

Review

Not peer-reviewed version

3D Bioprinting in Cardiovascular Disease: Current Status and Future Directions

[Zhonghua Sun Sun](#)^{*}, Jack Zhao, Emily Leung, Maria Flandes-Iparraguirre, [Michael Vernon Vernon](#), Jenna Silberstein, [Elena M. De-Juan-Pardo](#), Shirley Jansen

Posted Date: 2 June 2023

doi: 10.20944/preprints202306.0178.v1

Keywords: 3D bioprinting; cardiovascular disease; cells; tissues; 3D printing



Preprints.org is a free multidiscipline platform providing preprint service that is dedicated to making early versions of research outputs permanently available and citable. Preprints posted at Preprints.org appear in Web of Science, Crossref, Google Scholar, Scilit, Europe PMC.

Copyright: This is an open access article distributed under the Creative Commons Attribution License which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Review

3D Bioprinting in Cardiovascular Disease: Current Status and Future Directions

Zhonghua Sun ^{1,2,*}, Jack Zhao ³, Emily Leung ³, Maria Flandes-Iparraguirre ⁴, Michael Vernon ^{5,6,7}, Jenna Silberstein ¹, Elena M. De-Juan-Pardo ⁷ and Shirley Jansen ^{8,9,10,11}

¹ Discipline of Medical Radiation Science, Curtin Medical School, Curtin University, Perth, 6102, WA, Australia; jenna.beinart@student.curtin.edu.au

² Curtin Health Innovation Research Institute (CHIRI), Curtin University, Perth, 6102, WA, Australia

³ School of Medicine, Faculty of Health Sciences, The University of Western Australia, Perth, 6009, WA, Australia; jack.zhao@health.wa.gov.au (J.Z.); emily.leung@health.wa.gov.au (E.L.)

⁴ Regenerative Medicine Program, Cima Universidad de Navarra, Pamplona, Spain; mflandes@alumni.unav.es

⁵ T3mPLATE, Harry Perkins Institute of Medical Research, QEII Medical Centre and UWA Centre for Medical Research, The University of Western Australia, Perth, WA, 6009, Australia; michael.vernon@research.uwa.edu.au

⁶ Vascular Engineering Laboratory, Harry Perkins Institute of Medical Research, QEII Medical Centre and UWA Centre for Medical Research, The University of Western Australia, Perth, WA, 6009, Australia

⁷ School of Engineering, The University of Western Australia, Perth, WA, 6009 Australia; elena.juanpardo@uwa.edu.au

⁸ Curtin Medical School, Curtin University, Perth, 6102, WA, Australia; shirley.jansen@health.wa.gov.au

⁹ Department of Vascular and Endovascular Surgery, Sir Charles Gairdner Hospital, Perth, 6009, WA, Australia

¹⁰ Heart and Vascular Research Institute, Harry Perkins Medical Research Institute, Perth, 6009, WA, Australia

¹¹ School of Medicine, The University of Western Australia, Perth, 6009, WA, Australia

* Correspondence: z.sun@curtin.edu.au; Tel: +61 8 9266 7509

Abstract: 3D printing is playing an important role in cardiovascular disease by use of personalised models with replication of normal anatomy and pathology with high accuracy and reliability. While 3D printed heart and vascular models are shown to improve medical education, preoperative planning and simulation of cardiac procedures, and enhance communication with patients, 3D bioprinting however, represents a potential advancement of 3D printing technology by printing cellular or biological components, functional tissues and organs that can be used in a variety of applications in cardiovascular disease. Recent advances in bioprinting technology have shown the ability to support vascularisation of large scale constructs with enhanced biocompatibility and structural stability, thus creating opportunities to replace damaged tissues or organs. In this review, we provide an overview of the use of 3D bioprinting in cardiovascular disease with a focus on technologies and applications in cardiac tissues, vascular constructs and grafts, heart valves and myocardium. Limitations and future research directions are highlighted.

Keywords: 3D bioprinting; cardiovascular disease; cells; tissues; 3D printing

1. Introduction

Cardiovascular disease (CVD) is the leading cause of death and major contributor to disability worldwide, with a prevalence of 523 million people who suffer from CVD and an estimated 18.6 million deaths annually [1]. Clinical diagnosis and management of CVD largely relies on less invasive imaging modalities, such as use of computed tomography (CT), magnetic resonance imaging and echocardiography [2,3]. Despite being frequently used in daily practice, it is challenging to translate

two-dimensional images of cardiovascular anatomy and pathology to three-dimensional (3D) structures based on these imaging modalities and this is especially obvious when visualising complex cardiovascular structures and pathologies, such as congenital heart disease which usually involves multiple cardiac structures with a spectrum of abnormalities. Three-dimensional (3D) printing technology has overcome these limitations by providing realistic models, thus offering superior advantages over current image visualisations [4–10].

Use of 3D printing in cardiovascular disease has been increasingly reported in the literature with applications mainly focusing on these areas: education value for medical students and health professionals, preoperative planning and simulation of complex cardiovascular procedures, development of medical devices for improvement of outcomes, optimising CT protocols using 3D phantoms to minimise radiation exposure to patients during routine CT scans for diagnosis and follow up, bioprinting of scaffolds or cardiovascular constructs [11–18]. Figure 1 summarises these current applications. 3D printed models serve as valuable tools for medical education by enhancing understanding of anatomy and pathology compared to current education approaches. Nearly 50% of the current applications of 3D printing lie in pre-surgical planning of cardiovascular procedures, with results from single and multi-centre studies showing changes in surgical decision-making with the use of 3D printed models [7,14,19–27]. 3D printed models are also used to simulate interventional cardiology or radiology procedures, such as endovascular aneurysm repair, increasing confidence through training and practice [28–40].

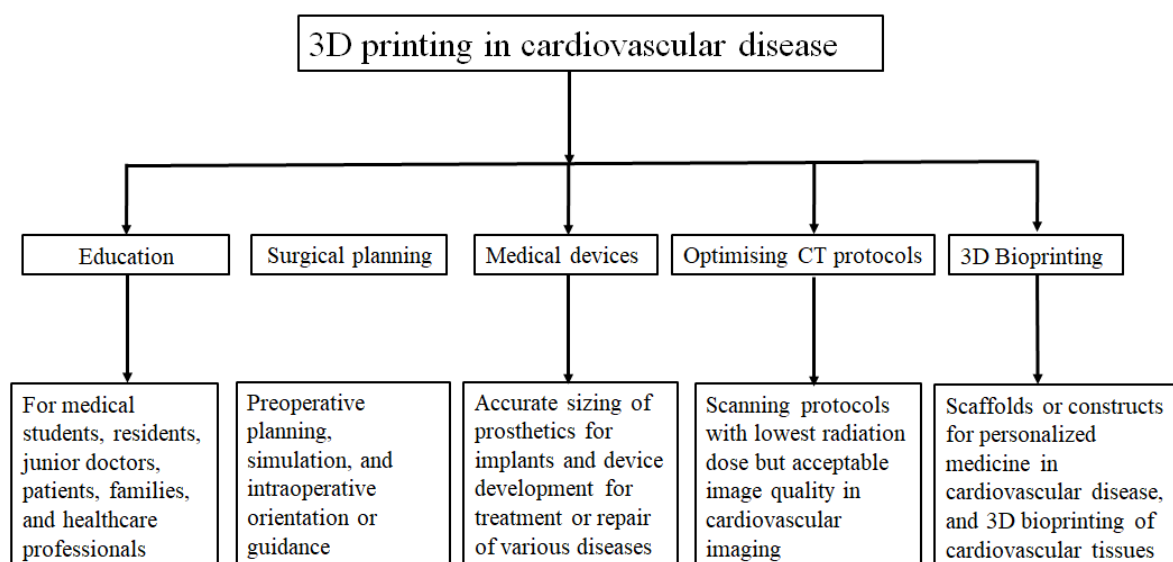


Figure 1. Flow chart showing applications of 3D printing including bioprinting in cardiovascular disease. Reprinted with permission under open access from Sun [9].

While there is sufficient evidence available to prove that 3D printed personalised heart and vascular models play an important role in clinical applications, 3D bioprinting represents a promising field that could revolutionise the future of treating cardiovascular disease. 3D bioprinting uses mostly the same additive manufacturing technologies but prints functional living structures from biological components such as living cells, biomaterials and growth factors [41,42]. This could eliminate the gap between the current animal study and human trials, and may in future contribute to repair or replace damage tissues or organ and optimise drug delivery techniques [43–47]. Although still in an early stage with many challenges to overcome, substantial progress has been made over the last decade in printing cardiovascular constructs as well as cardiovascular regeneration [48–50].

This review provides an overview of 3D bioprinting in cardiovascular disease, with a focus on the applications in cardiac tissues, vascular constructs and grafts, myocardium and heart valves.

2. 3D bioprinting: where are we now?

Heart transplant remains the gold standard treatment for selected people with end-stage heart failure [48]. However, the number of people requiring a heart transplant far outweighs the available donor hearts and the long-term outcome of heart transplantation remains uncertain, with acute rejection, cardiac allograft vasculopathy, malignancies, infection and the development of chronic kidney disease still significant complications [49,50].

Apart from organ transplantation, current treatment options for cardiac disease include cell therapy, implanted devices (stents or stent grafts), and bypass grafting [51–53]. Of these treatments, cell therapy using 3D bioprinting with patient-specific cells to repair damaged cardiac tissues or replace end-stage organ failure through tissue engineering is promising [48,54], and is theoretically able to bypass the limitations of poor biocompatibility, biofunctionality, immune rejection as well as the drastic shortage of organ donors [55–58].

Although recent advances in bioprinting methodologies and technologies have addressed limitations associated with bioprinting, it is still not clinically feasible to directly translate 3D printed cardiovascular tissues to patient therapy. The main limitation is the inability to construct thick tissues. Currently, only 1 mm tissues can be produced with incorporation of multiple cell types and induction of tissue differentiation of growth factors [59,60]. A further limitation is a lower density of cells compared to native tissues restricting the clinical value such as in myocardial ischemia [61].

3. 3D bioprinting-bioinks

3D bioprinting involves deposition of living cells, bioactive molecules and biomaterials in a layered pattern to allow generation of 3D structures [62,63]. Different from the standard 3D printing process which uses various printing materials from rigid to soft and elastic depending on usefulness or application, bioinks serve as a medium to deliver living cells [64]. Bioinks possess viscoelastic properties with higher water content to protect the cells during the printing procedure from external risk factors. Bioinks used for cardiac tissue engineering mainly refer to the use of hydrogels which mimic the 3D extracellular matrix, with its: biocompatibility, biodegradability and mechanical support [65].

