Disclaimer/Publisher's Note: The statements, opinions, and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions, or products referred to in the content.

Remieri

Newer Cancer Therapies and Implications for the Perioperative Period

Alessandro De Camilli and Gregory Fischer *

Memorial Sloan Kettering

* Correspondence: fischerg@mskcc.org

Abstract: Targeted cancer therapy has grown exponentially in the last decade, representing the largest frontier in the modern concept of precision medicine. This renaissance is largely due to our rapidly evolving ability to biochemically harness our immune systems to target cancer-specific antigens. Combined with an increasing armamentarium of surgical and non-invasive techniques for cancer mitigation we have upgraded cancer therapy from a non-specific assault on cell-division to a multi-modal, targeted approach. This renaissance has resulted in an increase in the perioperative footprint of patients with advanced cancer. The scope of the perioperative clinician has evolved from one of passive/reactive management of complications to a proactive approach that starts in the preoperative optimization phase and continues into the postoperative period. The perioperative world should not exist in a silo, and thus involvement of the oncology team is paramount. To provide safe oncologic care, it is essential to be up to date in newer cancer therapies, lest we act on erroneous and/or outdated information. Oncologists should weigh in on immune status, recent chemotherapeutic exposures, and potentially evolving toxicities. These might involve critical changes to perioperative infectious prophylaxis, steroid administration, and coagulopathy management. First, patients living beyond the typical life-expectancy for their cancer will carry a high burden of cancer-related downstream effects. General metabolic issues will include malnutrition, cachexia, and gastric outlet obstruction. Severe protein-calorie malnutrition, as well as hepatic and renal dysfunction can impact the metabolism and clearance of drugs. The cardiovascular, endocrinologic, pulmonary, and central nervous system implications of new cancer therapies will be discussed extensively below. Second, we have to enhance our ability to manage a cancer population who may be experiencing increasing novel chemotherapeutic toxicities. Newer agents differ from historical agents, which were blunt-force tools aimed at nonspecific destruction of cell-division. These drugs are efficacious but physiologically costly. Newer immunogenic therapies have a slightly improved acute safety profile, and a more complex chronic toxicity profile that may be overt or insidious. Chronic toxicities can present overtly or as a smoldering undertone to other active medical conditions such as sepsis, pneumonia, failure to thrive, or renal failure. The new therapies discussed in this review occur in the context of an overall increasingly multi-faceted cancer care machine. An example is screening for esophageal cancer in patients with Barrett's esophagus, or periodic chest CTs for patients with a strong history of tobacco use. These interventions are more likely to detect disease while it is still resectable, and resections will commonly accompany neoadjuvant therapy. These approaches commonly involve new immunotherapies whose efficacy is higher-yield with a lower disease burden. In a recent example, the immune-checkpoint inhibitors (ICIs) ipilimumab plus nivolumab have significantly increased survival rate for squamous cell esophageal cancer when combined with standard chemotherapy. Surgical resection has evolved to one that often accompanies neoadjuvant chemo and radiation. The increasing efficacy of this approach may de-necessitate many major resections, and increase the number of sub-anatomic resections and minimally invasive procedures. Improved techniques - notably for hepatectomy, whipple procedure, prostatectomy, nephrectomy - have increased the safety profile and decreased the perioperative physiologic burden. This has enabled a broader subset of patients (due to cancer burden or due to medical comorbidities) to qualify for the operation, and increased the involvement of anesthesia and their perioperative services. With an increase in surgical candidacy comes an increase in postsurgical complications, and post-surgical maintenance such as drains, embolizations, imaging studies, etc. In the non-OR anesthesia realm, there have been considerable advancements in cancer "micro-treatments" such as hepatic metastasis embolization, tumor ablation, and peri-tumor vascular embolization. There are an everincreasing number of IR-guided embolizations for bleeding tumors, aspiration of malignant effusions, and drainage of abscesses. This review serves to provide insight into new categories of chemotherapy and their perioperative implications.

1. Introduction—A New Paradigm of Perioperative Cancer Care

The last decade has brought about a revolution in cancer therapies that harness our immune system to target cancer-specific antigens. These novel agents increasingly play a role in neo-adjuvant therapy and have greatly improved quality of life for patients with advanced cancer. More than ever, it is necessary for the perioperative provider to be up to date in the physiology underlying these therapies, as well as their unique toxicity profiles.

Early generation chemotherapeutics relied on nonspecific destruction of cell-division, with toxicities mostly related to en-mass destruction of rapidly dividing cells. Newer immunotherapies have a more benign acute safety profile, but nonetheless can exert chronic toxicities that may be more insidious and require a higher index of suspicion for detection. The predominant cause of toxicities results from unwanted upregulated immune system activity, or molecular mimicry leading to destruction of "bystander" tissue.

To provide an example: Ipilumumab and nivolumab are two immune-checkpoint inhibitors (ICIs) that have recently been shown to improve survival when included as part of neoadjuvant therapy for squamous cell esophageal cancer [1]. However, the two drugs in combination can lead to a non-negligible incidence of ICI pneumonitis and ICI hypophysitis. It is thus becoming more important for thoracic surgeons and anesthesiologists who care for these patients to be aware of the potential risks of adrenal insufficiency and reduced pulmonary capacity. A thorough therapeutic history and close collaboration with oncologists are necessary to provide safe care.

The purpose of this review is to give the perioperative provider physiologic insight into new targeted immunotherapies, with a focus on their implications for safe perioperative care.

2. Immune Checkpoint Inhibitors

Immune checkpoint inhibitors have earned their keep as tremendously successful adjuncts to all manner of chemotherapeutic regimens. Our innate immune system relies on T cell activation and proliferation to destroy foreign cells. T cell activation requires both antigen-receptor coupling as well as co-stimulation by other immune effectors cells. This two-fold process underpins the versatility of our immune system but is also the process by which cancer cells can evade detection. T cell activation can be inhibited by two pathways: the cytotoxic T lymphocyte-associated antigen 4 (CTLA-4) pathway, and the programmed cell death protein (PD-LA1) pathway. This interplay of T cell activation and inhibition is referred to as an "immune checkpoint."

Cancer cells have evolved to thwart T cell activation by taking advantage of the inhibition role of this immune checkpoint, since CTLA-4 and PD-LA1 antigens are commonly found on the surfaces of tumor cells. Activation of these ligands by tumor cells allows them to evade detection.

Immune checkpoint inhibitors (ICIs), as a therapeutic target, prevent tumor cell evasion by acting as monoclonal antibodies against the CTLA-4 and PD-LA1 ligands. In preventing their ability to inactivate T cells, ICIs ensure that tumor cells are vulnerable to destruction (see Figure 1).



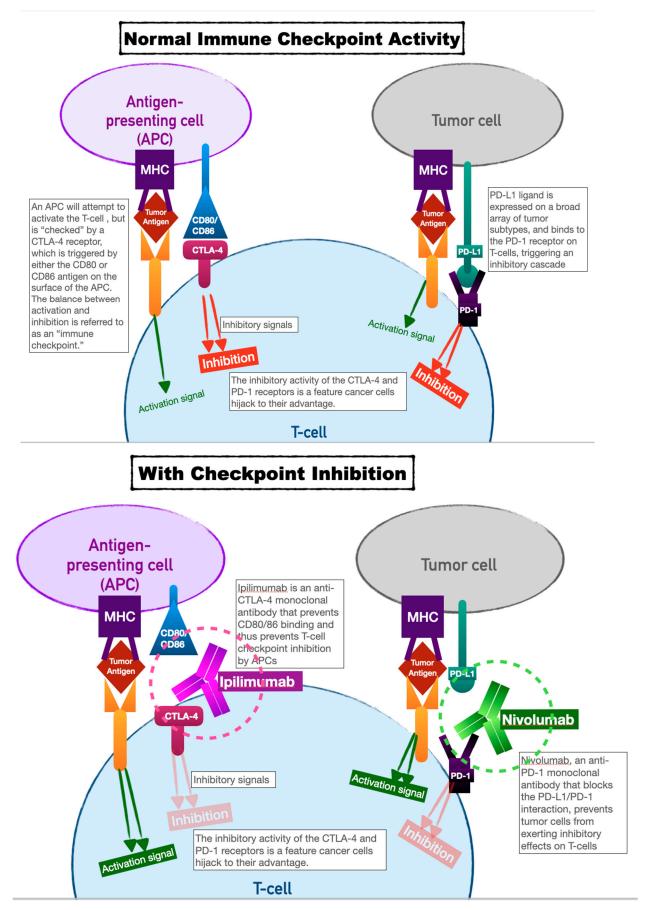


Figure 1. Above, normal immune checkpoint activity means that cancer (as tumor cells directly, or as tumor antigen presented via an APC) can inhibit T-cell activation via the CTLA-4 and PD-1 ligands. Below, immune checkpoint inhibitors prevent this inhibition, leading to T-cell activation.

