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## Article

# Differences in Bacterial Growth and Mortality between Seagrass Meadows and Adjacent Unvegetated Areas

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**Abstract:** A seagrass meadow is one of the most important ecosystems around the world, both economically and ecologically. An important feature of this ecosystem is the presence of large coastal seagrass beds, which dominate the primary production and contribute to the secondary productivity of the ecosystem. The microbial loop (consuming bacterial biomass by grazers and using seagrass-derived detritus by bacteria) may be an important mechanism for transferring seagrass-derived organic matter to aquatic food chain. The goal of this study is to gain a better understanding of how bacterial growth and mortality (grazing and viral lysis rates) differ in unvegetated meadow habitats and seagrass habitats. In a seagrass environment during the summer, we have found that viral lysis and grazing both result in similar mortality rates of bacteria during the summer season. It has been found, however, that bacterial production is controlled by the availability of resources (bottom-up control) in adjacent unvegetated waters, and is thus cycled internally within the bacteria-virus-DOC loop within those waters. The fate and cycling of organic matter in our study region may be impacted by this shift in organic matter fate and cycling.

**Keywords:** seagrass; microbial loop; viral lysis; mortality rates

## 1. Introduction

As one of the most important components of coastal benthic ecosystems, seagrass meadows play an important role since seagrasses are highly productive primary producers [1,2], providing food and habitat for various marine organisms [3,4]. Seagrasses have been shown to play an important role in capturing and storing carbon in the ocean, as well as acting as a carbon sink [5]. Through photosynthesis, they absorb carbon dioxide from the atmosphere and turn it into biomass by capturing carbon dioxide in the process. There is no doubt that seagrass ecosystems are one of the most significant natural carbon sinks on earth, due to the fact that they are able to sequester significant amounts of carbon, store it as organic carbon for long periods, and are capable of occurring worldwide [6]. In addition, seagrass meadows play an important role in absorbing carbon dioxide (blue carbon), helping to reduce ocean acidification [7].

The quality and quantity of dissolved organic carbon (DOC) in water is considered to be one of the main limiting factors for the growth of heterotrophic bacteria [8]. In areas of marine grass meadows, the amount of DOC is higher than in adjacent areas of unvegetated meadows [9]. There is also evidence to suggest that seagrass roots contain organic matter that can directly transfer to benthic bacteria through exudates, in addition to supporting productive epiphytic communities through exudates [10,11]. It is nonetheless noteworthy that Boschker et al. [12] conducted stable carbon isotope experiments in eutrophic environments, and discovered that the DOC derived from seagrasses in sediments from *Zostera* spp meadows played a relatively small role in bacterial growth, suggesting



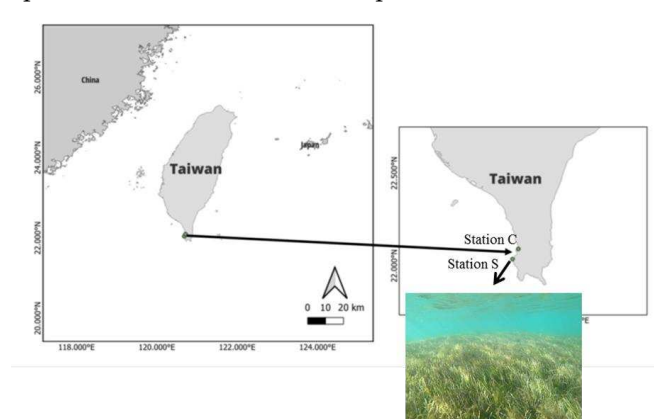
that benthic algae played the main role in supplying carbon to bacteria. However, seagrass-exuded DOC has been found to be a carbon source that contributes a significant amount of carbon to bacterial growth in the sediment of oligotrophic seagrass beds [10,13]. Seagrasses are also able to release DOC and oxygen as a result of their diel photosynthetic activity which also accounts for the amount of these substances they release [14–16]. Seagrass rhizospheres have been found to have twofold higher bacterial production rates during noon than at night, compared to non-vegetated sediments nearby [17]. In general, most information on bacterial biomass and production in seagrass systems is available in some studies [7,12–14], but no study on bacterial mortality, especially viral lysis on bacterial community in seagrass environments. There is no doubt that viral lysis and protozoan grazing are both crucial factors in controlling bacterial mortality in aquatic ecosystems. It is important to be aware that the process by which bacteria are lysed or grazed has different implications for the microbial food web. Moreover, it has impacts on biogeochemical cycling as well. In order to fully understand the complex dynamics of these systems, it is essential to understand and appreciate the role played by these different mechanisms. For understanding and quantifying the carbon cycle in seagrass-dominated ecosystems, it is essential to have a reliable measure of bacterial growth and mortality in the system.