4. 3D bioprinting technologies

Bioprinting methods include inkjet-based, extrusion-based, and light-based printing to create geometrically complex and scalable tissues.

4.1. Inkjet-based bioprinting

Inkjet bioprinters store bioink in a cartridge and printer head; printer heads are deformed by thermal, piezoelectric, electrostatic or electrohydrodynamic actuators to eject droplets in a controlled fashion [63]. This is a low-cost method with high printing speed [64] and around 80–90% for fibroblast cell viability [65,66], but is limited by a low range of printable viscosities (<30 mPa.s) [67].

4.2. Extrusion-based bioprinting

Extrusion-based bioprinting is the most common technique used. Bioink is loaded into a syringe, then printed as continuous strands using mechanical or pneumatic forces [68]. This can print biomaterials with a wide range of viscosities (up to $>6 \times 10^7$ mPa/s) [69] and a high level of structural integrity [70]. However, cell viability decreases with increasing pressure and/or decreasing nozzle diameter [71]. Although extrusion-based bioprinting is a convenient and inexpensive platform with reasonable costs, it has limited resolution which is inferior to other bioprinting techniques.

4.3. Light-based bioprinting

Laser-assisted bioprinting projects a laser beam onto a thin layer of bioink causing a precise droplet to fall onto the printing surface [72]. Xiong et al [73] successfully printed both straight and Y-

shaped vascular constructs; however, fibroblast cell viability was 68.1% and 70.8% for these constructs, respectively. The main advantages of laser-based bioprinting include high resolution, the ability to produce cardiac constructs with a high cell density (up to 10^8 cells per mL), and bioprinting with low viscosity bioinks. The main disadvantages are complexity in controlling laser pulses, challenges in fabricating cell-embedding hydrogel constructs before printing and high cost of the laser system.

5. 3D bioprinting cardiac tissues

5.1. Human pluripotent stem cells and cardiac tissue engineering

Recent advancements in bioprinting of the cardiovascular system includes areas involving human pluripotent stem cells (hPSCs) and cardiac tissue engineering [74]. Promising benefits have been shown in using current hPSCs and cardiac tissue engineering technologies to construct biocompatible materials for the repair and regeneration of diseased cardiac tissue. Traditionally, cardiomyocytes are among those that are known to be difficult to expand and culture. As reprogramming of differentiated human cells back into a pluripotent state has been made possible in recent decades, the potential for creating patient and disease-specific stem cells in nearly any patient is becoming a possibility for the future [75,76]. Additionally, hPSCs technologies show great potential over conventional 2D monolayer culture systems for studying the development of the human heart, the heart's response to therapeutic interventions in drug testing, and for disease modelling of acquired and inherited diseases [74,77,78]. Figure 2 provides a schematic view of the application of cardiac tissue engineering for disease modeling and drug screening in genetic heart diseases. Cardiovascular tissue engineering is a rapidly evolving field of medicine which involves multiple sophisticated components such as different cell types, biocompatible scaffold materials, cell differentiation factors and growth factors [79]. As this is an area of research that is still in its infancy, there are several issues which are yet to be solved.

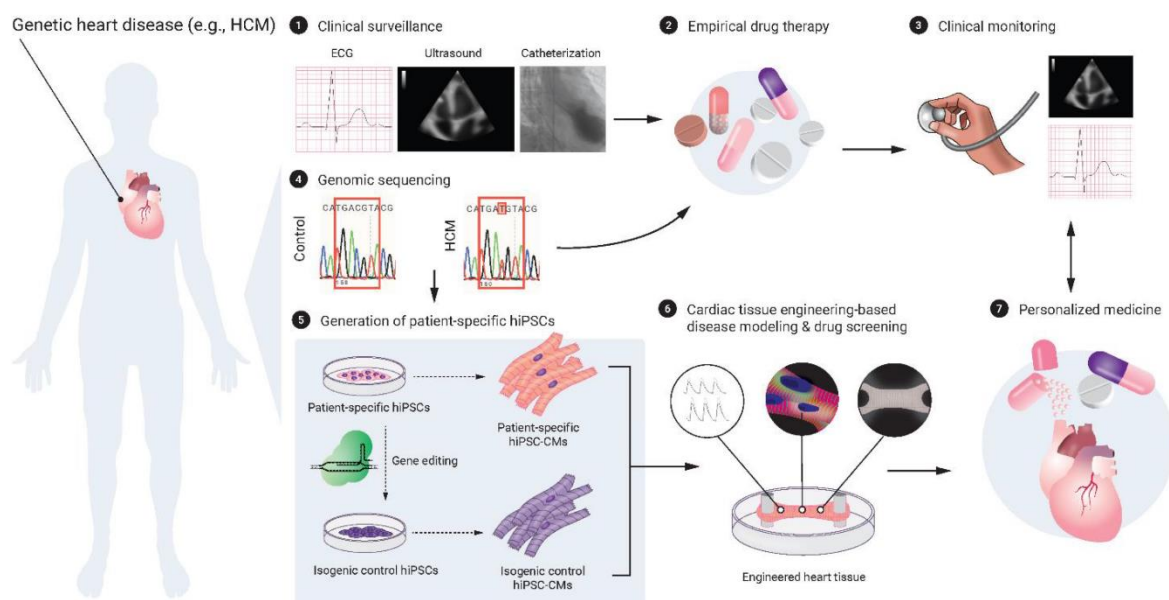


Figure 2. Schematic diagram highlighting the application of cardiac tissue engineering in the modeling of personalized medicine for genetic heart diseases. Reprinted with permission under open access from Häneke et al [74]. CM, cardiomyocyte; HCM; hypertrophic cardiomyopathy; hiPSC, human inducible pluripotent stem cell.

5.2. Cellular maturity

hPSCs such as human-embryonic stem cells (hESCs) and human-induced pluripotent stem cells (hiPSCs) are promising approaches for heart regeneration and have the potential to be differentiated efficiently into cardiomyocytes, vascular endothelial cells, and vascular smooth muscle cells [74]. Although there is much international research in this space, there are concerns over the immature properties of the tissues created [75]. The maturity of current hPSC-derived cardiomyocytes resemble characteristics of fetal cardiomyocytes [80].

To promote maturation, many approaches have been proposed, including the use of biochemical stimuli (such as hormonal treatments), mechanical strain stimulation, electrical stimulation, topographic cues such as patterned surfaces and substrate stiffness, and interactions with other cells or extracellular matrix [77,78,81]. However, the optimal state of cellular maturity for hPSC cardiomyocytes is yet to be determined; immature cardiomyocytes may cause ventricular arrhythmias and other electrical disturbances, but have increased survival compared to adult cardiomyocytes upon transplantation into a host myocardium [74].

5.3. Microvasculature constructs

Creating functional microvasculature constructs is a challenging area in 3D bioprinting of complex tissues, such as the heart which is highly vascularised to allow for adequate gas exchange, nutrient diffusion and waste disposal [82]. As diffusion of oxygen through tissues is limited to approximately 100-200 μm , careful incorporation of microvasculature into bioprinted materials is a fundamental step for tissue survival [82]. Characteristics which are fundamental for proper microvascular perfusion to tissues include the presence of a hollow and endothelialised lumen, a hierarchy-based branched vascular network, and a complex signalling milieu [83]. Many biofabrication technologies such as extrusion-based 3D bioprinting, induced sprouting, electrospinning, and lithographic approaches have been explored to address these issues, each with their own advantages and disadvantages [83,84]. These techniques generally involve microfluidics-based molding with numerous other strategies such as the addition of proangiogenic factors (e.g vascular endothelial growth factor [VEGF]) within the biomaterial scaffolding [85]. Figure 3 is a schematic framework proposed by Seymour et al. to address the current issues in biofabricating microvasculature [83].

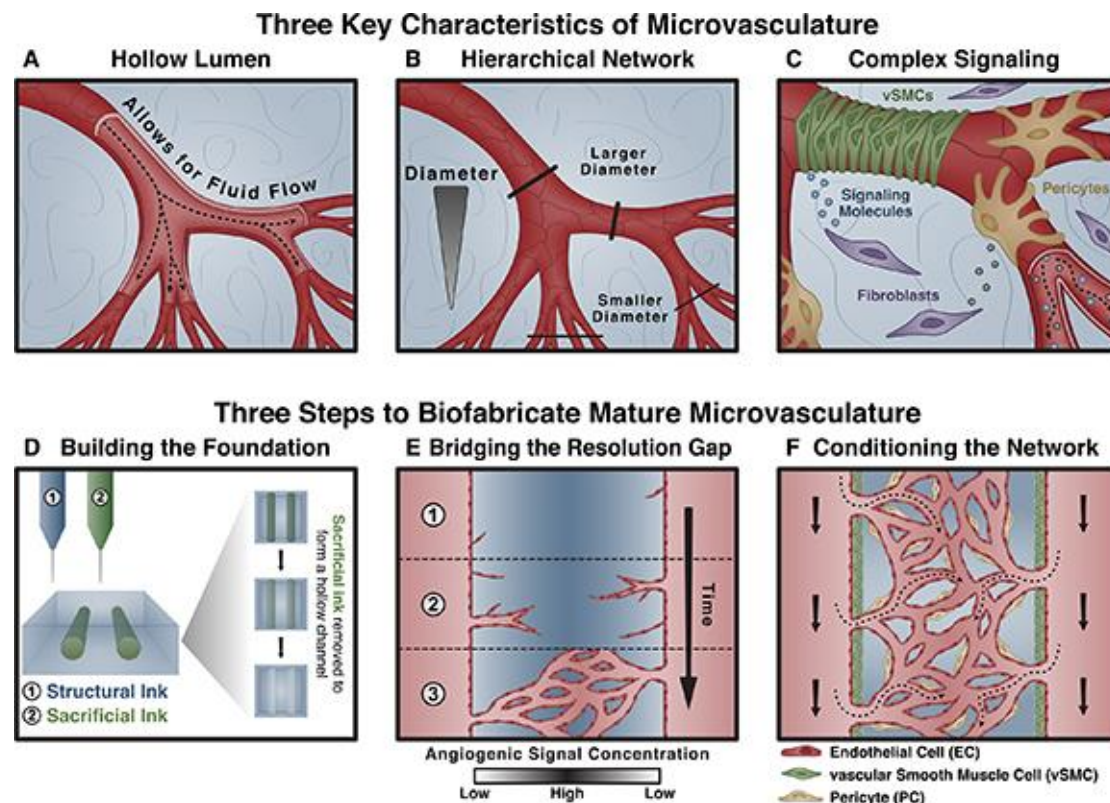


Figure 3. Schematic showing the key characteristics and steps to biofabricate mature microvasculature. (A) the presence of a hollow lumen to allow for blood flow; (B) hierarchal-based branched network; and (C) a complex signalling milieu. Seymour et al. proposes three steps to biofabricate mature microvasculature involving (D) an extrusion-based bioprinting technique to create the hollow lumen foundation, (E) use of proangiogenic signaling for capillary formation, and (F) stabilisation of the vascular network with supporting cell types. Reprinted with permission from Seymour et al [83].