This has proven to be a very effective therapeutic target, with successes in the treatment of melanoma, breast cancer, lymphoma, and prostate cancer.

Checkpoint inhibition technology has continued to advance, as new agents are developed targeting additional T cell pathways, and as existing ones are used on new tumor subtypes.

The first immune checkpoint inhibitors approved for human use were Ipilimumab and Nivolumab (2011 and 2014, respectively). Ipilimumab acts as a monoclonal antibody against CTLA-4; Nivolumab against PD-1. Ipilimumab was initially shown to dramatically improve survival in patients with metastatic melanoma [2]. The subsequent arrival of Nivolumab showed promise of the two used as combination therapy, both for metastatic melanoma and several other cancers - renal cell carcinoma, unresectable non-small-cell lung cancer (NSCL), prostate cancer, and esophageal cancer [3,4].

Immune checkpoint inhibitors have shown promise in treating tumors that have been historically resistant to other chemo or immunotherapeutic agents, but there are highly specific indications for the use of each. Up to 45% of patients with cancer are candidates for ICI therapy [5], so the perioperative footprint of these patients continues to rise. Their efficacy is determined by the rate of target PD-L1 or CTLA-4 receptor expression on the tumor cells, the mutational burden of tumors, and the relative inefficacy of classic chemotherapy agents for the tumor in question.

ICI therapy in the perioperative period is an area of ongoing study. Thus far, a few studies at major cancer centers have shown no association perioperative morbidity of any type [6], and in fact, most show greatly improved results when added as neo-adjuvant therapy [7,8].

2.1. Specific Toxicities and Perioperative Implications

Toxicity from ICIs occurs via multiple mechanisms. The up-regulation of T-cell activity in general can lead to lymphocyte infiltration in normal parenchyma and cause nonspecific tissue damage. ICIs can lead to a loss of immunologic self-tolerance, wherein more T and B cells will be activated regardless of the presence of tumor cells. Molecular mimicry can occur, wherein ICI targets may appear on non-tumor cells, which are inadvertently triggered for destruction. Direct drug toxicity is less common, owing to their relatively inert antibody structure [9].

2.1.1. Gastrointestinal

The increase in T-cell activation by ICIs is most clinically significant in sites that have a high number of T-cells: notably the lining of the GI tract, the airways, and skin. As such, the most prevalent condition is immune-checkpoint inhibitor induced enterocolitis, which can occur in up to 20% of patients and onset is typically within 2-3 months of therapy initiation. Mild cases of ICI enterocolitis (a few loose stools per day) are treated with low doses of oral corticosteroids. Endoscopy is typically obtained for patients with an increase in stool of over 4-6 more events over their baseline, to establish diagnosis and exclude other infectious or malabsorptive causes. These patients with moderate disease will be started on steroids and considered for biologic agents including infliximab. Repeat endoscopy is indicated after 2-3 months to gauge endoscopic response. For patients with mild symptoms, temporary cessation or low dose corticosteroids might suffice. For moderate to severe symptoms, ICI therapy is usually stopped permanently [10]. Surgical intervention is extremely rare, and reserved for cases of severe, refractory colitis with megacolon or perforation [11].

Perioperative implications of this condition track with disease severity. Patients on ICI should be screened for gastritis or other upper GI conditions that might affect anesthetic management - i.e.: rapid sequence induction. Patients with malabsorptive diarrhea warrant a comprehensive metabolic panel and screening for electrolyte analysis.

ICI hepatitis is a less common entity, but critical not to miss. It has an incidence of 5-10% [12], usually occurring within 2-3 months of therapy. It will typically manifest as an occult increase in liver function testing (LFTs). The pattern of liver dysfunction typically follows a hepatocellular pattern (AST and ALT elevation) and less commonly a cholestatic pattern (ALK elevation and hyperbilirubinemia). The risk increases with dual-ICI therapy [13]. CT imaging should be performed to rule out liver metastases, and abdominal ultrasound should be obtained to rule out biliary

obstruction in patients with hyperbilirubinemia. Workup should include hepatitis viral panels, history of alcohol use, and presence of concomitant hepatotoxins.

ICI hepatitis is graded on a scale: Grades 1 and 2 are defined by AST and ALT levels of 2.5 to 5 times normal. These cases are typically treated with corticosteroids, which can be tapered over a 4–6-week period once LFTs return to normal. Very mild cases in patients with known liver metastases may warrant observation alone. Grade 3 and 4 ICI hepatitis are defined by a 5 to 20-fold elevation of LFTs and the presence of any clinical signs of liver dysfunction (asterixis, encephalopathy, ascites). In addition to drug cessation and a course of IV corticosteroids (typically methylprednisolone 1 mg/kg/day), these patients should undergo a liver biopsy to confirm the diagnosis or evaluate for other causes of liver dysfunction. For patients with steroid-refractory liver dysfunction, mycophenolate mofetil and tacrolimus can be considered, in coordination with a hepatologist experienced in treating the condition [14]. Once treated and resolved, ICI therapy is only resumed for patients who achieved improvement from grade 2 toxicity or less.

To mitigate perioperative risk, patients requiring surgery on ICI therapy should be screened with recent LFTs. LFT elevation should prompt additional workup should including hepatitis viral panels, history of alcohol use, CT scan of the liver, and evaluation for concomitant hepatotoxic agents. Perioperative studies on patients with known ICI hepatitis are limited. It is recommended to postpone elective surgery on patients with hepatitis of any cause. Patients with ICI hepatitis should be treated as patients with liver dysfunction of any other cause. The Child-Pugh classification of liver dysfunction has been validated as a classification system of patients with cirrhosis to distinguish surgical mortality, with an increased score assigned to patients with signs of decompensated hepatic failure such as coagulopathy, ascites, and encephalopathy [15].

A less common effect of ICI therapy is pancreatic dysfunction. Approximately 4% of patients on ICIs will have an asymptomatically elevated lipase. The incidence of patients with symptomatic pancreatitis is <1% [16,17]. Symptoms of ICI pancreatitis are similar to those of all-cause pancreatitis, including epigastric pain, fever, and nausea. A CT scan should be obtained to rule out other causes of pancreatitis and biliary obstruction. Patients with asymptomatic lipase elevation are not recommended to receive any specific treatment other than monitoring. Symptomatic pancreatitis should be treated with usual care: fluid resuscitation, NPO status, and analgesia. It is notable that studies are extremely limited for ICI pancreatitis, including the lack of a standardized grading system. Most centers use steroids and ICI cessation for patients with severe symptoms requiring hospitalization [18,19]. As with usual pancreatitis, antibiotics are not routinely recommended in the absence of abscess or presence of another infectious source [20]. There is no established role for surgery in ICI pancreatitis, which in general is a last resort intervention for complicated pancreatitis of all causes.

New-onset type 1 diabetes is a rare result of acute autoimmune pancreatic exocrine dysfunction, with an incidence of <1% [21]. The most common presentation of this adverse effect is new-onset DKA. Destruction of islet cells renders patients permanently insulin-dependent in most cases, an outcome that is not treatable with steroids [22].

2.1.2. Pulmonary

Pulmonary toxicity from ICIs is referred to as ICI pneumonitis. The overall incidence is approximately 5% [23,24]. Cough and dyspnea are the typical initial symptoms, but the clinical syndrome falls on a wide spectrum of respiratory disease: acute, chronic, waxing/waning, permanently disabling.

The acute form of ICI pulmonary toxicity follows a pattern of acute hypersensitivity pneumonitis and can progress to full-blown acute respiratory distress syndrome (ARDS). Chronic ICI pneumonitis occurs within 3 months to one year after initiation of therapy [25]. In both conditions, alternative causes of respiratory symptoms should be ruled out with a history of exposures, toxins, smoking history, and CT scans with contrast should be obtained. Radiologic patterns of chronic pulmonary toxicity typically reveal ground glass opacities and peripheral multifocal consolidations, although there is no pathognomonic finding for ICI pneumonitis. Bronchoscopy is only

recommended for cases that do not respond to initial treatment, or for whom alternative diagnoses are being ruled out - i.e.: organizing infection, progression of malignancy, etc. Bronchoscopy specimens, if obtained, will show alveolar lymphocytic infiltration. Pathology specimens in ICI pneumonitis show a pattern of bronchiolitis obliterans and organizing pneumonia, with a lymphocytic predominance [26]. Smoking and prior radiation therapy increase the likelihood of ICI pneumonitis [27].

ICI pneumonitis is graded on a scale of grade 1 to 5 [28]. Patients with no symptoms and only mild radiographic findings are considered grade 1, and simply warrant observation with continuation of treatment. Patients with grade 2-5 (symptoms ranging from dyspnea/cough to fulminant respiratory failure) should be treated with glucocorticoids and cessation of treatment. Typical doses of prednisone range from 0.5-1mg/kg daily, and the typical course is 2-4 weeks. Addition of anti-IL-6 agents has shown some promising results in a subset of patients who are refractory to steroids. The prognosis of ICI pneumonitis varies, as the pattern of disease is heterogenous. Patients with grade 1-3 disease do well with early recognition and steroid treatment. Patients with grade 4 or above, or who have recurrent or treatment-refractory disease, have a significantly higher burden of inpatient care, oxygen dependence, and overall mortality. It is worth noting that radiographic findings can persist long after symptom resolution [29].

