

Review

Not peer-reviewed version

Inflammatory Cytokines in Psoriatic Arthritis: Understanding Pathogenesis and Implications for Treatment

Bong-Woo Lee and Su-Jin Moon *

Posted Date: 28 June 2023

doi: 10.20944/preprints202306.2029.v1

Keywords: psoriatic arthritis; inflammatory cytokines; tumor necrosis factor-alpha; interleukin-17; interleukin-23; JAK/STAT signaling pathway



Preprints.org is a free multidiscipline platform providing preprint service that is dedicated to making early versions of research outputs permanently available and citable. Preprints posted at Preprints.org appear in Web of Science, Crossref, Google Scholar, Scilit, Europe PMC.

Copyright: This is an open access article distributed under the Creative Commons Attribution License which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Disclaimer/Publisher's Note: The statements, opinions, and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions, or products referred to in the content.

Review

Inflammatory Cytokines in Psoriatic Arthritis: Understanding Pathogenesis and Implications for Treatment

Bong-Woo Lee 1 and Su-Jin Moon 2,*

- Division of Rheumatology, Department of Internal Medicine, Seoul St. Mary's Hospital, College of Medicine, The Catholic University of Korea, Seoul, Republic of Korea; tdomu1029@gmail.com
- ² Division of Rheumatology, Department of Internal Medicine, Yeouido St. Mary's Hospital, College of Medicine, The Catholic University of Korea, Seoul, Republic of Korea; prajna79@catholic.ac.kr
- * Correspondence: prajna79@catholic.ac.kr; Tel.: + 82 2-3779-2417 (SJM)

Abstract: Psoriatic arthritis (PsA) is a persistent, inflammatory disease that affects individuals with psoriasis, arthritis, and enthesitis. Research has demonstrated that inflammatory cytokines such as tumor necrosis factor-alpha (TNF- α), interleukin-23 (IL-23), and interleukin-17 (IL-17) play a pivotal role in both the onset and progression of PsA. These cytokines are generated by activated immune cells and stimulate the attraction of inflammatory cells to the synovium and joint tissues, resulting in the deterioration of cartilage and bone. The blocking of these cytokines has become a successful treatment strategy for PsA, as biological drugs that inhibit TNF- α , IL-23, and IL-17 have demonstrated notable clinical benefits. The association between PsA and other types of inflammatory cytokines or chemokines, excluding TNF- α , IL-23, and IL-17, has been extensively investigated in numerous studies. These findings may provide a chance for the discovery of novel therapeutic agents targeting other molecules, distinct from the currently approved biologics and targeted synthetic disease-modifying antirheumatic drugs. In this review, we discuss the current understanding of the role of inflammatory cytokines in PsA pathogenesis and clinical implications of targeting these cytokines for PsA treatment.

Keywords: psoriatic arthritis, inflammatory cytokines, tumor necrosis factor-alpha, interleukin-17, interleukin-23, JAK/STAT signaling pathway

1. Introduction

Psoriatic arthritis (PsA) is a persistent condition associated with immune-related inflammation that affects joints and other elements of the musculoskeletal system, in addition to skin manifestations. It is estimated that this disease affects approximately 1–2% of the worldwide population [1]. PsA is recognized as part of psoriatic diseases and belongs to a complex group of spondyloarthritides (SpA) [2–4].

PsA is identified by a varied range of clinical manifestations, such as peripheral joint inflammation, inflammatory back pain, uveitis, enthesitis, tenosynovitis, psoriasis, and nail changes [5]. Historically, PsA was regarded as a relatively benign condition. However, it is now widely acknowledged that the functional impact of PsA is comparable to that of other types of inflammatory arthritis, including rheumatoid arthritis (RA) and axial SpA (axSpA) [1]. In many individuals with PsA, the disease exhibits characteristics of progressive and destructive changes. Significant pathological changes occur already in the early stages of PsA, with approximately half of the patients displaying structural damage within 2 years from disease onset [6]. Research has demonstrated that a delay in diagnosing PsA is linked to unfavorable outcomes impacting the quality of life [7].

While the precise cause of PsA is not fully comprehended, it is believed to arise from a complex interplay of environmental, genetic, and immunological factors [2]. Specific human leukocyte antigen (HLA) alleles, namely HLA-B*27, HLA-B*08, HLA-B*38, and HLA-B*39, exhibit a strong correlation

with the prevalence of PsA [8]. It is hypothesized that these alleles may render individuals more susceptible to PsA by modifying the immune response to self-antigens. Furthermore, environmental factors, including infections and mechanical stress, can potentially act as triggers for PsA development in individuals with a genetic predisposition [9,10]. The microbiome of the gut and skin might have an impact on the development of PsA [11,12]. Immunological factors, including cytokines and chemokines, are believed to have a crucial role in PsA pathogenesis. Tumor necrosis factor-alpha (TNF- α), interleukin-17 (IL-17), and interleukin-23 (IL-23) have been recognized as significant contributors to the inflammatory response, tissue damage, and bone erosion associated with PsA. Therapies targeting these cytokines have been developed and used as treatments for patients with PsA. Despite advancements in comprehending the molecular mechanisms that cause PsA, the existing knowledge is not sufficient to meet significant clinical needs in effective treatments [13]. Overall, PsA pathogenesis is complex and multifactorial, involving genetic, environmental, and immunological factors.

The improved understanding of PsA pathogenesis has resulted in the development of powerful biologics and small-molecule drugs that selectively target cytokines and signaling pathways. However, due to the complex pathogenesis of PsA involving both innate and acquired immune responses, as well as the highly heterogeneous presentation of the disease among individuals, many current treatments fall short of achieving the desired outcome. For this reason, further research on the roles of other inflammatory cytokines in the pathogenesis of PsA could contribute to the search for valid diagnostic biomarkers, factors for predicting treatment response, and new target molecules for treating this disease. In this review, we summarize the latest information about inflammatory cytokines and possible therapeutic target molecules in PsA.

2. Immune cells in PsA pathogenesis

PsA is a chronic and complex condition characterized by the involvement of the skin, enthesis, and synovium. The degree of this involvement may vary among individuals and is influenced by genetic and environmental factors [14]. PsA is initiated by the stimulation of dendritic cells (DCs) and macrophages. These cells expose antigens to T cells through the toll-like receptor-2 (TLR-2) signaling pathway, facilitated by major histocompatibility complex (MHC) I. This activation occurs in response to a combination of genetic predisposition, environmental conditions, and biophysical factors. Consequently, the activation of both innate and adaptive immune responses leads to the increased secretion of various cytokines, such as IL-1, IL-6, TNF- α , IL-17, and IL-23 [15]. In addition, the activation and invasion of T cells and macrophages are considered essential in the initiation of inflammatory and destructive processes in the joints [16].

Specifically, DCs induce T cell differentiation by presenting antigens and secreting various proinflammatory cytokines. Increased expression of TLR-2 has been detected in immature DCs of PsA patients [17]. Patients with PsA exhibit an elevated proportion of immature myeloid DCs compared to plasmacytoid DCs in their synovial fluid [18]. In PsA, TLR stimulation leads to the polarization of T cells towards the Th1 subset, subsequently increasing the production of TNF α , IFN- γ , and IL-2 [17]. Plasmacytoid DCs generate cytokines, including IFN- γ , TNF- α , IL-12, and IL-23, which subsequently serve as signals for clonal proliferation of CD4+ and CD8+ T cells [19].

Activated macrophages participate in various proinflammatory processes within the synovium. A reduction in the quantity of CD68+ macrophages was observed in the synovium of patients with PsA who responded to treatment, which highlighted the role of these cells in PsA pathogenesis [20]. Skin inflammation induced by a TLR-7 ligand in murine models was associated with the infiltration of CD68+ macrophages and stimulation of inflammatory cytokines expression in the joints. This study implied that macrophages could contribute to the inflammatory process in PsA and may be a crucial factor in the skin-to-joint inflammation transition [21]. In RA, M1 macrophages become prevalent, which leads to the secretion of high levels of proinflammatory cytokines, activation of T and B cells through antigen presentation, and stimulation of bone resorption. Similarly, administration of a TLR-7 agonist increases the M1/M2 macrophage ratio in psoriatic skin lesions [22]. On the contrary, there

is no difference in M2 cytokine expression between PsA and RA, but PsA is marked by lower expression levels of M1 cytokines compared to those in RA [23].

Mast cells located in the synovium induce angiogenesis, neutrophil recruitment, and proliferation of synovial fibroblasts, indicating that these cells may actively contribute to the inflammatory arthritis [24]. In the synovium of peripheral SpA, mast cells are the predominant source of IL-17A, whereas only a small number of IL-17-positive T cells was identified [25]. Likewise, tissue-resident mast cells may increase inflammation by producing IL-17A. However, it remains to be established in more detail how the release of IL-17A from mast cells is controlled in the affected tissue during inflammation [26].

Other various cells in the synovial tissue could potentially participate in PsA pathogenesis. Notably, elevated levels of type 3 innate lymphoid cells (ILCs), which produce IL-17 and IL-22, have been observed in the synovial fluid of PsA patients [27]. Conversely, there was a decrease in the number of type 2 ILCs, which typically generate IL-4, IL-5, IL-9, and IL-13. Significantly, the ratio of type 2 ILCs to type 3 ILCs showed a strong correlation with both clinical indicators of disease activity and the radiographic evidence of arthritis and bony destruction. In addition, it has been shown that environmental stress signals activate natural killer (NK) cells, leading to their involvement in the harmful processes associated with PsA development [28,29]. Additionally, IL-15 effectively stimulates dormant NK cells within the synovium, causing them to transition into an effector phase [30].

In the development of PsA, innate immune cells contribute by producing IL-12 and IL-23, which stimulate the differentiation of T cells into distinct subtypes referred to as Th1 and Th17 helper T cells [5,31]. Consequently, IL-22 and IL-17 are secreted, which in turn increase the secretion of TNF- α [32]. T cells are the primary contributors to PsA pathogenesis, as evidenced by the elevation of CD4+ T17 cells that secrete IL-23 and IL-17 in both the peripheral blood and synovial fluid [33-35]. Furthermore, an accumulation of cells producing IL-17 was detected within the affected joints of PsA patients, and this group may also include IL-17 secreting CD8+ T cells [36]. Another study conducted on PsA patients unveiled a notable increase in the quantity of memory CD8+T cells within the synovial fluid, surpassing their levels detected in the peripheral blood. Additionally, these cells were identified as active CD8+ T cells expressing the CXCR3 receptor. Moreover, elevated levels of CXCR3 receptor ligands, such as CXCL9 and CXCL10, were also observed in the synovial fluid [37]. Th1, Th17, and CD8+ cytotoxic T cells have been recognized as crucial contributors to the inflammatory processes in PsA, suggesting the existence of distinct immune cellular pathways in this disease [13]. Generally, regulatory T cells (Tregs) are thought to participate in suppressing chronic inflammation and promoting an anti-inflammatory response. However, research on psoriasis and PsA has yielded conflicting findings regarding the relationship between Treg levels and disease activity [38-40]. The specific function of B cells is not fully understood; however, they can present antigens to T cells [5].

In summary, interactions between diverse types of immune cells lead to the production of various cytokines, such as IL-23, TNF- α , IL-17 (predominantly IL-17A isoform), and IL-22, which promote inflammation and stimulate tissue-resident cells in the joint and enthesis. These cells, including fibroblast-like synoviocytes, chondrocytes, osteoblasts, and osteoclasts, cause the degradation of cartilage, bone erosion, and joint destruction by secreting extracellular matrix degradation enzymes and receptor activator of nuclear factors κB ligand (RANKL). Furthermore, upon stimulation, these cells secrete proinflammatory mediators to attract additional immune cells, thereby establishing a sustained immune response in patients with active PsA [5].

3. Key inflammatory cytokines in PsA

In PsA, inflammation is triggered by cytokines released by activated immune cells (Table 1). Over the past few years, there has been a notable change in research focus regarding the key factors that contribute to the development of PsA. Previously, the focus was primarily on TNF- α , and cytokines associated with Th1 immune response. However, more recent studies have shifted their attention to Th17 cells, as well as IL-23 and IL-17 cytokines. The administration of monoclonal

antibodies targeting specific cytokines, such as IL-23, IL-17A, and TNF- α , has proven to effectively attenuate inflammation and ameliorate clinical manifestations of PsA. It is highly probable that each distinct cytokine possesses a unique pathophysiological role in the development and progression of the disease. IL-23 appears to play a role in initiating the development of the disease, whereas IL-17A primarily enhances the disease process. On the other hand, TNF- α plays a critical role in carrying out downstream effects in the pathogenesis of PsA [41].

