

Review

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Review

Exploration of Practical Biomarkers for Systemic Drug Therapy for Hepatocellular Carcinoma

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Simple Summary: A number of agents, including immune checkpoint inhibitors, have become available for the treatment of hepatocellular carcinoma, but the objective response rate of these drugs is currently only 30% to 40%. Therefore, the identification of new predictive biomarkers and an increased knowledge of the mechanisms of response or resistance to systemic chemotherapies are required.

Abstract: A number of agents, including immune checkpoint inhibitors, have become available for the treatment of hepatocellular carcinoma (HCC). However, the objective response rate of these drugs is currently only 30% to 40%, with a high incidence of side effects. There are also no practical biomarkers to predict their therapeutic effects. Most of the systemic therapies for HCC are performed in general hospitals without research facilities. Such hospitals can perform imaging tests, like CT and MRI, as well as pathological diagnosis using tumor tissue sampling and immunohistochemical staining. However, analyzing tumor genomic or transcriptomic profiles is difficult because of limitations in facilities, personnel, and cost. Therefore, in this review, we provide an overview of the wide range of research that has been conducted on HCC biomarkers from blood, tissue, or imaging information that can be used practically in general hospitals for predicting the therapeutic effect of systemic therapies before treatment begins. For general hospitals that treat HCC patients, we recommend conducting treatment after assessing the state within the tumor tissue as much as possible by collecting blood and tissue samples and performing pre-treatment MRI image evaluations.

Keywords: hepatocellular carcinoma; tyrosine kinase inhibitors; immune checkpoint inhibitors; biomarkers

1. Introduction

Liver cancer is a major cause of death worldwide, and the number of people diagnosed with liver cancer is expected to increase [1]. As the principal histologic type of liver cancer, hepatocellular carcinoma (HCC) is responsible for the great majority of liver cancer diagnoses and deaths, accounting for approximately 75% of the total [2]. Treatment options for localized HCC, such as surgical resection, ablation, liver transplantation, and transarterial chemoembolization (TACE), were established in the 20th century, but an effective drug therapy for advanced HCC did not emerge until 2007 [3]. Although many clinical trials of potential drug therapies for unresectable HCC were conducted before the introduction of tyrosine kinase inhibitors (TKIs), no chemotherapeutic drugs demonstrated any significant survival benefit, as shown in the meta-analysis by Mathurin et al. [4]. Subsequently, an advanced understanding of the mechanisms of tumor cell proliferation and angiogenesis supported the development of the TKI sorafenib [5]. In the clinical trial "SHARP trial" for unresectable HCC, sorafenib showed a clear survival benefit over placebo and became the standard treatment for unresectable HCC in 2007 [6]. Subsequently, the results of the "RESORSE trial", a clinical trial limited to unresectable HCC patients who have tolerated sorafenib led to the approval of regorafenib as a second-line therapy after sorafenib treatment in 2017 [7]. In 2018, lenvatinib demonstrated "non-inferiority" to sorafenib in the "REFLECT trial", leading to a choice of either sorafenib or lenvatinib as the first-line therapy [8]. In 2019, based on the results of the "REACH-2 trial" for cases with alpha-fetoprotein (AFP) levels of 400 ng/mL or more after sorafenib treatment,

ramucirumab (an anti-vascular endothelial growth factor (VEGF) receptor antibody) became available as a second-line therapy [9]

In the following years, immune checkpoint inhibitors (ICIs) emerged. In 2020, a combination therapy of atezolizumab and bevacizumab, which surpassed sorafenib in clinical trials, was introduced [10]. As a result, both sorafenib and lenvatinib were relegated to second-line or later therapies. Furthermore, a combination therapy of two ICIs, durvalumab and tremelimumab (STRIDE regimen), outperformed sorafenib in treatment results of the "HIMALAYA trial". Durvalumab monotherapy also showed non-inferiority to sorafenib [11], allowing these therapeutics to be added as new treatment options. Additionally, a TKI, cabozantinib, was developed as a second-line or later therapy [12].

Through these trials, a number of agents, including ICIs, have become available for treating HCC. However, the objective response rate (ORR) of these drugs is currently only 30% to 40%, with a high incidence of side effects [6–12]. Other than ramucirumab, which has the condition of AFP 400 ng/mL or more, there are no practical biomarkers to predict the therapeutic effects of these treatment methods.

Most of the systemic therapies for HCC are performed in general hospitals without research facilities. In such hospitals, it is possible to perform imaging tests, such as CT and MRI, and pathological diagnosis, including tumor tissue sampling and immunohistochemistry (IHC) staining. However, analyzing the tumor genomic or transcriptomic profiles is often difficult because of limitations in facilities, personnel, and cost, as well as increased medical costs. Therefore, in this review, we provide an overview of the wide range of research that has been conducted on HCC biomarkers from blood, tissue, or imaging information that can be used practically in general hospitals for predicting the therapeutic effect of systemic therapies before treatment begins. We also examine and describe future prospects in this field.

2. Exploring Biomarkers for Predicting the Therapeutic Effects of TKIs (Table 1)

2.1. Sorafenib Biomarkers

The first TKI, sorafenib, has remained the first-choice drug for a decade, leading to numerous early studies being conducted on biomarkers for predicting its therapeutic effect.

One of the objectives of the Phase III SHARP (Sorafenib HCC Assessment Randomized Protocol) trial was to investigate plasma biomarkers for predicting prognosis and therapeutic effect, with 10 plasma biomarkers measured at baseline and after 12 weeks of treatment [13]. As a result, baseline angiopoietin 2 (Ang-2) and VEGF concentrations were found to be independent predictors of survival in the entire advanced HCC patient population. However, these were not unique to the sorafenib cohort and were similar in the placebo cohort [13].

In a real-world study in Japan, Miyahara et al. measured the serum levels of eight pro-angiogenic cytokines (Ang-2, follistatin (FST), granulocyte colony-stimulating factor (G-CSF), hepatocyte growth factor (HGF), leptin, platelet-derived growth factor-BB (PDGF-BB), platelet endothelial cell adhesion molecule-1 (PECAM-1), and VEGF) in 120 consecutive HCC patients treated with sorafenib. They reported that high expression of Ang-2 or high expression of three or more pro-angiogenic cytokines was associated with poor progression-free survival (PFS) and overall survival (OS) in patients treated with sorafenib [14]. In addition, the presence of macrovascular invasion (MVI) was also shown to be associated with poor OS in clinical parameters [14].

Table 1. Factors influencing patient prognosis or efficacy for treating hepatocellular carcinoma with tyrosine kinase inhibitors.

Agent	Study design	Number of cases	Prognostic and predictive factors	Outcome	Statistical analysis	HR [95% CI]	P-value	Authors [reference no.]
Sorafenib	Retrospective, single-arm	120	[High serum Ang-2]	PFS OS	Univariate Multivariate	1.84 [1.21–2.81] 1.83 [1.12–2.98]	0.004 0.014	Miyahara K et al. [14]

			[High angiogenic group*]	PFS	Univariate	1.98 [1.30–3.06]	0.001	
			*: patients with > three serum cytokines (Ang-2, FST, G-CSF, HGF, Leptin, PDGF-BB, PECAM-1, or VEGF)	OS	Multivariate	1.76 [1.07–2.94]	0.023	
				OS	Multivariate	2.27 [1.36–3.72]	0.001	
			[MVI (present)]					
	Retrospective							
Sora-fenib	pooled analysis of two phase 3 trials (vs. placebo)	Sorafenib 448	[Without EHS]	OS	Multivariate	0.55 [0.42–0.72]	0.015	Bruix J et al. [15]
		Placebo 379	[With HCV]	OS	Multivariate	0.47 [0.32–0.69]	0.035	
			[Low NLR]	OS	Multivariate	0.59 [0.46–0.77]	0.0497	
Sora-fenib	Subgroup meta-analyses, single-arm	170	[Low NLR]	OS	Univariate	1.49 [1.17–1.91]	0.001	Qi X et al. [18]
Sora-fenib	Observational registry, single-arm	3,371	[Child-Pugh A]	OS	Kaplan-Meier	-	N/A	Marrero JA et al. [19]
			[Bilirubin]	OS	Univariate	1.71 [1.57–1.86]	N/A	
			[Albumin]	OS	Univariate	1.76 [1.63–1.89]	N/A	
Sora-fenib	Retrospective, single-arm, HCV patients only	103	[HCV eradication]	OS	Multivariate	0.46 [0.26–0.78]	0.004	Kuwano A et al. [20]
			[ALBI score]	OS	Multivariate	2.29 [1.20–4.37]	0.012	
Sora-fenib	Population-based retrospective cohort, HCV patients only, single-arm	1,684	[DAA user]	OS	Univariate PSM univariate	-	< 0.0001	Tsai H-Y et al. [21]
							< 0.0001	
Sora-fenib	Retrospective, single-arm	55	[FGF3/FGF4 amplification] (frozen tumor tissue)	CR/PR	Fisher's exact	-	0.006	Arao T et al. [26]
			[multiple lung metastases]	CR/PR	Fisher's exact	-	0.006	
Sora-fenib	Retrospective, single-arm	20	[High miR-224 expression] (FFPE tumor tissue)	PFS	Univariate	0.28 [0.09–0.92]	0.029	Gyöngyösi B et al. [28]
				OS	Univariate	0.24 [0.07–0.79]	0.012	
Sora-fenib	Retrospective, single-arm	Training coh. 26 Valid. coh. 58	[High miR-425-3p expression] (FFPE tumor tissue)]	TTP	Multivariate	0.4 [0.1–0.7]	0.002	Vaira V et al. [31]
				PFS	Multivariate	0.3 [0.1–0.7]	0.0012	
Sora-fenib	Retrospective validation of the pharmacogenomics panel, single-arm	54	[High serum DKK-1]	PFS	Univariate	-	0.0396	Qiu Z et al. [33]
				OS	Univariate	-	0.0171	

						1.52 [1.08–2.13]		
						*		
						1.21 [1.01–1.45]		
Lenva- tinib	Retrospective validation of the experimentally identified biomarker (vs. sorafenib)	Lenvatinib 65 (ST6GAL1 high 22, low 43) Sorafenib 31 (ST6GAL1 high 12, low 19)	[Serum ST6GAL1 high]	OS	Univariate		< 0.05	Myojin Y et al. [55]

HR, hazard ratio; CI, confidence interval; PFS, progression-free survival; OS, overall survival; CR, complete response; PR, partial response; TTP, Time to Progression; DCR, disease control rate; HBV, hepatitis B virus; HCV, hepatitis C virus; MVI, macrovascular invasion; EHS, extrahepatic spread; NLR, neutrophil to lymphocyte ratio. Note: In the 'Statistical analysis' section, 'univariate' typically refers to the Kaplan-Meier method and log-rank test, while the inclusion of HR indicates the use of Cox regression. Additionally, 'multivariate' typically refers to the utilization of the multivariate Cox regression model.

In an analysis of pooled data from the SHARP and Asia Pacific (AP) Phase III trials, a significantly greater OS benefit compared with placebo was observed in patients without extrahepatic spread (EHS; hazard ratio (HR), 0.55 vs. 0.84), with hepatitis C virus (HCV) (HR, 0.47 vs. 0.81), and a low neutrophil to lymphocyte ratio (NLR) (HR, 0.59 vs. 0.84) [15]. In this analysis, the NLR was divided into more than 3.0 and less than 3.0, which was the median in the sorafenib administration group. Although this was a retrospective study, it described certain factors that had significant differences compared with the placebo group. However, these findings were strictly predictions of the survival period and not of drug effectiveness.

The NLR reflects the inflammatory response to cancer, and its elevation is recognized as an indicator of poor prognosis [16,17]. Qi et al. conducted a systematic review and meta-analysis of 20,475 HCC patients from 90 articles to explore the prognostic role of NLR in HCC, finding that a lower baseline NLR was significantly associated with the survival period (HR, 1.80, 95% confidence interval (CI): 1.59–2.04, $P < 0.00001$) [18]. This study also conducted a subgroup meta-analysis for cases treated with sorafenib, and although it was not compared with a placebo group, low NLR was associated with better survival in 170 cases between high and low NLR groups [18].

The GIDEON trial, a large prospective observational registration study focused on assessing the safety of sorafenib treatment in the real world, showed better OS in Child-Pugh A patients compared with Child-Pugh B patients. Moreover, a univariate Cox regression analysis of each factor of the Child-Pugh score showed that albumin and bilirubin, which form the ALBI score, strongly influenced OS [19]. The authors did not utilize anything other than descriptive statistics that considered the impact of selection bias. Additionally, this study lacked a control group and was not randomized. However, liver functional reserve can clearly affect the patient survival period after treatment with sorafenib for HCC. For example, several studies have shown that in advanced HCC cases related to HCV, OS is extended as liver functional reserve improves following HCV eradication with interferon (IFN) or direct-acting antivirals (DAA) [20,21].

