Aliabadian, aliabadi@um.ac.ir (M.A.); Tel.: +98-51-3880-5507. Fax: 9177948974.

**Abstract:** Haemosporidian parasites, including *Haemoproteus*, *Plasmodium*, and *Leucocytozoon*, are intracellular parasites found in the blood and tissues of birds worldwide. The prevalence of these parasites varies depending on bird species and habitats. Humidity, influenced by water availability, is one of the factors affecting their prevalence. In this study, we collected 145 blood samples from wild songbirds in the western region of Iran from April to November 2021–2022. Blood smears were prepared using the push-slide method and stained with Giemsa. The stained smears were examined under a light microscope. Among the seven songbird families sampled, blood parasites were detected in 31 individuals from five families. Of these, 23 individuals (15.86%) were from humid areas, while 8 individuals (5.51%) were from dry areas. The highest infection rates were observed in the Paridae (9.65%) and Motacillidae (4.82%) families, while the lowest infection rates were found in the Corvidae (1.37%) and Acrocephalidae (1.37%) families. We recorded two new hosts for *Leucocytozoon* spp. (*Motacilla flava* and *Acrocephalus melanopogon*) for the first time in the world and four new host records (*Motacilla flava*, *Pica pica*, *Acrocephalus melanopogon*, and *Acrocephalus agricola*) for blood parasites in the west of Iran. Our findings highlight the high prevalence of avian blood parasites in songbirds inhabiting humid areas, suggesting that these environments provide suitable conditions for the growth and reproduction of invertebrate hosts.

**Keywords:** blood parasites; Iran; *Leucocytozoon* spp.; songbirds

## 1. Introduction

Blood parasites, specifically haemosporidians, are highly prevalent infections in birds, surpassing the prevalence observed in other vertebrates. The global expansion and substantial prevalence rates of various haemosporidian species highlight the need for their identification and classification. These parasites are primarily transmitted through biting insects such as *Simulium* and *Culex*, with Passeriformes being particularly susceptible compared to other bird orders [1,2]. The life cycle of avian blood parasites is complex, involving both sexual (gametogenesis and fertilization) and asexual (sporogony) reproduction in the invertebrate host, and asexual reproduction (merogony) in the vertebrate host [3–5]. Avian malaria-like diseases are caused by *Leucocytozoon*, *Plasmodium*, and *Haemoproteus* in birds [6]. *Haemoproteus* spp. is considered one of the most significant genera of parasitic organisms and can infect over 50% of bird hosts worldwide [2]. *Leucocytozoon* spp. is also prevalent among wild birds [7]. *Plasmodium* spp., with its more than 200 species in 14 subgenera, includes five subtypes of avian malaria [8]. Among bird species, songbirds have been found to have the highest prevalence of haemosporidian infections. For example, in Bulgaria, songbirds exhibited an infection rate of 58.04% [8], while in Austria, the rate was 43.7% [9]. Similar studies conducted in Iran reported a prevalence of haemosporidian infections of 35.75% in northern regions [10] and 51.1%

in southern parts of Iran [11]. The prevalence and transmission of blood parasites depend on factors such as vector presence and bird population density [12]. The absence of suitable habitat for vectors of haemosporidian in arid environments has been suggested as a reason for the absence of infections in those areas [13,14]. For example, black flies (family: Simuliidae), which are common vectors of *Leucocytozoon spp.*, require clean water for reproduction [15,16], making it challenging to establish infections in dry and semi-arid environments. Conversely, mosquitoes and midges, which are vectors for haemosporidian parasites, require standing water for reproduction, increasing the probability of infection in ponds, lakes [17–19], slow rivers [4], stream margins, fens, and bogs [20]. Another study, by Krama et al. (2015) found that the prevalence of haemosporidian infections decreased with increasing distance from forest lakes [21]. Despite these findings, no studies have yet been conducted on the prevalence of haemosporidian parasites in songbirds in western Iran, nor have there been any comparisons of infection rates between humid and arid areas. Therefore, the aim of this study is to evaluate the prevalence of blood parasites in songbirds in western Iran using light microscopy and investigate whether the prevalence of these parasites in songbirds is related to environmental humidity.

