

Review

Heterocyclic Iminoquinones and Quinones from National Cancer Institute (NCI, USA) COMPARE Analysis

Naemah Haji ^{1,†}, Masoma Faizi ^{1,†}, Panayiotis A. Koutentis ², Michael P. Carty ³ and Fawaz Aldabbagh ^{1,*}

¹ Department of Pharmacy, School of Life Sciences, Pharmacy and Chemistry, Kingston University, Penrhyn Road, Kingston upon Thames, KT1 2EE, UK; naemah.haji@gmail.com (N.H.); faizi.masoma2@gmail.com (M.F.)

² Department of Chemistry, University of Cyprus, PO Box 20537, 1678 Nicosia, Cyprus; koutenti@ucy.ac.cy

³ School of Biological and Chemical Sciences, University of Galway, University Road, H91 TK33 Galway, Ireland; michael.carty@universityofgalway.ie

* Correspondence: F.Aldabbagh@kingston.ac.uk

† These authors contributed equally to this work.

Abstract: This review uses the National Cancer Institute (NCI) COMPARE program to establish an extensive list of heterocyclic iminoquinones and quinones with similarities in differential growth inhibition across the 60-cell line panel of the NCI Developmental Therapeutic Program (DTP). Many natural products and synthetic analogues are revealed, as potential NAD(P)H:quinone oxidoreductase 1 (NQO1) substrates through correlations to dipyridoimidazo[5,4-*f*]benzimidazoleiminoquinone (DPIQ), and as potential thioredoxin reductase (TrxR) inhibitors, through correlations to benzo[1,2,4]triazin-7-ones and pleurotin. The strong correlation to NQO1 infers the enzyme has a major influence on the amount of active compound with benzo[*e*]perimidines, phenoxazinones, benz[*f*]pyrido[1,2-*a*]indole-6,11-quinones, seriniquinones, kalasinamide, indolequinones, and furano[2,3-*b*]naphthoquinones, hypothesized as prodrugs. Compounds with very strong correlations to known TrxR inhibitors had inverse correlations to the expression of both reductase enzymes, NQO1 and TrxR, including naphtho[2,3-*b*][1,4]oxazepane-6,11-diones, benzo[*a*]carbazole-1,4-diones, pyranonaphthoquinones (including kalafungin, nanaomycin A, and analogues of griseusin A), and discorhabdin C. Quinoline-5,8-dione scaffolds based on streptonigrin and laven-damycin can correlate to either reductase. Inhibitors of TrxR are not necessarily (imino)quinones, *e.g.*, parthenolides, while oxidizing moieties are essential for correlations to NQO1, as with the mitosenes. Herein, an overview of synthetic methods and biological activity of each family of heterocyclic imino(quinone) is provided.

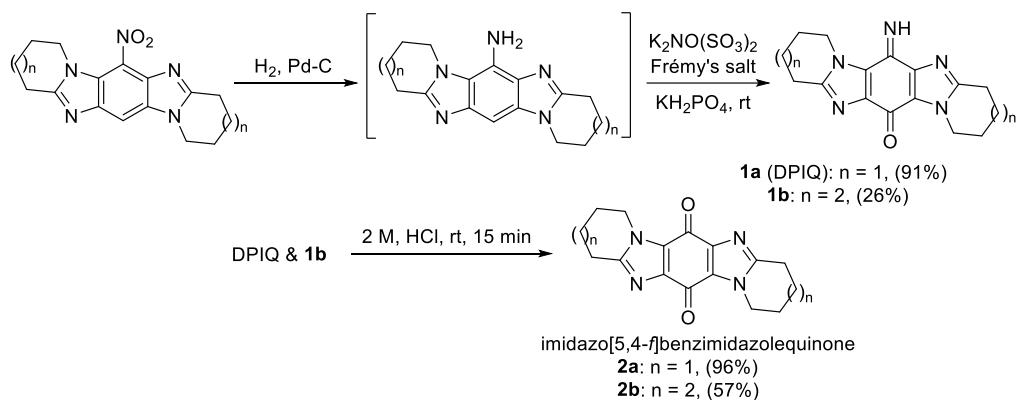
Keywords: bioreduction; heterocycles; natural products; NQO1; reductases; synthesis; thioredoxin.

1. Introduction

The National Cancer Institute (NCI, USA) Developmental Therapeutic Program (DTP) has supported the development of numerous clinical anticancer agents, including synthetic and natural compounds, vaccines, and antibodies [1-4]. The NCI human cancer 60-cell line panel consists of the nine major histological tissue types allowing high throughput screening of thousands of compounds and natural product extracts each month using the same assay under strictly identical conditions. Compounds showing high toxicity and variable patterns at an initial single dose (10 μ M) may be selected for five dose *in vitro* testing. The *in vitro* mean growth (inhibition) data against each cell line represents a pattern or “fingerprint” for the evaluated compound (the seed). The seed is identified using a designated NCI accession number (the Cancer Chemotherapy National Service Centre number, NSC number). Paull *et al.* transformed numerical cell line response data into the mean graph format of

visualizing differential growth inhibition [5]. After five dose testing, differential growth inhibition is depicted on log scale by bars (in delta units), which project either side of the mean (e.g., Figures S1-S82, in Supplementary Materials). The COMPARE algorithm is used to rank, in order of similarity to the seed, the activity of compounds in the huge NCI-DTP database, as well as the similarity of activity of the seed to expression of key cancer molecular targets in the panel of cell lines [1-3,5]. A molecular target is a protein, enzyme, gene, or any other cellular molecule whose presence within the 60-cell line panel has been identified and quantified by the NCI. For some test compounds, a single cellular component may determine activity, while in most cases cell sensitivity is complex and determined by gene expression, cell signaling, and repair pathways. The similarity of anti-cancer activity patterns to the seed is expressed quantitatively as a Pearson correlation coefficient (PCC). PCCs are between -1 and +1, with -1 indicating a perfect inverse correlation, zero indicating no correlation, and +1 indicating a perfect direct correlation. PCC of 0.3-0.5 is generally accepted as weak to moderate, 0.5-0.7 as being moderate to strong and above 0.7 as strong to very strong [1,5,6]. The toxicity evaluation service and COMPARE is available to the public free of charge from the NCI website (<https://dtp.cancer.gov/>), including as a new technology platform. Herein, we continue to assess similarities using the 50% growth inhibition (GI_{50}) parameter. GI_{50} is defined as the concentration that prevents half the proliferation from baseline compared to the maximal value of the untreated cells. Strong correlations may indicate similarities in the mechanisms of action of anti-cancer compounds, as well as structure-activity relationships. Two previous drug discovery investigations are revisited, that established two diverse heterocyclic iminoquinone scaffolds as potent anti-cancer agents [7-11]. Among the tools utilized to ascertain molecular targets was the NCI COMPARE program [7,10,11], which now reveals a more extensive list of correlated compounds. Other high throughput compound screening programs are not utilized in this review [3,4].

Aldabbagh and co-workers introduced ring-fused imidazo[5,4-*f*]benzimidazolequinones, *e.g.*, 2a and 2b, with iminoquinones 1a and 1b isolated from the Frémy oxidation of the amine intermediate in the presence of KH_2PO_4 acidic buffer (Scheme 1) [7,9].



Scheme 1. Iminoquinones via the synthesis of imidazo[5,4-*f*]benzimidazolequinones [7,9].

The dipyrido-fused iminoquinone 1a (DPIQ) was isolated in high yield (91%). DPIQ exhibited significant and variable cell growth inhibitory activity against the NCI-60 cell lines and was selected for five-dose testing [7]. The seven-membered (azepino) analogue 1b is inactive, and the hydrolyzed quinones 2a and 2b were less potent than DPIQ [7]. Notably, the isomeric iminoquinone of DPIQ, the imidazo[4,5-*f*]benzimidazole also exhibited low toxicity at the NCI [12]. COMPARE analysis gave a moderate correlation for DPIQ ($PCC = 0.51$) to NAD(P)H:quinone oxidoreductase 1 (NQO1, formerly known as DT-diaphorase) [7], an enzyme with heterogeneity of expression across the NCI-60 panel (Figure 1) [13]. COMPARE analysis of DPIQ against the huge library of synthetic compounds in the NCI-DTP database gave strong correlations in anti-cancer activity to two other heterocyclic iminoquinones 3a ($PCC = 0.87$) and 4 (0.77), indicating the compounds possibly possess similar mechanisms of action. Iminoquinone 3a ($PCC = 0.64$) gave a stronger correlation than DPIQ and 4 (0.47) to NQO1

expression in the NCI-DTP 60 cell lines. The higher PCC for 3a was supported by computation docking at the NQO1 active site, which predicted a higher affinity and a shorter distance for hydride reduction from the isoalloxazine ring of FADH₂ [7].

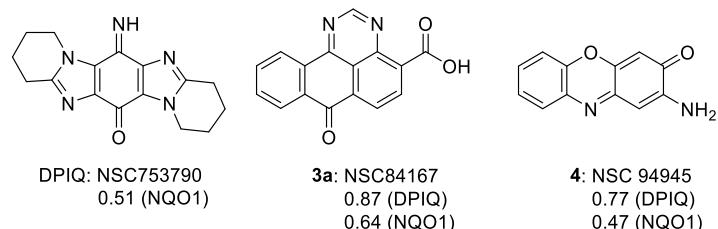


Figure 1. 6-Imino-1,2,3,4,8,9,10,11-octahydropyrido[1,2-a]pyrido[1',2':1,2]imidazo[5,4-f]benzimidazole-13-one (DPIQ), 3a and 4, and PCCs from COMPARE analysis [7].

A collaboration between the groups of Aldabbagh and Koutentis led to the discovery of anti-cancer activity for the benzo[e][1,2,4]triazin-7-ones, *e.g.*, 5a-d, (Figure 2) based on inhibition of the growth of cancer cell lines [10,11]. An earlier report demonstrated potency for 6-substituted 1,3-di-phenylbenzo[1,2,4]triazinones and 5-substituted [1,2,4]triazino[5,6,1-*j*k]carbazol-6-one derivatives 6, as multi-target inhibitors for Alzheimer's disease (AD) [14].

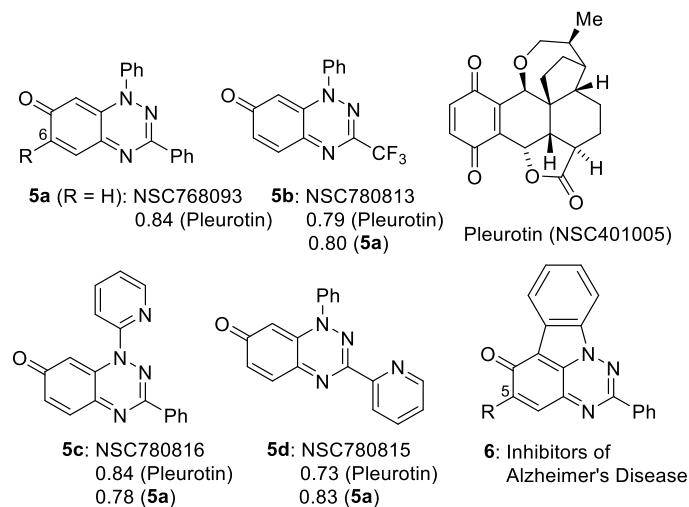
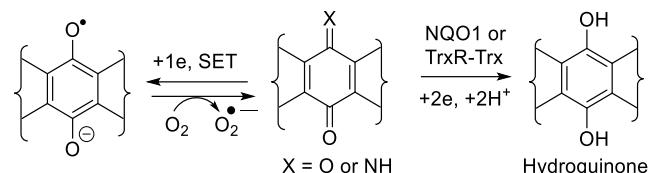


Figure 2. Benzo[1,2,4]triazin-7-ones 5a-d and PCCs to pleurotin [10,11], and anti-AD [1,2,4]triazino[5,6,1-*j*k]carbazol-6-one 6 [14].

The anti-cancer evaluation studies were stimulated by our interest in the iminoquinone motif, which is common to both DPIQ and the benzo[1,2,4]triazinones (Figures 1 and 2). The benzotriazinones 5a-d showed sufficient potency for selection by the NCI for five dose testing. The five-dose testing gave the GI₅₀ parameter used in COMPARE analysis. For 5a-d, the PCCs of ~0.8 indicates close to perfect direct correlations to the naturally occurring saturated benzoquinone, pleurotin [10,11]. Pleurotin possesses antibiotic and anti-cancer activity, including inhibiting hypoxia-induced factor 1α (HIF-1α) [15]. Pleurotin is a potent irreversible inhibitor of thioredoxin reductase (TrxR), and compounds 5a and 5b are reversible inhibitors with *K*_i values of 3.90 and 0.78 μM, respectively [10]. The greater enzyme inhibition by 5b was assumed to be related to the highly electron-withdrawing 3-CF₃ group increasing susceptibility to reductive activation and thereby increasing specificity towards cancer cell lines.

NQO1 and TrxR are obligatory 2-electron reductases, involved in regulating the production of toxic reactive oxygen species (ROS), including superoxide (O₂[•]) (Scheme 2). Although O₂[•] is used in cells of the immune system and provides defense against pathogens, excessive amounts are implicated in many physiological disorders, including cancer [16]. The 2-electron reduction overrides the single electron transfer (SET) by detoxifying the quinone, through the formation of a relatively stable

aromatic hydroquinone, which is eliminated through conjugation to glutathione, sulfate, or glucose [17].



Scheme 2. General mechanism for NQO1 and TrxR-Trx quinone detoxification.

NQO1 and TrxR, however, are over-expressed in many solid tumors and thus are viable molecular targets for cancer therapy [18,19]. In this context, the heterocyclic (imino)quinone is designed as a prodrug to form cytotoxic hydroquinone. For instance, in the case of the archetypical bioreductive quinone prodrug, mitomycin C, cytotoxicity occurs through quinone methide formation at C-1 and C-10, which react with nucleophilic DNA bases [20] (Figure 3). However, in the case of heterocyclic quinones and iminoquinones, such as DPIQ, which lack obvious reactive sites upon reduction, cell death is likely attributed to ROS formation with simultaneous reoxidation of the hydroquinone to the quinone, as observed in deoxynyboquinone (Fig. 3) [21].

Similar to the NQO1 reductive activation mechanism [22], NAD(P)H provides hydride reducing equivalents for transfer via a cascade in the TrxR-Trx (thioredoxin protein) system to the (imino)quinone [19]. Consequently, inhibition of TrxR leads to the accumulation of Trx protein and the activation of ROS-induced apoptosis pathways [16].

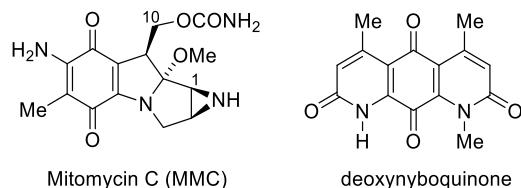


Figure 3. Bioreductive anti-cancer agents.