We aim to better understand how seagrass and unvegetated meadow habitats differ in terms of bacterial growth and mortality. Therefore, the primary focus of this study was determining bacterial growth and mortality using modified dilution experiments [18]. In this study, we hypothesized that exposition to seagrass-derived DOC would result in higher bacterial growth and mortality rates than phytoplankton in unvegetated meadows adjacent.

## 2. Materials and Methods

### 2.1. Study site and samplings

In June 2023, two established stations located in Kenting collected surface samples from the southern part of the Taiwan coastal area (Figure 1). The seagrass meadow (Station S) was compared against the control (Station C) at each site at a depth of about 1 m (Figure 1). We gently poured surface water into 10L polypropylene carboys after collecting it with a bucket. Following the casting of the sampling bucket, the temperature of the water was measured immediately, and all samples were brought to the laboratory within 30 minutes. According to our previous studies [19,20], we analyzed DOC concentration. To determine the DOC content, the water sample was filtered through a 0.7  $\mu\text{m}$  glass fiber filter (GF/F, Whatman). To prevent contamination from organic materials, the glass bottles for DOC analysis were pre-washed in 0.1 N HCl and pre-combusted for 4 hours at 450°C.



**Figure 1.** The sampling sites are illustrated on this map. Stations S and C represent seagrass meadows and adjacent unvegetated areas, respectively.



## 2.2. Modified dilution experiments

We examined how the seagrass environment affected surface waters during summer compared with adjacent unvegetated areas on bacterial growth and loss caused by grazing and viral infection. Our objective was to assess the growth, grazing, and viral lysis rates of bacteria by taking advantage of the modified dilution method [18] in order to achieve these results. In the first step, seawater waters (2 L) were passed through a 10  $\mu\text{m}$  mesh and filtered through Nuclepore 47 mm filters (type PC, pore size 0.2  $\mu\text{m}$ ) to produce the 'standard' diluent. Following the filtration of the seawater sample (10  $\mu\text{m}$ ), the 0.2  $\mu\text{m}$  filtered seawater was diluted using a 4-point dilution series of 25, 50, 75, and 100%. A dilution series using 30 kDa filtered seawater instead of 0.2  $\mu\text{m}$  filtered water altered grazing as well as viral mortality. A water bath set to the temperature of the in situ seawater at the time of sampling was used to incubate triplicates of 50 ml polycarbonate bottles under natural light for 24 hours. To determine the abundance of viruses and bacteria, triplicate subsamples were collected from  $t_0$ . At the end of the experimental period,  $t_{24}$ , final samples were taken from every bottle. We calculated the net growth rate for bacteria at each dilution within an experimental series (i.e. 0.2  $\mu\text{m}$  and 30 kDa) as  $\ln(N_{t_{24}}/N_0)/t$ .  $N_{t_{24}}$  and  $N_0$  represent the final and initial abundance of bacteria, respectively, and  $t$  represents the experimental duration (24 hours).