Despite rapid improvement in mimicking the vascular networks of the human cardiovascular system, there is yet to develop an approach which is able to address all the required characteristics of microvasculature in order to maintain perfusion and survival of tissues. Precisely, the spatiotemporal precision required for appropriate physiological function is still lacking [85]. Further exploration of biomaterials and bioprinting strategies for addressing the structural and physiological complexity of microvascular networks will be a key stepping-stone in the research field of 3D bioprinting of cardiovascular tissues.

5.4. Other issues

Other issues that are faced in 3D bioprinting of cardiovascular tissue include but are not limited to cellular heterogeneity in different anatomical regions of the heart [74], proliferation of primary cells in vitro entering senescence [81], foreign body reaction [86], and the risk of tumours such as teratomas in undifferentiated cell populations [87].

Although we are still far from a safe and functional 3D bioprinted cardiovascular construct, this field of research has been developing and improving rapidly. Future studies should focus on tackling the critical issues of cardiomyocyte maturation, improved microvasculature constructs as well as creating a safe microenvironment for cell culture and biocompatibility.

6. 3D bioprinting of vascular constructs and grafts

Vascular grafts are vital in surgical treatment of coronary artery disease and peripheral vascular disease; arterio-venous fistulae in haemodialysis; and repair of large-vessel aneurysms and congenital defects [88]. Autologous grafts remain the gold standard; however, 30% of patients lack

suitable grafts [89,90]. Synthetic grafts suffer significantly lower patency than autologous equivalents [91] and are limited in <5mm-vessel repairs by high incidence of thrombosis [90]. 3D bioprinting to create tissue-engineered vascular grafts (TEVGs) offers a promising future alternative to produce individualised grafts.

6.1. Requirements of a TEVG

TEVGs must be able to generate functional endothelium; be non-immunogenic; and possess similar mechanical properties to human vessels [92]. Vascular grafts should be tested for several mechanical properties. Burst pressure is the maximum pressure a vessel can withstand, and is measured with burst pressure or burst probe testing [93]. Vessels must be compliant in order to accommodate pulsatile blood flow. Suture-ability of grafts is measured with suture retention testing, the maximum force required to cause the suture or graft wall to fail [94]. TEVGs also require an intact layer of endothelial cells (ECs), tested using cell viability studies [95].

6.2. Methods of 3D bioprinting of TEVGs

Biological grafts require complex structures, including branched and cellular vessels. Inkjet-based printing is either 'vertical' or 'horizontal': if the nozzle's primary direction of movement is parallel to the vessel's circumference, it is vertical; in horizontal printing, the primary direction of movement is parallel to the vessel's longitudinal axis [96]. Christensen et al [65] successfully combined vertical and horizontal inkjet bioprinting to create vessels with multi-planar bifurcations. Fibroblast cell viability was >90% after 24-hour incubation. Cellular tubes ('zigzag' tubes), have also been successfully produced using inkjet techniques, with a cell viability of >82% after a 72-hour incubation [97].

Despite these advances in inkjet bioprinting, its use for TEVGs has been limited. Inkjet printer head orifices can be clogged by bioinks of higher viscosity; the use of lower viscosity bioinks is unfavourable for mechanical strength [98], a quality essential for vascular grafts.

Hinton et al [99] used extrusion-based bioprinting with a 'freeform reversible embedding of suspended hydrogels' (FRESH) support bath technique to create part of the right coronary artery, with accurate anatomical and mechanical fidelity to the 3D model. The support bath behaved as a rigid body at low shear stresses, but as a fluid at higher shear stresses, allowing for low resistance to the printer nozzle and high resistance to movement of the printed construct (Figure 4).

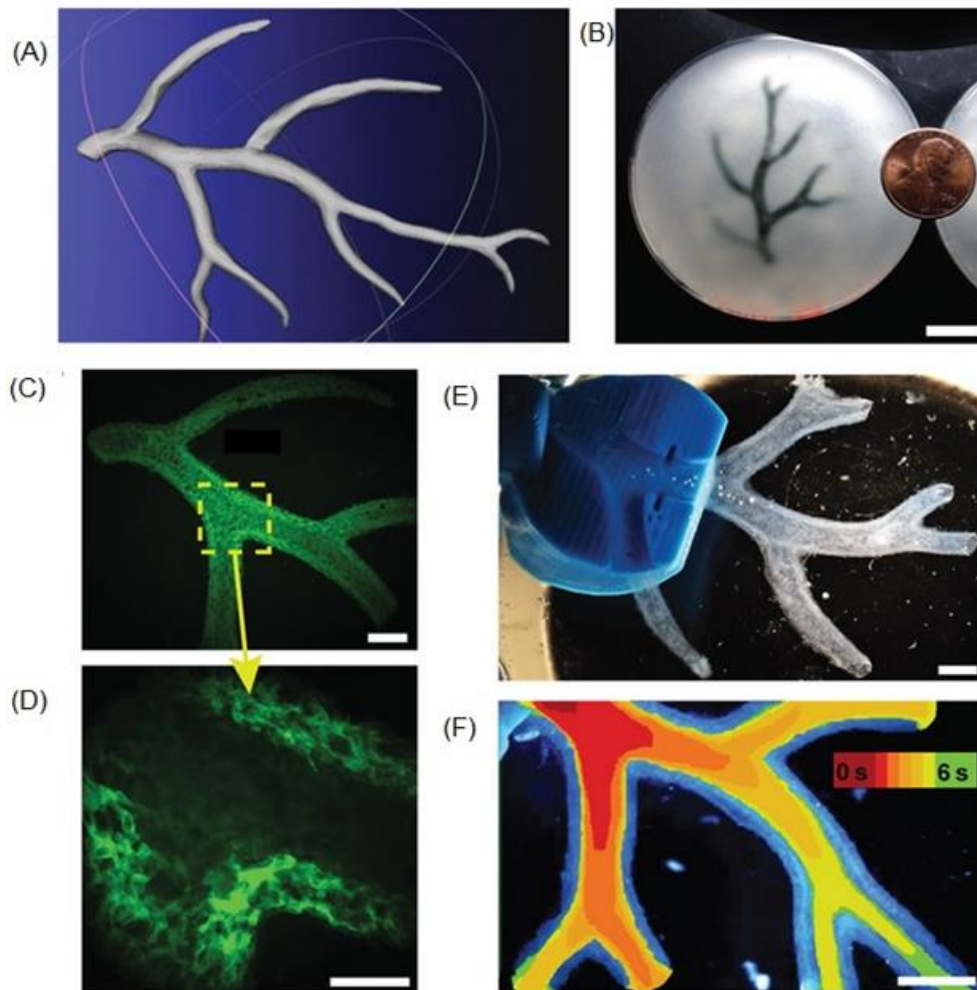


Figure 4. (A) model of a section of the right coronary arterial tree, printed with FRESH. (B) The arterial tree printed in alginate (black) and embedded in the support bath. (C) A section of the arterial trees printed in fluorescent alginate (green), demonstrating a hollow lumen. (D) A zoomed-in view, showing the vessel wall of <1mm thickness. (E) A dark-field image of the arterial tree mounted in a perfusion fixture to position a syringe in the root of the tree. (F) A time-lapse image of black dye perfused through the arterial tree demonstrating no leaks through the wall. Reprinted with permission under open access from Hinton et al [99].

Co-axial extrusion bioprinting utilises multiple distinct nozzles arranged coaxially, in order to print several concentric layers of biomaterial [100]. Hong et al [101] bioprinted vascular-like structures using a rapid-gelling gelatin-PEG-tyramine polymer with a dual coaxial nozzle. Viability of printed cells (fibroblast and endothelial cells) was 80–90%, and patency was confirmed with flushing of trypan-blue solution through the tube.

Composite bioinks can improve stability [102]; alginate and carboxymethylcellulose were combined by Milojević et al [103] to print vessels with good biocompatibility and sufficient mechanical stability (Figure 5). Wang et al [104] and Liu et al [105] (Figure 6) added gelatin methacryloyl (GelMA) to alginate, producing vessels with enhanced EC proliferation, adhesion, and spreading; the alginate sheath provided mechanical strength. Jia et al [106] used a novel bioink of GelMA, sodium alginate, and 4-arm poly(ethylene glycol)-tetra-acrylate. The combination of three bioinks stabilised the vessel and supported the maturation of vascular cells; their triaxial printing technique was able to achieve a wide range of diameters and wall thicknesses; and the printed vessels maintained high fidelity to their models, even when printing tortuous structures.

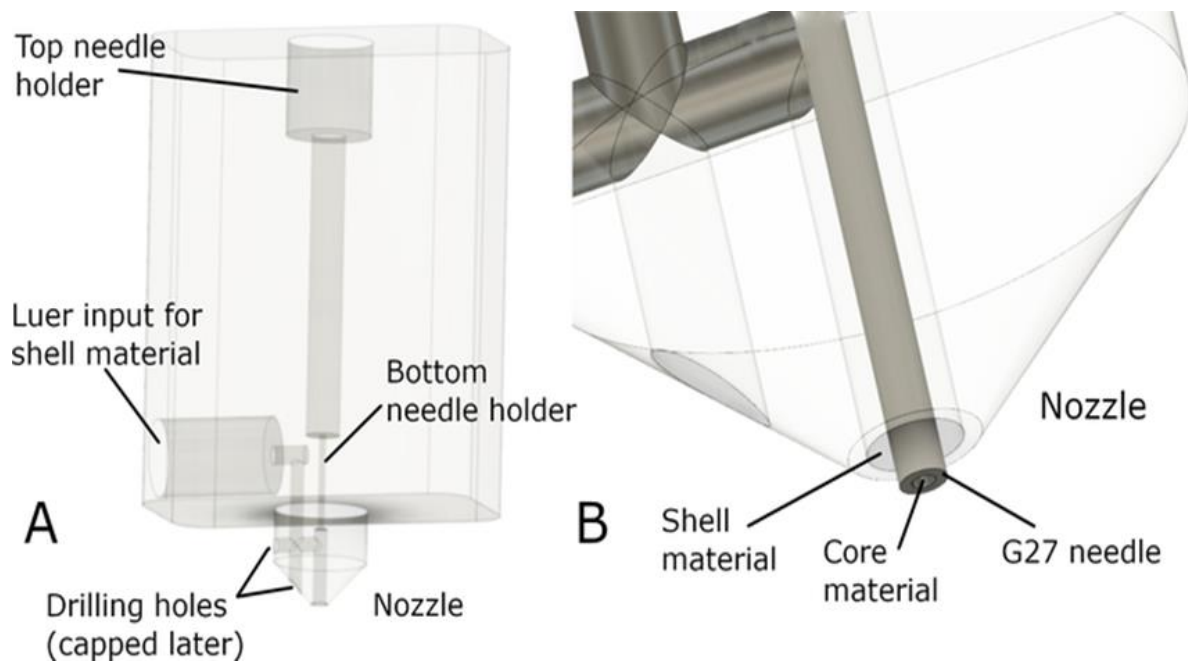


Figure 5. Core/Shell set-up for co-axial bioprinting using composite bioinks of alginate and carboxymethylcellulose. (A) Overall design. (B) Close-up of co-axial nozzle set-up. Reprinted with permission under open access from Milojevic et al [103].

