

Supportive information

Emergence and progression of behavioural motor deficits and skeletal muscle atrophy across adult lifespan of the rat

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Details of methods and chemicals used in this study according to previously published protocols by the authors [1; 2; 3; 4]

Microscopy and Stereology

Fig. S1

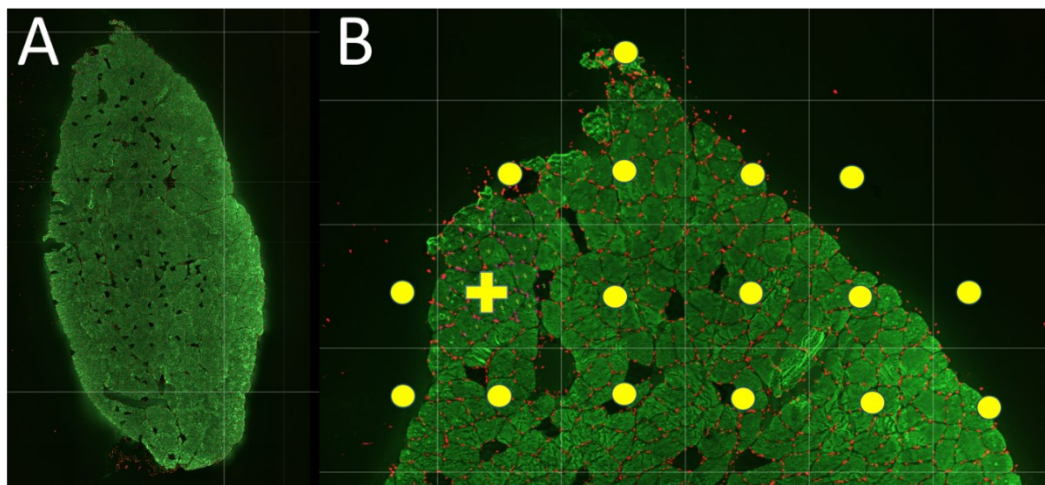


Fig. S1 Photomontage of confocal microscopic images covering the full cross-sectional area of m. soleus labelled for MyHC I (**A**). In (**B**) blow-up of the top part of the cross-section in (**A**). Overlay (in yellow) is a grid where each square corresponds to 300 kpixels to enable unbiased systematic sampling (**B**). The squares are counted (yellow dots) starting from the top, reading left to right (numbers). First square is generated from random numbers (1-10) and is in this example 7 (yellow cross) and then every 15th subsequent square is used until 200 fibers have been collected. Fibers completely inside the grid box or touching/stretching outside the top or left borders were included while fiber not confined inside of the right or lower borders of the grid box were excluded.

Fig. S2

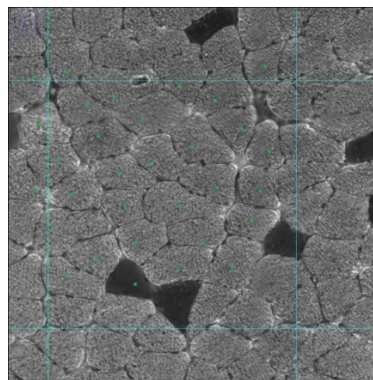


Fig. S2 Close up micrograph of a section through m. soleus stained with an antibody towards MyHC 1(slow). Superimposed is the grid (green lines) subdividing the cross sections into grid boxes used for an unbiased sampling of fibers to be included in the count. Fibers completely inside the grid box or touching/stretching outside the top or left borders were included (green dots) while fiber not confined inside of the right or lower borders of the grid box were excluded.

Hematoxylin and eosin staining protocol.

1. Air dry slides at room temperature.
2. Rehydrate tissue with 0.1M PBS (phosphate-buffered saline) for 1-2 minutes.
3. Immerse slides in Mayer's hematoxylin for 5 minutes.
4. Rinse slides in ordinary water for 3x10 minutes on Stovall Belly dancer 4702610 with two changes of water in between.
5. Immerse slides in eosin 0.2% for 8 minutes
6. Immerse slides in 75% and 95% ethanol for 2x1 minutes respectively.
7. Mount slides with 2.5% DABCO in glycerol.