We now employ NCI COMPARE analysis to examine heterocyclic quinones and iminoquinones, specifically focusing on their strong correlations with DPIQ, benzo[1,2,4]triazinone 5a, and pleurotin. The objective of this analysis is to determine whether these correlations in anti-cancer activity patterns reflect actual similarities in mechanisms of action and structure-activity relationships. In addition, to assessing the biological activities, we provide descriptions of selected syntheses utilized to obtain each heterocyclic iminoquinone and quinone scaffold. We believe that this review represents the first compilation of heterocyclic compounds derived from COMPARE analysis.

2. Compound Search Methods

Our previous COMPARE analyses were repeated [6,7,10,11,23], and new analyses were carried out under identical conditions to allow comparisons of Pearson correlation coefficients (PCCs). Public COMPARE (https://dtp.cancer.gov/public_compare/) was used to access NCI-60 data, which did not include one dose experiments. The mean graphs generated are listed along with the PCCs in the Supplementary Materials document (Figures S1-S82). Scifinderⁿ (<https://scifinder-n.cas.org>) was used to find relevant literature using chemical structure and citation searches with, where applicable, publication data refinements.

2.1. COMPARE using DPIQ and 5a as the Seed.

The seed NSC numbers for DPIQ (NCS753790) and benzo[1,2,4]triazinone **5a** (NCS768093) were used to search using the COMPARE algorithm screening data limited to GI₅₀ end points. The mean graph data was subjected to a standard COMPARE using GI₅₀ of the synthetic compound's as the

target set with compounds ranked according to the magnitude of the PCCs. The process was repeated using pleurotin (NSC401005) as the seed to confirm similarities in PCCs with **5a**.

2.2. COMPARE using Molecular Target Expression.

COMPARE analysis was carried out for selected compound NSC numbers against molecular target expression in the NCI-60. NAD(P)H:quinone oxidoreductase 1 (NQO1) used the MT22 MOLTID.MT.SERIES and thioredoxin reductase (TrxR) used the MT143 TXNRD1 MOLTID.MT.SERIES. A PCC was obtained using a matrix COMPARE by appending the GI₅₀ cell viability profile of a given NSC number against an appended molecular target expression across the NCI-60.

The strongest correlations to NQO1 (MT22 MOLTID.MT.SERIES) were found by submitting the molecular target expression data to a standard COMPARE using screening data limited to GI₅₀ end points. Compounds were listed according to the magnitude of the PCCs.

3. Discussion

3.1. COMPARE Analysis: Strong Correlations to DPIQ as the Seed.

This section reviews compounds **7-12** with strong to very strong correlations (PCC = 0.72-0.87) in anti-cancer activity to DPIQ (Figure 4). COMPARE gave planar fused heterocyclic compounds containing the oxidizing quinone or iminoquinone motif, apart from azaanthracenone **11b**, which is the *para*-dimethoxybenzene (bio)synthetic precursor (see Section 3.1.5).

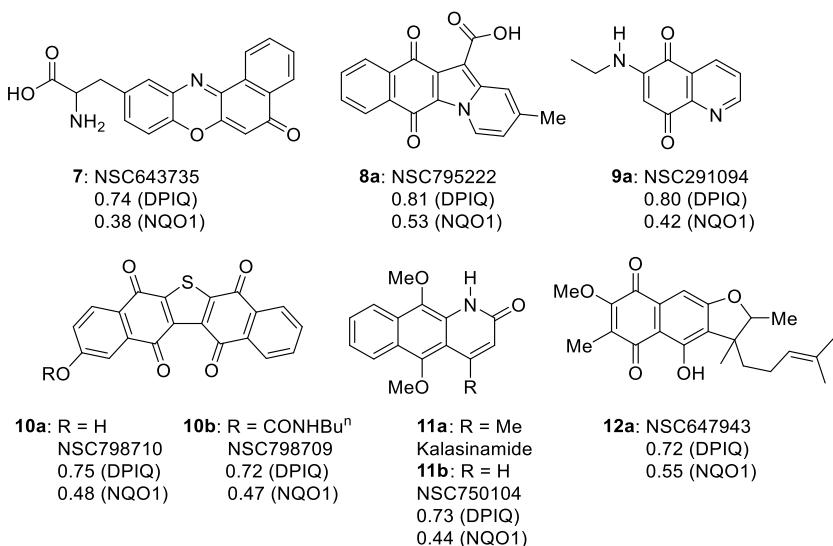


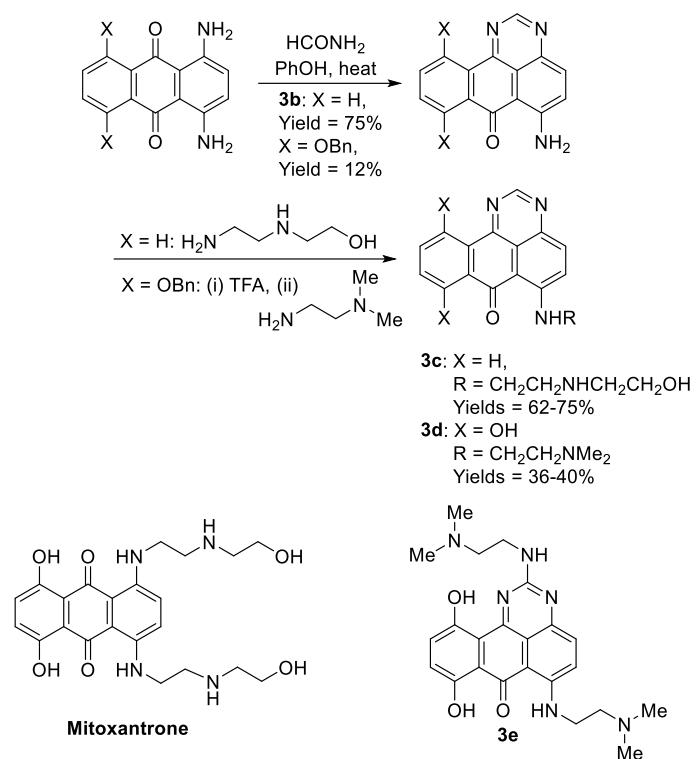
Figure 4. The strongest COMPARE correlations to DPIQ, excluding **3a** and **4**.

Weak to moderate correlations (PCC = 0.38-0.64) to NQO1 expression across the NCI-60 cell line panel were demonstrated, although benzo[e]perimidine-4-carboxylic acid **3a** is found to have a stronger correlation to NQO1 (PCC = 0.64, Figure 1). The following sub-sections review each heterocyclic scaffold, apart from quinoline-5,8-dione **9a**, since quinoline-5,8-diones also have strong correlations to the benzo[1,2,4]triazinone **5a** (reviewed in Section 3.3).

3.1.1. Benzo[e]perimidines

Commercially available benzo[e]perimidine-4-carboxylic acid **3a** displays the greatest similarity in anti-cancer activity to DPIQ with a PCC of 0.87 and a moderate to strong PCC of 0.64 to NQO1 expression across the NCI-60 cell line panel (Figure 1). Recently, iminoquinone **3a** was shown to induce apoptosis in NCI cell lines with high nuclear factor erythroid 2-related factor 2 (NRF2) activation [24]. NRF2 is a transcription factor over-expressed in pancreatic adenocarcinomas, which regulates expression of many redox enzymes, including NQO1 [25]. Compound **3a** induced apoptosis in *ex vivo*

cultures of pancreatic cancer xenografts with high NQO1/NRF2 activation, and inhibited the biosynthesis of amino acids, including asparagine and methionine [24]. The benzo[e]perimidine scaffold is obtained using the condensation of acetamide with 1,4-diamino-9,10-anthracenedione in molten phenol, with transamination providing 6-[(aminoalkyl)amino]-substituted examples **3c** and **3d** (Scheme 3) [26]. Substitutions mimic the well-known clinical anti-cancer agent, mitoxantrone, originally used to treat leukemia. Borowski and co-workers demonstrated the requirement of the 6-[(aminoalkyl)amino]-substituent in antileukemic activity, *e.g.*, **3c**, with the synthetic precursor **3b** inactive [26], and 8,11-dihydroxybenzo[e]perimidin-7-one **3d** displaying significant toxicity against NCI leukemic and solid tumor cell lines [27]. Compound **3d** displayed *in vivo* toxicity and overcomes multi-drug resistance tumor cells and like mitoxantrone induces cell cycle accumulation in the G2/M phase. The same group evaluated guanidine condensation adducts, with **3e** displaying the greatest *in vitro* toxicity [28].



Scheme 3. Synthesis and anti-cancer evaluation of benzo[e]perimidines **3b-e** [26-28].

3.1.2. Phenoxazinones

DPIQ displays a strong similarity in anti-cancer activity to the extensively studied and abundant natural chromophore, 2-aminophenoxazin-3-one **4** (PCC = 0.77, Figure 1), also known as the antibiotic, questrimycin A [29-33]. Iminoquinone **4** is isolated from the 6-electron oxidative coupling of two molecules of 2-aminophenol [33,34], mediated by the enzyme, phenoxazinone synthase [35]. The antibiotic has a weak to moderate correlation to NQO1 expression (PCC = 0.47) [7]. Anti-cancer activity of **4** includes cytotoxicity against a variety of human cancer cells lines with induction of apoptosis [31,36,37], and suppression of NO and prostaglandin E2 production leading to anti-inflammatory effects [38]. Cancer spread was monitored *in vivo* in mice, where the number of pulmonary metastasis of B6 melanoma cells remained the same [37]. The first reported antibiotic found to have anti-cancer activity is actinomycin D [39], containing 2-aminophenoxazin-3-one **4** at its core (Figure 5) [34]. Actinomycin D (also known as dactinomycin) has the trade name Cosmegen and is clinically used to treat a variety of solid tumors.

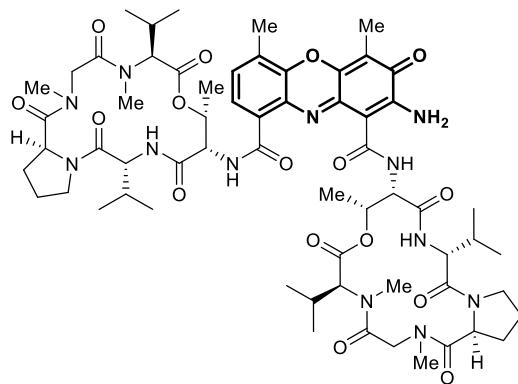


Figure 5. Actinomycin D, showing 2-aminophenoxazin-3-one **4** core in bold.

The phenoxazinone scaffold is contained in heterocyclic iminoquinone **7**, which also displays a strong PCC of 0.74 to DPIQ activity (Figure 4). Benzo[*a*]phenoxazin-5-one **7** was assembled by condensation of 1-nitroso-2-naphthol with *L*-tyrosine [40]. The iminoquinone was identified from the NCI diversity set of 2,000 compounds as an inhibitor of HIF-1 α induced by insulin-like growth factor-1 [41]. HIF-1 α is one of the sub-units of HIF-1, the transcription factor over-expressed in hypoxic tumor cells, and responsible for regulating anaerobic metabolism that leads to tumor progression and angiogenesis [42]. SET reversible by oxygen, mediated by NADPH-cytochrome (P450) reductase is associated with tumor hypoxia [16,43,44], and is prevented by the NQO1-mediated two-electron reduction (Scheme 2) [7,18,21,45]. This specificity towards hypoxia provides a rationale for the weak correlation of iminoquinone **7** (0.38) to NQO1 expression across the NCI 60 cell lines.

Increased cell signaling supports cancer proliferation and requires increased phosphorylation. Human cytoplasmic protein tyrosine phosphatases (HCPTPs) are overexpressed in hypophosphorylated breast cancer cells [46]. Benzo[*a*]phenoxazin-5-one **7** was identified as the third most potent inhibitor of HCPTP isoform B ($IC_{50} = 31 \mu M$), after *in silico* docking of the compounds of the NCI Diversity Set I and enzyme inhibition assays on five selected compounds [47].

3.1.3. Benz[f]pyrido[1,2-*a*]indole-6,11-quinone

Updating the 2012 COMPARE analysis on DPIQ [7], revealed the carboxylic acid **8a** of benz[f]pyrido[1,2-*a*]indole-6,11-quinone (Figure 4), has the second highest PCC of 0.81. Testing against the NCI-DTP cell panel revealed the *N,N*-dimethylethyl carboxamide **8b** to be the most potent amongst carboxamide derivatives evaluated (Figure 6), with cytotoxicity against the adriamycin-resistant breast tumor cell line (NCI/ADR-RES) at a concentration lower than clinical anticancer agents, daunorubicin and mitoxantrone [48]. More recently, methylester **8c** was found to inhibit indoleamine 2,3-dioxygenase 1 (IDO1) [49]. IDO1 depletes tryptophan, with deficiencies in this essential amino acid leading to suppression of immune response to tumors [50]. 12-Unsubstituted and carboxylate ester derivatives of **8a**, including ethylester **8d** exhibit anti-fungal activities [51]. Carboxylates **8e-h** display micromolar anti-bactericidal activity against the Erdman strain of *Mycobacterium tuberculosis* with the molecular target being a membrane-bound, iron-thiol reductase (IspQ) [52].

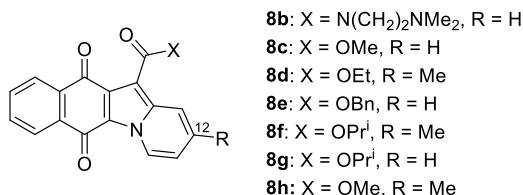
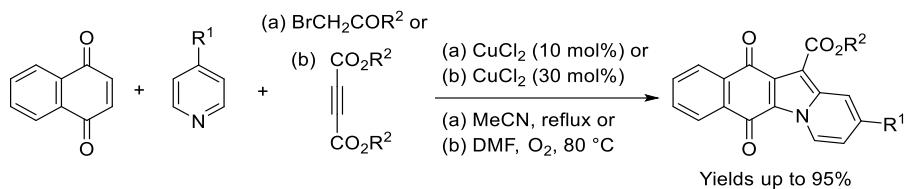


Figure 6. Biologically active benz[f]pyrido[1,2-*a*]indole-6,11-quinones [48,49,51,52].

For the biological evaluation purposes described above, the benz[f]pyrido[1,2-*a*]indole-6,11-quinone scaffold was obtained using adaptations of the traditional three-component one-pot condensation of 2,3-dichloro-1,4-naphthoquinone, an active methylene dicarbonyl and pyridine [48,49,51,52]. Yields are improved when using three-component reactions with 1,4-naphthoquinone and pyridine catalyzed by CuCl₂ with acyl bromides [53], or cleavage of butynedioates [54] (Scheme 4).



Scheme 4. Synthesis of benz[f]pyrido[1,2-*a*]indole-6,11-quinones [53,54].

3.1.4. Analogues of Seriniquinone

Dibenzo[*b,i*]thianthrene-5,7,12,14-tetrone was first synthesized in 1991 [55], and identified in 2014 as seriniquinone (Figure 7), derived from the marine bacterium, *Serinicoccus marinus* [56].