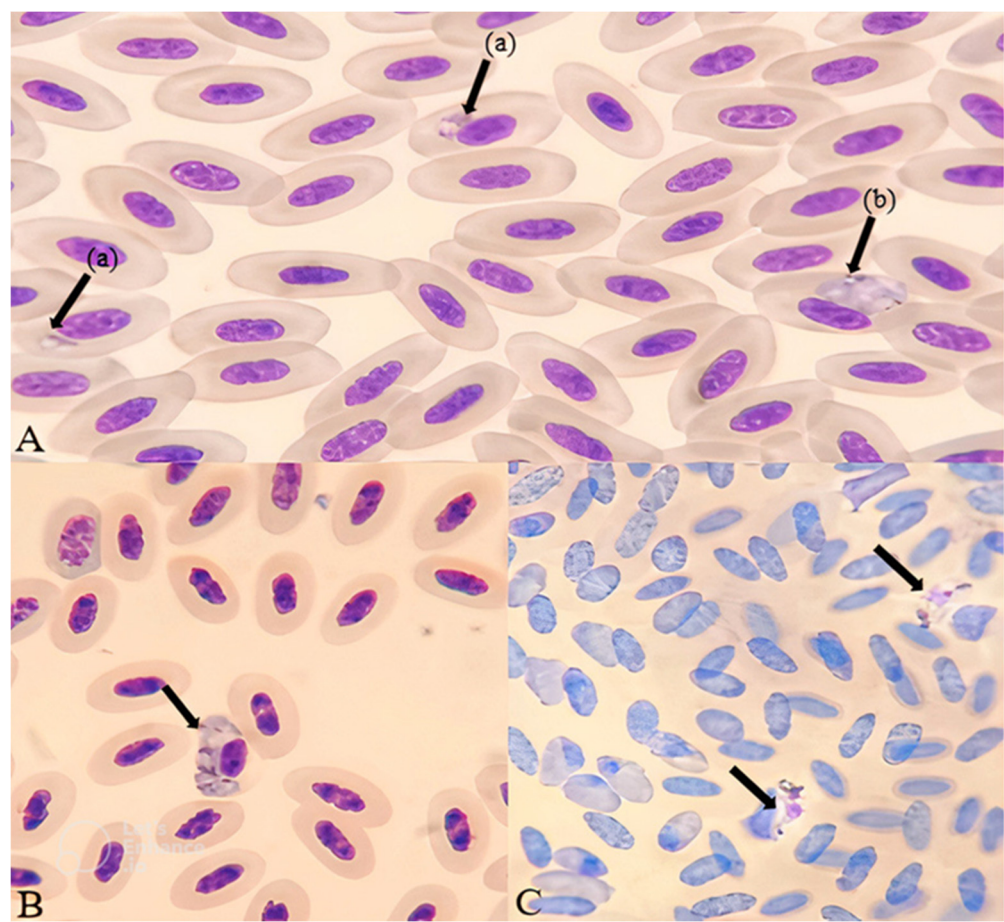
## 2. Materials and Methods

In this study, a total of 145 blood samples were collected from 145 songbirds representing nine species, seven genera, and seven families. The sampling took place in 13 different localities in the western region of Iran including Sanandaj (35°15'21.97'' N-47°00'58.86''E, 1385 m above sea level), Sarvabad (35°18'04.75''N-46°21'32.85''E, 1086 m above sea level), and Marivan (35°31'31.21''N-46°08'55.30''E, 1287 m above sea level). The sampling period extended from April to November 2021–2022, and the songbirds were captured using mist nets. To collect blood samples, an insulin syringe was used to draw blood from the brachial vein of each bird. The collected blood was then used to prepare smears using a push-slide method. After preparing the smears, they were air-dried, fixed with absolute methanol for five minutes, and dried again. Subsequently, the smears were stained with Giemsa stain at a pH of 7.1 for a duration of 20–25 minutes. The blood spreads were examined under a light microscope, following the method described by Valkiūnas (2005) [1]. To determine the humid and dry areas, the method proposed by Krama et al. (2015) was followed. Humid areas were identified based on the presence of rivers and lake margins, while dry areas were selected at least 4 kilometers away from any rivers or lakes. To assess the significant difference in blood parasite infection rates between individuals in humid and dry areas, we conducted a Fisher Exact Test ( $p < 0.05$ ) using SPSS version 22.0 (SPSS for Windows Inc., Chicago, Illinois, 2013) [22].