Perioperative management of patients with ICI pneumonitis varies depending on the context in which it arises. For patients on ICI therapy as neo-adjuvant to surgery, such as in non-small cell lung cancer [30], the benefits of therapy have been shown to outweigh the risks. However, it remains essential to screen patients for pneumonitis symptomatology, which will inform operative risk. A history of dyspnea or cough in any patient on ICIs should warrant a chest x ray, and their surgical risk should be stratified based on standard-of-care for patients with all-cause pulmonary disease. Recommendations for preoperative lung function testing are continually evolving, with recommendations including spirometry for all patients undergoing lung resection, and for patients with a history of severe obstructive pulmonary disease. Patients with forced expiratory volume in one second (FEV1) and diffusion capacity of the lungs for carbon monoxide (DLCO) that are both >80% of predicted are considered fit for surgery, while those with decreased scores may benefit from further workup. Stair climbing, ambulatory stress tests (such as walking 400m on level ground with dyspnea and pulse oximetry monitoring) have been evaluated, and there is increasing evidence supporting the use of cardio-pulmonary exercise testing for further risk stratification [31]. While further studies on outcomes specific to ICI pneumonitis are lacking, it is prudent to follow these existing guidelines.

2.1.3. Cardiac

ICI myocarditis is rare, with an incidence of less than 0.1% [32]. While low, the mortality is significantly higher than other ICI toxicities, and in cancer centers with a high relative volume of immunotherapy patients, its prevalence is not to be ignored. Toxicity mechanisms are unclear, but likely result from lymphocytic infiltration of myocardial tissue in the setting of upregulated T-cell activity [33,34]. Pembrolizumab is the most common causative agent; the incidence is higher when combined with nivolumab. The average time to onset of cardiac symptoms is 1-2 months.

Cardiac toxicity can present as a conduction system disorder (ranging from prolonged PR interval to complete heart block), pericarditis, myocarditis, or cardiomyopathy with reduced systolic function. A useful triage of myocarditis in the presence of cancer therapeutics was developed by Circulation in 2019 [35], grouping patients into three categories: definite myocarditis, probable myocarditis, and possible myocarditis. A positive cardiac MRI plus one of either 1) an elevation in biomarkers, 2) an ECG pattern showing diffuse ST elevations, or 3) the clinical syndrome of myocarditis are required for the diagnosis.

Mild cases of ICI myocarditis are treated with oral prednisone. More severe cases may require intravenous methylprednisolone in high doses, which mimics the treatment strategy of viral myocarditis. A taper regimen will generally last 4-6 weeks. Treatment with intravenous

immunoglobulins, mycophenolate, infliximab, and plasmapheresis have been considered for severe cases, and remain under investigation [36].

In addition to steroids, it is recommended to follow the standard American College of Cardiology/American Heart Association guidelines for the treatment of heart failure: beta-blockade (weighed against the risk of exacerbating a concurrent conduction system abnormality), ACE-inhibition, lifestyle modifications, and avoidance of volume overload.

Perioperative considerations for ICI myocarditis include a thorough cardiac history and a recent EKG to evaluate for conduction system abnormalities, particularly with a history of pembrolizumab exposure. There have been several case reports of complete heart block noted on a baseline preoperative EKG among patients on checkpoint inhibitors planned for cancer resection surgery [37]. Patients with dyspnea on exertion, chest pain, or unexplained hemodynamic compromise should be evaluated with an EKG, cardiac biomarkers, and an echocardiogram. Suspicions of myocarditis should prompt evaluation with a cardiologist, and consideration for a cardiac MRI.

2.1.4. Endocrine

Pituitary dysfunction is a well-established category of ICI toxicity. The incidence of hypophysitis is quite significant (10-15%), with the highest incidence found in patients on dual-ICI therapy with ipilimumab and nivolumab. The most common biochemical disturbances are ACTH deficiency, hypogonadism, and hypothyroidism. For patients not detected biochemically, common presenting symptoms are headache and fatigue, but anorexia, nausea, dizziness, and visual changes have also been described. Pituitary dysfunction typically present 2-3 months after therapy initiation, but onset can be as late as one year into therapy. In contrast to other adverse ICI effects, hypopituitarism is often permanent.

Patients on ICI therapy have quarterly thyroid function studies as part of maintenance therapy. Clinical signs of hypothyroidism include headache and fatigue, and laboratory findings include low-normal free T4 and inappropriately low or normal TSH.

A substantial portion of patients on ICI therapy have reduced ACTH and cortisol. Retrospective studies of ICI therapy cohorts have reported a wide range of incidences of low hormone values, ranging from one quarter to one half of patients. Most are associated with CTLA-4 inhibitors, but PD-1 inhibitors will also cause isolated ACTH deficiency [38,39].

Low serum cortisol resulting from central adrenocortical suppression can result in hypoaldosteronism with hyponatremia and hyperkalemia. It should be noted that hyponatremia is an extremely common adverse event in patients on any chemotherapeutics, including as a result of cancer related SIADH [40].

Hypogonadism on ICI therapy has an incidence of 10-15% and presents with decreased libido, hot flashes, or infertility. Laboratory studies will reveal low testosterone and inappropriately normal luteinizing hormone (LH) [41].

MRI findings in patients with ICI hypophysitis demonstrate pituitary enlargement in slightly more than half of cases, however MRI findings do not necessarily correlate with clinical symptom burden [42,43], and patients with significantly reduced ACTH or cortisol may have normal MRI findings. Pathology specimens for patients with this condition indicate lymphocytic infiltration and increased CTLA-4 antigen expression on pituitary endocrine cells (i.e.: molecular mimicry) [44] as culprit pathology.

Hyperthyroidism has a lower incidence (3-8% [45]) and can result both from a centrally mediated increase in TRH from hypophysitis, or a transient, local thyroiditis from molecular mimicry. Patients with locally mediated ICI thyroiditis will most commonly present with laboratory abnormalities (low TSH, elevated T3, T4), or present with symptoms of hyperthyroidism. Thyroiditis, after it subsides, will most often progress to hypothyroidism.

The perioperative provider should have a high index of suspicion for adrenal insufficiency. There are no studies thus far on the incidence of intraoperative hemodynamic instability related to ICI therapy, but anecdotal evidence exists for patients on ICI therapy to require higher doses of intraoperative vasopressors. Patients on ICI therapy should have recent thyroid-function testing and

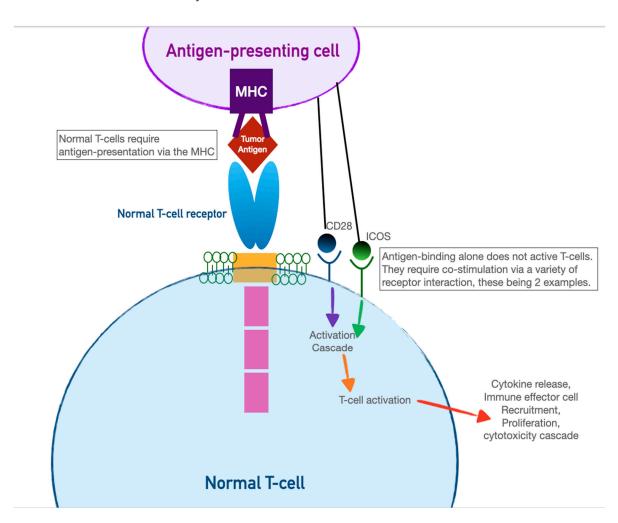
should be screened for headache and fatigue. If positive, it should be worth considering an underlying central adrenal insufficiency, and to treat empirically should hemodynamic instability occur. Hydration status should be optimized, and electrolytes should be checked pre and postoperatively for all patients on ICI therapy. Adrenally insufficient patients are more likely to decompensate in the setting of systemic infection, and benefit from early aggressive resuscitative strategy, as they have limited ability to compensate for physiologic stress.