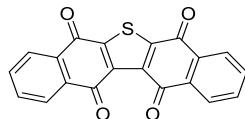
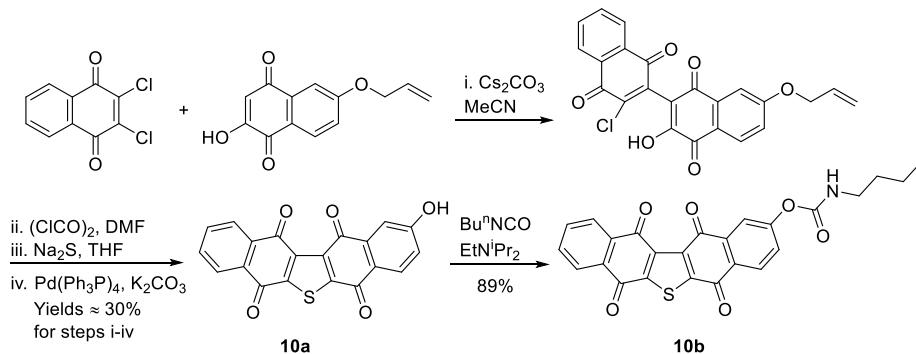


Figure 7. Seriniquinone.

Synthetic analogues of seriniquinone are required due to its poor water solubility, with **10a** and **10b** showing strong PCCs of 0.75 and 0.72 to DPIQ anti-cancer activity, but weak to moderate PCCs of 0.47 and 0.48 to NQO1 expression (Figure 4). Phenol **10a** and carbamate **10b** were prepared on a multi-gram scale via naphthalene-1,4-dione coupling, thiophene ring-formation, and Pd-induced deprotection (Scheme 5) [57]. Carbamate **10b** was shown to hydrolyze at room temperature to **10a** under phosphate-buffered saline (PBS) at pH 7.2. Analogues **10a** and **10b**, like DPIQ [7], possess specificity towards the NCI melanoma cell lines. Cytotoxicity of seriniquinones was demonstrated through binding to dermcidin [56,57], a small protein involved in cancer cell proliferation [58] and induction of cell death via autophagocytosis [56]. More recently, seriniquinones were shown to induce elevated levels of intracellular ROS, which trigger apoptotic events [59], which support the relatively weak correlations to NQO1. ROS is related to SET radical producing processes (Scheme 2), which are not mediated by NQO1.

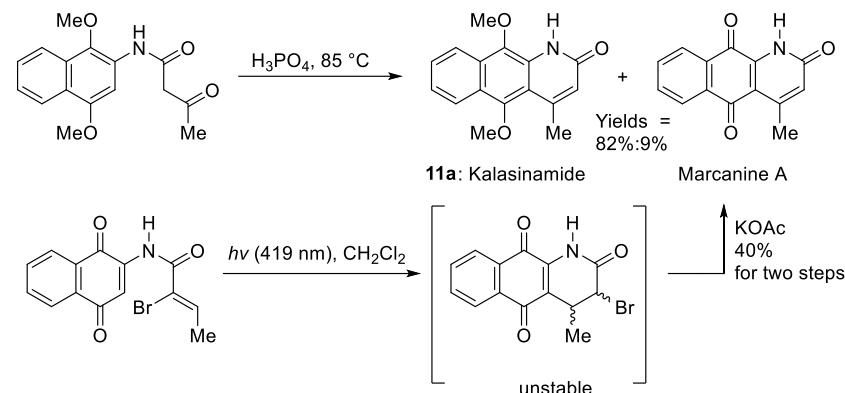


Scheme 5. Synthesis of seriniquinone analogues [57].

3.1.5. Azaanthracenone

Kalasinamide (**11a**) is an azaanthracenone first isolated from the tree, *Polyalthia suberosa* (Figure 4) [60]. Unreported analogue **11b** without the 4-methyl substituent was found by COMPARE analysis

to have a strong PCC to DPIQ of 0.73 and a weak to moderate PCC to NQO1 of 0.44. Kalasinamide is a photosensitizer thought to prevent the invasion of pathogens by generating singlet oxygen, and/or through conversion to the quinone, marcanine A [61]. Kalasinamide is prepared through acid-mediated substitutions onto the activated *p*-dimethoxybenzene [61,62], including using Knorr cyclization, which gives a mixture of **11a** and marcanine A, in the presence of oxygen and light (Scheme 6). Marcanine A is a plant natural product [61,63], also prepared through photochemical cyclization onto the naphthoquinone acrylamide substituent [64]. The potent anti-bacterial marcanine A [63], has cytotoxicity comparable to adriamycin using five different solid tumor cell lines [65].



Scheme 6. Syntheses of kalasinamide and marcanine A with yields [61,64].

3.1.6. Furano[2,3-*b*]naphthoquinone (FNQ)

The polyketide-isoprenoid **12a** was isolated from *Streptomyces cinnamonensis* (Figure 4) [66]. The biosynthesis of furano[2,3-*b*]naphthoquinone (FNQ) **12a** has attracted considerable attention [67], however there is no literature on antimicrobial and anti-cancer studies. COMPARE analysis shows **12a** exhibits a strong PCC to DPIQ of 0.72, and a moderate PCC of 0.55 to NQO1 expression across the NCI-60, which is comparable with DPIQ (0.51) (Figure 1). Related synthetic structures (FNQ13, Figure 8) induce mitochondrial swelling and apoptosis due to induction of ROS including H₂O₂ [68]. Furano[2,3-*b*]naphtho-4,9-dione **12b** isolated from *Tabebuia avellaneda* shows preference for inhibition of cancer cell growth over normal cells, with STAT3 inhibitory mechanisms proposed [69], while other FNQ natural products inhibit IDO1 (see Section 3.1.3) [70]. Inhibition of STAT3 may be important because the Janus kinase/signal transducers and activators of transcription (JAK/STAT) pathway plays a key role in membrane-to-nucleus signaling, which is critical in mediation of cancer and inflammation [71].

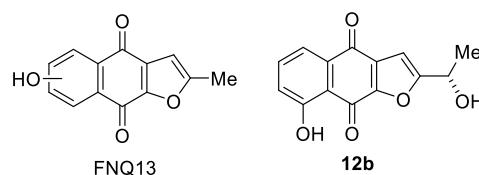
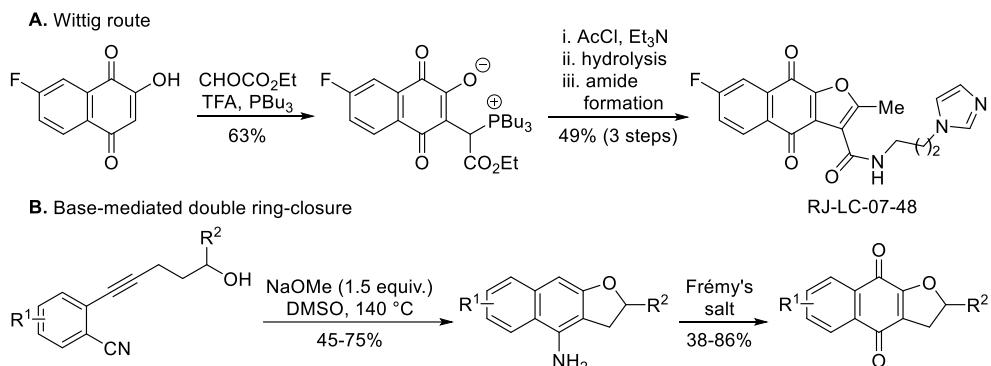


Figure 8. Anti-cancer FNQs.

Most syntheses of FNQ involve forming the furan ring from 2-hydroxy-1,4-naphthoquinone [70,72,73], including by an acetyl chloride/Et₃N-mediated Wittig reaction (Scheme 7A) [72,73]. The product (RJ-LC-07-48) is potent against drug-resistant non-small-cell lung cancer (NSCLC) cells by interaction with the minichromosomal maintenance protein MCM2, disrupting the formation of the MCM complex that is required for initiation of DNA replication [73]. Similar specificity towards non-small cell lung cancer cell lines was shown by DPIQ [7]. Wu and co-workers transformed 2-(5-hydroxy-1-pentynyl)benzonitriles to FNQ via a NaOMe-mediated ring-closure, followed by Frémy's salt oxidation (Scheme 7B) [74].



Scheme 7. Syntheses of furano[2,3-*b*]naphtho-4,9-diones [73,74].

3.2. COMPARE Analysis: Strong Correlations to Benzo[1,2,4]triazin-7-ones 5a as the Seed.

NCI COMPARE analysis using benzo[1,2,4]triazin-7-ones **5a** and pleurotin, as the seed, revealed very strong correlations to synthetic heterocyclic quinone scaffolds **13a-d** and **14a-f**, as well as to the natural products, pyranonaphthoquinones **15**, and discorhabdin C (PCC = 0.68-0.90, Figure 9). However, unlike DPIQ (above), very strong PCC of ~0.85 (to **5a**) are also found to non-quinone structures, notably synthetic derivatives of the anti-cancer sesquiterpene lactone, melampomagnolide B (MMB), isolated from *Magnolia grandiflora* [75,76]. MMB derivatives show specificity towards leukemic cell lines [76,77], with dimeric examples showing nanomolar activity against solid tumor cell lines in the NCI-60 panel [77]. Similarly, benzotriazin-7-ones **5a-d** also showed potent anti-proliferative effects against most NCI leukemic cell lines [10,11]. MMB is an extensively investigated scaffold that is thought to target the NFkB pathway through inhibition of the I κ B α /p65/p50 kinase complex (IKK) [78]. The following section reviews the heterocyclic quinones and iminoquinones having strong to very strong correlations to the anti-cancer activity of TrxR inhibitors **5a** and pleurotin. Quinoline-5,8-diones (e.g., **9b**) are reviewed separately in section 3.3.

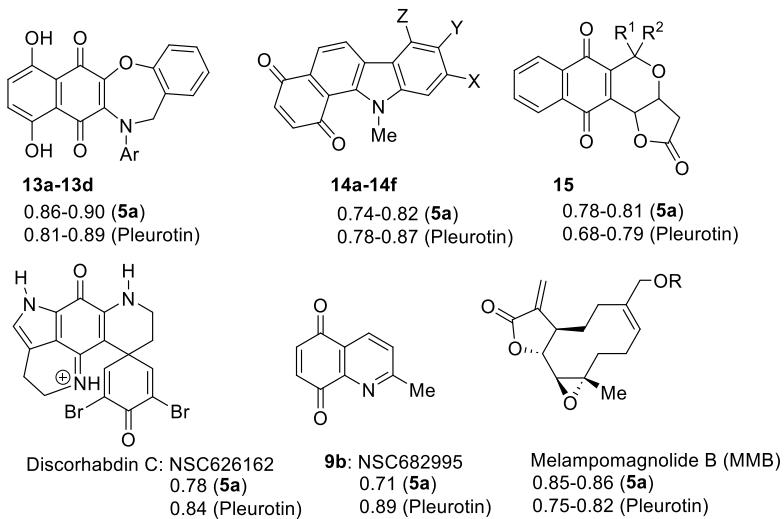


Figure 9. Very strong COMPARE correlations to benzo[1,2,4]triazinone **5a** and pleurotin.

3.2.1. Naphtho[2,3-b][1,4]oxazepine-6,11-dione

COMPARE gave almost perfect PCC of ~0.90 for the anti-cancer activity of benzo[1,2,4]triazinone **5a** to 1,4-benzoxazepine derivatives of 5,8-dihydroxy-1,4-naphthoquinones **13a-d** (related to Echinamines A and B) (Figure 10). Echinamines A and B are antioxidants isolated from the sea urchin *Scaphechinus mirabilis* [79], which suppress herpes simplex virus type 1 infection in the Vero monkey kidney cell line [80]. COMPARE analysis derived 1,4-benzoxazepines **13a-d** are unreported, giving analogous to **5a-d** (Figure 3) very strong correlations to pleurotin anticancer activity (PCC = 0.8-0.9).

The almost perfect correlations of **13a-d** to pleurotin are perhaps unsurprising given the common 1,4-naphthoquinone motif.

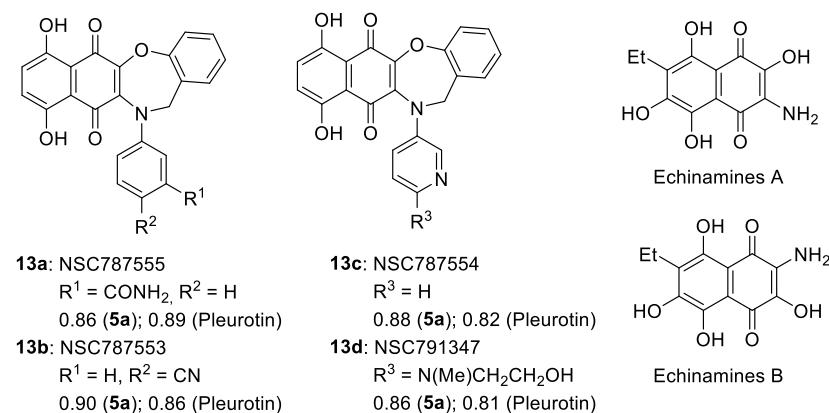
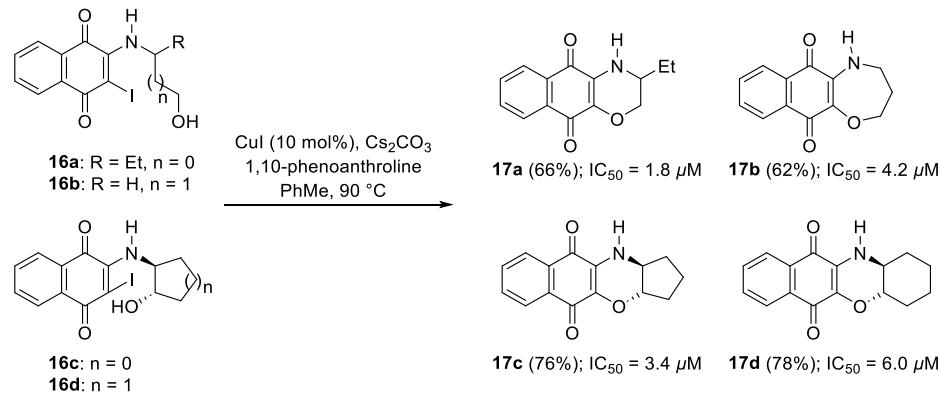


Figure 10. 1,4-Benzoxazepine derivatives of 1,4-naphthoquinones with PCCs and the related natural products Echinamine A and B.

You et al. synthesis of homomorpholine **17b**-fused and morpholine-fused, e.g., **17a**, **17c** and **17d**, naphthoquinone adducts used an intramolecular copper-catalyzed hydroxyl displacement of iodides (**16a-d**) with compounds giving moderate inhibition of lung A549 cell growth (Scheme 8) [81].



