

Review

Not peer-reviewed version

---

# Just Keep Rolling? – An Encompassing Review towards Accelerated Vaccine Product Life Cycles

---

[Janis Stiefel](#) <sup>\*</sup>, [Jeffrey Schloßhauer](#) <sup>‡</sup>, [Agnes Vosen](#) <sup>‡</sup>, [Sarah Kilz](#) <sup>‡</sup>, [Jan Zimmer](#) <sup>‡</sup>, [Sascha Balakin](#) <sup>‡</sup>

Posted Date: 20 July 2023

doi: [10.20944/preprints2023071341.v1](https://doi.org/10.20944/preprints2023071341.v1)

Keywords: vaccine adaption; product life cycle; nanocarrier; mRNA vaccines; vaccine market; protein structure prediction; digital twin



Preprints.org is a free multidiscipline platform providing preprint service that is dedicated to making early versions of research outputs permanently available and citable. Preprints posted at Preprints.org appear in Web of Science, Crossref, Google Scholar, Scilit, Europe PMC.

Copyright: This is an open access article distributed under the Creative Commons Attribution License which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Disclaimer/Publisher's Note: The statements, opinions, and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions, or products referred to in the content.

Review

# Just Keep Rolling? – An Encompassing Review towards Accelerated Vaccine Product Life Cycles

Janis Stiefel <sup>1,\*</sup>, Jeffrey Schloßhauer <sup>2,†</sup>, Agnes Vosen <sup>3,†</sup>, Sarah Kilz <sup>3,†</sup>, Jan Zimmer <sup>1,†</sup> and Sascha Balakin <sup>4,5,†</sup>

<sup>1</sup> Fraunhofer Institute for Microengineering and Microsystems IMM, Carl-Zeiss-Straße 18-20, 55129 Mainz, Germany

<sup>2</sup> Fraunhofer Institute for Cell Therapy and Immunology, Branch Bioanalytics and Bioprocesses IZI-BB, Am Mühlenberg 13, 14476 Potsdam, Germany

<sup>3</sup> Fraunhofer Center for International Management and Knowledge Economy IMW, Neumarkt 20, 04109 Leipzig, Germany

<sup>4</sup> Fraunhofer Institute for Ceramic Technologies and Systems IKTS Material Diagnostics, Bio- and Nanotechnology, Maria-Reiche-Straße 2, 01109 Dresden, Germany

<sup>5</sup> Max Bergmann Center of Biomaterials (MBC), Technical University of Dresden, Budapester Strasse 27, 01069 Dresden, Germany

\* Correspondence: janis.stiefel@imm.fraunhofer.de

† These authors contributed equally.

**Abstract:** In light of the recent pandemic, several COVID-19 vaccines were developed, tested and approved in a very short time, a process that otherwise takes many years. Above all, these efforts have also unmistakably revealed the capacity limits and potential for improvement in vaccine production. This review aims to emphasize recent approaches for targeted rapid adaptation and production of vaccines from an interdisciplinary multifaceted perspective. Using literature research, stakeholder analysis and a value proposition canvas, we reviewed technological innovations on the pharmacological level, formulation, validation and resilient vaccine production to supply bottlenecks and logistic networks. We identified four main drivers to accelerate the vaccine product life cycle: computerized candidate screening, modular production, digitized quality management and a resilient business model with corresponding transparent supply chains. In summary, the results presented here can serve as a guide and implementation tool for flexible, scalable vaccine production to respond swiftly to pandemic situations in the future.

**Keywords:** vaccine adaption; product life cycle; nanocarrier; mRNA vaccines; vaccine market; protein structure prediction; digital twin

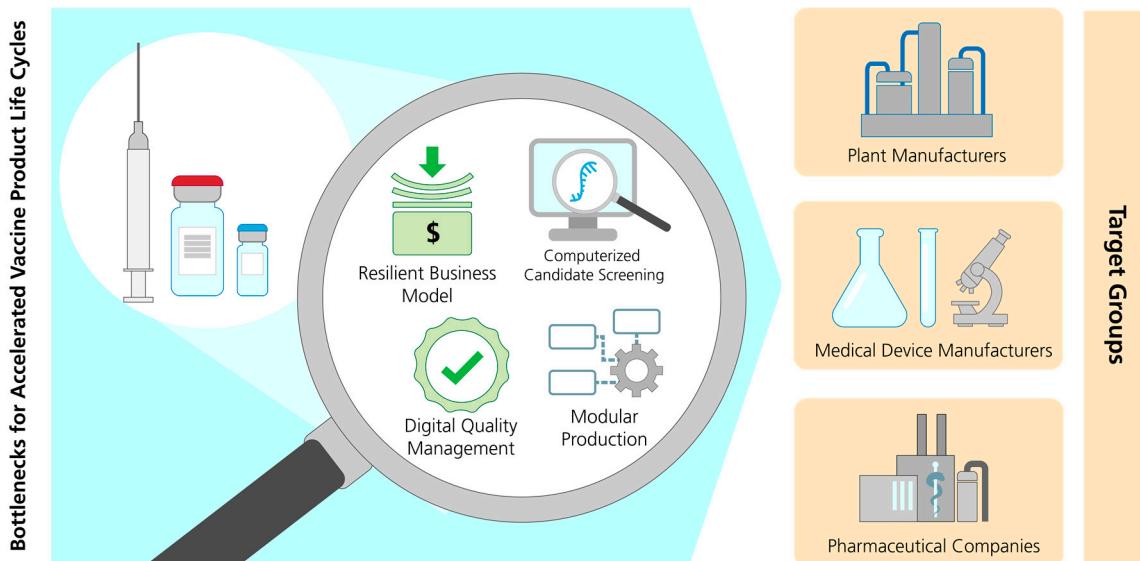
## 1. Introduction

Conventional development and manufacturing of vaccines against infectious diseases is complex and involves many suppliers, taking up to 15 years from the development to the approval of a vaccine candidate [1]. Younger companies such as Moderna (2010) and BioNTech (2008) have also been addressing vaccination in the field of cancer for years (BioNTech iNest BNT122; Moderna mRNA-4157/V940). They are thus setting a clear course towards personalized medicine, which has a significantly shorter product life cycle and involves greater product variability than previous vaccines and drugs. In view of the increasing competition due to the "New Era of Medicine" (mRNA drugs made popular by the pandemic), rapid adaptation of already existing vaccines is becoming increasingly important and driving the market situation to a new level.

In 2019, the global vaccine market made up approximately 2.5% of the total global pharmaceutical industry [2], making it one of the top ten therapeutic areas in terms of revenue [3]. At that time, the market comprised 5.5 billion vaccine doses with sales of \$33 billion [2]. About three quarters of global vaccine production took place and still takes place in Europe, with the majority of

vaccines produced being exported [4]. The largest customer regions are Southeast Asia, Africa and the USA [2].

In addition, the global vaccine market is characterized by an oligopolistic structure [5]. Since 1996, numerous mergers or acquisitions of pharmaceutical companies have led to a strong concentration in the vaccine market [6]. Therefore, with GlaxoSmithKline, Merck & Co, Sanofi, Pfizer, four key companies now take more than 91% of the global market. The high costs, time, and uncertainty of regulatory conditions related to the production of vaccines limit small and medium-sized companies, as they do not have the necessary skills and resources. However, since the emergence of the COVID-19 pandemic, the industry has experienced a boost in innovation: research and development investments as well as government participation in the market have increased significantly [7], which has also brought new players with innovative solutions to the market [8]. The APAC region in particular has been able to strengthen its position in the market [5] by rapidly adapting its vaccine development and production as well as its supply chain network to the new realities [7]. Sales of traditional vaccines are expected to continue to grow over the next five years [9] but market concentration will decline due to an increased number of competitors [5]. The increase in various infectious diseases [10] as well as investments in research and development of vaccines will be a driver for growth. In addition, investments in new therapeutic areas and improved production methods are expected to increase [7]. The demand for low-cost vaccines adapted to the needs of developing countries represents a major opportunity for vaccine developers [5]. Already, it is evident that vaccine manufacturers are pursuing diversified strategies to expand their geographic footprint and optimize their operations for more efficient and profitable vaccine production [7].



**Figure 1.** Graphical Abstract.

The quantum leaps in technology that accompanied the rapid market development have, not least, revealed much potential for improvement in accelerating the vaccine product life cycle. This review is intended to provide a holistic view on the vaccine adaptation process from the pharmacological level to production and logistical and economic issues with consideration of economic impacts and framework conditions. Therefore, the bottlenecks of the conventional vaccine production chain were identified and aligned with current innovation approaches based on literature research, ecosystem analysis and qualitative interviews with key players to bring the broad spectrum of possible actions closer to these very target groups.