Immune Checkpoint Inhibitor Toxicity Category	Manifestations	Treatment	Perioperative Considerations
Central Hypothyroidism	Fatigue, headache, low T4, low/normal TSH	Thyroid hormone replacement	Thyroid function tests within 3 months for all patients on ICI therapy
Hypogonadism	Hot flashes, decreased libido, decreased FSH and LH	Hormone replacement therapy	Screening for other signs of pituitary dysfunction
Pulmonary	Hypersensitivity pneumonitis, pulmonary fibrosis, organizing pneumonia, ARDS	Corticosteroids, supportive care.	Screening for respiratory symptoms, baseline CXR, pulmonary risk- stratification with baseline spirometry and cardio- pulmonary exercise testing.
Enterocolitis	Diarrhea, gastritis, hepatitis (manifests as occult rise in AST/ALT over first few months of therapy)	Low dose corticosteroids	Screening for reflux, gastritis symptoms. Recent LFTs if therapy was started within the last few months.
Pancreatic dysfunction	New-onset type 1 diabetes, ICI pancreatitis	Corticosteroids, resuscitative fluids; insulin therapy	Amylase and lipase levels for patients reporting abdominal pain, ruling out other causes of abdominal pain or pancreatitis; screening with preoperative blood glucose
Cardiac	Myocarditis, pericarditis, conduction system disorders.	Corticosteroids, standard AHA/ACC heart failure guidelines	Screening for symptoms of heart failure, and baseline EKGs for patients on ICI therapy, cardiac biomarkers and cardiac MRI for patients with symptoms of myocarditis

3. Chimeric Antigen-receptor T Cells (CAR-T)

CAR-T cells harness the adaptive immunity offered by our T-lymphocytes to target tumorspecific antigens. As the first foray into cellular therapeutics, they represent one of the most consequential discoveries in cancer therapy in the last two decades, owing to their relative ease of administration and robust efficacy against highly fatal hematologic malignancies. As discussed in the previous section, the immune system relies on T-cell activation via 1) antigen presentation via an MHC complex and 2) costimulation via other immune effector cells. Cancer cells have historically evaded this process of T-cell activation by thwarting antigen recognition. Enter CAR-T cells: genetically modified host T-cells that force this process to occur without the ability of tumor-cell evasion.

First, T-cells are extracted from the patient (or an allogeneic donor) and modified via a retroviral vector to express a new antigen receptor and co-stimulatory domains (see Figure 2). The term "chimeric" refers to the fact that this engineered receptor both *activates* and *co-stimulates* the T-cell [46]. The patient is then leukoreduced, to increase the relative proportion of CAR-T cells, and CAR-T cells are infused into the patient. The CAR-T cells will recognize antigens, activate, and recruit immune effector cells to destroy tumor cells.



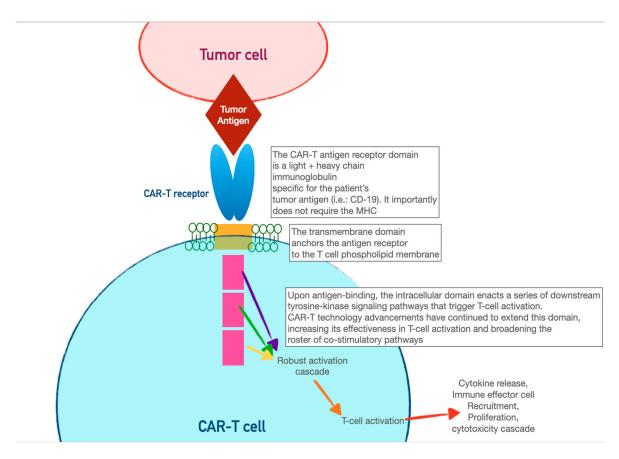


Figure 2. Above, normal T-cell activation requires antigen recognition and co-stimulation. Below, a re-engineered CAR-T cell enables simultaneous antigen recognition and co-stimulation, leading to T-cell mediated destruction of tumor cells.

The first CAR-T cells used clinically targeted the CD19 receptor, making them effective for the treatment of ALL or diffuse large B-cell lymphoma [47]. Trials have consistently shown a 50-90% remission rate for relapsed or refractory disease, which continues to improve with refinement of technique [48]. CAR-T for solid tumors is a growing area of research, where re-modeled CAR-T cells can be introduced intra-tumor or intra-pleural [49].

Once infused, CAR-T cell activity can be controlled via embedded biochemical switches. An "on" switch, or conversely, a "suicide gene" can induce CAR-T cell activation or apoptosis. These external controls allow CAR-T technology to be more pliable in the face of toxicities or other adverse events.

4. Bispecific T-cell Engager (BiTE) Therapy

BiTE therapy is one of the newest innovations in targeted immunomodulatory chemotherapy. BiTE molecules are an antibody with two domains (hence "bispecific"): one which recognizes tumor-specific antigens, and another which recognizes the universal CD-3 receptor on T cells. The binding sites are essentially two monoclonal antibodies bound together. In essence, the BiTE "forces" a recognition and activation of the T cell, so the tumor cell does not have a chance to present its own antigen and possibly release inhibitory signal receptors. Specific tumor antigen targets of BiTE molecules include CD19 (broadly expressed on B cell malignancies), B cell maturation antigen (expressed highly on malignant cells involved in multiple myeloma), CD33 (expressed in acute myeloid leukemia, myelodysplastic syndrome, and chronic myeloid leukemia). Under development are BiTE targets that include solid tumor antigens such as prostate-specific member antigen and delta-like protein 3 which is highly prevalent in small-cell lung cancer.

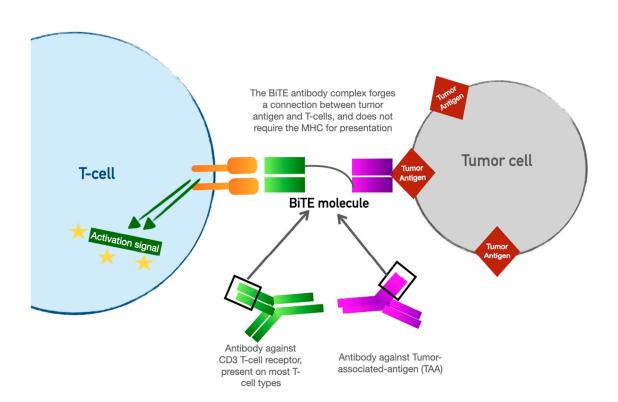
The most widely used BiTE agent at this time is Blinatumomab, approved for use in acute lymphoblastic leukemia (ALL). Trial response rates in children and adults with ALL have shown a

resounding 90% complete remission rate, with a 6-month event-free survival of 67%, and a 78% overall survival rate [50]. It is now approved for use around the globe. Solitumab is a BiTE molecule with a binding domain for epithelial cell adhesion molecule (EpCAM), which is being investigated for therapy against gastrointestinal, lung, and other solid tumors [51].

These molecules are administered as continuous IV infusion over several days, owing to their 2–4-hour half-life. Not unsurprisingly, owing to their similar mechanisms, they have an adverse event profile that parallels that of CAR-T cells. In the original study for Blinatumomab, adverse events most common were neutropenia, infection, elevated LFTs, neurotoxicity, and CRS [52]. CRS, neurotoxicity, and acute anaphylaxis are the most significant for the acute care and perioperative provider. There is perhaps a lower propensity than CAR-T for high-grade neurotoxicity [53]. Treatment for these adverse events follows the same protocol - corticosteroids, IL-6 inhibition, and supportive care.

5. Other Cellular Therapies

Several novel cellular therapies are under investigation: natural killer (NK) CAR-T cells, engineered T cell receptors, and tumor-infiltrated lymphocytes. The latter involves harvesting lymphocytes that have already successfully infiltrated tumor cells (isolated via biopsy or surgical resection), and proliferating and re-infusing them. The increasing armamentarium of cellular therapy agents warrants an understanding of their mechanisms and toxicities so the perioperative provider can provide safe and informed care.



6. Toxicity of Cellular Therapy and Perioperative Considerations

CAR-T recipients are monitored as inpatients for acute toxicity. Immediate post-infusion reactions include anaphylaxis and anaphylactoid reactions including transient vasodilation. The most clinically significant toxicities are sub-acute and include cytokine release syndrome (CRS) and neurotoxicity. Chief considerations for these conditions are summarized in Table 1.

Toxicity	Incidence	Symptoms	Mechanism	Evidence-based Treatments	Investigational treatments
Cytokine Release syndrome	Up to 50%	Vasodilation, confusion, fever, headache (see grading chart)	Systemic inflammation induced by mass-T- lymphocyte activation	Corticosteroids, Tocilizumab, time	Siltuximab, Anakinra
Neurotoxicity	3-5%	Encephalopathy, hypoactive delirium, lethargy, tremors, seizures (see ICANS scoring)	CNS infiltration of lymphocytes	Steroids, tocilizumab, anti- epileptics, time	Siltuximab, Anakinra

6.1. Cytokine Release Syndrome (CRS)

Around half of patients who have received CAR-T cells will develop some form of CRS [54]. This systemic inflammatory response results from en-masse activation of lymphocytes, and CAR-T induced cytokine release. Interleukin-6 (IL-6) is highly associated with CRS severity and can be useful when attempting to differentiate between CAR-T related CRS and CRS related to sepsis. CRS can occur within hours, but the peak onset-time is 4-7 days post-infusion.

CRS is grading occurs on a scale of 1-5. Grade 1 is defined as mild systemic symptoms such as low-grade fever, headache, and myalgias. Grade 2 is defined by the requirement of oxygen or pressor support. Grade 3 is multi-organ dysfunction requiring high-dose or multiple pressors. Grade 4 requires the presence of mechanical ventilation and advanced multi-organ dysfunction. Treatment for CRS is mostly supportive, and it is notable that with just time alone, patients will make a significant recovery.