3.2.2. Benzo[a]carbazole-1,4-dione

Benzo[a]carbazole-1,4-dione **14c** and others appear in the patent literature as inhibitors for the thioredoxin TrxR-Trx system (Figure 11) [82]. This supports the very strong anti-cancer activity correlations with TrxR inhibitors, benzo[e][1,2,4]triazin-7-one **5a** (~0.74-0.82) and Pleurotin (0.78-0.87). Benzo[a]carbazole-1,4-dione **14a** was one of six hits out of 2,000 in the NCI library to inhibit Plasmodium kinesin-5 from both human and malaria vivax [83]. Compounds **14a** and **14d-f** were amongst 35 hits out of 1597 compounds in the NCI Diversity SET III screen with >50% inhibitory activity at concentrations of 20 μM of the immunosuppressive enzyme, IDO1 [84]. Benzo[a]carbazole-1,4-dione **14a** is effective against vancomycin-resistant *Staphylococcus aureus* by targeting the cysteine thiol of bacterial MarR transcription factors [85].

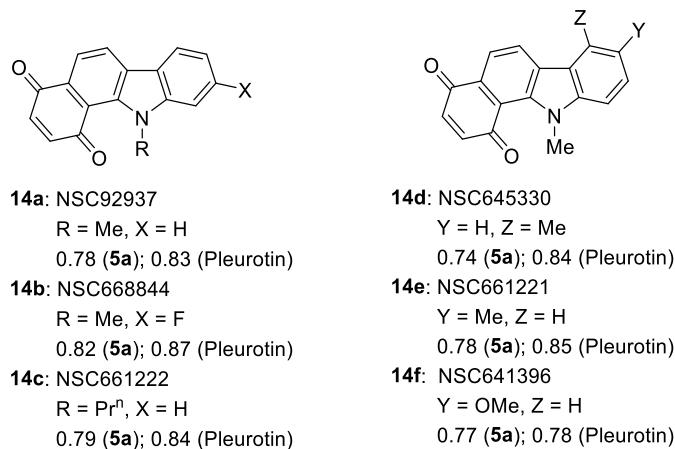
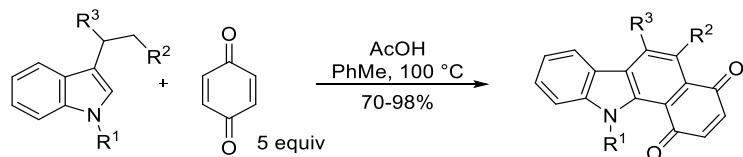


Figure 11. Benzo[a]carbazole-1,4-dione antibiotics and anti-cancer agents [82-85].

The traditional synthesis of the benzo[a]carbazole-1,4-dione scaffold (Figure 11) involves a Diels-Alder cycloaddition of 3-vinylindole with benzoquinone and oxidative dehydrogenation [86,87]. The 3-vinylindole can be generated in situ from 3-ethylindole using benzoquinone as the oxidizing agent, and the benzoquinone then acts as the dienophile to give the benzo[a]carbazole (Scheme 9) [87].



Scheme 9. Synthesis of benzo[a]carbazole-1,4-dione [87].

3.2.3. Pyranonaphthoquinones

Pyranonaphthoquinones are natural products possessing the naphtho[2,3-c]pyran-5,10-dione ring system (Figure 12) [88,89]. Benzotriazinone **5a** and pleurotin show strong to very strong PCCs of 0.72-0.83 to the antibiotics kalafungin and nanaomycin A first isolated from *Streptomyces tashiensis* strain Kala [90,91]. Nanaomycin D isolated from *Streptomyces rosa* is enantiomeric to kalafungin with nanaomycin A, the cleaved lactone [92]. Kalafungin and nanaomycin analogues show specificity towards breast cancer cell lines over-expressing cytochrome P450 oxidoreductase with cytotoxicity diminished under anoxia, where reactive oxygen production is inhibited [93]. The SET recycling mechanism is also important in antibacterial activity [94]. Conversely, kalafungin and analogues were reported inhibitors of the serine-threonine kinase AKT via a proposed two-electron reduction to the hydroquinone with alkylation of the quinone methide by a cysteine in the activation loop (T-loop) of the kinase domain of AKT [95]. Nanaomycin A was identified in a screening program as a DNA methyltransferase inhibitor, with biochemical assays revealing specificity towards DNMT3B with docking hypothesizing reduction by the sulfur atom of a cysteine in the catalytic site [96].

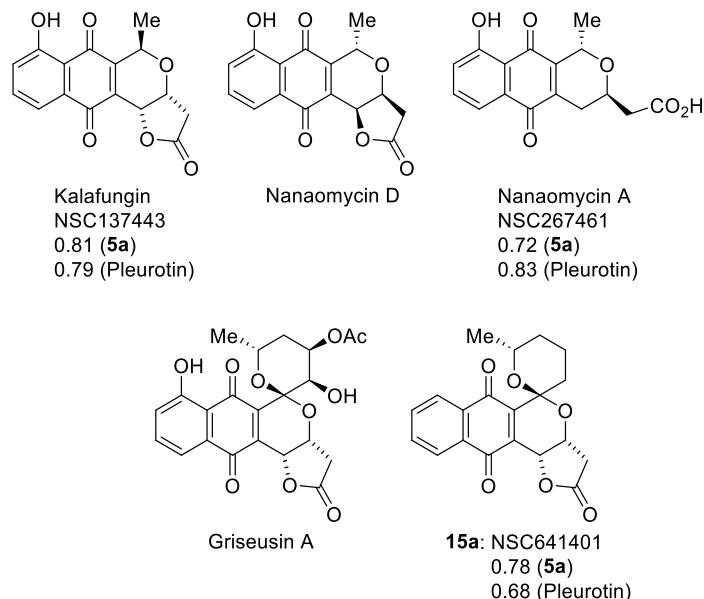
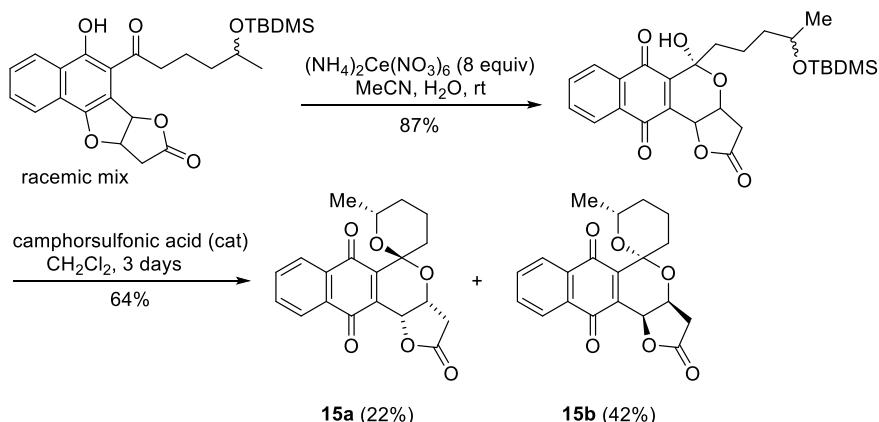


Figure 12. The pyranonaphthoquinones with COMPARE analysis PCCs.

Griseusin A, isolated from *Streptomyces griseus*, is a pyranonaphthoquinone antibiotic [97], that bears structural similarity to kalafungin, but possesses a 1,7-dioxaspiro[5,5]-undecane ring system. Synthetic griseusin **15a** exhibits a strong correlation in anti-cancer activity to benzotriazinone **5a** (PCC = 0.78) and pleurotin (0.68) (Figure 12). The multi-step synthesis of griseusin **15a** involves a cerium(IV) ammonium nitrate-mediated oxidative rearrangement and acid-mediated cyclization to the spiroacetal [98-101]. Brimble and co-workers have reported the separation of isomers **15a** and **15b** using flash chromatography (Scheme 10) [99-101].



Scheme 10. Brimble and co-workers synthesis of the griseusin scaffold [99,100].

3.2.4. Discorhabdin C

The pyrroloiminoquinone alkaloids, discorhabdins are isolated from numerous cold water marine sponges, with cytotoxic discorhabdin C from the New Zealand sponge, *Latrunculia Bocage* [102,103]. Discorhabdin C exhibits strong to very strong PCCs to benzotriazinone **5a** (0.78) and pleurotin (0.84) (Figure 9). Munro and co-workers reported selectivity towards the NCI colon and leukemia subpanels [104], while others reported in vitro anti-hepatitis virus C, antimalarial and antimicrobial activities [105]. Figg and co-workers performed high-throughput screens on crude natural product extracts and identified 3-dihydro-discorhabdin C as a HIF-1 α /p300 inhibitor [106], which interferes with the HIF-1 α and p300 protein–protein interaction to decrease HIF-1 α -related transcription (Figure 13) [43].

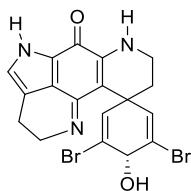
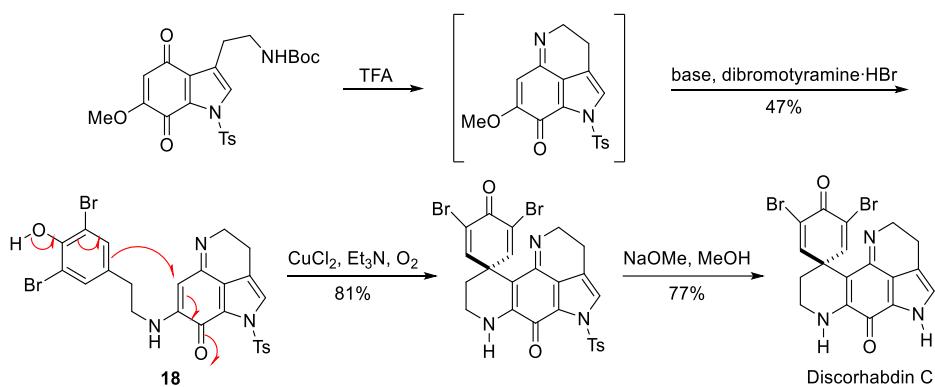


Figure 13. 3-Dihydrodiscorhabdin C.

In the 1990s, many multi-step total syntheses of discorhabdin C were reported, with approaches starting from the quinoline with formation of the fused pyrrole [107,108], and from the indole with formation of the six-membered imino-ring through condensation with the quinone [109-111]. One approach involves oxidation of the tyramine-substituted indoloquinonimine **18** in a nucleophilic addition onto the iminoquinone (Scheme 11) [111]. Kublak and Confalone had earlier used the para-phenoxy approach for alkylation of an adjacent naphthoquinone ethylamino-substituent by displacement of a mesylate [112].



Scheme 11. Aubart and Heathcock synthesis of discorhabdin C [111].

3.3. COMPARE Analysis: Strong Correlations to DPIQ and Benzo[1,2,4]triazin-7-one **5a** as the Seed: Quinoline-5,8-diones

Synthetic quinoline-5,8-diones scaffolds correlate strongly with the anti-cancer activity of DPIQ, benzotriazinone **5a**, and pleurotin. 6-Aminoethyl substituted derivative **9a** shows one of the strongest PCC to DPIQ of 0.80, with a relatively weak PCC to NQO1 expression of 0.42 (Figure 4). 2-Methyl substituted derivative **9b** shows a very strong PCC to pleurotin of 0.89, and a strong correlation to benzotriazinone **5a** of 0.71 (Figure 9). There is an abundance of synthetic and anti-cancer studies on quinoline-5,8-diones [113], with most stimulated by the *broad* range of cytotoxicity against solid tumors displayed by streptonigrin, a recognised substrate for NQO1 (Figure 14) [114]. Streptonigrin was isolated from *Streptomyces flocculus* [115], with lavendamycin a biosynthetically related antibiotic isolated from *Streptomyces lavendulae* [116]. Unfortunately, both streptonigrin and lavendamycin have proved too toxic for clinical use, although analogues of lavendamycin display potent HIV-reverse transcription inhibition [117], and improved specificity for NQO1 [118].

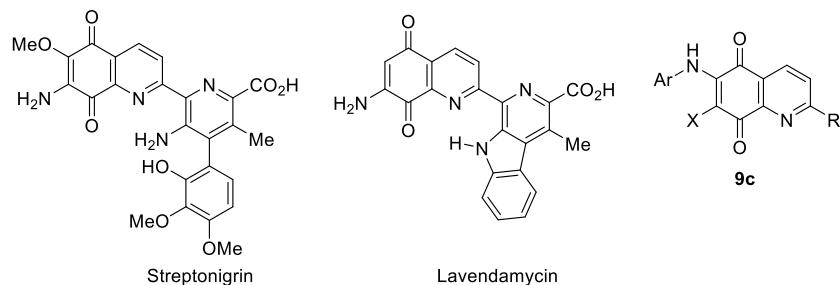


Figure 14. Popular quinoline-5,8-dione scaffolds for biological activity investigations.

N-Alkylamino compounds, analogous to dione **9a** also possess antimalarial activity [119], while compound **9b** is a reported inhibitor of Cdc25B ($IC_{50} = 4.6 \mu M$), a protein phosphatase involved in regulating cyclin-dependent kinase activity during the cell cycle [120]. Recent studies have reported 6-N-arylquinoline-5,8-diones **9c** (Figure 14) as inhibitors of *Mycobacterium tuberculosis* [121], as well as of Gram negative and positive bacteria [122]. 6-N-Arylquinoline-5,8-diones **9c** are also reported to cleave DNA, as an underlying mechanism for apoptosis induction in leukemic cell lines [123]. Nevertheless, the consensus is that there is a strong correlation between NQO1 bioreduction and anti-cancer activity for many quinoline-5,8-diones [118,124,125], given that NQO1 is strongly over-expressed in solid tumours [13] relative to normal tissues [126].

3.4. COMPARE Analysis using Molecular Target Expression.

In this section the COMPARE algorithm was used to derive PCCs for the similarity in expression of chosen cancer molecular targets, NQO1 and TrxR to compound growth inhibition patterns across the NCI-60 cell line panel. Section 3.4.1 deals with NQO1 expression and establishes the strongest correlations to compound cytotoxicity. Section 3.4.2 reveals the PCCs of compounds with almost perfect direct correlations to the anti-cancer activity of known TrxR inhibitors, benzo[1,2,4]triazinones **5a-d** and pleurotin.

3.4.1. Compound Correlations to NQO1 expression

Since most compounds that correlated strongly with DPIQ anti-cancer activity, gave modest PCCs of 0.38-0.55 to NQO1 expression, except for benzo[e]perimidine **3a**, which was noticeably stronger (PCC = 0.64, Figures 1 & 4), we searched for compounds with the strongest PCCs to NQO1 expression. The strongest compound correlations to NQO1 were of similar magnitude to the PCC for benzo[e]perimidine **3a** (Figure 15). Phenazine-5,10-dioxide **19**, 5-hydroxy-6-methoxy-8-nitroquinoline **20**, and indolequinones **21a** and **21b** gave PCCs of ~0.6-0.7 to NQO1 and DPIQ. Phenazine-5,10-dioxide **19** is unreported, and has a marginally lower PCC of 0.51 than the other NQO1 substrates to DPIQ.

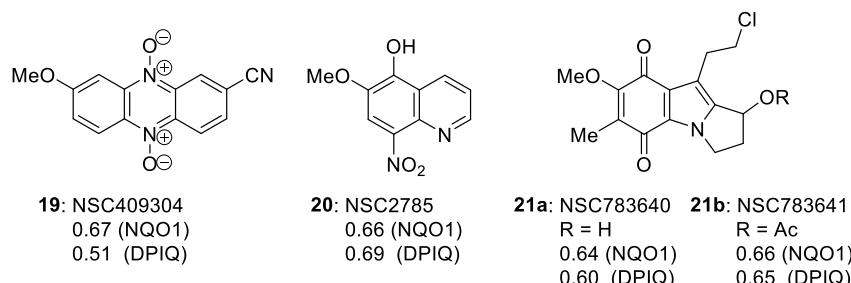


Figure 15. The three strongest PCCs to NQO1.

Analogues of phenazine **19** are reported as π -stacking DNA-intercalators with differential toxicity through DNA-damaging \cdot OH release under hypoxic conditions [127,128]. Phenazine-5,10-dioxides are designed to model 3-amino-1,2,4-benzotriazine-1,4-dioxide (tirapazamine, TPZ, Figure 16), which reached advanced clinical trials as a hypoxia-activated prodrug [129]. However, although there are successful Phase I and Phase II trials, Phase III randomized controlled trials showed no benefit of TPZ in chemotherapy without using an approach to ensure sustained tumor hypoxia [130]. Quinoline **20** is a synthetic precursor for 5-alkoxy derivatives of the clinical anti-malarial primaquine [131]. The biological activity of **20** is unreported, and, tentatively, the compound is metabolized to a quinoline-5,8-dione antibiotic upon reductive activation-oxidation (Section 3.3).

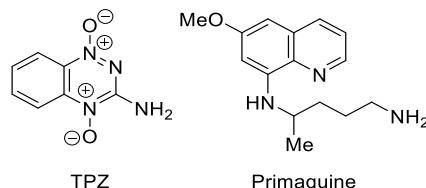
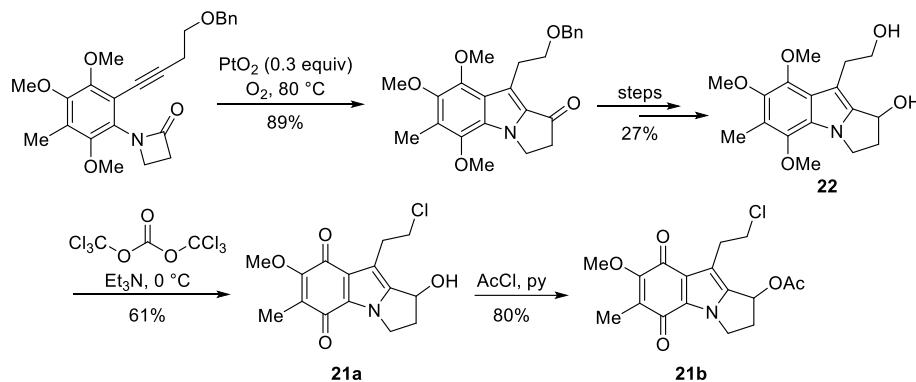


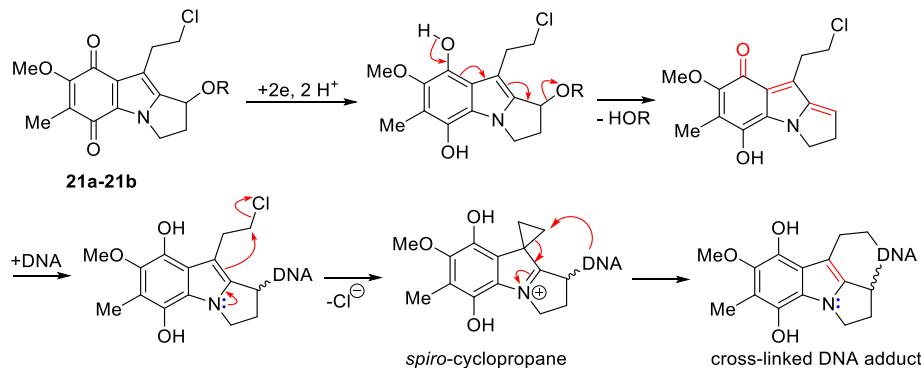
Figure 16. TPZ and primaquine related to **19** and **20**, respectively.

Pyrrolo[1,2-*a*]indoles are highly pursued synthetic targets [132], because they form the core of mitomycins, in particular MMC (Figure 3) [133]. The 7-methoxymitosene skeleton of **21a** and **21b** was accessed via a Pt-promoted cyclization of a β -lactam onto an internal acetylene (Scheme 12) [134]. Triphosgene and triethylamine preferentially chlorinate the primary alcohol over the secondary alcohol in **22** to give **21a** [135], with the substitution reported to be driven by steric demand [136].



Scheme 12. Cycloisomerization to the 7-methoxymitosene skeleton [134,135].

7-Methoxymitosenes **21a** and **22b** are designed to form an electrophilic *spiro*-cyclopropane intermediate upon reductive activation, which enables crosslinking possibly within the same DNA molecule (Scheme 13) [135]. Indolequinones **21a** and **21b** were evaluated using the prostate cancer cell line PPC-1 and the normal prostate cell line RWPE-1, with alcohol **21a** exhibiting the greater selectivity towards the cancer cell line [135].



Scheme 13. Hypothesis for cytotoxicity of 7-methoxymitosenes [135].

Interestingly, known TrxR inhibitors, benzo[1,2,4]triazinones **5a**, **5b** and pleurotin, as well as compounds that correlate very strongly to their anti-cancer activity, benzo[*a*]carbazole-1,4-dione **14b**, kalafungin, and discorhabdin C gave negative PCCs to NQO1 expression across the NCI-60 panel (PCC = -0.27 to -0.48, Table 1). This suggests that TrxR inhibitors may also act as inhibitors of other two-electron reductases over-expressed in solid tumors, namely NQO1 (see discussion below).

Table 1. COMPARE analysis derived PCCs to NQO1 and TrxR expression across the NCI-60 panel for TrxR inhibitors.¹

Compound	NQO1	TrxR
5a	-0.37	-0.24
5b	-0.30	-0.12
Pleurotin	-0.27	-0.21
14b	-0.48	-0.22
Kalafungin	-0.27	-0.24
Discorhabdin C	-0.29	-0.37

¹ See section 2 for the method [6,7,23].

3.4.1. Compound Correlations to TrxR

The negative correlation to TrxR (TXNRD1) expression by reported TrxR-Trx inhibitory compounds [10,11,15,19,137], as well as by compounds with very strong similarities in anti-cancer activity suggests that the cytotoxicity of these compounds is greater when TrxR levels are low (Table 1). Increased inhibition may thus prevent the reducing system's essential role in tumor development, such as inhibiting apoptosis, and angiogenesis promotion [19,137]. The TrxR-Trx system modulates cell signaling in particular through interactions with the tumor suppressor protein PTEN (protein tyrosine phosphatase and tensin homolog). The TrxR-Trx system activates PTEN through reduction [138], with oxidation of PTEN leading to inactivation [138,139]. Trx-1-PTEN interactions through disulfide bond formation between protein Cys active sites inhibits catalytic activity [140]. The inactivation of PTEN leads to activation of tumor propagating PI3K-AKT kinase signaling pathways [138]. The naturally occurring sesquiterpene lactone, parthenolide (structurally similar to MMB, Figure 9) is reported to inhibit TrxR by shifting the enzyme from antioxidant activity to ROS generation leading to promotion of apoptosis in HeLa cells (Figure 17) [141]. Cell death through ROS generation seems a common mode of action for most classes of compounds inhibiting TrxR [19,93,137,141,142]. Alternatively, the greater potency of compounds as cytotoxins when reductase expression is low, may be due to the TrxR-Trx or NQO1 systems detoxifying the heterocyclic (imino)quinones through bioreduction (Scheme 2). Further, COMPARE reveals a diverse range of chemical structures, including many structures without the (imino)quinone moiety, that have strong PCCs to the anti-cancer activity of TrxR inhibitors, **5a**, **5b** and pleurotin. Since benzotriazinones **5a** and **5b** exhibit reversible mixed and uncompetitive inhibition of TrxR respectively [10,137], with binding to positions other than the active site likely, it seems that bioreduction may not be directly involved in the inhibition or inactivation of TrxR by many compounds.

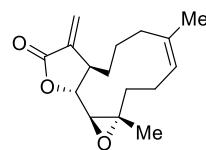


Figure 17. Parthenolide.

Importantly, specificity for NQO1 was demonstrated by DPIQ, benzo[e]perimidine **3a**, 2-amino-phenoxazinone **4** and phenazine **19**, which have no correlations to TrxR (TXNRD1: PCC = -0.15 to 0.12), but moderate to strong correlations to NQO1 expression across the NCI-60 cell line panel (PCC = 0.47-0.67, Table 2). This supports the premise that these compounds are specifically substrates for cellular NQO1, which facilitates reductively activated cytotoxicity. Further, MMC and other recognized indolequinone NQO1 substrates [7,13,143,144,145], are shown to irreversibly inhibit TrxR through formation of covalently bound adducts (Figure 18) [135,143,145]. Comparing mean growth (inhibition) graphs of NQO1 substrates, the greatest cytotoxicity is towards solid tumor cell lines, including most melanoma, non-small cell lung, and colon cancer cell lines, with negligible toxicity

towards leukemic cell lines, in contrast with the strong anti-leukemic activity of TrxR inhibitors (see Supplementary Materials).

Table 2. COMPARE analysis derived PCCs to NQO1 and TrxR expression across the NCI-60 panel for NQO1 substrates.¹

Compound	NQO1	TrxR
DPIQ	0.51	0.12
3a	0.64	0.11
4	0.47	-0.15
19	0.67	0.10

¹ See section 2 for the method [6,7,23], PCC for DPIQ, **3a** and **4** with NQO1 previously reported [7].

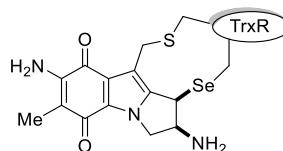


Figure 18. Inactivation of TrxR: MMC-TrxR adduct [143].

5. Conclusions

The NCI COMPARE program has enabled the composition of this review on the synthesis and biological activity of heterocyclic iminoquinones and quinones, where scaffolds are categorized according to similarities in their anti-cancer activity. COMPARE has enabled hypotheses of mechanisms of actions to be made, although it is important to emphasize that all correlations herein need to be verified experimentally [2]. We reveal several natural products with strong to very strong correlations in patterns of anti-cancer activity to DPIQ, benzo[1,2,4]triazin-7-one **5a**, and pleurotin. Most heterocyclic scaffolds exhibit significant biological activity to warrant extensive synthetic investigations.

Compounds strongly correlating to DPIQ, and showing specificity to NQO1 expression, have no correlation to the alternative two-electron reductase, TrxR. These “NQO1 specific prodrugs” are flat aromatic heterocycles with fused oxidizable, e.g., quinone or iminoquinone, moieties. The correlations to NQO1 expression, however, are modest, suggesting that NQO1 is not the sole cellular molecular component that influences cell sensitivity. NQO1 has a “Janus” effect in cancer biology [18], where it behaves as either a tumor suppressor or a tumor promotor, with the former based on the prevention of SET processes that lead to an accumulation of harmful ROS. However, the redox appears interchangeable, and NQO1-activated pathways can lead to ROS-induced apoptosis [18,21]. Ultimately, designed prodrugs for NQO1 may also effectively act as inhibitors of redox defensive signaling pathways that contribute to carcinogenesis, including the transcriptional regulators Nrf2 [25], HIF-1 α [43], and JAK/STAT [71].

There is a diverse range of compounds with almost direct correlations to the anti-cancer activity patterns of the TrxR inhibitors, benzotriazinone **5a** and pleurotin. These compounds do not always possess an oxidizing (iminoquinone or quinone) group, suggesting bioreduction is unnecessary, and compounds are more cytotoxic in the absence of the reductase. Among these compounds, there are similarities in biological activity, with many possessing antiviral, antimicrobial (including antimalarial), as well as anti-cancer activity. These compounds tend to be cytotoxic towards leukemic as well as solid tumor cell lines, unlike NQO1 substrates, which show specificity towards solid tumors over-expressing NQO1. There are similarities in the mechanism of action, with compounds correlating to NQO1 expression or acting as TrxR inhibitors, both showing inhibition of the monomeric oxidase, IDO1 [50]. 1,3-Diphenylbenzo[1,2,4]triazinones are inhibitors in AD [14], as well as cancer [10,11,137]. Reduced Trx protein binds to apoptosis signal-regulating kinase 1 (ASK1), thus inhibition of TrxR leads to oxidized Trx, which cannot bind to ASK1. The outcome is mitochondrial apoptosis through activation of ASK1, downstream JNK and mitogen-activated protein kinase (MAPK14) signaling

pathways [145], which lead to reduced inflammatory response and tumorigenesis or the initiating of neurodegenerative disorders, such as AD [137,142].

Supplementary Materials: The following supporting information can be downloaded at: www.mdpi.com/xxx/s1, Figures S1-S84.

Funding: Not applicable.

Author Contributions: Investigation and methodology: N.H. and M.F.; Review and editing: P.A.K and M.P.C.; Conceptualization, supervision, writing, review, and editing: F.A.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Not applicable.

Acknowledgments: F.A. is grateful to Drs. Vincent Fagan [6], Robert Coyle [23], and Martin Sweeney [137] for preliminary data. F.A. dedicates this review to Prof. William J. Spillane (University of Galway) on his 80th birthday.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Shoemaker, R. H. The NCI60 human tumour cell line anticancer drug screen. *Nat. Rev. Cancer* **2006**, *6*, 813-823. DOI
2. Holbeck, S. L. Update on NCI *in vitro* drug screen utilities. *Eur. J. Cancer* **2004**, *40*, 785-793. DOI
3. Sharma, S. V.; Haber D. A.; Settleman, J. Cell line-based platforms to evaluate the therapeutic efficacy of candidate anticancer agents. *Nat. Rev. Cancer* **2010**, *10*, 241-253. DOI
4. Holbeck, S. L.; Camalier, R.; Crowell, J. A.; Govindharajulu, J. P.; Hollingshead, M.; Anderson, L. W.; Polley, E.; Rubinstein, L.; Srivastava, A.; Wilsker, D.; Collins, J. M.; Doroshow, J. H. The National Cancer Institute ALMANAC: A Comprehensive screening resource for the detection of anticancer drug pairs with enhanced therapeutic activity. *Cancer Res.* **2017**, *77*, 3564-3576. DOI
5. Paull, K. D.; Shoemaker, R. H.; Hodes, L.; Monks, A.; Scudiero, D. A.; Rubinstein, L.; Plowman, J.; Boyd, M. R. Display and analysis of patterns of differential activity of drugs against human tumor cell lines: development of mean graph and COMPARE algorithm. *J. Natl. Cancer Inst.* **1989**, *81*, 1088-1092. DOI
6. Fagan, V. Synthesis and anti-cancer activity of novel imidazo[5,4-f]benzimidazolequinones. Ph.D., National University of Ireland Galway, Ireland, 2011.
7. Fagan, V.; Bonham, S.; Carty, M. P.; Saenz-Méndez, P.; Eriksson, L. A.; Aldabbagh, F. COMPARE analysis of the toxicity of an iminoquinone derivative of the imidazo[5,4-f]benzimidazoles with NAD(P)H:quinone oxidoreductase 1 (NQO1) activity and computational docking of quinones as NQO1 substrates. *Bioorg. Med. Chem.* **2012**, *20*, 3223-3232. DOI
8. Fagan, V.; Bonham, S.; McArdle, P.; Carty, M. P.; Aldabbagh, F. Synthesis and toxicity of new ring-fused imidazo[5,4-f]benzimidazolequinones and mechanism using amine N-oxide cyclization. *Eur. J. Org. Chem.* **2012**, 1967-1975. DOI
9. Fagan, V.; Bonham, S.; Carty, M. P.; Aldabbagh, F. One-pot double intramolecular homolytic aromatic substitution routes to dialycyclic ring fused imidazobenzimidazolequinones and preliminary analysis of anti-cancer activity. *Org. Biomol. Chem.* **2010**, *8*, 3149-3156. DOI
10. Sweeney, M.; Coyle, R.; Kavanagh, P.; Berezin, A. A.; Lo Re, D.; Zissimou, G. A.; Koutentis, P. A.; Carty, M. P.; Aldabbagh, F. Discovery of anti-cancer activity for benzo[1,2,4]triazin-7-ones: Very strong correlation to pleurotin and thioredoxin reductase inhibition. *Bioorg. Med. Chem.* **2016**, *24*, 3565-3570. DOI
11. Keane, L. -A. J.; Mirallai, S. I.; Sweeney, M.; Carty, M. P.; Zissimou, G. A.; Berezin, A. A.; Koutentis, P. A.; Aldabbagh, F. Anti-cancer activity of phenyl and pyrid-2-yl 1,3-substituted benzo[1,2,4]triazin-7-ones and stable free radical precursors. *Molecules* **2018**, *23*, 574. DOI
12. Conboy, D.; Aldabbagh, F. 6-Imino-1,2,3,4,8,9,10,11-octahydropyrido[1,2-a]pyridoimidazo[4,5-f]benzimidazole-13-one: synthesis and cytotoxicity evaluation. *Molbank* **2020**, (1), m1118. DOI

13. Fitzsimmons, S. A.; Workman, P.; Grever, M.; Paull, K.; Camalier, R.; Lewis, A. D. Reductase enzyme expression across the National Cancer Institute tumor cell line panel: correlation with sensitivity to mitomycin C and EO9. *J. Natl. Cancer Inst.* **1996**, *88*, 259-269. DOI

14. Catto, M.; Berezin, A. A.; Lo Re, D.; Loizou, G.; Demetriades, M.; De Stradis, A.; Campagna, F.; Koutentis, P. A.; Carotti, A. Design, synthesis and biological evaluation of benzo[e][1,2,4]triazin-7(1H)-one and [1,2,4]triazino[5,6,1-*j*]carbazol-6-one derivatives as dual inhibitors of beta-amyloid aggregation and acetyl/butyryl cholinesterase. *Eur. J. Med. Chem.* **2012**, *58*, 84-97. DOI

15. Welsh, S. J.; Williams, R. R.; Birmingham, A.; Newman, D. J.; Kirkpatrick, D. L.; Powis, G. The thioredoxin redox inhibitors 1-methylpropyl 2-imidazolyl disulfide and pleurotin inhibit hypoxia-induced factor 1 α and vascular endothelial growth factor formation. *Mol. Cancer Ther.* **2003**, *2*, 235-243. PMID: 12657718

16. Saikolappan, S.; Kumar, B.; Shishodia, G.; Koul, S.; Koul, H. K. Reactive oxygen species and cancer: a complex interaction. *Cancer Lett.* **2019**, *452*, 132-143. DOI

17. Bolton, J. L.; Dunlap, T. Formation and biological targets of quinones: Cytotoxic versus cytoprotective effects. *Chem. Res. Toxicol.* **2017**, *30*, 13-37. DOI

18. Zhang, K.; Chen, D.; Ma, K.; Wu, X.; Hao, H.; Jiang, S. NAD(P)H:quinone oxidoreductase 1 (NQO1) as a therapeutic and diagnostic target in cancer. *J. Med. Chem.* **2018**, *61*, 6983-7003. DOI

19. Bian, M.; Fan, R.; Zhao, S.; Liu, W. Targeting the thioredoxin system as a strategy for cancer therapy. *J. Med. Chem.* **2019**, *62*, 7309-7321. DOI

20. Tomasz, M.; Lipman, R.; Chowdary, D.; Pawlak, J.; Verdine, G. L.; Nakanishi, K. Isolation and structure of a covalent cross-link adduct between mitomycin C and DNA. *Science* **1987**, *235*, 1204-1208. DOI

21. Parkinson, E. I.; Bair, J. S.; Cismesia, M.; Hergenrother, P. J. Efficient NQO1 substrates are potent and selective anticancer agents. *ACS Chem. Biol.* **2013**, *8*, 2173-2183. DOI

22. Li, R.; Bianchet, M. A.; Talalay, P.; Amzel, L. M. NAD(P)H:quinone reductase, a flavoprotein involved in cancer chemoprotection and chemotherapy: mechanism of the two-electron reduction. *Proc. Natl. Acad. Sci. U. S. A.* **1995**, *92*, 8846-8850. DOI

23. Coyle, R. Barton esters for initiator-free radical cyclisation with heteroaromatic substitution and anti-cancer evaluation of benzo[e][1,2,4]triazin-7-ones. Ph.D., National University of Ireland Galway, Ireland, 2014.

24. Dai, B.; Augustine, J. J.; Kang, Y.; Roife, D.; Li, X.; Deng, J.; Tan, L.; Rusling, L. A.; Weinstein, J. N.; Lorenzi, P. L.; Kim, M. P.; Fleming, J. B. Compound NSC84167 selectively targets NRF2-activated pancreatic cancer by inhibiting asparagine synthesis pathway. *Cell Death Dis.* **2021**, *12*, 693. DOI

25. Lister, A.; Nedjadi, T.; Kitteringham, N. R.; Campbell, F.; Costello, E.; Lloyd, B.; Copple, I. M.; Williams, S.; Owen, A.; Neoptolemos, J. P.; Goldring, C. E.; Park, B. K. Nrf2 is overexpressed in pancreatic cancer: implications for cell proliferation and therapy. *Mol. Cancer* **2011**, *10*, 37. DOI

26. Stefańska, B.; Dzieduszycka, M.; Martelli, S.; Tarasiuk, J.; Bontemps-Gracz, M.; Borowski, E. 6-[(Aminoalkyl)amino]-substituted 7H-benzo[e]perimidin-7-ones as novel antineoplastic agents. Synthesis and biological evaluation. *J. Med. Chem.* **1993**, *36*, 38-41. DOI

27. Stefańska, B.; Dzieduszycka, M.; Bontemps-Gracz, M. M.; Borowski, E.; Martelli, S.; Supino, R.; Pratesi, G.; De Cesare, M.; Zunino, F.; Kuśnierszyk, H.; Radzikowski, C. 8,11-Dihydroxy-6-[(aminoalkyl)amino]-7H-benzo[e]perimidin-7-ones with activity in multidrug-resistant cell lines: synthesis and antitumor evaluation. *J. Med. Chem.* **1999**, *42*, 3494-3501. DOI

28. Dzieduszycka, M.; Martelli, S.; Arciemiu, M.; Bontemps-Gracz, M. M.; Kupieca, A.; Borowski, E. Effect of modification of 6-[(Aminoalkyl)amino]-7H-benzo[e]perimidin-7-ones on their cytotoxic activity toward sensitive and multidrug resistant tumor cell lines. Synthesis and biological evaluation. *Bioorg. Med. Chem.* **2002**, *10*, 1025-1035. DOI

29. Anzai, K.; Isono, K.; Okuma, K.; Suzuki, S. The new antibiotics, questiomycins A and B. *J. Antibiot. (Tokyo)* **1960**, *13*, 125-132. PMID: 13848826

30. Kozlovsky, A. G.; Zhelifonova, V. P.; Antipova, T. V.; Adanin, V. M.; Novikova, N. D.; Deshevaya, E. A.; Schlegel, B.; Dahse, H. M.; Gollmik, F.; Gafe, U. Penicillium expansum, a resident fungal strain of the orbital complex *Mir*, producing xanthocillin X and questiomycin A. *Appl. Biochem. Microbiol.* **2004**, *40*, 344-349. DOI

31. Bitzer, J.; Große, T.; Wang, L.; Lang, S.; Beil, W.; Zeeck, A. New aminophenoxazinones from a marine *Halomonas* sp.: fermentation, structure elucidation, and biological activity. *J. Antibiot.* **2006**, *59*, 86-92. DOI

32. Hanawa, T.; Osaki, T.; Manzoku, T.; Fukuda, M.; Kawakami, H.; Tomoda, A.; Kamiya, S. *In vitro* antibacterial activity of Phx-3 against *Helicobacter pylori*. *Biol. Pharm. Bull.* **2010**, *33*, 188-191. DOI

33. Guo, S.; Hu, H.; Bilal, M.; Zhang, X. Production of antibacterial queuomycin A in metabolically engineered *Pseudomonas chlororaphis* HT66. *J. Agric. Food Chem.* **2022**, *70*, 7742–7750. DOI

34. Barry III, C. E.; Nayar, P. G.; Begley, T. P. Phenoxazinone synthase: mechanism for the formation of the phenoxazinone chromophore of actinomycin1. *Biochem.* **1989**, *28*, 6323-6333. DOI

35. Le Roes-Hill, M.; Goodwin, C.; Burton, S. Phenoxazinone synthase: what's in a name? *Trends Biotechnol.* **2009**, *27*, 248-258. DOI

36. Nakachi, T.; Tabuchi, T.; Takasaki, A.; Arai, S.; Miyazawa, K.; Tomoda, A. Anticancer activity of phenoxazines produced by bovine erythrocytes on colon cancer cells. *Oncol. Rep.* **2010**, *23*, 1517-1522. DOI

37. Hongo, T.; Miyano-Kurosaki, N.; Kurosaki, K.; Hata, A.; Harigae, S.; Tomoda, A. 2-Aminophenoxazine-3-one prevents pulmonary metastasis of mouse B16 melanoma cells in mice. *J. Pharmacol. Sci.* **2010**, *114*, 63-68. DOI

38. Kohno, K.; Miyake, M.; Sano, O.; Tanaka-Kataoka, M.; Yamamoto, S.; Koya-Miyata, S.; Arai, N.; Fujii, M.; Watanabe, H.; Ushio, S.; Iwaki, K.; Fukuda, S. Anti-inflammatory and immunomodulatory properties of 2-amino-3H-phenoxazin-3-one. *Biol. Pharm. Bull.* **2008**, *31*, 1938—1945. DOI

39. Baidara, P.; Mandal, S. M. Bacteria and bacterial anticancer agents as a promising alternative for cancer therapeutics. *Biochimie* **2020**, *177*, 164-189. DOI

40. Bhansali, K. G.; Kook, A. M. Synthesis and characterization of a series of 5H-benzo[a]phenoxazin-5-one derivatives as potential antiviral/antitumor agents. *Heterocycles* **1993**, *36*, 1239-1251. DOI

41. Chau, N.-M.; Rogers, P.; Aherne, W.; Carroll, V.; Collins, I.; McDonald, E.; Workman, P.; Ashcroft, M. Identification of novel small molecule inhibitors of hypoxia-inducible factor-1 that differentially block hypoxia-inducible factor-1 activity and hypoxia-inducible factor-1 α induction in response to hypoxic stress and growth factors. *Cancer Res.* **2005**, *65*, 4918-4928. DOI

42. Lv, X.; Li, J.; Zhang, C.; Hu, T.; Li, S.; He, S.; Yan, H.; Tan, Y.; Lei, M.; Wen, M.; Zuo, J. The role of hypoxia-inducible factors in tumor angiogenesis and cell metabolism. *Genes Dis.* **2017**, *4*, 19-24. DOI

43. Li, Z.; You, Q.; Zhang, X. Small-Molecule modulators of the hypoxia-inducible factor pathway: development and therapeutic applications. *J. Med. Chem.* **2019**, *62*, 5725–5749. DOI

44. Sharma, A.; Arambula, J. F.; Koo, S.; Kumar, R.; Singh, H.; Sessler, J. L.; Kim, J.-S. Hypoxia-targeted drug delivery. *Chem. Soc. Rev.* **2019**, *48*, 771-813. DOI

45. Faig, M.; Bianchet, M. A.; Talalay, P.; Chen, S.; Winski, S.; Ross, D.; Amzel, L. M. Structures of recombinant human and mouseNAD(P)H:quinone oxidoreductases: species comparison and structural changes with substrate binding and release. *Proc. Natl. Acad. Sci. USA* **2000**, *97*, 3177-3182. DOI

46. Song, Y.; Wang, S.; Zhao, M.; Yang, X.; Yu, B. Strategies targeting protein tyrosine phosphatase SHP2 for cancer therapy. *J. Med. Chem.* **2022**, *65*, 3066-3079. DOI

47. Homan, K. T.; Balasubramaniam, D.; Zabell, A. P. R.; Wiest, O.; Helquist, P.; Stauffacher, C. V. Identification of novel inhibitors for a low molecular weight protein tyrosine phosphatase via virtual screening. *Bioorg. Med. Chem.* **2010**, *18*, 5449-5456. DOI

48. Defant, A.; Guella, G.; Mancini, I. Synthesis and *in vitro* cytotoxicity evaluation of novel naphthindolizinedione derivatives. *Arch. Pharm. Chem. Life Sci.* **2007**, *340*, 147–153. DOI

49. Yang, R.; Chen, Y.; Pan, L.; Yang, Y.; Zheng, Q.; Hu, Y.; Wang, Y.; Zhang, L.; Sun, Y.; Li, Z.; Meng, X. Design, synthesis and structure-activity relationship study of novel naphthoindolizine and indolizinoquinoline-5,12-dione derivatives as IDO1 inhibitors. *Bioorg. Med. Chem.* **2018**, *26*, 4886-4897. DOI

50. Feng, X.; Liao, D.; Liu, D.; Ping, A.; Li, Z.; Bian, J. Development of indoleamine 2,3-dioxygenase 1 inhibitors for cancer therapy and beyond: A recent perspective. *J. Med. Chem.* **2020**, *63*, 15115-15139. DOI

