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Article

Anti-Inflammatory and Antinociceptive Activity of Herbal Liposomes of *Pentaclethra macrophylla* (Fabaceae) Stem Bark Extract

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Abstract: **Purpose:** Inflammation of various degrees is common among humans. There are associated side effects with orthodox delivery systems and anti-inflammatory agents hence the study investigated the characteristics of herbal liposomes and the anti-inflammatory potency of the liposomes formulated from *Pentaclethra macrophylla* with the view to having a drug with a better delivery system and lesser side effects. **Methods:** Herbal liposomes were formulated using solidified reverse micellar solutions (SRMS) of P90H and goat fat and characterized for particle size and morphology, pH – time dependent analysis, encapsulation efficiency (EE%) and Fourier Transform infrared spectroscopy. The in vitro antinociceptive and anti-inflammatory study were carried out using membrane stabilization by hypotonicity induced haemolysis and determination of anti-platelet aggregatory activity models. The *in-vivo* anti-nociceptive and anti-inflammatory studies on egg albumin and formaldehyde induced arthritis models were done. Total white blood cell count and differential blood count were carried out on the rats. **Results:** The results showed that there was no change in pH for the PM unloaded liposomes and 2.5g of PM-loaded liposomes from Day 1 to Day 7, but mild variation in the rest of the formulations. The EE ranged from 35.2% to 94% increasing according to the drug concentration. The photomicrographs of the liposomes showed that the particles were spherical in shape. The particle sizes were within the acceptable range for liposomes. FTIR showed no interaction. In the arthritis study, PM-loaded liposomes inhibited edema consistently throughout the duration of observation. Inhibition of membrane increased steadily with an increase in concentration of PM in the liposomes and the standard drug. The platelet aggregatory inhibition decreased steadily with an increase in concentration of the PM in the liposomes as well as the standard. The T50 dose of PM had the highest percentage of WBC and it decreased as the treatment doses increased from T100 to T200. There was no significant differences among the Neutrophil counts of the different groups. **Conclusion:** The study, therefore, showed that the methanol extract of *Pentaclethra macrophylla* formed efficient herbal liposomes with antinociceptive and anti-inflammatory activities.

Keywords: solidified reverse micellar solutions (SRMS); *Pentaclethra macrophylla*; herbal liposome; antinociception; anti-inflammatory activity; stem bark extract

1. Introduction

Pentaclethra is a small genus of the Fabaceae family, comprising approximately three species of trees, distributed throughout tropical and subtropical regions of the world [1–4]. The species of the genus *Pentaclethra*, such as *Pentaclethra macroloba* and *P. eetveldeana* have been described as medicinal plants with their biological activities assessed [3,4]. *P. macrophylla* is the sole member of the genus occurring naturally in the lowlands of West Africa. It is abundantly found in the southern, Middle Belt and south eastern Nigeria [5–7], with no varietal characterization [8].

Nearly all parts of the *P. macrophylla* plant are used for various human and animal ailments. The fruits, seeds, stem bark, leaves and roots have been reportedly used in traditional herbal practices in different countries. The stem bark has been credited in Nigeria as an antiulcer [9], antidiabetic [10], anti-inflammatory [11], antigonorrhoea [12], antimicrobial [13], antinociceptive [14], anticonvulsant and antitussive [12]. In Nigeria, the seeds have been reported as anticancer [15] and abortifacient [10] whereas in Cameroon, it is used for infertility [16]. The root bark is a laxative, enema against dysentery, liniment against itching and abortifacient as an infusion in both Nigeria and Cameroon [14]. The leaf is used to treat diarrhoeal, fever, stomach ache and convulsion [8,16,17]. The plant has also many nutritional benefits [18,19]. In Ghana, it has been reported as an anthelmintic for leprosy sores [13], antiepileptic [20], antidiarrhoeal [17], antipruritus and for wound management [16]. Bergenin is an active constituent of the plant and research indicates that bergenin has multiple biological activities, including anti-inflammatory and immunomodulatory properties [5]. A large body of ethnobotanical evidence indicates that the traditional use of *Pentaclethra macrophylla* in Nigeria are most likely related to the plant's anti-nociceptive and anti-inflammatory actions. However, since the pharmacological properties of *P. macrophylla* have rarely been reported, the use of this plant have continued without solid scientific evidence to support them. The present study was carried out to evaluate the anti-nociceptive and anti-inflammatory activities of liposomes prepared with the methanol extract of *P. macrophylla* and administered orally in animal models of pain and inflammation.

2. Materials and Methods

2.1. Collection and Identification of *Pentaclethra macrophylla* Stem Bark

The stem bark of *Pentaclethra macrophylla* tree was collected from Ejuona Obukpa Nsukka, Enugu State, Nigeria in March 2022 and authenticated by a plant taxonomist Mr. Alfred Ozioko of the International Centre for Ethno medicine Development (InterCEED) Nsukka, Enugu State, Nigeria. The Voucher specimen (PCG/UNN/ 014/507) was assigned to the sample which was deposited in the Herbarium of the Department of Pharmacognosy and Environmental Medicines, University of Nigeria, Nsukka, Nigeria. The plant material was collected in line with the ethical guide of the University of Nigeria, Nsukka, on herbal research.

2.2. Maceration and Extraction of Plant Material

The fleshy stem bark of *P. macrophylla* was harvested, thoroughly rinsed with distilled water, and air-dried at room temperature ($28 \pm 2^\circ\text{C}$). Dried plant portion (2 kg) was mechanically pulverized using a hammer mill (Henan Always Machinery, Zhengzhou, China). Methanol (MeOH, 98%) extraction was carried out according to an earlier method [9], at room temperature for 72 h, with vigorous shaking at intervals. The mixture was filtered using a muslin cloth, and the resulting filtrate was further filtered through multiple fluted filter-papers (Whatman No. 1 filter paper (Sigma-Aldrich, Johannesburg, South Africa) and the final filtrate concentrated *in vacuo* at 40°C to obtain dry crude methanol extract. Extracts were stored in sterile containers until further use.

2.3. Extraction of goat fat from *Capra hircus*

The adipose tissue of freshly slaughtered *Capra hircus* (goat) was obtained from the abattoir in Ikpa market Nsukka and freed of extraneous materials. The fat was extracted from the adipose tissue according to an earlier method with modification [21]. Briefly, the adipose tissue was grated and

boiled with half its weight of water on a water bath for 45 min. The molten fat was separated from the aqueous phase using a muslin cloth. The fat was further purified by heating the lipid with a 2% w/w suspension of Activated charcoal and bentonite (1:9) ratio blend at 100°C for 1 h. Thereafter the suspension was vacuum filtered using a Buchner funnel. The fat was stored in the refrigerator at 4°C.

2.4. Preparation of Lipospheres

The lipid matrices were first prepared according to Table 1. Briefly, Phospholipon 90H® (GmbH, Köln, Germany) and goat fat corresponded to 1.25 % (0.75:0.5) respectively. A 0.5g quantity of goat fat was carefully weighed into a beaker and melted at 80°C using a hot water bath. A 0.75g of Phospholipon 90H® was transferred into the melted goat fat and both were stirred continuously with a glass stirrer to ensure adequate mixing until a homogenous transparent liquid was obtained. Consequently, the aqueous phase consisting of Tween 80, sorbitol, sorbic acid and graded concentration of PM stem bark extract (0, 2.5, 5.0, 7.5 and 10 g) were equally heated to the same temperature and later added to the lipid phase. Lipospheres were formulated by high shear homogenization (Ultra-Turrax, T18 basic, IKA, Staufen, Germany) at 18,000 rpm for 5 min. The resultant lipospheres were cooled at room temperature and stored in amber-coloured bottles firmly secured for further analysis.