Patients with CRS in the perioperative period require close collaboration with the oncology services. Nearly all patients with CRS will receive glucocorticoids to blunt the overactive immune response. Grade 3 and 4 CRS should be treated with tocilizumab. Originally approved as a disease modifying agent for rheumatoid arthritis, tocilizumab is a monoclonal antibody which prevents IL-6 from binding to its receptor [55]. It is administered on a three-times daily dosing schedule and is commonly combined with corticosteroids. Siltuximab is an anti-IL 6 monoclonal antibody that has shown safety and equivalence to Tocilizumab [56]. Siltuximab should be noted as a strong CYP-450 inducer. Anakinra is an IL-1 antagonist which has been explored as an off-label agent to mitigate CRS and CAR-T neurotoxicity [57,58]. It can also be used to treat hemophagocytic lymphohistiocytosis (HLH), a severe complication of CAR-T therapy in which lymphocyte over-activation leads to destruction of all cell lines, liver dysfunction, coagulopathy, fever, and rash [59].

It is important to note that none of the CRS-targeted therapies discussed here will significantly impact the efficacy of CAR-T cell therapy.

6.2. Neurotoxicity

3-5% of patients who receive CAR-T cells will develop neurotoxicity [60]. Commonly referred to as immune effector cell-associated neurotoxicity syndrome (ICANS), the etiology of this condition is multifactorial and not yet fully understood. A likely early contributor is the breakdown of cerebral endothelial integrity by increased circulating cytokines, like the pathophysiology underlying septic encephalopathy [61,62]. The typical timeframe for onset of ICANS is 5 days post-infusion, but can be delayed by up to 2 weeks. Neurologic symptoms that are specific to early ICANS are outlined in

Tables 2 and 3, and include word-finding difficulty, tremors, hypoactive delirium, lethargy, and impaired handwriting. Moderate ICANS can progress to hyperactive delirium, myoclonus, obtundation, and ataxia. Severe ICANS can result in neurologic catastrophe including seizures, coma, and death. The median duration of this syndrome is 4 days. Mortality is related to the degree of their resultant critical illness - ARDS, malnutrition, degree of shock, etc.

Neuroimaging and video-electroencephalography (vEEG) obtained from ICANS patients will demonstrate nonspecific patterns of cerebral edema, and generalized slowing, respectively. Lumbar puncture will reveal elevated lymphocytes in CSF, as well as the presence of CAR-T cells.

Time and supportive care are the mainstay of treatment. ICANS is less responsive to the typical IL-6-targeted therapies as compared to CRS. Corticosteroids are beneficial, but come with significant iatrogenic harm - immunosuppression, gastrointestinal bleeding, and increased delirium. There is some evidence showing seizure prophylaxis with leveliracetam to be useful for anyone with grade 2 or higher ICANS.

CRS and ICANS mortality are more closely linked to the degree of organ dysfunction and iatrogenic exposure (prolonged ICU stay, mechanical ventilation, pressor use, prolonged steroid use and immunosuppression, etc.) than the degree of the toxicity per se. With supportive care, a relatively high proportion of patients experience complete recovery [63,64]. The overall mortality of CAR-T cells remains low, at 5.4% [65].

6.3. Perioperative Considerations

The increasing efficacy and safety profile of cellular therapy has placed them at the forefront of cancer therapeutics at a rapidly expanding array of academic and non-academic centers. At this time, around 150 centers across the country offer CAR-T therapy, and the technology has spread across the globe to Canada, China, Australia, Singapore, Israel, and Europe. Evidence for specific perioperative treatment is lacking, as are studies on outcomes for emergent surgery done in the immediate post-CAR-T period.

It is crucial to involve the critical care team for patients with high-grade CRS or who are at risk for developing high-grade CRS, to enable high-quality supportive care for respiratory and hemodynamic compromise.

Perioperative care of the CAR-T cell patient has several important implications. Centers that offer CAR-T therapy are always equipped with services that are familiar with post-infusion monitoring, and downstream complications such as CRS and neurotoxicity. It is essential to involve a multi-disciplinary team of oncologists, anesthesiologists, and critical care physicians in perioperative decision making. For patients with uneventful CAR-T therapy who have had no signs of CRS, neurotoxicity, or infectious complications: usual care should be pursued for urgent or emergent surgery. Elective surgery should ideally be deferred until hematologic dyscrasias have normalized, typically by 3-4 weeks. A discussion with the oncology team about intraoperative steroid dosing should be considered. For patients within a week of CAR-T infusion who need urgent surgery, there should be a low threshold for pre-emptive postoperative ICU admission so that acute changes in hemodynamics and neurologic status will not be missed. Coagulation profiles should be obtained prior to neuraxial anesthesia, and if no thrombocytopenia or elevated INR is present, there is no contraindication to regional anesthesia.

The physiologic milieu of CRS will render patients sensitive to the vasodilatory effects of anesthetics. They should be treated as patients with severe sepsis, with slow dosing of induction agents along with supplemental vasopressor support. Widespread endothelial dysfunction will lead to increased capillary leak, which portends to intravascular depletion and increased third spacing. Fevers and prolonged NPO status (in the setting of CRS or neurotoxicity) will exacerbate this. Insensible losses will be higher, so fluid-sparing strategies should be used with caution. As with any patient experiencing compromised end-organ perfusion, attention should be paid to not further worsen tissue oxygenation. Hypoxemia and hypercarbia should be avoided. Sedation without a protected airway should be administered cautiously in the presence of metabolic acidosis, and there should be a low threshold to provide an endotracheal airway. The presence of severe acute kidney

injury (AKI) is uncommon in CRS, and most patients recover to their baseline renal function [66]. In such cases, dosing of paralytic and analgesics should be done with attention to GFR.

Neurotoxic patients should be treated as at-risk for having elevations in intracranial pressure. Caution should be taken to avoid exacerbations of ICP (hypercarbia, hypoxemia, trendelenburg position), normo-glycemia, normothermia, normal sodium homeostasis. Anticonvulsants should be continued. The ICANS scoring system should be used pre and immediately postoperative, so post-procedure neurologic changes can be evaluated [67].

7. Conclusion

Targeted cancer therapy is undergoing a seismic shift from one that exerts an imprecise attack on cell division to one that targets specific tumor antigens, engaging our innate immune system to assist in the process. With these more precise and efficient therapies comes an increasingly complex toxicity profile. Toxicities can occur chronically, and masquerade beneath other neoplastic and paraneoplastic conditions. These agents increasingly play a role in multimodal cancer care, which is significantly prolonging life expectancy and quality of life for patients with cancer. As the depth and breadth of cancer procedures increases, and as the footprint of these therapies increases, the perioperative provider plays a more central role in managing cancer therapies and drug toxicities. The above review serves to update the perioperative clinician on guidelines for screening, diagnosing, and treating toxicities associated with novel cellular therapies and immune-checkpoint therapies.