## 2. Vaccine Types

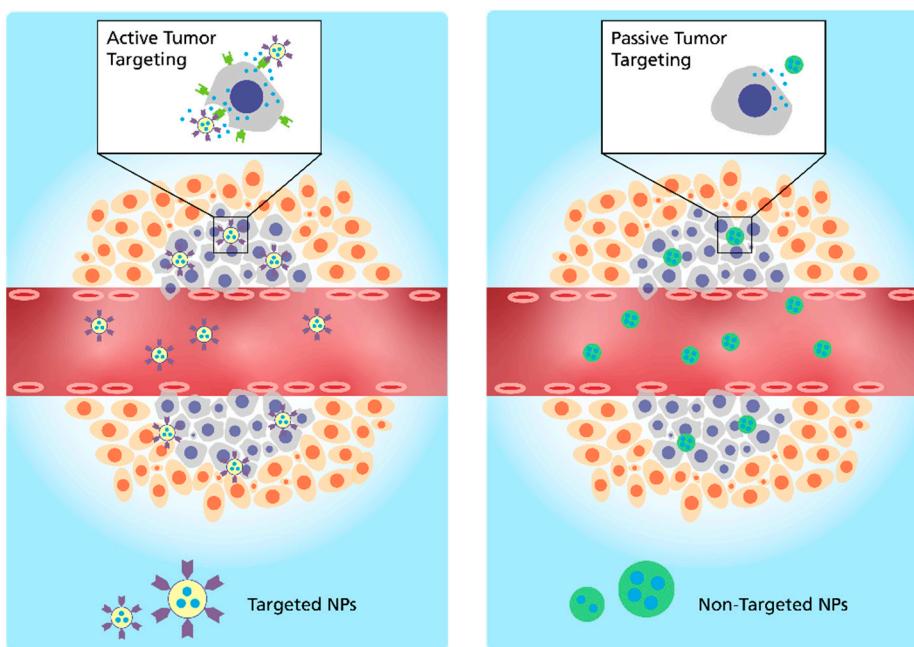
Nowadays, there is a broad spectrum of vaccine types to fight well-known diseases and emerging pathogens. This spectrum includes inactivated, live-attenuated, subunit, recombinant,

polysaccharide, conjugate toxoid and viral vector vaccines, among others [11]. In view of recent breakthroughs, this review will focus on messenger ribonucleic acid (mRNA) vaccines.

## 2.1. Administration and Targeting Mechanism

Various administration approaches for mRNA lipid nanoparticles (LNPs) have been reported, such as inhalation [12,13], oral [14–16], subcutaneous [17], intravenous [18–21] and intramuscular [22–24]. Following administration, the intended targeting mechanism is of great interest. Targeting of mRNA LNPs is beneficial because it can reduce the dose of mRNA required to elicit an immune response and facilitate vaccine production and distribution. To target a specific tissue, the LNP can be modified by introducing targeting ligands directly into the formulation, chemically conjugating them to the LNP surface, or by modifying the composition of the lipids in the formulation.

Two different types of targeting are possible. Active targeting can be executed with LNPs containing a target-specific ligand in the formulation, whereas passive targeting can be executed without targeting moiety [25]. Targeting moieties for active targeting mechanisms include antibodies [26], aptamers [27], small molecules [28] and proteins or peptides [29]. Passive targeting (EPR-effect) is influenced by factors such as the size and charge of the LNP, which depend on the molar composition of the different types of lipids used in the formulation [30]. Both mechanisms of active and passive targeting are illustrated in Figure 2.



**Figure 2.** Targeting mechanisms of mRNA-LNPs: (left) Active tumor targeting executed with a target-specific ligand in the formulation (antibodies, aptamers, small molecules and proteins or peptides), (right) passive tumor targeting based on the EPR-effect (updated from [31]).

It is possible to target secondary lymphoid organs by adding phosphatidylserine into the standard four-component MC3-based LNP formulation to facilitate the cellular uptake of immune cells beyond the charge-driven targeting principle commonly used today. As a result, the LNP achieved efficient protein expression in both lymph nodes and the spleen after IV administration [32]. Also, reticuloendothelial system (RES)-targeted LNPs were developed by modifying one lipid within the formulation of Onpattro, an approved lipid nanoparticle-based short interfering RNA drug for the treatment of polyneuropathies induced by hereditary transthyretin amyloidosis [33], to switch the surface charge of the LNP from neutral to anionic [31]. Furthermore, it was found that LNPs containing an amide bond in the tail are capable of selectively delivering mRNA to the mouse lung, in contrast to LNPs containing an ester bond in the tail, which tended to deliver mRNA to the liver

[32]. Once the cell has been targeted by the LNP, endocytosis is the most investigated internalization mechanism [33].

### 2.2. *Endosomal Escape*

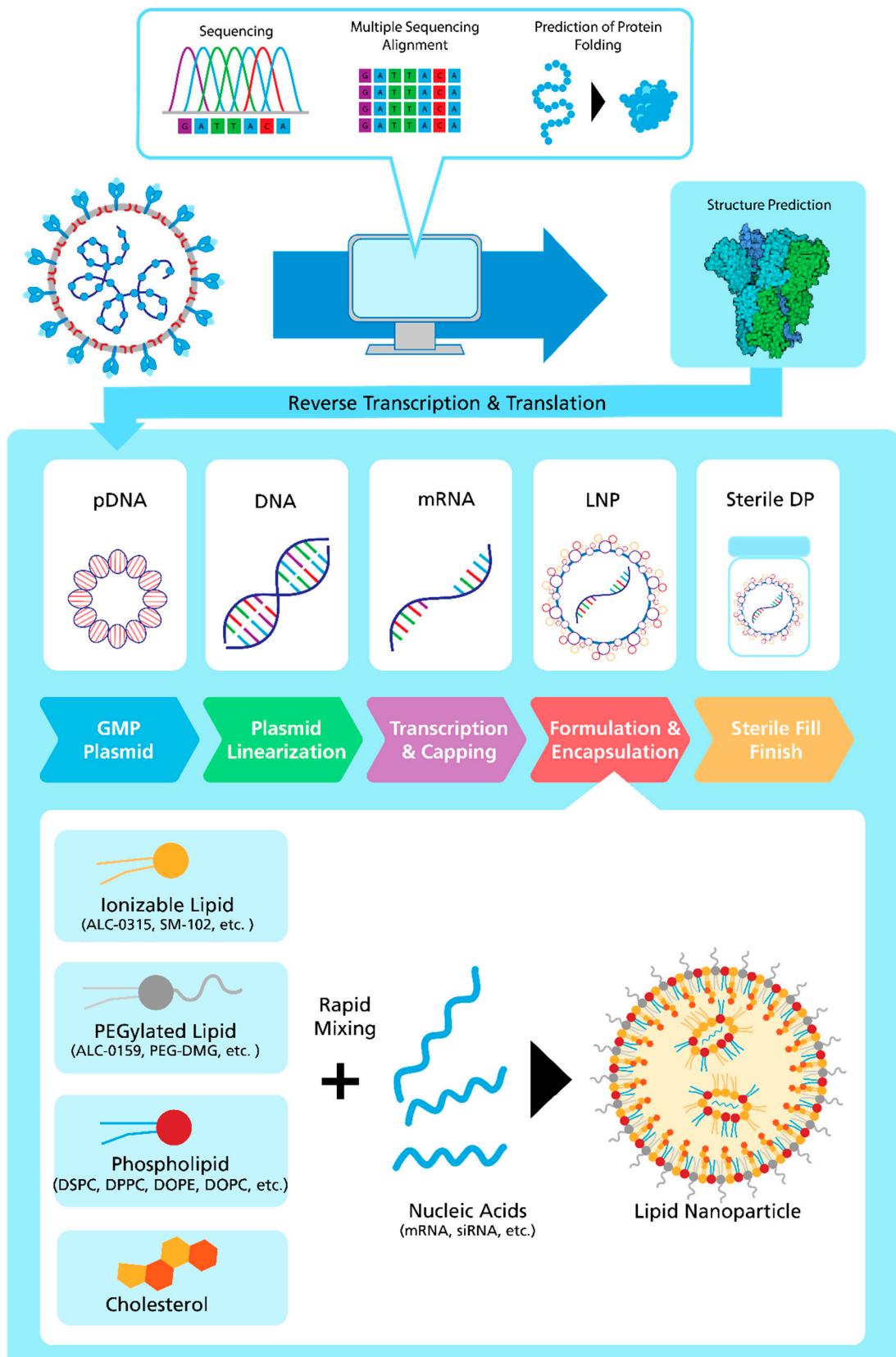
To induce efficient release of the mRNA to the cytoplasm after endocytosis, a detailed understanding of the endosomal escape is of major interest. Materials that have the ability to escape from the endosome/lysosome into the cytosol are called endosomolytic agents (e.g. peptides, proteins, toxins, polymers, small chemical compounds) [34]. Modification of LNPs to facilitate endosomal escape is an extensively studied nanotechnology tool for delivering therapeutics into cells. However, endosomal degradation of LNPs is a major hurdle [35].

Endosomal escape mechanisms include membrane destabilization [36], membrane fusion, proton sponge effect and photochemical internalization. Cationic polymers can escape from endosomes via the proton sponge effect. The downside is that they are covalently linked and usually have low biodegradability and high cytotoxicity [37–39]. Lysosomal cargo escape can be enhanced by excitation, using a lamp or a laser, to induce a more efficient leakage of siRNA into the cytoplasm. This is the mechanism of photochemical internalization [40,41].

### 2.3. *RNA as Active Pharmaceutical Ingredient*

There are three main types of RNA vaccines: non-amplifying mRNA molecules (mRNA), base-modified, non-amplifying mRNA molecules (bmRNA), which incorporate chemically modified nucleotides, and self-amplifying mRNA (saRNA or replicons) which maintain self-replicative activity and are derived from an RNA virus vector. Thus, saRNA is comparable to mRNA vaccines in terms of the benefits of rapid development, modular design, and cell-free synthesis, but requires a lower dose of RNA due to its self-replicative properties. Drug substance and product can be produced more quickly and are advantageous in the context of a pandemic response, as a larger percentage of the population could be vaccinated in a shorter period of time [42–44].

mRNA is synthesized via in vitro transcription (IVT) with a plasmid DNA (pDNA) template (Figure 3) [45]. In the following, mRNA requires the addition of an inverted triphosphate cap such as N7-methylated guanosine to the 5' end of the mRNA molecule to enhance biological activity [46]. The mRNA is then purified to remove impurities generated during IVT using chromatography techniques or affinity purification. Finally, a long poly-A tail is attached to increase protein expression [47,48].