Table 1. Composition of lipospheres.

PM Batches	Goat fat	Phospholipon 90H	Sorbitol	Sorbic acid	Tween 80	Distilled water (qs)
(g)	(g)	(g)	(g)	(g)	(g)	w/w
0	0.5	0.75	4	0.05	1.5	100
2.5	0.5	0.75	4	0.05	1.5	100
5.0	0.5	0.75	4	0.05	1.5	100
7.5	0.5	0.75	4	0.05	1.5	100
10.0	0.5	0.75	4	0.05	1.5	100

2.5. Characterization of Lipospheres

2.5.1. Determination of Particle size and morphology of lipospheres

Microscopic evaluation and particle size analysis of the lipospheres were performed at $\times 100$ magnification. A 5 mg of liposphere from each batch was placed on a slide, covered with a cover slip and observed using a compound phase-contrast microscope (Motic B3; Motic, Carlsbad, CA, USA). All digital micrographs were captured using Moticam 2.0 image system (Motic, Carlsbad, CA, USA). Each sample was assessed five times and representative measurements were made for each sample. The particle dimensions (height, width, area and perimeter) were measured in micrometer (μm).

2.5.2. pH – time dependent analysis

After the lipospheres preparation the pH of the batches were determined using a pH meter (Hanna, H198108). The pH analysis and stability of the loaded and unloaded lipospheres were determined in a time dependent manner (24 h, 7 and 30 days).

2.5.3. Encapsulation efficiency (EE %)

The entrapment efficiency of the liposphere formulations was calculated by using the indirect method of ultrafiltration using Amicon filter tubes (Germany), which consisted of a filter membrane with a molecular weight cut-off of 10,000 at the base of the sample donor chamber. For this, the liposphere (1 ml aliquot) of undiluted sample was placed in the upper chamber and the sample recovery chamber was fitted below the membrane in the lower compartment. The unit was closed and centrifuged at 2,500 rpm for 5 min using a centrifuge (Model 420R Rotina Hettich, Germany) at 25 °C in order to separate microparticles from the untrapped drug and the free drug in the

supernatant was quantified afterwards using HPLC. The encapsulation efficiency was calculated through the formula;

$$EE (\%) = \frac{\text{Theoretical drug content} - \text{actual drug content}}{\text{Theoretical drug content}} \times 100 \quad (1)$$

The loading capacity was determined in relation to the total weight of the lipids using the formula:

$$LC (\%) = \frac{W_a - W_s}{W_a - W_s + W_l} \times 100 \quad (2)$$

W_a is the weight of the *P. macrophylla* added to the formulation and W_s is the actual amount of *P. macrophylla* encapsulated in the liposomes. W_l is the weight of the lipid added in the formulation.

2.5.4. HPLC method

HPLC determination of *P. macrophylla* in methanol was performed using a (ASI-100 automated sample injector) equipped with UV-vis detectors operating at 274 nm. Samples were chromatographed on a stainless steel C18 reverse phase column (Luna (2) 150x4.6 mm) packed with 5 mm particles (Lichrospher1 100 RP-18). For the separations, a gradient of mobile phase A (0.1 v/v % TFA in acetonitrile) and mobile phase B (0.1 v/v % TFA in water) was used. TFA was chosen to provide pH 2 in order to achieve narrow peak shapes. The flow rate was 1.0 mL min⁻¹, the column temperature was 40 °C and the injection volume was 10 µL. UV detection wavelength was 274 nm. The detector cell temperature was 40 °C. A calibration curve was plotted for the extract in the concentration range of 2–10 mg/ml. A good linear relationship was observed between the concentration of extract and its peak area with a percentage correlation coefficient of 99.37%. The required studies were carried out to estimate the precision and accuracy of the HPLC method.

2.6. FTIR analysis

The unloaded and loaded liposphere formulations (0, 2.5, 5, 7.5 and 10g) were analyzed for FTIR at spectra range of 4000-500cm⁻¹. A 0.4g of KBr was weighed and ground to powder. 0.001g of the samples containing *Pentaclethra macrophylla* (2.5, 5, 7.5 and 10 g) was separately weighed into the ground KBr and both were thoroughly mixed and moulded into a disc. The disc was inserted into the sample compartment of the instrument, and the IR spectrum was generated (Shimadzu FTIR. Model IR Affinity-1. Japan). The same procedure was repeated for the unloaded (placebo) liposphere.

2.7. *In vitro* Anti-nociceptive and Anti-inflammatory study

2.7.1. Assay of Membrane Stabilization by Hypotonicity induced hemolysis

The sample batches of the *P. macrophylla* loaded liposomes, unloaded liposomes and acetyl salicylic acid (Aspirin) were dissolved in distilled water (hypotonic solution). A 5ml hypotonic solution containing graded doses of the liposomes (100, 200, 400, 600 and 800 µg/ml) were made into duplicate pairs (per dose) of the centrifuge tubes. 5ml of the isotonic solution containing graded doses of the unloaded and loaded liposomes (100-800 µg/ml) were prepared and put into duplicate pairs (per dose) of the centrifuge tubes. The control tubes contained 5ml of the vehicle (distilled water) and 5 ml of 200 µg/ml of Aspirin respectively. Erythrocyte suspension (0.1 ml) was added to each of the tubes and mixed gently [22]. The mixtures were incubated for 1h at room temperature (37 °C), and then, centrifuged for 5 min at 1300 rpm. Absorbance (OD) of the hemoglobin content of the supernatant was estimated at 540 nm using a UV spectrophotometer (Milton Roy). The percentage haemolysis was calculated by assuming the haemolysis produced in the presence of distilled water was 100%.

The percentage inhibition of hemolysis was calculated using the following relation:

$$\% \text{ Inhibition of hemolysis} = 1 - \frac{\text{OD}_2 - \text{OD}_1}{\text{OD}_3 - \text{OD}_1} \times 100 \quad (3)$$

where OD_1 = absorbance of test sample in isotonic solution
 OD_2 = absorbance of test sample in hypotonic solution
 OD_3 = absorbance of control sample in hypotonic solutions

2.7.2. Determination of Anti-platelet aggregatory activity

This method of antiplatelet activity determination is a modification of Born and Cross [23]. Briefly, a 5ml of fresh blood sample was withdrawn intravenously from healthy volunteers using 5ml plastic syringes and put into plastic tube containing 0.01ml of 1% EDTA as the anticoagulant. The EDTA plastic tubes were centrifuged at 3000 rpm for 10 min. The blood sample separated and the supernatant was collected, diluted twice using normal saline and then used as platelet-rich plasma [24]. Absorption changes of the platelet rich plasma (PRP) was determined. A 0.2 ml of the platelet rich plasma, 0.4 ml of the 2M $CaCl_2$, and varying concentrations of the *Pentaclethra macrophylla* loaded liposomes (50, 100, 150, 200 mg representing 2.5, 5, 7.5 and 10 g formulations respectively), unloaded liposomes and normal saline were incubated. The absorbances of the solutions were measured at 520 nm using UV spectrophotometer (Milton Roy). The changes in absorbances of the solutions at 520 nm were taken at time intervals; (0, 30, 60, 90 and 120 s).