References

- Doki Y, Ajani JA, Kato K, Xu J, Wyrwicz L, Motoyama S, Ogata T, Kawakami H, Hsu CH, Adenis A, El Hajbi F, Di Bartolomeo M, Braghiroli MI, Holtved E, Ostoich SA, Kim HR, Ueno M, Mansoor W, Yang WC, Liu T, Bridgewater J, Makino T, Xynos I, Liu X, Lei M, Kondo K, Patel A, Gricar J, Chau I, Kitagawa Y; CheckMate 648 Trial Investigators. Nivolumab Combination Therapy in Advanced Esophageal Squamous-Cell Carcinoma. N Engl J Med. 2022 Feb 3;386(5):449-462. doi: 10.1056/NEJMoa2111380. PMID: 35108470.
- 2. Hodi FS, O'Day SJ, McDermott DF, Weber RW, Sosman JA, Haanen JB, Gonzalez R, Robert C, Schadendorf D, Hassel JC, Akerley W, van den Eertwegh AJ, Lutzky J, Lorigan P, Vaubel JM, Linette GP, Hogg D, Ottensmeier CH, Lebbé C, Peschel C, Quirt I, Clark JI, Wolchok JD, Weber JS, Tian J, Yellin MJ, Nichol GM, Hoos A, Urba WJ. Improved survival with ipilimumab in patients with metastatic melanoma. N Engl J Med. 2010 Aug 19;363(8):711-23. doi: 10.1056/NEJMoa1003466. Epub 2010 Jun 5. Erratum in: N Engl J Med. 2010 Sep 23;363(13):1290. PMID: 20525992; PMCID: PMC3549297.
- 3. "Opdivo- nivolumab injection". DailyMed. 17 December 2019. Retrieved 11 March 2020.
- 4. Hellmann, Matthew D.; Paz-Ares, Luis; Caro, Reyes Bernabe; Zurawski, Bogdan; Kim, Sang-We; Costa, Enric Carcereny; Park, Keunchil; Alexandru, Aurelia; Lupinacci, Lorena; Jimenez, Emmanuel de la Mora; Sakai, Hiroshi (2019-09-28). "Nivolumab plus Ipilimumab in Advanced Non–Small-Cell Lung Cancer". *New England Journal of Medicine*. **381** (21): 2020–2031.
- 5. Alyson Haslam, PhD; Jennifer Gill, MS; Vinay Prasad, MD, MPH Estimation of the Percentage of US Patients With Cancer Who Are Eligible for Immune Checkpoint Inhibitor Drugs Author Affiliations Article Information *JAMA Netw Open.* 2020;3(3):e200423. doi:10.1001/jamanetworkopen.2020.0423
- Elias AW, Kasi PM, Stauffer JA, Thiel DD, Colibaseanu DT, Mody K, Joseph RW, Bagaria SP. The Feasibility and Safety of Surgery in Patients Receiving Immune Checkpoint Inhibitors: A Retrospective Study. Front Oncol. 2017 Jun 12;7:121. doi: 10.3389/fonc.2017.00121. PMID: 28660171; PMCID: PMC5466999.
- 7. Cloughesy, T.F., Mochizuki, A.Y., Orpilla, J.R. *et al.* Neoadjuvant anti-PD-1 immunotherapy promotes a survival benefit with intratumoral and systemic immune responses in recurrent glioblastoma. *Nat Med* 25, 477–486 (2019). https://doi.org/10.1038/s41591-018-0337-7
- 8. Patel SP, Othus M, Chen Y, et al. Neoadjuvant–adjuvant or adjuvant-only pembrolizumab in advanced melanoma. N Engl J Med 2023;388:813-23. DOI: 10.1056/NEJMoa2211437.
- 9. Morad G, Helmink BA, Sharma P, Wargo JA. Hallmarks of response, resistance, and toxicity to immune checkpoint blockade. Cell. 2021 Oct 14;184(21):5309-5337. doi: 10.1016/j.cell.2021.09.020. Epub 2021 Oct 7. Erratum in: Cell. 2022 Feb 3;185(3):576. PMID: 34624224; PMCID: PMC8767569.
- 10. Schneider BJ, Naidoo J, Santomasso BD, Lacchetti C, Adkins S, Anadkat M, Atkins MB, Brassil KJ, Caterino JM, Chau I, Davies MJ, Ernstoff MS, Fecher L, Ghosh M, Jaiyesimi I, Mammen JS, Naing A, Nastoupil LJ, Phillips T, Porter LD, Reichner CA, Seigel C, Song JM, Spira A, Suarez-Almazor M, Swami U, Thompson JA, Vikas P, Wang Y, Weber JS, Funchain P, Bollin K. Management of Immune-Related Adverse Events in Patients Treated With Immune Checkpoint Inhibitor Therapy: ASCO Guideline Update. J Clin Oncol. 2021

- Dec 20;39(36):4073-4126. doi: 10.1200/JCO.21.01440. Epub 2021 Nov 1. Erratum in: J Clin Oncol. 2022 Jan 20;40(3):315. PMID: 34724392.
- 11. Tunio, Nahel A. MD; Desai, Aakash MD; Dalal, Shaman MD; Kurin, Michael MD; Waghray, Nisheet MD. S218 Prevalence and Outcomes of Immune Checkpoint Inhibitor (ICI) Associated Colitis: A Population-Based Cohort Study. The American Journal of Gastroenterology 117(10S):p e157, October 2022. | DOI: 10.14309/01.ajg.0000857512.69949.fc
- 12. Weber JS, Kahler KC, Hauschild A. Management of immune-related adverse events and kinetics of response with ipilimumab. *J Clin Oncol* 2012;30:2691-7. 10.1200/JCO.2012.41.6750
- 13. Wolchok JD, Chiarion-Sileni V, Gonzalez R, Rutkowski P, Grob JJ, Cowey CL, Lao CD, Wagstaff J, Schadendorf D, Ferrucci PF, Smylie M, Dummer R, Hill A, Hogg D, Haanen J, Carlino MS, Bechter O, Maio M, Marquez-Rodas I, Guidoboni M, McArthur G, Lebbé C, Ascierto PA, Long GV, Cebon J, Sosman J, Postow MA, Callahan MK, Walker D, Rollin L, Bhore R, Hodi FS, Larkin J. Overall Survival with Combined Nivolumab and Ipilimumab in Advanced Melanoma. N Engl J Med. 2017 Oct 5;377(14):1345-1356. doi: 10.1056/NEJMoa1709684. Epub 2017 Sep 11. Erratum in: N Engl J Med. 2018 Nov 29;379(22):2185. PMID: 28889792; PMCID: PMC5706778.
- 14. Omori G, Takada K, Murase K, Hayasaka N, Nakamura H, Iyama S, Ohnuma H, Miyanishi K, Fukuta F, Tanaka T, Masumori N, Kato J. Successful mycophenolate mofetil treatment of a patient with severe steroid-refractory hepatitis evoked by nivolumab plus ipilimumab treatment for relapsed bladder cancer. Clin Case Rep. 2020 Dec 4;9(2):654-659. doi: 10.1002/ccr3.3597. PMID: 33598220; PMCID: PMC7869331.
- 15. Mahmud N, Fricker Z, Hubbard RA, Ioannou GN, Lewis JD, Taddei TH, Rothstein KD, Serper M, Goldberg DS, Kaplan DE. Risk Prediction Models for Post-Operative Mortality in Patients With Cirrhosis. Hepatology. 2021 Jan;73(1):204-218. doi: 10.1002/hep.31558. Epub 2020 Dec 10. PMID: 32939786; PMCID: PMC7902392
- 16. George J, Bajaj D, Sankaramangalam K, Yoo JW, Joshi NS, Gettinger S, Price C, Farrell JJ. Incidence of pancreatitis with the use of immune checkpoint inhibitors (ICI) in advanced cancers: A systematic review and meta-analysis. Pancreatology. 2019 Jun;19(4):587-594. doi: 10.1016/j.pan.2019.04.015. Epub 2019 May 2. PMID: 31076344.
- 17. George, J., Yoo, J. W., Joshi, N., & Farrell, J. J. (2018). Mo1236 Incidence of acute pancreatitis with the use of immune checkpoint inhibitors (ICI) in solid tumors: A systematic review and meta-analysis. *Gastroenterology*, 154(6 suppl 1), S–714. 10.1016/S0016-5085(18)32506-X
- 18. Ikeuchi, K., Okuma, Y., & Tabata, T. (2016). Immune-related pancreatitis secondary to nivolumab in a patient with recurrent lung adenocarcinoma: A case report. *Lung Cancer*, 99, 148–150. 10.1016/j.lungcan.2016.07.001
- 19. National Comprehensive Cancer Network (2020). NCCN Clinical Practice Guidelines in Oncology: Management of immunotherapy-related toxicities. V1.2020.
- 20. Waller, A., Long, B., Koyfman, A., & Gottlieb, M. (2018). Acute pancreatitis: Updates for emergency clinicians. *Journal of Emergency Medicine*, 55(6), 769–779. 10.1016/j.jemermed.2018.08.009
- 21. Barroso-Sousa R, Barry WT, Garrido-Castro AC, Hodi FS, Min L, Krop IE, Tolaney SM. Incidence of Endocrine Dysfunction Following the Use of Different Immune Checkpoint Inhibitor Regimens: A Systematic Review and Meta-analysis. JAMA Oncol. 2018 Feb 1;4(2):173-182. doi: 10.1001/jamaoncol.2017.3064. PMID: 28973656; PMCID: PMC5838579.
- 22. Clotman K, Janssens K, Specenier P, Weets I, De Block CEM. Programmed Cell Death-1 Inhibitor-Induced Type 1 Diabetes Mellitus. J Clin Endocrinol Metab. 2018 Sep 1;103(9):3144-3154. doi: 10.1210/jc.2018-00728. PMID: 29955867.
- 23. Banavasi H, Kim S, Alkassis S, Daoud A, Laktineh A, Nagasaka M, Sukari A, Soubani AO. Immune Checkpoint Inhibitor-induced Pneumonitis: Incidence, Clinical Characteristics, and Outcomes. Hematol Oncol Stem Cell Ther. 2023 Jan 17;16(2):144-150. doi: 10.1016/j.hemonc.2021.09.005. PMID: 34688626.
- 24. Poulose V. Incidence of Immune Checkpoint Inhibitor-Related Pneumonitis in Lung Cancer. Chest. 2022 Mar;161(3):e196-e197. doi: 10.1016/j.chest.2021.10.041. PMID: 35256099.
- 25. Naidoo J, Wang X, Woo KM, et al.. Pneumonitis in patients treated with anti–programmed death-1/programmed death ligand 1 therapy. *JCO* 2017;35:709–17. 10.1200/JCO.2016.68.2005
- Naidoo J, Cottrell TR, Lipson EJ, Forde PM, Illei PB, Yarmus LB, Voong KR, Feller-Kopman D, Lee H, Riemer J, Wang D, Taube JM, Brahmer JR, Lin CT, Danoff SK, D'Alessio FR, Suresh K. Chronic immune checkpoint inhibitor pneumonitis. J Immunother Cancer. 2020 Jun;8(1):e000840. doi: 10.1136/jitc-2020-000840. Erratum in: J Immunother Cancer. 2020 Nov;8(2): PMID: 32554618; PMCID: PMC7304886.
- 27. Barrón F, Sánchez R, Arroyo-Hernández M, Blanco C, Zatarain-Barrón ZL, Catalán R, Ramos-Ramírez M, Cardona AF, Flores-Estrada D, Arrieta O. Risk of Developing Checkpoint Immune Pneumonitis and Its Effect on Overall Survival in Non-small Cell Lung Cancer Patients Previously Treated With Radiotherapy. Front Oncol. 2020 Sep 29;10:570233. doi: 10.3389/fonc.2020.570233. PMID: 33117699; PMCID: PMC7550759.
- 28. Schneider BJ, Naidoo J, Santomasso BD, Lacchetti C, Adkins S, Anadkat M, Atkins MB, Brassil KJ, Caterino JM, Chau I, Davies MJ, Ernstoff MS, Fecher L, Ghosh M, Jaiyesimi I, Mammen JS, Naing A, Nastoupil LJ,