According to Evans et al. [18], growth, grazing, and viral-induced mortality coefficients were calculated for bacteria as described in this paper. In order to calculate the growth rates of bacteria ( $\mu$ ) without both factors of mortality, a dilution series regression of 30 kDa was used. In this study, the nanoflagellate grazing coefficient ( $mg$ ) was calculated using a slope of the 0.2  $\mu\text{m}$ -filtered seawater dilution series. Additionally, the slope of the 30 kDa-filtered seawater dilution series was used to derive the coefficients of nanoflagellate grazing and viral-induced mortality ( $mg+mv$ ). It thus follows that the virus-induced bacterial mortality rate is equal to the difference between these two regression slopes, i.e.,  $mv = [(mv + mg) - mg]$ .

## 2.3. Flow cytometric analyses

An analysis of virus samples and bacteria samples was performed using a CytoFLEX S flow cytometer (Beckman Coulter, Indianapolis) that was equipped with an argon-ion laser at 488 nm, a filter at 525 nm, and a SYBR signal trigger in order to measure the presence of viruses and bacteria. To reduce interference from high particle density due to interference from high particle density, TE buffer (pH 8.0, EM grade) was used to dilute virus samples 1:10 prior to staining. After diluting the samples, incubate them for 10 minutes at 80°C with SYBR Green I (final concentration 50,000 of commercial stock). Following staining, samples were cooled in an ice bath to 25 °C and processed through FCM according to Brussaard [21]. It was determined that blank controls were run on TE buffer stained with the same concentrations of SYBR Green I as those used in the experiment, in order to detect and eliminate noise in the buffer. As described by Hammes and Egli [22], bacteria samples were stained with SYBR Green I (final concentration 1:10,000) for 15 min in the dark, before processing through FCM.

## 2.4. Statistical analysis

It was determined by a linear regression analysis of Model 1 to determine if there is a relationship between the net growth rate of bacteria and each of the series of dilution experiments (30 kDa and 0.2  $\mu\text{m}$ ). An analysis of variance (ANOVA) was used to examine the significance of the slope based on the data. The significance of the difference between the slopes of the 30 kDa and 0.2  $\mu\text{m}$  regression lines was determined by F-test. All statistical operations were carried out using the STATISTICA 7.0 software. The significance of the results was determined by a probability value of  $< 0.05$ .



3. Results

3.1. Station characteristics

In June, both stations recorded a temperature of approximately 28°C. Station S was situated in a seagrass environment and had a higher level of DOC (1685 µg L<sup>-1</sup>) compared to station C (as shown in Table 1). According to Table 1, the bacterial abundance in the seawater collected in June measured 7.3 and 5.2 × 10<sup>5</sup> cells mL<sup>-1</sup> at station C and S, respectively. Virus abundance was 3.4 × 10<sup>6</sup> mL<sup>-1</sup> and 8.9 × 10<sup>6</sup> cells mL<sup>-1</sup> at stations C and S, respectively (Table 1). Virus-to-bacterial ratios (VBR) were 4.7 and 17.1 at stations C and S, respectively (Table 1). Station C had the lower VBR.

**Table 1.** The data of dissolved organic carbon, bacterial abundance, viral abundance, virus-to-bacteria ratio (VBR), bacterial growth rate, grazing and viral lysis at the sampling stations. \*: growth rate of bacteria was calculated from the fractionated series of 0.2-µm. nd: not determined.

	DOC µg L <sup>-1</sup>	bacteria 10 <sup>5</sup> cells mL <sup>-1</sup>	viruses 10 <sup>6</sup> viruses mL <sup>-1</sup>	VBR	growth rate (d <sup>-1</sup> )	grazing rate (d <sup>-1</sup> )	viral lysis (d <sup>-1</sup> )
Station C	1196	7.3	3.4	4.7	1.35*	0.007	nd
Station S	1685	5.2	8.9	17.1	2.05	0.007	0.005