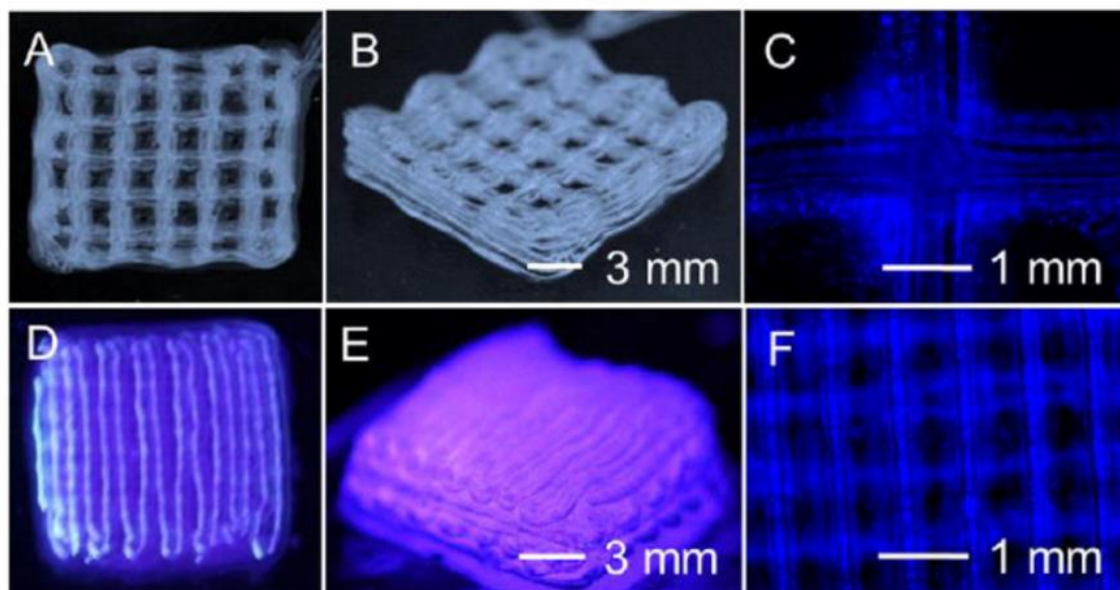


Figure 6. Bioprinting of 3D hollow microfibrous constructs. (A-F) A 10-layer construct with a distance between the continuous microfibers of (A)-(C) 2.3 mm, and (D)-(F) 0.0 mm. GelMA and alginate producing hollow perfusable vessels with good biocompatibility and mechanical strength. Reprinted with permission from Liu et al [105].

Mandrel-based printing uses a rotating rod as the support structure for the surrounding printed vessel. Gao et al [107] used coaxial nozzles to print hollow alginate filaments in a spiral configuration along the rotating support rod, creating a macro-channel (equivalent to the vessel lumen) and microchannels within the vessel wall. This approach produced biocompatible blood vessels with relatively strong mechanical strength. In a recent study, Jin et al [108] used a novel approach: nanofibre electrospinning to create an inner EC tube, and mandrel-based extrusion printing to create an outer layer of smooth muscle (SM) cells (Figure 7). This process yielded >90% cell viability, tensile strain sufficient to withstand blood pulsation, and suture retention strength greater than the common

carotid artery. The combination of biocompatibility and mechanical strength render these processes promising for clinical application.

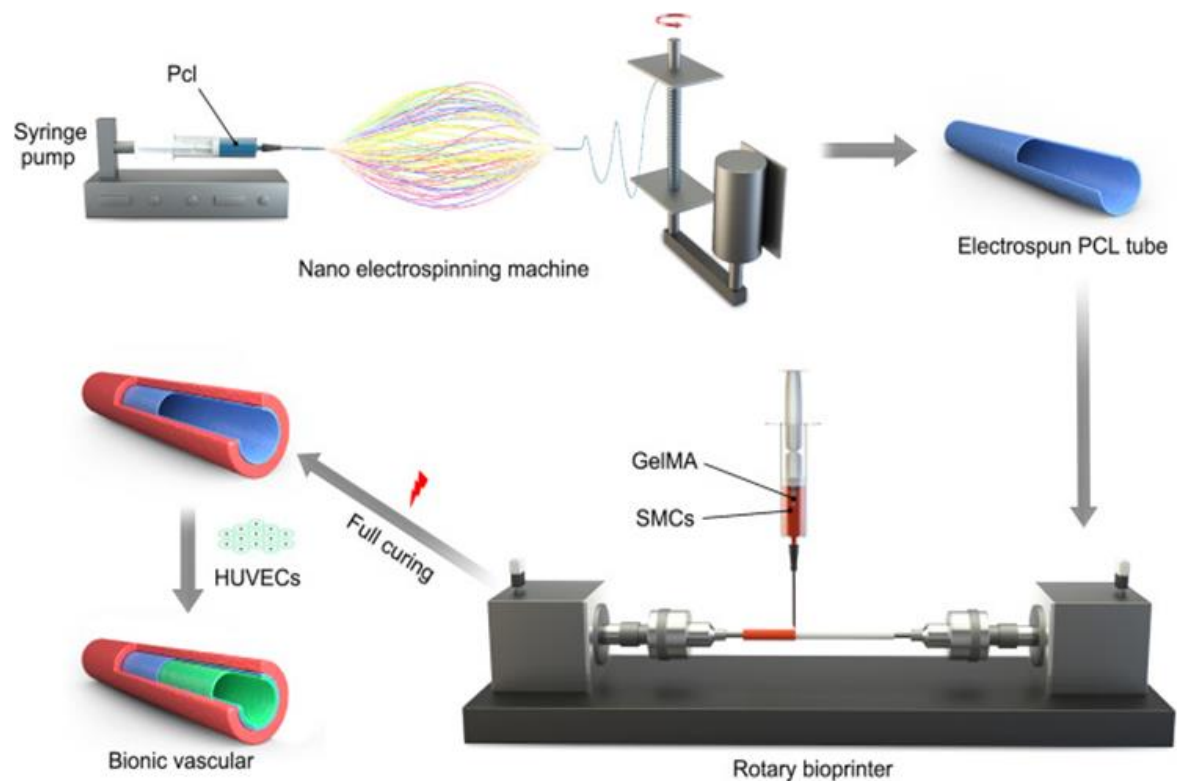


Figure 7. Schematic showing the novel approach for fabricating small-diameter bionic vascular tissue by combining nanofiber electrospinning and rotary bioprinting. Reprinted with permission under the open access from Jin et al [108].

Xu et al [109] created a vascular construct with both good biocompatibility and mechanical strength using a novel integrated tissue-organ printer [110]. Two syringes of bioink were used, one for the inner layer of ECs and the other for the outer SM cells. The porosity of the GelMA bioink was varied, creating different-sized pores for each layer, essential for the different sizes of endothelial and SM cells. Gold et al [111] disposed with a supporting sacrificial structure, and used direct extrusion printing to fabricate vessels with a novel bioink: nanoengineered extracellular matrix (nECM). Their nECM comprised GelMA, poly (ethylene glycol) diacrylate, and two dimensions nanosilicates. This nECM had high printability and fidelity, cell viability >80%, and mimicked the thromboinflammatory response of native vessels.

Stereolithographic bioprinting uses patterned UV light to photocrosslink entire layers of bioink at once. In contrast to extrusion or inkjet printing, it does not print in points; as such, it produces larger-scale vascular structures more easily [112]. Krishnamoorthy et al [113] used this layer-by-layer approach to print vascular constructs using fibroblasts encapsulated within GelMA bioink. Cell viability after 48-hour incubation was 80%. They identified an ideal cure depth (photocrosslinking thickness of an individual layer) range of 200 μm for this printing process, which gave the best shape fidelity and resolution. Thomas et al [114] trialled multiple photoink formulations to find the ideal sacrificial photoink in which to embed ECs. 1.5% methacrylated hyaluronic acid (HAMA) was superior due to its rapid digestion time, resulting in a faster release of ECs. They created a sophisticated bifurcating vessel with sufficient EC proliferation (Figure 8).

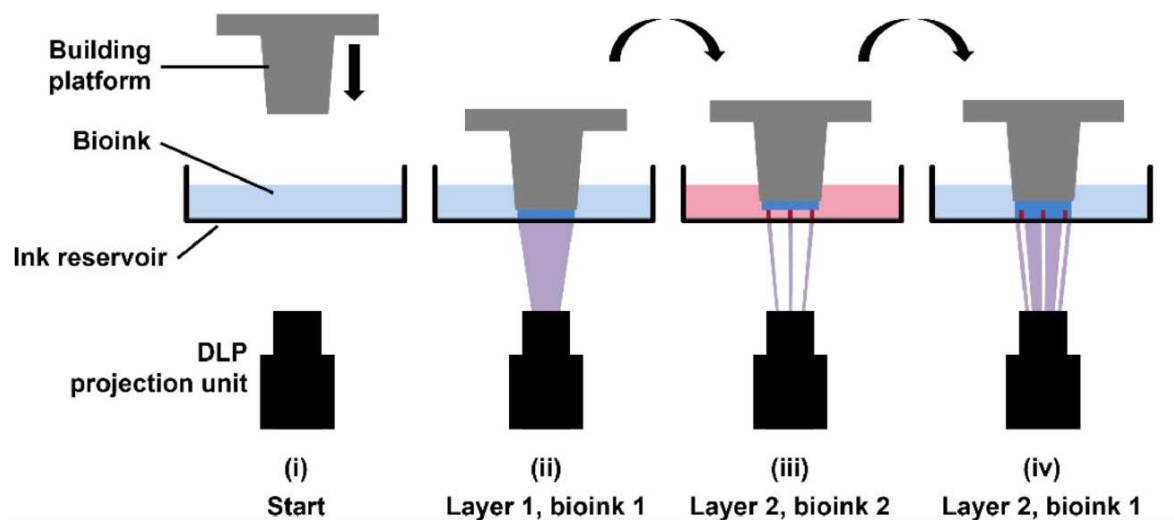


Figure 8. Pictorial representation of stereolithographic bioprinting with multiple inks to create the different components of the vessel wall. DLP: digital light processing. Reprinted with permission under the open access from Thomas et al [114].