51. Ryu, C.-K.; Yoon, J.-H.; Song, A. L.; Im, H. A.; Kim, J. Y.; Kim, A. Synthesis and antifungal evaluation of pyrido[1,2-a]indole-1,4-diones and benzo[f]pyrido[1,2-a]indole-6,11-diones. *Bioorg. Med. Chem.* **2012**, *22*, 497-499. DOI

52. Székely, R.; Rengifo-Gonzalez, M.; Singh, V.; Riabova, O.; Benjak, A.; Piton, J.; Gimino, M.; Kornobus, E.; Mizrahi, V.; Johnsson, K.; Manina, G.; Makarov, V.; Cole, S. T. 6,11-Dioxobenzo[f]pyrido[1,2-a]indoles kill *mycobacterium tuberculosis* by targeting iron–sulfur protein Rv0338c (IspQ), a putative redox sensor. *ACS Infect. Dis.* **2020**, *6*, 3015-3025. DOI

53. Liu, Y.; Sun, J.-W. Copper(II)-catalyzed synthesis of benzo[f]pyrido[1,2-a]indole-6,11-dione derivatives via naphthoquinone difunctionalization reaction. *J. Org. Chem.* **2012**, *77*, 1191-1197. DOI

54. Sun, J.; Wang, F.; Hu, H.; Wang, X.; Wu, H.; Liu, Y. Copper(II)-catalyzed carbon–carbon triple bond cleavage of internal alkynes for the synthesis of annulated indolizines. *J. Org. Chem.* **2014**, *79*, 3992-3998. DOI

55. Matsuoka, M.; Iwamoto, A.; Kitao, T. Reaction of 2,3-dichloro-1,4-naphthoquinone with dithiooxamide. Synthesis of dibenzo[*b,i*]thianthrene-5,7,12,14-tetrone. *J. Heterocycl. Chem.* **1991**, *28*, 1445-1447. DOI

56. Trzoss, L.; Fukuda, T.; Costa-Lotufo, L. V.; Jimenez, P.; La Clair, J. J.; Fenical, W. Seriniquinone, a selective anticancer agent, induces cell death by autophagocytosis, targeting the cancer-protective protein dermcidin. *Proc. Natl. Acad. Sci. USA* **2014**, *111*, 14687-14692. DOI

57. Hammons, J. C.; Trzoss, L.; Jimenez, P. C.; Hirata, A. S.; Costa-Lotufo, L. V.; La Clair, J. J.; Fenical, W. Advance of seriniquinone analogues as melanoma agents. *ACS Med. Chem. Lett.* **2019**, *10*, 186-190. DOI

58. Burian, M.; Schittek, B. The secrets of dermcidin action. *Int. J. Med. Microbiol.* **2015**, *305*, 283-286. DOI

59. Nagao, H.; Ninomiya, M.; Sugiyama, H.; Itabashi, A.; Uno, K.; Tanaka, K.; Koketsu, M. Comparative analysis of *p*-terphenylquinone and seriniquinone derivatives as reactive oxygen species-modulating agents. *Bioorg. Med. Chem. Lett.* **2022**, *76*, 128992. DOI

60. Tuchinda, P.; Pohmakotr, M.; Munyoo, B.; Reutrakul, V.; Santisuk, T. An azaanthracene alkaloid from *Polyalthia suberosa*. *Phytochem.* **2000**, *53*, 1079-1082. DOI

61. Gandy, M. N.; Piggott, M. J. Synthesis of kalasinamide. A putative plant defense photoxin. *J. Nat. Prod.* **2008**, *71*, 866-868. DOI

62. Lang, S.; Groth, U. Total syntheses of cytotoxic, naturally occurring kalasinamide, geovanine, and marcanine A. *Angew Chem. Int. Ed.* **2009**, *48*, 911-913. DOI

63. Zhang, Y.-J.; Kong, M.; Chen, R.-Y.; Yu, D. Q. Alkaloids from the roots of *Goniothalamus griffithii*. *J. Nat. Prod.* **1999**, *62*, 1050-1052. DOI

64. Yin, J.; Landward, M. B.; Rainier, J. D. Photoelectrocyclization reactions of amidonaphthoquinones. *J. Org. Chem.* **2020**, *85*, 4298-4311. DOI

65. Soonthornchareonnon, N.; Suwanborirux, K.; Bavorada, R.; Patarapanich, C.; Cassady, J. M. New cytotoxic 1-azaanthraquinones and 3-aminonaphthoquinone from the stem bark of *Goniothalamus marcanii*. *J. Nat. Prod.* **1999**, *62*, 1390-1394. DOI

66. Sedmera, P.; Pospisil, S.; Novak, J. New furanonaphthoquinone from *Streptomyces cinnamomensis*. *J. Nat. Prod.* **1991**, *54*, 870-872. DOI

67. Bringmann, G.; Haagen, Y.; Gulder, T. A. M.; Gulder, T.; Heide, L. Biosynthesis of the isoprenoid moieties of furanonaphthoquinone I and endophenazine A in *Streptomyces cinnamomensis* DSM 1042. *J. Org. Chem.* **2007**, *72*, 4198-4204. DOI

68. Simamura, E.; Hirai, K.-I.; Shimada, H.; Koyama, J.; Niwa, Y.; Shimizu, S. Furanonaphthoquinones cause apoptosis of cancer cells by inducing the production of reactive oxygen species by the mitochondrial voltage-dependent anion channel. *Cancer Biol. Ther.* **2006**, *5*, 1523-1529. DOI

69. Tahara, T.; Watanabe, A.; Yutani, M.; Yamano, Y.; Sagara, M.; Nagai, S.; Saito, K.; Yamashita, M.; Ihara, M.; Iida, A. STAT3 inhibitory activity of naphthoquinones isolated from *Tabebuia avellanedae*. *Bioorg. Med. Chem.* **2020**, *28*, 115347. DOI

70. Feng, X.; Qiu, X.; Huang, H.; Wang, J.; Xu, X.; Xu, P.; Ge, R.; Liu, X.; Li, Z.; Bian, J. Palladium(II)-catalyzed reaction of Lawesones and propargyl carbonates: construction of 2,3-furanonaphthoquinones and evaluation as potential indoleamine 2,3-dioxygenase inhibitors. *J. Org. Chem.* **2018**, *83*, 8003-8010. DOI

71. Hu, X.; Li, J.; Fu, M.; Zhao, X.; Wang, W. The JAK/STAT signaling pathway: from bench to clinic. *Sig. Transduct. Target Ther.* **2021**, *6*, 402. DOI

72. Wu, Z.-Z.; Jang, Y.-J.; Lee, C.-J.; Lin, W. A versatile and practical method for the regioselective synthesis of polysubstituted furanonaphthoquinones. *Org. Biomol. Chem.* **2013**, *11*, 828-834. DOI

73. Lin, C.-Y.; Wu, H.-Y.; Hsu, Y.-L.; Cheng, T.-J. R.; Liu, J.-H.; Huang, R.-J.; Hsiao, T.-H.; Wang, C.-J.; Hung, P.-F.; Lan, A.; Pan, S.-H.; Chein, R.-J.; Wong, C.-H.; Yang, P.-C. Suppression of drug-resistant non-small-cell lung cancer with inhibitors targeting minichromosomal maintenance protein. *J. Med. Chem.* **2020**, *63*, 3172-3187. DOI

74. Tsai, C.-J.; Chen, C.-C.; Tsai, C.-W.; Wu, M. J. Base-mediated cyclization reaction of 2-(5-hydroxy-1-pentynyl)benzonitriles to 4-amino-2,3-dihydroronaphtho[2,3-*b*]furanes and synthesis of furanonaphthoquinones. *J. Org. Chem.* **2016**, *81*, 3882-3889. DOI

75. El-Feraly, F. S. Melampolides from *Magnolia grandiflora*. *Phytochem. (Elsevier)* **1984**, *23*, 2372-2374. DOI

76. Nasim, S.; Pei, S.; Hagen, F. K.; Jordan, C. T.; Crooks, P. A. Melampomagnolide B: a new antileukemic sesquiterpene. *Bioorg. Med. Chem.* **2011**, *19*, 1515-1519. DOI

77. Janganati, V.; Ponder, J.; Jordan, C. T.; Borrelli, M. J.; Penthala, N. R.; Crooks, P. A. Dimers of melampomagnolide B exhibit potent anticancer activity against hematological and solid tumor cells. *J. Med. Chem.* **2015**, *58*, 8896-8906. DOI

78. Pentala, N. R.; Balasubramaniam, M.; Dachavaram, S. S.; Morris, E. J.; Bhat-Nakshatri, P.; Ponder, J.; Jordan, C. T.; Nakshatri, H.; Crooks, P. A. Antitumor properties of novel sesquiterpene lactone analogs as NF κ B inhibitors that bind to the IKK β ubiquitin-like domain (ULD). *Eur. J. Med. Chem.* **2021**, *224*, 113675. DOI

79. Mischenko, N. P.; Fedoreyev, S. A.; Pokhilo, N. D.; Anufriev, V. P.; Denisenko, V. A.; Glazunov, V. P. Echinamines A and B, first aminated hydroxynaphthazarins from the sea urchin *scaphechinus mirabilis*. *J. Nat. Prod.* **2005**, *68*, 1390–1393. DOI

80. Mischenko, N. P.; Krylova, N. V.; Iunikhina, O. V.; Vasileva, E. A.; Likhatskaya, G. N.; Pislyagin, E. A.; Tarbeeva, D. V.; Dmitrenok, P. S.; Fedoreyev, S. A. Antiviral potential of sea urchin aminated spinochromes against herpes simplex virus type 1. *Mar. Drugs* **2020**, *18*, 550. DOI

81. You, H.; Vegi, S. R.; Lagishetti, C.; Chen, S.; Reddy, R. S.; Yang, X.; Guo, J.; Wang, C.; He, Y. Synthesis of bioactive 3,4-dihydro-2H-naphtho[2,3-*b*][1,4]oxazine-5,10-dione and 2,3,4,5-tetrahydro-1H-naphtho[2,3-*b*]azepine-6,11-dione derivatives via the copper-catalyzed intramolecular coupling reaction. *J. Org. Chem.* **2018**, *83*, 4119–4130. DOI

82. Kirkpatrick, D. L.; Powis, G. Inhibitors of redox signaling for restoration of apoptosis and inhibition of abnormal cell proliferation. WO2000006088 A2, October 2, 2000.

83. Liu, L.; Richard, J.; Kim, S.; Wojcik, E. J. Small molecule screen for candidate antimalarials targeting *plasmodium* kinesin-5. *J. Biol. Chem.* **2014**, *289*, 16601–16614. DOI

84. Tomek, P.; Palmer, B. D.; Flanagan, J. U.; Sun, C.; Raven, E. L.; Ching, L.-M. Discovery and evaluation of inhibitors to the immunosuppressive enzyme indoleamine 2,3-dioxygenase 1 (IDO1): probing the active site-inhibitor interactions. *Eur. J. Med. Chem.* **2017**, *126*, 983–996. DOI

85. Kulkarni, A.; Soni, I.; Kelkar, D. S.; Dharmaraja, A. T.; Sankar, R. K.; Beniwal, G.; Rajendran, A.; Tamhankar, S.; Chopra, S.; Kamat, S. S.; Chakrapani, H. Chemoproteomics of an indole-based quinone epoxide identifies druggable vulnerabilities in vancomycin-resistant *staphylococcus aureus*. *J. Med. Chem.* **2019**, *62*, 6785–6795. DOI

86. Bergamasco, R.; Porter, Q. N.; Yap, C. Vinylindenes and some heteroanalogues in the Diels-Alder reaction. V. 3-Vinylindoles and quinones. *Aust. J. Chem.* **1978**, *31*, 1841–1844. DOI

87. Kuo, C.-W.; Konala, A.; Lin, L.; Chiang, T.-T.; Huang, C.-Y.; Yang, T.-H.; Kavala, V.; Yao, C.-F. Synthesis of benzo[*a*]carbazole derivatives from 3-ethylindoles by exploiting the dual character of benzoquinone as an oxidizing agent and dienophile. *Chem. Commun.* **2016**, *52*, 7870–7873. DOI

88. Brimble, M. A.; Duncalf, L. J.; Nairn, M. R. Pyranonaphthoquinone antibiotics—isolation, structure and biological activity. *Nat. Prod. Rep.* **1999**, *16*, 267–281. DOI

89. Naysmith, B. J.; Hume, P. A.; Sperry, J.; Brimble, M. A. Pyranonaphthoquinones – isolation, biology and synthesis: an update. *Nat. Prod. Rep.* **2017**, *34*, 25–61. DOI

90. Bergy, M. E. Kalafungin, a new broad spectrum antibiotic: isolation and characterization. *J. Antibiot.* **1968**, *21*, 454–457. DOI

91. Kakinuma, S.; Ikeda, H.; Omura, S.; Hopwood, D. A. Biosynthesis of kalafungin in *Streptomyces tanashiensis*. *J. Antibiot.* **1990**, *43*, 391–396. DOI

92. Omura, S.; Tanaka, H.; Okada, Y.; Marumo, H. Isolation and structure of nanaomycin D, an enantiomer of the antibiotic kalafungin. *J. Chem. Soc., Chem. Commun.* **1976**, 320–321. DOI

93. Heapy, A. M.; Patterson, A. V.; Smaill, J. B.; Jamieson, S. M. F.; Guise, C. P.; Sperry, J.; Hume, P. A.; Rathwell, K.; Brimble, M. A. Synthesis and cytotoxicity of pyranonaphthoquinone natural product analogues under bioreductive conditions. *Bioorg. Med. Chem.* **2013**, *21*, 7971–7980. DOI

94. Tanaka, H.; Minami-Kakinuma, S.; Omura, S. Biosynthesis of nanaomycin. III. Nanaomycin A formation from nanaomycin D by nanaomycin D reductase via a hydroquinone. *J. Antibiot.* **1982**, *35*, 1565–1570. DOI

95. Salaski, E. J.; Krishnamurthy, G.; Ding, W.-D.; Yu, K.; Insaf, S. S.; Eid, C.; Shim, J.; Levin, J. I.; Tabei, K.; Toral-Barza, L.; Zhang, W.-G.; McDonald, L. A.; Honores, E.; Hanna, C.; Yamashita, A.; Johnson, B.; Li, Z.; Laakso, L.; Powell, D.; Mansour, T. S. Pyranonaphthoquinone lactones: A new class of AKT selective kinase inhibitors alkylate a regulatory loop cysteine. *J. Med. Chem.* **2009**, *52*, 2181–2184. DOI

96. Kuck, D.; Caulfield, T.; Lyko, F.; Medina-Franco, J. L. Nanaomycin A selectively inhibits DNMT3B and reactivates silenced tumor suppressor genes in human cancer cells. *Mol. Cancer Ther.* **2010**, *9*, 3015–3023. DOI

97. Tsuji, N.; Kobayashi, M.; Wakisaka, Y.; Kawamura, Y.; Mayama, M.; Matsumoto, K. New antibiotics, griseusins A and B. Isolation and characterization. *J. Antibiot.* **1976**, *29*, 7–9. DOI