**Figure 3.** Process for mRNA-LNP vaccine development: (top) Computer-assisted prediction of coronavirus spike protein (PDB: 5I08), (middle) mRNA-LNP production [48] starting from manufacturing of plasmid DNA (pDNA), mRNA and LNP to sterile drug product (DP) (adapted from [49]), (bottom) Lipid composition of mRNA-LNPs (adapted from [50]).

#### 2.4. Role of Adjuvants and LNPs

Adjuvants are introduced in the formulation of vaccines to increase their immunogenicity. An ideal adjuvant should have the following characteristics of biological activity: The formulation should be safe and effective for all ages. Immune responses should be increased in very young, elderly, or immunocompromised populations, where the adjuvant activity should be regional and transient, and adjuvants should not directly affect lymphocytes and should not be associated with non-specific B and T cellular responses [51]. Many human vaccines utilize non-biological materials, such as organic oil, aluminum salt, squalene, liposomes or LNPs to form micro-sized or nano-sized particles as delivery vehicles for antigens [52]. The LNPs in mRNA COVID-19 vaccines consist of four main components: (1) a neutral phospholipid, (2) cholesterol, (3) a polyethylene-glycol (PEG)-lipid, and (4) an ionizable cationic lipid. The latter contains positively charged ionizable amine groups (at low pH) to interact with the anionic mRNA during particle formation (Figure 3) and to facilitate membrane fusion during internalization. In addition, PEG-lipid is used to control the particle size and act as a steric barrier to prevent aggregation during storage. Together with the mRNA, these components form particles of about 60–100 nm in size [53]. The size of the LNP, pKa of the ionizable lipid, and lipid gradients, affect the tissue and cell specificity of the mRNA vaccine [54]. Ionizable lipid-containing LNP drives the production of interleukin 6, which leads to a prone follicular helper T cell and germinal center B cell responses to mRNA and recombinant protein vaccines [55].

#### 2.5. Biomaterials enabled long-term Storage

The two main focuses in the formulation development for vaccines are the stability of the final product, which will influence the storage conditions, and the addition of adjuvants to increase their immunogenicity [56]. To enable long-term storage of mRNA vaccines, which can help to increase their availability and accessibility in global health initiatives, the development of new biomaterials and technologies is essential. Cryoprotectants, lyophilization, and polymer-based formulations are researched to increase the self-life of vaccines. Cryoprotectants are substances that protect biological material from damage during freezing and thawing. Trehalose and glycerol are well known candidates to stabilize mRNA vaccines during long-term storage at low temperature [57].

LNPs have rapidly gained public attention as the delivery platform for mRNA vaccines. LNPs can be lyophilized and stored at ambient temperature for 12 weeks and at 4°C for at least 24 weeks without substantial changes to their physical properties or mRNA delivery efficiency [58]. Compared with LNPs, silica nanoparticles have several advantages, which warrant further clinical studies; for example, by modulating their structural and physiochemical properties, such as size, charge, surface functionality and shape, silica nanoparticles can deliver drugs across biological barriers. In addition, silica nanoparticles are stable in harsh biological settings, for example, in the acidic environment of the stomach, in which liposomes usually degrade, limiting their applicability in oral delivery [59]. The lyophilized complex of mesoporous silica nanoparticles with miRNA remained functional after 6 months of storage [60]. Polymer-based formulations, such as hydrogels and microparticles, are used to encapsulate mRNA vaccines and protect them from degradation during long-term storage. These formulations can also be designed to release the mRNA slowly over time, which can improve the immune response and reduce the need for booster shots. The immune response of mRNA LNPs encapsulated in hyaluronan hydrogels was maintained after being stored at room temperature for two weeks [61]. PCL/PLGA/PLLA microspheres were used to deliver SARS-CoV-2 antigens, where no visible particle degradation or changes in porosity patterns were observed during storage at 4°C for 180 days [62]. For example, a potent self-amplifying RNA (saRNA) vaccine against SARS-CoV-2 that is stable at room temperature has been developed. This saRNA vaccine is formulated with a nanostructured lipid carrier (NLC), which provides stability, ease of manufacture, and protection against degradation. Notably, the saRNA/NLC platform demonstrated thermostability when stored lyophilized at room temperature for at least 6 months and at refrigerated temperatures for at least 10 months [63].

### 3. *In Silico* Tools for Vaccine Development

The three-dimensional structure of a protein determines its properties to a crucial extent. Therefore, techniques to resolve the protein structure are indispensable. However, the experimental determination of a protein structure is very costly and time-consuming, so that alternatively modelling of the structures has gained outstanding importance in basic research. Computer-assisted predictions can be used to model higher-order three-dimensional protein structures based on amino acid sequence data. Acceleration in genome sequencing and the associated identification of protein-coding sequences using diverse bioinformatics methods has increased the available amount of protein data for protein structure prediction.

### 3.1. Epitope Prediction in Immunoinformatics

In pandemic situations, the spread of viruses can be significantly reduced by active immunization, hence accelerated development based on *in silico* predictions of envelope protein structures are important, as surface proteins of viruses form the first contact with the immune system. Due to the complexity of the innate and adaptive immune system, many different tools exist to train the immune cells to novel immunogens. Prediction of B-cell and T-cell epitopes have been utilized methods in immunoinformatics for many years [64]. Modelling of B-cell epitopes is based on the charged exposed surface area, secondary structure, and on hydrophilicity, as B-cell receptors have primarily hydrophobic binding sites [65]. However, many servers can only identify linear epitopes and not epitopes in which the amino acid residues are in physical contact but are separated in the primary structure [66–68]. While the 3D structure of the antigen is used for the identification of these conformational epitopes, the underlying antigen structure must be resolved [69,70]. In contrast, T-cells recognize intracellularly processed antigens in the form of peptide - major histocompatibility complex class (MHC) complexes. The prediction of T-cell epitopes is performed by a variety of methods, including docking models, hidden Markov models, decision trees, and artificial neural networks [71,72].

### 3.2. AI-based Protein Complex Predictions

Although machine learning models have been significantly improved in recent years, the prediction of optimal B-cell and T-cell epitopes is often limited due to a low amount of training data [73]. Chen et al. were able to obtain strong T-cell responses by take advantage of a neural network for predicting antigens in the context of specific human MHC II alleles, called MARIA (major histocompatibility complex analysis with recurrent integrated architecture) [74]. The combination of the mentioned epitope prediction tools and an additional approach, which utilizes good quality data sets to predict 3D protein structures based on its amino acid sequence would be of significance to adequately target emerging pathogens (Figure 3). Since the accurate determination of protein structure based on predictions is considered the “holy grail” of protein biochemistry, the Critical Assessment of Structure Prediction (CASP) event, which takes place every 2 years, introduced the AlphaFold neural network in 2020, which appears to master the major challenges of protein structure prediction. AlphaFold combines biological and physical information about protein structures and utilizes multi-sequence alignments to build a deep learning approach, which can predict various protein structures with high accuracy [75]. This neural network developed by DeepMind efficiently learns from the growing experimental data available in the Protein data bank, while the AlphaFold prediction significantly accelerates the experimental structure resolution, resulting in an increase in training data [76,77]. In addition to AlphaFold, the neural network RoseTTAFold provides another related approach to solve modeling problems and accurately predict protein complexes [78]. ESMFold and OmegaFold are two more recently published machine learning methods that allow precise predictions of atomic-level protein structures [79,80]. Although AlphaFold was initially limited in predicting multimer complex formation, which is essential for the development of potential vaccine candidates, the published source code was rapidly improved after the release to end up with the prediction tool AlphaFold Multimer [81]. In collaboration with EMBL’s European Bioinformatics Institute, DeepMind made the predictions of over 200 million UniProt entries available, thereby revolutionizing basic and applied research [82]. Whereas computerized identification of antigens can

significantly accelerate vaccine development, the stability of the potentially highly immunogenic protein can complicate expression. Recently, it was shown that the computational method PROSS can be utilized to increase the stability of the SARS-CoV-2 spike protein while maintaining high immunogenicity [83].