6.3. Summary

3D bioprinting represents an important potential solution to the shortage of biocompatible vascular grafts. Multiple techniques have been trialled *in vitro*, with extrusion-based printing being the most common approach. These studies demonstrate excellent viability and proliferation of printed ECs. Relatively few studies have tested the mechanical properties of printed structures which need to be robust under arterial pressure and will inform *in vivo* trials [103,107–109].

7. 3D bioprinting of heart valves

Valvular heart disease is a serious global health problem, affecting 3.1% of the adult population and projected to claim 23.6 million lives by 2030 [115,117]. Of the four valves in the heart, the aortic valve bears the greatest hemodynamic load, pumping 3-5 liters per minute and undergoing 30-40 million cycles per year [117]. Degenerative aortic valve disease is the most common cause of valvular heart disease representing at least 65.2% of cases (aortic stenosis 47.2%, aortic regurgitation 18.0%) [115,118]. The current clinical solution for a diseased valve is replacement with a prosthetic one which can be classified as either mechanical (made from metal or carbon) or bioprosthetic (made from chemically fixed animal cardiac tissue). These prosthetic valves have transformed the treatment of valvular heart disease, however, are not without their problems. Although mechanical valves are the most durable they require lifelong anticoagulation therapy [119,120]. Conversely, bioprosthetic valves do not require anticoagulation therapy but suffer from limited durability [122,123]. Furthermore, both types of prosthetic valves require replacement (mechanical valves ~20 years, bioprosthetic valves as short as 5 years in younger patients) which is of growing concern as the population ages, and as younger patients are being treated with the adoption of lower-risk procedures such as transcatheter valve implantation [116,124].

Tissue engineered heart valves (TEHVs) present a possible solution to this problem, with the potential to provide a valve that does not require anticoagulation therapy and could last a lifetime [125–127]. However, a successful TEHV has complex requirements including the ability to encourage cells to infiltrate, differentiate and proliferate in a highly specific manner, as well as exhibit adequate mechanical and haemodynamic functionality [128–130]. These, along with complex regulatory approvals and patient to patient variability means that TEHVs are yet to reach the clinic.

Bioprinting is one possible avenue to creating a successful TEHV, with the ability to construct patient-specific, hierarchical, cell-laden constructs that could also match the complex microstructure of a native valve. A handful of attempts have been made using extrusion-based, light-based and bioplotting bioprinting techniques and are described below.

7.1. Extrusion based TEHVs

Extrusion-based printing can rapidly fabricate anatomically correct heterogeneous TEHVs using two types of cell-free poly-ethylene glycol-diacrylate (PEGDA) which are able to support porcine aortic valve interstitial cells (PAVICs) after 21 days [131]. Furthermore, the same researchers incorporated SM cells and valve interstitial cells (VICs) into anatomically correct extrusion bioprinted valve roots and leaflets made from alginate and gelatin (Figure 9A) [132]. Both SMCs and VICs were viable (>80%) after 7 days and expressed elevated alpha-smooth muscle actin (aSMA) and vimentin respectively. Cell-laden constructs initially exhibited weaker ultimate tensile strength, failure strain and Young's modulus than their cell-free counterparts, although over 7 days the cell-laden valves maintained their mechanical strength as opposed to cell-free constructs which weakened. Subsequently, human aortic valvular interstitial cells (HAVIC) were extrusion bioprinted into a simplified heart valve geometry (Figure 9B) [133]. Different ratios of methacrylated hyaluronic acid (Me-HA) and methacrylated gelatin (Me-Gel) impacted cell spreading, mechanical properties, and printability of constructs. After 3 days in a static culture tube, the encapsulated cells below the surface started to remodel the hydrogel by depositing collagen and glycosaminoglycans (GAGs).

These initial studies demonstrated the feasibility of extrusion bioprinting for the creation of TEHVs, however did not address concerns regarding the extreme mechanical requirements. Accordingly, there have been multiple investigations to reinforce the mechanical integrity of cell-laden hydrogels. These include the incorporation of nanoparticles such as nanocrystalline cellulose [134], variations in photocrosslinking methods [135], and reinforcement with electrospun polyacrylonitrile (PAN) fibers [136]. Despite demonstrating the ability to tailor mechanical properties, none of these studies have shown mechanical properties sufficient to match those of native heart valves.

7.2. Light-based TEHVs

Stereolithography printing was the first technique to successfully 3D print a TEHV, albeit cell-free [137]. However, since then there has only been one attempt at light-based bioprinting which used PEGDA:GelMA seeded with cardiac fibroblasts [138]. The valve showed 80% cell viability after 7 days although was not tested for haemodynamic or mechanical properties. Since then there have been no further attempts at light-based bioprinted TEHV, perhaps due to the lack of material heterogeneity achievable with the method, or more likely due to the relatively low mechanical strength.

7.2. Bioplotting TEHVs

More recently, TEHVs have been fabricated using a novel bioplotting method called freeform reversible embedding of suspended hydrogels (FRESH). Notably, this method has been used to fabricate an entire neo-natal scale human heart exhibiting multiple anatomical structures such as the ventricular chambers and aortic valves (Figure 9D) [139]. In the same study, tri-leaflet valves were fabricated using a cell-free collagen based ink. While this valve strictly speaking, did not contain cells, the same technique was used to print cell-laden vessels with diameters of 8-50 μm . It must be noted however, that prior to mechanical testing the valves were decellularized to enhance mechanical properties. The valve was tested in a physiological flow loop setting as per ISO 5840 and demonstrated adequate total regurgitation fraction, although insufficiently high effective orifice area and transvalvular pressure gradient for the aortic setting.

The same technique has been used to evaluate the recellularization potential of simplified cell-laden TEHVs in a subcutaneous rat model (Figure 9C) [140]. Rat mesenchymal stem cells (rMSCs) were printed using a bioink of highly concentrated Type I collagen hydrogels (Lifeink 200) and implanted for 2, 4, 8, and 12 weeks to assess cell infiltration, inflammation, and uni-axial tensile mechanical properties. The scaffolds expressed aSMA and vimentin biomarkers, demonstrating an acceptable inflammatory response although there was limited cellular infiltration into the scaffold. Tensile mechanical properties varied throughout the duration of the 12 weeks potentially showing variations of scaffold resorption and remodelling, although statistically insignificant. Following this,

a similar study was conducted accompanied by finite element computational analysis in an attempt to characterise the remodelling of mechanical properties [141]. The study concluded that subcutaneous culture was insufficient for replicating native tissue mechanical properties and a dynamic cellularization environment such as a bioreactor would be necessary.