2.8. *In-vivo* antinociceptive and anti-inflammatory study

The animal experimental protocols were in accordance with the guidelines for conducting animal experiments stipulated by our Institution's Animal Ethics Committee and in compliance with the Federation of European Laboratory Animal Science Association and the European Community Council Directive of November 24, 1986 (86/609/EEC). Briefly, the rats were divided into groups and acclimatized to the laboratory environment a week before the study.

2.8.1. Egg albumin induced rat paw edema inflammatory model.

Fresh egg albumen induced rat hind paw oedema was used as model for acute inflammation. Some 28 adult Wistar albino rats of either sexes were weighed and divided into 7 groups of 4 rats each. The animals were allowed to acclimatize to the environment for 7 days before the experiment. The animals were then fasted and deprived of water for 18 h before the experiment. Deprivation of water was to ensure uniform hydration and to minimize variability in edematous response [24]. Immediately after the deprivation, their right hind paw volumes were taken at time zero ($t = 0$) volume ($V = V_0$) using a Vernier caliper. The first group of four animals were induced with edema but didn't receive any treatment, the second group received 20 mg/kg of Diclofenac (positive control group). The third group received 50 mg/kg of blank liposome containing no drug while the fourth to seventh groups received different doses (50, 100, 150 and 200 mg/kg body weight corresponding to 2.5, 5, 7.5 and 10 g) of the *P. macrophylla* loaded liposomes respectively. One hour after the administration of the test substances, 0.1 ml of undiluted fresh egg albumin was injected into the sub-plantar region of the right hind paw of the rats to induce acute inflammation. After the egg albumin administration, the paw volumes were measured using a vernier caliper at 0, 1, 2, 3, 4 and 5 h. Formation of oedema was then assessed in terms of the difference in the zero-time paw edema volume of the injected paw and its paw edema volume at different times after egg albumin injection of the right hind paw. Average inflammation, % inflammation and % inhibition of inflammation were calculated for each dose of the loaded liposome containing *P. macrophylla* using the following relationships:

(i)

$$\text{Average inflammation} = (V_T - V_0) \quad (4)$$

where V_0 = Volume of oedema at time zero (initial time); V_T = Volume of oedema at time T (0.5, 1, 2, 3, 4 and 5h)

(ii)

$$\% \text{ inflammation} = \frac{\text{Average inflammation of treated groups at time T}}{\text{Average inflammation of control at time T}} \times 100 \quad (5)$$

then

$$\% \text{ inflammation} = \frac{(V_T - V_0) \text{ groups 1,2,3,4,5}}{(V_T - V_0) \text{ control}} \times 100 \quad (6)$$

$$\% \text{ inhibition} = \frac{\text{Average inflammation of control} - \text{average inflammation of treated}}{\text{Average inflammation of control}} \times 100 \quad (7)$$

Average inflammation of control

$$\% \text{ inhibition of inflammation} = \frac{(V_T - V_0) \text{ control} - (V_T - V_0) \text{ treated groups}}{(V_T - V_0) \text{ control}} \times 100 \quad (8)$$

2.8.2. 2% Formaldehyde induced arthritis model

This experiment was carried out using adult Wistar albino rats weighing 60-100 g. The animals were placed at random and allocated to treatment groups. They were housed in clean cages and allowed free access to food and water while they acclimatized to the laboratory environment one week prior to the experiment. A total of 32 animals were used and were divided into 8 groups of 4 rats each (n=4). Group I received the vehicle water and served as the control group. Group II were induced with 0.1ml of 2 % formaldehyde but received no treatment. Group III received the standard diclofenac (20 mg/kg/body weight), group IV received blank liposphere while groups V, VI, VII, VIII received doses of *Pentaclethra macrophylla* lipospheres (50, 100, 150, 200 mg/kg bodyweight) respectively representing PM liposphere batches 2.5, 5, 7.5 and 10 g. One hour after the administration of oral doses of the vehicle and drugs, 0.1 ml of formaldehyde (2 %v/v) was used to induce arthritis in the sub-plantar region of the right hind paw of all the rats [25].

After the intravenous 0.1ml formaldehyde (2 % v/v) administration, the volume of the paws were measured using a vernier caliper at 0, 1, 2, 3, 4, 5 and 48 h. Formation of oedema was then assessed in terms of the difference in the zero-time paw edema volume of the injected paw and its paw edema volume at different times after 0.1ml formaldehyde injection of the right hind paw. % inhibition of the paw edema was calculated using the following formula:

$$\% \text{ inhibition of paw oedema} = 1 - \frac{\Delta V_{\text{treated}}}{\Delta V_{\text{untreated}}} \times 100 \quad (9)$$

where ΔV treated= mean change in paw volume edema of treated rat

ΔV untreated = mean change in paw volume of untreated rat

After 48 h, 2ml of blood was collected by retro-orbital cavity under the influence of anesthesia. Aliquot of the collected blood was used to perform hematological studies to estimate total blood count, differential blood count including (eosinophils, basophils, lymphocytes, monocytes).

2.8.2.1. Total White blood cell count

This was carried out following standard hematological procedures [26]. Briefly, well mixed anticoagulated blood was diluted 1:20 with Truk's solution (2% glacial acetic acid) in a test tube. This was loaded into an improved Neubauer counting chamber. Appropriate squares were counted and added up to determine the total red cell count.

2.8.2.2. Differential blood count

Drops of blood (2 drops) were placed on a dry slide and spread on the surface using another slide (blood film method) [26]. The Leishman's stain was carefully dropped onto the blood film and was covered using a smaller slide. The stain was allowed to act on the blood for 2-3 min. After that distilled water was used to wash off the stain until a pink color was obtained. Excess water was shaken off, under the slide cleaned and the slide allowed to dry upright. The slide (smear side up) was placed on the microscope stage. The blood smear was examined under the microscope with x10

lens resolution. The area with plenty WBCs was selected and an immersion oil was placed on that area. The resolution was changed to the oil immersion objective (100x). The differential cell count was then carried out and the morphology of the WBC noted.

Interventionary studies involving animals or humans, and other studies that require ethical approval, must list the authority that provided approval and the corresponding ethical approval code.

3. Results

3.1. pH time dependent analysis

Time dependent pH analysis of liposomes is presented in Table 2. There was no change in pH for the PM unloaded liposomes and 2.5g of PM-loaded liposomes from Day 1 to Day 30. However, in higher PM loadings, there were only slight changes in pH from Day 1 to Day 7. This change was highest in the 7.5g PM liposomes.

Table 2. Properties of the formulated liposomes.