3.1. Role of TNF- α in PsA

TNF- α , a pro-inflammatory cytokine produced by cells like macrophages and monocytes, is prominently present during inflammatory processes [42]. It serves as a key component of pro-inflammatory immune responses. TNF- α interacts with two distinct receptors, TNFR1 and TNFR2, initiating separate signaling pathways that trigger apoptosis, cell differentiation, proliferation, and migration. As a result, TNF- α contributes to the development of inflammatory reactions. Additionally, TNF- α plays a crucial role in promoting bone resorption by directly activating osteoclast precursor cells via the RANKL signaling pathway [43]. At high concentrations, TNF- α causes adverse effects, including uncontrolled inflammatory responses, increased formation of osteoclast precursors, and enhanced osteoclast activity, ultimately resulting in bone resorption [44].

TNF- α plays a pivotal role in the pathogenesis of SpA manifestations, encompassing skin, enthesis, joint, and spine involvement, which can be considered as the secondary effects of hyperactivated TNF α -dependent inflammation due to the activation of both innate and adaptive immune responses [45]. The evidence supporting the involvement of TNF- α in inflammatory forms of arthritis is particularly compelling [43]. Elevated levels of TNF- α are detected in the synovial fluid and tissues of patients with RA and PsA. Various cell types, including macrophages and T cells, produce TNF- α , which binds to specific receptors on target cells, thereby activating multiple intracellular signaling pathways. Consequently, the production of other pro-inflammatory cytokines, such as IL-1 and IL-6, is induced, along with the recruitment and activation of immune cells, including neutrophils and monocytes [46].

TNF- α is recognized as a key contributor to the initiation and persistence of joint inflammation and damage in PsA. It facilitates production of enzymes that degrade the cartilage and bone, thereby leading to joint destruction. Moreover, TNF- α induces angiogenesis, which further fuels the progression of joint inflammation [2]. Furthermore, TNF- α plays a crucial role in the development of erosions through the induction of receptor activator of nuclear factor- κ B (NF- κ B) ligand expression in the synovium, a pivotal factor in osteoclast differentiation. Additionally, TNF- α stimulates synovial fibroblasts to express Dickkopf-related protein 1, which inhibits osteoblast differentiation, thereby exacerbating the formation of erosions [47].

3.1. Role of IL-23 in PsA

IL-23 is a cytokine composed of two subunits, p19 and p40, with the latter being shared with IL-12. Its main function is to facilitate the activation, differentiation, and proliferation of Th17 cells, which are responsible for producing effector cytokines like IL-17A and IL-22. However, it is worth noting that IL-17 production can also occur independently of IL-23 [48]. The injection of IL-23 causes a skin disease like psoriasis in mice with normal expression of IL-17, while IL-17 knockout mice do not exhibit such disease manifestations. Moreover, pretreatment of wild-type mice with anti-IL-17A antibodies effectively prevents IL-23-mediated disease development [49]. These findings indicate that IL-23 functions as an upstream regulator of IL-17A, whereas IL-17A, acting downstream, directly impacts tissue physiology.

The physiological processes governed by the IL-23 signaling strongly support its crucial role in maintaining the homeostasis of the intestinal barrier. Intestinal inflammation is linked to the increased permeability of the gut barrier, which facilitates the infiltration of pathogen-associated molecular patterns into the host system. The compromised intestinal barrier leads to a persistent

activation of the host's defense mechanisms, which subsequently triggers an excessive production of IL-23, the cytokine that plays a pivotal role in the development of SpA pathogenesis [50].

IL-23 demonstrates a remarkable capacity to induce inflammation in various anatomical sites, including the gut, skin, enthesis, and joints, with high efficiency. Notably, IL-23 alone could directly induce the characteristic clinical features of psoriasis and PsA in mice. Experimental evidence has shown that IL-23 expression in mice results in the emergence of enthesitis, osteitis, new bone formation, and bony erosion [51]. In mice overexpressing IL-23 specifically in the skin, initial manifestation of psoriatic-like skin lesions was observed, followed by the development of suggestive musculoskeletal features, including enthesitis, synovitis, and dactylitis [52]. IL-23 and its receptor IL-23R appear to play a central role in the pathogenesis of PsA and SpA in general, as they are present in various anatomical sites affected by these conditions [2]. Consequently, various types of immune cells that express IL-23R, including Th17 cells, γ/δ T cells, ILC3s, and CD8*IL-23R* mucosal-associated invariant T cells, are elevated in the joints and entheses of SpA and PsA patients [53]. IL-23 plays a crucial role in mediating inflammation in the skin and gut, acting as a significant link between these pathological conditions and peripheral joints. However, its involvement in axial inflammation seems to be relatively minimal [54].

3.2. Role of IL-17 in PsA

IL-17, a group of pro-inflammatory cytokines comprising six known members (IL-17 A/B/C/D/E/F), is mainly generated by Th17 T cells, and its presence has been associated with the development of psoriatic skin lesions and enthesitis [55]. IL-17 is secreted by multiple cell populations, including Th17 helper cells, γ/δ T cells, interstitial lymphoid cells, neutrophils, and mast cells [2]. Several studies have suggested that a combination of cytokines, including IL-23, TGF- β , IL-6, IL-1 β , and IL-21, can induce the differentiation of naïve T cells into Th17 cells [56]. IL-23 stimulates activated Th17 cells, leading to the release of IL-17. However, clinical trials conducted in human subjects have demonstrated limited efficacy of IL-23 inhibitors in the treatment of axSpA, suggesting the involvement of other cytokines in IL-17 production. This observation implies that IL-17 can also be generated independently of IL-23 [56].

While many members of the IL-17 family contribute to inflammation and participate in immune-related diseases and defense against pathogens, they also have various functions in preserving mucosal integrity, responding to allergens, supporting anti-inflammatory responses, and regulating the function of lymphocytes [57]. Among these members, IL-17A and IL-17F are particularly significant in PsA. IL-17A and IL-17F have the ability to form both homo- and heterodimers, which can bind to the IL-17 receptor and stimulate signaling pathways downstream. This activation leads to the release of pro-inflammatory cytokines. In PsA, IL-17A is considered the main pro-inflammatory cytokine, although IL-17F likely also has some effects [58].

IL-17A has a multitude of effects on diverse immune cells in the skin and joints, stimulating inflammation, coagulation, and causing damage to bones and joints. IL-17A exerts significant biological effects by attracting neutrophils, Th17 cells, and ILC3s to the affected tissues, particularly the skin and entheses. This recruitment process contributes to the generation of the immune response [48]. IL-17A acts as a mediator that aggravates tendon inflammation by initiating neutrophil responses. This inflammatory process clinically presents as enthesitis and tendinitis, which are characteristic features of PsA [59]. Both preclinical and clinical pieces of evidence indicate that bone erosion and new bone formation coexist at various anatomical sites in SpA, and IL-17A appears to have a multifaceted involvement in these mechanisms [60]. IL-17A stimulates stromal cells and macrophages to produce pro-inflammatory cytokines such as TNF, IL-1, and IL-6, which contribute to osteoclastogenesis. Overall, by virtue of its stimulatory effect on osteoclastogenesis, IL-17A plays a catabolic role in bone metabolism. This phenomenon helps elucidate the mechanisms underlying the formation of bone erosions and occurrence of osteopenia and/or osteoporosis that are commonly observed in patients with PsA [61].

3.3. Other cytokines or chemokines in PsA

While the therapeutic efficacy of apremilast, a phosphodiesterase 4 (PDE4) inhibitor, in PsA is linked to its capacity to suppress the production of TNF- α , IL-17, and IL-23, the effectiveness of Janus kinase (JAK) inhibition in PsA treatment suggests the participation of other cytokines in the inflammatory mechanisms underlying this condition [62,63].

IL-22 is a cytokine generated by Th22 cells, a recently identified subset of T helper cells expressing chemokine receptor (CCR)10. Serum levels of IL-22 are notably elevated in patients with PsA compared to those in patients with psoriasis only [64]. IL-32 activates monocytes, promoting their differentiation into macrophages or DC, and also triggers the production of pro-inflammatory cytokines such as TNF-α, IL-1β, IL-6, and IL-8 through the inflammatory signaling pathway involving NF-κB and p38 mitogen-activated protein kinase [65]. Plasma levels of IL-32 are elevated in patients with PsA compared to those in healthy controls, and notably higher than in patients with psoriasis alone. These findings indicate that plasma IL-32 levels in patients with psoriasis only increase prior to PsA onset, and further elevate upon PsA aggravation [66]. IL-33, a member of the IL-1 family, interacts with the ST2 receptor and activates various immune cells, including ILC2 and Th2 cells. Furthermore, the IL-33/ST2 axis appears to play a role in regulating the Th17 response [67]. Several studies demonstrated the involvement of IL-33 in both psoriasis pathogenesis and PsA progression [68]. Consistently, serum levels of IL-33 in patients with PsA are elevated [69]. However, there is limited evidence suggesting a significant involvement of IL-6, which is key cytokine in RA, in the development of PsA [70].

Prior to treatment, patients with PsA exhibit notably higher expression of CXCL12 compared to that in patients with psoriasis vulgaris [71]. The significant up-regulation of myeloid cells in PsA has generated increased attention of the researchers towards myeloid-derived pro-inflammatory mediators, including osteopontin and CCL2, found in the synovial fluid of patients with PsA [72]. Elevated expression levels of the *CCL1*, *CCL20*, *CXCL5*, and *CX3CL1* genes were detected in both the synovial fluid and peripheral blood cells of individuals with PsA [73]. The levels of CXCL4 in the synovial fluid of patients with PsA were found to correlate significantly with IL-17 and IL-22 levels. Collectively, these findings provided evidence that CXCL4 enhances the production of pro-inflammatory cytokines, particularly IL-17, by human CD4* T cells, suggesting a potential involvement of CXCL4 in the pathology of PsA [74].

Table 1. Inflammatory cytokines in PsA.

Cytokine	Main source	Function in PsA	
TNF-α	Macrophages, T	Elevation in the levels of cytokines, chemokines, matrix	
	cells, NK cells,	metalloproteinases, and adhesion molecules from various immune cells	
	mast cells	Induction of osteoclasts to promote the degradation of cartilage and bone	
IL-23	Dendritic cells,	Promotion of Th17 cell differentiation and GM-CSF production	
	macrophages		
IL-12	Dendritic cells,	Promotion of Th1 cell differentiation through STAT4	
	macrophages		
IL-17A/F	Th17 cells, NK	Stimulation of fibroblast-like synoviocytes, chondrocytes, and osteoclasts	
	cells, type 3 innate	Elevation in the levels of proinflammatory cytokines and matrix	
	lymphoid cells	metalloproteinases	
		Induction of neutrophil recruitment	
IL-21	T cells, NK cells	Promotion of T cell differentiation (Specifically, into Th17 cells)	
IL-22	T cells, innate	Activation of fibroblast-like synoviocytes;	
	lymphoid cells	Promotion of osteoclast and bone degradation	
IL-32	NK cells, T cells,	Potentiation of inflammation through the activation of NF-kB signaling	
	monocytes,	and promotion of osteoclast differentiation	
	epithelial cells		

IL-33	Macrophages,	Activation of Th1/Th17-mediated inflammation	
	dendritic cells,		
	mast cells,		
	epithelial cells		
IFN γ	Th1 cells, type 1	Activation of macrophage and T-cell	
	innate lymphoid	Promotion of RANKL secretion	
	cells, NK cells		
GM-CSF	Macrophages, T	Recruitment of various immune cells	
	cells, synovial		
	fibroblasts		
IL-9	Th9 cells	Promotion of Th17-associated inflammation and IL-17A production	
IL-6	Macrophages, T	Stimulation of STAT3 signaling to increase the production of	
	cells, endothelial	proinflammatory cytokines	
	cells		
IL-15	Macrophages	Promotion of T cell proliferation, natural killer cell activation, and	
		production of IFN-γ, TNF-α, and IL-17	
IL-1α	Macrophages,	Induction of IL-17, IL-21 and IL-22 expression by γδT cells in combination	
	dendritic cells	with IL-23	

GM-CSF; Granulocyte-macrophage colony-stimulating factor; IFN; interferon; IL; Interleukin; RANKL; Receptor activator of nuclear factors κB ligand, STAT; Signal transducer and activator of transcription; TNF; Tumor necrosis factor.