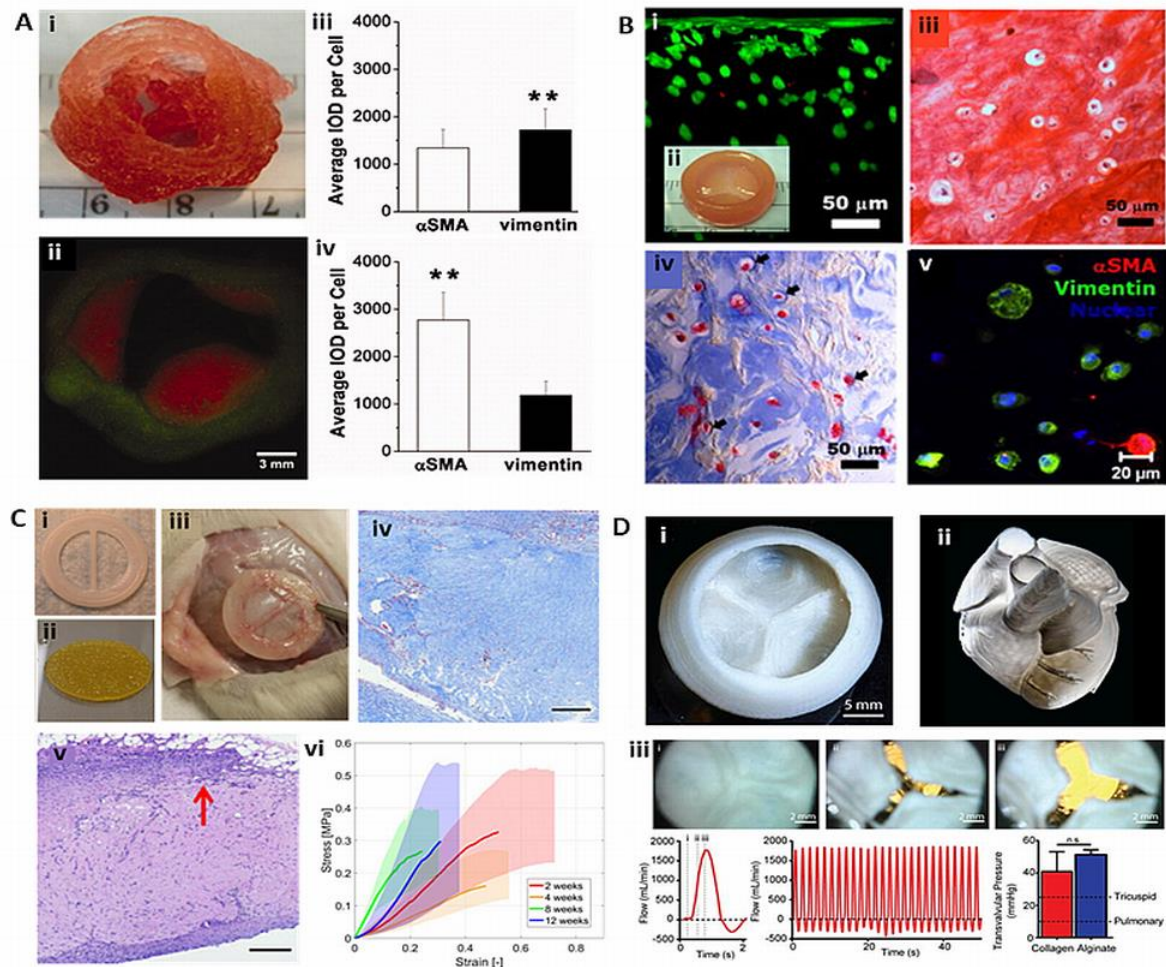


Figure 9. Bioprinted heart valves **A:** i) Extrusion-based bioprinted heart valve containing alginate/gelatin encapsulated with VIC/SMC respectively with ii) fluorescent cell labelling of SMCs in green and VICs in red and expression of α SMA and vimentin after 7 day culture and inset live/dead assay for encapsulated iii) VIC and iv) SMC. Reprinted with permission from Duan et al [132]. **B:** Simplified extrusion-bioprinted heart valve printed from Me-Gel encapsulated with HAVIC with i) Live/Dead staining showing cell viability through cross section of valve from surface to 300 μ m, ii) inset showing simplified morphology, iii-v) histological staining of bioprinted leaflets after 7-day culture. iii) Safranin-O staining was used to stain the GAGs red, which also stained the Me-HA within the hydrogel red, iv) Masson's Trichrome staining was used to stain collagen blue, which also stained the Me-Gel within the hydrogel blue. The arrow shows the newly secreted collagen around the HAVIC, indicating the remodeling process, vi) representative image of immunohistochemical staining for α SMA (green) and vimentin (red), and Draq 5 counterstaining for cell nuclei (blue). Reprinted with permission from Duan et al [133]. **C:** Simplified bioplotting valve made from i) PCL frame and ii) bioplotting type I collagen hydrogels and rMSCs, with iii) showing subcutaneous explant from Sprague-Dawley rat after 12 weeks, iv) H&E staining showing increase in host cellular concentration found at the periphery (red arrow) and infiltrating within the scaffold, v) Masson's trichrome showing a diffuse blue expression representative of collagen, scale bars = 300 μ m, vi) stress-strain plot of the heart valve scaffolds at 2, 4, 8, and 12 weeks. Reprinted with permission under the open access from Maxson et al [140]. **D:** i) decellularized bioplotting TEHV from collagen-based bioink, ii) entire collagen heart, (iii) sequence of valve opening under pulsatile flow over ~1 s with doppler flow velocimetry of a single cycle and multiple cycles, and maximum transvalvular pressure of printed

alginate and collagen valves compared to operating pressure for native valves. Reprinted with permission from Lee et al [141].

Thus, despite the promise shown in printing of heterogeneous cell-laden valves with patient-specific geometries, bioprinting of purely bioink-based TEHV's have insufficient mechanical properties. Novel methods of mechanical reinforcement such as incorporation of nanoparticles [142,143], new methods of physical or chemical crosslinking [144], and soft network composites may help [145,146].

8. 3D bioprinting of myocardium and heart

Injury to the heart in the form of myocardial infarction (MI) or heart failure, can be treated with medication, angioplasty and stenting, bypass surgery and lifestyle changes, however, the only remaining option for end stage disease is a heart transplant. With the well-known drawbacks that transplants entail, including donor shortage and procedure-associated risks, tissue engineering and regenerative medicine (TE&RM) approaches have been gaining ground as potential solutions.

Current views in TE&RM support the notion that the best way to achieve adequate engineered-tissue capabilities is by trying to replicate native composition, structure and properties as closely as possible [147,148]. Bioprinting technologies, with their capability to pattern cells and biomaterials with great spatial resolution, are standing out as an unparalleled tool for myocardial TE&RM and will be examined in the following paragraphs and also summarised in Table 1.

Many myocardial bioprinting studies focus on optimisation of printing conditions, bioink formulation and/or functionalisation to incorporate additional benefits. Hence, one research group formulated a bioink based on cardiac extracellular matrix (ECM) [149], while another incorporated gold nanorods (GNRs) under the hypothesis that it would beneficially contribute to the electrical conduction of the construct [150]. For their part, Erdem and co-workers attempted to create an oxygen-releasing bioink by adding calcium peroxide to their GelMA-based formulation, which improved survival and function of the construct under hypoxic conditions [151]. All reported good feasibility and cell viability, together with the desired effect albeit to a limited degree.

More recently, Ahrens et al focused on imparting cardiomyocyte (CM) alignment in their bioprinted cardiac constructs. The particularity of this approach relies on the fact that the cellular component of the bioink is not dissociated cells, but rather preassembled aggregates of tens of thousands of cardiac cells, denominated anisotropic organ building blocks (OOBs), and generated by seeding and culturing a mixture of CMs derived from iPSCs, human neonatal dermal fibroblasts and a collagen hydrogel in a micropillar array. This resulted in highly aligned, densely cellularized ($+200 \cdot 10^6$ cells/ml) cardiac struts and sheets.

The anisotropy of the bioprinted constructs came from both the auxotonic mechanical conditioning of the OOBs by the micropillars, and the shear stress direction imparted by the extrusion through the nozzle. The cardiac constructs contracted synchronously and with greater force than spheroid-based controls and could sustain pacing up to 3 Hz [152].

Similarly, Ong et al also chose to print with cell aggregates (spheroids) rather than cell suspensions, but they did not add any supporting hydrogel under the assumption that it may hinder cell-cell interactions. Their constructs beat spontaneously and synchronously, and responded to electrical pacing. When varying the ratio between CMs, ECs and cardiac fibroblasts, results suggested that the latter can delay and even block electrical conduction if present in too high quantity [153]. When implanted in a rat MI model, the cardiac patch resulted in a smaller scar area and greater vascularization than the control (omental patch), as well as better functional output (ejection fraction and cardiac output), however these were not statistically significant [154].

Interestingly, bioprinting of cells can be complemented by the simultaneous deposition of other materials that will perform a specific function. For instance, one study incorporated a polycaprolactone (PCL) frame around the bioprinted construct to subject it to auxotonic stress during culture, as well as micro-springs that enabled measurement of the patches' contractile force. After 4 weeks in culture, samples showed the presence of aligned and dense sarcomeres, as well as connexin

43. Moreover, the cardiac constructs displayed physiologic response when subjected to drugs known to alter contraction force and frequency, namely epinephrine and carbachol [44].

In another example, Asulin and co-workers combined the bioprinting of three distinct components to create a cardiac construct with integrated electronics allowing for both sensing of the patch's activity and its pacing. One bioink consisted of a mixture of CMs and ECM, while the other two material inks were made of polydimethylsiloxane (PDMS) and either graphite flakes for conduction, or surfactant for passivation (passivation is essential for achieving accurate spatial sensing and stimulation while reducing noise) (Figure 10) [155].

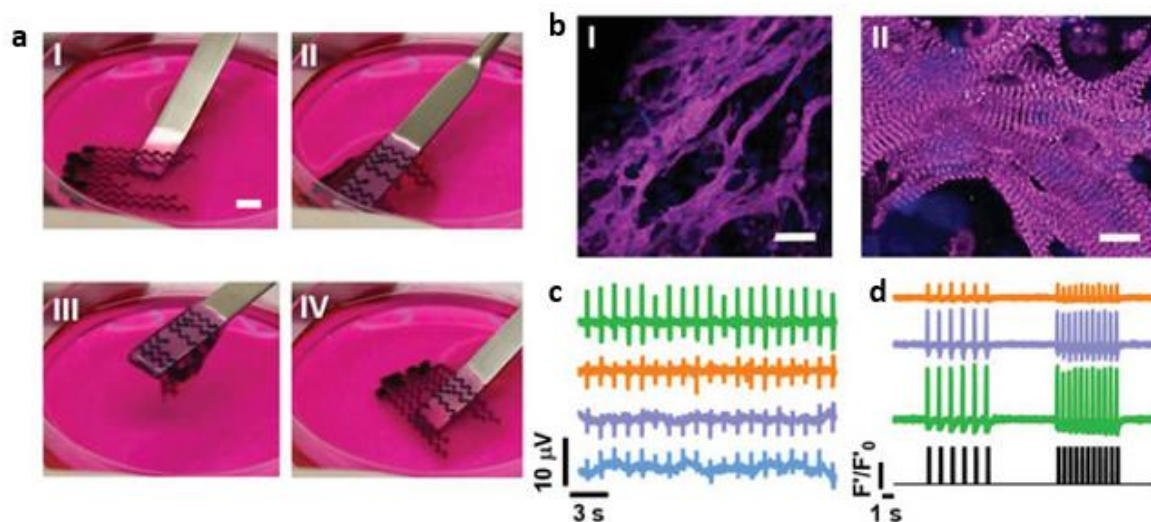


Figure 10. Bioprinted cardiac patch with integrated electrodes. a) the patch is soft and flexible, electrodes can be seen in black. I-IV show the patch being picked up and returned to the culture plate. b) immunostaining for sarcomeric actinin (pink) and nuclei (blue). Scale bars are 50 and 10 μm for I and II respectively. c) recordings of action potentials from four distinct locations. d) calcium transients from three distinct locations after pacing at 7 V and 1 and 2 Hz. The pacing pattern is shown at the bottom of the panel. Reprinted with permission under the open access from Asulin et al [155].

In a major step towards implant personalization, Tal Dvir and team used patient omental biopsies to create patient-specific bioinks [156]. These consisted of a mixture of the patient's own cells, reprogrammed into iPSCs and later differentiated into the desired phenotypes (endothelial and CMs), and the patient's extracellular matrix, turned into a thermoresponsive collagen hydrogel. Using these they bioprinted thick (2 mm) vascularized cardiac patches, made of CMs and perfusable channels 300 μm in diameter, lined with endothelial cells. Furthermore, they sought to print constructs in a more architecturally relevant, larger size, for which they developed a support medium allowing bioink curing and construct extraction without damage to the cells. This allowed them to bioprint an anatomically accurate, miniaturized heart (20x18 mm) (Figure 11) [139]. The heart displayed two hollow chambers as ventricles, and was surrounded by perfusable channels modelled to imitate coronary arteries.

Similarly, Lee et al also utilized a support bath to print human CMs derived from embryonic stem cells and collagen into an ellipsoidal shell, which contracted in unison and could be paced up to 2 Hz (Figure 12) [157]. Finally, Kupfer and co-workers focused on bioprinting complex structures, this time adapting an MRI scan of the heart to contain an inlet and outlet channel through which unidirectional flow could be propagated. A key aspect of this work is that they chose to bioprint hiPSCs, in situ differentiating them to cardiac lineages after construct formation. This way the rupture of CM-to-CM connections for bioink generation is avoided, which the authors argue results in enhanced cell density and tissue connectivity [158].