#### 4. Digitization and Regulation of Vaccine Quality Management

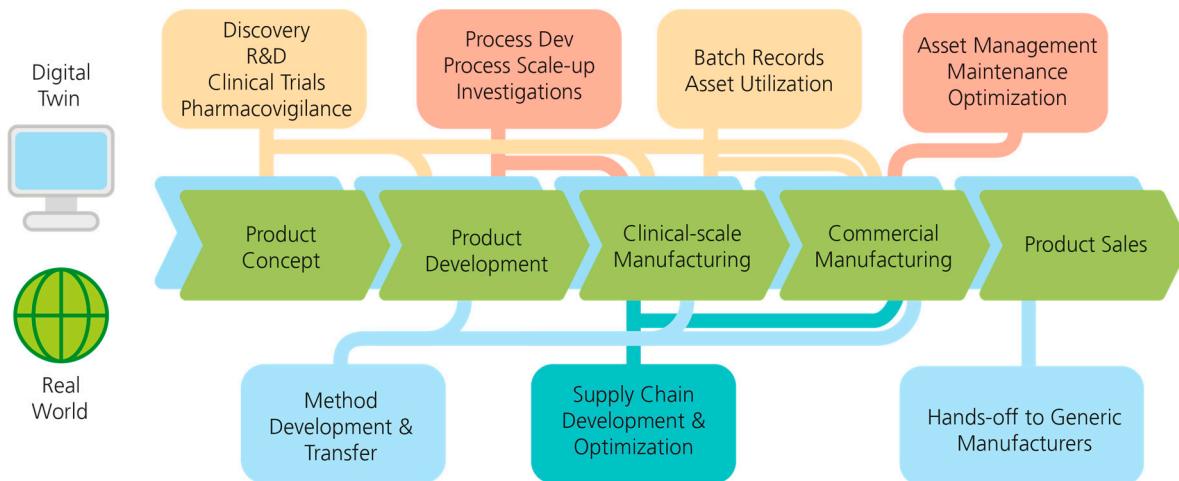
As described in the previous chapters, in mRNA-based vaccines, the protein-coding sequence is introduced directly into the cells, causing them to express and present the antigen themselves. In contrast to DNA vaccines, recombinant changes in the recipient genome are almost impossible due to the chemical structure of the mRNA sequences [45] and their fast degradation. During the Covid19 pandemic, more than 90 percent of the 224 million delivered vaccine doses in Germany were based on mRNA systems (by BioNTech/Pfizer 164.6 M doses, Moderna 37.7 M doses) until today [84]. As mRNA contact triggers both humoral and cellular immune response while entering only the cytoplasm but not to the nucleus, mRNA vaccination is seen as highly efficient [85]. The state-of-the-art for industrial production of the relatively new class of mRNA vaccines is based on enzymatically catalyzed in vitro transcription, a fundamental molecular biology technique. First, plasmids carrying the relevant DNA sequence are produced and linearized. In mRNA manufacturing, the linearized DNA is then enzymatically transcribed into mRNA. After complete processing, the mRNA molecules are encapsulated in lipid nanoparticles and dosed into storable units during fill-and-finish. Along this manufacturing chain, two quality control steps are implemented regarding DNA quality after linearization and hydrophobic interaction chromatography and mRNA purity after sterile filtration.

##### 4.1. Challenges and Benefits of Digital Vaccine Quality Management

Despite its finally recognized medical and economic potential, mRNA vaccine production, like that of most chemical and medicinal products, is still carried out in batches, which limits scalability and associated production capacity. However, the transition to continuous production, as also sought and encouraged by regulatory bodies such as EMA and FDA due to the benefits of agility, quality, flexibility, and cost, will only be possible through digital process development and control and digital quality assurance. With digitalization approaches in so-called eQMS (electronic quality management systems e.g., Veeva Systems, ValGenesis or SAP HANA) on the rise, the trend of digitized research and development, with its increasingly rapid breakthroughs, has also unmistakably impacted the field of pharmaceutical quality control. However, the necessary disruptions in the pharmaceutical industry are lacking, as they already existed in other industries years ago. One challenge could be that established companies ("Big Pharma") have already invested in existing research, study and manufacturing processes resulting in a slow drive to invest in new solutions as the risk of low pay-off in time is seen as too high. Pfizer, Sanofi, and Bayer helped the respective start-ups BioNTech, Moderna and CureVac to transfer their research on mRNA drugs into Covid-19 vaccines. Vice versa they provided them with the appropriate infrastructure e.g., pharmacovigilance. In contrast, changing to digital quality management systems from the start of the product life cycle might be able to reduce of overall product development costs of up to 20.000 times [86] and promote flexible decision-making towards societal and economical changes, especially underlined by the current pandemic. The pharmaceutical life cycle is composed of the following: Product concept, product development, clinical scale manufacturing, commercial manufacturing, and market sales. Clinical trials are part of this product lifecycle [87] being most important to sort out "correlates of immunity/protection" (number of neutralizing antibodies) whereas study design and cohort recruitment is key to a fast approval. In reality, difficulties in recruitment causes delays in 80 % of all clinical trials. This might be since not all patients are treated with the drug in the trials (placebo). With recruiting the appropriate patients in sufficient numbers being difficult, they also represent a small portion of the large and diverse population and, accordingly, are not accurate even though safety and effectiveness of drugs are crucial to be evaluated in clinical trials.

##### 4.2. Digital Twin Implementations in Vaccine Product Life Cycles

In line with the concept of Quality-by-Design (QbD), so-called digital twins of a patient could provide different real-time simulations of patients and are thus more comprehensive faster, safer, and more accurate. QbD gives the required design spaces to sharpen the quality target product profile (QTPP), related to the quality, safety, and efficacy of the active ingredient to avoid out of specification batches based on advanced process control using a digital twin. There are only limited studies made public addressing a digital twin for continuous in vitro-transcription that explicitly discuss QbD-compliant model validation and the requirements for a digital twin in good manufacturing process (GMP)-compliant production [88]. A digital twin is a virtual representation of an object or system that spans its lifecycle, is updated from real-time data, and uses simulation, machine learning and reasoning to help decision-making [89]. This way, the entire production including quality attributes, process parameters, critical aspects, equipment, processes such as changes, deviations, CAPAs (Corrective Actions, Preventive Actions) can be translated in a virtual image in the digital world (Figure 4).



**Figure 4.** Real and digital production. A digital twin offers actionable process steps at all stages of the production process chain compared to the real world scenario (modified from [90]).

As in real life, changes and CAPAs are made to this virtual image and deviations occur. The entire life cycle of a product is mapped digitally from the idea generation and conceptualization and a continuous open optimization loop for product and production is introduced, as the entire product lifecycle is integrated into factory or plant lifecycle and performance data [91]. As an example, periodic reviews of equipment and plants and systems are required by regulation at defined intervals. The evaluation of all changes made during the period under review is considered to assess whether there is an impact on the system. At present, this aspect is treated "stepmotherly" and there is no detailed evaluation of the changes made. In a digital twin, the status could be reviewed and evaluated whether changes had an impact. The operator/system owner is thus supported in the implementation of regulatory requirements. The industry is reacting to the momentum of the global digital twin market, estimated at \$ 3.1 billion in 2020, projected to reach \$ 48.2 billion by 2026, resulting in highly funded endeavors like the \$1.78M „Smart Design and Manufacturing Pilot” project between FDA and Siemens to showcase advanced digital design and manufacturing [92].

#### 4.3. Regulatory Considerations

Not least, the transition to a digital twin will also have an enormous positive impact on the speed of regulatory processes and approvals. On the surface, the relatively swift approval of Covid-19 vaccines under „Accelerated Assessment” with the aid of the centralized “Rolling Review” (RR), where pharmaceutical and non-clinical development data packages are evaluated before the clinical data is fully available, could be dealt as the future of medicinal product approval. However, RR comes with less predictable study outcome, an overload of regulatory resources and more

uncertainties as safety issues may be postponed to the post-marketing phase. Further, accelerated approval was not a new phenomenon of the current Covid pandemic. Laws for accelerated vaccine approval were in power before. Conditional marketing authorization (CMA) in e.g., Germany was introduced with the adaption of regulation (EG) Nr. 726/2004, article 14(7) in 2006 and applied to for Influenza vaccine approval in 2010 and 2016. Using digital quality assurance, data is recorded without gaps at every point in time, leading to an always up-to-date version of documentation without printing or active distribution necessary and less training effort for the employee. Recording and evaluation could take place in real time [93] and inline continuous documentation of processes leads to clearer comprehension and faster approval through authorities.

## 5. Discussion

In this review, we identified the bottlenecks of the conventional vaccine production chain to map them against current innovation approaches (Figure 5). To do this, we first conducted an ecosystem and stakeholder analysis and laid out the pains and gains of the key players in a value proposition canvas. As a result, we derived a concept that focuses on four main drivers to accelerate the vaccine product life cycle: computerized candidate screening, modular production, digitized quality management and a resilient business model with corresponding transparent supply chains.

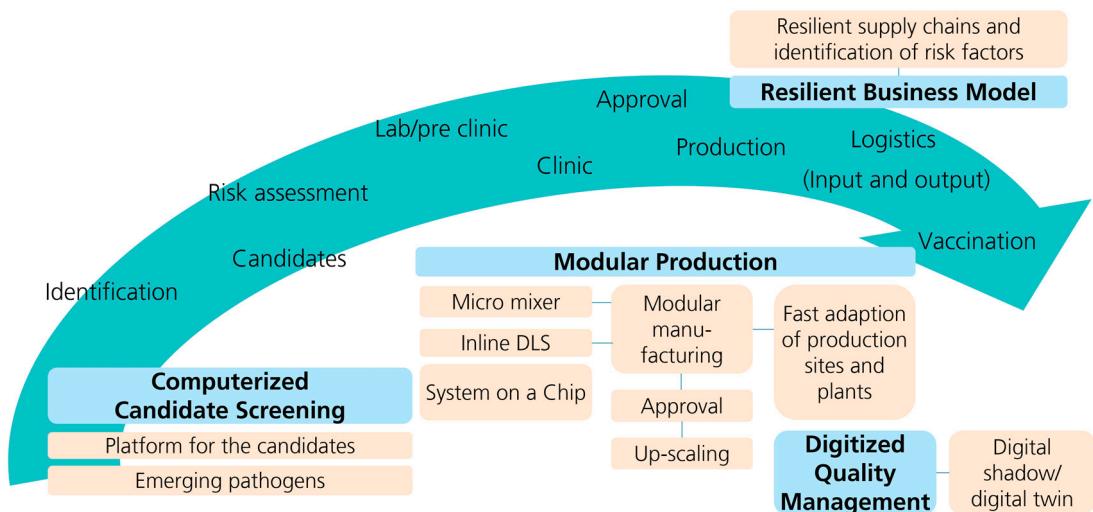
First, we highlighted the advantages and disadvantages of mRNA-LNP-vaccines, nevertheless it is important to point out that there is no “panacea”, and it will be essential to rely on a variety of vaccine-types to encounter future pathogens.

The need for rapid adaptation to a rapidly changing environment makes mRNA-LNP vaccines a promising tool due to their modular production capabilities in terms of formulation and drug delivery. It is possible to tailor the individual components such as the mRNA for expression of identified antigens (or proteins in general) to elicit an immune response and the lipid components to target specific tissues by passive or even active targeting mechanisms to minimize off-target effects and encapsulate the mRNA cargo for prolonged circulation in the bloodstream.

In addition, it is possible to produce mRNA-LNP vaccines on a large scale by simply mixing the required components for rapid distribution of sufficient vaccine for global use. As the development of vaccines that are stable at elevated temperatures continues, production time and costs will decrease, and distribution routes will be simplified.

The incorporation of improved micro-mixing technology and in-line DLS (Dynamic Light Scattering) into the modular production platform enables the production of precisely tailored LNPs in terms of particle size and particle size distribution, with the possibility of upscaling. Also, this technology can be used to produce small quantities of drug product for efficient vaccine candidate screening.

Further research should focus on understanding the mechanisms of internalization. This may not only help to improve drug delivery and targeting and to reduce the required drug dose to minimize cost and production time but may also reduce off-target effects.



**Figure 5.** Main drivers to accelerate the vaccine product life cycle: computerized candidate screening, modular production, digitized quality management and a resilient business model with corresponding transparent supply chains.

During the Corona pandemic, effective vaccines against the SarsCov2 virus could be developed in a relatively short time. A key factor was the knowledge of the resolved protein structure of the SARS-CoV virus, which could be transferred to SARS-CoV2 [94]. For this purpose, a 2P modification was used, which utilizes two proline mutations to stabilize the pre-fusion conformation resulting in an efficient neutralizing antibody response [95].

Because habitats of currently separated species will overlap in future and pathogens can consequently infect new hosts, pandemic situations and thus novel viruses will play a major role. In addition, the rapid evolution of viruses and their adaptation to selection pressures is a particular challenge to vaccine development, so accelerated development and adaptation of vaccines are urgently needed. Therefore, the recently developed approaches for accurate protein structure prediction can make a substantial contribution by rapidly responding to future pandemics by modelling potential vaccine candidates.

In terms of regulatory aspects and quality management, various emergency tools for accelerated vaccine approval were given already before the current pandemic but are in constant transit in view of the relatively new mRNA technology and emerging pathogens. “Accelerated Assessment” under Rolling Review makes sense as an instrument in emergencies where benefits outweigh risks but remains questionable as a standard instrument. This would mean that much more human capacity (headcounts) would have to be mobilized in order not to permanently overburden resources on the part of the authorities. Systemic and digitalized solutions i.e. digital twin technology both in companies and authorities should be suggested.

The EMA aims towards regulatory research and innovation drive in their “EMA Regulatory Science to 2025 – Strategic reflection” and the International Council for Harmonization of Technical Requirements for Pharmaceuticals for Human Use (ICH) promotes digital and fast exchange of approval documents to be accepted among the authorities that take part in the ICH. In the case of the BioNTech Covid vaccine, the EMA gave data to the FDA. RR could also be supported by digital solutions/automated document uploads and headcounts would only be necessary for decision-making. In conclusion, digital twin integration of equipment and process is most urgent today to increasing the productivity and profitability of enterprises and the quality of products. Nowadays, many companies are engaged in methodological issues of building digital twins, determining a set of controlled parameters/signals and their classification. Further research in the field of digital twins should be aimed at organizing communication between a real industrial facility and its digital model i.e., the exchange of data between real and virtual controllers. Thus, the modern approach to quality management is increasingly becoming a digital version of the quality management system, which is

driven by artificial intelligence. To adapt to the short product lifecycle but not overburden employees beyond their capacity, we need to move away from human-driven production and support the trend of digitalized process control and QA.

## 6. Conclusions

In reality, vaccine development and adaptation, especially in the area of mRNA technology, is not necessarily a truly circular or linear product life cycle as suspected at the beginning. Rather, our investigations showed that the communication interfaces between the individual instances within the process fields require major improvements. Comparable to other industries, the concatenation of already existing innovations and the associated digital knowledge transfer between basic research and application is the key to rapid, demand-driven vaccine production. In the future, this review is to give thought and aid at combining these multifaceted technological breakthroughs.

**Author Contributions:** Conceptualization, J.S., J.Sc., A.V., S.K., J.Z. and S.B.; methodology, J.S., J.Sc., A.V., S.K., J.Z. and S.B.; validation, J.S., J.Sc., A.V., S.K., J.Z. and S.B.; formal analysis, J.S., J.Sc., A.V., S.K., J.Z. and S.B.; investigation, J.S., J.Sc., A.V., S.K., J.Z. and S.B.; resources, J.S., J.Sc., A.V., S.K., J.Z. and S.B.; data curation, J.S., J.Sc., A.V., S.K., J.Z. and S.B.; writing—original draft preparation, J.S., J.Sc., A.V., S.K., J.Z. and S.B.; writing—review and editing, J.S., J.Sc., A.V., S.K., J.Z. and S.B.; visualization, J.S., J.Sc., J.Z. and S.B.; supervision, J.S. and S.K.; project administration, J.S., J.Sc., A.V., S.K., J.Z. and S.B.; funding acquisition, J.S., J.Sc., A.V., S.K., J.Z. and S.B. All authors have read and agreed to the published version of the manuscript.

**Funding:** This research received no external funding. The APC was funded by the Fraunhofer Society for the Advancement of Applied Research, Germany.

**Institutional Review Board Statement:** Not applicable.

**Informed Consent Statement:** Not applicable.

**Data Availability Statement:** No new data were created or analyzed in this study. Data sharing is not applicable to this article.

**Acknowledgments:** We would like to thank Maria Fläschner for her support and Lisa Pokropp for illustration (both Fraunhofer IMM).

**Conflicts of Interest:** The authors declare no conflict of interest. The funders had no role in the design of the study; in the collection, analyses, or interpretation of data; in the writing of the manuscript; or in the decision to publish the results.

## References

1. Deutsche Akademie der Naturforscher Leopoldina e. V. – German National Academy of Sciences –. Vaccine Development, Testing, and Approval. Available online: <https://www.leopoldina.org/en/topics/vaccinations/vaccine-development-and-recommendations/> (accessed on 30 May 2023).
2. Nord LB. *Pharmamarkt in der Pandemie – Der Gewinner der Krise?*
3. Statista. Leading 10 therapeutic areas worldwide by sales in 2019. Available online: <https://www.statista.com/statistics/407971/projected-revenue-of-top-therapeutic-areas-worldwide/>.
4. vfa - Verband Forschender Arzneimittelhersteller e.V. Europas Impfstoffindustrie. Wir versorgen die Welt – verlässlich und innovativ. Available online: <https://www.vfa.de/download/faktenblatt-impfstoffindustrie-europa.pdf>.
5. Frost & Sullivan. *Growth Opportunities in Global Vaccines Market, Forecast to 2024: Increased Investment in Adult Immunization and Improved Mammalian Cell-culture Expression Systems will Drive Growth and Profitability* ME98-52, 2019.
6. Gahr, M.; Heininger, U.; Bartmann, P.; Huppertz, H.I.; Kinet, M.; Klein, R. & Korenke, C. Folgen der Monopolisierung in der Pharmaindustrie für die Bereitstellung von Impfstoffen. *Monatsschr Kinderheilkd* **2013**, *161*, 554–558, doi:10.1007/s00112-013-2912-9.
7. Frost & Sullivan. *Global Vaccine Growth Opportunities: Capability Expansion in Nucleic Acid-based Vaccines is Accelerating the Disruption in Vaccines Globally* PCCF-52, 2022.
8. Statista. Impfstoffe – Umsatz. Available online: <https://de.statista.com/outlook/hmo/pharmazeutika/impfstoffe/weltweit#umsatz>.
9. Statista. Marktanteile führender Pharmaunternehmen im Segment Impfstoffe im Jahr 2017 und Prognose für das Jahr 2024. Available online: <https://de.statista.com/outlook/hmo/pharmazeutika/impfstoffe/weltweit#umsatz>.

<https://de.statista.com/statistik/daten/studie/312609/umfrage/marktanteile-fuehrender-pharmaunternehmen-im-segment-impfstoffe/>.

- 10. Statista. Global vaccine market revenues from 2014 to 2020 (in billion U.S. dollars)\*. Available online: <https://www.statista.com/statistics/265102/revenues-in-the-global-vaccine-market/>.
- 11. U.S. Department of Health and Human Services. Vaccine Types. Available online: <https://www.hhs.gov/immunization/basics/types/index.html> (accessed on 5 June 2023).
- 12. Lokugamage, M.P.; Vanover, D.; Beyersdorf, J.; Hatit, M.Z.C.; Rotolo, L.; Echeverri, E.S.; Peck, H.E.; Ni, H.; Yoon, J.-K.; Kim, Y.; et al. Optimization of lipid nanoparticles for the delivery of nebulized therapeutic mRNA to the lungs. *Nat. Biomed. Eng.* **2021**, *5*, 1059–1068, doi:10.1038/s41551-021-00786-x.
- 13. Yin, B.; Chan, C.K.W.; Liu, S.; Hong, H.; Wong, S.H.D.; Lee, L.K.C.; Ho, L.W.C.; Zhang, L.; Leung, K.C.-F.; Choi, P.C.-L.; et al. Intrapulmonary Cellular-Level Distribution of Inhaled Nanoparticles with Defined Functional Groups and Its Correlations with Protein Corona and Inflammatory Response. *ACS Nano* **2019**, *13*, 14048–14069, doi:10.1021/acsnano.9b06424.
- 14. Sung, J.; Alghoul, Z.; Long, D.; Yang, C.; Merlin, D. Oral delivery of IL-22 mRNA-loaded lipid nanoparticles targeting the injured intestinal mucosa: A novel therapeutic solution to treat ulcerative colitis. *Biomaterials* **2022**, *288*, 121707, doi:10.1016/j.biomaterials.2022.121707.
- 15. Yang, C.; Long, D.; Sung, J.; Alghoul, Z.; Merlin, D. Orally Administered Natural Lipid Nanoparticle-Loaded 6-Shogaol Shapes the Anti-Inflammatory Microbiota and Metabolome. *Pharmaceutics* **2021**, *13*, doi:10.3390/pharmaceutics13091355.
- 16. El-Mayta, R.; Zhang, R.; Shepherd, S.J.; Wang, F.; Billingsley, M.M.; Dudkin, V.; Klein, D.; Lu, H.D.; Mitchell, M.J. A Nanoparticle Platform for Accelerated In Vivo Oral Delivery Screening of Nucleic Acids. *Adv. Therap.* **2021**, *4*, 2000111, doi:10.1002/adtp.202000111.
- 17. Davies, N.; Hovdal, D.; Edmunds, N.; Nordberg, P.; Dahlén, A.; Dabkowska, A.; Arteta, M.Y.; Radulescu, A.; Kjellman, T.; Höijer, A.; et al. Functionalized lipid nanoparticles for subcutaneous administration of mRNA to achieve systemic exposures of a therapeutic protein. *Mol. Ther. Nucleic Acids* **2021**, *24*, 369–384, doi:10.1016/j.omtn.2021.03.008.
- 18. An, D.; Schneller, J.L.; Frassetto, A.; Liang, S.; Zhu, X.; Park, J.-S.; Theisen, M.; Hong, S.-J.; Zhou, J.; Rajendran, R.; et al. Systemic Messenger RNA Therapy as a Treatment for Methylmalonic Acidemia. *Cell Rep.* **2017**, *21*, 3548–3558, doi:10.1016/j.celrep.2017.11.081.
- 19. Ramaswamy, S.; Tonnu, N.; Tachikawa, K.; Limphong, P.; Vega, J.B.; Karmali, P.P.; Chivukula, P.; Verma, I.M. Systemic delivery of factor IX messenger RNA for protein replacement therapy. *Proc. Natl. Acad. Sci. U. S. A.* **2017**, *114*, E1941–E1950, doi:10.1073/pnas.1619653114.
- 20. Richner, J.M.; Himansu, S.; Dowd, K.A.; Butler, S.L.; Salazar, V.; Fox, J.M.; Julander, J.G.; Tang, W.W.; Shresta, S.; Pierson, T.C.; et al. Modified mRNA Vaccines Protect against Zika Virus Infection. *Cell* **2017**, *168*, 1114–1125.e10, doi:10.1016/j.cell.2017.02.017.
- 21. Schrom, E.; Huber, M.; Aneja, M.; Dohmen, C.; Emrich, D.; Geiger, J.; Hasenpusch, G.; Herrmann-Janson, A.; Kretzschmann, V.; Mykhailyk, O.; et al. Translation of Angiotensin-Converting Enzyme 2 upon Liver- and Lung-Targeted Delivery of Optimized Chemically Modified mRNA. *Mol. Ther. Nucleic Acids* **2017**, *7*, 350–365, doi:10.1016/j.omtn.2017.04.006.
- 22. Alberer, M.; Gnad-Vogt, U.; Hong, H.S.; Mehr, K.T.; Backert, L.; Finak, G.; Gottardo, R.; Bica, M.A.; Garofano, A.; Koch, S.D.; et al. Safety and immunogenicity of a mRNA rabies vaccine in healthy adults: an open-label, non-randomised, prospective, first-in-human phase 1 clinical trial. *Lancet* **2017**, *390*, 1511–1520, doi:10.1016/S0140-6736(17)31665-3.
- 23. Hassett, K.J.; Benenato, K.E.; Jacquinet, E.; Lee, A.; Woods, A.; Yuzhakov, O.; Himansu, S.; Deterling, J.; Geilich, B.M.; Ketova, T.; et al. Optimization of Lipid Nanoparticles for Intramuscular Administration of mRNA Vaccines. *Mol. Ther. Nucleic Acids* **2019**, *15*, 1–11, doi:10.1016/j.omtn.2019.01.013.
- 24. Sedic, M.; Senn, J.J.; Lynn, A.; Laska, M.; Smith, M.; Platz, S.J.; Bolen, J.; Hoge, S.; Bulychev, A.; Jacquinet, E.; et al. Safety Evaluation of Lipid Nanoparticle-Formulated Modified mRNA in the Sprague-Dawley Rat and Cynomolgus Monkey. *Vet. Pathol.* **2018**, *55*, 341–354, doi:10.1177/0300985817738095.
- 25. Kularatne, R.N.; Crist, R.M.; Stern, S.T. The Future of Tissue-Targeted Lipid Nanoparticle-Mediated Nucleic Acid Delivery. *Pharmaceutics (Basel)* **2022**, *15*, doi:10.3390/ph15070897.
- 26. Mucker, E.M.; Thiele-Suess, C.; Baumhof, P.; Hooper, J.W. Lipid nanoparticle delivery of unmodified mRNAs encoding multiple monoclonal antibodies targeting poxviruses in rabbits. *Mol. Ther. Nucleic Acids* **2022**, *28*, 847–858, doi:10.1016/j.omtn.2022.05.025.
- 27. Liu, Y.; Qian, X.; Ran, C.; Li, L.; Fu, T.; Su, D.; Xie, S.; Tan, W. Aptamer-Based Targeted Protein Degradation. *ACS Nano* **2023**, *17*, 6150–6164, doi:10.1021/acsnano.2c10379.
- 28. Lin, C.; Mostafa, A.; Jans, A.; Wolters, J.C.; Mohamed, M.R.; van der Vorst, E.P.C.; Trautwein, C.; Bartneck, M. Targeting Ligand Independent Tropism of siRNA-LNP by Small Molecules for Directed Therapy of Liver or Myeloid Immune Cells. *Adv. Healthc. Mater.* **2023**, *e2202670*, doi:10.1002/adhm.202202670.

29. Qin, J.; Xue, L.; Gong, N.; Zhang, H.; Shepherd, S.J.; Haley, R.M.; Swingle, K.L.; Mitchell, M.J. RGD peptide-based lipids for targeted mRNA delivery and gene editing applications. *RSC Adv.* **2022**, *12*, 25397–25404, doi:10.1039/d2ra02771b.

30. Nakamura, T.; Kawai, M.; Sato, Y.; Maeki, M.; Tokeshi, M.; Harashima, H. The Effect of Size and Charge of Lipid Nanoparticles Prepared by Microfluidic Mixing on Their Lymph Node Transitivity and Distribution. *Mol. Pharm.* **2020**, *17*, 944–953, doi:10.1021/acs.molpharmaceut.9b01182.

31. Silva, F.; Cabral Campello, M.P.; Paulo, A. Radiolabeled Gold Nanoparticles for Imaging and Therapy of Cancer. *Materials (Basel)* **2020**, *14*, doi:10.3390/ma14010004.

32. Luozhong, S.; Yuan, Z.; Sarmiento, T.; Chen, Y.; Gu, W.; McCurdy, C.; Gao, W.; Li, R.; Wilkens, S.; Jiang, S. Phosphatidylserine Lipid Nanoparticles Promote Systemic RNA Delivery to Secondary Lymphoid Organs. *Nano Lett.* **2022**, *22*, 8304–8311, doi:10.1021/acs.nanolett.2c03234.

33. Akinc, A.; Maier, M.A.; Manoharan, M.; Fitzgerald, K.; Jayaraman, M.; Barros, S.; Ansell, S.; Du, X.; Hope, M.J.; Madden, T.D.; et al. The Onpattro story and the clinical translation of nanomedicines containing nucleic acid-based drugs. *Nat. Nanotechnol.* **2019**, *14*, 1084–1087, doi:10.1038/s41565-019-0591-y.

34. Varkouhi, A.K.; Scholte, M.; Storm, G.; Haisma, H.J. Endosomal escape pathways for delivery of biologicals. *J. Control. Release* **2011**, *151*, 220–228, doi:10.1016/j.jconrel.2010.11.004.

35. Chan, C.-L.; Ewert, K.K.; Majzoub, R.N.; Hwu, Y.-K.; Liang, K.S.; Leal, C.; Safinya, C.R. Optimizing cationic and neutral lipids for efficient gene delivery at high serum content. *J. Gene Med.* **2014**, *16*, 84–96, doi:10.1002/jgm.2762.

36. Benedikt Junglas; Amelie Axt; Carmen Siebenaller; Hilal Sonel; Nadja Hellmann; Stefan A.L. Weber; Dirk Schneider. Membrane destabilization and pore formation induced by the Synechocystis IM30 protein. *Biophysical Journal* **2022**, *121*, 3411–3421, doi:10.1016/j.bpj.2022.08.014.

37. Bus, T.; Traeger, A.; Schubert, U.S. The great escape: how cationic polyplexes overcome the endosomal barrier. *J. Mater. Chem. B* **2018**, *6*, 6904–6918, doi:10.1039/c8tb00967h.

38. Su, D.; Coste, M.; Diaconu, A.; Barboiu, M.; Ulrich, S. Cationic dynamic covalent polymers for gene transfection. *J. Mater. Chem. B* **2020**, *8*, 9385–9403, doi:10.1039/d0tb01836h.

39. Lee, J.; Sands, I.; Zhang, W.; Zhou, L.; Chen, Y. DNA-inspired nanomaterials for enhanced endosomal escape. *Proc. Natl. Acad. Sci. U. S. A.* **2021**, *118*, doi:10.1073/pnas.2104511118.

40. Ali, L.M.A.; Gary-Bobo, M. Photochemical Internalization of siRNA for Cancer Therapy. *Cancers (Basel)* **2022**, *14*, doi:10.3390/cancers14153597.

41. Ali, L.M.A.; Gary-Bobo, M. Photochemical Internalization of siRNA for Cancer Therapy. *Cancers (Basel)* **2022**, *14*, doi:10.3390/cancers14153597.

42. Rappaport, A.R.; Hong, S.-J.; Scallan, C.D.; Gitlin, L.; Akoopie, A.; Boucher, G.R.; Egorova, M.; Espinosa, J.A.; Fidanza, M.; Kachura, M.A.; et al. Low-dose self-amplifying mRNA COVID-19 vaccine drives strong protective immunity in non-human primates against SARS-CoV-2 infection. *Nat. Commun.* **2022**, *13*, 3289, doi:10.1038/s41467-022-31005-z.

43. Vogel, A.B.; Lambert, L.; Kinney, E.; Busse, D.; Erbar, S.; Reuter, K.C.; Wicke, L.; Perkovic, M.; Beissert, T.; Haas, H.; et al. Self-Amplifying RNA Vaccines Give Equivalent Protection against Influenza to mRNA Vaccines but at Much Lower Doses. *Mol. Ther.* **2018**, *26*, 446–455, doi:10.1016/j.ymthe.2017.11.017.

44. Blakney, A.K.; Ip, S.; Geall, A.J. An Update on Self-Amplifying mRNA Vaccine Development. *Vaccines (Basel)* **2021**, *9*, doi:10.3390/vaccines9020097.

45. Sahin, U.; Karikó, K.; Türeci, Ö. mRNA-based therapeutics--developing a new class of drugs. *Nat. Rev. Drug Discov.* **2014**, *13*, 759–780, doi:10.1038/nrd4278.

46. Shuman, S. Catalytic activity of vaccinia mRNA capping enzyme subunits coexpressed in Escherichia coli. *Journal of Biological Chemistry* **1990**, *265*, 11960–11966, doi:10.1016/S0021-9258(19)38494-7.

47. Holtkamp, S.; Kreiter, S.; Selmi, A.; Simon, P.; Koslowski, M.; Huber, C.; Türeci, O.; Sahin, U. Modification of antigen-encoding RNA increases stability, translational efficacy, and T-cell stimulatory capacity of dendritic cells. *Blood* **2006**, *108*, 4009–4017, doi:10.1182/blood-2006-04-015024.

48. Gebre, M.S.; Brito, L.A.; Tostanoski, L.H.; Edwards, D.K.; Carfi, A.; Barouch, D.H. Novel approaches for vaccine development. *Cell* **2021**, *184*, 1589–1603, doi:10.1016/j.cell.2021.02.030.

49. Arranta Bio. An integrated solution for mRNA vaccine manufacturing. Available online: <https://arrantabio.com/mrna-manufacturing-vaccines/> (accessed on 22 June 2023).

50. Jung, H.N.; Lee, S.-Y.; Lee, S.; Youn, H.; Im, H.-J. Lipid nanoparticles for delivery of RNA therapeutics: Current status and the role of in vivo imaging. *Theranostics* **2022**, *12*, 7509–7531, doi:10.7150/thno.77259.

51. Nooraei, S.; Sarkar Lotfabadi, A.; Akbarzadehmoallemkolaei, M.; Rezaei, N. Immunogenicity of Different Types of Adjuvants and Nano-Adjuvants in Veterinary Vaccines: A Comprehensive Review. *Vaccines (Basel)* **2023**, *11*, 453, doi:10.3390/vaccines11020453.

52. Kobiyama, K.; Ishii, K.J. Making innate sense of mRNA vaccine adjuvanticity. *Nat Immunol* **2022**, *23*, 474–476, doi:10.1038/s41590-022-01168-4.

53. Schoenmaker, L.; Witzigmann, D.; Kulkarni, J.A.; Verbeke, R.; Kersten, G.; Jiskoot, W.; Crommelin, D.J.A. mRNA-lipid nanoparticle COVID-19 vaccines: Structure and stability. *International Journal of Pharmaceutics* **2021**, *601*, 120586, doi:10.1016/j.ijpharm.2021.120586.

54. Li, C.; Lee, A.; Grigoryan, L.; Arunachalam, P.S.; Scott, M.K.D.; Trisal, M.; Wimmers, F.; Sanyal, M.; Weidenbacher, P.A.; Feng, Y.; et al. Mechanisms of innate and adaptive immunity to the Pfizer-BioNTech BNT162b2 vaccine. *Nat Immunol* **2022**, *23*, 543–555, doi:10.1038/s41590-022-01163-9.

55. Alameh, M.-G.; Tombácz, I.; Bettini, E.; Lederer, K.; Ndeupen, S.; Sittplangkoon, C.; Wilmore, J.R.; Gaudette, B.T.; Soliman, O.Y.; Pine, M.; et al. Lipid nanoparticles enhance the efficacy of mRNA and protein subunit vaccines by inducing robust T follicular helper cell and humoral responses. *Immunity* **2021**, *54*, 2877–2892.e7, doi:10.1016/j.jimmuni.2021.11.001.

56. D'Amico, C.; Fontana, F.; Cheng, R.; Santos, H.A. Development of vaccine formulations: past, present, and future. *Drug Deliv. Transl. Res.* **2021**, *11*, 353–372, doi:10.1007/s13346-021-00924-7.

57. Li, H.; Bian, Y.-L.; Schreurs, N.; Zhang, X.-G.; Raza, S.H.A.; Fang, Q.; Wang, L.-Q.; Hu, J.-H. Effects of five cryoprotectants on proliferation and differentiation-related gene expression of frozen-thawed bovine calf testicular tissue. *Reprod. Domest. Anim.* **2018**, *53*, 1211–1218, doi:10.1111/rda.13228.

58. Young, R.E.; Hofbauer, S.I.; Riley, R.S. Overcoming the challenge of long-term storage of mRNA-lipid nanoparticle vaccines. *Mol. Ther.* **2022**, *30*, 1792–1793, doi:10.1016/j.mtthe.2022.04.004.

59. Janjua, T.I.; Cao, Y.; Yu, C.; Popat, A. Clinical translation of silica nanoparticles. *Nat. Rev. Mater.* **2021**, *6*, 1072–1074, doi:10.1038/s41578-021-00385-x.

60. Hosseinpour, S.; Cao, Y.; Liu, J.; Xu, C.; Walsh, L.J. Efficient transfection and long-term stability of mono-miRNA-26a-5p for osteogenic differentiation by large pore sized mesoporous silica nanoparticles. *J. Mater. Chem. B* **2021**, *9*, 2275–2284, doi:10.1039/D0TB02756A.

61. Jia, F.; Huang, W.; Yin, Y.; Jiang, Y.; Yang, Q.; Huang, H.; Nie, G.; Wang, H. Stabilizing RNA Nanovaccines with Transformable Hyaluronan Dynamic Hydrogel for Durable Cancer Immunotherapy. *Adv Funct Materials* **2023**, *33*, 2204636, doi:10.1002/adfm.202204636.

62. Shahjin, F.; Patel, M.; Machhi, J.; Cohen, J.D.; Nayan, M.U.; Yeapuri, P.; Zhang, C.; Waight, E.; Hasan, M.; Abdelmoaty, M.M.; et al. Multipolymer microsphere delivery of SARS-CoV-2 antigens. *Acta Biomater.* **2023**, *158*, 493–509, doi:10.1016/j.actbio.2022.12.043.

63. Voigt, E.A.; Gerhardt, A.; Hanson, D.; Jennewein, M.F.; Battisti, P.; Reed, S.; Singh, J.; Mohamath, R.; Bakken, J.; Beaver, S.; et al. A self-amplifying RNA vaccine against COVID-19 with long-term room-temperature stability. *NPJ Vaccines* **2022**, *7*, 136, doi:10.1038/s41541-022-00549-y.

64. Oli, A.N.; Obialor, W.O.; Ifeanyichukwu, M.O.; Odimegwu, D.C.; Okoyeh, J.N.; Emechebe, G.O.; Adejumo, S.A.; Ibeano, G.C. Immunoinformatics and Vaccine Development: An Overview. *Immunotargets Ther.* **2020**, *9*, 13–30, doi:10.2147/ITT.S241064.

65. Kazi, A.; Chuah, C.; Majeed, A.B.A.; Leow, C.H.; Lim, B.H.; Leow, C.Y. Current progress of immunoinformatics approach harnessed for cellular- and antibody-dependent vaccine design. *Pathog. Glob. Health* **2018**, *112*, 123–131, doi:10.1080/20477724.2018.1446773.

66. Chen, J.; Liu, H.; Yang, J.; Chou, K.-C. Prediction of linear B-cell epitopes using amino acid pair antigenicity scale. *Amino Acids* **2007**, *33*, 423–428, doi:10.1007/s00726-006-0485-9.

67. Saha, S.; Raghava, G.P.S. Prediction of continuous B-cell epitopes in an antigen using recurrent neural network. *Proteins* **2006**, *65*, 40–48, doi:10.1002/prot.21078.

68. Singh, H.; Ansari, H.R.; Raghava, G.P.S. Improved Method for Linear B-Cell Epitope Prediction Using Antigen's Primary Sequence. *PLoS One* **2013**, *8*, doi:10.1371/journal.pone.0062216.

69. Krügel, J.V.; Lundegaard, C.; Lund, O.; Nielsen, M. Reliable B cell epitope predictions: impacts of method development and improved benchmarking. *PLoS Comput. Biol.* **2012**, *8*, e1002829, doi:10.1371/journal.pcbi.1002829.

70. Ponomarenko, J.; Bui, H.-H.; Li, W.; Fusseder, N.; Bourne, P.E.; Sette, A.; Peters, B. ElliPro: a new structure-based tool for the prediction of antibody epitopes. *BMC Bioinformatics* **2008**, *9*, 1–8, doi:10.1186/1471-2105-9-514.

71. Tong, J.C.; Ren, E.C. Immunoinformatics: current trends and future directions. *Drug Discov. Today* **2009**, *14*, 684–689, doi:10.1016/j.drudis.2009.04.001.

72. Schaap-Johansen, A.-L.; Vujošić, M.; Borch, A.; Hadrup, S.R.; Marcatili, P. T Cell Epitope Prediction and Its Application to Immunotherapy. *Front. Immunol.* **2021**, *12*, 712488, doi:10.3389/fimmu.2021.712488.

73. Meysman, P.; Barton, J.; Bravi, B.; Cohen-Lavi, L.; Karnaughov, V.; Lilleskov, E.; Montemurro, A.; Nielsen, M.; Mora, T.; Pereira, P.; et al. Benchmarking solutions to the T-cell receptor epitope prediction problem: IMMREP22 workshop report, 2022.

74. Chen, B.; Khodadoust, M.S.; Olsson, N.; Wagar, L.E.; Fast, E.; Liu, C.L.; Muftuoglu, Y.; Sworder, B.J.; Diehn, M.; Levy, R.; et al. Predicting HLA class II antigen presentation through integrated deep learning. *Nat. Biotechnol.* **2019**, *37*, 1332–1343, doi:10.1038/s41587-019-0280-2.

75. Jumper, J.; Evans, R.; Pritzel, A.; Green, T.; Figurnov, M.; Ronneberger, O.; Tunyasuvunakool, K.; Bates, R.; Žídek, A.; Potapenko, A.; et al. Highly accurate protein structure prediction with AlphaFold. *Nature* **2021**, *596*, 583–589, doi:10.1038/s41586-021-03819-2.

76. Kryshtafovych, A.; Moult, J.; Albrecht, R.; Chang, G.A.; Chao, K.; Fraser, A.; Greenfield, J.; Hartmann, M.D.; Herzberg, O.; Josts, I.; et al. Computational models in the service of X-ray and cryo-electron microscopy structure determination. *Proteins* **2021**, *89*, 1633–1646, doi:10.1002/prot.26223.

77. Tai, L.; Zhu, Y.; Ren, H.; Huang, X.; Zhang, C.; Sun, F. 8 Å structure of the outer rings of the *Xenopus laevis* nuclear pore complex obtained by cryo-EM and AI. *Protein Cell* **2022**, *13*, 760–777, doi:10.1007/s13238-021-00895-y.

78. Baek, M.; DiMaio, F.; Anishchenko, I.; Dauparas, J.; Ovchinnikov, S.; Lee, G.R.; Wang, J.; Cong, Q.; Kinch, L.N.; Schaeffer, R.D.; et al. Accurate prediction of protein structures and interactions using a three-track neural network. *Science* **2021**, *373*, 871–876, doi:10.1126/science.abj8754.

79. Wu, R.; Ding, F.; Wang, R.; Shen, R.; Zhang, X.; Luo, S.; Su, C.; Wu, Z.; Xie, Q.; Berger, B.; et al. *High-resolution de novo structure prediction from primary sequence*, 2022.

80. Lin, Z.; Akin, H.; Rao, R.; Hie, B.; Zhu, Z.; Lu, W.; Smetanin, N.; Verkuil, R.; Kabeli, O.; Shmueli, Y.; et al. Evolutionary-scale prediction of atomic-level protein structure with a language model. *Science* **2023**, *379*, 1123–1130, doi:10.1126/science.ade2574.

81. Evans, R.; O'Neill, M.; Pritzel, A.; Antropova, N.; Senior, A.; Green, T.; Žídek, A.; Bates, R.; Blackwell, S.; Yim, J.; et al. *Protein complex prediction with AlphaFold-Multimer*, 2021.

82. Callaway, E. 'The entire protein universe': AI predicts shape of nearly every known protein. *Nature* **2022**, *608*, 15–16, doi:10.1038/d41586-022-02083-2.

83. Williams, J.A.; Biancucci, M.; Lessen, L.; Tian, S.; Balsaraf, A.; Chen, L.; Chesterman, C.; Maruggi, G.; Vandepaer, S.; Huang, Y.; et al. Structural and computational design of a SARS-CoV-2 spike antigen with improved expression and immunogenicity. *Sci. Adv.* **2023**, *9*, eadg0330, doi:10.1126/sciadv.adg0330.

84. Federal Ministry of Health. Current vaccination status. Available online: <https://impfdashboard.de/en/> (accessed on 17 May 2023).

85. Oğuz, F.; Atmaca, H. mRNA as a Therapeutics: Understanding mRNA Vaccines. *Adv. Pharm. Bull.* **2021**, *12*, 274–282, doi:10.34172/apb.2022.028.

86. Rodionov, N.; Tatarnikova, L. Digital twin technology as a modern approach to quality management. *E3S Web Conf.* **2021**, *284*, 4013, doi:10.1051/e3sconf/202128404013.

87. Forbes. Meet Your Digital Twin: The Coming Revolution In Drug Development. Available online: <https://www.forbes.com/sites/ganeskesari/2021/09/29/meet-your-digital-twin-the-coming-revolution-in-drug-development/> (accessed on 17 May 2023).

88. Helgers, H.; Hengelbrock, A.; Schmidt, A.; Strube, J. Digital Twins for Continuous mRNA Production. *Processes* **2021**, *9*, 1967, doi:10.3390/pr9111967.

89. IBM. What is a digital twin? Available online: <https://www.ibm.com/topics/what-is-a-digital-twin> (accessed on 17 May 2023).

90. Maher, C. QUALITY & SPEED: EMPLOYING DIGITAL TWINS TO ACCELERATE THE PRODUCT LIFECYCLE. Available online: [https://www.ispeboston.org/download/educational\\_presentations/2022/2022-02-16-Digital-Tools-Charlie-Maher.pdf](https://www.ispeboston.org/download/educational_presentations/2022/2022-02-16-Digital-Tools-Charlie-Maher.pdf) (accessed on 17 May 2023).

91. SIEMENS. Als Digital Enterprise die digitale Transformation beschleunigen. Available online: [https://www.siemens.com/de/de/produkte/automatisierung/themenfelder/digital-enterprise.html?gclid=EAIAjQobChMI6ITaq7WN\\_AIVBp3VCh3tGAFuEAMYAyAAEgKFYPD\\_BwE&acz=1](https://www.siemens.com/de/de/produkte/automatisierung/themenfelder/digital-enterprise.html?gclid=EAIAjQobChMI6ITaq7WN_AIVBp3VCh3tGAFuEAMYAyAAEgKFYPD_BwE&acz=1) (accessed on 17 May 2023).

92. U.S. Food And Drug Administration. Smart Design and Manufacturing Pilot. Available online: <https://www.fda.gov/emergency-preparedness-and-response/ocet-advanced-manufacturing/smart-design-and-manufacturing-pilot> (accessed on 17 May 2023).

93. kpibench GmbH. kpibench Qualitätsmanagement: digitale Qualitätssicherung in Echtzeit. Available online: <https://www.kpibench.com/de/2021/02/kpibench-qualitätsmanagement-digitale-qualitäts-sicherung-in-echtzeit/> (accessed on 17 May 2023).

94. Wrapp, D.; Wang, N.; Corbett, K.S.; Goldsmith, J.A.; Hsieh, C.-L.; Abiona, O.; Graham, B.S.; McLellan, J.S. Cryo-EM structure of the 2019-nCoV spike in the prefusion conformation. *Science* **2020**, *367*, 1260–1263, doi:10.1126/science.abb2507.

95. Higgins, M.K. Can We AlphaFold Our Way Out of the Next Pandemic? *J. Mol. Biol.* **2021**, *433*, 167093, doi:10.1016/j.jmb.2021.167093.

**Disclaimer/Publisher's Note:** The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.