Immunohistochemistry protocol

Tissues were dried in RT for 30 min. A well was created using a DAKO pen. The tissue was then fixated in 4% formaldehyde for 10 minutes then rinsed in cold tap water for 10 minutes repeated three times. Blocking was performed with 5% normal donkey serum for one hour in RT. Primary antibody was incubated for either 1h at 37 °C or overnight at 4 °C. In case of dual epitope labeling, this step was repeated following washing. After primary antibody incubation, the washing sequence above was repeated. Secondary antibody was incubated for one hour at RT covered from light. The samples were mounted with DAPI-antifade (Vectashield, H-1200) and left to incubate covered from exposure to light for 20 min at RT before imaging or storage in 8-12 °C. DAPI served as unspecific nuclear stain in microscopical multicolor analyses.

Table S1 of primary and secondary antibodies used.

Source	Species	Type of ab	Dilution
sMHC (MyHC I)	Mo	Novocastra	1:10
fMHC (MyHC II)	Mo	Novocastra	1:50
BF-G6(MyHC 3)	Mo	Developmental Studies Hybridoma Bank, Ohio 2011	3µg/ml
MyHC 3	Mo	Novocastra	1:50
Laminin PC128	Sheep	The Binding Site	0.7-1ug/ml
Laminin L9393	Rb	Sigma	1:800
Pax7	Mo	DS Hybridoma Bank	1-3 ug/ml
Ki67	Rb	Novocastra	1:1000

RNA extraction, cyber-green PCR, and primers

12-24mg of tissue

⅓ of 2ml vial filled with zirconia beads

1ml Trizol to each screw top vial

Add biopsy

Homogenate 15sec/round, check that all samples dissolved, if still solid tissue re-run only that vial

Incubate 5min at RT

Add 200ul of chloroform to each vial

Shake 15 sec, all at once kept in the rack

Incubate 3 min at RT (when doing several samples incubate on ice until all samples are processed → initial phase separation starts)

Spin at 12000g 15 min at 4 °C (samples separated into upper RNA and lower DNA+protein layer)

Collect upper aqueous phase and place in RNA-vial. Do not disturb the interphase!

Add 500ul of isopropanol (2-propanol) to the RNA-phase → immediately invert tube manually and vortex full speed (about 30 sec)

Incubate at RT for 10 min

Spin at 12000g for 10 min at 4 °C

Prepare 75% ethanol (dilute 100% EtOH with milliQwater → pre-cool on ice

Gently pour off the liquid (in hazard wastes in the hood), careful not to lose the pellet. The last part of fluid is removed with 100ul pipette

Add 1 ml (ice-cold) 75% ethanol to the pellet and wash by vortexing gently (the pellet should loosen from the bottom of tube)

Spin at 7500g for 10 min at 4 °C

Discard supernatant, remove the last amount of liquid using pipette.

Semi-dry the pellet 5 min at RT

add 20ul MilliQ water on top of the RNA pellet

Leave at RT for 5-10 min

Vortex carefully for about 1 min to dissolve pellet. Quick spin if necessary to collect drops on walls

Store at -80 °C

Concentration measurement

Was done with PicoDrop; Picopet01, Picodrop Ltd, Cambridge UK

RNA integrity

RNA integrity was measured with Experion™ RNA StdSens & HighSens Analysis Kits

Reverse Transcription

Applied Biosystems High-Capacity cDNA Reverse Transcription Kit. 300ng RNA in a 20ul reaction volume diluted 1:20 for a final calculated cDNA-concentration of 0,75ng/μl.

Incubate at 25 °C for 10 min

Transcription 2h at 37 °C

Inactivate at 85 °C for 5 s

PCR

Enzymes Dynamo Colorflash SYBR Green qPCR Kit

Primer (20uM) U+L 0,5ul / well

cDNA (0,75ng/μl) 2,5ul / well

2X Mastermix 6,25μl / well

Nucleotide free H₂O 3,25ul / well

Reaction volume 12,5 μl

Holding stage 7 min 95 °C

Cycling stage 10s 95 °C, 30s 60C X40

Melting Curve 15s 95 °C, 1 min 60 C, slow incremental heating up to 95 C

Table S2 of primers used in qPCR

Gene	Accession number	Forward primer	Reverse primer
GPx1	NM_030826	CCGGGACTACACCGAAATGA	CTGATGTCCGAACTGATTGCA
β Actin			
AChR γ (CHRN γ)	NM_009604	GATGCAATGGTGCGACTATCGC	GCCTCCGGGTCAATGAAGATCC
MuRF1 (TRIM63)	NM_080903.1	TGTCTGGAGGTCGTTTCCG	ATGCCGGTCCATGATCACTT
Atrogin (MAFbx/FBXO32)	NM_133521	CCATCAGGAGAAGTGGATCTATGT	GCTTCCCCCAAAGTGCAGTA
Smad3	NM_013095	CATCTACTGCCGCTTGTGG	TCATGTTGAAGGCGAACTCAC
Myogenin (MYOG)	NM_017115.2	ACCATGCCCAACTGAGATT	TGGGCAGGGTGTTAGTCTT
MyoD (MYF3, MYOD1)	M84176	GACGCCGCCTACTACAGTGAG	CCACCCCTCAGTGCTCCTGCAGG
TGF- β 1	NM_021578	CTGAACCAAGGAGACGGAATAC	CCAGGTCACCTCGACGTT
β -Catenin (CTNNB1)	NM_053357	TCACCACGCTGCATAATC	ATAGCCAAGAACTTCACGTTT
Glial-derived neurotrophic factor (GDNF)	NM_012528	AAATCGGGGGTGCGTCTT	ATGCCTGGCCTACCTTGTC
Neural cell adhesion molecule (NCAM, CD56)	NM_031521	CAGCGGATCTCAGTGGTGTG	TCTCGGCCTTTGTGTTCCAG

Confocal microscopy

Microscopy and Image Processing. Representative images were acquired from one airy unit pinhole on an LSM700 confocal laser scanning microscope (Carl Zeiss) equipped with an objective with a magnification of x5, an EC Plan-Neofluar objective with a magnification of x10 and N.A. of 0.30, a Plan-Apochromat M27 objective with a magnification of x20 and N.A. of 0.80. Emission spectra for each dye were filtered as follows: FITC (505–540 nm), Cy3 (560–610 nm), and Cy5 (>640 nm). Images were processed using ZEN2012 software (Zeiss). Multi panel figures were assembled using Adobe Photoshop CS6 software (Adobe Systems). Fiber type imaging was done according to Zhang et al 2014[5] adjusted for our circumstances. We thank Dr Jan Mulder for generously allowing us to use the equipment.

Supportive information of Results

Fig S3

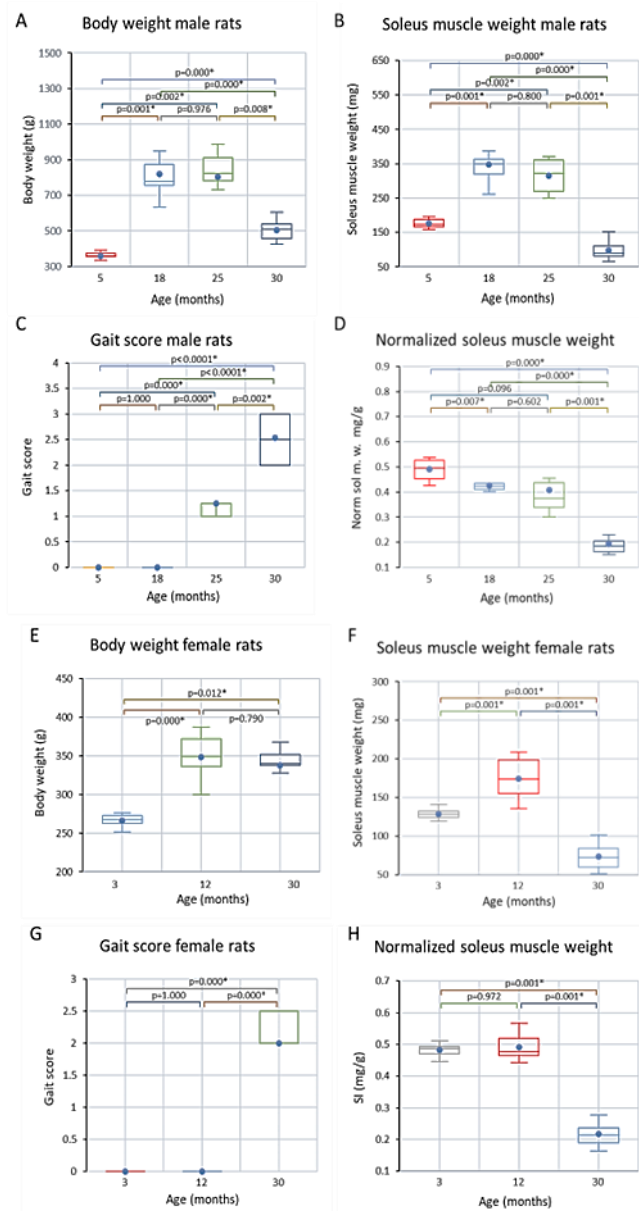


Fig. S3 Boxplots of male (A-D) and female (E-H) rats whole body weight (A, E), m. soleus muscle weight (B, F), gait score (C, G) and soleus muscle weight normalized to body weight (D, H) across adult lifespan. The variation across lifespan was significant for each parameter in both sexes (KW < 0.001). In the plots, the outcome of pairwise post hoc testing has been indicated.

Fig. S4

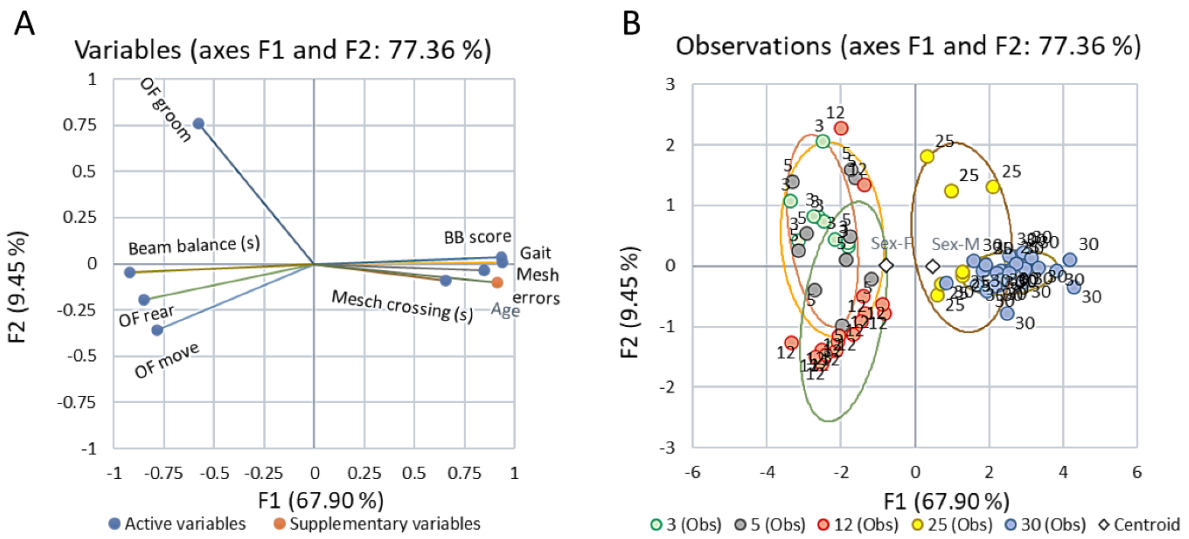


Fig S4 PCA analysis of (A) activity, motor, and coordination indicis in (B) male and female age cohorts (3-, 5-, 12- and 30-month-old) when also observations on early aged males (25-month-old; yellow markers) were included. The 25-month-old males formed a group of observations positioned in between middle aged and 30-month-old rats. Ovals indicate the 70% confidence limit of each age group.

Fig S5

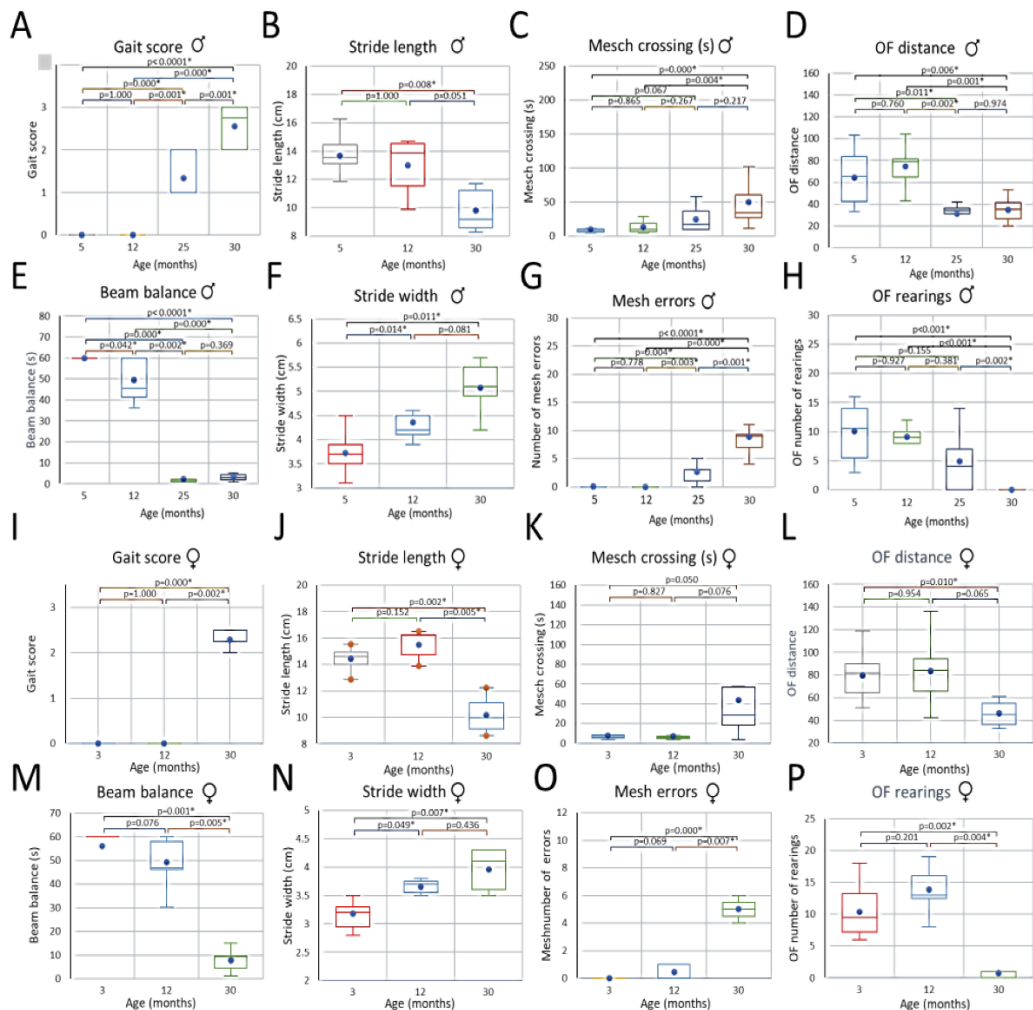


Fig S5 A-G (males) and I-P (females) show boxplots of the behavioral variables assessed across adult life span. In each panel the outcomes of the post hoc testing have been indicated.

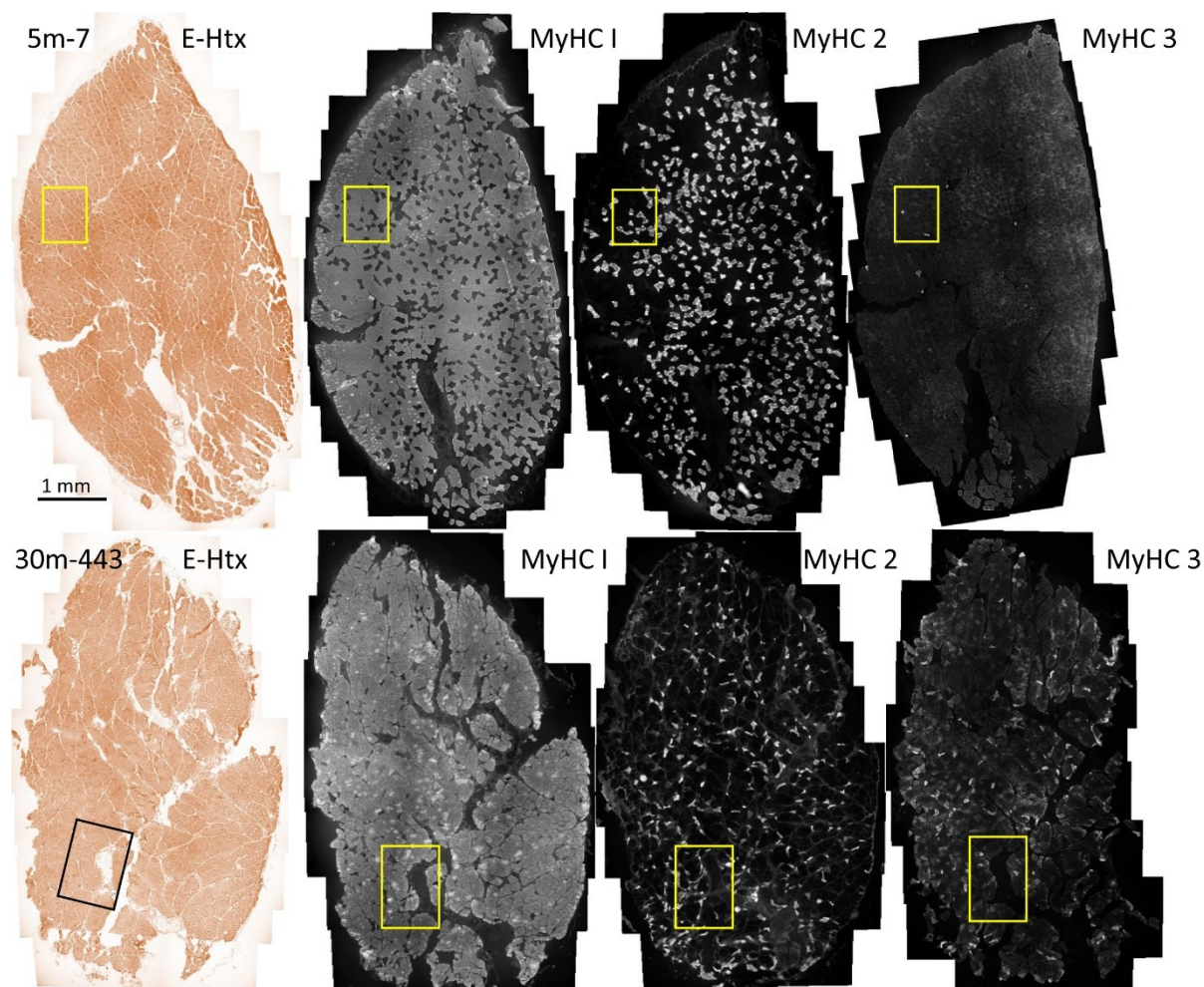


Table S3

Fig S7

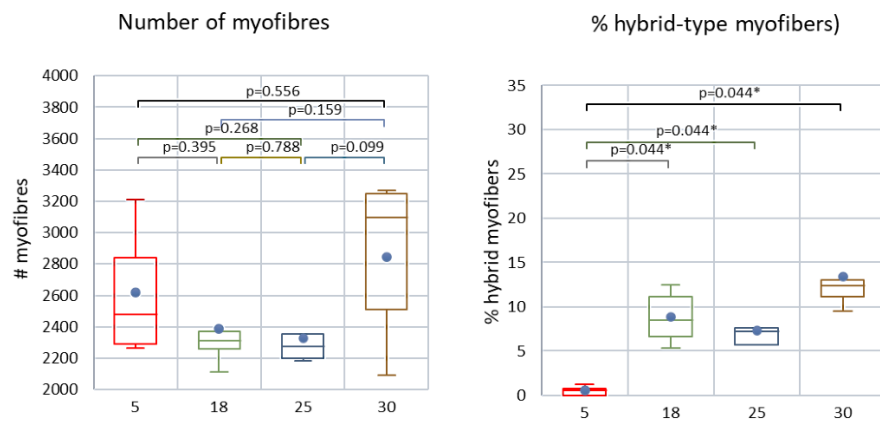


Fig. S7 Boxplots of metrics (A-B) on male m soleus during aging. Age groups have been indicated on the abscissa and metric on the ordinate. In A-B the pairwise post hoc testing results have been indicated.

Fig S8

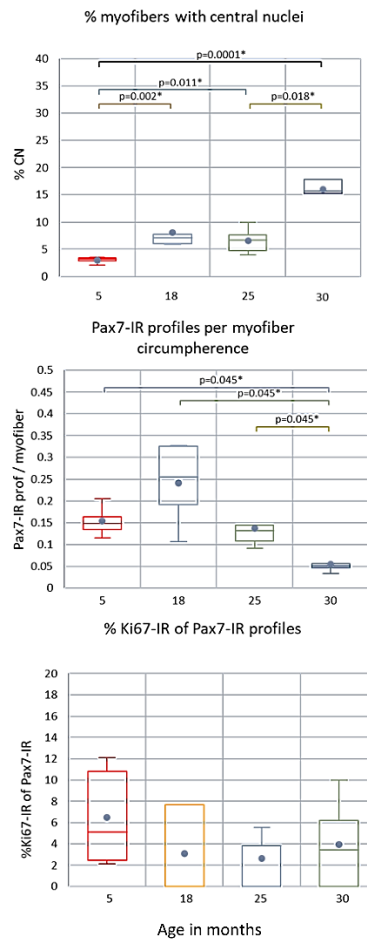


Fig. S8 Boxplots of metrics on male m soleus during aging. Age groups have been indicated on the abscissa and metric on the ordinate of the panels. Kruskal-Wallis analysis of variance indicated that each metric except prevalence of Ki67-IR in Pax7-IR profiles (lower panel) changed significantly during aging. Top and middle panels show pairwise post hoc testing results indicated when significant.

Fig. S9

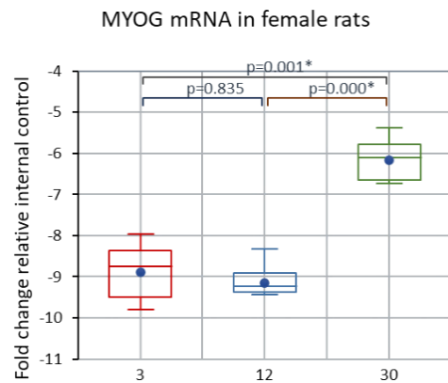


Fig.S9 Boxplots of MYOG mRNA abundance in m soleus during aging of female rats. Age groups have been indicated on the abscissa and fold difference relative internal control on the ordinate. Kruskal-Wallis analysis of variance indicated that mRNA changed in a significant way during aging. Pairwise post hoc testing results have been indicated where they were significant.

Cited literature

- [1] M. Grönholdt-Klein, M. Altun, M. Becklén, E. Dickman Kahm, A. Fahlström, E. Rullman, and B. Ulfhake, Muscle atrophy and regeneration associated with behavioural loss and recovery of function after sciatic nerve crush. *Acta Physiol (Oxf)* 227 (2019) e13335.
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