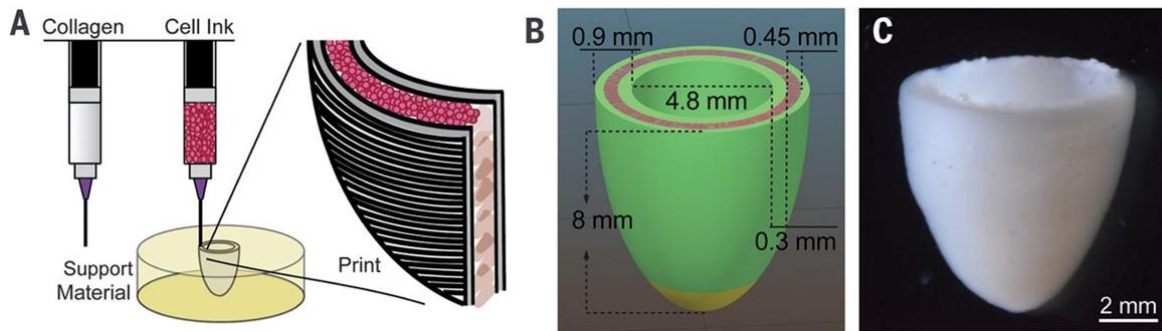


Figure 11. 3D printed ventricle model a) schematic of the dual-material printing in a support medium. b) ventricle model with the cell compartment depicted in pink, while the green represents a collagen-based bioink. c) micrograph of the bioprinted ventricle. Reprinted with permission from Lee et al [139].

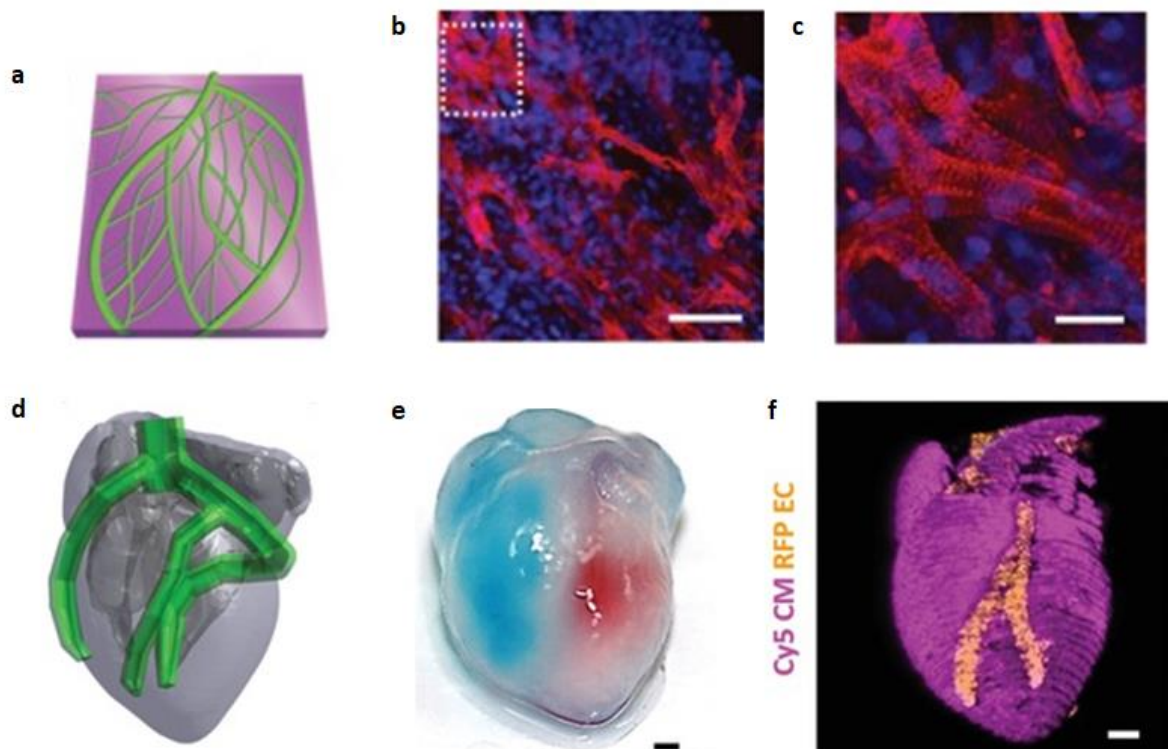


Figure 12. Patient-derived cardiac patch and whole heart model. a) model of the vascularised patch, the two bioinks shown in different colorus. b, c) immunostaining for sarcomeric actinin (red) and nuclei (blue) from the patch; c is a higher magnification of the dotted area in b. Scale bars correspond to 50 and 25 μm in b and c respectively. d) computerized model of the miniaturized heart. e) after printing, the two compartments were injected with blue and red dyes to demonstrate the presence of hollow chambers and a septum in between, scale bar=1 mm. f) confocal image of the printed heart, with CMs in pink and ECs in orange. Scale bar corresponds to 1 mm. Reprinted with permission from Noor et al [157].

Very recently, to tackle the issues of construct size and cell survival during and after bioprinting, a 6 degrees-of-freedom robotic arm was modified to serve as a bioprinter. This surpasses the capabilities of any cartesian printer, which only permits bottom-up manufacturing, and allows for printing in any direction. In this approach, tubular scaffolds in the manner of vascular conduits, through which culture medium is constantly circulated, act as a backbone onto which layers of endothelial cells and CMs are deposited. The process is iterative, consisting of rounds of bioprinting and culturing, to give time for cell attachment between the new layer and the previously deposited ones, and vascular network growth. The printing is performed in a chamber, filled either with an oil bath to ensure adhesion of the bioink to complex-shaped surfaces, or culture media. Although the

authors did not demonstrate the printing of a full heart, this work provides proof of concept and shows clear potential to generate complex tissues and entire organs (Figure 13) [159].

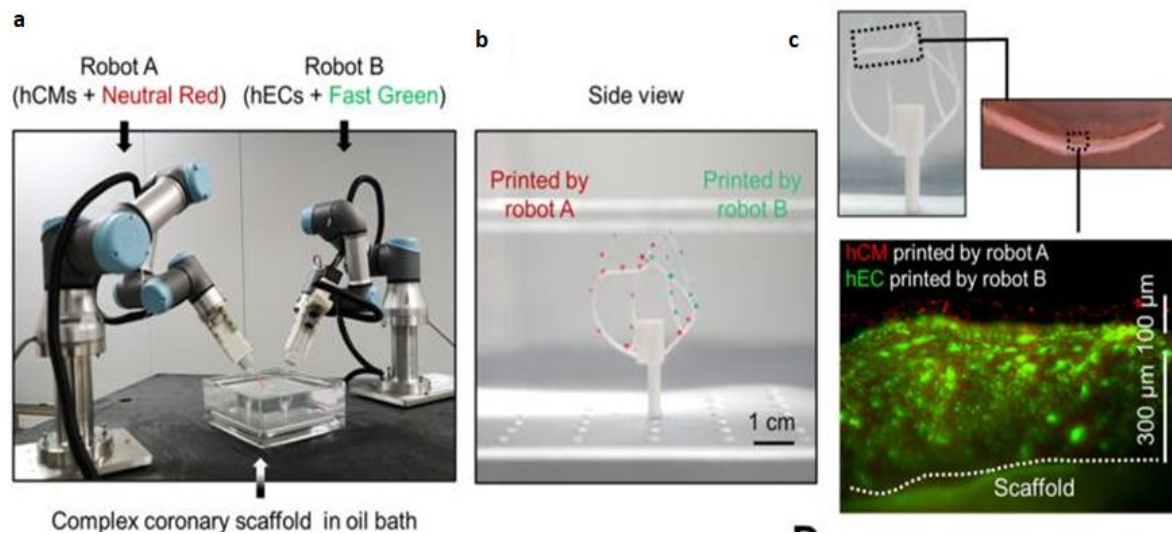


Figure 13. 6 degree-of-freedom robotic arms are able to bioprint in complex shapes. a) setup with two robotic arms, one bioprinting red-stained CMs and the other green-stained ECs on top of a mold resembling the coronary tree. b) all zones of the template are accessible for the robots, as demonstrated with the selectively deposited bioink droplets. c) CMs and ECs are assembled in two distinct layers bioprinted on one branch of the coronary template. Reprinted with permission under the open access from Zhang et al [159].

Table 1. Summary of myocardial bioprinting approaches. Abbreviations: hiPSC, human induced pluripotent stem cells; CM, cardiomyocytes; CB, cardiac fibroblasts; EC, endothelial cells; GelMA, gelatin methacryloyl; GNR, gold nanorods; Cx43, connexin 43; hCPCs, human cardiac progenitor cells; ECM, extracellular matrix; PCL, poly-ε-caprolactone; hESC, human embryonic stem cells; CPO, calcium peroxide; ColMA, collagen methacryloyl; SMC, smooth muscle cells; PDMS, polydimethylsiloxane.