4. Cytokine-targeted therapies in PsA

In recent years, the development of innovative drugs that specifically inhibit cytokines and intracellular signaling pathways has led to new treatment paradigms for PsA (Table 2). The heterogenous features of PsA pose a challenge for rheumatologists trying to select the optimal treatment for individual. In order to address these challenges, global organizations including the Group for Research and Assessment of Psoriasis and Psoriatic Arthritis (GRAPPA) and the European League Against Rheumatism (EULAR) have released guidelines to provide guidance for healthcare providers [75,76]. These guidelines prioritize the initial use of conventional synthetic disease-modifying antirheumatic drugs (csDMARDs) with subsequent options including therapy with biological disease-modifying antirheumatic drugs (bDMARDs) or targeted synthetic DMARDs (tsDMARDs), such as JAK inhibitors (JAKi) and PDE4 inhibitors.

For a considerable period, TNF inhibitors (TNFi) have been recognized as the most efficacious treatment for managing the various clinical manifestations of PsA. However, a significant percentage of patients did not achieve a satisfactory response to TNFi. The wide range of novel therapies derived from understanding of the IL-23/IL-17 axis in PsA pathogenesis has provided additional treatment choices for PsA patients. However, on a population scale, these therapies have not demonstrated superior overall outcomes compared to the effects of TNFi. As a next therapeutic approach, JAKi may effectively suppress the pro-inflammatory actions of various cytokines and growth factors.

4.1. TNF-α inhibitors in PsA

TNFi, a class of pharmaceuticals that antagonize TNF- α activity, have shown efficacy in the management of PsA. These medications, including etanercept, certolizumab pegol, infliximab, adalimumab, and golimumab, decrease joint inflammation, improve physical function, and prevent joint damage in patients with PsA [77]. TNFi has shown significant efficacy in treating all SpA manifestations and is recommended by the GRAPPA and EULAR for the management of enthesitis and dactylitis [76]. Phase III trials showed TNFi efficacy in PsA with acceptable safety profiles. Significant improvements according to the American College of Rheumatology criteria (ACR20 response) were observed for all TNFi compared to the effects of placebo in PsA. Moreover, all five TNFi inhibited radiographic progression [78].

Among TNFi, etanercept was less effective than TNFi representing monoclonal antibodies in the treatment of uveitis or inflammatory bowel disease, which are extra-musculoskeletal manifestations of PsA [79,80]. On the other hand, etanercept exhibits lower immunogenicity compared to other TNFi and does not necessitate concurrent methotrexate administration to maintain its long-term efficacy. In a randomized, controlled phase III trial, both etanercept monotherapy and combination therapy with etanercept and methotrexate had superior efficacy compared to that achieved with methotrexate monotherapy in patients with PsA. Furthermore, the addition of methotrexate did not synergistically enhance the efficacy of etanercept [81].

Nevertheless, there can be adverse effects associated with TNFi, including higher susceptibility to infections, so they necessitate careful supervision by a healthcare professional. The TNFi drugs do not prevent the development of new bone in patients with PsA. In fact, these results suggest that cytokines other than TNF- α may play a crucial role in the abnormal bone growth in PsA [82].

4.2. IL-17 inhibitors in psoriatic arthritis

The prominent involvement of IL-17 in the immune-related mechanism of PsA validates the use of IL-17 inhibitors for all forms of psoriatic disease. Secukinumab became the first IL-17 inhibitor to receive approval for the treatment of PsA. After that, ixekizumab was approved, and bimekizumab and brodalumab are also being studied for their potential efficacy in PsA.

Secukinumab is a human IgG1k class monoclonal antibody that specifically targets IL-17A. The FUTURE trials have shown its superior efficacy over placebo in multiple domains of PsA, including spondylitis, peripheral arthritis, enthesitis, dactylitis, psoriasis, and nail disease [83,84]. Moreover, secukinumab reduced radiographic progression compared to the effect of placebo. In a head-to-head clinical trial comparing secukinumab and adalimumab, the former did not show superiority in musculoskeletal domain outcomes in patients with PsA. However, therapy with secukinumab was associated with better skin outcomes and a higher treatment retention rate [85].

Ixekizumab is a IgG4 κ subclass monoclonal antibody, produced using recombinant technology, which specifically targets IL-17A. In both TNFi refractory and naïve patients, ixekizumab exhibited superiority over placebo in terms of ACR responses, inhibition of radiographic progression, and various clinical aspects, similar to the effects observed with secukinumab [86,87]. Ixekizumab was superior to adalimumab in achieving complete remission of psoriasis, as indicated by the psoriasis area and severity index (PASI)100 response at week 24 in a head-to-head clinical trial. However, there were no significant differences in the clinical effect on musculoskeletal domain manifestations between ixekizumab and adalimumab [88].

Bimekizumab, a humanized IgG1κ monoclonal antibody targeting IL-17A and IL-17F, demonstrated notable improvements in the ACR50 response compared to placebo in PsA patients treated for 12 weeks, as evidenced by data from the randomized controlled trial (BE ACTIVE). Moreover, sustained improvements in pain and fatigue were observed over a period of 3 years [89,90]. In a recent randomized, double-blind, placebo-controlled, phase 3 trial (BE COMPLETE), treatment with bimekizumab resulted in more significant enhancements in joint and skin efficacy measures at week 16 compared to the effects of placebo in patients with PsA who had previously experienced inadequate response or intolerance to TNFi [91].

Brodalumab, a human IgG2 antibody, specifically binds to IL-17 receptor A, leading to the inhibition of IL-17A, IL-17F, and IL-17E. In multicenter phase III clinical trials (AMVISION-1 and 2), brodalumab demonstrated a superior ACR response compared to the placebo group at week 24. Additionally, significant improvements in dactylitis, enthesitis, and psoriasis resolution rates were observed in the brodalumab treatment group [92].

IL-17 inhibitors are recommended for the management of most manifestations in PsA. A recent meta-analysis showed similar effectiveness of anti-IL-17A agents and TNFi in the treatment of arthritis, with superior cutaneous responses to anti-IL-17A agents [93]. However, patients with PsA and inflammatory bowel disease (IBD) are an exception, as IL-17 inhibitors in this group are not recommended owing to the lack of demonstrated efficacy and potential exacerbation of IBD

manifestations [94]. The production of IL-17 in the gut epithelium occurs independently of IL-23 stimulation and is involved in tissue repair processes and local tissue homeostasis. Therefore, blocking IL-17 may paradoxically trigger or worsen IBD symptoms, making the use of IL-17 inhibitors contraindicated in patients with IBD [95]. Furthermore, in patients with recurrent uveitis, monoclonal antibodies targeting TNF- α are generally preferred over IL-17 inhibitors, as the evidence of the efficacy of the latter treatment is limited [95].

4.3. IL-23 inhibitors in PsA

The first IL-23 inhibitor introduced to the market was ustekinumab, which reduces the activities of both IL-23 and IL-12 by specifically targeting their shared p40 subunit. Furthermore, monoclonal antibodies guselkumab, risankizumab, and tildrakizumab specifically target the p19 subunit of IL-23 [96].

Ustekinumab effectively inhibits the differentiation of Th1 and Th17 cells. Phase 3 clinical trials (PSUMMIT trials 1 and 2) showed that this drug effectively treats skin manifestations, peripheral arthritis, enthesitis, and dactylitis in both TNFi-experienced and TNFi-naïve patients with PsA [15]. The use of ustekinumab resulted in a higher rate of achieving the primary outcome, defined as an ACR20 response at week 24, compared to the placebo group [97]. Positive responses to the ustekinumab treatment were sustained over a period of 2 years, and there was a significant reduction in radiographic progression in patients with active PsA [98,99]. In the open-label randomized controlled trial ECLIPSA, patients with PsA showed better improvement of enthesitis at week 24 after ustekinumab than after TNFi [100]. Furthermore, the results of an observational prospective realworld study indicated that TNFi and ustekinumab yielded comparable responses in terms of achieving low disease activity or minimal disease activity after 6 months. Additionally, the study revealed similar safety outcomes for both treatments [101]. In the management of psoriasis, ustekinumab was more effective than etanercept, a soluble TNF receptor inhibitor. Nevertheless, subsequent studies have suggested that targeting IL-23 through p19 inhibition had superior effectiveness in comparison to the effects of p40 inhibition for psoriasis treatment. These findings imply that IL-12 may play a protective role in psoriasis. Due to the favorable outcomes observed with p19 inhibition in psoriasis, it has become the preferred approach for inhibiting IL-23 in dermatology practice. This shift in preference is a result of the significant efficacy of p19 inhibition in treating psoriasis [96,102].

Guselkumab, a monoclonal antibody that specifically binds to the p19 subunit and blocks the activity of IL-23, has obtained approval for the treatment of PsA. In clinical trials (DISCOVER 1&2), ACR20 response to guselkumab was significantly higher than to placebo at week 24 both in TNFi naïve and TNFi-treated PsA patients [103–105]. The safety and clinical improvements were consistently maintained over 2 years [106]. Guselkumab rapidly decreased the levels of type 17 effector cytokines as early as on week 4 after the start of the treatment. After 24 weeks of the treatment with guselkumab, the cytokine levels reached the levels similar to those found in healthy individuals without PsA, indicating normalization of the IL-23/IL-17 effector cytokines. Notably, the decreases in IL-17A and IL-17F levels were more pronounced in patients treated with guselkumab than in those who received ustekinumab [107].

Risankizumab, a monoclonal antibody targeting IL-23, has received the U.S Food and Drug Administration (FDA) approval for the treatment of PsA. The clinical trials conducted on both bDMARD-naïve and bDMARD-experienced patients with PsA showed that significantly more patients achieved an ACR20 response at week 24 after the treatment with risankizumab compared to the effects of placebo [108,109]. Tildrakizumab, another monoclonal antibody targeting IL-23, has shown promising efficacy against PsA in a phase 2 trial [110]. However, although patients in the tildrakizumab groups demonstrated higher response rates in skin-related outcomes, there were no positive outcomes for enthesitis and dactylitis [110].

Neither of the IL-23 inhibitors has met the primary endpoint to be a considered as a treatment option for axSpA [15]. However, a *post hoc* analysis indicated potential efficacy of guselkumab in

treating axial involvement in PsA [111]. It should be noted that IL-23 inhibitors generally have a good safety profile, but further evidence from long-term extension studies is needed to fully assess their benefits and risks.

4.4. JAKi in PsA

The Janus kinase-signal transducer and activator of transcription (JAK-STAT) pathway plays a crucial role in the activation of the immune system in response to cytokines and growth factors, facilitating the transmission of signals from cell-membrane receptors to the nucleus. This pathway involves four JAK proteins, namely JAK1, JAK2, JAK3, and tyrosine-protein kinase 2 (TYK2). These molecules intricately interact with different members of the signal transducers and activators of transcription (STAT) family, ultimately regulating the transcription of genes downstream of the pathway [112]. These signaling pathways promote the proliferation and activation of T helper cells and stimulate the expression of IL-17 and IL-23 receptors in the skin and joints. This emphasizes their significance in the development of PsA.

Tofacitinib is a small molecule that selectively blocks the activities of JAK1 and JAK3. In two separate phase III trials with a randomized, double-blind design for PsA patients, ACR20 response rates and improvement in the health assessment questionnaire—disability index scores were significantly greater after tofacitinib than after placebo [113,114]. The tofacitinib group showed improved clinical outcomes of peripheral arthritis, dactylitis, enthesitis, and psoriasis compared to the outcomes in the placebo group during the first 6 months of therapy [115]. Furthermore, in the long-term extension study, tofacitinib attenuated radiographic progression in patients with axial involvement [115]. In terms of safety, the ORAL Surveillance study sparked controversy by suggesting that the use of tofacitinib in patients with RA may increase the risk of cancer and cardiovascular diseases [116]. Consequently, both the European Medicines Agency (EMA) and the FDA have warned against the initial use of JAKi in patients with a high risk of cardiovascular event or malignancy.