## 3. Results

In our study, we found that out of 145 host individuals, 31 (21.37%) were infected with avian blood parasites. Among the positive individuals, 14 birds were found to be infected with two genera (Co-infection) of blood parasites, namely *Haemoproteus* and *Leucocytozoon*. The overall prevalence of avian blood parasites was determined to be 30.33%, with *Haemoproteus* accounting for 17.93%, *Leucocytozoon* for 11.03%, and *Plasmodium* for 1.37% of the infections (Figure 1). Interestingly, we observed that 23 individuals (15.86%) were found in humid areas such as Sirwan River and Zarivar Lake, while 8 individuals (5.51%) were detected in dry areas within the city of Marivan and Sanandaj. Importantly, based on the Fisher Exact Test, we found a significant difference ( $\text{sig} = 0.00 < 0.05$ ) in the prevalence of blood parasites between humid and dry areas. The highest infection rate was observed in species belonging to the Paridae (9.65%) and Motacillidae (4.82%) families, while the lowest infection rate was observed in the Corvidae (1.37%) and Acrocephalidae (1.37%) families (Table 1). Among the 145 captured host songbirds, we recorded two new hosts for *Leucocytozoon spp.* (*Motacilla flava* and *Acrocephalus melanopogon*), which represents the first documentation of their infection worldwide (Figure 1C). Additionally, we identified four new hosts in the western region of Iran, including *Motacilla flava*, *Pica pica*, and *Acrocephalus melanopogon* for *Leucocytozoon spp.*, *Pica pica* and *Motacilla flava* for *Haemoproteus spp.*, and *Acrocephalus agricola* for *Plasmodium spp.* (Table 1). Furthermore, we detected 14 cases (9.65%) of co-infection with *Haemoproteus spp.*

and *Leucocytozoon* spp., with the Paridae family exhibiting the highest co-infection rate (eight individuals), while the Acrocephalidae family had the lowest co-infection rate (one individual).



**Figure 1.** A) Early (left (a)) and late (right (b)) trophozoites of *Plasmodium* spp. in *Acrocephalus agricola*, B): *Haemoproteus* spp. in *Motacilla flava*, and C) *Leucocytozoon* spp. in *Motacilla flava*. (Stained with the Giemsa stain, Magnification 1000×).

**Table 1.** Prevalence of the avian haemosporidian parasite in songbirds from the west of Iran.

| Family and Species for Host        | NEx | N Inf | Inf (%) | H | P | L | Hm | Pm | Lm |
|------------------------------------|-----|-------|---------|---|---|---|----|----|----|
| <b>Corvidae</b>                    |     |       |         |   |   |   |    |    |    |
| <i>Pica pica</i> +                 | 4   | 2     | 50%     | 2 | 0 | 2 | 2  | 1  | 2  |
| <b>Pycnonotidae</b>                |     |       |         |   |   |   |    |    |    |
| <i>Pycnonotus leucotis</i>         | 1   | 0     | 0 %     | 0 | 0 | 0 | 0  | 0  | 0  |
| <b>Passeridae</b>                  |     |       |         |   |   |   |    |    |    |
| <i>Passer domesticus</i>           | 74  | 6     | 8.10 %  | 3 | 1 | 2 | 16 | 40 | 12 |
| <b>Motacillidae</b>                |     |       |         |   |   |   |    |    |    |
| <i>Motacilla flava</i> *+          | 10  | 7     | 70 %    | 7 | 0 | 3 | 5  | 6  | 0  |
| <b>Acrocephalidae</b>              |     |       |         |   |   |   |    |    |    |
| <i>Acrocephalus agricola</i> +     | 3   | 1     | 33.33 % | 0 | 0 | 0 | 5  | 5  | 0  |
| <i>Acrocephalus melanopogon</i> *+ | 1   | 1     | 100 %   | 1 | 0 | 1 | 1  | 2  | 0  |
| <b>Turdidae</b>                    |     |       |         |   |   |   |    |    |    |
| <i>Turdus merula</i>               | 1   | 0     | 0       | 0 | 0 | 0 | 10 | 25 | 22 |
| <b>Paridae</b>                     |     |       |         |   |   |   |    |    |    |
| <i>Parus major</i>                 | 22  | 6     | 27 %    | 5 | 0 | 4 | 18 | 18 | 48 |