For pathological biomarkers in tumor tissues, the Phase II trial results indicated that phosphorylated ERK could potentially be useful as a biomarker for predicting the prognosis of patients treated with sorafenib. This protein is located downstream of the Raf kinase in the MAPK cascade, which is a major target of sorafenib [22]. While there have been studies in the clinical setting that have shown favorable efficacy [23], several have also shown unfavorable efficacy [24,25]. Therefore, no consensus has been reached regarding phosphorylated ERK.

Arao et al. reported that in 13 cases that showed significant tumor shrinkage with sorafenib treatment, fibroblast growth factor (FGF)3/FGF4 amplification was observed in the tumor genome. Additionally, multiple lung metastases in poorly differentiated histological types were seen as

clinical pathological features. Although the sample size was relatively small, FGF3/FGF4-amplified tumors were frequently observed in responders to sorafenib [26]. Although this study examined biomarkers for sorafenib treatment efficacy, no further research with additional cases to support these findings was conducted.

Tumor tissues can also be used to investigate microRNA (miRNA) expression. MiRNAs are small endogenous non-coding RNAs that can inhibit translation or support cleavage of mRNAs to negatively regulate gene expression. These molecules are highly stable and can be reliably detected in stored clinical samples and cell cytology specimens, making them ideal biomarker candidates [27–29]. Various miRNAs are also mechanistically involved in the development, proliferation, and progression of HCC and can be detected in serum and plasma samples, suggesting they could potentially be used as diagnostic markers [30].

Gyöngyösi et al. investigated the expression levels of 14 miRNAs in 20 HCC cases where tumor tissue samples were collected by fine needle aspiration before sorafenib administration, with the data demonstrating that high expression of miR-224 was associated with increased PFS and OS rates [28].

Vaira et al. conducted a comprehensive profiling of approximately 700 miRNAs in a series of 26 HCC patients treated with sorafenib (training set) using tumor tissues collected prior to treatment, then verified the results in an independent series of 58 patients (validation set) [31]. As a result, six miRNAs were found to be significantly associated with clinical variables in the training set. Of these, only miR-425-3p was significant in the validation set, with high miR-425-3p levels being associated with longer time to progression (TTP) and PFS [31]. However, no follow-up studies have been conducted. In general, miRNA-related cancer research is primarily focused on the development of therapies that target specific miRNAs or use these molecules as a tool for early cancer detection [32].

Other studies have constructed high-throughput assay systems in completely different ways. Qiu et al. created a Liver Cancer Model Repository (LIMORE) panel of 81 cell lines by creating 50 patient-derived liver cancer cell lines, in addition to 31 existing cell lines, to model HCC heterogeneity. The authors examined the sensitivity of the cells to a total of 90 drugs. By using this panel, which has verified gene mutations and gene expression characteristics, it is possible to identify gene-drug interactions of therapeutic methods and biomarker candidates. When predicting the effect of sorafenib treatment, a molecule called Dickkopf-1 (DKK-1) was identified as a potential useful biomarker [33]. Interestingly, DKK-1 is a secreted protein that antagonizes Wnt signal transduction, which is known to affect ICI efficacy [34]. Because DKK-1 is a serum protein, it may be relatively easy to verify its potential as a biomarker in existing cohorts with preserved serum samples.

2.2. Regorafenib Biomarker Studies

Although insufficient results were obtained from the biomarker studies in the Phase III SHARP trial for sorafenib, a far more comprehensive exploratory biomarker analysis was conducted for patients in the RESORCE trial at the DNA, RNA, and protein levels [35]. Of the 266 proteins studied in baseline plasma samples, decreases in five, Ang-1, cystatin B, the latency-associated peptide of transforming growth factor beta 1 (LAP TGF- β 1), oxidized low-density lipoprotein receptor 1 (LOX-1), and C-C motif chemokine ligand 3 (MIP-1 α), were found to be associated with extended TTP and OS. Moreover, nine plasma miRNAs, miR-30a, miR-122, miR-125b, miR-200a, miR-374b, miR-15b, miR-107, miR-320, and miR-645, were related to OS, although none were associated with TTP. Furthermore, there was no apparent correlation between the AFP or c-MET protein expression levels and the OS or TTP benefits of regorafenib treatment, causing them to be excluded as potential predictive biomarkers [35]. Currently, with treatments including ICIs becoming the standard of care as first-line treatments, the number of cases where regorafenib is used after sorafenib treatment is likely not substantial. This could make planning prospective validation studies challenging. However, as these potential biomarkers are plasma proteins and miRNAs, it is hoped that they can be validated in existing cohorts using blood samples that have been preserved.

2.3. Signaling Pathways as Biomarkers for TKIs: Insights from Trials with mTOR and MET Inhibitors

In several cancer types other than HCC, there are driver gene mutations in signaling pathways that strongly promote tumor growth. This has led to established biomarker-driven treatment concepts for drug selection and predicting treatment outcomes [36–38]. In HCC, signaling pathways such as RAS, mammalian target of rapamycin (mTOR), MET, and FGF-19 have been considered as potential therapeutic targets. Several clinical trials were conducted using inhibitors targeting these pathways, but many unfortunately ended with disappointing results [39]. For example, mTOR signaling is activated in about half of all HCC cases and is associated with worse outcomes [40]. Despite this strong theoretical basis for using the mTOR inhibitor everolimus for treating HCC, the final results from a phase III trial did not suggest any trend of prolonged OS (everolimus vs. placebo, 7.6 vs. 7.3 months) [41].

The potential cause of failure in these clinical trials may be the inclusion of all patients with unresectable HCC. It has been considered desirable to incorporate molecular selection factors into prospective research, as in clinical trials for other cancers [42]. One of the drugs for which there was hope for biomarker-driven treatment in HCC was tivantinib. Tivantinib, a MET inhibitor, was investigated in a phase II trial against placebo as a second-line treatment. Although no significant effect was observed in all cases, it improved survival rates in patients with high tumor MET expression levels [43]. From these results, phase III trials (METIV-HCC, JET-HCC) were conducted comparing tivantinib and placebo only in patients with high MET expression [44,45]. However, in both trials, no statistically significant treatment effect was observed for tivantinib compared with placebo.

Cabozantinib, which targets several TKs including MET, VEGF, and AXL, was successful in a phase III trial (CELESTIAL) [12]. In this trial, baseline plasma levels of MET, AXL, VEGFR2, HGF, GAS6, VEGF-A, PIGF, IL-8, EPO, ANG2, IGF-1, VEGF-C, and c-KIT were evaluated as biomarkers, but none of them could predict the treatment effect of cabozantinib on OS or PFS [46].

HCC has diverse causes, including viral infection, toxin exposure, and metabolic disorders. From the results of large-scale genomic analyses, it has become clear that gene mutations in HCC are centered on non-drug targetable mutations, such as TERT, CTNNB1, TP53, that are diverse [47,48]. Furthermore, HCC is heterogeneous even within an individual patient, and sequencing analysis of a single lesion cannot fully characterize the genomic features of HCC in certain cases [49]. The failure to use the expression patterns of specific therapeutic target molecules or activated signaling as biomarkers in TKI clinical trials may indirectly suggest that the heterogeneity and diversity of HCC results in a lack of full dependence on the several signaling pathways that have been considered [50].

2.4. New Approaches for Biomarker Discovery in Lenvatinib Treatment

Lenvatinib is a multi-kinase inhibitor that inhibits vascular endothelial growth factor receptors (VEGFR) 1–3, fibroblast growth factor receptors (FGFR) 1–4, platelet-derived growth factor receptor (PDGFR) α , and oncogenes RET and KIT [51]. Preclinical studies have shown that lenvatinib has strong anti-angiogenic activity, primarily through inhibition of the VEGF and FGF signaling pathways [52].

In a subgroup analysis of the REFLECT trial, patients with HBV infection or alcohol as underlying factors showed better PFS rates with lenvatinib treatment than with sorafenib [8]. However, it seems that no biomarker exploration beyond subgroup analysis was planned in the REFLECT trial.

Tada et al. focused on the associations between outcomes in HCC patients treated with lenvatinib and the NLR. In a multivariate analysis of a cohort of 237 individuals, an NLR ≥ 4 was independently associated with OS and PFS. There was also a significant difference in the disease control rate between patients with low NLR (< 4) and high NLR (≥ 4) (85.5% vs. 67.3%, $P = 0.007$). A spline curve analysis showed that an NLR of approximately 3.0 to 4.5 was an appropriate cutoff value related to OS (53). In addition, in the retrospective RELEVANT study from 23 other facilities, data were collected for 1,325 patients treated with lenvatinib. In the multivariate analysis of OS, HBsAg positivity, NLR > 3 , and AST > 38 were independently associated with poor prognosis in all three

models. Furthermore, nonalcoholic fatty liver disease (NAFLD)/nonalcoholic steatohepatitis (NASH)-related etiology was independently associated with a good prognosis. The multivariate analysis showed that NAFLD/NASH, Barcelona Clinic Liver Cancer (BCLC) stage, NLR, and AST were independent prognostic factors for PFS in cases treated with lenvatinib [54]. However, these studies did not make comparisons with placebo or other drug treatments. Essentially, a control group or placebo group is necessary to identify predictors of the therapeutic effect of a certain drug, so it is necessary to consider the limitations of such single-arm observational studies when evaluating predictive markers.

Qiu et al., who identified DKK-1 as a potential predictor of sorafenib effectiveness, confirmed with their panel of 81 HCC cell lines, LIMORE, that FGFR inhibitors like lenvatinib have a favorable effect on HCC strains with amplification of FGFR and FGF. They suggested that the amplification of FGF19 and FGFR could potentially serve as biomarkers for lenvatinib effectiveness [33].

Myojin et al. developed a new HCC mouse model that reproduces the diversity of tumor driver genes by introducing a pooled cancer gene cDNA library using transposon-based intrahepatic delivery. This could be used to simultaneously evaluate the individual effects of various genetic drivers on TKI sensitivity in HCC in vivo. This model revealed that tumors expressing FGF19 are sensitive to lenvatinib in vivo. They comprehensively evaluated tumor secretory proteins to discover biomarkers for FGF19-driven HCC, identifying a correlation between FGF19 and the secretory protein ST6 β -galactoside α -2,6-sialyltransferase 1 (ST6GAL1) in HCC cells. This provided clinical evidence that ST6GAL1 may be a useful serum biomarker for selecting HCC patients who may benefit more from lenvatinib than sorafenib treatment [55]. This study strongly focused on the exploration of biomarkers for the therapeutic effect of lenvatinib in cancer cells, but validation in a prospective cohort is necessary for clinical application. Because ST6GAL1 protein levels can be measured in serum samples, it would be a very useful biomarker if validated.

Lenvatinib reportedly has higher selectivity for FGFR compared with other kinase inhibitors [51,56]. Therefore, biomarkers related to FGF-FGFR signaling may be more promising than those related to other signaling pathways that have been previously studied.

3. AFP as a Predictive Biomarker for Ramucirumab Treatment

VEGF and VEGFR2 signaling plays a crucial role in angiogenesis and tumor growth [57]. Multi-kinase inhibitors, such as sorafenib and lenvatinib that have been shown to be effective against HCC, always target VEGFR2. Ramucirumab is a human IgG1 monoclonal antibody that inhibits ligand activation of VEGFR2 [58]. In a Phase II trial of ramucirumab as a first-line therapy for HCC, ramucirumab demonstrated an overall response rate (ORR) and OS that surpassed the sorafenib administration group in the SHARP trial [59]. In this trial, an exploratory study of biomarkers measured circulating VEGF, soluble VEGFR1 (sVEGFR1), sVEGFR2, and several cytokines and growth factors in serum samples after ramucirumab administration. Among them, a potential correlation was suggested between reduced serum sVEGFR1 levels until day 8 post-administration and prolonged PFS and OS [58]. However, in the 'REACH trial', a Phase III trial of ramucirumab vs. placebo as a second-line therapy after sorafenib treatment, a significant improvement in OS was not achieved in the ramucirumab group compared with the placebo group [60].

Apart from the initially explored biomarkers, a subgroup analysis of the REACH trial revealed that OS in the ramucirumab group was significantly better when limited to cases with AFP \geq 400 ng/mL [60]. Therefore, the 'REACH-2 trial' was planned, which was a Phase III trial of ramucirumab vs. placebo as a second-line therapy restricted to cases with AFP \geq 400 ng/mL after sorafenib treatment [9]. As expected, the REACH-2 trial results indicated that OS was significantly extended in the ramucirumab treatment group compared with the placebo group. This trial became the first successful Phase III trial for advanced HCC treatment that selected target cases using a biomarker [9].

Because AFP was shown to be a predictive biomarker for the therapeutic effect of ramucirumab, to investigate the molecular profile differences of tumors using AFP levels, an analysis of 520 HCC

cases with known baseline AFP values was conducted. The data suggested that tumors in cases with AFP > 400 ng/mL showed significant activation of VEGF signaling [61].

4. Exploration of Biomarkers for Predicting the Therapeutic Efficacy of Single-agent ICIs and Combined Immunotherapy (Table 2)

The pharmacotherapy of HCC has shifted from being dominated by TKIs to ICIs and combined immunotherapies. Correspondingly, research into biomarkers for predicting therapeutic effectiveness has transitioned from focusing on those related to tumor growth signals to those focusing on the tumor microenvironment and tumor immune environment [62].

Studies on biomarkers for predicting the therapeutic effects of ICIs for HCC began with the validation of biomarkers discovered in other cancer types, such as melanoma, non-small cell lung cancer, and colorectal cancer. However, despite the demonstrated response rate of about 20% for HCC cases to single-agent therapies such as nivolumab and pembrolizumab, these treatments could not become standard first-line or second-line therapies [63–67]. The established combination therapy of atezolizumab and bevacizumab, which are anti-PD-L1 and anti-VEGF-A antibodies, respectively, became the standard treatment. This resulted in research on biomarkers for predicting the therapeutic effects of ICIs to primarily focus on this type of combined immunotherapy.

Table 2. Factors influencing patient prognosis or efficacy for treating hepatocellular carcinoma with immune checkpoint inhibitors.

Agents	Study design	Number of cases	Prognostic and predictive factors	Outcome	analysis	HR [95%CI]	P-value	Author (reference no)
Anti-PD-(L)1-based immunotherapy	Meta-analyses of 3 phase 3 trials: Checkmate 459 (Nivo vs Sora), IMbrave 150 (Ate/Bev vs Sora), KEYNOTE-240 (Pembro vs PBO) Retrospective (ICI single arm)	ICI 985	[HBV]	OS	univariate	0.64 [0.49-0.83]	0.0008	Pfister D et. al. [75]
		Nivo 371	[HCV]	OS	univariate	0.68 [0.48-0.97]	0.04	
Ate/Bev	retrospective	Non-viral cohort	[Lenvatinib]	OS	multivariate	0.65 [0.44-0.95]	0.0268	Rimini M et. al. [77]
		Ate/Bev 190		PFS	multivariate	0.67 [0.51-0.86]	0.035	
Lenva (Sora)	retrospective	Len 569	[Lenvatinib]	OS	multivariate	0.46 [0.26-0.84]	0.011	[77]
		NAFLD/NASH cohort		PFS	multivariate	0.55 [0.38-0.82]	0.031	
Anti-PD-(L)1 mono-therapy	retrospective, single arm	18	[hyperintensity tumor (RER* ≥ 0.9) on EOB-MRI]	PFS	multivariate	7.78 [1.59–38.1]	0.011	Aoki T et. al. [82]
		Non-viral HCC 30		[Steatotic HCC]	PFS	univariate		<0.05

		Ate/Bev 35	[heterogenous tumor on EOB-MRI]						
Ate/Bev	retrospective, separate single arm (not vs Len)		[hyperintensity tumor (RER _‡ ≥ 0.9) on EOB-MRI]	PFS	univariate		0.007	Sasaki R et.al.	
Lenva				PFS	univariate		0.012	[86]	
		Len 33	(no significant factor)						
Anti-PD-(L)1-based immuno-therapy	retrospective, single arm	24	[20 gene inflamed signature] (CCL5, CD2, CD3D, CD48, CD52, CD53, CXCL9, CXCR4, FYB, GZMA, GZMB, GZMK, IGHG1, IGHG3, LAPTM5, LCP2, PTPRC, SLA, TRAC, TRBC2)	PR	Wilcoxon rank-sum		0.047	Montironi C et.al.	[91]
		Anti-PD-(L)1-based immuno-therapy: training 190 (anti-PD-(L)1 mono 110, Ate/Bev 75, Others 5)	[Child-Pugh A]	OS	multivariate	2.3 (1.5-3.4)	<0.001		
			[ECOG PS 0]	OS	multivariate	2.1 (1.4-3.2)	<0.001		
			[AFP<100]	OS	multivariate	1.7 (1.2-2.6)	0.007		
			[CRP<1]	OS	multivariate	1.7 (1.2-2.6)	0.007		
			[CRAFITY score [†]]	OS	univariate		0.001		
Anti-PD-(L)1-based immuno-therapy	retrospective, separate single arm (not vs Sora)	validation 102 (anti-PD-(L)1 mono 68, Ate/Bev 25, Anti-PD-(L)1 + TKI 7, Others 2)	CRAFITY low			1			
			CRAFITY int.			2.0 [1.1-3.4]			
			CRAFITY high			3.6 [2.1-6.2]			
			[CRAFITY score [†]]	ORR	Chi square	-	0.001	Scheiner B	[93]
			[CRAFITY score [†]]	DCR	Chi square	-	<0.001		
Sora			[CRAFITY score [†]]	OS	univariate	-	0.001		
				DCR	Chi square	-	0.037		
		Sora 204	[CRAFITY score [†]]	OS	univariate	-	<0.001		
				PFS	multivariate	-	<0.001		
			[AFP<100]	OS	multivariate	-	0.028		
Ate/Bev	retrospective, single arm	297	[CRP<1]	PFS	multivariate	-	<0.001	Hatanaka T et.al.	[94]
			[CRAFITY score [†]]	OS	multivariate	-	0.032		
				PFS	univariate	-	<0.001		
				OS	univariate	-			
				DCR	Chi square	-	0.029		
Ate/Bev	retrospective, single arm	40	[NLR > 3.21]	PFS	univariate	-	<0.0001	Eso Y et.al	[99]
Ate/Bev	retrospective, single arm	249	[NLR > 3]	OS	multivariate	3.37 [1.02-11.08]	0.001	Tada T et.al.	[100]
Ate/Bev	retrospective pooled analysis	GO30140 arm A	<Transcriptome > [ABRS ^a high]	PFS	univariate	0.51 [0.3-0.87]	0.013	Zhu AX et. al.	

Sora of the phase 1b GO30140 (single arm) and the phase 3 trial IMbrave 150 (Ate/Bev vs Sora)		Ate/Bev 90 single arm)	[CD274 ^b high]	PFS	univariate	0.42 [0.25-0.72]	0.0011	[72]	
			[Teff ^c high]	PFS	univariate	0.46 [0.27-0.78]	0.0035		
			<In situ analyses>						
			[CD8+Tcell density]	CR/PR	Student T	-	0.007		
			[CD3+Tcell density]	CR/PR	Student T	-	0.039		
			[CD3+GZMB+Tcell density]	CR/PR	Student T	-	0.044		
			[MHC1+ tumor cells]	CR/PR	Student T	-	0.0087		
			IMbrave 150 (Ate/Bev119 Sora 58)	<Transcriptome >	PFS	multivariate	0.49 [0.25-0.97]	0.041	
			[ABRS ^a high]	OS	multivariate	0.26 [0.11-0.58]	0.0012		
			[CD274 ^b high]	OS	multivariate	0.3 [0.14-0.64]	0.002		
[Teff ^c high]	OS	multivariate	0.24 [0.11-0.5]	0.0002					
[Treg ^d /Teff ^c low]	OS	multivariate	0.24 [0.11-0.54]	0.0006					
[GPC3 low]	OS	multivariate	0.29 [0.13-0.62]	0.002					
[AFP low]	OS	multivariate	0.49 [0.28-0.87]	0.014					
			<In situ >	OS	multivariate	0.29 [0.14-0.61]	0.0011		
			[CD8+Tcell high dens.]	PFS	multivariate	0.54 [0.29-1.00]	0.053		
			<Genetic profiling>	OS	multivariate	0.42 [0.19-0.91]	3×10 ⁻⁴		
			[CTNNB1 WT]	PFS	multivariate	0.45 [0.27-0.86]	0.0086		
			[TERT Mut]	OS	multivariate	0.38 [0.16-0.89]	7.8×10 ⁻⁵		
				PFS	multivariate	0.61 [0.33-1.10]	0.047		
Ate/Bev	retrospective, single arm	34	[high plasma IL-6]	PFS	univariate	-	<0.05	Myojin Y et.al.	
				OS	univariate	2.785 [1.216-6.38]	0.01	[103]	
							<0.05		
Ate/Bev	retrospective, separate single arm (not vs Len)	Ate/Bev 24	[High-level CD8+ TILs]	PFS	univariate	-	0.041	Kuwano A et.al.	
Len		Len 15	(no significant factor)	ORR	Chi square	-	0.012	[104]	
				DCR	Chi square	-	0.031		

HR, hazard ratio; CI, confidence interval; PFS, progression-free survival; OS, overall survival; PD-1, Programmed death receptor 1; PD-L1, Programmed cell Death ligand 1; HBV, hepatitis B virus; HCV, hepatitis C virus; NAFLD, nonalcoholic fatty liver disease; NASH, Nonalcoholic steatohepatitis; RER, relative enhancement ratio; EOB-MRI, gadolinium ethoxybenzyl diethylenetriaminepentaacetic acid-enhanced magnetic resonance imaging; HCC, hepatocellular carcinoma; PR, partial response; ECOG, Eastern Cooperative Oncology Group; AFP, α -fetoprotein; CRP, c-reactive protein; ORR, overall response rate; DCR, disease control

rate;NLR, neutrophil to lymphocyte ratio; CR, complete response; PR, partial response. Note: In the 'Statistical analysis' section, 'univariate' typically refers to the Kaplan-Meier method and log-rank test, while the inclusion of HR indicates the use of Cox regression. Additionally, 'multivariate' typically refers to the utilization of the multivariate Cox regression model. ‡: (nodule SI/parenchyma SI on hepatobiliary phase images)/(nodule SI/parenchyma SI on precontract images) SI: signal intensity. †: CRAFITY-low: AFP<100 & CRP<1, intermediate: AFP≥100 ng/ml or CRP≥1 mg/dl, high: AFP ≥ 100 ng/mL & CRP ≥ 1 mg/dL. a: ABR5, atezolizumab + bevacizumab response signature (including CXCR2P1, ICOS, TIMD4, CTLA4, PAX5, KLRC3, FCRL3, AIM2, GBP5, and CCL4). b: CD274, PD-L1 mRNA. c: Teff, T effector (including CXCL9, PRF1, and GZMB). d: Treg, T regulatory (including CCR8, BATF, CTSC, TNFRSF4, FOXP3, TNFRSF18, IKZF2, and IL2RA).

4.1. Known Predictive Markers for the Efficacy of Single-Agent ICI and Combined Immunotherapies for HCC: PD-L1 Expression, Tumor Mutation Burden (TMB), and Microsatellite Instability (MSI)

The discovery of immune checkpoint proteins, such as PD-1/PD-L1 and CTLA-4, represents a significant breakthrough in the cancer immunotherapy field [68,69]. Currently, anti-PD-1 antibodies, anti-PD-L1 antibodies, and anti-CTLA-4 antibodies are used to treat HCC, including in combination therapies. For other types of cancer, PD-L1 expression, TMB, and MSI have been reported as biomarkers for predicting the therapeutic effects of these ICIs [70]. However, when considering the practicality of these as biomarkers, the frequency of PD-L1 expression, TMB-High, and MSI-High becomes an issue. According to a large cohort study by Ang et al., the incidence of MSI-High in HCC was extremely limited, with only one case among 542. Additionally, only six cases (0.8%) among 755 cases had a TMB of 20 mutations/Mb or more [71].

Zhu et al. conducted comprehensive analyses of transcriptomics, genomics, and IHC staining of patient samples collected in Phase Ib GO30140 trials and Phase III IMbrave150 trials to explore biomarkers for atezolizumab and bevacizumab combination therapy [72]. Either whole exome sequencing (WES) or FoundationOne panel profiling was performed to evaluate TMB, resulting in median TMBs of 5.6 mutations/Mb and 4.4 mutations/Mb, respectively. TMB was categorized as low, medium, or high, and its associations with response rates and survival times were verified. However, the relationship between TMB and response rate or survival benefit did not coincide in the GO30140 trial arm A and IMbrave 150 [72].

PD-L1 expression merits further investigation. In the phase I/II CheckMate 040 trial of nivolumab monotherapy for advanced HCC, PD-L1 expression in tumor tissues was examined in 174 out of 214 cases in the dose-expansion phase. Of these, 34 cases (20%) showed PD-L1 expression ≥ 1% in tumor cells via IHC, and these cases demonstrated an ORR of 9/34 (26%; 95% CI 13–44). However, even in 140 cases with PD-L1 < 1%, an OR was observed in 26/140 cases (19%; 95% CI 13–26), suggesting that therapeutic responses were observed regardless of PD-L1 expression status [63].

In the phase III Checkmate459 trial of nivolumab vs. sorafenib, PD-L1 expression ≥ 1% in tumor cells was found in 71 of 366 cases (19%) in the nivolumab group and in 64 of 362 cases (18%) in the sorafenib group. In patients who were administered nivolumab, a higher ORR was indicated if PD-L1 ≥ 1% than if PD-L1 < 1% (PD-L1 ≥ 1% ORR 20/71 (28%; 18–40); PD-L1 < 1% ORR 36/295 (12%; 9–17)). However, in the sorafenib group, there was no difference in ORR whether PD-L1 ≥ 1% or PD-L1 < 1% (PD-L1 ≥ 1% ORR 6/64 (9%; 4–19); PD-L1 < 1% ORR 20/300 (7%; 4–10)). A comparison between the nivolumab group's PD-L1 ≥ 1% and the sorafenib group's PD-L1 ≥ 1% showed a trend that favored nivolumab, with a median OS of 16.1 months (95% CI 8.4–22.3) vs. 8.6 months (95% CI 5.7–16.3), but the difference was not statistically significant (HR 0.80 (0.54–1.19)) [64].

In the phase II Keynote224 trial of pembrolizumab monotherapy, the conventional positive cell rate of PD-L1 in tumor cells (tumor proportion score (TPS)) and the combined positive score (combined positive score (CPS)) were calculated by dividing the number of PD-L1 positive cells in tumor cells and immune cells by the total number of surviving tumor cells and multiplying by 100 [73]. Of 52 cases, 22 cases (42%) were CPS positive and only seven cases (13%) were TPS positive. Significant differences were observed in the response rates and PFS between CPS positive and negative cases, but not between TPS positive and negative cases [73].

According to the comprehensive analysis by Zhu et al., patients with high CD274 (PD-L1 mRNA) expression showed a longer PFS with the atezolizumab-bevacizumab combination therapy than those with low expression. However, IHC data for PD-L1 protein levels indicated that there was only a potential correlation between PD-L1 expression and response [72].

As these results suggest, PD-L1 expression is somewhat related to the efficacy of anti-PD-1/PD-L1 immunotherapy in HCC and could potentially serve as a biomarker for predicting therapeutic effects. However, there are some uncertainties, as it is difficult to definitively make these predictions at the protein level, it is not yet determined whether to use TPS or CPS scoring, and there is an issue of heterogeneity associated with IHC staining assays [74].

4.2. NASH as a Background Liver Disease

One potential biomarker that could predict the lack of efficacy of ICI monotherapy is NASH/NAFLD as a background liver disease. Pfister et al. suggested that in a mouse model of NASH-induced HCC, CD8+/PD-1+ T cells could promote the progression of NASH. Administration of ICIs could release the brakes on these NASH-promoting cells, resulting in a potential exacerbation of NASH and increased HCC occurrence [75]. The authors conducted a meta-analysis of the cohorts from three phase III trials where ICIs were administered, namely Checkmate 459, IMbrave 150, and KEYNOTE-240. This analysis showed that while the ICI treatment significantly prolonged OS compared with the control in HBV-related and HCV-related HCC cases, prognosis did not improve in non-viral HCC cases. Furthermore, in two separate retrospective cohorts treated with anti-PD-1 or anti-PD-L1 antibodies, HCC cases caused by NAFLD showed reduced OS compared with those with other etiologies [75]. As demonstrated in this study, in a subgroup analysis of the phase III IMbrave 150 trial of atezolizumab-bevacizumab combination therapy and sorafenib, non-viral HCC including NASH did not show superiority, with a median OS of 17.0 months in the atezolizumab-bevacizumab combination group compared with 18.1 months in the sorafenib group [76].

Moreover, in a multicenter study involving 36 facilities in four countries (Italy, Japan, South Korea, and the UK), a retrospective analysis of 759 cases of advanced non-viral HCC revealed that when lenvatinib and atezolizumab + bevacizumab treatments were compared, lenvatinib showed significantly better results for OS and PFS rates in non-viral HCC overall. When non-viral HCC was divided into NAFLD/NASH and non-NAFLD/NASH, lenvatinib treatment was associated with a significant survival benefit compared with atezolizumab + bevacizumab in patients with NAFLD/NASH HCC [77]. These studies suggest the potential for the background liver disease in HCC to serve as a biomarker when deciding if ICI therapy is appropriate.

4.3. Wnt/ β -Catenin Mutations as a Biomarker and MRI Findings as Imaging Biomarkers

Spranger et al. reported that the presence of Wnt/ β -catenin mutations in melanoma results in the exclusion of T cell infiltration and resistance to ICIs [78]. Using The Cancer Genome Atlas (TCGA), Luke et al. demonstrated that tumors lacking the genetic expression signature of T cell-mediated inflammation, including 31 types of solid cancers like melanoma, show activated Wnt/ β -catenin signaling [79]. Furthermore, Harding et al. conducted a genomic analysis of 127 HCC tumor tissues, reporting that Wnt/ β -catenin mutations were found in 45% of cases. While the presence or absence of these mutations did not affect PFS with sorafenib treatment, their presence significantly shortened PFS with ICI treatment (2.0 vs. 7.4 months, $P < 0.0001$) [80]. In addition, in a comprehensive study by Zhu et al., patients with a wild-type CTNNB1 genotype in the IMbrave150 trial showed a greater therapeutic effect with atezolizumab + bevacizumab compared with sorafenib treatment, but no significant difference was observed between the treatments in cases with CTNNB1 mutations [72].

Ueno et al. focused on the differences in HCC findings in the hepatobiliary phase of gadolinium ethoxybenzyl diethylenetriamine pentaacetic acid (Gd-EOB-DTPA) enhanced MRI, comprehensively examined the transporter of Gd-EOB-DTPA, and analyzed the molecular regulatory mechanism. In clinical samples, it was demonstrated that high expression levels of OATP1B3 were strongly correlated with greater enhancement in the hepatobiliary phase of Gd-EOB-DTPA enhanced MRI.

Additionally, activated Wnt/ β -catenin signaling is closely associated with OATP1B3 expression in HCC cell cultures [81].

Aoki et al. analyzed 18 HCC cases that had received anti-PD-1 or anti-PD-L1 monotherapy and had Gd-EOB-DTPA enhanced MRI taken before treatment. As a result, in cases with high signal nodules in the hepatobiliary phase ($n = 8$), the median PFS was 2.7 months, while in cases with low signal nodules ($n = 10$), it was 5.8 months ($P = 0.007$). There was also a significant difference in the period until tumor enlargement, indicating that the hepatobiliary phase of Gd-EOB-DTPA enhanced MRI is a promising imaging biomarker for predicting the therapeutic effect of anti PD-1/PD-L1 monotherapy [82].

Moreover, it should be noted that multiple studies have reported that the efficacy of lenvatinib treatment for HCC is not affected by the signal intensity of the hepatobiliary phase of EOB-MRI [83,84].

Murai et al. focused on the increasingly prevalent non-viral HCC cases. They extracted the genomic DNA and total RNA from tumor tissues for profiling, then compared them with the pathological findings to identify sensitivity to immunotherapy. Steatotic HCC accounted for 23% of non-viral HCC cases, which showed an immune-rich, yet immune-exhausted, tumor immune microenvironment characterized by T cell exhaustion, infiltration of M2 macrophages and cancer-associated fibroblasts (CAFs), high expression of immune PD-L1, and activation of TGF- β signaling. Histological fatty deposition of resected HCC tissue and Fat Fraction Corrected for Spectral Complexity and Inhomogeneities (FFCSI) measured by MRI were strongly correlated. When 30 HCC patients evaluated by MRI before atezolizumab-bevacizumab combination therapy were retrospectively reviewed, a significantly longer PFS was confirmed in patients with steatotic HCC [85].

Whether MRI findings are useful as imaging biomarkers for predicting the efficacy of systemic therapy for HCC remains to be demonstrated with prospective validation results. However, as the number of cohorts of systemic therapy using ICIs, such as atezolizumab-bevacizumab combination therapy, increases in clinical practice, we believe a consensus will form.

4.4. Problems with Wnt/ β -catenin mutations as a biomarker and MRI findings as imaging biomarkers

With the combination therapy of atezolizumab and bevacizumab, Sasaki et al. reported that HCC patients with high-intensity EOB-MRI hepatobiliary phase suggesting Wnt/ β -catenin signal activation had a shorter PFS than the low-intensity HCC patients in the atezolizumab + bevacizumab group. This MRI finding was not associated with the treatment effect of lenvatinib [86]. However, in a study by Kuwano et al. using not EOB-MRI, but pretreatment tumor biopsy, there was no significant difference in the treatment effect or PFS of those receiving atezolizumab/bevacizumab combination therapy that depended on the presence or absence of Wnt/ β -catenin activation [87]. These discrepancies may be from biases resulting from each study being retrospective and having a small number of cases, but some research results may suggest otherwise.

First, previous research results have indicated that there may be a discrepancy between EOB-MRI hepatocellular phase uptake findings and Wnt/ β -catenin mutations. The transcription factor HNF4 α , which plays a role in maintaining mature hepatocyte function, decreases in dedifferentiated HCC. A decrease in HNF4 α during HCC dedifferentiation lead to decreased expression of OATP1B3 regardless of Wnt/ β -catenin mutations. This then results in a loss of gadoteric acid uptake in the hepatocyte phase, which may cause a mismatch [88,89].

Another question is if the presence of Wnt/ β -catenin mutations in HCC always results in a suppressed immune response. Sia et al. reported that about 25% of HCC cases are a subtype of immune class characterized by immune activation, with an overexpression of adaptive immune response genes, such as CD8A, CD3E, IFNG, CXCL9, and others, in the active immune response subtype. Additionally, immunosuppressive signals, like TGF- β , and M2 macrophages are present in the exhausted immune response subtype. The authors also stated that a better response to ICI treatment is expected in this immune class [90].

This research group further investigated the immune characteristics of HCC cases outside this immune class. As a result, they found that about 10% of HCC cases have an immune-like class characterized by high interferon (IFN) signaling, cytokines, and a diverse T cell repertoire, despite significant activation of Wnt/ β -catenin signaling by CTNNB1 mutation. This led them to classify HCC into an inflamed class, which includes the immune class and the immune-like class, and the other non-inflamed class. They designed an 'inflamed signature' consisting of 20 genes that can accurately indicate the inflamed class and confirmed that there was a significant overexpression of this signature in a group of patients who showed PR (partial response) with ICI treatment in an external cohort compared with a group of patients who showed SD (stable disease) and PD (progressive disease) [91].

When evaluating the tumor microenvironment, immunostaining in the inflamed class showed an enrichment of intratumoral CD8+ T cells (CD8 \geq 1%, 58% vs. 30%, $P=0.08$) and PD-L1 (PD-L1 \geq 1%, 21% vs. 4%, $P=0.19$) compared with the non-inflamed class. Additionally, analysis using CIBERSORT, which estimates the presence and ratios of immune cell subsets within tissues from gene expression data, showed a significantly higher proportion of CD8+ T cells ($P=3.51 \times 10^{-7}$) and M1 macrophages ($P=1.82 \times 10^{-4}$). However, in the immune-like class, M2 macrophages were significantly excluded ($P=1.78 \times 10^{-6}$) [91].

Furthermore, the authors created a 13-protein signature for identifying the inflamed class using a cohort with blood samples, as liquid biopsy-based biomarkers. They suggested that in treating HCC, it is worth considering whether to distinguish between the inflamed and non-inflamed classes using the 20-gene signature in tumor tissues or the liquid biopsy-based signature [91].

Interestingly, a study using a dataset of over 9,000 solid cancer cases across 31 types from TCGA found that activation of the Wnt/ β -catenin pathway is often associated with reduced T cell infiltration in most human cancers. A significant inverse correlation was observed between β -catenin protein levels and T cell inflammatory gene expression in 177 HCC cases [92]. According to this study, the degree of the inverse correlation between β -catenin protein levels and T cell inflammatory gene expression varied by cancer type. Furthermore, in certain types of cancer, such as colorectal and rectal cancers, examples where T cell inflammation and activation of the Wnt/ β -catenin pathway coexist are not rare [92]. This suggests that while activation of the Wnt/ β -catenin pathway often hinders T cell inflammation in human cancers, it is not always the case. However, it seems that subclasses, like the immune-like class, have not been proposed for other types of cancers.

Several studies have indicated the possibility that the hepatocyte phase of EOB-MRI may not match with Wnt/ β -catenin mutations, and that there may be HCC cases of the immune-like class that do not suppress immune responses against the tumor, even with Wnt/ β -catenin mutations. There may be potential in using findings from the hepatobiliary phase of EOB-MRI as biomarkers to predict the therapeutic effect of ICIs or the combination therapy of atezolizumab and bevacizumab. However, these findings do not necessarily indicate the presence or absence of Wnt/ β -catenin mutations. When using the presence or absence of Wnt/ β -catenin mutations as biomarkers to predict the therapeutic effect of ICIs, it is necessary to consider the existence of the immune-like class.

4.5. Blood Sample Biomarkers for Predicting the Therapeutic Effect of ICI Therapy: CRAFTY Score and NLR

Scheiner et al. created a training set of 190 cases and a validation set of 102 cases from a database of HCC cases in Europe that had received PD-L1/PD-1-based immunotherapy. Seventy-five cases (40%) of the training set and 25 cases (25%) of the validation set were patients who had received atezolizumab and bevacizumab combination therapy. In the training set, the investigated baseline parameters were etiology, whether immunotherapy was primary or after other treatments, Childs-Pugh class, ECOG performance status, radiological criteria, including the presence of major vessel invasion and extrahepatic metastasis, and serum AFP and C-reactive protein (CRP) levels. Serum AFP < 100 vs. ≥ 100 ng/mL and CRP < 1 vs. ≥ 1 mg/dL were identified as independent prognostic factors in the multivariate analysis, and the CRAFTY score was developed using this [93].

Patients with a score of 0 (CRAFITY-low: AFP < 100 ng/mL and CRP < 1 mg/dL) had the longest OS, followed by those with a score of 1 (CRAFITY-intermediate: either AFP \geq 100 ng/mL or CRP \geq 1 mg/dL) and those with a score of 2 (CRAFITY-high: both AFP \geq 100 ng/mL and CRP \geq 1 mg/dL). Similarly, the best treatment effect was seen in patients with a low CRAFTY score. This study also validated a cohort of 204 cases of sorafenib administration. The CRAFTY score was associated with the survival of the individuals, but not with the therapeutic effect [93]. The C-statistics, one of the statistical indicators to evaluate the performance and predictive ability of the model in the CRAFTY score, was 0.62 for both the derivation and validation cohorts. Although not highly accurate, it is very outstanding in its ease of use in routine practice and may be useful for predicting responses to ICI.

In a multi-institutional retrospective study in Japan, the CRAFTY score of 297 patients who received atezolizumab and bevacizumab combination therapy was analyzed. The median PFS in the CRAFTY score 0, 1, and 2 groups was 11.8 months, 6.5 months, and 3.2 months, respectively ($p < 0.001$). The median OS in patients with CRAFTY score 0, 1 and 2 was not reached, 14.3 months, and 11.6 months, respectively. This study showed the CRAFTY score could be useful for predicting therapeutic outcomes [94].

The pre-treatment NLR reflects the inflammatory response to cancer and is reportedly associated with patient prognosis and response to ICI treatment in various tumors [95–98]. Eso et al. analyzed the course of 40 HCC patients who received atezolizumab and bevacizumab combination therapy and found that the NLR value was significantly lower in the complete response (CR), PR and SD group than in the PD group (2.47 vs. 4.48, $P = 0.013$). Using the optimal NLR cut-off value (3.21) determined by receiver operating characteristic curve analysis for predicting response, they also found that patients with $NLR \leq 3.21$ showed significantly better PFS than patients with $NLR > 3.21$ [99].

A similar examination was conducted in a multi-institutional joint study in Japan, and the cumulative OS rate was significantly different between patients with low NLR (< 3.0) and high NLR (≥ 3.0) ($P = 0.001$). Conversely, there was no difference in cumulative PFS or response between patients with low and high NLR values. In Cox proportional hazards modeling analysis using inverse probability weighting, having an NLR of at least 3.0 was found to be significantly associated with OS [100].

Because the CRAFTY score and NLR are parameters that can be easily obtained from blood samples, it is necessary to rigorously validate whether they should be used as biomarkers in routine practice.

4.6. Biomarkers Predicting the Therapeutic Effect of Atezolizumab and Bevacizumab Combination Therapy

As will be discussed below, the results of the HIMALAYA trial have made it possible to administer a combination of durvalumab and tremelimumab as a first-line treatment. However, comparisons of treatment outcomes have indicated that the first-line treatment of choice for unresectable HCC cases is currently thought to be atezolizumab and bevacizumab combination therapy [101,102].

Myojin et al. measured the levels of 34 baseline plasma proteins in patients with advanced HCC who received atezolizumab + bevacizumab therapy, finding that plasma IL-6 levels were a significant predictor of non-response to this therapy. They confirmed that the PFS and OS were significantly shorter in the high IL-6 group than in the low IL-6 group [103].

As mentioned previously, Zhu et al. used transcriptome analysis to derive an atezolizumab + bevacizumab response signature (ABRS) comprising 10 genes associated with a response to atezolizumab + bevacizumab (defined as CR or PR). High expression of the ABRS, as well as the existing immune gene CD274 (PD-L1 mRNA) or the Teff sign (CXCL9, PRFI, and GZMB), was associated with longer PFS in patients treated with atezolizumab + bevacizumab. The Treg signature (CCR8, BATF, CTSC, TNFRSF4, FOXP3, TNFRSF18, IKZF2, and IL2RA) was also related to improved PFS and OS when the Treg/Teff signature ratio was low in IMbrave150, which compared atezolizumab + bevacizumab with sorafenib treatment [72].

Multiplex IHC analysis in this study showed that in the GO30140 cohort A, responding patients (CR/PR) had a higher density of infiltrating CD8+ T cells, CD3+ T cells, and GZMB+/CD3+ T cells in tumor areas than non-responders (SD/PD). Furthermore, the density of tumor-infiltrating CD8+ T cells in IMbrave 150 baseline tumor samples was analyzed. Patients with a high density of tumor-infiltrating CD8+ T cells (defined by the split median) had significantly longer OS when treated with atezolizumab + bevacizumab combination therapy compared with those treated with sorafenib [72]. These studies highlight an important point: the state of T cell immunity in the tumor microenvironment before treatment ultimately influences the treatment effect of atezolizumab + bevacizumab combination therapy for HCC.

Kuwano et al. examined the relationship between tumor infiltration of CD8+ T cells detected by IHC staining of liver tumor biopsies before treatment initiation and the therapeutic effect of drug therapy. In cases with a high level of CD8+ T cell tumor infiltration, the PFS of patients treated with atezolizumab + bevacizumab combination therapy was significantly extended and the response rate was also significantly improved compared with cases with low levels. However, in patients receiving lenvatinib, there was no association between CD8+ T cell tumor infiltration and the response rate or PFS [104]. Although this study included only a limited number of patients, it suggests that evaluating CD8+ T cell tumor infiltration alone, without any transcriptomics analysis or genomic profiling, may serve as a useful biomarker for predicting the treatment method choice and therapeutic response to drug therapy in HCC.

4.7. Biomarkers for Durvalumab and Tremelimumab Combination Therapy

The combination therapy of the anti-PD-L1 antibody durvalumab with the anti-CTLA-4 antibody tremelimumab for unresectable HCC cases has surpassed the control drug sorafenib in the HIMALAYA trial and has been approved as a first-line therapy [11]. Among the factors studied as potential biomarkers in the HIMALAYA trial, the only results currently available relate to the PD-L1 status of the tumor prior to therapy. According to the subgroup analysis, there was no difference in benefits of a combination of the two ICIs compared with sorafenib, regardless of whether PD-L1 expression was positive or negative [11].

Another interesting finding was that the combination therapy showed clear advantages over sorafenib treatment in cases of HBV-related and non-viral HCC, but not in cases of HCV-related HCC. A similar trend was observed with durvalumab monotherapy [11]. As mentioned in the sorafenib section of this review, this may be because sorafenib has greater benefits for HCV-related HCC [15]. Nevertheless, there is no clear biomarker candidate for the durvalumab and tremelimumab combination therapy. The CRAFTY score and NLR can be easily validated, but as of now, no reports have confirmed their effectiveness with this treatment.

5. Conclusions and Future Directions

In this review, we have discussed the published research findings on biomarkers for predicting the therapeutic effects of drugs available for unresectable HCC tumors. As TKIs will continue to be used as secondary therapies, the search for biomarkers must continue. Among the factors mentioned, hepatic function, underlying hepatic disease, and the NLR may serve as vague indications of utility, but they are far from being decisive in drug selection. Because it is considered difficult to carry out prospective trials of existing TKIs in the future, we believe it would be worthwhile to proactively investigate if biomarker candidates like DKK-1, ST6GAL1, and regorafenib hold value, as they have been indicated in several studies. We encourage this investigation within existing cohorts that have retained blood samples.

For the current standard treatment of atezolizumab and bevacizumab combination therapy, routine examinations such as underlying hepatic disease, CRAFTY score, and the NLR seem to provide some guidance. Furthermore, imaging diagnostics, such as the evaluation of fat deposition through EOB-MRI hepatobiliary phase or FFCSI and information from MRI examinations, can be useful. At present, the most important factor is believed to be an accurate assessment of the state of T cell immunity in the tumor microenvironment prior to treatment. The literature suggests that

evaluating the tumor microenvironment and immune environment is more achievable compared with the diversity and heterogeneity of tumor cells. Assessing CD8+ T cell infiltration by collecting tumor tissues can be performed relatively easily, even in general hospitals. Thus, this has a high potential to become a practical biomarker.

In general hospitals that treat HCC patients, it is not necessarily wrong to administer drug therapy for advanced HCC cases according to guidelines based only on the results of blood tests and CT scans. However, we recommend conducting treatment after assessing the state within the tumor tissue as much as possible by collecting blood and tissue samples and performing MRI image evaluations before treatment.

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References

1. Rungay H.; Arnold M.; Ferlay J.; Lesi O.; Cabasag C.J.; Vignat J.; Laversanne M.; McGlynn K.A.; Soerjomataram I. Global burden of primary liver cancer in 2020 and predictions to 2040. *J. Hepatol.* 2022, 77, 1598–1606. <https://doi.org/10.1016/j.jhep.2022.08.021>
2. McGlynn K.A.; Petrick J.L.; El-Serag H.B. Epidemiology of Hepatocellular Carcinoma. *Hepatology.* 2021, 73, 4–13. <https://doi.org/10.1002/hep.31288>
3. Llovet J.M.; Kelley R.K.; Villanueva A.; Singal A.G.; Pikarsky E.; Roayaie S.; Lencioni R.; Koike K.; Zucman-Rossi J.; Finn R.S. Hepatocellular carcinoma. *Nat. Rev. Dis. Primers.* 2021, 7, 6. <https://doi.org/10.1038/s41572-020-00240-3>
4. Mathurin P.; Rixe O.; Carbonell N.; Bernard B.; Cluzel P.; Bellin M.F.; Khayat D.; Opolon P.; Poynard T. Overview of medical treatments in unresectable hepatocellular carcinoma—an impossible meta-analysis? *Aliment Pharmacol. Ther.* 1998, 12, 111–126. <https://doi.org/10.1046/j.1365-2036.1998.00286.x>
5. Wilhelm S.M.; Carter C.; Tang L.; Wilkie D.; McNabola A.; Rong H.; Chen C.; Zhang X.; Vincent P.; McHugh M.; et al. BAY 43-9006 exhibits broad spectrum oral antitumor activity and targets the RAF/MEK/ERK pathway and receptor tyrosine kinases involved in tumor progression and angiogenesis. *Cancer Res.* 2004, 64, 7099–7109. <https://doi.org/10.1158/0008-5472.CAN-04-1443>
6. Llovet J.M.; Ricci S.; Mazzaferro V.; Hilgard P.; Gane E.; Blanc J-F.; de Oliveira A.C.; Santoro A.; Raoul J-L.; Forner A.; et al. Sorafenib in advanced hepatocellular carcinoma. *N. Engl. J. Med.* 2008, 359, 378–390. <https://doi.org/10.1056/NEJMoa0708857>
7. Bruix J.; Qin S.; Merle P.; Granito A.; Huang Y-H.; Bodoky G.; Pracht M.; Yokosuka O.; Rosmorduc O.; Breder V.; et al. Regorafenib for patients with hepatocellular carcinoma who progressed on sorafenib treatment (RESORCE): a randomised, double-blind, placebo-controlled, phase 3 trial. *Lancet.* 2017, 389, 56–66. [https://doi.org/10.1016/S0140-6736\(16\)32453-9](https://doi.org/10.1016/S0140-6736(16)32453-9)
8. Kudo M.; Finn R.S.; Qin S.; Han K-H.; Ikeda K.; Piscaglia F.; Baron A.; Park J-W.; Han G.; Jassem J.; et al. Lenvatinib versus sorafenib in first-line treatment of patients with unresectable hepatocellular carcinoma: a randomised phase 3 non-inferiority trial. *Lancet.* 2018, 391, 1163–1173. [https://doi.org/10.1016/S0140-6736\(18\)30207-1](https://doi.org/10.1016/S0140-6736(18)30207-1)
9. Zhu A.X.; Kang Y-K.; Yen C-J.; Finn R.S.; Galle P.R.; Llovet J.M.; Assenat E.; Brandi G.; Pracht M.; Lim H.Y.; et al. Ramucirumab after sorafenib in patients with advanced hepatocellular carcinoma and increased α -fetoprotein concentrations (REACH-2): a randomised, double-blind, placebo-controlled, phase 3 trial. *Lancet Oncol.* 2019, 20, 282–296. [https://doi.org/10.1016/S1470-2045\(18\)30937-9](https://doi.org/10.1016/S1470-2045(18)30937-9)
10. Finn R.S.; Qin S.; Ikeda M.; Galle P.R.; Ducreux M.; Kim T-Y.; Kudo M.; Breder V.; Merle P.; Kaseb A.O.; et al. Atezolizumab plus Bevacizumab in Unresectable Hepatocellular Carcinoma. *N. Engl. J. Med.* 2020, 382, 1894–1905. <https://doi.org/10.1056/NEJMoa1915745>

11. Abou-Alfa G.K.; Lau G.; Kudo, M.; Chan S.L.; Kelley R.K.; Furuse J.; Sukeepaisarnjaroen W.; Kang Y-K.; Dao T.V.; De Toni E.N.; et al. Tremelimumab plus Durvalumab in Unresectable Hepatocellular Carcinoma. *NEJM Evid.* 2022, 1, EVIDo2100070.
12. Abou-Alfa G.K.; Meyer T.; Cheng A-L.; El-Khoueiry A.B.; Rimassa L.; Ryoo B-Y.; Cicin I.; Merle P.; Chen Y.H.; Park J-W.; et al. Cabozantinib in Patients with Advanced and Progressing Hepatocellular Carcinoma. *N. Engl. J. Med.* 2018, 379, 54–63. <https://doi.org/10.1056/NEJMoa1717002>
13. Llovet J.M.; Peña C.E.A.; Lathia C.D.; Shan M.; Meinhardt G.; Bruix J; SHARP Investigators Study Group (2012). Plasma biomarkers as predictors of outcome in patients with advanced hepatocellular carcinoma. *Clin. Cancer. Res.* 2012, 18, 2290–2300. <https://doi.org/10.1158/1078-0432.CCR-11-2175>
14. Miyahara K.; Nouse K.; Morimoto Y.; Takeuchi Y.; Hagihara H.; Kuwaki K.; Onishi H.; Ikeda F.; Miyake Y.; Nakamura S.; et al. Pro-angiogenic cytokines for prediction of outcomes in patients with advanced hepatocellular carcinoma. *Br. J. Cancer.* 2013, 109, 2072–2078. <https://doi.org/10.1038/bjc.2013.554>
15. Bruix J.; Cheng A-L.; Meinhardt G.; Nakajima K.; De Sanctis Y.; Llovet J. Prognostic factors and predictors of sorafenib benefit in patients with hepatocellular carcinoma: Analysis of two phase III studies. *J. Hepatol.* 2017, 67, 999–1008. <https://doi.org/10.1016/j.jhep.2017.06.026>
16. Guthrie G.J.K.; Charles K.A.; Roxburgh C.S.D.; Horgan P.G.; McMillan D.C.; Clarke S.J. The systemic inflammation-based neutrophil-lymphocyte ratio: experience in patients with cancer. *Crit. Rev. Oncol. Hematol.* 2013, 88, 218–230. <https://doi.org/10.1016/j.critrevonc.2013.03.010>
17. Templeton A.J.; McNamara M.G.; Šeruga B.; Vera-Badillo F.E.; Aneja P.; Ocaña A.; Leibowitz-Amit R.; Sonpavde G.; Knox J.J.; Tran B.; et al. Prognostic role of neutrophil-to-lymphocyte ratio in solid tumors: a systematic review and meta-analysis. *J. Natl. Cancer Inst.* 2014, 106, dju124. <https://doi.org/10.1093/jnci/dju124>
18. Qi X.; Li J.; Deng H.; Li H.; Su C.; Guo X. Neutrophil-to-lymphocyte ratio for the prognostic assessment of hepatocellular carcinoma: A systematic review and meta-analysis of observational studies. *Oncotarget.* 2016, 7, 45283–45301. <https://doi.org/10.18632/oncotarget.9942>
19. Marrero J.A.; Kudo M.; Venook A.P.; Ye S-L.; Bronowicki J-P.; Chen X-P.; Dagher L.; Furuse J.; Geschwind J-F.H.; Ladrón de Guevara L.; et al. Observational registry of sorafenib use in clinical practice across Child-Pugh subgroups: The GIDEON study. *J. Hepatol.* 2016, 65, 1140–1147. <https://doi.org/10.1016/j.jhep.2016.07.020>
20. Kuwano A.; Yada M.; Nagasawa S.; Tanaka K.; Morita Y.; Masumoto A.; Motomura K. Hepatitis C virus eradication ameliorates the prognosis of advanced hepatocellular carcinoma treated with sorafenib. *J. Viral Hepat.* 2022, 29, 543–550. <https://doi.org/10.1111/jvh.13681>
21. Tsai H-Y.; Chang H-P.; Chen C-J.; Hsu W-L.; Huang L-Y.; Lee P-C. Effects of direct-acting antiviral therapy for patients with advanced hepatocellular carcinoma and concomitant hepatitis C-A population-based cohort study. *Eur. Rev. Med. Pharmacol. Sci.* 2021, 25, 7543–7552. https://doi.org/10.26355/eurrev_202112_27454
22. Abou-Alfa G.K.; Schwartz L.; Ricci S.; Amadori D.; Santoro A.; Figer A.; De Greve J.; Douillard J.Y.; Lathia C.; Schwartz B.; et al. Phase II study of sorafenib in patients with advanced hepatocellular carcinoma. *J. Clin. Oncol.* 2006, 24, 4293–4300. <https://doi.org/10.1200/JCO.2005.01.3441>
23. Chen D.; Zhao P.; Li S.Q.; Xiao W.K.; Yin X.Y.; Peng B.G.; Liang L.J. Prognostic impact of pERK in advanced hepatocellular carcinoma patients treated with sorafenib. *Eur. J. Surg. Oncol.* 2013, 39, 974–980. <https://doi.org/10.1016/j.ejso.2013.06.018>
24. Negri F.; Bello B.D.; Porta C.; Campanini N.; Rossi S.; Tinelli C.; Poggi G.; Missale G.; Fanella S.; Salvagni S.; et al. Expression of pERK and VEGFR-2 in advanced hepatocellular carcinoma and resistance to sorafenib treatment. *Liver Int.* 2015, 35, 2001–2008. <https://doi.org/10.1111/liv.12778>
25. Personeni N.; Rimassa L.; Pressiani T.; Destro A.; Ligorio C.; Tronconi M.C.; Bozzarelli S.; Carnaghi C.; Di Tommaso L.; Giordano L.; et al. Molecular determinants of outcome in sorafenib-treated patients with hepatocellular carcinoma. *J. Cancer Res. Clin. Oncol.* 2013, 139, 1179–1187. <https://doi.org/10.1007/s00432-013-1429-x>
26. Arao T.; Ueshima K.; Matsumoto K.; Nagai T.; Kimura H.; Hagiwara S.; Sakurai T.; Haji S.; Kanazawa A.; Hidaka H.; et al. FGF3/FGF4 amplification and multiple lung metastases in responders to sorafenib in hepatocellular carcinoma. *Hepatology.* 2013, 57, 1407–1415. <https://doi.org/10.1002/hep.25956>
27. Chen X.; Ba Y.; Ma L.; Cai X.; Yin Y.; Wang K.; Guo J.; Zhang Y.; Chen J.; Guo X.; et al. Characterization of microRNAs in serum: a novel class of biomarkers for diagnosis of cancer and other diseases. *Cell Res.* 2008, 18, 997–1006. <https://doi.org/10.1038/cr.2008.282>
28. Gyöngyösi B.; Végh É.; Járay B.; Székely E.; Fassan M.; Bodoky G.; Schaff Z.; Kiss A. Pretreatment MicroRNA Level and Outcome in Sorafenib-treated Hepatocellular Carcinoma. *J. Histochem. Cytochem.* 2014, 62, 547–555. <https://doi.org/10.1369/0022155414537277>
29. Giordano S.; Columbano A. MicroRNAs: new tools for diagnosis, prognosis, and therapy in hepatocellular carcinoma? *Hepatology.* 2013, 57, 840–847. <https://doi.org/10.1002/hep.26095>

30. Ghidini M.; Braconi C. Non-Coding RNAs in Primary Liver Cancer. *Front. Med. (Lausanne)*. 2015, 2, 36. <https://doi.org/10.3389/fmed.2015.00036>
31. Vaira V.; Roncalli M.; Carnaghi C.; Fav ersani A.; Maggioni M.; Augello C.; Rimassa L.; Pressiani T.; Spagnuolo G.; Di Tommaso L.; et al. MicroRNA-425-3p predicts response to sorafenib therapy in patients with hepatocellular carcinoma. *Liver Int.* 2015, 35, 1077–1086. <https://doi.org/10.1111/liv.12636>
32. Shi Y.; Liu Z.; Lin Q.; Luo Q.; Cen Y.; Li J.; Fang X.; Gong C. MiRNAs and Cancer: Key Link in Diagnosis and Therapy. *Genes (Basel)*. 2021, 12, 1289. <https://doi.org/10.3390/genes12081289>
33. Qiu Z.; Li H.; Zhang Z.; Zhu Z.; He S.; Wang X.; Wang P.; Qin J.; Zhuang L.; Wang W.; et al. A Pharmacogenomic Landscape in Human Liver Cancers. *Cancer Cell*. 2019, 36, 179–193.e11. <https://doi.org/10.1016/j.ccell.2019.07.001>
34. Semenov M.V.; Zhang X.; He X. DKK1 antagonizes Wnt signaling without promotion of LRP6 internalization and degradation. *J. Biol. Chem.* 2008, 283, 21427–21432. <https://doi.org/10.1074/jbc.M800014200>
35. Teufel M.; Seidel H.; Köchert K.; Meinhardt G.; Finn R.S.; Llovet J.M.; Bruix J. Biomarkers Associated With Response to Regorafenib in Patients With Hepatocellular Carcinoma. *Gastroenterology*. 2019, 156, 1731–1741. <https://doi.org/10.1053/j.gastro.2019.01.261>
36. Piccart-Gebhart M.J.; Procter M.; Leyland-Jones B.; Goldhirsch A.; Untch M.; Smith I.; Gianni L.; Baselga J.; Bell R.; Jackisch C.; et al. Trastuzumab after adjuvant chemotherapy in HER2-positive breast cancer. *N. Engl. J. Med.* 2005, 353, 1659–1672. <https://doi.org/10.1056/NEJMoa052306>
37. Cataldo V.D.; Gibbons D.L.; Pérez-Soler R.; Quintás-Cardama A. Treatment of non-small-cell lung cancer with erlotinib or gefitinib. *N. Engl. J. Med.* 2011, 364, 947–955. <https://doi.org/10.1056/NEJMct0807960>
38. Solomon B.J.; Mok T.; Kim D.W.; Wu Y.L.; Nakagawa K.; Mekhail T.; Felip E.; Cappuzzo F.; Paolini J.; Usari T.; et al. First-line crizotinib versus chemotherapy in ALK-positive lung cancer. *N. Engl. J. Med.* 2014, 371, 2167–2177. <https://doi.org/10.1056/NEJMoa1408440>
39. Llovet J.M.; Hernandez-Gea V. Hepatocellular carcinoma: reasons for phase III failure and novel perspectives on trial design. *Clin. Cancer Res.* 2014, 20, 2072–2079. <https://doi.org/10.1158/1078-0432.CCR-13-0547>
40. Villanueva A.; Chiang D.Y.; Newell P.; Peix J.; Thung S.; Alsinet C.; Tovar V.; Roayaie S.; Minguez B.; Sole M.; et al. Pivotal role of mTOR signaling in hepatocellular carcinoma. *Gastroenterology*. 2008, 135, 1972–1983. <https://doi.org/10.1053/j.gastro.2008.08.008>
41. Zhu A.X.; Kudo M.; Assenat E.; Cattani S.; Kang Y.K.; Lim H.Y.; Poon R.T.; Blanc J.F.; Vogel A.; Chen C.L.; et al. Effect of everolimus on survival in advanced hepatocellular carcinoma after failure of sorafenib: the EVOLVE-1 randomized clinical trial. *JAMA*. 2014, 312, 57–67. <https://doi.org/10.1001/jama.2014.7189>
42. Zhang B.; Finn R.S. Personalized Clinical Trials in Hepatocellular Carcinoma Based on Biomarker Selection. *Liver Cancer*. 2016, 5, 221–232. <https://doi.org/10.1159/000367763>
43. Santoro A.; Rimassa L.; Borbath I.; Daniele B.; Salvagni S.; Van Laethem J.L.; Van Vlierberghe H.; Trojan J.; Kolligs F.T.; Weiss A.; et al. Tivantinib for second-line treatment of advanced hepatocellular carcinoma: a randomised, placebo-controlled phase 2 study. *Lancet Oncol.* 2013, 14, 55–63. [https://doi.org/10.1016/S1470-2045\(12\)70490-4](https://doi.org/10.1016/S1470-2045(12)70490-4)
44. Rimassa L.; Assenat E.; Peck-Radosavljevic M.; Pracht M.; Zagonel V.; Mathurin P.; Rota Caremoli E.; Porta C.; Daniele B.; Bolondi L.; et al. Tivantinib for second-line treatment of MET-high, advanced hepatocellular carcinoma (METIV-HCC): a final analysis of a phase 3, randomised, placebo-controlled study. *Lancet Oncol.* 2018, 19, 682–693. [https://doi.org/10.1016/S1470-2045\(18\)30146-3](https://doi.org/10.1016/S1470-2045(18)30146-3)
45. Kudo M.; Morimoto M.; Moriguchi M.; Izumi N.; Takayama T.; Yoshiji H.; Hino K.; Oikawa T.; Chiba T.; Motomura K.; et al. A randomized, double-blind, placebo-controlled, phase 3 study of tivantinib in Japanese patients with MET-high hepatocellular carcinoma. *Cancer Sci.* 2020, 111, 3759–3769. <https://doi.org/10.1111/cas.14582>
46. Rimassa L.; Kelley R.K.; Meyer T.; Ryoo B.Y.; Merle P.; Park J.W.; Blanc J.F.; Lim H.Y.; Tran A.; Chan Y.W.; et al. Outcomes Based on Plasma Biomarkers for the Phase 3 CELESTIAL Trial of Cabozantinib versus Placebo in Advanced Hepatocellular Carcinoma. *Liver Cancer*. 2021, 11, 38–47. <https://doi.org/10.1159/000519867>
47. Totoki Y.; Tatsuno K.; Covington K.R.; Ueda H.; Creighton C.J.; Kato M.; Tsuji S.; Donehower L.A.; Slagle B.L.; Nakamura H.; et al. Trans-ancestry mutational landscape of hepatocellular carcinoma genomes. *Nat. Genet.* 2014, 46, 1267–1273. <https://doi.org/10.1038/ng.3126>
48. Schulze K.; Imbeaud S.; Letouzé E.; Alexandrov L.B.; Calderaro J.; Rebouissou S.; Couchy G.; Meiller C.; Shinde J.; Soysouvanh F.; et al. Exome sequencing of hepatocellular carcinomas identifies new mutational signatures and potential therapeutic targets. *Nat. Genet.* 2015, 47, 505–511. <https://doi.org/10.1038/ng.3252>
49. Xue R.; Li R.; Guo H.; Guo L.; Su Z.; Ni X.; Qi L.; Zhang T.; Li Q.; Zhang Z.; et al. Variable Intra-Tumor Genomic Heterogeneity of Multiple Lesions in Patients With Hepatocellular Carcinoma. *Gastroenterology*. 2016, 150, 998–1008. <https://doi.org/10.1053/j.gastro.2015.12.033>

50. Pinter M.; Peck-Radosavljevic M. Review article: systemic treatment of hepatocellular carcinoma. *Aliment Pharmacol. Ther.* 2018, 48, 598–609. <https://doi.org/10.1111/apt.14913>
51. Tohyama O.; Matsui J.; Kodama K.; Hata-Sugi N.; Kimura T.; Okamoto K.; Minoshima Y.; Funahashi Y. Antitumor activity of lenvatinib (e7080): an angiogenesis inhibitor that targets multiple receptor tyrosine kinases in preclinical human thyroid cancer models. *J. Thyroid Res.* 2014, 2014, 638747. <https://doi.org/10.1155/2014/638747>
52. Matsuki M.; Hoshi T.; Yamamoto Y.; Ikemori-Kawada M.; Minoshima Y.; Funahashi Y.; Matsui J. Lenvatinib inhibits angiogenesis and tumor fibroblast growth factor signaling pathways in human hepatocellular carcinoma models. *Cancer Med.* 2018, 7, 2641–2653. <https://doi.org/10.1002/cam4.1517>
53. Tada T.; Kumada T.; Hiraoka A.; Michitaka K.; Atsukawa M.; Hirooka M.; Tsuji K.; Ishikawa T.; Takaguchi K.; Kariyama K.; et al. Neutrophil-to-lymphocyte ratio is associated with survival in patients with unresectable hepatocellular carcinoma treated with lenvatinib. *Liver Int.* 2020, 40, 968–976. <https://doi.org/10.1111/liv.14405>
54. Casadei-Gardini A.; Rimini M.; Kudo M.; Shimose S.; Tada T.; Suda G.; Goh M.J.; Jefremow A.; Scartozzi M.; Cabibbo G.; et al. Real Life Study of Lenvatinib Therapy for Hepatocellular Carcinoma: RELEVANT Study. *Liver Cancer.* 2022, 11, 527–539. <https://doi.org/10.1159/000525145>
55. Myojin Y.; Kodama T.; Maesaka K.; Motooka D.; Sato Y.; Tanaka S.; Abe Y.; Ohkawa K.; Mita E.; Hayashi Y.; et al. ST6GAL1 Is a Novel Serum Biomarker for Lenvatinib-Susceptible FGF19-Driven Hepatocellular Carcinoma. *Clin. Cancer Res.* 2021, 27, 1150–1161. <https://doi.org/10.1158/1078-0432.CCR-20-3382>
56. Wilson L.J.; Linley A.; Hammond D.E.; Hood F.E.; Coulson J.M.; MacEwan D.J.; Ross S.J.; Slupsky J.R.; Smith P.D.; Evers P.A.; et al. The biology of VEGF and its receptors. *Nat. Med.* 2003, 9, 669–676. <https://doi.org/10.1038/nm0603-669>
57. Spratlin J.L.; Cohen R.B.; Eadens M.; Gore L.; Camidge D.R.; Diab S.; Leong S.; O'Bryant C.; Chow L.Q.M.; Serkova N.J.; et al. Phase I pharmacologic and biologic study of ramucirumab (IMC-1121B), a fully human immunoglobulin G1 monoclonal antibody targeting the vascular endothelial growth factor receptor-2. *J. Clin. Oncol.* 2010, 28, 780–787. <https://doi.org/10.1200/JCO.2009.23.7537>
58. Zhu A.X.; Finn R.S.; Mulcahy M.; Gurtler J.; Sun W.; Schwartz J.D.; Dalal R.P.; Joshi A.; Hozak R.R.; Xu Y.; et al. A phase II and biomarker study of ramucirumab, a human monoclonal antibody targeting the VEGF receptor-2, as first-line monotherapy in patients with advanced hepatocellular cancer. *Clin. Cancer Res.* 2013, 19, 6614–6623. <https://doi.org/10.1158/1078-0432.CCR-13-1442>
59. Zhu A.X.; Park J.O.; Ryoo B.Y.; Yen C.J.; Poon R.; Pastorelli D.; Blanc J.F.; Chung H.C.; Baron A.D.; Pfiffer T.E.F.; et al. Ramucirumab versus placebo as second-line treatment in patients with advanced hepatocellular carcinoma following first-line therapy with sorafenib (REACH): a randomised, double-blind, multicentre, phase 3 trial. *Lancet Oncol.* 2015, 16, 859–870. [https://doi.org/10.1016/S1470-2045\(15\)00050-9](https://doi.org/10.1016/S1470-2045(15)00050-9)
60. Spratlin J.L.; Cohen R.B.; Eadens M.; Gore L.; Camidge D.R.; Diab S.; Leong S.; O'Bryant C.; Chow L.Q.M.; Serkova N.J.; et al. Phase I pharmacologic and biologic study of ramucirumab (IMC-1121B), a fully human immunoglobulin G1 monoclonal antibody targeting the vascular endothelial growth factor receptor-2. *J. Clin. Oncol.* 2010, 28, 780–787. <https://doi.org/10.1200/JCO.2009.23.7537>
61. Montal R.; Andreu-Oller C.; Bassaganyas L.; Esteban-Fabré R.; Moran S.; Montironi C.; Moeini A.; Pinyol R.; Peix J.; Cabellos L.; et al. Molecular portrait of high alpha-fetoprotein in hepatocellular carcinoma: implications for biomarker-driven clinical trials. *Br. J. Cancer.* 2019, 121, 340–343. <https://doi.org/10.1038/s41416-019-0513-7>
62. Donne R.; Lujambio A. The liver cancer immune microenvironment: Therapeutic implications for hepatocellular carcinoma. *Hepatology.* 2023, 77, 1773–1796. <https://doi.org/10.1002/hep.32740>
63. El-Khoueiry A.B.; Sangro B.; Yau T.; Crocenzi T.S.; Kudo M.; Hsu C.; Kim T.-Y.; Choo S.-P.; Trojan J.; Welling Rd T.H.; et al. Nivolumab in patients with advanced hepatocellular carcinoma (CheckMate 040): an open-label, non-comparative, phase 1/2 dose escalation and expansion trial. *Lancet.* 2017, 389, 2492–2502. [https://doi.org/10.1016/S0140-6736\(17\)31046-2](https://doi.org/10.1016/S0140-6736(17)31046-2)
64. Yau T.; Park J.-W.; Finn R.S.; Cheng A.-L.; Mathurin P.; Edeline J.; Kudo M.; Harding J.J.; Merle P.; Rosmorduc O.; et al. Nivolumab versus sorafenib in advanced hepatocellular carcinoma (CheckMate 459): a randomised, multicentre, open-label, phase 3 trial. *Lancet Oncol.* 2022, 23, 77–90. [https://doi.org/10.1016/S1470-2045\(21\)00604-5](https://doi.org/10.1016/S1470-2045(21)00604-5)
65. Zhu A.X.; Finn R.S.; Edeline J.; Cattani S.; Ogasawara S.; Palmer D.; Verslype C.; Zagonel V.; Fartoux L.; Vogel A.; et al. Pembrolizumab in patients with advanced hepatocellular carcinoma previously treated with sorafenib (KEYNOTE-224): a non-randomised, open-label phase 2 trial. *Lancet Oncol.* 2018, 19, 940–952. [https://doi.org/10.1016/S1470-2045\(18\)30351-6](https://doi.org/10.1016/S1470-2045(18)30351-6)
66. Finn R.S.; Ryoo B.-Y.; Merle P.; Kudo M.; Bouattour M.; Lim H.Y.; Breder V.; Edeline J.; Chao Y.; Ogasawara S.; et al. Pembrolizumab As Second-Line Therapy in Patients With Advanced Hepatocellular Carcinoma in KEYNOTE-240: A Randomized, Double-Blind, Phase III Trial. *J. Clin. Oncol.* 2020, 38, 193–202. <https://doi.org/10.1200/JCO.19.01307>

67. Qin S.; Chen Z.; Fang W.; Ren Z.; Xu R.; Ryoo B.-Y.; Meng Z.; Bai Y.; Chen X.; Liu X.; et al. Pembrolizumab Versus Placebo as Second-Line Therapy in Patients From Asia With Advanced Hepatocellular Carcinoma: A Randomized, Double-Blind, Phase III Trial. *J. Clin. Oncol.* 2023, 41, 1434–1443. <https://doi.org/10.1200/JCO.22.00620>
68. Leach D.R.; Krummel M.F.; Allison J.P. Enhancement of antitumor immunity by CTLA-4 blockade. *Science.* 1996, 271, 1734–1736. <https://doi.org/10.1126/science.271.5256.1734>
69. Iwai Y.; Ishida M.; Tanaka Y.; Okazaki T.; Honjo T.; Minato N. Involvement of PD-L1 on tumor cells in the escape from host immune system and tumor immunotherapy by PD-L1 blockade. *Proc. Natl. Acad. Sci. USA.* 2002, 99, 12293–12297. <https://doi.org/10.1073/pnas.192461099>
70. Shiravand Y.; Khodadadi F.; Kashani S.M.A.; Hosseini-Fard S.R.; Hosseini S.; Sadeghirad H.; Ladwa R.; O'Byrne K.; Kulasinghe A. Immune Checkpoint Inhibitors in Cancer Therapy. *Curr. Oncol.* 2022, 29, 3044–3060. <https://doi.org/10.3390/currncol29050247>
71. Ang C.; Klemptner S.J.; Ali S.M.; Madison R.; Ross J.S.; Severson E.A.; Fabrizio D.; Goodman A.; Kurzrock R.; Suh J.; Millis S.Z. Prevalence of established and emerging biomarkers of immune checkpoint inhibitor response in advanced hepatocellular carcinoma. *Oncotarget.* 2019, 10, 4018–4025. <https://doi.org/10.18632/oncotarget.26998>
72. Zhu A.X.; Abbas A.R.; de Galarreta M.R.; Guan Y.; Lu S.; Koeppen H.; Zhang W.; Hsu C.H.; He A.R.; Ryoo B.Y.; et al. Molecular correlates of clinical response and resistance to atezolizumab in combination with bevacizumab in advanced hepatocellular carcinoma. *Nat. Med.* 2022, 28, 1599–1611. <https://doi.org/10.1038/s41591-022-01868-2>
73. Kudo M. Pembrolizumab for the Treatment of Hepatocellular Carcinoma. *Liver Cancer.* 2019, 8, 143–154. <https://doi.org/10.1159/000500143>
74. Pinato D.J.; Mauri F.A.; Spina P.; Cain O.; Siddique A.; Goldin R.; Victor S.; Pizio C.; Akarca A.U.; Boldorini R.L.; et al. Clinical implications of heterogeneity in PD-L1 immunohistochemical detection in hepatocellular carcinoma: the Blueprint-HCC study. *Br. J. Cancer.* 2019, 120, 1033–1036. <https://doi.org/10.1038/s41416-019-0466-x>
75. Pfister D.; Núñez N.G.; Pinyol R.; Govaere O.; Pinter M.; Szydłowska M.; Gupta R.; Qiu M.; Deczkowska A.; Weiner A.; et al. NASH limits anti-tumour surveillance in immunotherapy-treated HCC. *Nature.* 2021, 592, 450–456. <https://doi.org/10.1038/s41586-021-03362-0>
76. Cheng A.L.; Qin S.; Ikeda M.; Galle P.R.; Ducreux M.; Kim T.Y.; Lim H.Y.; Kudo M.; Breder V.; Merle P.; et al. Updated efficacy and safety data from IMbrave150: Atezolizumab plus bevacizumab vs. sorafenib for unresectable hepatocellular carcinoma. *J. Hepatol.* 2022, 76, 862–873. <https://doi.org/10.1016/j.jhep.2021.11.030>
77. Rimini M.; Rimassa L.; Ueshima K.; Burgio V.; Shigeo S.; Tada T.; Suda G.; Yoo C.; Cheon J.; Pinato D.J.; et al. Atezolizumab plus bevacizumab versus lenvatinib or sorafenib in non-viral unresectable hepatocellular carcinoma: an international propensity score matching analysis. *ESMO Open.* 2022, 7, 100591. <https://doi.org/10.1016/j.esmoop.2022.100591>
78. Spranger S.; Bao R.; Gajewski T.F. Melanoma-intrinsic β -catenin signalling prevents anti-tumour immunity. *Nature.* 2015, 523, 231–235. <https://doi.org/10.1038/nature14404>
79. Luke J.J.; Bao R.; Sweis R.F.; Spranger S.; Gajewski T.F. WNT/ β -catenin Pathway Activation Correlates with Immune Exclusion across Human Cancers. *Clin. Cancer Res.* 2019, 25, 3074–3083. <https://doi.org/10.1158/1078-0432.CCR-18-1942>
80. Harding J.J.; Nandakumar S.; Armenia J.; Khalil D.N.; Albano M.; Ly M.; Shia J.; Hechtman J.F.; Kundra R.; El Dika I.; et al. Prospective Genotyping of Hepatocellular Carcinoma: Clinical Implications of Next-Generation Sequencing for Matching Patients to Targeted and Immune Therapies. *Clin. Cancer Res.* 2019, 25, 2116–2126. <https://doi.org/10.1158/1078-0432.CCR-18-2293>
81. Ueno A.; Masugi Y.; Yamazaki K.; Komuta M.; Effendi K.; Tanami Y.; Tsujikawa H.; Tanimoto A.; Okuda S.; Itano O.; et al. OATP1B3 expression is strongly associated with Wnt/ β -catenin signalling and represents the transporter of gadoxetic acid in hepatocellular carcinoma. *J. Hepatol.* 2014, 61, 1080–1087. <https://doi.org/10.1016/j.jhep.2014.06.008>
82. Aoki T.; Nishida N.; Ueshima K.; Morita M.; Chishina H.; Takita M.; Hagiwara S.; Ida H.; Minami Y.; Yamada A.; et al. Higher Enhancement Intrahepatic Nodules on the Hepatobiliary Phase of Gd-EOB-DTPA-Enhanced MRI as a Poor Responsive Marker of Anti-PD-1/PD-L1 Monotherapy for Unresectable Hepatocellular Carcinoma. *Liver Cancer.* 2021, 10, 615–628. <https://doi.org/10.1159/000518048>
83. Kubo A.; Suda G.; Kimura M.; Maehara O.; Tokuchi Y.; Kitagataya T.; Ohara M.; Yamada R.; Shigesawa T.; Suzuki K.; et al. Characteristics and Lenvatinib Treatment Response of Unresectable Hepatocellular Carcinoma with Iso-High Intensity in the Hepatobiliary Phase of EOB-MRI. *Cancers (Basel).* 2021, 13, 3633. <https://doi.org/10.3390/cancers13143633>
84. Kuwano A.; Tanaka K.; Yada M.; Nagasawa S.; Morita Y.; Masumoto A.; Motomura K. Therapeutic efficacy of lenvatinib for hepatocellular carcinoma with iso-high intensity in the hepatobiliary phase of Gd-EOB-DTPA-MRI. *Mol. Clin. Oncol.* 2022, 16, 53. <https://doi.org/10.3892/mco.2021.2486>

85. Murai H.; Kodama T.; Maesaka K.; Tange S.; Motooka D.; Suzuki Y.; Shigematsu Y.; Inamura K.; Mise Y.; Saiura A.; et al. Multiomics identifies the link between intratumor steatosis and the exhausted tumor immune microenvironment in hepatocellular carcinoma. *Hepatology*. 2023, 77, 77–91. <https://doi.org/10.1002/hep.32573>
86. Sasaki R.; Nagata K.; Fukushima M.; Haraguchi M.; Miura S.; Miyaaki H.; Soyama A.; Hidaka M.; Eguchi S.; Shigeno M.; et al. Evaluating the Role of Hepatobiliary Phase of Gadoteric Acid-Enhanced Magnetic Resonance Imaging in Predicting Treatment Impact of Lenvatinib and Atezolizumab plus Bevacizumab on Unresectable Hepatocellular Carcinoma. *Cancers (Basel)*. 2022, 14, 827. <https://doi.org/10.3390/cancers14030827>
87. Kuwano A.; Yada M.; Narutomi F.; Nagasawa S.; Tanaka K.; Kurosaka K.; Ohishi Y.; Masumoto A.; Motomura K. Therapeutic efficacy of atezolizumab plus bevacizumab for hepatocellular carcinoma with WNT/ β -catenin signal activation. *Oncol. Lett.* 2022, 24, 216. <https://doi.org/10.3892/ol.2022.13337>
88. Yamashita T.; Kitao A.; Matsui O.; Hayashi T.; Nio K.; Kondo M.; Ohno N.; Miyati T.; Okada H.; Yamashita T.; et al. Gd-EOB-DTPA-enhanced magnetic resonance imaging and alpha-fetoprotein predict prognosis of early-stage hepatocellular carcinoma. *Hepatology*. 2014, 60, 1674–1685. <https://doi.org/10.1002/hep.27093>
89. Hagiwara S.; Nishida N.; Kudo M. Advances in Immunotherapy for Hepatocellular Carcinoma. *Cancers (Basel)*. 2023, 15, 2070. <https://doi.org/10.3390/cancers15072070>
90. Sia D.; Jiao Y.; Martinez-Quetglas I.; Kuchuk O.; Villacorta-Martin C.; Castro de Moura M.; Putra J.; Camprecios G.; Bassaganyas L.; Akers N.; et al. Identification of an Immune-specific Class of Hepatocellular Carcinoma, Based on Molecular Features. *Gastroenterology*. 2017, 153, 812–826. <https://doi.org/10.1053/j.gastro.2017.06.007>
91. Montironi C.; Castet F.; Haber P.K.; Pinyol R.; Torres-Martin M.; Torrens L.; Mesropian A.; Wang H.; Puigvehi M.; Maeda M.; et al. Inflamed and non-inflamed classes of HCC: a revised immunogenomic classification. *J. Hepatol.* 2023, 72, 129–140. <https://doi.org/10.1136/gutjnl-2021-325918>
92. Luke J.J.; Bao R.; Sweis R.F.; Spranger S.; Gajewski T.F. WNT/ β -catenin Pathway Activation Correlates with Immune Exclusion across Human Cancers. *Clin. Cancer Res.* 2019, 25, 3074–3083. <https://doi.org/10.1158/1078-0432.CCR-18-1942>
93. Scheiner B.; Pomej K.; Kirstein M.M.; Hucke F.; Finkelmeier F.; Waidmann O.; Himmelsbach V.; Schulze K.; von Felden J.; Fründt T.W.; et al. Prognosis of patients with hepatocellular carcinoma treated with immunotherapy - development and validation of the CRAFTY score. *J. Hepatol.* 2022, 76, 353–363. <https://doi.org/10.1016/j.jhep.2021.09.035>
94. Hatanaka T.; Kakizaki S.; Hiraoka A.; Tada T.; Hirooka M.; Kariyama K.; Tani J.; Atsukawa M.; Takaguchi K.; Itobayashi E.; et al. Prognostic impact of C-reactive protein and alpha-fetoprotein in immunotherapy score in hepatocellular carcinoma patients treated with atezolizumab plus bevacizumab: a multicenter retrospective study. *Hepatol. Int.* 2022, 16, 1150–1160. <https://doi.org/10.1007/s12072-022-10358-z>
95. Capone M.; Giannarelli D.; Mallardo D.; Madonna G.; Festino L.; Grimaldi A.M.; Vanella V.; Simeone E.; Paone M.; Palmieri G.; et al. Baseline neutrophil-to-lymphocyte ratio (NLR) and derived NLR could predict overall survival in patients with advanced melanoma treated with nivolumab. *J. Immunother. Cancer.* 2018, 6, 74. <https://doi.org/10.1186/s40425-018-0383-1>
96. Bilen M.A.; Dutcher G.M.A.; Liu Y.; Ravindranathan D.; Kissick H.T.; Carthon B.C.; Kucuk O.; Harris W.B.; Master V.A. Association Between Pretreatment Neutrophil-to-Lymphocyte Ratio and Outcome of Patients With Metastatic Renal-Cell Carcinoma Treated With Nivolumab. *Clin. Genitourin. Cancer.* 2018, 16, e563–e575. <https://doi.org/10.1016/j.clgc.2017.12.015>
97. Ogata T.; Satake H.; Ogata M.; Hatachi Y.; Inoue K.; Hamada M.; Yasui H. Neutrophil-to-lymphocyte ratio as a predictive or prognostic factor for gastric cancer treated with nivolumab: a multicenter retrospective study. *Oncotarget*. 2018, 9, 34520–34527. <https://doi.org/10.18632/oncotarget.26145>
98. Bagley S.J.; Kothari S.; Aggarwal C.; Bauml J.M.; Alley E.W.; Evans T.L.; Kosteva J.A.; Ciunci C.A.; Gabriel P.E.; Thompson J.C.; et al. Pretreatment neutrophil-to-lymphocyte ratio as a marker of outcomes in nivolumab-treated patients with advanced non-small-cell lung cancer. *Lung Cancer*. 2017, 106, 1–7. <https://doi.org/10.1016/j.lungcan.2017.01.013>
99. Eso Y.; Takeda H.; Taura K.; Takai A.; Takahashi K.; Seno H. Pretreatment Neutrophil-to-Lymphocyte Ratio as a Predictive Marker of Response to Atezolizumab Plus Bevacizumab for Hepatocellular Carcinoma. *Curr. Oncol.* 2021, 28, 4157–4166. <https://doi.org/10.3390/curroncol28050352>
100. Tada T.; Kumada T.; Hiraoka A.; Hirooka M.; Kariyama K.; Tani J.; Atsukawa M.; Takaguchi K.; Itobayashi E.; Fukunishi S.; et al. Neutrophil-lymphocyte ratio predicts early outcomes in patients with unresectable hepatocellular carcinoma treated with atezolizumab plus bevacizumab: a multicenter analysis. *Eur. J. Gastroenterol. Hepatol.* 2022, 34, 698–706. <https://doi.org/10.1097/MEG.0000000000002356>
101. Reig M.; Forner A.; Rimola J.; Ferrer-Fàbrega J.; Burrel M.; Garcia-Criado Á.; Kelley R.K.; Galle P.R.; Mazzaferro V.; Salem R.; et al. BCLC strategy for prognosis prediction and treatment recommendation: The 2022 update. *J. Hepatol.* 2022, 76, 681–693. <https://doi.org/10.1016/j.jhep.2021.11.018>

102. Kudo M. Durvalumab Plus Tremelimumab: A Novel Combination Immunotherapy for Unresectable Hepatocellular Carcinoma. *Liver Cancer*. 2022, 11, 87–93. <https://doi.org/10.1159/000523702>
103. Myojin Y.; Kodama T.; Sakamori R.; Maesaka K.; Matsumae T.; Sawai Y.; Imai Y.; Ohkawa K.; Miyazaki M.; Tanaka S.; et al. Interleukin-6 Is a Circulating Prognostic Biomarker for Hepatocellular Carcinoma Patients Treated with Combined Immunotherapy. *Cancers (Basel)*. 2022, 14, 883. <https://doi.org/10.3390/cancers14040883>
104. Kuwano A.; Yada M.; Miyazaki Y.; Tanaka K.; Kurosaka K.; Ohishi Y.; Masumoto A.; Motomura K. Tumor-infiltrating CD8+ T cells as a biomarker for chemotherapy efficacy in unresectable hepatocellular carcinoma. *Oncol. Lett.* 2023, 25, 259. <https://doi.org/10.3892/ol.2023.13845>

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