3.2. Bacterial growth rates

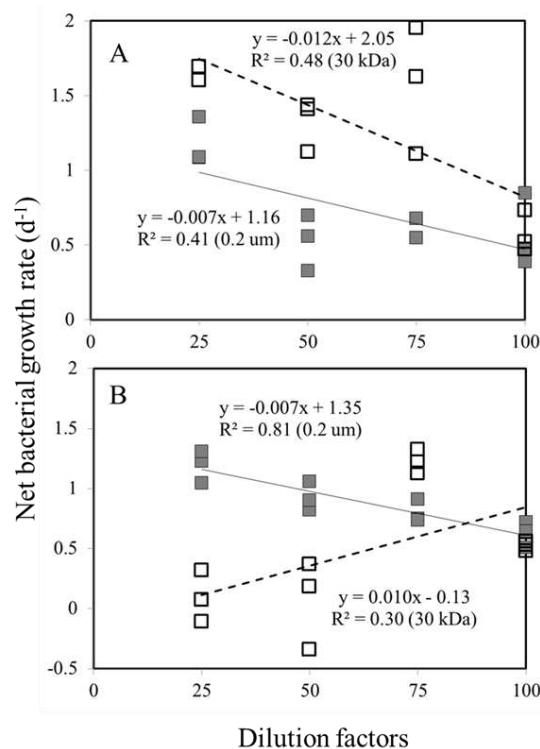
Using least-square regression, it was determined whether net growth rates increased directly with the dilution factor (Figure 2). For station S, there was a highly significant relationship (ANOVA,  $p < 0.05$ ) between bacterial net growth rate and dilution level in both series of 0.2 m and 30kDa (Figure 2A). According to Figure 2A, the y-intercepts of these regression lines were 1.16 and 2.05 d<sup>-1</sup> for 0.2 m and 30 kDa fractionated series, respectively. An instantaneous growth rate of 2.05 d<sup>-1</sup> is calculated without lytic or grazing pressure (Table 1).

In the adjacent unvegetated areas experiments, we also found a significant relationship (ANOVA,  $p < 0.05$ ) between the bacterial net growth rate and the level of dilution in both the 0.2-µm and 30-kDa series (Figure 2B), but a positive slope was found in the 30 kDa dilution series (Figure 2B), and so it was impossible to quantify their specific potential growth rates. Figure 2B shows that the y-intercept of these regression lines is 1.35 d<sup>-1</sup> for fractionated series of 0.2-µm. In the absence of grazing pressure, this value represents the instantaneous growth rate (Table 1).

3.3. Bacterial mortality

The regression slopes of the 0.2-µm and the 30-kDa dilution series were clearly different ( $F$ -test,  $p < 0.05$ ), indicating that the gradient in grazers and viral abundance created in the 0.2 µm and 30-kDa dilution series had a significant impact on the mortality of bacteria at station S. Thus, the regression coefficients (slope) at station S were -0.007 and -0.012 d<sup>-1</sup> for the 0.2 µm and 30 kDa fractionated series, respectively (Figure 2A). The difference between the 2 slope values of 0.005 d<sup>-1</sup>, indicated bacterial mortality due to viruses, lytic mortality accounted for was 42% of the total mortality at station S (Table 1). According to standard dilution experiments (0.2 m fractionated series), the bacterial mortality rate due to nanoflagellate grazing at station C was 0.007 d<sup>-1</sup> (Figure 2B); however, in this study, we were unable to estimate the viral mortality of bacteria at station C (Figure 2B).





**Figure 2.** Bacterial net growth rate vs. fraction of sample water in the dilution series experiments incubated at station S (A) and C (B), respectively. Seawater sample (<10  $\mu\text{m}$ ) diluted with the 30 kDa filtered seawater (open square), seawater sample (<10  $\mu\text{m}$ ) diluted with the 0.2  $\mu\text{m}$  filtered seawater (filled square).