Upadacitinib, a highly specific inhibitor of JAK1, has recently been approved as the latest drug in its class for the treatment of PsA. The phase 3 clinical trial SELECT-PsA 1 provided evidence of the effectiveness of upadacitinib in achieving a significant ACR20 response when compared to the effects of placebo. Significant improvements in secondary endpoints, such as decreased radiographic progression, attainment of minimal disease activity, resolution of dactylitis, and improvement in enthesitis, were observed in both the 15 and 30 mg upadacitinib treatment groups, surpassing the results seen in the placebo group. Furthermore, upadacitinib at doses of 15 and 30 mg had comparable or higher efficacy than adalimumab. However, the 30 mg dose of upadacitinib was associated with a higher incidence of severe adverse events. Additionally, both the upadacitinib and adalimumab treatment groups had comparable PsA manifestations, such as psoriasis, enthesitis, physical function deficit, fatigue, and lower quality of life [117–119]. Similarly, patients who were non-responsive or intolerant to TNFi (SELECT-PsA2), had significantly greater ACR20 responses to upadacitinib and achieved minimal disease activity compared to that noted in the placebo group [120].

Filgotinib, another selective JAK1 inhibitor, had notable superiority over placebo in the randomized, double-blind, phase 2 trial EQUATOR for PsA. That study revealed that considerably higher proportion of patients that received filgotinib achieved the primary endpoint of ACR20 response at week 16. Additionally, filgotinib administration led to superior improvements in peripheral arthritis, enthesitis, psoriasis, physical functioning, fatigue, and pain [121]. However, preclinical studies have raised concerns regarding potential adverse effects on male patients' reproductive function [122]. Some results from the safety studies will be needed to relieve this concern.

Deucravacitinib is a small molecule that selectively and non-competitively inhibits TYK2. By binding to the regulatory domain of TYK2, it modulates the immune inflammatory response to IL-12, IL-23, and type I IFNs [123]. In a phase II clinical trial involving patients with active PsA, significant improvements in arthritis, enthesitis, dactylitis, and psoriasis domains were observed in

participants receiving deucravacitinib at two different doses (6 mg and 12 mg QD) compared to the effects in patients receiving placebo. A *post hoc* analysis conducted on data from phase 2 trial of deucravacitinib examined the ACR20 response at week 16 in patients with PsA, considering their prior exposure to csDMARDs. These findings revealed that all patients treated with deucravacitinib, regardless of their prior exposure to csDMARDs, demonstrated comparable clinical improvements [124]. Currently, a phase 3 randomized controlled trial of deucravacitinib for PsA is ongoing.

Brepocitinib is an orally administered drug with dual inhibitory activity toward TYK2 and JAK1. New findings from a phase 2 randomized controlled trial indicate that both 30 mg and 60 mg doses of brepocitinib administered once daily were more effective in alleviating PsA signs and symptoms than placebo. Over the course of a 52-week study, brepocitinib had a favorable safety profile and was well tolerated by the participants [125].

Recent advancements in our understanding of the JAK/STAT pathway have enabled the development of two JAK inhibitors (JAKi) that have received approval for the treatment of PsA: tofacitinib and upadacitinib. These JAKi showed both a favorable safety profile and efficacy, leading to rapid improvements in PsA symptoms, quality of life, and attenuation of radiographic progression. Notably, when compared to other b/tsDMARDs, JAKi offer several advantages, namely short half-life, oral route of administration, and better patient compliance [123]. TYK2 inhibitors, in particular, have shown a favorable safety profile, possibly attributed to their highly selective mode of action [124]. It will be of utmost importance in the future to study these treatments in a real-world setting to address the unmet requirements of all individuals with PsA and provide answers to questions that clinical trial data alone may not fully address.

4.5. Other pharmacological treatments

Apremilast, an orally administered small molecule that functions as a PDE4 inhibitor, obtained FDA approval in 2014 for the management of PsA. Inhibition of PDE4 elevates intracellular cyclic AMP levels, which subsequently reduces the synthesis of pro-inflammatory cytokines and enhances the production of anti-inflammatory cytokines such as IL-10 [15]. The efficacy of apremilast in PsA has been shown in the PALACE trials [62,126–128]. The GRAPPA guidelines suggest the use of apremilast for patients presenting with psoriasis, nail involvement, peripheral arthritis, enthesitis, and dactylitis.

Although numerous drugs may effectively treat PsA, some patients fail to respond, lose response over time, or experience drug toxicity, resulting in discontinuation of the medication or the need for combination therapies. Consequently, there remains a significant demand for new alternative treatments [129].

Remtolumab is a dual variable domain immunoglobulin that binds to both TNF- α and IL-17A. It was expected that a drug targeting two cytokines target would be able to treat PsA more effectively. Nonetheless, the ACR20 and PASI90 responses to remtolumab and adalimumab were comparable. As a result, the approach of simultaneously inhibiting both the TNF- α and IL-17A pathways was abandoned, even though treatment with dual variable domain immunoglobulin appeared promising for patients with challenging to treat PsA [130].

Nanobodies, which are antibodies consisting of recombinant variable domain on a heavy chain, could be another promising tool for new drug development. Nanobodies possess several useful characteristics such as small size, good solubility, high stability, easy production, fast elimination from the bloodstream, and effective penetration into deep tissues. These properties make nanobodies highly valued as therapeutic tools, and they are also being explored for use in chimeric antigen receptors and targeted drug delivery systems [131]. A phase 3 clinical trial for netakimab, a recombinant IgG1 nanobody targeting IL-17A, is ongoing in patients with PsA [132]. Sonelokimab is an innovative trivalent nanobody that consists of three distinct parts. The C-terminal segment of this drug attaches to both IL-17A and IL-17F subtypes, while the middle segment binds to serum albumin. The N-terminal segment, on the other hand, selectively targets IL-17F [133]. Although the

effectiveness and safety of these two drugs have been demonstrated in patients with plaque psoriasis, there is currently no published data evaluating their effects specifically on patients with PsA.

Table 2. Current cytokine-targeting treatment for psoriatic arthritis.

b/tsDMARDs	Target	Dose and administration route	FDA or EMA
			approval
Etanercept	TNF-α	50 mg weekly, SC	FDA, EMA
Infliximab	TNF-α	5 mg/kg at weeks 0, 2, and 6; every 8	FDA, EMA
		weeks thereafter, IV	
Adalimumab	TNF-α	40 mg every 2 weeks, SC	FDA, EMA
Golimumab	TNF-α	50 mg weekly, SC	FDA, EMA
Certolizumab	TNF-α	200 mg every 2 weeks or 400 mg	FDA, EMA
pegol		every 4 weeks, SC	
Ustekinumab	p40 subunit of IL-12 and IL-23	(BW<100 kg)	FDA, EMA
		45 mg/kg at weeks 0, 4, and 12; every	
		12 weeks thereafter, SC	
		(BW>100kg)	
		90 mg/kg at weeks 0, 4, and 12; every	
		12 weeks thereafter, SC	
Guselkumab	p19 subunit of IL-23	100 mg at weeks 0 and 4; every 8	FDA, EMA
		weeks thereafter, SC	
Risankizumab	p19 subunit of IL-23	150 mg at weeks 0 and 4; every 12	FDA, EMA
		weeks thereafter, SC	
Tildrakizumab	p19 subunit of IL-23	100 mg at weeks 0 and 4; every 12	-
		weeks thereafter, SC	
Secukinumab	IL-17A	150 mg weekly for 4 weeks; monthly	FDA, EMA
		thereafter, SC	
Ixekizumab	IL-17A	160 mg at weeks 0 and 80 mg at	FDA, EMA
		weeks 2, 4, 6, 8, 10, and 12; 80 mg	
		every 4 weeks thereafter, SC	
Bimekizumab	IL-17A/F	320 mg every 4 weeks for 16 weeks,	EMA
		every 8 weeks thereafter, SC	
Brodalumab	IL-17 receptor	210 mg at weeks 0, 1, and 2; every 2	-
		weeks thereafter, SC	
Tofacitinib	JAK1/3	5 mg twice daily, orally	FDA, EMA
Upadacitinib	JAK1	15 mg once daily, orally	FDA, EMA
Filgotinib	JAK1	200 mg once daily, orally	-
Deucravacitinib	TYK2	6 mg once daily, orally	-
Brepocitinib	JAK1/TYK2	30 mg or 60 mg once daily, orally	-

b/tsDMARDs, biological/targeted synthetic disease-modifying antirheumatic drugs; EMA, the European Medicines Agency; FDA; the U.S Food and Drug Administration; IV, intravenously; SC; subcutaneously.

5. Future perspectives and conclusion

The progress in the elucidation of the impacts of inflammatory cytokines on PsA pathogenesis has enabled exploration of the novel molecular mechanisms that can be precisely targeted by therapeutic approaches. The activation of immunoinflammatory pathways in PsA involves multiple cell types and proinflammatory cytokines, such as TNF- α , IL-17, and IL-23. Providing timely and effective treatment is of utmost importance in preventing the spread of joint damage and improving the overall well-being of individuals diagnosed with PsA. Despite the advancements in

understanding PsA pathogenesis and development of new drugs that target specific cytokines, managing PsA continues to pose challenges for rheumatologists.

The pathogenesis and current therapeutic targets of PsA are shown in figure 1. Our review shows that the ongoing studies investigating PsA pathogenesis offer a potential to reveal more pertinent therapeutic targets and broaden the array of available treatment options. Furthermore, there is a burgeoning interest in utilizing new biotechnology tools and combination therapies aimed at targeting multiple pathogenic pathways to achieve more effective disease control. Although there is little reliable clinical evidence of new drugs surpassing previous b/tsDMARDs in efficacy, significant advancements achieved in recent years in this field offer a promising outlook for the future.

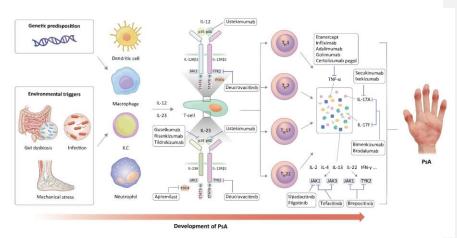


Figure 1. Pathogenesis of PsA and therapeutic targets.

Author Contributions: BWL wrote the original draft and edited and reviewed the final version of the manuscript. SJM edited and reviewed the original draft and the final version of the manuscript. Both authors have read and agreed to the version of the manuscript to be published.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: No new data were created or analyzed in this study. Data sharing is not applicable to this article.

Conflicts of Interest: The authors declare no conflict of interest.

References

- Ritchlin C.T.; Colbert R.A.; Gladman D.D. Psoriatic arthritis. N. Engl. J. Med. 2017, 376, 957-970 [doi:10.1056/NEIMra1505557].
- Schett G.; Rahman P.; Ritchlin C.; McInnes I.B.; Elewaut D.; Scher J.U. Psoriatic arthritis from a mechanistic perspective. Nat. Rev. Rheumatol. 2022, 18, 311-325 [doi:10.1038/s41584-022-00776-6].
- López-Medina C.; Molto A.; Sieper J.; Duruöz T.; Kiltz U.; Elzorkany B.; Hajjaj-Hassouni N.; Burgos-Vargas R.; Maldonado-Cocco J.; Ziade N.; et al. Prevalence and distribution of peripheral musculoskeletal manifestations in spondyloarthritis including psoriatic arthritis: results of the worldwide, cross-sectional ASAS-PerSpA study. RMD Open 2021, 7 [doi:10.1136/rmdopen-2020-001450].
- 4. Qi F.; Tan Y.; Yao A.; Yang X.; He Y. Psoriasis to psoriatic arthritis: the application of proteomics technologies. Front. Med. (Lausanne) 2021, 8, 681172 [doi:10.3389/fmed.2021.681172].
- Veale D.J.; Fearon U. The pathogenesis of psoriatic arthritis. *Lancet* 2018, 391, 2273-2284 [doi:10.1016/S0140-6736(18)30830-4].

- Kane D.; Stafford L.; Bresnihan B.; FitzGerald O. A prospective, clinical and radiological study of early psoriatic arthritis: an early synovitis clinic experience. *Rheumatol. (Oxf. Engl.)* 2003, 42, 1460-1468 [doi:10.1093/rheumatology/keg384].
- Haroon M.; Gallagher P.; FitzGerald O. Diagnostic delay of more than 6 months contributes to poor radiographic and functional outcome in psoriatic arthritis. *Ann. Rheum. Dis.* 2015, 74, 1045-1050 [doi:10.1136/annrheumdis-2013-204858].
- Queiro R.; Morante I.; Cabezas I.; Acasuso B. HLA-B27 and psoriatic disease: a modern view of an old relationship. Rheumatol. (Oxf. Engl.) 2016, 55, 221-229 [doi:10.1093/rheumatology/kev296].
- Muto M.; Date Y.; Ichimiya M.; Moriwaki Y.; Mori K.; Kamikawaji N.; Kimura A.; Sasazuki T.; Asagami C. Significance of antibodies to streptococcal M protein in psoriatic arthritis and their association with HLAa*0207. Tissue Antigens 1996, 48, 645-650 [doi:10.1111/j.1399-0039.1996.tb02687.x].
- Thorarensen S.M.; Lu N.; Ogdie A.; Gelfand J.M.; Choi H.K.; Love T.J. Physical trauma recorded in primary care is associated with the onset of psoriatic arthritis among patients with psoriasis. *Ann. Rheum. Dis.* 2017, 76, 521-525 [doi:10.1136/annrheumdis-2016-209334].
- 11. Scher J.U.; Ubeda C.; Artacho A.; Attur M.; Isaac S.; Reddy S.M.; Marmon S.; Neimann A.; Brusca S.; Patel T.; et al. Decreased bacterial diversity characterizes the altered gut microbiota in patients with psoriatic arthritis, resembling dysbiosis in inflammatory bowel disease. *Arthritis Rheumatol.* 2015, 67, 128-139 [doi:10.1002/art.38892].
- 12. Boix-Amorós A.; Badri M.H.; Manasson J.; Blank R.B.; Haberman R.H.; Neimann A.L.; Girija P.V.; Jimenez Hernandez A.; Heguy A.; Koralov S.B.; et al. Alterations in the cutaneous microbiome of patients with psoriasis and psoriatic arthritis reveal similarities between non-lesional and lesional skin. *Ann. Rheum. Dis.* **2022** [doi:10.1136/ard-2022-223389].
- 13. Jadon D.R.; Stober C.; Pennington S.R.; FitzGerald O. Applying precision medicine to unmet clinical needs in psoriatic disease. *Nat. Rev. Rheumatol.* **2020**, *16*, 609-627 [doi:10.1038/s41584-020-00507-9].
- 14. McGonagle D. Enthesitis: an autoinflammatory lesion linking nail and joint involvement in psoriatic disease. *J. Eur. Acad. Dermatol. Venereol.* **2009**, 23 suppl 1, 9-13 [doi:10.1111/j.1468-3083.2009.03363.x].
- Azuaga A.B.; Ramírez J.; Cañete J.D. Psoriatic arthritis: pathogenesis and targeted therapies. Int. J. Mol. Sci. 2023, 24, 4901.
- Fitzgerald O.; Winchester R. Psoriatic arthritis: from pathogenesis to therapy. Arthritis Res. Ther. 2009, 11, 214 [doi:10.1186/ar2580].
- Candia L.; Marquez J.; Hernandez C.; Zea A.H.; Espinoza L.R. Toll-like receptor-2 expression is upregulated in antigen-presenting cells from patients with psoriatic arthritis: a pathogenic role for innate immunity? *J. Rheumatol.* 2007, 34, 374-379.
- 18. Jongbloed S.L.; Lebre M.C.; Fraser A.R.; Gracie J.A.; Sturrock R.D.; Tak P.P.; McInnes I.B. Enumeration and phenotypical analysis of distinct dendritic cell subsets in psoriatic arthritis and rheumatoid arthritis. *Arthritis Res. Ther.* 2006, 8, R15 [doi:10.1186/ar1864].
- Lew W.; Bowcock A.M.; Krueger J.G. Psoriasis vulgaris: cutaneous lymphoid tissue supports T-cell activation and "Type 1" inflammatory gene expression. Trends Immunol. 2004, 25, 295-305 [doi:10.1016/j.it.2004.03.006].
- 20. van Mens L.J.J.; van de Sande M.G.H.; Menegatti S.; Chen S.; Blijdorp I.C.J.; de Jong H.M.; Fluri I.A.; Latuhihin T.E.; van Kuijk A.W.R.; Rogge L.; et al. Brief report: interleukin-17 blockade with secukinumab in peripheral spondyloarthritis impacts synovial immunopathology without compromising systemic immune responses. Arthritis Rheumatol. 2018, 70, 1994-2002 [doi:10.1002/art.40581].
- Van Raemdonck K.; Umar S.; Palasiewicz K.; Romay B.; Volkov S.; Arami S.; Sweiss N.; Shahrara S. TLR7
 endogenous ligands remodel glycolytic macrophages and trigger skin-to-joint crosstalk in psoriatic
 arthritis. Eur. J. Immunol. 2021, 51, 714-720 [doi:10.1002/eji.202048690].
- Lu C.H.; Lai C.Y.; Yeh D.W.; Liu Y.L.; Su Y.W.; Hsu L.C.; Chang C.H.; Catherine Jin S.L.; Chuang T.H. Involvement of M1 macrophage polarization in endosomal toll-like receptors activated psoriatic inflammation. *Mediators Inflamm.* 2018, 2018, 3523642 [doi:10.1155/2018/3523642].
- 23. Vandooren B.; Noordenbos T.; Ambarus C.; Krausz S.; Cantaert T.; Yeremenko N.; Boumans M.; Lutter R.; Tak P.P.; Baeten D. Absence of a classically activated macrophage cytokine signature in peripheral spondylarthritis, including psoriatic arthritis. Arthritis Rheum. 2009, 60, 966-975 [doi:10.1002/art.24406].
- Nigrovic P.A.; Lee D.M. Mast cells in inflammatory arthritis. Arthritis Res. Ther. 2005, 7, 1–11 [doi:10.1186/ar1446].
- Noordenbos T.; Yeremenko N.; Gofita I.; van de Sande M.; Tak P.P.; Caňete J.D.; Baeten D. Interleukin-17positive mast cells contribute to synovial inflammation in spondylarthritis. *Arthritis Rheum.* 2012, 64, 99109 [doi:10.1002/art.33396].
- 26. Chen S.; Noordenbos T.; Blijdorp I.; van Mens L.; Ambarus C.A.; Vogels E.; Te Velde A.; Alsina M.; Cañete J.D.; Yeremenko N.; et al. Histologic evidence that mast cells contribute to local tissue inflammation in peripheral spondyloarthritis by regulating interleukin-17A content. *Rheumatol. (Oxf. Engl.)* 2019, 58, 617-627 [doi:10.1093/rheumatology/key331].

- 27. Leijten E.F.; van Kempen T.S.; Boes M.; Michels-van Amelsfort J.M.; Hijnen D.; Hartgring S.A.; van Roon J.A.; Wenink M.H.; Radstake T.R. Brief report: enrichment of activated group 3 innate lymphoid cells in psoriatic arthritis synovial fluid. *Arthritis Rheumatol.* 2015, 67, 2673-2678 [doi:10.1002/art.39261].
- Soare A.; Weber S.; Maul L.; Rauber S.; Gheorghiu A.M.; Luber M.; Houssni I.; Kleyer A.; von Pickardt G.; Gado M.; et al. Cutting edge: homeostasis of innate lymphoid cells is imbalanced in psoriatic arthritis. *J. Immunol.* 2018, 200, 1249-1254 [doi:10.4049/jimmunol.1700596].
- Sakkas L.I.; Bogdanos D.P. Are psoriasis and psoriatic arthritis the same disease? The IL-23/IL-17 axis data. Autoimmun. Rev. 2017, 16, 10-15 [doi:10.1016/j.autrev.2016.09.015].
- 30. Tang F.; Sally B.; Ciszewski C.; Abadie V.; Curran S.A.; Groh V.; Fitzgerald O.; Winchester R.J.; Jabri B. Interleukin 15 primes natural killer cells to kill via NKG2D and cPLA2 and this pathway is active in psoriatic arthritis. *PLOS ONE* **2013**, *8*, e76292 [doi:10.1371/journal.pone.0076292].
- 31. Qu N.; Xu M.; Mizoguchi I.; Furusawa J.; Kaneko K.; Watanabe K.; Mizuguchi J.; Itoh M.; Kawakami Y.; Yoshimoto T. Pivotal roles of T-helper 17-related cytokines, IL-17, IL-22, and IL-23, in inflammatory diseases. Clin. Dev. Immunol. 2013, 2013, 968549 [doi:10.1155/2013/968549].
- 32. Lories R.J.; McInnes I.B. Primed for inflammation: enthesis-resident T cells. Nat. Med. 2012, 18, 1018-1019 [doi:10.1038/nm.2854].
- 33. Veale D.J.; Ritchlin C.; FitzGerald O. Immunopathology of psoriasis and psoriatic arthritis. *Ann. Rheum. Dis.* **2005**, *64* suppl 2, ii26–ii29 [doi:10.1136/ard.2004.031740].
- Diani M.; Altomare G.; Reali E. T cell responses in psoriasis and psoriatic arthritis. Autoimmun. Rev. 2015, 14, 286-292 [doi:10.1016/j.autrev.2014.11.012].
- 35. Kirkham B.W.; Kavanaugh A.; Reich K. Interleukin-17A: a unique pathway in immune-mediated diseases: psoriasis, psoriatic arthritis and rheumatoid arthritis. *Immunology* **2014**, *141*, 133-142 [doi:10.1111/imm.12142].
- 36. Menon B.; Gullick N.J.; Walter G.J.; Rajasekhar M.; Garrood T.; Evans H.G.; Taams L.S.; Kirkham B.W. Interleukin-17+CD8+T cells are enriched in the joints of patients with psoriatic arthritis and correlate with disease activity and joint damage progression. *Arthritis Rheumatol.* **2014**, *66*, 1272-1281 [doi:10.1002/art 38376]
- Penkava F.; Velasco-Herrera M.D.C.; Young M.D.; Yager N.; Nwosu L.N.; Pratt A.G.; Lara A.L.; Guzzo C.; Maroof A.; Mamanova L.; et al. Single-cell sequencing reveals clonal expansions of pro-inflammatory synovial CD8 T cells expressing tissue-homing receptors in psoriatic arthritis. *Nat. Commun.* 2020, 11, 4767 [doi:10.1038/s41467-020-18513-6].
- Kawashiri S.Y.; Kawakami A.; Okada A.; Koga T.; Tamai M.; Yamasaki S.; Nakamura H.; Origuchi T.; Ida H.; Eguchi K. CD4+CD25(high)CD127(low/-) Treg cell frequency from peripheral blood correlates with disease activity in patients with rheumatoid arthritis. J. Rheumatol. 2011, 38, 2517-2521 [doi:10.3899/jrheum.110283].
- 39. Yun W.J.; Lee D.W.; Chang S.E.; Yoon G.S.; Huh J.R.; Won C.H.; Lee M.W.; Kim S.E.; Kim B.J.; Moon K.C.; et al. Role of CD4CD25FOXP3 regulatory T cells in psoriasis. *Ann. Dermatol.* **2010**, 22, 397-403 [doi:10.5021/ad.2010.22.4.397].
- Sugiyama H.; Gyulai R.; Toichi E.; Garaczi E.; Shimada S.; Stevens S.R.; McCormick T.S.; Cooper K.D. Dysfunctional blood and target tissue CD4+CD25high regulatory T cells in psoriasis: mechanism underlying unrestrained pathogenic effector T cell proliferation. *J. Immunol.* 2005, 174, 164-173 [doi:10.4049/jimmunol.174.1.164].
- 41. Schett G.; McInnes I.B.; Neurath M.F. Reframing immune-mediated inflammatory diseases through signature cytokine hubs. N. Engl. J. Med. 2021, 385, 628-639 [doi:10.1056/NEJMra1909094].
- Saddala M.S.; Huang H. Identification of novel inhibitors for TNFα, TNFR1 and TNFα-TNFR1 complex using pharmacophore-based approaches. J. Transl. Med. 2019, 17, 215 [doi:10.1186/s12967-019-1965-5].
- 43. Zamri F.; de Vries T.J. Use of TNF inhibitors in rheumatoid arthritis and implications for the periodontal status: for the benefit of both? *Front. Immunol.* **2020**, *11*, 591365 [doi:10.3389/fimmu.2020.591365].
- 44. de Vries T.J.; El Bakkali I.; Kamradt T.; Schett G.; Jansen I.D.C.; D'Amelio P. What are the peripheral blood determinants for increased osteoclast formation in the various inflammatory diseases associated with bone loss? Front. Immunol. 2019, 10, 505 [doi:10.3389/fimmu.2019.00505].
- Kaaij M.H.; van Tok M.N.; Blijdorp I.C.; Ambarus C.A.; Stock M.; Pots D.; Knaup V.L.; Armaka M.; Christodoulou-Vafeiadou E.; van Melsen T.K.; et al. Transmembrane TNF drives osteoproliferative joint inflammation reminiscent of human spondyloarthritis. J. Exp. Med. 2020, 217 [doi:10.1084/jem.20200288].
- 46. Silvagni E.; Missiroli S.; Perrone M.; Patergnani S.; Boncompagni C.; Bortoluzzi A.; Govoni M.; Giorgi C.; Alivernini S.; Pinton P.; et al. From bed to bench and back: TNF-α, IL-23/IL-17A, and JAK-dependent inflammation in the pathogenesis of psoriatic synovitis. Front. Pharmacol. 2021, 12, 672515 [doi:10.3389/fphar.2021.672515].
- Ritchlin C.T.; Haas-Smith S.A.; Li P.; Hicks D.G.; Schwarz E.M. Mechanisms of TNF-alpha- and RANKL-mediated osteoclastogenesis and bone resorption in psoriatic arthritis. *J. Clin. Invest.* 2003, 111, 821-831 [doi:10.1172/JCI16069].

- 48. Blauvelt A.; Chiricozzi A. The immunologic role of IL-17 in psoriasis and psoriatic arthritis pathogenesis. Clin. Rev. Allergy Immunol. 2018, 55, 379-390 [doi:10.1007/s12016-018-8702-3].
- Rizzo H.L.; Kagami S.; Phillips K.G.; Kurtz S.E.; Jacques S.L.; Blauvelt A. IL-23-mediated psoriasis-like epidermal hyperplasia is dependent on IL-17A. J. Immunol. 2011, 186, 1495-1502 [doi:10.4049/jimmunol.1001001].
- Łukasik Z.; Gracey E.; Venken K.; Ritchlin C.; Elewaut D. Crossing the boundaries: il-23 and its role in linking inflammation of the skin, gut and joints. *Rheumatol. (Oxf. Engl.)* 2021, 60 Supplement 4, iv16-iv27 [doi:10.1093/rheumatology/keab385].
- 51. Sherlock J.P.; Cua D.J. Interleukin-23 in perspective. *Rheumatol. (Oxf. Engl.)* **2021**, *60* Supplement 4, iv1-iv3 [doi:10.1093/rheumatology/keab461].
- Chen L.; Deshpande M.; Grisotto M.; Smaldini P.; Garcia R.; He Z.; Gulko P.S.; Lira S.A.; Furtado G.C. Skin expression of IL-23 drives the development of psoriasis and psoriatic arthritis in mice. Sci. Rep. 2020, 10, 8259 [doi:10.1038/s41598-020-65269-6].
- Raychaudhuri S.K.; Abria C.; Mitra A.; Raychaudhuri S.P. Functional significance of MAIT cells in psoriatic arthritis. Cytokine 2020, 125, 154855 [doi:10.1016/j.cyto.2019.154855].
- 54. Baeten D.; Østergaard M.; Wei J.C.; Sieper J.; Järvinen P.; Tam L.S.; Salvarani C.; Kim T.H.; Solinger A.; Datsenko Y.; et al. Risankizumab, an IL-23 inhibitor, for ankylosing spondylitis: results of a randomised, double-blind, placebo-controlled, proof-of-concept, dose-finding phase 2 study. *Ann. Rheum. Dis.* 2018, 77, 1295-1302 [doi:10.1136/annrheumdis-2018-213328].
- Huynh D.; Kavanaugh A. Psoriatic arthritis: current therapy and future approaches. *Rheumatol. (Oxf. Engl.)* 2015, 54, 20-28 [doi:10.1093/rheumatology/keu237].
- Maeda S.; Hayami Y.; Naniwa T.; Ueda R. The Th17/IL-23 axis and natural immunity in psoriatic arthritis. Int. J. Rheumatol. 2012, 2012, 539683 [doi:10.1155/2012/539683].
- Tam H.K.J.; Robinson P.C.; Nash P. Inhibiting IL-17A and IL-17F in rheumatic disease: therapeutics help to elucidate disease mechanisms. Curr. Rheumatol. Rep. 2022, 24, 310-320 [doi:10.1007/s11926-022-01084-4].
- 58. Iznardo H.; Puig L. Dual inhibition of IL-17A and IL-17F in psoriatic disease. *Ther. Adv. Chronic Dis.* **2021**, *12*, 20406223211037846 [doi:10.1177/20406223211037846].
- 59. Millar N.L.; Akbar M.; Campbell A.L.; Reilly J.H.; Kerr S.C.; McLean M.; Frleta-Gilchrist M.; Fazzi U.G.; Leach W.J.; Rooney B.P.; et al. IL-17A mediates inflammatory and tissue remodelling events in early human tendinopathy. *Sci. Rep.* **2016**, *6*, 27149 [doi:10.1038/srep27149].
- McGonagle D.G.; McInnes I.B.; Kirkham B.W.; Sherlock J.; Moots R. The role of IL-17A in axial spondyloarthritis and psoriatic arthritis: recent advances and controversies. *Ann. Rheum. Dis.* 2019, 78, 1167-1178 [doi:10.1136/annrheumdis-2019-215356].
- 61. Gravallese E.M.; Schett G. Effects of the IL-23–IL-17 pathway on bone in spondyloarthritis. *Nat. Rev. Rheumatol.* **2018**, *14*, 631-640 [doi:10.1038/s41584-018-0091-8].
- 62. Kavanaugh A.; Mease P.J.; Gomez-Reino J.J.; Adebajo A.O.; Wollenhaupt J.; Gladman D.D.; Lespessailles E.; Hall S.; Hochfeld M.; Hu C.; et al. Treatment of psoriatic arthritis in a phase 3 randomised, placebocontrolled trial with apremilast, an oral phosphodiesterase 4 inhibitor. *Ann. Rheum. Dis.* **2014**, 73, 1020-1026 [doi:10.1136/annrheumdis-2013-205056].
- 63. McInnes I.B.; Anderson J.K.; Magrey M.; Merola J.F.; Liu Y.; Kishimoto M.; Jeka S.; Pacheco-Tena C.; Wang X.; Chen L.; et al. Trial of upadacitinib and adalimumab for psoriatic arthritis. *N. Engl. J. Med.* **2021**, 384, 1227-1239 [doi:10.1056/NEJMoa2022516].
- 64. Miyagawa I.; Nakayamada S.; Ueno M.; Miyazaki Y.; Iwata S.; Kubo S.; Sonomoto K.; Anan J.; Ohkubo N.; Inoue Y.; et al. Impact of serum interleukin-22 as a biomarker for the differential use of molecular targeted drugs in psoriatic arthritis: a retrospective study. *Arthritis Res. Ther.* 2022, 24, 86 [doi:10.1186/s13075-022-02771-4].
- Kwon O.C.; Park M.C.; Kim Y.G. Interleukin-32 as a biomarker in rheumatic diseases: A narrative review. Front. Immunol. 2023, 14, 1140373 [doi:10.3389/fimmu.2023.1140373].
- Al-Shobaili H.A.; Farhan J.; Zafar U.; Rasheed Z. Functional role of human interleukin-32 and nuclear transcription factor-kB in patients with psoriasis and psoriatic arthritis. Int. J. Health Sci. (Qassim) 2018, 12, 29-34.
- Dong Y.; Zhong J.; Dong L. IL-33 in rheumatic diseases. Front. Med. (Lausanne) 2021, 8, 739489 [doi:10.3389/fmed.2021.739489].
- Cannavò S.P.; Bertino L.; Di Salvo E.; Papaianni V.; Ventura-Spagnolo E.; Gangemi S. Possible roles of IL-33 in the innate-adaptive immune crosstalk of psoriasis pathogenesis. Mediators Inflamm. 2019, 2019, 7158014 [doi:10.1155/2019/7158014].
- 69. Li J.; Liu L.; Rui W.; Li X.; Xuan D.; Zheng S.; Yu Y.; Zhang J.; Kong N.; Zhu X.; et al. New interleukins in psoriasis and psoriatic arthritis patients: the possible roles of interleukin-33 to Interleukin-38 in disease activities and bone erosions. *Dermatology* **2017**, 233, 37-46 [doi:10.1159/000471798].
- 70. Sokolova M.V.; Simon D.; Nas K.; Zaiss M.M.; Luo Y.; Zhao Y.; Rech J.; Schett G. A set of serum markers detecting systemic inflammation in psoriatic skin, entheseal, and joint disease in the absence of C-reactive

Commented [오전1]: The journal title differs from PubMed. Suggestion: 'Mediators Inflamm.'

Al translation:

저널 제목이 PubMed 와 다릅니다. 제안: 'Mediators

Inflamm.

- protein and its link to clinical disease manifestations. Arthritis Res. Ther. 2020, 22, 26 [doi:10.1186/s13075-020-2111-8].
- Abdelaal N.H.; Elhefnawy N.G.; Abdulmonem S.R.; Sayed S.; Saleh N.A.; Saleh M.A. Evaluation of the expression of the stromal cell-derived factor-1 alpha (CXCL 12) in psoriatic patients after treatment with methotrexate. *J. Cosmet. Dermatol.* 2020, 19, 253-258 [doi:10.1111/jocd.12994].
- Yager N.; Cole S.; Lledo Lara A.; Maroof A.; Penkava F.; Knight J.C.; Bowness P.; Al-Mossawi H. Ex vivo mass cytometry analysis reveals a profound myeloid proinflammatory signature in psoriatic arthritis synovial fluid. *Ann. Rheum. Dis.* 2021, 80, 1559-1567 [doi:10.1136/annrheumdis-2021-220280].
- 73. Abji F.; Pollock R.A.; Liang K.; Chandran V.; Gladman D.D. Th17 gene expression in psoriatic arthritis synovial fluid and peripheral blood compared to osteoarthritis and cutaneous psoriasis. *Clin. Exp. Rheumatol.* **2018**, 36, 486-489.
- 74. Affandi A.J.; Silva-Cardoso S.C.; Garcia S.; Leijten E.F.A.; van Kempen T.S.; Marut W.; van Roon J.A.G.; Radstake T.R.D.J. CXCL4 is a novel inducer of human Th17 cells and correlates with IL-17 and IL-22 in psoriatic arthritis. *Eur. J. Immunol.* 2018, 48, 522-531 [doi:10.1002/eji.201747195].
- 75. Gossec L.; Baraliakos X.; Kerschbaumer A.; de Wit M.d.; McInnes I.; Dougados M.; Primdahl J.; McGonagle D.G.; Aletaha D.; Balanescu A.; et al. EULAR recommendations for the management of psoriatic arthritis with pharmacological therapies: 2019 update. *Ann. Rheum. Dis.* 2020, 79, 700-712 [doi:10.1136/annrheumdis-2020-217159].
- Coates L.C.; Soriano E.R.; Corp N.; Bertheussen H.; Callis Duffin K.; Campanholo C.B.; Chau J.; Eder L.; Fernández-Ávila D.G.; FitzGerald O.; et al. Group for Research and Assessment of Psoriasis and Psoriatic Arthritis (GRAPPA): updated treatment recommendations for psoriatic arthritis 2021. *Nat. Rev. Rheumatol.* 2022, 18, 465-479 [doi:10.1038/s41584-022-00798-0].
- 77. Sundanum S.; Orr C.; Veale D. Targeted therapies in psoriatic arthritis-an update. *Int. J. Mol. Sci.* 2023, 24 [doi:10.3390/ijms24076384].
- 78. Mantravadi S.; Ogdie A.; Kraft W.K. Tumor necrosis factor inhibitors in psoriatic arthritis. Expert Rev. Clin. Pharmacol. 2017, 10, 899-910 [doi:10.1080/17512433.2017.1329009].
- 79. Wendling D.; Joshi A.; Reilly P.; Jalundhwala Y.J.; Mittal M.; Bao Y. Comparing the risk of developing uveitis in patients initiating anti-tumor necrosis factor therapy for ankylosing spondylitis: an analysis of a large US claims database. Curr. Med. Res. Opin. 2014, 30, 2515-2521 [doi:10.1185/03007995.2014.969368].
- 80. Levin A.D.; Wildenberg M.E.; van den Brink G.R. Mechanism of action of anti-TNF therapy in inflammatory bowel disease. *J. Crohns Colitis* **2016**, *10*, 989-997 [doi:10.1093/ecco-jcc/jjw053].
- 81. Mease P.J.; Gladman D.D.; Collier D.H.; Ritchlin C.T.; Helliwell P.S.; Liu L.; Kricorian G.; Chung J.B. Etanercept and methotrexate as monotherapy or in combination for psoriatic arthritis: primary results from a randomized, controlled Phase III trial. *Arthritis Rheumatol.* **2019**, *71*, 1112-1124 [doi:10.1002/art.40851].
- 82. Lories R.J.; Schett G. Pathophysiology of new bone formation and ankylosis in spondyloarthritis. *Rheum. Dis. Clin. North Am.* **2012**, *38*, 555-567 [doi:10.1016/j.rdc.2012.08.003].
- 83. McInnes I.B.; Mease P.J.; Kirkham B.; Kavanaugh A.; Ritchlin C.T.; Rahman P.; van der Heijde D.; Landewé R.; Conaghan P.G.; Gottlieb A.B.; et al. Secukinumab, a human anti-interleukin-17A monoclonal antibody, in patients with psoriatic arthritis (FUTURE 2): a randomised, double-blind, placebo-controlled, phase 3 trial. *Lancet* 2015, 386, 1137-1146 [doi:https://doi.org/10.1016/S0140-6736(15)61134-5].
- 84. Nash P.; Mease P.J.; McInnes I.B.; Rahman P.; Ritchlin C.T.; Blanco R.; Dokoupilova E.; Andersson M.; Kajekar R.; Mpofu S.; et al. Efficacy and safety of secukinumab administration by autoinjector in patients with psoriatic arthritis: results from a randomized, placebo-controlled trial (FUTURE 3). *Arthritis Res. Ther.* **2018**, 20. 47 [doi:10.1186/s13075-018-1551-x].
- 85. McInnes I.B.; Behrens F.; Mease P.J.; Kavanaugh A.; Ritchlin C.; Nash P.; Masmitja J.G.; Goupille P.; Korotaeva T.; Gottlieb A.B.; et al. Secukinumab versus adalimumab for treatment of active psoriatic arthritis (EXCEED): a double-blind, parallel-group, randomised, active-controlled, phase 3b trial. *Lancet* 2020, 395, 1496-1505 [doi:10.1016/S0140-6736(20)30564-X].
- 86. Mease P.J.; van der Heijde D.; Ritchlin C.T.; Okada M.; Cuchacovich R.S.; Shuler C.L.; Lin C.Y.; Braun D.K.; Lee C.H.; Gladman D.D.; et al. Ixekizumab, an interleukin-17A specific monoclonal antibody, for the treatment of biologic-naive patients with active psoriatic arthritis: results from the 24-week randomised, double-blind, placebo-controlled and active (adalimumab)-controlled period of the phase III trial SPIRIT-P1. Ann. Rheum. Dis. 2017, 76, 79-87 [doi:10.1136/annrheumdis-2016-209709].
- 87. Nash P.; Kirkham B.; Okada M.; Rahman P.; Combe B.; Burmester G.R.; Adams D.H.; Kerr L.; Lee C.; Shuler C.L.; et al. Ixekizumab for the treatment of patients with active psoriatic arthritis and an inadequate response to tumour necrosis factor inhibitors: results from the 24-week randomised, double-blind, placebo-controlled period of the SPIRIT-P2 phase 3 trial. *Lancet* 2017, 389, 2317-2327 [doi:10.1016/S0140-6736(17)31429-0].
- 88. Mease P.J.; Smolen J.S.; Behrens F.; Nash P.; Liu Leage S.; Li L.; Tahir H.; Gooderham M.; Krishnan E.; Liu-Seifert H.; et al. A head-to-head comparison of the efficacy and safety of ixekizumab and adalimumab in

- biological-naïve patients with active psoriatic arthritis: 24-week results of a randomised, open-label, blinded-assessor trial. *Ann. Rheum. Dis.* 2020, 79, 123-131 [doi:10.1136/annrheumdis-2019-215386].
- 89. Ritchlin C.T.; Kavanaugh A.; Merola J.F.; Schett G.; Scher J.U.; Warren R.B.; Gottlieb A.B.; Assudani D.; Bedford-Rice K.; Coarse J.; et al. Bimekizumab in patients with active psoriatic arthritis: results from a 48-week, randomised, double-blind, placebo-controlled, dose-ranging phase 2b trial. *Lancet* 2020, 395, 427-440 [doi:10.1016/S0140-6736(19)33161-7].
- 90. Mease P.J.; Asahina A.; Gladman D.D.; Tanaka Y.; Tillett W.; Ink B.; Assudani D.; de la Loge C.; Coarse J.; Eells J.; et al. Effect of bimekizumab on symptoms and impact of disease in patients with psoriatic arthritis over 3 years: results from BE ACTIVE. *Rheumatol.* (Oxf. Engl.) 2023, 62, 617-628 [doi:10.1093/rheumatology/keac353].
- 91. Merola J.F.; Landewé R.; McInnes I.B.; Mease P.J.; Ritchlin C.T.; Tanaka Y.; Asahina A.; Behrens F.; Gladman D.D.; Gossec L.; et al. Bimekizumab in patients with active psoriatic arthritis and previous inadequate response or intolerance to tumour necrosis factor-α inhibitors: a randomised, double-blind, placebocontrolled, phase 3 trial (BE COMPLETE). *Lancet* 2023, 401, 38-48 [doi:10.1016/S0140-6736(22)02303-0].
- 92. Mease P.J.; Helliwell P.S.; Hjuler K.F.; Raymond K.; McInnes I. Brodalumab in psoriatic arthritis: results from the randomised phase III AMVISION-1 and AMVISION-2 trials. *Ann. Rheum. Dis.* **2021**, *80*, 185-193 [doi:10.1136/annrheumdis-2019-216835].
- 93. Mease P.J.; McInnes I.B.; Tam L.S.; Eaton K.; Peterson S.; Schubert A.; Chakravarty S.D.; Parackal A.; Karyekar C.S.; Nair S.; et al. Comparative effectiveness of guselkumab in psoriatic arthritis: results from systematic literature review and network meta-analysis. *Rheumatol.* (Oxf. Engl.) 2021, 60, 2109-2121 [doi:10.1093/rheumatology/keab119].
- 94. Colombel J.F.; Sendid B.; Jouault T.; Poulain D. Secukinumab failure in Crohn's disease: the yeast connection? *Gut* 2013, 62, 800-801 [doi:10.1136/gutjnl-2012-304154].
- 95. Cuthbert R.J.; Watad A.; Fragkakis E.M.; Dunsmuir R.; Loughenbury P.; Khan A.; Millner P.A.; Davison A.; Marzo-Ortega H.; Newton D.; et al. Evidence that tissue resident human enthesis γδT-cells can produce IL-17A independently of IL-23R transcript expression. *Ann. Rheum. Dis.* 2019, 78, 1559-1565 [doi:10.1136/annrheumdis-2019-215210].
- 96. Fragoulis G.E.; Siebert S. The role of IL-23 and the use of IL-23 inhibitors in psoriatic arthritis. *Musculoskelet. Care* 2022, 20 suppl 1, S12–S21 [doi:10.1002/msc.1694].
- 97. McInnes I.B.; Kavanaugh A.; Gottlieb A.B.; Puig L.; Rahman P.; Ritchlin C.; Brodmerkel C.; Li S.; Wang Y.; Mendelsohn A.M.; et al. Efficacy and safety of ustekinumab in patients with active psoriatic arthritis: 1 year results of the phase 3, multicentre, double-blind, placebo-controlled PSUMMIT 1 trial. *Lancet* 2013, 382, 780-789 [doi:10.1016/S0140-6736(13)60594-2].
- 98. Kavanaugh A.; Ritchlin C.; Rahman P.; Puig L.; Gottlieb A.B.; Li S.; Wang Y.; Noonan L.; Brodmerkel C.; Song M.; et al. Ustekinumab, an anti-IL-12/23 p40 monoclonal antibody, inhibits radiographic progression in patients with active psoriatic arthritis: results of an integrated analysis of radiographic data from the phase 3, multicentre, randomised, double-blind, placebo-controlled PSUMMIT-1 and PSUMMIT-2 trials. Ann. Rheum. Dis. 2014, 73, 1000-1006 [doi:10.1136/annrheumdis-2013-204741].
- 99. Kavanaugh A.; Puig L.; Gottlieb A.B.; Ritchlin C.; Li S.; Wang Y.; Mendelsohn A.M.; Song M.; Zhu Y.; Rahman P.; et al. Maintenance of clinical efficacy and radiographic benefit through two years of ustekinumab therapy in patients with active psoriatic arthritis: results from a randomized, placebo-controlled Phase III trial. *Arthritis Care Res.* (Hoboken) 2015, 67, 1739-1749 [doi:10.1002/acr.22645].
- 100. Araujo E.G.; Englbrecht M.; Hoepken S.; Finzel S.; Kampylafka E.; Kleyer A.; Bayat S.; Schoenau V.; Hueber A.; Rech J.; et al. Effects of ustekinumab versus tumor necrosis factor inhibition on enthesitis: results from the enthesial clearance in psoriatic arthritis (ECLIPSA) study. Semin. Arthritis Rheum. 2019, 48, 632-637 [doi:10.1016/j.semarthrit.2018.05.011].
- 101. Gossec L.; Siebert S.; Bergmans P.; de Vlam K.; Gremese E.; Joven-Ibáñez B.; Korotaeva T.V.; Lavie F.; Noël W.; Nurmohamed M.T.; et al. Persistence and effectiveness of the IL-12/23 pathway inhibitor ustekinumab or tumour necrosis factor inhibitor treatment in patients with psoriatic arthritis: 1-year results from the real-world PsABio Study. Ann. Rheum. Dis. 2022, 81, 823-830 [doi:10.1136/annrheumdis-2021-221640].
- 102. Gordon K.B.; Strober B.; Lebwohl M.; Augustin M.; Blauvelt A.; Poulin Y.; Papp K.A.; Sofen H.; Puig L.; Foley P.; et al. Efficacy and safety of risankizumab in moderate-to-severe plaque psoriasis (UltIMMa-1 and UltIMMa-2): results from two double-blind, randomised, placebo-controlled and ustekinumab-controlled phase 3 trials. *Lancet* 2018, 392, 650-661 [doi:10.1016/S0140-6736(18)31713-6].
- 103. Deodhar A.; Helliwell P.S.; Boehncke W.H.; Kollmeier A.P.; Hsia E.C.; Subramanian R.A.; Xu X.L.; Sheng S.; Agarwal P.; Zhou B.; et al. Guselkumab in patients with active psoriatic arthritis who were biologic-naive or had previously received TNFα inhibitor treatment (DISCOVER-1): a double-blind, randomised, placebocontrolled phase 3 trial. *Lancet* 2020, 395, 1115-1125 [doi:https://doi.org/10.1016/S0140-6736(20)30265-8].
- 104. Mease P.J.; Rahman P.; Gottlieb A.B.; Kollmeier A.P.; Hsia E.C.; Xu X.L.; Sheng S.; Agarwal P.; Zhou B.; Zhuang Y.; et al. Guselkumab in biologic-naive patients with active psoriatic arthritis (DISCOVER-2): a