References	Construct shape	Bioinks		Key aspect of study	Examined benefits
		Cells	Hydrogel		
[153]	Patch	Spheroids of hiPSC-derived CM, CB and EC	-	Biomaterial-free	Spontaneous contraction, ability to pace constructs, rudimentary vascularization, in vivo engraftment
[150]	Grid	Neonatal rat CM + CF	GelMA + alginate + GNR	GNR to improve electrical conduction	Higher Cx43 expression, higher synchronous contractile frequency than constructs without GNR
[149]	Grid, patch	hCPCs	ECM + GelMA	Cardiac-ECM specific bioink	Higher cardiac and endothelial-specific gene expression than GelMA-only constructs, retention and vascularization after in vivo implantation
[44]	Patch	Neonatal rat CM	Fibrinogen + gelatin	PCL frame to impart auxotonic mechanical stress	Cell alignment, physiologic response to drugs altering force and frequency of contraction
[139]	Ellipsoid	hESC-CM	Collagen	Ventricle-like shape	Spontaneous, synchronous contraction, pacing at 1 and 2 Hz.
[157]	Patch, two-chambered ellipsoid	hiPSC-derived CM and EC	ECM	Patient specificity, vascularization, shape	Cardiac patch with perfusable, vascular-like channels. Spontaneous and synchronous contraction
[154]	Patch	Spheroids of hiPSC-derived CM, CF and EC	-	In vivo study of patch described in (7)	Smaller scar, greater vascularization than control (omentum patch). Greater ejection fraction and cardiac output, although not significant

[151]	Grid	Neonatal rat CM + mouse fibroblasts	GelMA + CPO	Oxygen-releasing bioink	Enhanced viability and function under hypoxic conditions
[158]	Chambered ellipsoid	hiPSCs	GelMA + ColMA	Ventricular-like shape, pump-like function, differentiation after printing	Differentiation to CM, SMC and EC. Spontaneous and synchronous contraction, physiologic response to isoproterenol, for up to 6 weeks in culture
[155]	Patch	Neonatal rat CM/hiPSC-CM	ECM, PDMS + graphite, PDMS + surfactant	Integrated electrodes for sensing and pacing	Good cell viability, spontaneous contraction and actinin expression. Sensing and pacing at 1 and 2 Hz
[152]	Struts, patch	hiPSC-CM microtissues	Fibrinogen + gelatin	High cellular density, alignment	Higher directionality, conduction velocity and force generation than spheroid-based constructs
[159]	Lining of vascular model	hESC-CM + EC	-	Ability to print in any direction	No damage in viability or activity after printing, evidence of vasculogenesis, synchronous and spontaneous contraction

9. Summary and concluding remarks

Recent developments in 3D printing technology have revolutionized medical practice by using personalised 3D printed heart and vascular models in enhancing diagnosis and treatment of cardiovascular disease. Technological advancement in 3D bioprinting has great potential and is a powerful tool utilising tissue engineering principles to bioprint cardiac cells, vascular and tissue constructs. New advancements in 3D bioprinting have made it possible to construct organ-mimicking myocardial or cellular structures with high precision and flexibility. A variety of biomaterials in combination with various bioprinting methods have been shown to demonstrate great potential in 3D bioprinting cardiac constructs and grafts, myocardium and heart valves. Despite promising results and rapid progress further research and investigation is required to better understand the cross over between a native organ and amalgamation of bioprinted heterogeneous tissue constructs. Although mechanical properties are in some cases being reproduced, novel methods of mechanical reinforcement are essential to enable translation in vivo. Whilst the area of valve replacement is advancing rapidly, we see less exploration of bioprinting of vascular grafts. Endovascular minimally invasive treatments have rendered bypass surgery redundant in many patients, in both the coronary and peripheral circulations, however, bespoke patient derived bioprinted arteries may in fact sway the pendulum back from minimally invasive treatments by providing a more durable solution to occlusive arterial disease by virtue of biocompatibility, bespoke sizing and the reproduction of arterial mechanical properties.

Author Contributions: Conceptualization, Z.S. and S.J.; methodology: Z.S.; writing—original draft preparation, Z.S., J.Z., E.L., M.F., M.V., J.S.; writing—review and editing: Z.S., E.DJP., S.J. Project administration, Z.S. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Institutional Review Board Statement: Not applicable as this is a review article.

Informed Consent Statement: Not applicable.

Conflicts of Interest: EDJP is Co-Founder and Director of CoraMetix Pty Ltd. Other authors declare no conflict of interest.

References

1. Virani, S.S.; Alvonso, A.; Aparicio, J.E.; Benjamin, J.E.; Bittencourt, M.; Callaway, C.; Clifton, W.; et al. Heart disease and stroke statistics-2021 update: a report from the American Heart Association. *Circulation*. **2021**, *143*, e254-e743.
2. Sun, Z.; Al Moudi, M.; Cao, Y. CT angiography in the diagnosis of cardiovascular disease: a transformation in cardiovascular CT practice. *Quant. Imaging. Med. Surg.* **2014**, *4*, 376-396.
3. Russo, V.; Lovato, L.; Ligabue, G. Cardiac MRI: technical basis. *Radiol. Med.* **2020**, *125*, 1040-1055.
4. Giannopoulos, A.A.; Steigner, M.L.; George, E.; Barile, M.; Hunsaker, A.R.; Rybicki, F.J.; Mitsouras, D. Cardiothoracic applications of 3-dimensional printing. *J. Thorac. Imaging* **2016**, *31*, 253-272.
5. Lau, I.; Sun, Z. Dimensional accuracy and clinical value of 3D printed models in congenital heart disease: A systematic review and meta-analysis. *J. Clin. Med.* **2019**, *8*, 1483.
6. Lee, S.; Squelch, A.; Sun, Z. Quantitative assessment of 3D printed model accuracy in delineating congenital heart disease. *Biomolecules* **2021**, *11*, 270.
7. Valverde, I.; Gomez-Ciriza, G.; Hussain, T.; Suarez-Mejias, C.; Velasco-Forte, M.N.; Byrne, N.; Ordonex, A.; Gonzalez-Calle, A.; Anderson, D.; Hazekamp, M.G.; et al. Three dimensional printed models for surgical planning of complex congenital heart defects: An international multicenter study. *Eur. J. Cardiothorac. Surg.* **2017**, *52*, 1139-1148.
8. Sun, Z.; Wee, C. 3D printed models in cardiovascular disease: An exciting future to deliver personalized medicine. *Micromachines* **2022**, *13*, 1575.
9. Sun, Z. Clinical applications of patient-specific 3D printed models in cardiovascular disease: Current status and future directions. *Biomolecules* **2020**, *10*, 1577.
10. Sun, Z.; Wong, Y.H.; Yeong, C.H. Patient-specific 3D printed low-cost models in medical education and clinical practice. *Micromachines* **2023**, *14*, 464.

11. Anwar, S.; Singh, G.K.; Miller, J.; Sharma, M.; Manning, P.; Billadello, J.J.; Eghtesady, P.; Woodard, P.K. 3D printing is a transformative technology in congenital heart disease. *JACC. Basic. Transl. Sci.* **2018**, *3*, 294-312.
12. Gallo, M.; D'Onofrio, A.; Tarantini, G.; Nocerino, E.; Remondino, F.; Gerosa, G. 3D-printing model for complex aortic transcatheter valve treatment. *Int. J. Cardiol.* **2016**, *210*, 139-40.
13. Ripley, B.; Kelil, T.; Cheezum, M.K.; Goncalves, A.; Di Carli, M.F.; Rybicki, F.J.; Steigner, M.; Mitsouras, D.; Blankstein, R. 3D printing based on cardiac CT assists anatomic visualization prior to transcatheter aortic valve replacement. *J. Cardiovasc. Comput. Tomogr.* **2016**, *10*, 28-36.
14. Kiraly, L.; Shah, N.C.; Abdullah, O.; Al-Ketan, O.; Rowshan, R. Three-dimensional virtual and printed prototypes in complex congenital and pediatric cardiac surgery-A multidisciplinary team-learning experience. *Biomolecules.* **2021**, *11*, 1703.
15. Meyer-Szary, J.; Luis, M.S.; Mikulski, S.; Patel, A.; Schulz, F.; Tretiakow, D.; Fercho, J.; Jaguszewska, K.; Frankiewicz, M.; Pawłowska, E.; Targoński, R.; Szarpak, Ł.; Dądela, K.; Sabiniewicz, R.; Kwiatkowska, J. The role of 3D printing in planning complex medical procedures and training of medical professionals-cross-sectional multispecialty review. *Int. J. Environ. Res. Public. Health.* **2022**, *19*(6), 3331.
16. Yamasaki, T.; Toba, S.; Sanders, S.P.; Carreon, C.K. Perfusion-distension fixation of heart specimens: a key step in immortalizing heart specimens for wax infiltration and generating 3D imaging data sets for reconstruction and printed 3D models. *Cardiovasc. Pathol.* **2022**, *58*, 107404.
17. Ghosh, R.M.; Jolley, M.A.; Mascio, C.E.; Chen, J.M.; Fuller, S.; Rome, J.J.; Silvestro, E.; Whitehead, K.K. Clinical 3D modeling to guide pediatric cardiothoracic surgery and intervention using 3D printed anatomic models, computer aided design and virtual reality. *3D. Print. Med.* **2022**, *8*, 11.
18. Yoo, S.J.; Spray, T.; Austin, E.H.; Yun, T.J.; van Arsdell, G.S. Hands-on surgical training of congenital heart surgery using 3-dimensional print models. *J. Thorac. Cardiovasc. Surg.* **2017**, *153*, 15301-15540.
19. Gomez-Ciriza, G.; Gomez-Cia, T.; Rivas-Gonzalez, J.A.; Velasco Forte, M.N.; Valverde, I. Affordable three-dimensional printed heart models. *Front. Cardiovasc. Med.* **2021**, *8*, 642011.
20. Guo, H.; Wang, Y.; Dai, J.; Ren, C.; Li, J.; Lai, Y. Application of 3D printing in the surgical planning of hypertrophic obstructive cardiomyopathy and physician-patient communication: A preliminary study. *J. Thorac. Dis.* **2018**, *10*, 867-873.
21. Ryan, J.; Plasencia, J.; Richardson, R.; Velez, D.; Nigro, J.J.; Pophal, S.; Frakes, D. 3D printing for congenital heart disease: A single site's initial three-year experience. *3D. Print. Med.* **2018**, *4*, 10.
22. Zhao, L.; Zhou, S.; Fan, T.; Li, B.; Liang, W.; Dong, H. Three-dimensional printing enhances preparation for repair of double outlet right ventricular surgery. *J. Card. Surg.* **2018**, *33*, 24-27.
23. Fan, Y.; Yang, F.; Cheung, S.H.; Chan, A.K.Y.; Wang, D.D.; Lam, Y.Y.; Chow, M.C.K.; Leong, M.C.W.; Kam, K.K.; So, K.C.Y.; et al. Device sizing guided by echocardiography-based three-dimensional printing is associated with superior outcome after percutaneous left atrial appendage occlusion. *J. Am. Soc. Echocardiogr.* **2019**, *32*, 708-719.
24. Hell, M.H.; Achenbach, S.; Yoo, I.S.; Franke, J.; Blackutzik, F.; Roether, J.; Graf, V.; Raaz-Schrauder, D.; Marvann, M.; Schlundt, C. 3D printing for sizing left atrial appendage closure device: Head-to head comparison with computed tomography and transoesophageal echocardiography. *EuroIntervention* **2017**, *13*, 1234-1241.
25. Li, H.; Yao, Q.; Shen, B.; Shu, M.; Zhong, L.; Wang, X.; Song, Z. Application of 3D printing technology to left atrial appendage occlusion. *Int. J. Cardiol.* **2017**, *231*, 258-263.
26. Conti, M.; Marconi, S.; Muscogiuri, G.; Guglielmo, M.; Baggiano, A.; Italiano, G.; Mancini, M.E.; Auricchio, F.; Andreini, D.; Rabbat, M.G.; et al. