

Article

Not peer-reviewed version

Expanding the Heteroaromatic and 2-Aminosugar Chemical Space Accessible from the Biopolymer Chitin

Thais A Rossa , Jessica C Neville , Seongmin Paul Jun , [Tilo Söhnel](#) , [Jonathan Sperry](#) *

Posted Date: 9 August 2023

doi: [10.20944/preprints202308.0782.v1](https://doi.org/10.20944/preprints202308.0782.v1)

Keywords: chitin; biomass; bio-based



Preprints.org is a free multidiscipline platform providing preprint service that is dedicated to making early versions of research outputs permanently available and citable. Preprints posted at Preprints.org appear in Web of Science, Crossref, Google Scholar, Scilit, Europe PMC.

Copyright: This is an open access article distributed under the Creative Commons Attribution License which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Disclaimer/Publisher's Note: The statements, opinions, and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions, or products referred to in the content.

Article

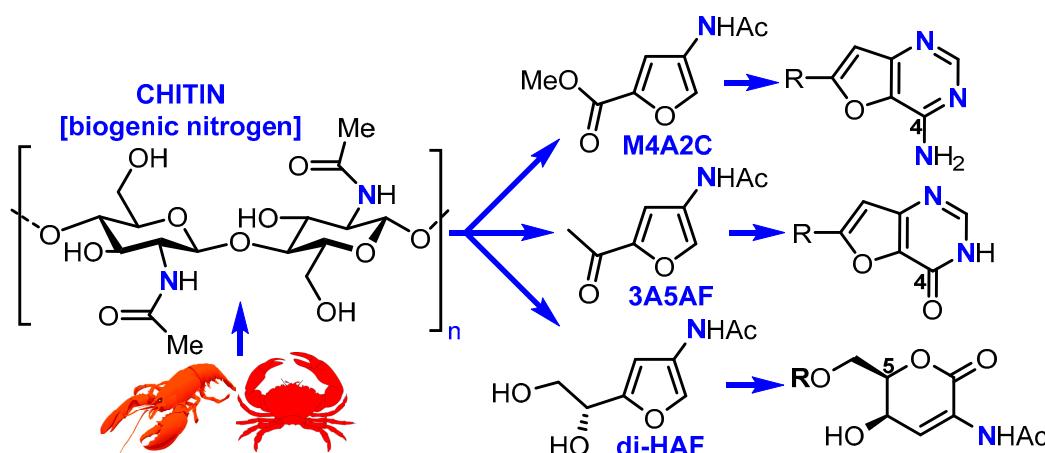
Expanding the Heteroaromatic and 2-Aminosugar Chemical Space Accessible from the Biopolymer Chitin

Thaís A. Rossa, Jessica C. Neville, Seongmin Paul Jun, Tilo Söhnel and Jonathan Sperry *

Centre for Green Chemical Science, University of Auckland, 23 Symonds Street, Auckland 1010, New Zealand; throssa@gmail.com (T.A.R.); jnev410@aucklanduni.ac.nz (J.C.N.); sjun310@aucklanduni.ac.nz (S.P.J.); t.soehnel@auckland.ac.nz (T.S.)

* Correspondence: j.sperry@auckland.ac.nz

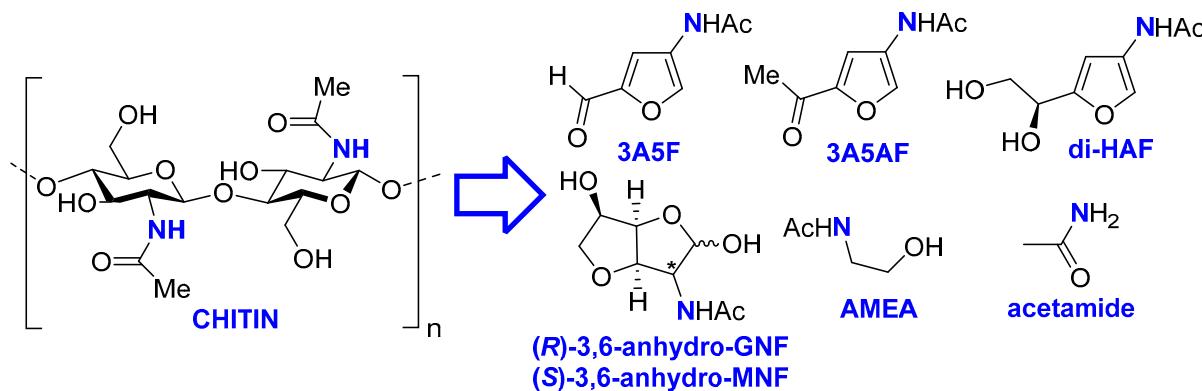
Abstract: Herein we report the expansion of the chemical space available from the chitin, accessible via the biogenic N-platforms 3A5AF, M4A2C and di-HAF. The biologically active heteroaromatics furo[3,2-*d*]pyrimidin-4-one and furo[3,2-*d*]pyrimidin-4-amine can be selectively accessed from 3A5AF and M4A2C, respectively. The chiral pool synthon di-HAF is a viable substrate for the Achmatowicz rearrangement, providing streamlined access to 2-aminosugars possessing a versatile hydroxymethyl group at C5.



Keywords: chitin; biomass; bio-based

1. Introduction

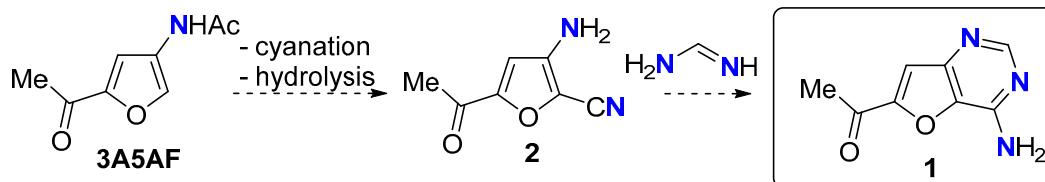
The nitrogen (N) atom in fine and commodity chemicals is derived from the commodity ammonia, a base chemical produced on an enormous scale using the energy-intensive Haber process [1–4]. To reduce the carbon footprint of organonitrogen chemicals, their manufacture can be conducted in a Haber-independent fashion by sourcing nitrogen from the huge quantities of biogenic nitrogen available on earth, with one accessible source being the biopolymer chitin [5,6]. Indeed, several reports describing the valorisation of chitin (or its monomer, *N*-acetyl-D-glucosamine; GlcNAc) into biogenic N-platforms have appeared [7–12], including the functionally rich furans 3-acetamido-5-furyl aldehyde (3A5F) [13], 3-acetamido-5-acetyl furan (3A5AF) [14] and dihydroxyethyl acetamidofuran (di-HAF) [15] (Scheme 1).



Scheme 1. Selected *N*-platforms available from chitin.

Over the past few years, we have showcased the utility of 3A5AF in the Haber-independent synthesis of natural product proximicin A [16], 3-azafurans [17,18], new heteroaromatic scaffolds [19], 2-amino sugars [20] and in a diversity-oriented synthesis (DOS) programme that furnished a number of structurally distinct *N*-heterocycles [21]. We recently demonstrated that the inherent chirality present in chitin can be transferred to the natural product *epi*-leptosphaerin A via the chiral pool di-HAF platform [22]. Other research groups have shown that 3A5AF (and derivatives) [23,24] and di-HAF [25] can serve as dienes in Diels-Alder, while 3A5F shows significant promise as a platform chemical, including as a bioconjugation handle for *N*-cysteine modification [13].

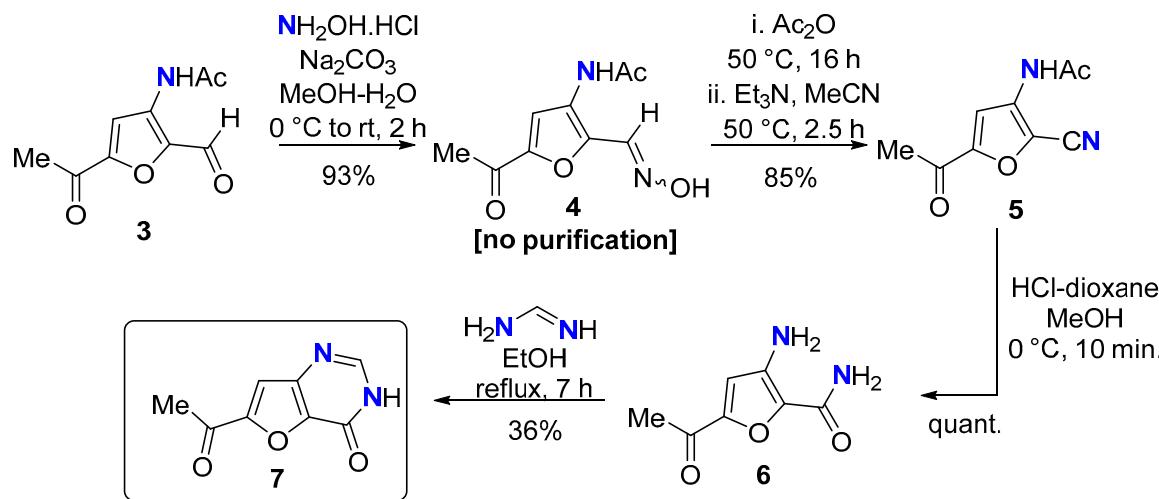
Our ongoing efforts in this area are focused on using the aforementioned *N*-platforms to access chemical intermediates used within the pharmaceutical and/or agrochemical sectors. One target that fulfils this criteria is furo[3,2-*d*]pyrimidin-4-amine **1**, a heteroaromatic scaffold found in the C-nucleoside antibiotic pyrrolosine [26–28] and employed in a multitude of medicinal chemistry programmes against a diverse range of targets, including spleen tyrosine kinases [29], G protein-coupled receptor 119 [30], p110δ PI3 kinase [31], and FYVE-type finger-containing phosphoinositide kinases [32]. We envisaged that the heteroaromatic ring system **1** could be accessible from 3A5AF by cyanation at C2 followed by acetamide hydrolysis to give aminonitrile **2** followed by heteroannulation with formamidine [27,28,33,34] (Scheme 2). Although only one nitrogen atom in **1** is sourced from chitin, our aim was that the other *N*-atoms would be sourced from *N*-compounds present in Nature (q.v.).



Scheme 2. Proposed synthesis of furo[3,2-*d*]pyrimidin-4-amine **1**.

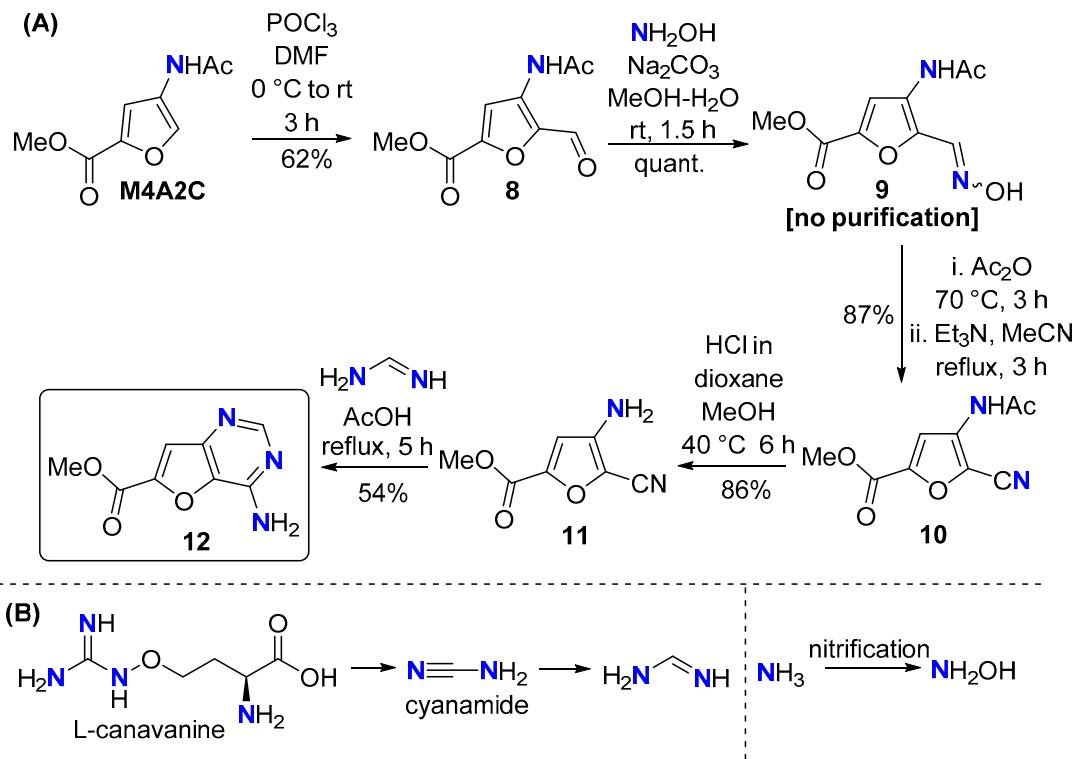
The 3A5AF-derived furfural **3** [18] was converted to the oxime **4** (no purification) that upon dehydration gave 2-cyano-3A5AF **5** in excellent yield over the three steps (Scheme 3). In our experience, acetamide hydrolysis of 3A5AF derivatives does not proceed well under aqueous conditions, often leading to extensive amounts of degradation due to the elevated temperatures required. We have found anhydrous acid-mediated methanolysis to be more reliable given it proceeds at lower temperatures. Methanolysis of the acetamide was successful to give **6**, but NMR spectroscopic analysis revealed hydrolysis of the nitrile had also occurred, promoted by the water produced upon reaction of HCl with MeOH. Despite this unpredicted result, we subjected **6** to heteroannulation with formamidine to give the furo[3,2-*d*]pyrimidin-4-one **7**. While not the originally

intended target, furo[3,2-*d*]pyrimidin-4-ones have found utility in medicinal programmes targeting the kinesin spindle protein [35] and ubiquitin proteasome system 7 (USP7) [36].



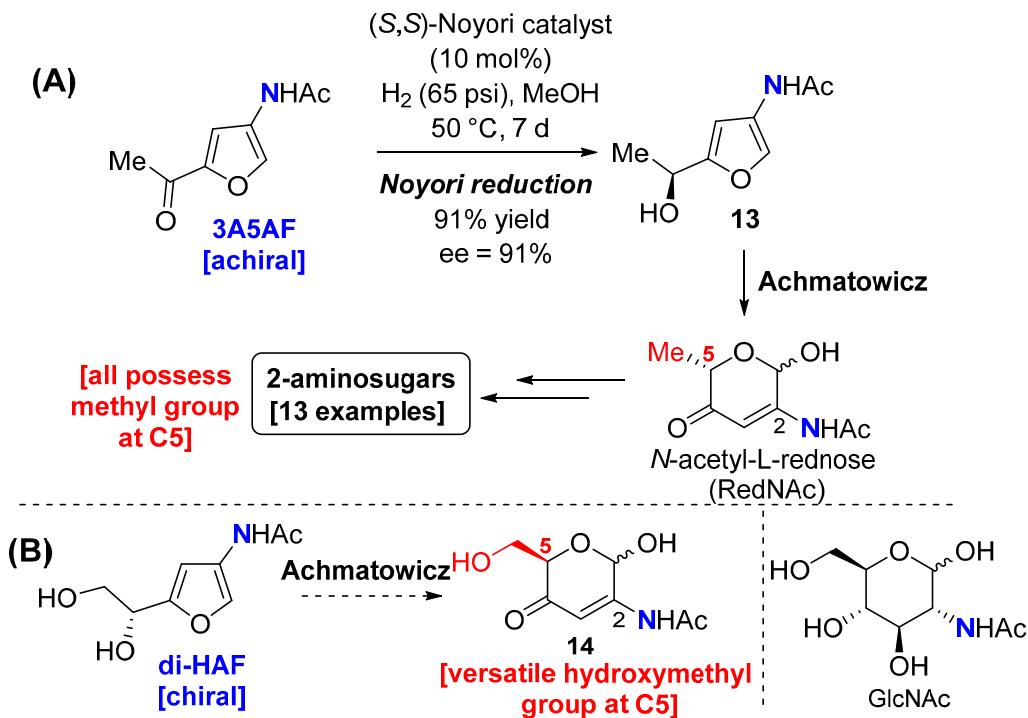
Scheme 3. Synthesis of furo[3,2-*d*]pyrimidin-4-one 7.

The original plan contained weaknesses as we did not anticipate facile hydrolysis of the nitrile in **5**. In pursuit of the original target furo[3,2-*d*]pyrimidin-4-amine scaffold, the chitin-derived amino acid derivative methyl 4-acetamido-2-furanone (M4A2C) [16] was chosen as the substrate, anticipating the less-electron withdrawing ester would reduce the reactivity of the nitrile making it less susceptible to hydrolysis (Scheme 4A). Vilsmeier formylation of M4A2C gave the furfural **8**, which was converted to the 2-cyanofuran **10** (via oxime **9**) in excellent overall yield. The impact of the ester in M4A2C was compelling; the methanolysis selectively cleaved the acetamide in **9**, leaving the cyano group intact. Finally, heteroannulation with formamidine gave the furo[3,2-*d*]pyrimidin-4-amine **12**. In the products **7** and **12**, only one nitrogen atom is sourced from chitin. However, hydroxylamine can be considered biogenic; it is a product of the nitrification process and widely distributed throughout Nature [37]. Formamidine is available from the natural product cyanamide [38,39], itself biosynthesised from the amino acid L-canavanine [40] (Scheme 4B). It is also possible that biogenic formamidine could be produced upon conversion of the urea to thiourea, followed by reductive desulfurisation [41].



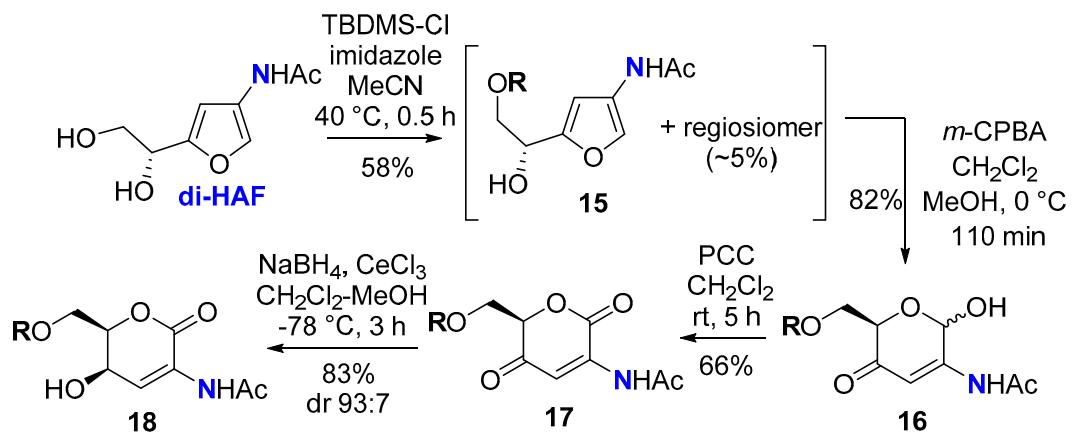
Scheme 4. (A) Synthesis of furo[3,2-d]pyrimidin-4-amine **12** from M4A2C; (B) formamidine and hydroxylamine are *N*-compounds present in Nature.

The synthesis of furo[3,2-*d*]pyrimidin-4-amine **12** and furo[3,2-*d*]pyrimidin-4-one **7** from 3A5AF expands the heteroaromatic chemical space available from chitin. We would also like to report our preliminary results on the oxidative ring expansion of the chiral pool synthon, di-HAF. We previously reported the synthesis of enantioenriched 2-amino sugars from 3A5AF [20], but this method contains some drawbacks (Scheme 5A). The introduction of artificial chirality using a Noyori reduction was cumbersome (50 °C, 1 week reaction time) to give furfuryl alcohol **13** that upon oxidative ring expansion gave *N*-acetyl-L-rednose (RedNAc). From here, several 2-amino sugars were available, but they all contain a methyl group at C5 that restricted modifications at this site. It was anticipated these drawbacks would be overcome if the chitin-derived, chiral pool synthon di-HAF successfully underwent an Achmatowicz reaction; not only would the natural chirality present in chitin be transferred to the product (thus eliminating the need for a Noyori reduction), but the resulting 2-amino sugar **14** would possess a versatile hydroxymethyl group at the C5 position (Scheme 5B). Moreover, **14** is a novel 2-amino sugar scaffold that would be challenging to prepare from *N*-acetyl-D-glucosamine (GlcNAc) [42–45].



Scheme 5. (A) Synthesis of 2-aminosugars from 3A5AF; (B) proposed oxidative ring expansion of di-HAF. Structure of GlcNAc shown for comparison.

To facilitate easy handling of the anticipated products, di-HAF was monosilylated at the primary alcohol to give **15**, alongside small quantities of its regiosiomer that could not be separated by column chromatography (Scheme 6). The Achmatowicz rearrangement proceeded smoothly to give the somewhat unstable 2-aminosugar **16** in good yield. Oxidation of the lactol helped to stabilise the scaffold, affording dione **17**. Luche reduction of the C4-ketone gave **18** with good diastereoselectivity, and the structure of the major *syn*-diastereomer was confirmed by X-ray crystallographic analysis (Figure 1).



Scheme 6. Application of di-HAF in the Achmatowicz rearrangement ($\mathbf{R} = \text{tert-butyldimethylsilyl}$; TBDMS).

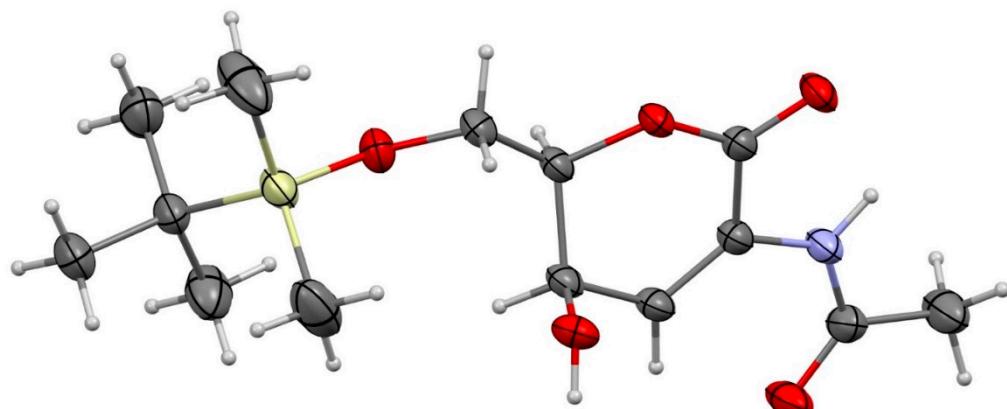


Figure 1. Molecular structure of 2-amino sugar **18** (CCDC 2285900). Atomic displacement parameters are drawn at the 50% probability level.

To conclude, further biogenic N-chemical space has been accessed from the chitin-derived platforms 3A5AF, M4A2C and di-HAF. The high-value heteroaromatic scaffolds furo[3,2-*d*]pyrimidin-4-one **7** and furo[3,2-*d*]pyrimidin-4-amine **12** can be selectively obtained from biogenic N-platforms 3A5AF and M4A2C, respectively. Moreover, the chiral di-HAF scaffold is a viable substrate for the Achmatowicz rearrangement, generating new, enantioenriched 2-amino sugar chemicals possessing chiral centres traceable back to the chitin biopolymer, with all products possessing a versatile 5-hydroxymethyl handle at C5.

2. Experimental

General Information

Commercially available starting materials, reagents, and solvents were used as received unless otherwise noted. In case anhydrous conditions were applied, the reaction was performed under an atmosphere of dry nitrogen in oven-dried (100 °C) glassware and the solvent was dried by passage through a column of activated alumina under nitrogen using an LC Technology solvent purification system. Thin layer chromatography (TLC) was performed using F254 0.2 mm silica plates, followed by visualisation with UV irradiation at 254 nm and 366 nm, and staining with ethanolic vanillin solution. Flash column chromatography was performed using 63–100 µm silica gel. Melting points were determined on a Kofler hot-stage apparatus and are uncorrected. High-resolution mass spectra were recorded on a micrOTOF Q mass spectrometer operated in the positive ion mode. The standard electrospray ion (ESI) source was used to generate the ions. The instrument was operated in the *m/z* 50–1000 range. Infrared (IR) spectra were recorded using a Perkin-Elmer Spectrum One Fourier Transform IR spectrometer with a universal attenuated total reflectance (ATR) attachment installed. Absorption maxima are expressed as wavenumber (cm⁻¹). NMR spectra were recorded at room temperature in CDCl₃, (CD₃)₂SO or (CD₃)₂CO solutions using a spectrometer operating at 400 MHz for ¹H nuclei and 100 MHz for ¹³C nuclei. All chemical shifts are reported in parts per million (ppm) scale and were measured relative to the protium solvent in which the sample was analysed: CDCl₃ (δ 7.26 ppm for ¹H NMR and δ 77.16 ppm for ¹³C NMR), (CD₃)₂SO (δ 2.50 ppm for ¹H NMR and δ 39.52 ppm for ¹³C NMR) or (CD₃)₂CO (δ 2.05 ppm for ¹H NMR and δ 29.84 ppm for ¹³C NMR). Coupling constants, *J*, are reported in Hertz [Hz] where applicable. Multiplicities are reported as “s” (singlet), “d” (doublet), and “br s” (broad singlet). X-ray diffraction measurements of single crystals were performed on a Rigaku Oxford Diffraction XtaLAB-Synergy-S single-crystal diffractometer with a PILATUS 200K hybrid pixel array detector using Cu K α radiation ($\lambda = 1.54184$ Å). The data were processed with the SHELX2018-3 and Olex2 software packages [46–48]. All non-hydrogen atoms were refined anisotropically. Hydrogen atoms were inserted at calculated positions or located directly and refined with a riding model or without restrictions. Mercury 2020.3.1 [49] was used to

visualize the molecular structure. Crystal growth for X-ray crystallographic analysis purposes was achieved using slow evaporation or slow vapour diffusion.

2-Hydroxyimino-3-acetamido-5-acetyl furan (4)

A solution of hydroxylamine hydrochloride (195 mg, 2.8 mmol) and sodium carbonate (148 mg, 1.4 mmol) in water (10 mL) was added dropwise to a solution of 2-formylfuran **3** [18] (488 mg, 2.5 mmol) in methanol (25 mL) at 0 °C. The solution was stirred at room temperature for 2 h, then concentrated *in vacuo*, diluted in ethyl acetate (80 mL), and washed with water (40 mL) and brine (40 mL). The organic layer was dried over Na₂SO₄, filtered, and concentrated *in vacuo*. The *title compound* (489 mg, 2.3 mmol, 93%; *dr* 10:1) which was used in the next step without any purification.

2-Cyano-3-acetamido-5-acetyl furan (5)

A solution of oxime **4** (250 mg, 1.2 mmol) in acetic anhydride (5 mL) was stirred at 50 °C for 16 h. The reaction mixture was concentrated *in vacuo* to give a the acetoxyimino furan which was dissolved in acetonitrile (5 mL). Triethylamine (0.50 mL, 3.6 mmol) was added, and the solution was stirred at 50 °C for 2.5 h. The solution was concentrated *in vacuo* and purified by flash column chromatography on silica gel eluting with ethyl acetate-light petroleum (2:3) to give the *title compound* (196 mg, 1.0 mmol, 85%) as a yellow solid; mp 126.0–127.5 °C; HRMS [ESI, (M + Na)⁺]: calcd. for [C₉H₈N₂O₃ + Na]⁺ 215.0433, found 215.0427 ν_{max} /cm⁻¹ (ATR) 3286, 3236, 3088, 2227, 1690, 1559, 1528, 1366, 1276, 1223, 1141, 928; ¹H NMR (400 MHz, acetone-*d*₆): δ 9.86 (1 H, br s, NH), 7.67 (1 H, s, ArH), 2.50 (3 H, s, Me), 2.17 (3 H, s, Me); ¹³C NMR (100 MHz, acetone-*d*₆): δ 186.7 (C), 169.2 (C), 154.4 (C), 136.8 (C), 111.7 (CH), 111.4 (C), 26.3 (Me), 23.3 (Me), 1 x C not observed.

5-Acetyl-3-amino-2-carboxamido furan (6)

To a solution of 2-cyanofuran **5** (50 mg, 0.26 mmol) in dry methanol (0.6 mL) was added a solution of HCl in dioxane (4 M, 0.6 mL) at 0 °C. The reaction mixture was stirred at 0 °C for 10 min, quenched with saturated NaHCO₃ (15 mL) then extracted with ethyl acetate (3 × 15 mL). The organic layer was dried with anhydrous Na₂SO₄, filtered, and concentrated *in vacuo*. The *title compound* (44 mg, 0.26 mmol, 100%) was obtained as a yellow solid, which was used in the next step without any purification; mp 192.5–193.5 °C; HRMS [ESI, (M + Na)⁺]: calcd. for [C₇H₈N₂O₃ + Na]⁺ 191.0427, found 191.0429; ν_{max} /cm⁻¹ (ATR) 3427, 3352, 3152, 1648, 1610, 1570, 1359, 1319, 1194, 940, 775; ¹H NMR (400 MHz, DMSO-*d*₆): δ 7.30 (2 H, br s, NH₂), 6.86 (1 H, s, ArH), 5.47 (2 H, s, NH₂), 2.43 (3 H, s, Me); ¹³C NMR (100 MHz, DMSO-*d*₆): δ 187.3 (C), 162.0 (C), 149.3 (C), 141.4 (C), 129.0 (C), 110.2 (CH), 25.8 (Me).

6-Acetyl furo[3,2-*d*]pyrimidin-4-one (7)

To a solution of 2-carboxamido furan **6** (20 mg, 0.12 mmol) in ethanol (4 mL) was added formamidine acetate (125 mg, 1.2 mmol), and the reaction mixture was stirred at reflux for 7 h. The solution was concentrated *in vacuo*, diluted in ethyl acetate (20 mL), and washed with saturated Na₂CO₃ (10 mL) and brine (10 mL). The organic layer was dried with anhydrous Na₂SO₄, filtered, and concentrated *in vacuo*. The residue was purified by flash chromatography on silica gel eluting with ethyl acetate to give the *title compound* (7.7 mg, 0.043 mmol, 36%) as a yellow oil; HRMS [ESI, (M + Na)⁺]: calcd. for [C₈H₆N₂O₃ + Na]⁺ 201.0271, found 201.0271; ¹H NMR (400 MHz, DMSO-*d*₆): δ 12.83 (1 H, br s, NH), 8.14 (1 H, s, ArH), 7.84 (1 H, s, ArH), 2.57 (3 H, s, Me); ¹³C NMR (100 MHz, DMSO-*d*₆): δ 187.7 (C), 154.5 (C), 152.1 (C), 147.3 (C), 147.1 (CH), 139.7 (C), 113.6 (CH), 26.5 (Me).

Methyl 4-acetamido-5-formyl furan-2-carboxylate (8)

A solution of phosphoryl chloride (0.28 mL, 3.0 mmol) in dry dimethylformamide (2.0 mL) at 0 °C was added dropwise to a solution of **M4A2C** [16] (366 mg, 2.0 mmol) in dry dimethylformamide (2 mL). The reaction mixture was warmed to room temperature and stirred for 3 h. Water (10 mL) was added at 0 °C and the mixture was stirred for another 10 min. The aqueous phase was neutralized with NaHCO₃ and extracted with ethyl acetate (3 × 30 mL), dried with anhydrous Na₂SO₄, filtered, and concentrated *in vacuo*. The residue was purified by flash chromatography on silica gel eluting with ethyl acetate-light petroleum (1:1) to give the *title compound* (262 mg, 1.2 mmol, 62%) as a yellow

solid; mp 175.0–176.5 °C; HRMS [ESI, (M + Na)⁺]: calcd. for [C₉H₉NO₅ + Na]⁺ 234.0373, found 234.0368; ν_{max} /cm⁻¹ (ATR) 3341, 3197, 1754, 1697, 1667, 1590, 1434, 1329, 1219, 1199, 984, 792, 769, 741; ¹H NMR (400 MHz, acetone-*d*₆) δ 9.87 (1 H, s, CH), 9.52 (1 H, br s, NH), 7.93 (1 H, s, ArH), 3.93 (3 H, s, Me), 2.24 (3 H, s, Me); ¹³C NMR (100 MHz, acetone-*d*₆) δ 181.3 (C), 169.8 (C), 158.8 (C), 147.2 (C), 140.7 (C), 135.0 (C), 113.3 (CH), 52.9 (Me), 23.9 (Me).

Methyl 4-acetamido-5-[(hydroxyimino)methyl]furan-2-carboxylate (9)

A solution of hydroxylamine hydrochloride (83 mg, 1.2 mmol) and sodium carbonate (64 mg, 0.6 mmol) in water (4 mL) was added dropwise to a solution of **8** (211 mg, 1.0 mmol) in methanol (8 mL) at room temperature. The reaction mixture was stirred at room temperature 1.5 h. The methanol was removed *in vacuo* and reaction mixture diluted with ethyl acetate (40 mL), washed with water (20 mL) and brine (20 mL). The aqueous phase was extracted with ethyl acetate (40 mL) and the combined organic phases dried with anhydrous Na₂SO₄, filtered, and concentrated *in vacuo*. The *title compound* (224 mg, 0.99 mmol, 99%, *dr* 10:1) was obtained as a yellow solid which was used in the next step without further purification.

Methyl 4-acetamido-5-cyanofuran-2-carboxylate (10)

A solution of the oxime **9** (226 mg, 1 mmol) in acetic anhydride (3 mL) was stirred at 70 °C for 3 h. After formation of the acetoxyiminofuran, the acetic anhydride was concentrated *in vacuo*. To the residue was added acetonitrile (4 mL) and triethylamine (0.3 mL, 2 mmol) and the resulting solution stirred at reflux for 3 h. The reaction mixture was concentrated under reduced pressure and purified by flash chromatography on silica gel eluting with ethyl acetate-light petroleum (1:1) to give the *title compound* (181 mg, 0.87 mmol, 87%) as a yellow solid; mp 219.5–220.5 °C; HRMS [ESI, (M + Na)⁺]: calcd. for [C₉H₈N₂O₄ + Na]⁺ 231.0376, found 231.0376; ν_{max} /cm⁻¹ (ATR) 3285, 3087, 2229, 1737, 1696, 1533, 1320, 1226, 1149, 985, 815, 769; ¹H NMR (400 MHz, DMSO-*d*₆): δ 10.82 (1 H, s, NH), 7.41 (1 H, s, ArH), 3.86 (3 H, s, Me), 2.10 (3 H, s, Me); ¹³C NMR (100 MHz, DMSO-*d*₆): δ 168.7 (C), 156.9 (C), 145.8 (C), 134.6 (C), 116.1 (C), 112.2 (CH), 111.0 (C), 52.7 (Me), 22.9 (Me).

Methyl 4-amino-5-cyanofuran-2-carboxylate (11)

To a solution of **10** (62 mg, 0.30 mmol) in dry methanol (3 mL) at 0 °C was added hydrochloric acid (4 M in dioxane, 0.2 mL). The solution was stirred at 40 °C for 6 h. The reaction mixture was neutralised with solid NaHCO₃ at 0 °C, followed by addition of ethyl acetate (15 mL) and water (15 mL). The layers were separated, and the aqueous phase extracted with ethyl acetate (2 × 15 mL). The combined organic phases were dried over Na₂SO₄, filtered, and concentrated *in vacuo*. The crude mixture was purified by flash chromatography on silica gel eluting with ethyl acetate-light petroleum (1:1) to give the *title compound* (43 mg, 0.26 mmol, 86%) as a yellow solid; mp 165.0–166.0 °C; HRMS [ESI, (M + Na)⁺]: calcd. for [C₇H₆N₂O₃ + Na]⁺ 189.0271, found 189.0270; ν_{max} /cm⁻¹ (ATR) 3430, 3343, 3232, 2210, 1720, 1607, 1329, 1224, 1176, 765; ¹H NMR (400 MHz, DMSO-*d*₆): δ 6.85 (1 H, s, ArH), 6.17 (2 H, br s, NH₂), 3.81 (3 H, s, Me); ¹³C NMR (100 MHz, DMSO-*d*₆): δ 157.2 (C), 147.0 (C), 145.8 (C), 112.7 (C), 110.8 (CH), 109.1 (C), 52.4 (Me).

Methyl 4-aminofuro[3,2-*d*]pyrimidine-6-carboxylate (12)

To a solution of 4-amino-5-cyanofuran **11** (37 mg, 0.22 mmol) in acetic acid (0.2 mL) was added formamidine acetate (70 mg, 0.69 mmol) and the reaction mixture was stirred at reflux for 5 h. The solution was diluted with ethyl acetate (20 mL), washed with brine (10 mL) and saturated NaHCO₃ (10 mL). The aqueous layer was extracted with ethyl acetate (2 × 10 mL) and the combined organic layers dried over Na₂SO₄, filtered, and concentrated *in vacuo*. The residue was purified by flash chromatography on silica gel eluting with ethyl acetate-light petroleum (7:3) to give the *title compound* (23 mg, 0.12 mmol, 54%) as a colourless solid; mp 218 °C (dec); HRMS [ESI, (M + H)⁺]: calcd. for [C₈H₇N₃O₃ + H]⁺ 194.0560, found 194.0564; ν_{max} /cm⁻¹ (ATR) 3434, 3284, 3008, 2960, 1732, 1660, 1301, 1203, 1094, 969, 761; ¹H NMR (400 MHz, DMSO-*d*₆): δ 8.31 (1 H, s, ArH), 7.71 (2 H, br s, NH₂), 7.67 (1 H, s, ArH), 3.92 (3 H, s, Me); ¹³C NMR (100 MHz, DMSO-*d*₆): δ 158.6 (C), 154.5 (CH), 150.4 (C), 148.2 (C), 147.3 (C), 135.4 (C), 113.6 (CH), 52.6 (Me).

(*R*)-*N*-(5-(2-((*tert*-Butyldimethylsilyl)oxy)-1-hydroxyethyl)furan-3-yl)acetamide (15)

To a solution of di-HAF [15] (55 mg, 0.297 mmol) in acetonitrile (3.5 mL) was added imidazole (80.9 mg, 1.19 mmol, 4 eq.) and *tert*-butyldimethylsilyl chloride (42.5 mg, 0.282 mmol, 0.95 eq.) at room temperature. The reaction mixture was heated to 40 °C for 30 min, then concentrated *in vacuo*. The residue was purified by flash chromatography eluting with light petroleum/acetone (5:1) to give the *title compound* (51 mg, 0.170 mmol, 58%) embedded within an inseparable mixture alongside its regioisomer (95:5); m.p. 58.6 – 59.8 °C; HRMS (ESI) *m/z* [M + Na]⁺, calcd for C₁₄H₂₅NO₄SiNa 322.1445, found 322.1435; $\nu_{\text{max}}/\text{cm}^{-1}$ (ATR) 3279, 2929, 2857, 1659, 1569, 1463, 1378, 1254, 1119, 1071, 837, 779; ¹H NMR (400 MHz, (CD₃)₂CO): δ 9.18 (1 H, br s, NH), 7.89 (1 H, s, CH), 6.28 (1 H, s, CH), 4.60 (1 H, q, *J* 5.5, CH), 4.35 (1 H, d, *J* 5.5, OH), 3.83 (2 H, dq, *J* 5.5, 6.4, CH₂), 2.02 (3 H, s, Ac), 0.87 (9 H, s, (Me)₃), 0.05 (3 H, s, Me), 0.04 (3 H, s, Me); ¹³C NMR (100 MHz, (CD₃)₂CO): δ 162.8 (C), 155.0 (C), 131.2 (CH), 126.7 (C), 102.3 (CH), 69.3 (CH), 67.0 (CH₂), 26.2 (3 x Me), 23.0 (COMe), 18.8 (C), -5.2 (Me), -5.3 (Me)

N-((6*R*)-6-(((*tert*-Butyldimethylsilyl)oxy)methyl)-2-hydroxy-5-oxo-5,6-dihydro-2*H*-pyran-3-yl)acetamide (16)

To a solution of **15** (150 mg, 0.501 mmol) in dichloromethane-methanol (12 mL: 3 mL) at 0 °C was added *m*-CPBA (77%, 123.5 mg, 0.551 mmol, 1.1 eq.) and the reaction mixture was stirred at 0 °C for 2 h. Na₂CO₃ (15 mL) was added to the solution and stirred for 30 min. A saturated solution of NaHCO₃ (15 mL) was added to the reaction mixture and stirred for another 30 min. The mixture was extracted with ethyl acetate (200 mL), washed with cold Na₂CO₃-NaHCO₃ (1:1, 4 x 50 mL), cold brine (2 x 30 mL), dried (Na₂SO₄), filtered and concentrated *in vacuo*. The residue was purified by flash column chromatography on silica gel eluting with light petroleum-ethyl acetate (1:1) to afford the *title compound* (130 mg, 0.412 mmol, 82%, dr 1:0.6) as a colourless solid; m.p. 145.8 – 146.8 °C; HRMS (ESI) *m/z* [M + Na]⁺, calcd for C₁₄H₂₅NO₅SiNa 338.1394, found 338.1385; $\nu_{\text{max}}/\text{cm}^{-1}$ (ATR) 3290, 2930, 2858, 1719, 1642, 1628, 1518, 1472, 1369, 1228, 1205, 1121, 1043, 958, 872, 825, 780, 729, 708 ¹H NMR (400 MHz, (CD₃)₂CO) Major diastereomer: δ 9.21 (1 H, br s, NH), 6.79 (1 H, s, CH), 5.60 (1 H, s, CH), 4.41 (1 H, q, *J* 2.6, CH), 4.00 (2 H, dq, *J* 4.5, 2.5 CH₂), 2.13 (3 H, s, Ac), 0.86 (9 H, s, (Me)₃), 0.05 (3 H, s, Me), 0.03 (3 H, s, Me); ¹³C NMR (100 MHz, (CD₃)₂CO) 195.2 (C), 171.0 (C), 153.5 (C), 108.3 (CH), 89.2 (CH), 76.0 (CH), 64.1 (CH₂), 26.2 ((Me)₃), 24.6 (Ac), 18.90 (COMe), -5.1 (SiMe), -5.2 (SiMe); Minor diastereomer: (400 MHz, (CD₃)₂CO) δ 8.93 (1 H, br s, NH), 6.91 (1 H, s, CH), 5.48 (1 H, s, CH), 4.21 (1 H, dd, *J* 2.8, CH), 4.00 (2 H, dq, *J* 4.4, 2.7, CH₂), 2.17 (3 H, s, Ac), 0.87 (9 H, s, (Me)₃), 0.08 (3 H, s, Me), 0.05 (3 H, s, Me); ¹³C NMR (100 MHz, (CD₃)₂CO): δ 194.8 (C), 171.1 (C), 154.3 (C), 109.0 (CH), 89.6 (CH), 80.6 (CH), 64.8 (CH₂), 26.1 ((Me)₃), 24.7 (Ac), 18.87 (COMe), -5.29 (SiMe), -5.34 (SiMe)

(R)-N-(6-(((*tert*-Butyldimethylsilyl)oxy)methyl)-2,5-dioxo-5,6-dihydro-2*H*-pyran-3-yl)acetamide (17)

To a stirring suspension of Celite (1 g) and pyridinium chlorochromate (502 mg, 2.33 mmol, 2 eq.) in dry dichloromethane (20 mL) at 0 °C was added **16** (367 mg, 1.16 mmol) in one portion. The mixture was warmed to room temperature and stirred for 5 hours. The mixture was filtered through a plug of Celite and concentrated *in vacuo*. The residue was purified by flash column chromatography on silica gel eluting with light petroleum-ethyl acetate (2:1) to give the *title compound* as a colourless solid (241 mg, 0.770 mmol, 66%), m.p. 112 – 113.6 °C; $[\alpha]_D^{23} +56.0$ (*c* 0.1, CH₂Cl₂); HRMS (ESI) *m/z* [M + Na]⁺, calcd for C₁₄H₂₃NO₅SiNa 336.1238, found 336.1240; $\nu_{\text{max}}/\text{cm}^{-1}$ (ATR) 3302, 2957, 2930, 2884, 2858, 1708, 1662, 1626, 1485, 1378, 1332, 1299, 1250, 1209, 1109, 1075, 1022, 997, 918, 879, 836, 778, 733; ¹H NMR (400 MHz, CDCl₃): δ 8.30 (1 H, br s, NH), 7.63 (1 H, s, CH), 4.88 (1 H, dd, *J* 1.7, CH), 4.03 (2 H, dq, CH₂), 2.24 (3 H, s, Ac), 0.78 (9 H, s, C(Me)₃), 0.00 (3 H, s, Si-Me), -0.02 (3 H, s, Si-Me); ¹³C NMR (100 MHz, CDCl₃): δ 192.5 (C), 169.4 (C), 160.2 (C), 138.3 (C), 117.2 (CH), 85.1 (CH), 64.7 (CH₂), 25.6 (COMe), 25.0 (Ac), 18.1 (C), -5.6 (Si-Me), -5.7 (Si-Me).

N-((5*R*,6*R*)-6-(((*tert*-Butyldimethylsilyl)oxy)methyl)-5-hydroxy-2-oxo-5,6-dihydro-2*H*-pyran-3-yl)acetamide (18)

A solution of **17** (120 mg, 0.382 mmol) in dichloromethane-methanol (1.8:2.6 mL) was cooled to -78 °C and cerium trichloride (4.7 mg, 0.0191, 5 mol%) was added, followed by sodium borohydride (22 mg, 0.574 mmol, 1.5 eq.). The mixture was stirred at -78 °C for 3 h, then extracted with ethyl

acetate (80 mL). The organic layer was washed with water (2 x 20 mL), brine (20 mL), dried (Na_2SO_4), filtered and concentrated *in vacuo*. The residue was purified by flash column chromatography on silica gel eluting with light petroleum-ethyl acetate (3:1) to give the *title compound* as a colourless solid (dr 93:7) (100 mg, 0.317 mmol, 83%), m.p. 144.8 – 146.0 °C; HRMS (ESI) m/z [M + Na]⁺, calcd for $\text{C}_{14}\text{H}_{25}\text{NO}_5\text{SiNa}$ 338.1394, found 338.1394; $\nu_{\text{max}}/\text{cm}^{-1}$ (ATR) 3322, 2955, 2930, 2858, 1725, 1671, 1552, 1472, 1384, 1340, 1242, 1191, 1123, 1087, 1006, 974, 941, 895, 835, 776, 745, 668; ^1H NMR (400 MHz, CDCl_3) Major diastereomer: δ 7.83 (1 H, s, NH), 7.65 (1 H, d, J 6.8, CH), 4.54 (1 H, dd, J 2.5, CH), 4.41 (1 H, dq, J 2.5, 1.1, CH), 4.03 (2 H, dq, J 6.5, 5.3, CH_2), 2.14 (3 H, s, Ac), 0.91 (9 H, s, $\text{C}(\text{Me})_3$), 0.121 (3 H, s, Si-Me), 0.117 (3 H, s, Si-Me), OH not observed; ^{13}C NMR (100 MHz, CDCl_3): δ 169.4 (C), 162.1 (C), 127.1 (C), 121.2 (CH), 79.8 (CH), 61.9 (CH_2), 61.8 (CH), 25.9 (3 x Me), 24.7 (Ac), 18.3 ($\text{C}(\text{Me})_3$), -5.3 (Si-Me), -5.4 (Si-Me).

Supplementary Materials: The following supporting information can be downloaded at the website of this paper posted on Preprints.org. Experimental procedures and NMR spectra for all novel compounds.

Acknowledgments: We thank the University of Auckland for the award of a Doctoral Scholarship (J. C. N.). We are grateful to Timothy Christopher for collecting the single crystal X-ray diffraction data.

References

1. Kyriakou, V.; Garagounis, I.; Vourros, A.; Vasileiou, E.; Stoukides, M. An Electrochemical Haber-Bosch Process. *Joule* **2020**, *4*, 142–158. <https://doi.org/10.1016/j.joule.2019.10.006>.
2. Song, X.; Basheer, C.; Zare, R.N. Making Ammonia from Nitrogen and Water Microdroplets. *Proc. Natl. Acad. Sci.* **2023**, *120*, e2301206120. <https://doi.org/10.1073/pnas.2301206120>.
3. Soloveichik, G. Electrochemical Synthesis of Ammonia as a Potential Alternative to the Haber–Bosch Process. *Nat. Catal.* **2019**, *2*, 377–380. <https://doi.org/10.1038/s41929-019-0280-0>.
4. Wang, M.; A. Khan, M.; Mohsin, I.; Wicks, J.; H. Ip, A.; Z. Sumon, K.; Dinh, C.-T.; H. Sargent, E.; D. Gates, I.; Golam Kibria, M. Can Sustainable Ammonia Synthesis Pathways Compete with Fossil-Fuel Based Haber–Bosch Processes? *Energy Environ. Sci.* **2021**, *14*, 2535–2548. <https://doi.org/10.1039/D0EE03808C>.
5. Yan, N.; Chen, X. Sustainability: Don't Waste Seafood Waste. *Nature* **2015**, *524*, 155–157. <https://doi.org/10.1038/524155a>.
6. Hülsey, M.J.; Yang, H.; Yan, N. Sustainable Routes for the Synthesis of Renewable Heteroatom-Containing Chemicals. *ACS Sustain. Chem. Eng.* **2018**, *6*, 5694–5707. <https://doi.org/10.1021/acssuschemeng.8b00612>.
7. Dai, J.; Li, F.; Fu, X. Towards Shell Biorefinery: Advances in Chemical-Catalytic Conversion of Chitin Biomass to Organonitrogen Chemicals. *ChemSusChem* **2020**, *13*, 6498–6508. <https://doi.org/10.1002/cssc.202001955>.
8. Osada, M.; Kikuta, K.; Yoshida, K.; Totani, K.; Ogata, M.; Usui, T. Non-Catalytic Synthesis of Chromogen I and III from N-Acetyl-D-Glucosamine in High-Temperature Water. *Green Chem.* **2013**, *15*, 2960–2966. <https://doi.org/10.1039/C3GC41161C>.
9. Techikawara, K.; Kobayashi, H.; Fukuoka, A. Conversion of N-Acetylglucosamine to Protected Amino Acid over Ru/C Catalyst. *ACS Sustain. Chem. Eng.* **2018**, *6*, 12411–12418. <https://doi.org/10.1021/acssuschemeng.8b02951>.
10. Bobbink, F.D.; Zhang, J.; Pierson, Y.; Chen, X.; Yan, N. Conversion of Chitin Derived N-Acetyl-D-Glucosamine (NAG) into Polyols over Transition Metal Catalysts and Hydrogen in Water. *Green Chem.* **2015**, *17*, 1024–1031. <https://doi.org/10.1039/C4GC01631A>.
11. Nikahd, M.; Mikusek, J.; Yu, L.-J.; Coote, M.L.; Banwell, M.G.; Ma, C.; Gardiner, M.G. Exploiting Chitin as a Source of Biologically Fixed Nitrogen: Formation and Full Characterization of Small-Molecule Hetero- and Carbocyclic Pyrolysis Products. *J. Org. Chem.* **2020**, *85*, 4583–4593. <https://doi.org/10.1021/acs.joc.9b03438>.
12. Banwell, M.G.; Pollard, B.; Liu, X.; Connal, L.A. Exploiting Nature's Most Abundant Polymers: Developing New Pathways for the Conversion of Cellulose, Hemicellulose, Lignin and Chitin into Platform Molecules (and Beyond). *Chem. – Asian J.* **2021**, *16*, 604–620. <https://doi.org/10.1002/asia.202001451>.
13. Gomes, R.F.A.; Gonçalves, B.M.F.; Andrade, K.H.S.; Sousa, B.B.; Maulide, N.; Bernardes, G.J.L.; Afonso, C.A.M. Unlocking the Potential of Bio-Based Nitrogen-Rich Furanic Platforms as Biomass Synthons. *Angew. Chem. Int. Ed.* **2023**, *62*, e202304449. <https://doi.org/10.1002/anie.202304449>.
14. Padovan, D.; Kobayashi, H.; Fukuoka, A. Facile Preparation of 3-Acetamido-5-Acetyl furan from N-Acetyl-d-Glucosamine by Using Commercially Available Aluminum Salts. *ChemSusChem* **2020**, *13*, 3594–3598. <https://doi.org/10.1002/cssc.202001068>.
15. Loo, C.H.M. van der; G. Borst, M.L.; Pouwer, K.; J. Minnaard, A. The Dehydration of N -Acetylglucosamine (GlcNAc) to Enantiopure Dihydroxyethyl Acetamidofuran (Di-HAF). *Org. Biomol. Chem.* **2021**, *19*, 10105–10111. <https://doi.org/10.1039/D1OB02004H>.

16. Sadiq, A.D.; Chen, X.; Yan, N.; Sperry, J. Towards the Shell Biorefinery: Sustainable Synthesis of the Anticancer Alkaloid Proximicin A from Chitin. *ChemSusChem* **2018**, *11*, 532–535. <https://doi.org/10.1002/cssc.201702356>.
17. Pham, T.T.; Lindsay, A.C.; Chen, X.; Gözaydin, G.; Yan, N.; Sperry, J. Transferring the Biorenewable Nitrogen Present in Chitin to Several N-Functional Groups. *Sustain. Chem. Pharm.* **2019**, *13*, 100143. <https://doi.org/10.1016/j.scp.2019.100143>.
18. Pham, T.T.; Lindsay, A.C.; Kim, S.-W.; Persello, L.; Chen, X.; Yan, N.; Sperry, J. Two-Step Preparation of Diverse 3-Amidofurans from Chitin. *ChemistrySelect* **2019**, *4*, 10097–10099. <https://doi.org/10.1002/slct.201902765>.
19. Pham, T.T.; Chen, X.; Yan, N.; Sperry, J. A Novel Dihydrodifuroypyridine Scaffold Derived from Ketones and the Chitin-Derived Heterocycle 3-Acetamido-5-Acetyl furan. *Monatsh. Chem.* **2018**, *149*, 857–861. <https://doi.org/10.1007/s00706-017-2112-8>.
20. Pham, T.T.; Gözaydin, G.; Söhnle, T.; Yan, N.; Sperry, J. Oxidative Ring-Expansion of a Chitin-Derived Platform Enables Access to Unexplored 2-Amino Sugar Chemical Space. *Eur. J. Org. Chem.* **2019**, *2019*, 1355–1360. <https://doi.org/10.1002/ejoc.201801683>.
21. Pham, T.T.; Chen, X.; Söhnle, T.; Yan, N.; Sperry, J. Haber-Independent, Diversity-Oriented Synthesis of Nitrogen Compounds from Biorenewable Chitin. *Green Chem.* **2020**, *22*, 1978–1984. <https://doi.org/10.1039/D0GC00208A>.
22. Neville, J.C.; Lau, M.Y.; Söhnle, T.; Sperry, J. Haber-Independent, Asymmetric Synthesis of the Marine Alkaloid Epi-Leptosphaerin from a Chitin-Derived Chiral Pool Synthon. *Org. Biomol. Chem.* **2022**, *20*, 6562–6565. <https://doi.org/10.1039/D2OB01251K>.
23. Pereira, J.G.; Ravasco, J.M.J.M.; Vale, J.R.; Queda, F.; Gomes, R.F.A. A Direct Diels–Alder Reaction of Chitin-Derived 3-Acetamido-5-Acetyl furan. *Green Chem.* **2022**, *24*, 7131–7136. <https://doi.org/10.1039/D2GC00253A>.
24. Santos, C.S.; Rodini Mattioli, R.; Soares Baptista, J.; Menezes da Silva, V.H.; Browne, D.L.; Pastre, J.C. Nitrogenated Aromatics from Chitin. *Green Chem.* **2023**, *25*, 5059–5067. <https://doi.org/10.1039/D3GC00272A>.
25. van der Loo, C.H.M.; Schim van der Loeff, R.; Martín, A.; Gomez-Sal, P.; Borst, M.L.G.; Pouwer, K.; Minnaard, A.J. π -Facial Selectivity in the Diels–Alder Reaction of Glucosamine-Based Chiral Furans and Maleimides. *Org. Biomol. Chem.* **2023**, *21*, 1888–1894. <https://doi.org/10.1039/D2OB02221D>.
26. Ikegami, S.; Isomura, H.; Tsuchimori, N.; Hamada, K.; KOBAYASHI, H.; Kojima, Y.; Osano, Y.T.; Kumazawa, S.; Matsuzaki, T. Crystal Structure of an Inhibitor of Starfish Embryonic Development, 4-Oxo-7-(β -D-Ribofuranosyl)-3H-Furo[3, 2-d]Pyrimidine: Revision of Pyrrolosine Structure. *Anal. Sci.* **1992**, *8*, 897–898. <https://doi.org/10.2116/analsci.8.897>.
27. Bhattacharya, B.K.; Lim, M.-I.; Otter, B.A.; Klein, R.S. Synthesis of Furo[3,2-d]Pyrimidine Nucleosides: A Novel c-Nucleoside Isostere of Adenosine. *Tetrahedron Lett.* **1986**, *27*, 815–818. [https://doi.org/10.1016/S0040-4039\(00\)84108-8](https://doi.org/10.1016/S0040-4039(00)84108-8).
28. Bhattacharya, B.K.; Otter, B.A.; Berens, R.L.; Klein, R.S. Studies on the Synthesis of Furo[3,2-d]Pyrimidine C-Nucleosides: New Inosine Analogues with Antiprotozoan Activity. *Nucleosides Nucleotides* **1990**, *9*, 1021–1043. <https://doi.org/10.1080/07328319008046060>.