- double-blind, randomised, placebo-controlled phase 3 trial. Lancet 2020, 395, 1126-1136 [doi:https://doi.org/10.1016/S0140-6736(20)30263-4].
- 105. Coates L.C.; Gossec L.; Theander E.; Bergmans P.; Neuhold M.; Karyekar C.S.; Shawi M.; Noël W.; Schett G.; McInnes I.B. Efficacy and safety of guselkumab in patients with active psoriatic arthritis who are inadequate responders to tumour necrosis factor inhibitors: results through one year of a phase IIIb, randomised, controlled study (COSMOS). Ann. Rheum. Dis. 2022, 81, 359–369 [doi:10.1136/annrheumdis-2021-220991].
- 106. McInnes I.B.; Rahman P.; Gottlieb A.B.; Hsia E.C.; Kollmeier A.P.; Xu X.L.; Jiang Y.; Sheng S.; Shawi M.; Chakravarty S.D.; et al. Long-term efficacy and safety of guselkumab, a monoclonal antibody specific to the p19 subunit of interleukin-23, through two years: results from a Phase III, randomized, double-blind, placebo-controlled study conducted in biologic-naive patients with active psoriatic arthritis. *Arthritis Rheumatol.* 2022, 74, 475-485 [doi:10.1002/art.42010].
- 107. Sweet K.; Song Q.; Loza M.J.; McInnes I.B.; Ma K.; Leander K.; Lakshminarayanan V.; Franks C.; Cooper P.; Siebert S. Guselkumab induces robust reduction in acute phase proteins and type 17 effector cytokines in active psoriatic arthritis: results from phase 3 trials. RMD Open 2021, 7 [doi:10.1136/rmdopen-2021-001679].
- 108. Kristensen L.E.; Keiserman M.; Papp K.; McCasland L.; White D.; Lu W.; Wang Z.; Soliman A.M.; Eldred A.; Barcomb L.; et al. Efficacy and safety of risankizumab for active psoriatic arthritis: 24-week results from the randomised, double-blind, phase 3 KEEPsAKE 1 trial. Ann. Rheum. Dis. 2022, 81, 225–231 [doi:10.1136/annrheumdis-2021-221019].
- 109. Östör A.; Van den Bosch F.; Papp K.; Asnal C.; Blanco R.; Aelion J.; Alperovich G.; Lu W.; Wang Z.; Soliman A.M.; *et al.* Efficacy and safety of risankizumab for active psoriatic arthritis: 24-week results from the randomised, double-blind, phase 3 KEEPsAKE 2 trial. *Ann. Rheum. Dis.* **2022**, *81*, 351–358 [doi:10.1136/annrheumdis-2021-221048].
- 110. Mease P.J.; Chohan S.; Fructuoso F.J.G.; Luggen M.E.; Rahman P.; Raychaudhuri S.P.; Chou R.C.; Mendelsohn A.M.; Rozzo S.J.; Gottlieb A. Efficacy and safety of tildrakizumab in patients with active psoriatic arthritis: results of a randomised, double-blind, placebo-controlled, multiple-dose, 52-week phase IIb study. *Ann. Rheum. Dis.* **2021**, *80*, 1147–1157 [doi:10.1136/annrheumdis-2020-219014].
- 111. Mease P.J.; Helliwell P.S.; Gladman D.D.; Poddubnyy D.; Baraliakos X.; Chakravarty S.D.; Kollmeier A.P.; Hsia E.C.; Xu X.L.; Sheng S.; et al. Efficacy of guselkumab on axial involvement in patients with active psoriatic arthritis and sacroiliitis: a post-hoc analysis of the phase 3 DISCOVER-1 and DISCOVER-2 studies. *The Lancet Rheumatol.* **2021**, *3*, e715-e723 [doi:https://doi.org/10.1016/S2665-9913(21)00105-3].
- 112. Campanaro F.; Batticciotto A.; Zaffaroni A.; Cappelli A.; Donadini M.P.; Squizzato A. JAK inhibitors and psoriatic arthritis: A systematic review and meta-analysis. *Autoimmun. Rev.* **2021**, *20*, 102902 [doi:10.1016/j.autrev.2021.102902].
- 113. Strand V.; de Vlam K.; Covarrubias-Cobos J.A.; Mease P.J.; Gladman D.D.; Graham D.; Wang C.; Cappelleri J.C.; Hendrikx T.; Hsu M.A. Tofacitinib or adalimumab versus placebo: patient-reported outcomes from OPAL Broaden-a phase III study of active psoriatic arthritis in patients with an inadequate response to conventional synthetic disease-modifying antirheumatic drugs. *RMD Open* **2019**, *5*, e000806 [doi:10.1136/rmdopen-2018-000806].
- 114. Strand V.; de Vlam K.; Covarrubias-Cobos J.A.; Mease P.J.; Gladman D.D.; Chen L.; Kudlacz E.; Wu J.; Cappelleri J.C.; Hendrikx T.; et al. Effect of tofacitinib on patient-reported outcomes in patients with active psoriatic arthritis and an inadequate response to tumour necrosis factor inhibitors in the phase III, randomised controlled trial: OPAL Beyond. RMD Open 2019, 5, e000808 [doi:10.1136/rmdopen-2018-000808].
- 115. Nash P.; Coates L.C.; Kivitz A.J.; Mease P.J.; Gladman D.D.; Covarrubias-Cobos J.A.; FitzGerald O.; Fleishaker D.; Wang C.; Wu J.; et al. Safety and efficacy of tofacitinib in patients with active psoriatic arthritis: interim analysis of OPAL balance, an open-label, long-term extension study. *Rheumatol. Ther.* 2020, 7. 553-580 [doi:10.1007/s40744-020-00209-4].
- 116. Ytterberg S.R.; Bhatt D.L.; Mikuls T.R.; Koch G.G.; Fleischmann R.; Rivas J.L.; Germino R.; Menon S.; Sun Y.; Wang C.; et al. Cardiovascular and cancer risk with tofacitinib in rheumatoid arthritis. N. Engl. J. Med. 2022, 386, 316-326 [doi:10.1056/NEJMoa2109927].
- 117. Mease P.J.; Lertratanakul A.; Papp K.A.; van den Bosch F.E.; Tsuji S.; Dokoupilova E.; Keiserman M.W.; Bu X.; Chen L.; McCaskill R.M.; et al. Upadacitinib in patients with psoriatic arthritis and inadequate response to biologics: 56-week data from the randomized controlled Phase 3 SELECT-PsA 2 study. *Rheumatol. Ther.* **2021**, *8*, 903-919 [doi:10.1007/s40744-021-00305-z].
- 118. Muensterman E.; Engelhardt B.; Gopalakrishnan S.; Anderson J.K.; Mohamed M.F.Upadacitinib pharmacokinetics and exposure-response analyses of efficacy and safety in psoriatic arthritis patients Analyses of phase III clinical trials. Clin. Transl. Sci. 2022, 15, 267-278 [doi:10.1111/cts.13146].
- 119. McInnes I.B.; Kato K.; Magrey M.; Merola J.F.; Kishimoto M.; Pacheco-Tena C.; Haaland D.; Chen L.; Duan Y.; Zueger P.; et al. Upadacitinib in patients with psoriatic arthritis and an inadequate response to non-

- biological therapy: 56-week data from the phase 3 SELECT-PsA 1 study. RMD Open 2021, 7 [doi:10.1136/rmdopen-2021-001838].
- 120. Mease P.J.; Lertratanakul A.; Anderson J.K.; Papp K.; Van den Bosch F.; Tsuji S.; Dokoupilova E.; Keiserman M.; Wang X.; Zhong S.; et al. Upadacitinib for psoriatic arthritis refractory to biologics: SELECT-PsA 2. Ann. Rheum. Dis. 2021, 80, 312–320 [doi:10.1136/annrheumdis-2020-218870].
- 121. van der Heijde D.; Baraliakos X.; Gensler L.S.; Maksymowych W.P.; Tseluyko V.; Nadashkevich O.; Abi-Saab W.; Tasset C.; Meuleners L.; Besuyen R.; et al. Efficacy and safety of filgotinib, a selective Janus kinase 1 inhibitor, in patients with active ankylosing spondylitis (TORTUGA): results from a randomised, placebo-controlled, phase 2 trial. *Lancet* 2018, 392, 2378-2387 [doi:10.1016/S0140-6736(18)32463-2].
- 122. Hellstrom W.J.G.; Dolhain R.J.E.M.; Ritter T.E.; Watkins T.R.; Arterburn S.J.; Dekkers G.; Gillen A.; Tonussi C.; Gilles L.; Oortwijn A.; et al. MANTA and MANTA-RAy: rationale and design of trials evaluating effects of filgotinib on semen parameters in patients with inflammatory diseases. *Adv. Ther.* **2022**, *39*, 3403-3422 [doi:10.1007/s12325-022-02168-4].
- 123. Časo F.; Costa L.; Triggianese P.; Maione F.; Bertolini N.; Vastarella M.; Chimenti M.S.; Tasso M. Recent developments for new investigational JAK inhibitors in psoriatic arthritis. *Expert Opin. Investig. Drugs* **2023**, 1-11 [doi:10.1080/13543784.2023.2207737].
- 124. Mease P.J.; Deodhar A.A.; van der Heijde D.; Behrens F.; Kivitz A.J.; Neal J.; Kim J.; Singhal S.; Nowak M.; Banerjee S. Efficacy and safety of selective TYK2 inhibitor, deucravacitinib, in a phase II trial in psoriatic arthritis. *Ann. Rheum. Dis.* **2022**, *81*, 815-822 [doi:10.1136/annrheumdis-2021-221664].
- 125. Mease P.; Helliwell P.; Silwinska-Stanczyk P.; Miakisz M.; Ostor A.; Peeva E.; Vincent M.S.; Sun Q.; Sikirica V.; Winnette R.; et al. Efficacy and safety of tyrosine kinase 2/Janus kinase 1 inhibitor brepocitinib for active psoriatic arthritis: A Phase IIb randomized controlled trial. *Arthritis Rheumatol.* 2023 [doi:10.1002/art.42519].
- 126. Cutolo M.; Myerson G.E.; Fleischmann R.M.; Lioté F.; Díaz-González F.; Van den Bosch F.; Marzo-Ortega H.; Feist E.; Shah K.; Hu C.; et al. A Phase III, randomized, controlled trial of apremilast in patients with psoriatic arthritis: results of the PALACE 2 trial. *J. Rheumatol.* 2016, 43, 1724–1734 [doi:10.3899/jrheum.151376].
- 127. Edwards C.J.; Blanco F.J.; Crowley J.; Birbara C.A.; Jaworski J.; Aelion J.; Stevens R.M.; Vessey A.; Zhan X.; Bird P. Apremilast, an oral phosphodiesterase 4 inhibitor, in patients with psoriatic arthritis and current skin involvement: a phase III, randomised, controlled trial (PALACE 3). *Ann. Rheum. Dis.* **2016**, 75, 1065–1073 [doi:10.1136/annrheumdis-2015-207963].
- 128. Wells A.F.; Edwards C.J.; Kivitz A.J.; Bird P.; Nguyen D.; Paris M.; Teng L.; Aelion J.A. Apremilast monotherapy in DMARD-naive psoriatic arthritis patients: results of the randomized, placebo-controlled PALACE 4 trial. *Rheumatology (Oxford)* **2018**, *57*, 1253-1263 [doi:10.1093/rheumatology/key032].
- 129. Orr C.; Veale D.J. Is there a need for new agents with novel mechanisms of action in psoriatic arthritis? *Ann. Rheum. Dis.* **2014**, *73*, 951-953 [doi:10.1136/annrheumdis-2013-204934].
- 130. Mease P.J.; Genovese M.C.; Weinblatt M.E.; Peloso P.M.; Chen K.; Othman A.A.; Li Y.; Mansikka H.T.; Khatri A.; Wishart N.; et al. Phase II study of ABT-122, a tumor necrosis factor- and Interleukin-17A-Targeted dual variable domain immunoglobulin, in patients with psoriatic arthritis with an inadequate response to methotrexate. *Arthritis Rheumatol.* **2018**, *70*, 1778-1789 [doi:10.1002/art.40579].
- 131. Sun S.; Ding Z.; Yang X.; Zhao X.; Zhao M.; Gao L.; Chen Q.; Xie S.; Liu A.; Yin S.; et al. Nanobody: A small antibody with big implications for tumor therapeutic strategy. *Int. J. Nanomedicine* **2021**, *16*, 2337-2356 [doi:10.2147/IJN.S297631].
- 132. Puig L.; Bakulev A.L.; Kokhan M.M.; Samtsov A.V.; Khairutdinov V.R.; Morozova M.A.; Zolkin N.A.; Kuryshev I.V.; Petrov A.N.; Artemeva A.V.; et al. Efficacy and safety of netakimab, A novel anti-IL-17 monoclonal antibody, in patients with moderate to severe plaque psoriasis. Results of A 54-week randomized double-blind placebo-controlled PLANETA clinical trial. *Dermatol. Ther.* (Heidelb) 2021, 11, 1319-1332 [doi:10.1007/s13555-021-00554-4].
- 133. Papp K.A.; Weinberg M.A.; Morris A.; Reich K. IL17A/F nanobody sonelokimab in patients with plaque psoriasis: a multicentre, randomised, placebo-controlled, phase 2b study. *Lancet* **2021**, 397, 1564-1575 [doi:10.1016/S0140-6736(21)00440-2].