- Phillips T, Porter LD, Reichner CA, Seigel C, Song JM, Spira A, Suarez-Almazor M, Swami U, Thompson JA, Vikas P, Wang Y, Weber JS, Funchain P, Bollin K. Management of Immune-Related Adverse Events in Patients Treated With Immune Checkpoint Inhibitor Therapy: ASCO Guideline Update. J Clin Oncol. 2021 Dec 20;39(36):4073-4126. doi: 10.1200/JCO.21.01440. Epub 2021 Nov 1. Erratum in: J Clin Oncol. 2022 Jan 20;40(3):315. PMID: 34724392.
- 29. Tiu BC, Zubiri L, Iheke J, *et al*Real-world incidence and impact of pneumonitis in patients with lung cancer treated with immune checkpoint inhibitors: a multi-institutional cohort study *Journal for ImmunoTherapy of Cancer* 2022;**10:**e004670. doi: 10.1136/jitc-2022-004670
- 30. Qiu B, Cai K, Chen C, Chen J, Chen KN, Chen QX, Cheng C, Dai TY, Fan J, Fan Z, Hu J, Hu WD, Huang YC, Jiang GN, Jiang J, Jiang T, Jiao WJ, Li HC, Li Q, Liao YD, Liu HX, Liu JF, Liu L, Liu Y, Long H, Luo QQ, Ma HT, Mao NQ, Pan XJ, Tan F, Tan LJ, Tian H, Wang D, Wang WX, Wei L, Wu N, Wu QC, Xiang J, Xu SD, Yang L, Zhang H, Zhang L, Zhang P, Zhang Y, Zhang Z, Zhu K, Zhu Y, Um SW, Oh IJ, Tomita Y, Watanabe S, Nakada T, Seki N, Hida T, Sasada S, Uchino J, Sugimura H, Dermime S, Cappuzzo F, Rizzo S, Cho WC, Crucitti P, Longo F, Lee KY, De Ruysscher D, Vanneste BGL, Furqan M, Sieren JC, Yendamuri S, Merrell KW, Molina JR, Metro G, Califano R, Bongiolatti S, Provencio M, Hofman P, Gao S, He J. Expert consensus on perioperative immunotherapy for local advanced non-small cell lung cancer. Transl Lung Cancer Res. 2021 Sep;10(9):3713-3736. doi: 10.21037/tlcr-21-634. PMID: 34733623; PMCID: PMC8512472.
- 31. Brunelli A, Charloux A, Bolliger CT, Rocco G, Sculier JP, Varela G, Licker M, Ferguson MK, Faivre-Finn C, Huber RM, Clini EM, Win T, De Ruysscher D, Goldman L; European Respiratory Society and European Society of Thoracic Surgeons joint task force on fitness for radical therapy. ERS/ESTS clinical guidelines on fitness for radical therapy in lung cancer patients (surgery and chemo-radiotherapy). Eur Respir J. 2009 Jul;34(1):17-41. doi: 10.1183/09031936.00184308. Erratum in: Eur Respir J. 2009 Sep;34(3):782. PMID: 19567600.
- 32. Lyon AR, Yousaf N, Battisti NML, Moslehi J, Larkin J. Immune checkpoint inhibitors and cardiovascular toxicity. Lancet Oncol. 2018 Sep;19(9):e447-e458. doi: 10.1016/S1470-2045(18)30457-1. PMID: 30191849.
- 33. Johnson DB, Balko JM, Compton ML, Chalkias S, Gorham J, Xu Y, Hicks M, Puzanov I, Alexander MR, Bloomer TL, Becker JR, Slosky DA, Phillips EJ, Pilkinton MA, Craig-Owens L, Kola N, Plautz G, Reshef DS, Deutsch JS, Deering RP, Olenchock BA, Lichtman AH, Roden DM, Seidman CE, Koralnik IJ, Seidman JG, Hoffman RD, Taube JM, Diaz LA, Anders RA, Sosman JA, Moslehi JJ. Fulminant myocarditis with combination immune checkpoint blockade.
- 34. Nishimura H, Okazaki T, Tanaka Y, Nakatani K, Hara M, Matsumori A, Sasayama S, Mizoguchi A, Hiai H, Minato N, Honjo T. Autoimmune dilated cardiomyopathy in PD-1 receptor-deficient mice. Science. 2001; 291:319–322. doi: 10.1126/science.291.5502.319.
- 35. Bonaca Marc P, Olenchock Benjamin A, Salem J-E, Wiviott Stephen D, Ederhy S, Cohen A, Stewart Garrick C, Choueiri Toni K, Di Carli M, Allenbach Y, Kumbhani Dharam J, Heinzerling L, Amiri-Kordestani L, Lyon Alexander R, Thavendiranathan P, Padera R, Lichtman A, Liu Peter P, Johnson Douglas B, Moslehi J. Myocarditis in the setting of cancer therapeutics. Circulation. 2019; 140:80–91.
- 36. Norwood TG, Westbrook BC, Johnson DB, Litovsky SH, Terry NL, McKee SB, Gertler AS, Moslehi JJ, Conry RM. Smoldering myocarditis following immune checkpoint blockade. J Immunother Cancer. 2017; 5:91
- 37. Vartanov A, Kalotra A, Varughese J, Gautam S, Kandel S, Hosmer W. Immunotherapy-associated complete heart block in a patient with NSCLC: A case report and literature review. Respir Med Case Rep. 2021 Mar 18;33:101390. doi: 10.1016/j.rmcr.2021.101390. PMID: 33786301; PMCID: PMC7994778.
- 38. Chang LS, Barroso-Sousa R, Tolaney SM, Hodi FS, Kaiser UB, Min L. Endocrine toxicity of cancer immunotherapy targeting immune checkpoints. *Endocr Rev.* 2019;40:17-65.
- 39. Cui K, Wang Z, Zhang Q, Zhang X. Immune checkpoint inhibitors and adrenal insufficiency: a large-sample case series study. Ann Transl Med. 2022 Mar;10(5):251. doi: 10.21037/atm-21-7006. PMID: 35402601; PMCID: PMC8987884.
- 40. Pelletier K, Škrtić M, Kitchlu A. Cancer therapy-induced hyponatremia: A case-illustrated review. *Journal of Onco-Nephrology*. 2021;5(1):70-78.
- 41. Özdemir BC. Immune checkpoint inhibitor-related hypogonadism and infertility: a neglected issue in immuno-oncology. J Immunother Cancer. 2021 Feb;9(2):e002220. doi: 10.1136/jitc-2020-002220. PMID: 33589529; PMCID: PMC7887354.
- 42. Jessel S, Weiss SA, Austin M, Mahajan A, Etts K, Zhang L, Aizenbud L, Perdigoto AL, Hurwitz M, Sznol M, Herold KC, Kluger HM. Immune Checkpoint Inhibitor-Induced Hypophysitis and Patterns of Loss of Pituitary Function. Front Oncol. 2022 Mar 8;12:836859. doi: 10.3389/fonc.2022.836859. PMID: 35350573; PMCID: PMC8958012.
- 43. Caturegli P, Newschaffer C, Olivi A, Pomper MG, Burger PC, Rose NR. Autoimmune hypophysitis. *Endocr Rev.* 2005;26:599-614.
- 44. Caturegli P, Di Dalmazi G, Lombardi M, Grosso F, Larman HB, Larman T, Taverna G, Cosottini M, Lupi I. Hypophysitis Secondary to Cytotoxic T-Lymphocyte-Associated Protein 4 Blockade: Insights into