#### 4. Discussion

Various physical and biological phenomena interact with each other to determine how the dynamics of a population of any given group of organisms proceed. As a result of advection and diffusion, bacteria are subjected to two types of physical processes. As well as cell division and cell death, grazing, and viral lysis are some of the biological processes that affect them. As far as I know, the modified dilution method is the only method available that will allow the simultaneous measurement of bacteria growth and mortality rates simultaneously. The effect of viruses on marine bacteria in our study area has not been studied to the best of our knowledge, therefore the rate of viral lysis of marine bacteria in the southern part of the Taiwan coastal area remains largely unknown. As the first study to identify the mortality effects of both viral lysis and grazing on bacteria in a seagrass zone as well as an adjacent unvegetated zone, the results of the study were compared. Although both viral lysis and grazing caused similar mortality rates of bacteria in seagrass in summer, grazing mortality of bacteria prevailed in adjacent unvegetated zones where grazing mortality dominated. Moreover, our data indicate that bacteria in adjacent unvegetated waters are controlled by available resources (bottom-up control) and are cycling within the bacteria-virus-DOC loop that takes place in adjacent unvegetated waters. It is possible that this shift in the fate and cycling of organic matter may have implications for the fate and cycling of organic matter in the regions we studied.

##### 4.1. Bacterial growth and mortality

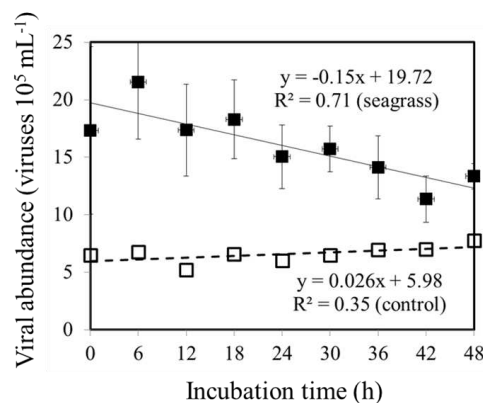
In terms of the amount of productivity that benthic producers generate as well as the fate of that productivity, the studies of benthic producers can be of importance to climate studies. It has been estimated that there are approximately  $6 \times 10^5 \text{ km}^2$  of seagrass covering the global seafloor, and their net primary production (NPP) is estimated to be approximately 0.5 PgC per year. As a result, this represents roughly 1% of NPP of the phytoplankton in the ocean [23]. Although benthic producers



play an important role in both the local ecology and biogeochemical cycle of a particular area, the extent to which they contribute to this process also depends on the extent to which production is left unremineralized by herbivores and decomposers to accumulate on the seabed [24]. During photosynthesis by seagrass roots and rhizomes, the organic carbon that is produced is released, and some of it can be consumed by bacteria in order to satisfy their nutritional needs [25]. The study also found that sources of organic carbon outside of the seagrass community are able to contribute to increased bacterial productivity [25]. A substantial amount of bacterial production has been reported in ecosystems dominated by seagrass in previous studies. It has been reported that more than half of the total primary production in seagrass meadows in Australia could be attributed to heterotrophic bacteria (sediments and the water column) [26]. As a result of the current study, the DOC value of the seagrass was found to be higher than that of the adjacent meadow habitats (Table 1). Microbial loops (the utilization of seagrass-derived detritus by bacteria, and the consumption of bacterial biomass by grazers) might contribute to the transfer of seagrass-derived organic matter to aquatic ecosystems. In seagrass-dominated ecosystems, the question of whether bacteria are carbon sinks or carbon links remains open.

The technique of dilution has been successfully used on a variety of prey as well as in a variety of environments [27–29]. Moreover, Evans et al. [18] also applied a dilution series that included microzooplankton and viruses to determine the mortality caused by virus lysis in the laboratory. Based on the regression coefficient for apparent growth rate versus dilution factor of 0.2  $\mu\text{m}$  dilution series, only microzooplankton grazing rate (mg) was considered. Using 30 kDa filtered seawater as a diluent, the regression slope represented both the loss of bacteria due to grazing, as well as the loss of viruses (mg + mv). The viral mortality rate of bacteria was calculated, based on the difference between the regression slopes drawn for the two dilution series, using the difference between the slopes of the regression equation. Compared to other methods for estimating viral impact on host mortality, the dilution method provides quantitative insight into both viral lysis and microzooplankton grazing without requiring conversion factors. There are, however, two critical assumptions associated with using the dilution technique in many cases, which are not satisfied. The first point to keep in mind is that, based on the assumptions of the dilution method, plankton growth is exponential, without being influenced by the dilution factor [30]. As a consequence, nutrient availability as well as other factors of the environment should not restrict the growth of plankton in any way. It is interesting to note that the results of Ayukai [31] differ from the findings of Landry and Hassett [30] concerning the effects of limiting nutrients on their study. In an experiment performed by Ayukai [31], *Synechococcus* spp. growth was reduced by most diluted treatments. Similar to Ayukai [31], we found that the positive slope of the fractionated 30 kDa series is contrary to expectations, and we were unable to measure the rate of growth and mortality of bacteria at station C by the same method (Figure 2B). According to the dilution method, a second assumption is made that the mortality rate of plankton resulting from predation is proportional to the amount of dilution seen on the abundance of parasites (viruses) and predators (grazers) [32]. The dilution gradient established at the beginning of the experiment should, therefore, remain constant throughout the experiment because predators or viruses should remain prevalent during the experiment. In our study, we observed that the abundances of the viruses significantly differed between the start and end of each experiment, which did not satisfy this assumption. During the present experiments at station S, we observed a decrease in viral abundance after the viral dilution series were observed over time in a 25% dilution factor of 30 kDa fractionated series (Figure 3). In seagrass-dominated ecosystems, bacteria are more likely to be infected by viruses at a 25% dilution factor of 30 kDa fractionated series within 24 hours.





**Figure 3.** Temporal variations of viral abundance during 24 h incubations for 25% diluted factor in the 30 kDa filtered seawater in seagrass meadows (filled black square) and adjacent unvegetated areas (open square), respectively.

The present study showed that there were two types of responses to the modified dilution experiment at station C (Figure 2B). It is expected that there will be a negative slope in a regression line of apparent growth versus a fraction of 0.2- $\mu$ m filtered water when there is a significant mortality impact from grazing. In spite of this, the slope generated from the fractionated 30 kDa series is the opposite of what one would expect, making it impossible to calculate the growth and mortality rates for bacteria. Studies suggest that viral lysis products, such as cell content and fragments of cell walls, enter the organic matter pool, which bacteria primarily use. As a result of this viral shunt [33,34], bacterial production increases, and the bacteria-nanoflagellate relationship is reduced. There is a possibility that a reduction in nutrient remineralization might have caused a decrease in bacterial instantaneous growth rate due to the dilution of heterotrophic bacteria and viruses from the 30 kDa fractionated series as a result of the decreased mineralization. It seems based on our findings that nutrients were limited in the experiments, even for those cases in which positive-linear responses were detected. We believe that this was due to the fact that dilution of the original water may disrupt the supply of nutrients and DOC to bacteria, resulting in a lower growth rate in the most diluted treatments based on these findings. There is still a great deal of work to be done in order to understand how viruses may lead to nutrient regeneration within aquatic environments. The issue of nutrient regeneration in microbial communities remains challenging, however, as it is difficult to directly measure it. There have been some experiments in the field with viral-host systems that have provided evidence that the regeneration of nutrients by viral lysis may be an important ecological function in some environments [35].

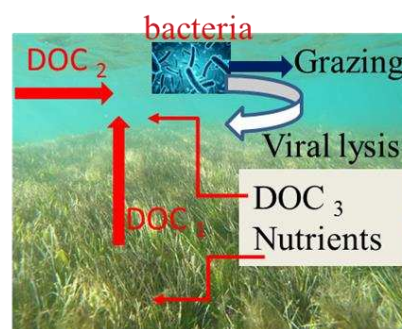
#### 4.2. Microbial ecology in seagrass meadows

A further study is needed for the estimation of carbon and energy flux through heterotrophic microorganisms as well as the possible role bacteria play in the trophic dynamics of ecosystems. A debate has been going on about whether bacterial production and biomass are controlled by available resources (bottom-up control) or predators/viruses (top-down control). The present study does not identify what factors may be responsible for the variation in growth and mortality of seagrass meadows and adjacent non-vegetated areas, but it is possible to speculate that there are some possible explanations. The carbon sources available for bacterial growth are mostly likely to come from three important sources in the seagrass meadow environment. During the photosynthesis process, DOC is released by seagrasses as the first step (shown as DOC<sub>1</sub>) (Figure 4). Also during phytoplankton photosynthesis, another DOM is released extracellularly (DOC<sub>2</sub>). As a third effect, viral lysis of bacteria would release relatively labile dissolved organic carbon into the marine DOC pool (DOC<sub>3</sub>). As a result of the fact that carbon is not only fixed photosynthetically but is also recycled through bacteria-virus interactions, it is difficult to estimate the percentage of primary productivity



contributed by bacterial productivity in this study. A previous study in seagrass meadows of Australia estimated that heterotrophic bacteria (sediments and the water column) accounted for nearly half of total primary production [36]. However, a number of studies have shown that seagrass growth is limited by nutrient availability, and marine seagrass may depend on bacteria associated with it for nutrient availability [37,38]. Hence, this “viral loop” might not only control bacterial production but also supply recycled nutrients for seagrass growth (Figure 4).

In the present study, we found that bacterial growth rates were significantly lower in treatments with reduced viral abundance in the 30 kDa filtered series in adjacent unvegetated water, the presence of active viruses stimulated bacterial growth, as indicated by Figure 2B. Moreover, the DOC pool in the oligotrophic unvegetated waters was significantly influenced by viral lysis of bacteria, based on our results in adjacent unvegetated waters. Aside from this, even small amounts of organic phosphorus, nitrogen, and carbon are likely to increase bacterial growth in the presence of bacterial lysis products, which bacteria could readily utilize [39]. Our data show that bacteria production is controlled by resource availability (bottom-up control) and cycled within the bacteria-virus-DOC loop in adjacent unvegetated water.



**Figure 4.** Image the interaction of seagrass meadows and bacteria. DOC<sub>1</sub>: DOC is released by seagrasses during photosynthesis. DOC<sub>2</sub>: DOC is released extracellularly from phytoplankton photosynthesis. DOC<sub>3</sub>: viral lysis of bacteria release relatively labile dissolved organic carbon into the marine DOC pool.

## 5. Conclusions

In summary, since viral lysis and microzooplankton grazing impact the bacterial community differently in seagrass meadows and adjacent unvegetated areas, it is crucial to assess their importance relative to each other. The current study found similar rates of bacterial mortality caused by viral lysis and grazing in a seagrass environment, whereas grazing mortality predominated in adjacent unvegetated regions. Also, our data indicate that bacteria production is controlled by available resources (bottom-up control) and cycled in the bacteria-virus-DOC loop in unvegetated waters. In the regions that we studied, this shift may have important implications for organic matter fate and cycling.

**Author Contributions:** Conceptualization: A.-Y.T.; methodology: A.-Y.T.; P.W.-Y.C.; M.O.; validation: A.-Y.T.; formal analysis: P.W.-Y.C., A.-Y.T., and M.O.; investigation: A.-Y.T.; P.W.-Y.C.; M.O.; resources: A.-Y.T.; W. C.C.; data curation: A.-Y.T.; R. F. S.; Writing—Original draft preparation: A.-Y.T.; P.W.-Y.C.; Writing—Review and editing: A.-Y.T. and V.M.; funding acquisition: A.-Y.T.; V.M. W.C. C. All authors have read and agreed to the published version of the manuscript.

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