Liposome batches (g)	Particle size (μm)	Encapsulation efficiency (EE %)	Loading capacity (LC)	pH time dependence (days)		
				1	7	30
0	12.56 ± 2.18	-	-	3.5	3.5	3.5
2.5	26.87 ± 4.21	35.2	4.4	3.8	3.8	3.8
5.0	42.42 ± 3.64	75.6	1.89	3.8	3.9	3.9
7.5	82.27 ± 6.24	89.4	3.4	3.7	3.9	3.9
10.0	98.67 ± 10.23	94.1	4.7	3.7	3.8	3.8

3.2. Encapsulation Efficiency (EE) and Loading capacity (LC)

Results of EE and LC are also presented in Table 2. EE ranged from 35 % to 94 %. It increased consistently and progressively from 35.2 % for 2.5 g PM-loaded liposomes to 94% in 10 g PM liposomes. This trend however, was not obtainable in the LC. An increase in PM-loading from 2.5 g to 5.0 g led to a sharp fall in LC from 4.4 to 1.89; on further increase of PM-loading to 7.5 g, LC quickly rose back to 3.4 and finally to 4.7 at 10 g PM-loading.

3.3. Particle size and morphology of liposomes

The photomicrographs of the liposomes in Figure 1 show that the particles were spherical in shape. The particle sizes were within the acceptable range for liposomes. The particle size of unloaded liposomes was lower than those of the PM-loaded liposomes. The result also shows that particle size increased with increase in the amount of PM loaded in the liposomes.

3.4. FTIR analysis

FTIR measures a sample's absorbance of infrared light at various wavelengths to determine the material's molecular composition and structure. The results of the FTIR of the unloaded and loaded samples of the liposomes are shown in Figure 2. The unloaded sample maintained peaks at 3623.14cm⁻¹ (N-H groups), 27706cm⁻¹ (C=O groups), 2009.92cm⁻¹ (C-O groups) and 832.9cm⁻¹ (C-H groups). The fingerprint region of the extract, PM-unloaded showed spectra with absorbance at 1457.4 cm⁻¹ and 1379.1 cm⁻¹. These peaks at this region corresponded to the asymmetric and symmetric bending of the C-H bond of methylene. Asymmetric and symmetric stretching of the C-H bond of methylene also occurred between 2935–2915 cm⁻¹ and 2865–2845 cm⁻¹ in all the formulated liposomes samples. The loaded liposomes had more sharp peaks at 2609cm⁻¹ (C=O

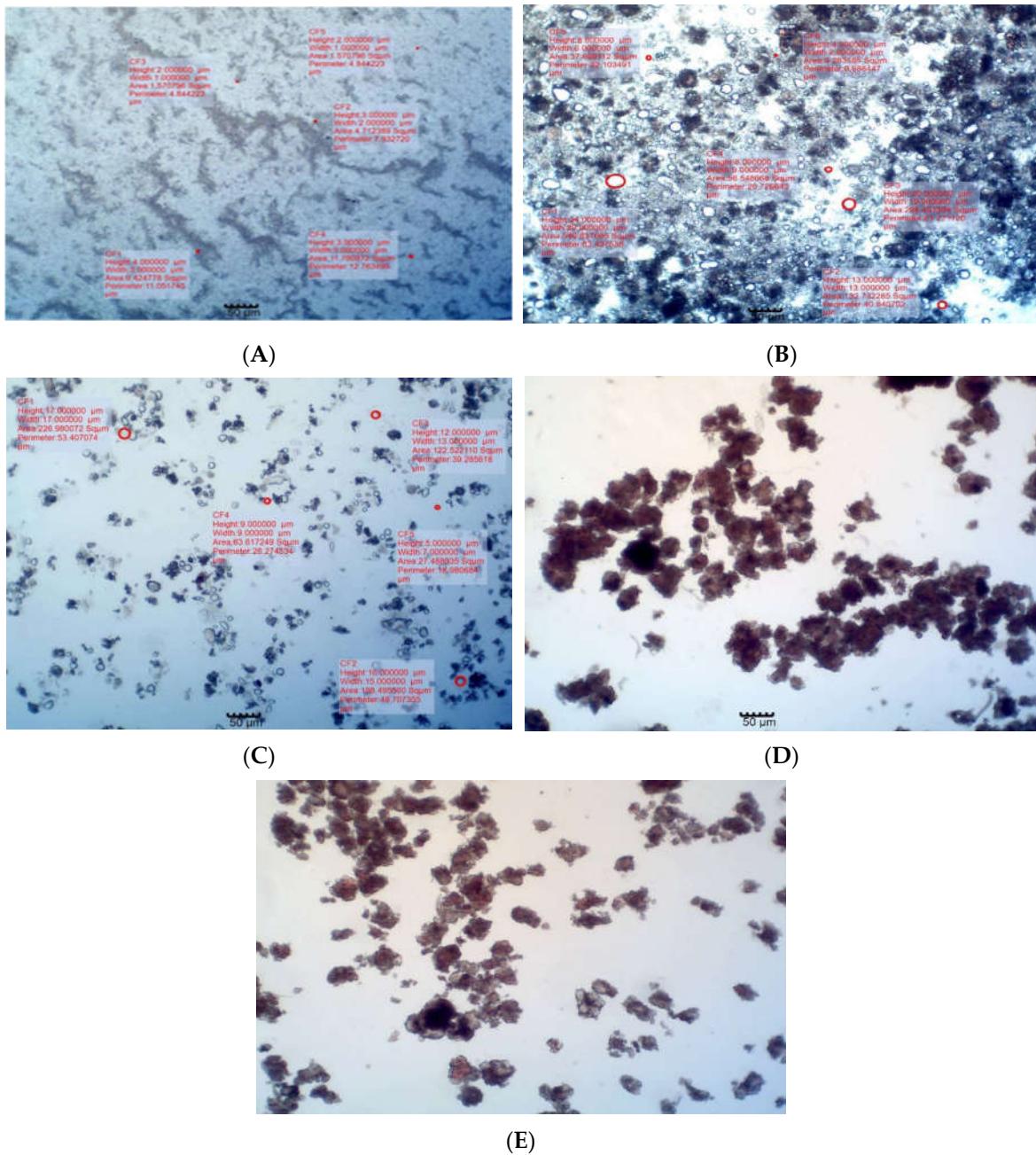
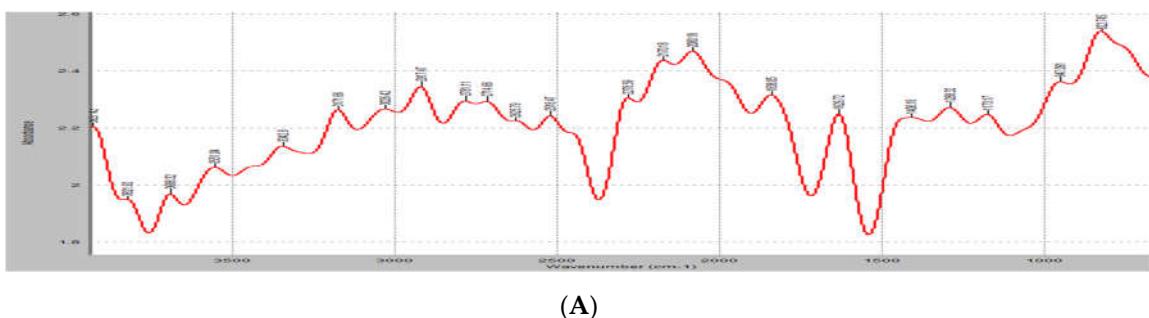


Figure 1. Morphology of the lipospheres. (A) = unloaded (placebo) liposphere; (B) = 2.5 g PM loaded liposphere; (C) = 5.0 g PM loaded liposphere; (D) = 7.5 g PM loaded liposphere; (E) = 10.0 g PM loaded liposphere.