29. Hoemann, M.; Wilson, N.; Argiriadi, M.; Banach, D.; Burchat, A.; Calderwood, D.; Clapham, B.; Cox, P.; Duignan, D.B.; Konopacki, D.; et al. Synthesis and Optimization of Furano[3,2-d]Pyrimidines as Selective Spleen Tyrosine Kinase (Syk) Inhibitors. *Bioorganic Med. Chem. Lett.* **2016**, *26*, 5562–5567. <https://doi.org/10.1016/j.bmcl.2016.09.077>.

30. Koshizawa, T.; Morimoto, T.; Watanabe, G.; Watanabe, T.; Yamasaki, N.; Sawada, Y.; Fukuda, T.; Okuda, A.; Shibuya, K.; Ohgiya, T. Optimization of a Novel Series of Potent and Orally Bioavailable GPR119 Agonists. *Bioorganic Med. Chem. Lett.* **2017**, *27*, 3249–3253. <https://doi.org/10.1016/j.bmcl.2017.06.034>.

31. Hancox, T.C.; Pegg, N.A.; Nadin, A.J.; Price, S. Pharmaceutical Compounds.

32. Rhodes, J.; Mighdoll, M.; Choi, I.Y.; Kopec, B. Methods and Treatment of Viral Infection with Substituted Furo-Pyrimidines.

33. Kim, S.; Hong, J.H. Synthesis of Novel 4'-Trifluoromethyl-5'-Norcarbocyclic C-Nucleoside Phosphonic Acids as Potent Anti-Leukemic Agents. *Nucleosides Nucleotides Nucleic Acids* **2015**, *34*, 848–865. <https://doi.org/10.1080/15257770.2015.1079327>.

34. Butora, G.; Olsen, D.B.; Carroll, S.S.; McMasters, D.R.; Schmitt, C.; Leone, J.F.; Stahlhut, M.; Burlein, C.; MacCoss, M. Synthesis and HCV Inhibitory Properties of 9-Deaza- and 7,9-Dideaza-7-Oxa-2'-C-Methyladenosine. *Bioorganic Med. Chem.* **2007**, *15*, 5219–5229. <https://doi.org/10.1016/j.bmc.2007.05.020>.

35. Theoclitou, M.-E.; Aquila, B.; Block, M.H.; Brassil, P.J.; Castriotta, L.; Code, E.; Collins, M.P.; Davies, A.M.; Deegan, T.; Ezhuthachan, J.; et al. Discovery of (+)-N-(3-Aminopropyl)-N-[1-(5-Benzyl-3-Methyl-4-Oxo-[1,2]Thiazolo[5,4-d]Pyrimidin-6-Yl)-2-Methylpropyl]-4-Methylbenzamide (AZD4877), a Kinesin Spindle Protein Inhibitor and Potential Anticancer Agent. *J. Med. Chem.* **2011**, *54*, 6734–6750. <https://doi.org/10.1021/jm200629m>.

36. O'Dowd, C.R.; Helm, M.D.; Rountree, J.S.S.; Flasz, J.T.; Arkoudis, E.; Miel, H.; Hewitt, P.R.; Jordan, L.; Barker, O.; Hughes, C.; et al. Identification and Structure-Guided Development of Pyrimidinone Based USP7 Inhibitors. *ACS Med. Chem. Lett.* **2018**, *9*, 238–243. <https://doi.org/10.1021/acsmmedchemlett.7b00512>.

37. Zhang, D.; Li, W.; Huang, X.; Qin, W.; Liu, M. Removal of Ammonium in Surface Water at Low Temperature by a Newly Isolated Microbacterium Sp. Strain SFA13. *Bioresour. Technol.* **2013**, *137*, 147–152. <https://doi.org/10.1016/j.biortech.2013.03.094>.

38. Odo, K.; Ichikawa, E.; Shirai, K.; Sugino, K. Notes - A New Method for the Preparation of Formamidine. *J. Org. Chem.* **1957**, *22*, 1715–1715. <https://doi.org/10.1021/jo01363a620>.

39. Kamo, T.; Hiradate, S.; Fujii, Y. First Isolation of Natural Cyanamide as a Possible Allelochemical from Hairy Vetch Vicia Villosa. *J. Chem. Ecol.* **2003**, *29*, 275–283. <https://doi.org/10.1023/A:1022621709486>.

40. Kamo, T.; Sakurai, S.; Yamanashi, T.; Todoroki, Y. Cyanamide Is Biosynthesized from L-Canavanine in Plants. *Sci. Rep.* **2015**, *5*, 10527. <https://doi.org/10.1038/srep10527>.

41. Brown, D.J. A New Synthesis of Formamidine. *J. Appl. Chem.* **1952**, *2*, 202–203. <https://doi.org/10.1002/jctb.5010020408>.

42. Pfrengle, F.; Reissig, H.-U. Amino Sugars and Their Mimetics via 1,2-Oxazines. *Chem. Soc. Rev.* **2010**, *39*, 549–557. <https://doi.org/10.1039/B914356D>.

43. Emmadi, M.; Kulkarni, S.S. Recent Advances in Synthesis of Bacterial Rare Sugar Building Blocks and Their Applications. *Nat. Prod. Rep.* **2014**, *31*, 870–879. <https://doi.org/10.1039/C4NP00003J>.

44. Skarbek, K.; Milewska, M.J. Biosynthetic and Synthetic Access to Amino Sugars. *Carbohydr. Res.* **2016**, *434*, 44–71. <https://doi.org/10.1016/j.carres.2016.08.005>.

45. Yang, J.; Xie, D.; Ma, X. Recent Advances in Chemical Synthesis of Amino Sugars. *Molecules* **2023**, *28*. <https://doi.org/10.3390/molecules28124724>.

46. Sheldrick, G.M. A Short History of SHELX. *Acta Crystallogr. Sect. A* **2008**, *64*, 112–122. <https://doi.org/10.1107/S0108767307043930>.

47. Bourhis, L.J.; Dolomanov, O.V.; Gildea, R.J.; Howard, J.A.K.; Puschmann, H. The Anatomy of a Comprehensive Constrained, Restrained Refinement Program for the Modern Computing Environment – Olex2 Dissected. *Acta Crystallogr. Sect. A* **2015**, *71*, 59–75. <https://doi.org/10.1107/S2053273314022207>.

48. Dolomanov, O.V.; Bourhis, L.J.; Gildea, R.J.; Howard, J.A.K.; Puschmann, H. OLEX2: A Complete Structure Solution, Refinement and Analysis Program. *J. Appl. Crystallogr.* **2009**, *42*, 339–341. <https://doi.org/10.1107/S0021889808042726>.

49. Macrae, C.F.; Sovago, I.; Cottrell, S.J.; Galek, P.T.A.; McCabe, P.; Pidcock, E.; Platings, M.; Shields, G.P.; Stevens, J.S.; Towler, M.; et al. Mercury 4.0: From Visualization to Analysis, Design and Prediction. *J. Appl. Crystallogr.* **2020**, *53*, 226–235. <https://doi.org/10.1107/S1600576719014092>.

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.