- Pathogenesis from an Autopsy Series. Am J Pathol. 2016 Dec;186(12):3225-3235. doi: 10.1016/j.ajpath.2016.08.020. Epub 2016 Oct 15. PMID: 27750046; PMCID: PMC5225294.
- 45. El Sabbagh R, Azar NS, Eid AA, Azar ST. Thyroid Dysfunctions Due to Immune Checkpoint Inhibitors: A Review. Int J Gen Med. 2020 Nov 4;13:1003-1009. doi: 10.2147/IJGM.S261433. PMID: 33177863; PMCID: PMC7650809.
- 46. Zhang H, Zhao P, Huang H (December 2020). "Engineering better chimeric antigen receptor T cells". Experimental Hematology & Oncology. 9 (1): 34. doi:10.1186/s40164-020-00190-2. PMC 7709221. PMID 33292660.
- 47. Kochenderfer JN, Wilson WH, Janik JE, Dudley ME, Stetler-Stevenson M, Feldman SA, et al. (November 2010). "Eradication of B-lineage cells and regression of lymphoma in a patient treated with autologous T cells genetically engineered to recognize CD19". *Blood.* 116 (20): 4099–4102. doi:10.1182/blood-2010-04-281931. PMC 2993617. PMID 20668228.
- 48. Xian Zhang, Xin-an Lu, Junfang Yang, Gailing Zhang, Jingjing Li, Lisong Song, Yunchao Su, Yanze Shi, Min Zhang, Jiujiang He, Dan Song, Fanyong Lv, Wenqian Li, Yan Wu, Hui Wang, Hongxing Liu, Xiaosu Zhou, Ting He, Peihua Lu; Efficacy and safety of anti-CD19 CAR T-cell therapy in 110 patients with B-cell acute lymphoblastic leukemia with high-risk features. *Blood Adv* 2020; 4 (10): 2325–2338. doi: https://doi.org/10.1182/bloodadvances.2020001466
- 49. Kyte JA. Strategies for Improving the Efficacy of CAR T Cells in Solid Cancers. Cancers (Basel). 2022 Jan 23;14(3):571. doi: 10.3390/cancers14030571. PMID: 35158839; PMCID: PMC8833730.
- 50. Maude SL, Frey N, Shaw PA, et al. Chimeric antigen receptor T cells for sustained remissions in leukemia. *N Engl J Med*. 2014;371:1507-1517.
- 51. Helwick C (June 2008). "Novel BiTE antibody mediates contact between T cells and cancer cells". *Oncology NEWS International.* **17** (6).
- 52. Dombret H, Topp MS, Schuh AC, Wei AH, Durrant S, Bacon CL, Tran Q, Zimmerman Z, Kantarjian H. Blinatumomab versus chemotherapy in first salvage or in later salvage for B-cell precursor acute lymphoblastic leukemia. Leuk Lymphoma. 2019 Sep;60(9):2214-2222. doi: 10.1080/10428194.2019.1576872. Epub 2019 Apr 5. PMID: 30947585.
- 53. Diefenbach C, Assouline S, Bosch F, et al. An individualized risk mitigation approach for safety: experience from the mosunetuzumab (CD20/CD3 bispecific antibody) development program in relation to neurotoxicity risk [abstract]. *Blood*. 2019;134(suppl 1):4728.
- 54. Lee DW, Santomasso BD, Locke FL, Ghobadi A, Turtle CJ, Brudno JN, et al. ASTCT Consensus Grading for Cytokine Release Syndrome and Neurologic Toxicity Associated with Immune Effector Cells. *Biol Blood Marrow Transplant* (2019) 25(4):625–38. doi: 10.1016/j.bbmt.2018.12.758
- 55. Si S, Teachey DT. Spotlight on Tocilizumab in the Treatment of CAR-T-Cell-Induced Cytokine Release Syndrome: Clinical Evidence to Date. Ther Clin Risk Manag. 2020 Aug 4;16:705-714. doi: 10.2147/TCRM.S223468. PMID: 32801727; PMCID: PMC7414980.
- 56. Seema Patel, Danielle Cenin, Donna Corrigan, Betty K. Hamilton, Matt Kalaycio, Ronald M. Sobecks, Faiz Anwer, Jack Khouri, Robert M. Dean, Allison Winter, Deepa Jagadeesh, Brad Pohlman, Brian T. Hill, Craig S. Sauter, Paolo F. Caimi; Siltuximab for First-Line Treatment of Cytokine Release Syndrome: A Response to the National Shortage of Tocilizumab. *Blood* 2022; 140 (Supplement 1): 5073–5074. doi: https://doi.org/10.1182/blood-2022-169809
- 57. Wehrli M, Gallagher K, Chen Y, *et al* Single-center experience using anakinra for steroid-refractory immune effector cell-associated neurotoxicity syndrome (ICANS) *Journal for ImmunoTherapy of Cancer* 2022;**10**:e003847. doi: 10.1136/jitc-2021-003847
- 58. Wehrli M, Gallagher K, Chen YB, Leick MB, McAfee SL, El-Jawahri AR, DeFilipp Z, Horick N, O'Donnell P, Spitzer T, Dey B, Cook D, Trailor M, Lindell K, Maus MV, Frigault MJ. Single-center experience using anakinra for steroid-refractory immune effector cell-associated neurotoxicity syndrome (ICANS). J Immunother Cancer. 2022 Jan;10(1):e003847. doi: 10.1136/jitc-2021-003847. PMID: 34996813; PMCID: PMC8744112.
- 59. Daver N, McClain K, Allen CE, Parikh SA, Otrock Z, Rojas-Hernandez C, Blechacz B, Wang S, Minkov M, Jordan MB, La Rosée P, Kantarjian HM. A consensus review on malignancy-associated hemophagocytic lymphohistiocytosis in adults. Cancer. 2017 Sep 1;123(17):3229-3240. doi: 10.1002/cncr.30826. Epub 2017 Jun 16. PMID: 28621800; PMCID: PMC5568927.
- 60. Gust J, Hay KA, Hanafi LA, et al. Endothelial activation and blood-brain barrier disruption in neurotoxicity after adoptive immunotherapy with CD19 CAR-T cells. *Cancer Discov.* 2017;7(12):1404–1419. doi: 10.1158/2159-8290.CD-17-0698
- 61. Ziaja M. Septic encephalopathy. Curr Neurol Neurosci Rep. 2013 Oct;13(10):383. doi: 10.1007/s11910-013-0383-y. PMID: 23954971; PMCID: PMC3779311.
- 62. Gust J, Hay KA, Hanafi LA, et al. Endothelial Activation and Blood-Brain Barrier Disruption in Neurotoxicity after Adoptive Immunotherapy with CD19 CAR-T Cells. Cancer Discov 2017;7:1404–19.

- 63. Cohen, A.D., Parekh, S., Santomasso, B.D. *et al.* Incidence and management of CAR-T neurotoxicity in patients with multiple myeloma treated with ciltacabtagene autoleucel in CARTITUDE studies. *Blood Cancer J.* 12, 32 (2022). https://doi.org/10.1038/s41408-022-00629-1
- 64. Shah NN, Highfill SL, Shalabi H, Yates B, Jin J, Wolters PL et al. CD4/CD8 T-Cell Selection Affects Chimeric Antigen Receptor (CAR) T-Cell Potency and Toxicity: Updated Results From a Phase I Anti-CD22 Car T-Cell Trial. *J Clin Oncol* 2020; : JCO.19.03279.
- 65. Cai C, Tang D, Han Y, Shen E, Abdihamid O, Guo C, Shen H, Zeng S. A comprehensive analysis of the fatal toxic effects associated with CD19 CAR-T cell therapy. Aging (Albany NY). 2020 Sep 24;12(18):18741-18753. doi: 10.18632/aging.104058. Epub 2020 Sep 24. PMID: 32973124; PMCID: PMC7585129.
- Gutgarts V, Jain T, Zheng J, Maloy MA, Ruiz JD, Pennisi M, Jaimes EA, Perales MA, Sathick J. Acute Kidney Injury after CAR-T Cell Therapy: Low Incidence and Rapid Recovery. Biol Blood Marrow Transplant. 2020 Jun;26(6):1071-1076. doi: 10.1016/j.bbmt.2020.02.012. Epub 2020 Feb 20. PMID: 32088364; PMCID: PMC8375362.
- 67. Lee DW, Santomasso BD, Locke FL, Ghobadi A, Turtle CJ, Brudno JN, Maus MV, Park JH, Mead E, Pavletic S, Go WY, Eldjerou L, Gardner RA, Frey N, Curran KJ, Peggs K, Pasquini M, DiPersio JF, van den Brink MRM, Komanduri KV, Grupp SA, Neelapu SS. ASTCT Consensus Grading for Cytokine Release Syndrome and Neurologic Toxicity Associated with Immune Effector Cells. Biol Blood Marrow Transplant. 2019 Apr;25(4):625-638. doi: 10.1016/j.bbmt.2018.12.758. Epub 2018 Dec 25. PMID: 30592986.

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.