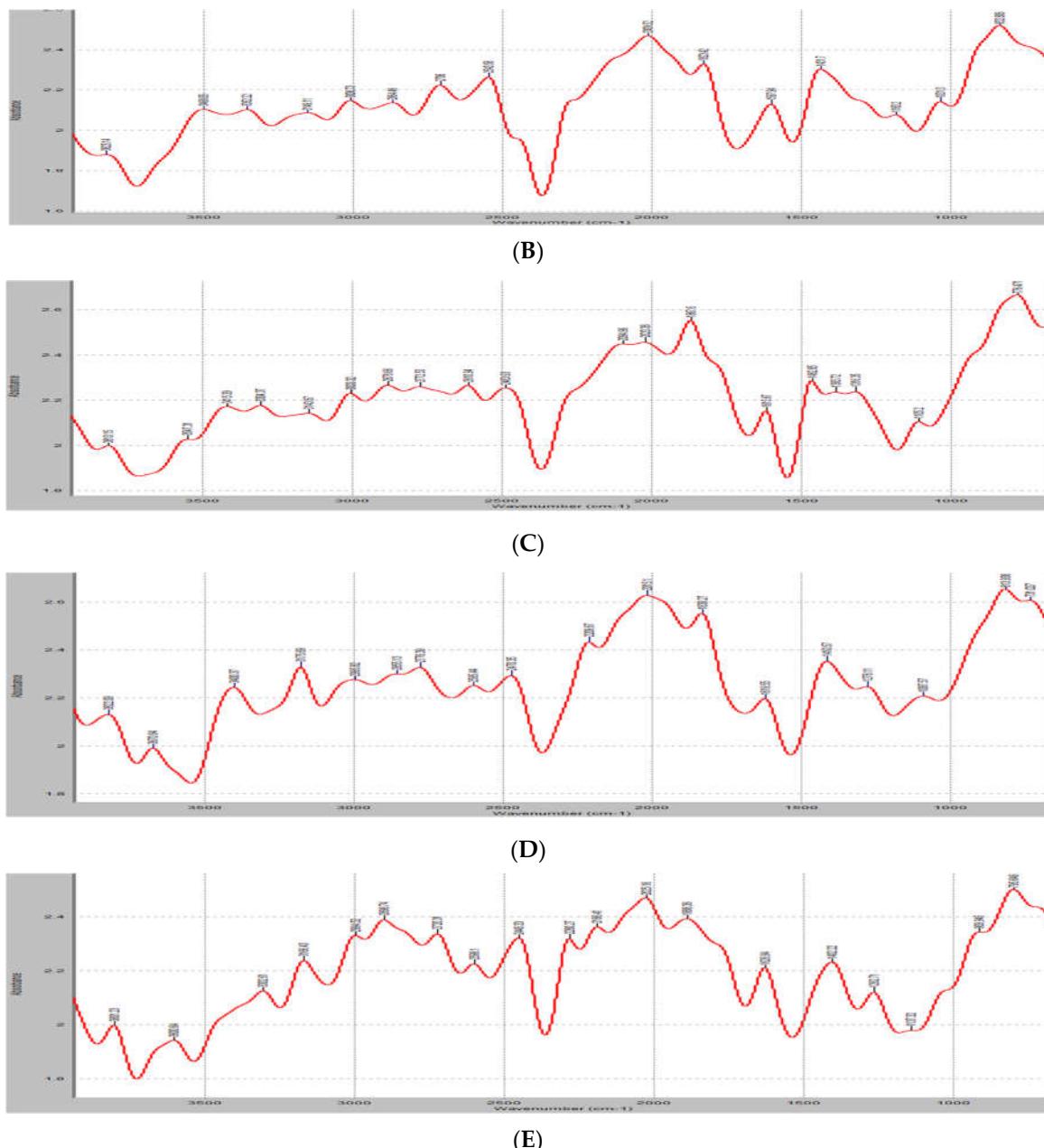


Figure 2. FTIR spectra of liposomes. (A-E) represents 0, 2.5, 5, 7.5, 10 g PM loaded liposomes groups) and 2002.75cm⁻¹ (C-O groups). Hence, the excipients used were compatible with the extract and there was no chemical interaction thereby improving the stability of the formulations.

3.5. *In-vitro* anti-inflammatory studies

3.5.1. Membrane stabilization

In Table 2, the percentage (%) inhibition of membrane stabilization by the samples and the standard drug were shown. Inhibition of membrane increased steadily with an increase in concentration of PM in the liposomes and the standard drug. However, after 400 µg/ml of sample, percentage inhibition fell drastically. The higher the percentage inhibition the more potent the concentration to stop the lysing of the red blood cells. The integrity of the cells is needed to maintain proper cellular function. In inflammation, lysis of some cellular component leads to release of inflammatory mediators. Stabilizing the red blood cell is a model to demonstrate the ability of the sample to stabilize those membranes of the stores of inflammatory mediators. Antioxidants play good role in this. The sample contains antioxidant activity which could account for the anti-inflammatory

activity [27]. Membrane stabilization assay of erythrocytes is a very popular tool to investigate the anti-inflammatory potential of a plant extract. When erythrocytes are exposed to hypotonic medium, heat, methyl salicylate, phenylhydrazine, among others, the lysis of their membrane occurs. It results in the leakage of serum protein and fluids into the tissues instigating inflammation [27]. Membrane stabilization leads to prevention of leakage of serum protein and fluids into the tissues during a period of increased permeability caused by inflammatory mediators [27]. So, the compounds capable of membrane stabilization might be very suitable as anti-inflammatory agents [22,28].

Table 2. Membrane stabilization of the Samples and the standard drug.

Sample (μg/ml)	% inhibition of membrane
50	24.60
100	51.70
200	100.0
400	300.0
800	200.0
Aspirin (μg/ml)	
50	89.00
100	5.90
200	100.00
400	100.00
800	106.25

In other words, the crude methanol extract of *Pentaclethra macrophylla* which contains copious amounts of bergenin demonstrated noticeable membrane stabilizing property [5]. Several flavonoids and triterpenes have been reported earlier to have anti-inflammatory activity [29]. As flavonoids and triterpenes are also present in *Pentaclethra macrophylla* methanol extract [30], it might be a reason for its membrane stabilizing anti-inflammatory potential.

3.5.2. Anti-platelet aggregation activity

In Table 3, the percentage anti-platelet aggregatory activity of platelets by the samples and the standard drug were shown. The platelet aggregatory inhibition decreased steadily with an increase in concentration of the PM in the liposomes as well as the standard. However, after 800 μg/ml of sample, percentage inhibition fell drastically without any effect on platelet. Antiplatelets prevent them from coming together as seen in inflammation (there is aggregation of platelets in inflammation).

Table 3. Anti-platelet aggregation activity.

Sample (μg/ml)	Change 1	Change 2	Change 3	Change 4
	Mean ± SEM	Mean ± SEM	Mean ± SEM	Mean ± SEM
50	0.007 ± 0.0025 ^{cd}	0.0043 ± 0.0013 ^{bcd}	0.0017 ± 0.0007 ^{bcd}	0.001 ± 0.00 ^b
100	0.004 ± 0.001 ^{abcd}	0.0047 ± 0.0017 ^{bcd}	0.005 ± 0.0021 ^{ab}	0.0037 ± 0.002 ^a
200	0.0013 ± 0.009 ^{ab}	0.002 ± 0.0006 ^{ab}	0.007 ± 0.0026 ^a	0.0023 ± 0.009 ^b
400	0.0017 ± 0.002 ^{ab}	0.0033 ± 0.0007 ^{bcd}	0.0007 ± 0.0003 ^c	0.0057 ± 0.002 ^a
800	0.000 ± 0.00 ^a	0.000 ± 0.00 ^a	0.00 ± 0.00 ^c	0.001 ± 0.001 ^b
Aspirin (μg/ml)				
50	0.005 ± 0.001 ^{abcd}	0.0063 ± 0.0018 ^c	0.0023 ± 0.009 ^{bcd}	0.002 ± 0.006 ^b
100	0.0033 ± 0.0007 ^{abc}	0.002 ± 0.0006 ^{ab}	0.0010 ± 0.00 ^c	0.0010 ± 0.006 ^b
200	0.004 ± 0.0021 ^{abcd}	0.003 ± 0.001 ^{abc}	0.003 ± 0.00 ^{bcd}	0.0013 ± 0.0007 ^b
400	0.0057 ± 0.002 ^{bcd}	0.0043 ± 0.0015 ^{abc}	0.0027 ± 0.0007 ^{bcd}	0.0013 ± 0.0003 ^b
800	0.009 ± 0.0012 ^d	0.0067 ± 0.0015 ^c	0.0027 ± 0.0007 ^{bcd}	0.0023 ± 0.0009 ^b

SEM=standard error of mean (n=5). In each column, values with same superscript do not differ significantly from each other at $p<0.05$. Change 1 is absorbance at 30 sec minus absorbance at zero sec; Change 2 is absorbance at 60 sec minus absorbance at 30 sec; Change 3 is absorbance at 90 sec minus absorbance at 60 sec; Change 4 is absorbance at 120 sec minus absorbance at 90 sec.

The aim is to enable platelet generate some inflammatory mediators that increases inflammation. Then the mechanism of anti-platelet aggregation could be by indirectly inhibiting cyclooxygenase-2 or by blockade of calcium influx through membrane calcium channels [31–33]. The model used showed that PM blocked calcium influx at acute time as shown by the result in Table 3.

3.6. *In-vivo* anti-inflammatory studies

3.6.1. 2% Formaldehyde induced arthritis model

The effect of *Pentaclethra macrophylla* herbal lipospheres on arthritis induced model of inflammation is as shown in Table 4. The PM-loaded lipospheres inhibited edema consistently throughout the duration of observation. Among the extracts, the 150 mg/kg (corresponding to the 7.5 g PM-loaded liposphere) had the highest inhibition of edema. There were significant differences between the test and positive controls. Arthritis which is marked by pains around the joints could be as a result of auto-immunity, which could be genetic or by excessive deposition of metabolite leading to gout. *Pentaclethra macrophylla* has been reported to have anti-inflammatory property. Farrukh et al [34] have implicated down-regulation of the expression of genes for COX-2, PGE2, IL-1 β , IL-6, TNF- α and NF- κ B and up-regulation of the expression of IL-4 and IL-10.

3.6.2. Egg albumin induced rat paw edema inflammatory model

Table 5 shows the effect of *Pentaclethra macrophylla* herbal lipospheres on egg albumin induced inflammation in mice. The test substances at 50, and 100mg/kg did not inhibit the inflammation throughout the observation time. This is clearly shown by a negative percentage inhibition across all the time intervals. At dose of 150mg/kg, there was slight inhibition initially, then after 2h, the inhibition disappeared as shown by the negative percentage inhibition at 3h. This same trend was seen in the negative control which shows no justifiable and significant differences in activity when compared to the test samples at the different doses. *Pentaclethra* has been reported to have anti-inflammatory property [33,34], and Farrukh et al [34] has implicated down-regulation of the expression of genes for COX-2, PGE2, IL-1 β , IL-6, TNF- α and NF- κ B and up-regulation of the expression of IL-4 and IL-10.

3.6.3. Blood Cells Counts of infected mice

The effect of *Pentaclethra macrophylla*-loaded lipospheres on the blood cells profile of infected mice is as shown in Table 6. The T50 dose of PM had the highest percentage of WBC and it decreased as the treatment doses increased from T100 to T200. There were no significant differences among the Neutrophil counts of the different groups. Similar trends was observed in the lymphocytes and Eosinophil counts. The monocytes and the basophils were undetected.

Table 4. The effect of *Pentaclethra macrophylla* loaded liposomes on arthritis model of inflammation in mice.

Treatments (mg/kg)	Volume of Edema (ml) (Percentage inhibition of Edema) over time (h)						
	0	1	2	3	4	5	48
T50	0.375 ± 0.025 (0.00)	0.750 ± 0.029 (7.14)	0.650 ± 0.029 ^a (0.00)	0.525 ± 0.025 ^a (0.00)	0.500 ± 0.00 ^b (16.7)	0.500 ± 0.00 ^b (0.00)	0.425 ± 0.048 (-6.2)
T100	0.300 ± 0.00 (6.7)	0.700 ± 0.00 (14.3)	0.600 ± 0.00 ^b (14.3)	0.525 ± 0.025 ^a (0.00)	0.525 ± 0.025 ^b (12.5)	0.475 ± 0.025 ^c (0.00)	0.400 ± 0.00 (0.00)
T150	0.325 ± 0.025 (10.0)	0.675 ± 0.025 (10.0)	0.575 ± 0.025 ^b (17.8)	0.525 ± 0.025 ^a (0.00)	0.525 ± 0.025 ^b (12.5)	0.500 ± 0.00 ^b (0.00)	0.425 ± 0.025 (-6.25)
T200	0.325 ± 0.025 (3.33)	0.725 ± 0.025 (14.29)	0.600 ± 0.00 ^b (14.29)	0.500 ± 0.00 ^b (4.8)	0.525 ± 0.025 ^b (12.5)	0.550 ± 0.029 ^b (10.0)	0.425 ± 0.025 (-6.25)
PC	0.325 ± 0.025 (6.7)	0.70 ± 0.04 (6.7)	0.60 ± 0.04 ^b (6.7)	0.50 ± 0.041 ^b (4.77)	0.475 ± 0.00 ^b (21.0)	0.40 ± 0.00 ^a (20.0)	0.40 ± 0.00 (0.00)
NC	0.30 ± 0.00	0.75 ± 0.041	0.70 ± 0.029 ^a (0.00)	0.525 ± 0.025 ^a (0.00)	0.60 ± 0.00 ^a (0.00)	0.50 ± 0.00 ^b (0.00)	0.40 ± 0.00 (0.00)
Normal	0.35 ± 0.05	0.75 ± 0.05	0.65 ± 0.00 ^a (0.00)	0.60 ± 0.00 ^a (0.00)	0.5 ± 0.025 ^b (0.00)	0.475 ± 0.025 ^c (0.00)	0.40 ± 0.00 (0.00)

T50, T100, T150, T200 = Treatments corresponding to the PM-loaded liposphere containing 2.5, 5, 7.5 and 10g of PM; PC = Positive Control (Diclofenac 20mg/kg); NC = Negative Control; Normal = No induction plus Vehicle. SEM=standard error of mean (n=5). In each column, values with same superscript do not differ significantly from each other at p<0.05.

Table 5. Effect of *Pentaclethra macrophylla* loaded liposomes on Egg albumin induced inflammation in mice.

Treatments (mg/kg)	Volume of Edema (ml) (Percentage inhibition of Edema) over time (h)					
	0	1	2	3	4	5
T50	0.350 ± 0.029 (-4.17)	0.625 ± 0.025 (-4.17)	0.550 ± 0.029 ^{abc} (-4.7)	0.500 ± 0.058 ^a (-17.65)	0.475 ± 0.048 ^a (-15.79)	0.650 ± 0.029 ^a (-8.33)
T100	0.275 ± 0.025 (-4.17)	0.625 ± 0.025 (-4.17)	0.600 ± 0.00 ^{ab} (-14.29)	0.525 ± 0.025 ^a (-23.5)	0.525 ± 0.025 ^b (-31.25)	0.650 ± 0.029 ^a (-4.17)
T150	0.300 ± 0.00 (16.6)	0.500 ± 0.00 (16.6)	0.500 ± 0.058 ^c (4.76)	0.433 ± 0.033 ^b (-1.88)	0.433 ± 0.033 ^a (-8.25)	0.633 ± 0.033 ^a (-5.5)
T200	0.267 ± 0.033 (0.00)	0.600 ± 0.058 (0.00)	0.533 ± 0.033 ^{ac} (-1.5)	0.500 ± 0.00 ^a (-17.64)	0.533 ± 0.033 ^b (-33.25)	0.700 ± 0.00 ^a (-6.0)
PC	0.30 ± 0.00 (4.17)	0.625 ± 0.025 (4.17)	0.500 ± 0.00 ^c (4.76)	0.425 ± 0.025 ^b (0.00)	0.425 ± 0.025 ^a (-6.25)	0.600 ± 0.00 ^b (0.00)
NC	0.30 ± 0.00	0.6 ± 0.00	0.525 ± 0.025 ^b (0.00)	0.425 ± 0.029 ^a (0.00)	0.400 ± 0.00 ^a (0.00)	0.60 ± 0.00 ^a (0.00)

Normal	0.275 ± 0.025	0.55 ± 0.029	0.625 ± 0.025 ^{ac}	0.575 ± 0.025 ^b	0.50 ± 0.041 ^a	0.65 ± 0.029 ^b
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T50, T100, T150, T200 = Treatments corresponding to the PM-loaded liposphere containing 2.5, 5, 7.5 and 10g of PM; PC = Positive Control (Diclofenac 20mg/kg); NC = Negative Control; Normal = No induction plus Vehicle. SEM=standard error of mean (n=5). In each column, values with same superscript do not differ significantly from each other at p<0.05.

Table 6. Effect of *Pentaclethra macrophylla* herbal lipospheres on the Blood Cells Counts of infected mice.

Treatments (mg/kg)	Blood Cell Count (%)					
	WBC	Neutrophil	Lymphocytes	Eosinophil	Monocyte	Basophil
T50	77.50 ± 3.59	65.50 ± 1.50 ^{bc}	34.00 ± 1.414 ^{abc}	0.50 ± 0.5	0.00	0.00
T100	70.00 ± 1.83	58.50 ± 1.71 ^a	40.00 ± 1.63 ^c	1.50 ± 0.5	0.00	0.00
T150	70.50 ± 1.71	61.50 ± 1.71 ^{abc}	37.50 ± 2.22 ^{abc}	1.00 ± 0.57	0.00	0.00
T200	67.50 ± 3.50	61.00 ± 1.29 ^b	38.50 ± 0.96 ^{bc}	0.50 ± 0.5	0.00	0.00
PC	71.00 ± 3.512	63.50 ± 3.403 ^{abc}	36.00 ± 3.747 ^{abc}	0.50 ± 0.50	0.00	0.00
NC	67.00 ± 2.646	67.25 ± 0.479 ^c	31.00 ± 0.577 ^a	1.75 ± 0.629	0.00	0.00
Normal 1	72.00 ± 4.320	61.50 ± 1.50 ^{abc}	36.50 ± 0.751 ^{abc}	2.0 ± 0.816	0.00	0.00
Normal 2	68.80 ± 2.062	65.00 ± 1.915 ^{bc}	32.00 ± 1.414 ^{ab}	2.50 ± 0.500	0.00	0.00

T50, T100, T150, T200 = Treatments corresponding to the PM-loaded liposphere containing 2.5, 5, 7.5 and 10g of PM; PC = Positive Control (Diclofenac 20mg/kg); NC = Negative Control; Normal 1 = No induction, no treatment; Normal 2 = No induction plus Vehicle. SEM=standard error of mean (n=5). In each column, values with same superscript do not differ significantly from each other at p<0.05.

4. Conclusions

There are associated side effects with delivery system and anti-inflammatory agents. There was therefore a need to investigate the potency of herbal liposphere from *Pentaclethra macrophylla*. The excipients used were compatible with the extract and there was no chemical interaction thereby improving the stability of the formulations. The characterization of the herbal lipospheres formed presented good qualities and hence a promising delivery system for the herbal formulation of *Pentaclethra macrophylla* which is a potent drug that ameliorates different ailments. The study showed that *Pentaclethra macrophylla* was able to produce efficient herbal lipospheres with antinociceptive and anti-inflammatory activities.

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References

1. Viana FA, Pouliquen YBM, Andrade-Neto M, et al. Complete ¹H and ¹³C NMR assignments for two new monodesmoside saponins from *Pentaclethra macroloba* (Willd.) Kuntze. *Magn Reson Chem.* 2004; 42(8):695-699.
2. Pertuit D, Kapundu M, Mitaine-Offer AC, Miyamoto T, Tanaka C, Delaude C, and Lacaille-Dubois M-A. Triterpenoid Saponins From the Stem Bark of *Pentaclethra eetveldeana*. *Natural Product Communications* 2019; 1-5.
3. Babady-Byla, Herz W. Triterpenes and 1-(ω -hydroxyceratyl) glycerols from *Pentaclethra eetveldeana* root bark. *Phytochemistry.* 1996; 42(2):501-504.
4. Viana FA, Braz-Filho R, Pouliquen YBM, Andrade Neto M, Santiago GMP, Rodrigues-Filho E. Triterpenoid saponins from stem bark of *Pentaclethra macroloba*. *J Braz Chem Soc.* 2004; 15(4):595-602.
5. Chinaka C. Nnennaya, Garuba A. Sunday and Augustine A. Ahmadu. Chemical Constituents from the Stem Bark of *Pentaclethra macrophylla* Benth (Fabaceae). *Nig.J.Pharm. Res.* 2017; 13 (1):37-44.
6. Folefoc GN, Bissek JP, Fomum ZT and Bodo B. A new Secokaurane diterpenoid and O-glucoside from seeds of *Pentaclethra macrophylla*. *J Cameroon Academy of Science* 2004; 4(3): 227-231.
7. Akindahunsi, A. A. (2004). Physicochemical studies on African oil bean (*Pentaclethra macrophylla* Benth.) seed. *Journal of Food Agriculture and Environment*, 2: 14 - 17.
8. Asoegwu S, Ohanyere S, Kanu O. and Iwueke C. "Physical Properties of African Oil Bean Seed (*Pentaclethra macrophylla*)" Agricultural Engineering International: the CIGR Ejournal. Manuscript FP 05 006. Vol. VIII. August, 2006.
9. Nnamani PO, Kenechukwu FC, Asogwa FO, Momoh MA, Lehr C-M, Attama AA. Novel Anti-Ulcer Phytosomal Formulation of Ethanol Extract of *Pentaclethra macrophylla* Stem-Bark. *Trop J Nat Prod Res.* 2020; 4(8):385-391. doi.org/10.26538/tjnpnpr/v4i8.11
10. Igbe I. and Osigwe C. Hypoglycaemic activity of aqueous extract of *Pentaclethra macrophylla* Benth. (Fabaceae) stem bark in streptozotocin-induced diabetic rats. *J of Pharmacy and Bioresources.* 2012; Vol. 9 no. 1, pp. 39-44.
11. Ukor B., Chike C.P.R, Olorunfemi O.J. Effects of Aqueous Extract of *Pentaclethra macrophylla* Leaves on Blood Pressure and Oxidative Stress in Albino Wistar Rats. *International Journal of Scientific Research and Engineering Development* 2020; Volume 3 Issue 2.
12. Iwu, I.C., Uchegbu R. Isolation and characterization of 4, 15- Cholestene-3-benzoate from the stem bark of *Pentaclethra macrophylla* (P. Benth), *International Journal of Emerging Knowledge.* 2014; 1(10):185-192

13. Tico Etnobotanical Dictionary. 2005. *Pentaclethra macrophylla* (Benth). <http://www.ars-grin.gov/duke/dictionary/tico/p.html>
14. Okunrobo L. O., Ching F. P. and Ifijeh F. Antinociceptive activity of methanol extract and aqueous fraction of the stem bark of *Pentaclethra macrophylla* Benth (Mimosaceae). Journal of Medicinal Plants Research 2009; 3(3):101-104.
15. Chidozie JI. African oil bean seed as an essential food supplement in the control of cancer and tobacco related diseases. Proceedings from the U1CC World Cancer Congress, July 8-12, Washington DC, 2006..
16. Balogun, B.I. Evaluation of the Nutritional Potentials of Fermented Oil Beans Seed *Pentaclethra macrophylla* Benth. PAT December, 2013; 9 (2): 73-87 ISSN: 0794-5213.
17. Akah, P.A., Aguwa, C.N., Agu, R.U. Studies on the antidiarrhoeal properties of *Pentaclethra macrophylla* leaf extracts. Phytother. Res. 1999; 13 (4):292-295
18. Alinnor, I. J. and Oze, R.. Chemical Evaluation of the Nutritive Value of *Pentaclethra macrophylla* (African Oil Bean) seed. Pakistan Journal of nutrition, 2011; 10: 355 - 359.
19. Osabor, VN, Pkpnkwo, PC and Ikeuba AI. Chemical profile of leaves and seeds of *Pentaclethra macrophylla* Benth. J Medinal Plant and Herbal Therapy Research 2017; 5:11-17.
20. Githens, TS. African Handbooks – Drug Plants of Africa. University of Pennsylvania Press. 1948: P. 64.
21. Nnamani, P. O., Ibezim, E. C., Attama, A. A. and Adikwu, M. U. Surface modified solid lipid microparticles based on homolipids and softisan® 142: preliminary characterization. Asian Pacif. J. Trop. Med. 2010; 3(3)205-210. doi.org/10.1016/S1995-7645(10)60010-7
22. Yesmin, S., Paul, A., Naz, T., Rahman, A. B. M. A., Akhter, S. F., Wahed, M. I. I., Emran, T. bin,& Siddiqui, S. A. Membrane stabilization as a mechanism of the antiinflammatory activity of ethanolic root extract of Choi (Piper chaba). Clinical Phytoscience, 2020 6(1). <https://doi.org/10.1186/s40816-020-00207-7>
23. Sur T. K, Biswas T. K., Ali L., Mukherjee B. Anti-inflammatory and anti-platelet aggregation activity of human placental extract. Acta Pharmacol Sin. 2003; 24(2):187-92.
24. Ying-Ying Zheng, Ting-Ting Wu, Yi Yang, Xian-Geng Hou, Ying Gao, You Chen, Yi-Ning Yang, Xiao-Mei Li, Xiang Ma, Yi-Tong Ma, Xiang Xie. Personalized antiplatelet therapy guided by a novel detection of platelet aggregation function in stable coronary artery disease patients undergoing percutaneous coronary intervention: a randomized controlled clinical trial. Eur Heart J Cardiovasc Pharmacother 2020 Jul 1;6(4):211-221. doi: 10.1093/ehjcvp/pvz059.
25. Nnamani, P.O., Attama, A.A., Kenechukwu, F.C., Ibezim, E.C. and Adikwu, M.U. Pharmacodynamics of Piroxicam from Novel Solid Lipid Microparticles Formulated with Homolipids from *Bos indicus*. Current Drug Delivery 2013;10(6) 645-655. DOI: 10.2174/156720181006131125145712
26. Ochei,J.and Kolhatkar,A. Medical LaboratoryScience Theory and Practice. Tata McGraw Hill Publishing Company Limited.New Dell.2008
27. Halliwell B, Whiteman M. Measuring reactive species and oxidative damage in vivo and in cell culture: How should you do it and what do the results mean? Br J Pharmacol. 2004;142:231–55.
28. Chaitanya R, Sandhya S, Banji D, Ravindran V and Murali S. "HRBC Membrane Stabilizing Property of Root, Stem and Leaf of *Glochidion elutinum*". International Journal of Research in Pharmaceutical and Biomedical Sciences 2.1 (2011): 256-259
29. Shinde UA, Phadke AS, Nair AM, Mungantiwar AA, Dikshit VJ, Saraf MN. Membrane stabilizing activity – a possible mechanism of action for the anti-inflammatory activity of *Cedrus deodara*wood oil. Fitoterapia.2008;70:251–7.
30. Serafini M, Peluso I, Raguzzini A. Flavonoids as anti-inflammatory agents. Proc Nutr Soc. 2010;69:273–8.
31. Beknal AK, Korwar PG, Halkai M, Kulkarni U, Patil BS, Soodam SR. Phytochemical investigation and antioxidant activity study of *Drynaria quercifolia* Linn. rhizome. Int J Curr Pharmaceut Res. 2010;4:36–9.
32. Shah, B. H., Safdar, B., Virani, S. S., Nawaz, Z., Saeed, S. A. & Gilani, A. H. The anti-platelet aggregatory activity of *Acacia nilotica* is due to blockade of calcium influx through membrane calcium channels. Gen Pharmacol. 1997;29(2): 251-5
33. Masafumi, Z., Yuhei, H., Shinji, N., Muneaki, M., Ichiro, F. & Eiichi, I. Thromboxane synthesis is increased by upregulating of cytosolic phospholipase A2 and cyclooxygenase-2 in peripheral polymorphonuclear leucocytes during bacterial infection in childhood. Am J Haematol 2003; 72 (2): 115-20.
34. Farrukh M., Saleem U., Qasim, M., Manan, M., Shah, MA. *Sarcococca saligna* extract attenuates formaldehyde-induced arthritis in Wistar rats via modulation of pro-inflammatory and inflammatory biomarkers. Inflammopharmacology 2022; 30(2): 579-597.

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