98. Masamoto, K.; Takeuchi, Y.; Takeda, K.; Yoshii, E. Synthesis of pentacyclic structure of griseusin A. *Heterocycles* **1981**, *16*, 1659-1664. DOI

99. Brimble, M. A.; Nairn, M. R. Synthesis of the Griseusin A ring system. *J. Chem. Soc., Perkin Trans. 1* **1990**, 169-171. DOI

100. Brimble, M. A.; Nairn, M. R. Synthesis of a pyranonaphthoquinone-spiroacetal. *J. Chem. Soc., Perkin Trans. 1* **1992**, 579-583. DOI

101. Brimble, M. A.; Nairn, M. R.; Park, J. Synthesis of Analogues of Griseusin A. *Org. Lett.* **1999**, *1*, 1459-1462. DOI

102. Perry, N. B.; Blunt, J. W.; McCombs, J. D.; Munro, M. H. G. Discorhabdin C, a highly cytotoxic pigment from a sponge of the genus *Latrunculia*. *J. Org. Chem.* **1986**, *51*, 5476-5478. DOI

103. Hu, J.-F.; Fan, H.; Xiong, J.; Wu, S.-B. Discorhabdins and pyrroloiminoquinone-related alkaloids. *Chem. Rev.* **2011**, *111*, 5465-5491. DOI

104. Copp, B. R.; Fulton, K. F.; Perry, N. B.; Blunt, J. W.; Munro, M. H. G. Natural and synthetic derivatives of discorhabdin C, a cytotoxic pigment from the New Zealand sponge *Latrunculia* cf. *bocagei*. *J. Org. Chem.* **1994**, *59*, 8233-8238. DOI

105. Na, M. K.; Ding, Y.; Wang, B.; Tekwani, B. L.; Schinazi, R. F.; Franzblau, S.; Kelly, M.; Stone, R.; Li, X.-C.; Ferreira, D.; Hamann, M. T. Anti-infective discorhabdins from a deep-water Alaskan sponge of the genus *Latrunculia*. *J. Nat. Prod.* **2010**, *73*, 383-387. DOI

106. Goey, A. K. L.; Chau, C. H.; Sissung, T. M.; Cook, K. M.; Venzon, D. J.; Castro, A.; Ransom, T. R.; Henrich, C. J.; McKee, T. C.; McMahon, J. B.; Grkovic, T.; Cadelis, M. M.; Copp, B. R.; Gustafson, K. R.; Figg, W. D. Screening and biological effects of marine pyrroloiminoquinone alkaloids: potential inhibitors of the HIF-1 α /p300 interaction. *J. Nat. Prod.* **2016**, *79*, 1267-1275. DOI

107. Mąkosza, M.; Stalewski, J.; Maslennikova, O. S. Synthesis of 7,8-dimethoxy-4-oxo-1,3,4,5-tetrahydro-pyrrolo[4,3,2-*de*]quinoline. A key intermediate *en route* to makaluvamines, discorhabdin C and other marine alkaloids of this group via vicarious nucleophilic substitution of hydrogen. *Synthesis* **1997**, 1131-1133. DOI

108. Roberts, D.; Joule, J. A.; Bros, M. A.; Alvarez, M. Synthesis of pyrrolo[4,3,2-*de*]quinolines from 6,7-dimethoxy-4-methylquinoline. Formal total syntheses of damirones A and B, batzelline C, isobatzelline C, discorhabdin C, and makaluvamines A-D. *J. Org. Chem.* **1997**, *62*, 568-577. DOI

109. Kita, Y.; Tohma, H.; Inagaki, M.; Hatanaka, K.; Yakura, T. Total synthesis of discorhabdin C: a general aza spiro dienone formation from *O*-silylated phenol derivatives using a hypervalent iodine reagent. *J. Am. Chem. Soc.* **1992**, *114*, 2175-2180. DOI

110. Sadanandan, E. V.; Pillai, S. K.; LakshmiKantham, M. V.; Billimoria, A. D.; Culpepper, J. S.; Cava, M. P. Efficient syntheses of the marine alkaloids makaluvamine D and discorhabdin C: The 4,6,7-trimethoxyindole approach. *J. Org. Chem.* **1995**, *60*, 1800-1805. DOI

111. Aubart, K. M.; Heathcock, C. H. A Biomimetic approach to the discorhabdin alkaloids: total syntheses of discorhabdins C and E and dethiadicorhabdin D. *J. Org. Chem.* **1999**, *64*, 16-22. DOI

112. Kublak, G. G.; Confalone, P. N. The preparation of the aza-spirobicyclic system of discorhabdin C via an intramolecular phenolate alkylation. *Tetrahedron Lett.* **1990**, *31*, 3845-3848. DOI

113. Kadela-Tomanek, M.; Bebenek, E.; Chrobak, E.; Boryczka, S. 5,8-Quinolinedione scaffold as a promising moiety of bioactive agents. *Molecules* **2019**, *24*, 4115. DOI

114. Bolzán, A. D.; Bianchi, M. S. Genotoxicity of streptonigrin: a review. *Mutat. Res.* **2001**, *488*, 25-37. DOI

115. Rao, K. V.; Cullen, W. P. Streptonigrin, an antitumor substance, in: H. Welch, F. Martilhanez (Eds.), *Antibiotics Annual: 1959-1960*, Medical Encyclopedia, Inc., New York, 1960, pp. 950-953.

116. Doyle, T. W.; Balitz, D. M.; Grulich, R. E.; Nettleton, D. E.; Gould, S. J.; Tann, C.-H.; Moews, A. E. Structure determination of lavendamycin, a new antitumor antibiotic from *Streptomyces lavendulae*. *Tetrahedron Lett.* **1981**, *22*, 4595-4598. DOI

117. Behforouz, M.; Cai, W.; Stocksdale, M. G.; Lucas, J. S.; Jung, J.; Briere, D.; Wang, A.; Katen, K. S.; Behforouz, N. C. Novel lavendamycin analogues as potent HIV-reverse transcriptase inhibitors: Synthesis and evaluation of anti-reverse transcriptase activity of amide and ester analogues of lavendamycin. *J. Med. Chem.* **2003**, *46*, 5773-5780. DOI

118. Hassani, M.; Cai, W.; Koelsch, K. H.; Holley, D. C.; Rose, A. S.; Olang, F.; Lineswala, J. P.; Holloway, W. G.; Gerdes, J. M.; Behforouz, M.; Beall, H. D. Lavendamycin antitumor agents: Structure-based design, synthesis, and NAD(P)H:quinone oxidoreductase 1 (NQO1) model validation with molecular docking and biological studies. *J. Med. Chem.* **2008**, *51*, 3104-3115. DOI

119. Porter, T. H.; Skelton, F. S.; Folkers, K. Synthesis of new alkylamino- and alkylaminomethyl-5,8-quinolin-equinones as inhibitors of Coenzyme Q and as antimalarials. *J. Med. Chem.* **1971**, *14*, 1029-1033. DOI

120. Cossy, J.; Belotti, D.; Brisson, M.; Skoko, J. J.; Wipf, P.; Lazo, J. S. Biological evaluation of newly synthesized quinoline-5,8-quinones as Cdc25B inhibitors. *Bioorg. Med. Chem.* **2006**, *14*, 6283-6287. DOI

121. Santoso, K. T.; Menorca, A.; Cheung, C.-Y.; Cook, G. M.; Stocker, B. L.; Timmer, M. S. M. The synthesis and evaluation of quinolinequinones as anti-mycobacterial agents. *Bioorg. Med. Chem.* **2019**, *27*, 3532-3545. DOI

122. Egu, S. A.; Ibezim, A.; Onoabedje, E. A.; Okoro, U. C. Biological and *in silico* evaluation of quinolinedione and naphthoquinone derivatives as potent antibacterial agents. *ChemistrySelect* **2017**, *2*, 92222-9226. DOI

123. Ciftci, H. I.; Bayrak, N.; Yıldız, M.; Yıldırım, H.; Sever, B.; Tateishi, H.; Otsuka, M.; Fujita, M.; Tuyun, A. F. Design, synthesis and investigation of the mechanism of action underlying anti-leukemic effects of the quinolinequinones as LY83583 analogs. *Bioorg. Chem.* **2021**, *114*, 105160. DOI

124. Ryu, C.-K.; Jeong, H.-J.; Lee, S. K.; You, H. J.; Choi, K. U.; Shim, J. Y.; Heo, Y. H.; Lee, C.-O. Effects of 6-arylamino-5,8-quinolinediones and 6-chloro-7-arylamino-5,8-isoquinolinediones on NAD(P)H:quinone oxidoreductase (NQO1) activity and their cytotoxic potential. *Arch. Pharm. Res.* **2001**, *24*, 390-396. DOI

125. Fryatt, T.; Pettersson, H. I.; Gardipee, W. T.; Bray, K. C.; Green, S. J.; Slawin, A. M. Z.; Beall, H. D.; Moody, C. J. Novel quinolinequinone antitumor agents: structure-metabolism studies with NAD(P)H:quinone oxidoreductase (NQO1). *Bioorg. Med. Chem.* **2004**, *12*, 1667-1687. DOI

126. Schlager, J. J.; Powis, G. Cytosolic NAD(P)H:(quinone-acceptor)oxidoreductase in human normal and tumor tissue: effects of cigarette smoking and alcohol. *Int. J. Cancer* **1990**, *45*, 403-409. DOI

127. Cerecetto, H.; González, M.; Lavaggi, M. L.; Azqueta, A.; Lopez de Cerain, A.; Monge, A. Phenazine 5,10-dioxide derivatives as hypoxic selective cytotoxins. *J. Med. Chem.* **2005**, *48*, 21-23. DOI

128. Lavaggi, M. L.; Cabrera, M.; Pintos, C.; Arredondo, C.; Pachon, G.; Rodriguez, J.; Raymondo, S.; Pacheco, J. P.; Cascante, M.; Olea-Azar, C.; Lopez de Cerain, A.; Monge, A.; Cerecetto, H.; Gonzalez, M. Novel phenazine 5,10-dioxides release •OH in simulated hypoxia and induce reduction of tumour volume *in vivo*. *ISRN Pharmacol.* **2011**, 314209. DOI

129. Reddy, S. B.; Williamson, S. K. Tirapazamine: a novel agent targeting hypoxic tumor cells. *Expert Opin. Invest. Drugs* **2009**, *18*, 77-87. DOI

130. Abi-Jaoudeh, N.; Dayyani, F.; Chen, P. J.; Fernando, D.; Fidelman, N.; Javan, H.; Liang, P. -C.; Hwang, J. -I.; Imagawa, D. K. Phase I trial on arterial embolization with hypoxia activated tirapazamine for unresectable hepatocellular carcinoma. *J. Hepatocell. Carcinoma* **2021**, *8*, 421-434. DOI

131. Chen, E. H.; Tanabe, K.; Saggiomo, A. J.; Nodiff, E. A. Modifications of primaquine as antimalarials. 4. 5-Alkoxy derivatives of primaquine. *J. Med. Chem.* **1987**, *30*, 1193-1199. DOI

132. Shelke, Y. G.; Hande, P. E.; Gharpure, S. J. Recent advances in the synthesis of pyrrolo[1,2-*a*]indoles and their derivatives. *Org. Biomol. Chem.* **2021**, *19*, 7544-7574. DOI

133. Bass, P. D.; Gubler, D. A.; Judd, T. C.; Williams, R. M. Mitomycinoid alkaloids: Mechanism of action, biosynthesis, total syntheses, and synthetic approaches. *Chem. Rev.* **2013**, *113*, *8*, 6816-6863. DOI

134. Liu, L.; Wang, Y.; Zhang, L. Formal synthesis of 7-methoxymitosene and synthesis of its analog via a key PtCl₂-catalyzed cycloisomerization. *Org. Lett.* **2012**, *14*, 3736-3739. DOI

135. Zheng, Z.; Touve, M.; Barnes, J.; Reich, N.; Zhang, L. Synthesis-enabled probing of mitosene structural space leads to improved IC₅₀ over mitomycin C. *Angew. Chem. Int. Ed.* **2014**, *53*, 9302-9305. DOI

136. Ayala, C. E.; Villalpando, A.; Nguyen, A. L.; McCandless, G. T.; Kartika, R. Chlorination of aliphatic primary alcohols via triphosgene-triethylamine activation. *Org. Lett.* **2012**, *14*, 3676-3679. DOI

137. Sweeney, M. Hydrogen peroxide and hydrohalic acid mediated synthesis of halogenated benzimidazolequinones and anti-cancer evaluation of benzotriazinones. Ph.D., National University of Ireland Galway, Ireland, 2019.

138. Lee, S.-R.; Yang, K.-S.; Kwon, J.; Lee, C.; Jeong, W.; Rhee, S. G. Reversible inactivation of the tumor suppressor PTEN by H₂O₂. *J. Biol. Chem.* **2002**, *277*, 20336-20342. DOI

139. Smith, S. L.; Pitt, A. R.; Spickett, C. M. Approaches to investigating the protein interactome of PTEN. *J. Proteome Res.* **2021**, *20*, 60-77. DOI

140. Meuillet, E. J.; Mahadevan, D.; Berggren, M.; Coon, A.; Powis, G. Thioredoxin-1 binds to the C2 domain of PTEN inhibiting PTEN's lipid phosphatase activity and membrane binding: a mechanism for the functional loss of PTENs tumor suppressor activity. *Arch. Biochem. Biophys.* **2004**, *429*, 123-133. DOI

141. Duan, D.; Zhang, J.; Yao, J.; Liu, Y.; Fang, J. Targeting thioredoxin reductase by parthenolide contributes to inducing apoptosis of HeLa Cells. *J. Biol. Chem.* **2016**, *291*, 10021-10031. DOI

142. Madkour, M. M.; Anbar, H. S.; El-Gamal, M. I. Current status and future prospects of p38a/MAPK14 kinase and its inhibitors. *Eur. J. Med. Chem.* **2021**, *213*, 113216. DOI

143. Paz, M. M.; Zhang, X.; Lu, J.; Holmgren, A. A new mechanism of action for the anticancer drug mitomycin C: mechanism-based inhibition of thioredoxin reductase. *Chem. Res. Toxicol.* **2012**, *25*, 1502–1511. DOI

144. Newsome, J. J.; Swann, E.; Hassani, M.; Bray, K. C.; Slawin, A. M. Z.; Beall, H. D.; Mood, C. J. Indolequinone antitumour agents: correlation between quinone structure and rate of metabolism by recombinant human NAD(P)H:quinone oxidoreductase. *Org. Biomol. Chem.* **2007**, *5*, 1629-1640. DOI

145. Yan, C.; Siegel, D.; Newsome, J. J.; Chilloux, A.; Moody, C. J.; Ross, D. Antitumor indolequinones induced apoptosis in human pancreatic cancer cells via inhibition of thioredoxin reductase and activation of redox signaling. *Mol. Pharmacol.* **2012**, *81*, 401-410. DOI