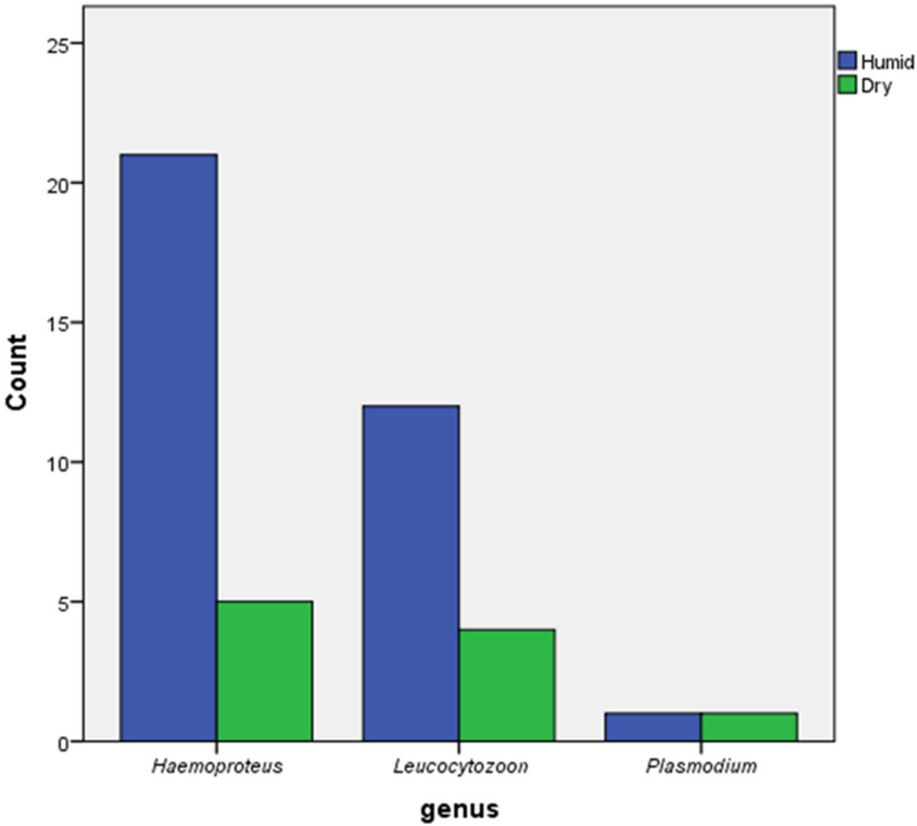
|                            |     |       |         |   |   |   |        |      |        |
|----------------------------|-----|-------|---------|---|---|---|--------|------|--------|
| <i>Cyanistes caeruleus</i> | 29  | 8     | 27.58 % | 8 | 0 | 4 | 20     | 22   | 26     |
| <b>Total</b>               | 145 | 31    | -----   |   |   |   | 26     | 2    | 16     |
| <b>Prevalence, %</b>       |     | 21.37 | ----    |   |   |   | 17.39/ | 1.37 | /11.03 |

<sup>1</sup> NEx = Number examined, N Inf = number infection, Inf % = Infection %, H = *Haemoproteus* spp.; P = *Plasmodium* spp.; L = *Leucocytozoon* spp, Hm= number of a recorded lineage of *Haemoproteus* in MalAvi database, Pm= number of a recorded lineage of *Plasmodium* in MalAvi database, Lm= number of a recorded lineage of *Leucocytozoon* in MalAvi database, + New host records for Iran, \* new host record for the world.

4. Discussion

This study showed a range of infection rates among different avian species, with the highest prevalence of *Haemoproteus* parasite infection being 17.93% and the lowest prevalence of *Plasmodium* spp. infection at 1.37%. The overall prevalence of infection varied between 8.10% and 100% across different avian species (Table 1). Notably, *Cyanistes caeruleus* had the highest infection rate (eight individuals), followed by *Motacilla flava* (seven individuals) and *Parus major* (six individuals). Additionally, *Motacilla flava* showed the highest prevalence of *Haemoproteus* spp., while *Parus major* and *Cyanistes caeruleus* showed the highest prevalence of *Leucocytozoon* spp.

Our findings indicated that humid areas had the significantly highest infection rates with haemosporidian parasites (Figure 2). These results align with a study conducted in Baikal Lake, where the prevalence of blood parasites was reported to be as high as 63.9% [23]. Similarly, high prevalence rates were observed in north-western Siberia (76.3%) [24], Bulgaria (58.04%) [8], Lake Superior in the USA (54%) [25], and southern Iran (51.1%) [11]. In contrast, dry areas demonstrated lower prevalence rates of haemosporidian parasites (5.51%) (Figure 2), which is consistent with a study conducted in Almeria City (3.45%) [26].



**Figure 2.** Comparison between individuals infected with haemosporidian parasites in humid (blue) and dry (green) areas in this study.

The overall prevalence of infection in our study aligns with the findings in north Iran [10], which may be due to similar weather conditions between the regions. However, the high prevalence in

southern Iran [11] can be attributed to the relatively high temperature and humidity in that area. Furthermore, our study found a high prevalence of *Haemoproteus* spp., consistent with other studies [8,11]. However, some studies have reported *Leucocytozoon* spp. as the most prevalent blood parasite in subarctic regions, likely due to the higher abundance of *Leucocytozoon* hosts in those areas [25], or due to higher elevations and colder conditions [27–32]. Temperature is known to be a determining factor for the prevalence of *Plasmodium* spp., typically more prevalent in hot areas [33]. However, our study and the findings in north Iran [10] revealed a low infection rate with the *Plasmodium* parasite (1.37% and 1.26%, respectively), contrasting with the higher rates observed in Bulgaria (24.7%) southern Iran (14.5%) and Austria (13.8%) [8, 9, 11]. In humid areas of our study, the infection rates for *Haemoproteus* spp. and *Leucocytozoon* spp. were higher compared to dry areas, while the infection rate for the *Plasmodium* parasite was similar in both dry and wet areas (Figure 2). This suggests that the prevalence of *Plasmodium* spp. is more influenced by temperature.

In humid areas, we observed 12 cases of co-infection with *Haemoproteus* spp. and *Leucocytozoon* spp., while there were only two cases in dry areas. The highest occurrence of co-infection was found in *Parus major* and *Cyanistes caeruleus*, while *Acrocephalus melanopogon* had the lowest occurrence. However, we did not observe any co-infection between *Haemoproteus* spp. and *Plasmodium* spp., or between *Plasmodium* spp. and *Leucocytozoon* spp. This finding is consistent with previous studies [10, 11].

The overall co-infection rate of 14 individuals in our study aligns with the results of a study conducted by Ghaemitalab et al. (2021), which reported 13 cases of co-infection with *Haemoproteus* spp. and *Leucocytozoon* spp. [11]. However, in the study by Nourani et al. (2018) only one sample was found to be co-infected with *Haemoproteus* spp. and *Leucocytozoon* spp. [10]. The higher occurrence of co-infection in the Paridae family in our study is consistent with a study by van Rooyen et al. (2013), which reported a high incidence of co-infection in great tits [34]. In sum, our results demonstrate a high prevalence of haemosporidian parasites and co-infection in humid areas, indicating that suitable environmental conditions for vectors may play an important role. In future research, it would be valuable to investigate the impact of temperature and other factors on blood parasite infection in these areas.

## 5. Conclusions

In conclusion, our study reveals a higher prevalence of haemosporidian parasite infections in songbirds residing in humid areas compared to dry areas. This difference can be attributed to the greater abundance and movement of blood parasite vectors in humid regions. Furthermore, our findings suggest that humidity plays a significant role as a determining factor in the prevalence of these parasites.

**Author Contributions:** Conceptualization: M.A., O.M., M.A.; Methodology: M.A., M.A., O.M.; formal analysis and investigation: M.A., M.A.; Writing-original draft preparation: M.A., M.A.; Writing review and editing: M.A., M.A., O.M.; Supervisions: M.A., O.M. All authors have read and agreed to publish this manuscript.

**Funding:** This research was supported by the Ferdowsi University of Mashhad (M.A., project number 3/56606).

**Institutional Review Board Statement:** This study was carried out according to the protocol approved by a committee on the ethics of animal experiments of the Ferdowsi University Mashhad (Ethical Certificate No. IR.UM.REC.1400.372). Furthermore, all field works were carried out with permits approved by the Department of the Environment. (Permits No. 2/1177/ 1401).

**Acknowledgments:** We thank the Vice-Chancellor for Research of the Ferdowsi University of Mashhad and the Department of Environment in Kurdistan.

**Conflicts of Interest:** The authors declare that they have no conflict of interest.

## References

1. Valkiūnas G, Anwar AM, Atkinson CT, Greiner EC, Paperna I, Peirce MA. What distinguishes malaria parasites from other pigmented haemosporidians? *Trends Parasitol.* **2005**; 21(8):357-358.
2. Valkiūnas G. *Avian Malaria Parasites and Other Haemosporidia*. CRC press; 2004.

3. Scheuerlein A, Ricklefs RE. Prevalence of blood parasites in European passeriform birds. *Proc R Soc B Biol Sci.* **2004**; 271(1546):1363-1370. doi:10.1098/rspb.2004.2726
4. Wood MJ, Cosgrove CL, Wilkin TA, Knowles SCL, Day KP, Sheldon BC. Within-population variation in prevalence and lineage distribution of avian malaria in blue tits, *Cyanistes caeruleus*. *Mol Ecol.* **2007**; 16(15):3263-3273.
5. Jenkins T, Owens IPF. Biogeography of avian blood parasites (*Leucocytozoon* spp.) in two resident hosts across Europe: phylogeographic structuring or the abundance–occupancy relationship? *Mol Ecol.* **2011**; 20(18):3910-3920.
6. Medeiros MCI, Hamer GL, Ricklefs RE. Host compatibility rather than vector–host-encounter rate determines the host range of avian *Plasmodium* parasites. *Proc R Soc B Biol Sci.* **2013**; 280(1760):20122947.
7. Peirce MA. Distribution and host-parasite check-list of the haematozoa of birds in Western Europe. *J Nat Hist.* **1981**; 15(3):419-458.
8. Dimitrov D, Zehndtjiev P, Bensch S. Genetic diversity of avian blood parasites in SE Europe: Cytochrome b lineages of the genera *Plasmodium* and *Haemoproteus* (Haemosporida) from Bulgaria. *Acta Parasitol.* **2010**; 55:201-209.
9. Himmel T, Harl J, Matt J, Weissenböck H. A citizen science-based survey of avian mortality focusing on haemosporidian infections in wild passerine birds. *Malar J.* **2021**; 20(1). doi:10.1186/s12936-021-03949-y
10. Nourani L, Aliabadian M, Mirshamsi O, Dinparast Djadid N. Molecular detection and genetic diversity of avian haemosporidian parasites in Iran. *PLoS One.* **2018**; 13(11):e0206638.
11. Ghaemitalab V, Mirshamsi O, Valkiūnas G, Aliabadian M. Prevalence and genetic diversity of avian haemosporidian parasites in southern Iran. *Pathogens.* **2021**; 10(6):645.
12. Bennett GF, Squires-Parsons D, Siikamäki P, Huhta E, Allander K, Hillström L. A comparison of the blood parasites of three Fenno-Scandian populations of the pied flycatcher *Ficedula hypoleuca*. *J Avian Biol.* Published online **1995**:33-38.
13. Laird M. A lack of avian and mammalian haematozoa in the Antarctic and Canadian Arctic. *Can J Zool.* **1961**; 39(2):209-213.
14. Merino S, Barbosa A, Moreno J, Potti J. Absence of haematozoa in a wild chinstrap penguin *Pygoscelis antarctica* population. *Polar Biol.* **1997**; 18:227-228.
15. Little RM, Earlé RA. Sandgrouse (Pterocleididae) and sociable weavers *Philetarius socius* lack avian haematozoa in semi-arid regions of south Africa. *J Arid Environ.* **1995**; 30(3):367-370. doi:10.1016/S0140-1963(05)80011-4
16. Earlé RA. Absence of haematozoa in some Charadriiformes breeding in the Taimyr Peninsula, Russia. *Ardea.* **1993**; 81:21-24.
17. Kettle DS. *Medical and Veterinary Entomology*. Croom Helm Ltd.; 1984.
18. Krams I, Cirule D, Krama T, et al. Effects of forest management on haematological parameters, blood parasites, and reproductive success of the Siberian tit (*Poecile cinctus*) in northern Finland. *Ann Zool Fennici.* **2010**; 47(5):335-346. doi:10.5735/086.047.0504
19. Krams I, Suraka V, Rattiste K, et al. Comparative analysis reveals a possible immunity-related absence of blood parasites in Common Gulls (*Larus canus*) and Black-headed Gulls (*Chroicocephalus ridibundus*). *J Ornithol.* **2012**; 153(4):1245-1252. doi:10.1007/s10336-012-0859-6
20. Hendry G. *Midges in Scotland*. Aberdeen University Press; 1989.
21. Krama T, Krams R, Cirule D, Moore FR, Rantala MJ, Krams IA. Intensity of haemosporidian infection of parids positively correlates with proximity to water bodies, but negatively with host survival. *J Ornithol.* **2015**; 156(4):1075-1084. doi:10.1007/s10336-015-1206-5
22. Statistics IS. IBM Corp. Released 2013. IBM SPSS Statistics for Windows, Version 22.0. Armonk, NY: IBM Corp. *Google Search*. Published online **2013**.
23. Palinauskas V, Efremov VD, Leonid V, Markovets MY, Kosarev V V, Sokolov L V. Occurrence of avian haematozoa in Ekaterinburg and Irkutsk districts of Russia. *Ekologija.* **2005** ;(4):8-12.
24. Yusupova DA, Schumm YR, Sokolov AA, Quillfeldt P. Haemosporidian blood parasites of passerine birds in north-western Siberia. *Polar Biol.* Published online **2023**:1-15.
25. Ferrer MM. Avian Haemosporidian Blood Parasite Diversity, Prevalence, and Distribution in Michigan's Western Upper Peninsula. Published online **2022**.

26. Valera F, Carrillo CM, Barbosa A, Moreno E. Low prevalence of haematozoa in Trumpeter finches *Bucanetes githagineus* from south-eastern Spain: additional support for a restricted distribution of blood parasites in arid lands. *J Arid Environ.* **2003**; 55(2):209-213.
27. Cuevas E, Vianna JA, Botero-Delgadillo E, et al. Latitudinal gradients of haemosporidian parasites: prevalence, diversity and drivers of infection in the Thorn-tailed Rayadito (*Aphrastura spinicauda*). *Int J Parasitol Parasites Wildl.* **2020**; 11:1-11.
28. Fecchio A, Bell JA, Bosholn M, et al. An inverse latitudinal gradient in infection probability and phylogenetic diversity for *Leucocytozoon* blood parasites in New World birds. *J Anim Ecol.* **2020**; 89(2):423-435.
29. Haas M, Lukáň M, Kisková J, Hrehová Z. Occurrence of blood parasites and intensity of infection in *Prunella modularis* in the montane and subalpine zone in the Slovak Carpathians. *Acta Parasitol.* **2012**; 57(3):221-227. doi:10.2478/s11686-012-0041-6
30. Oakgrove KS, Harrigan RJ, Loiseau C, Guers S, Seppi B, Sehgal RNM. Distribution, diversity and drivers of blood-borne parasite co-infections in Alaskan bird populations. *Int J Parasitol.* **2014**; 44(10):717-727.
31. Galen SC, Witt CC. Diverse avian malaria and other haemosporidian parasites in Andean house wrens: Evidence for regional co-diversification by host-switching. *J Avian Biol.* **2014**; 45(4):374-386. doi:10.1111/jav.00375
32. Gonzalez AD, Matta NE, Ellis VA, Miller ET, Ricklefs RE, Gutierrez HR. Mixed species flock, nest height, and elevation partially explain avian haemoparasite prevalence in Colombia. *PLoS One.* **2014**; 9(6):e100695.
33. Loiseau C, Harrigan RJ, Bichet C, et al. Predictions of avian *Plasmodium* expansion under climate change. *Sci Rep.* **2013**; 3(1):1126.
34. Van Rooyen J, Lalubin F, Glaizot O, Christe P. Avian haemosporidian persistence and co-infection in great tits at the individual level. *Malar J.* **2013**; 12(1). doi:10.1186/1475-2875-12-40

**Disclaimer/Publisher's Note:** The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.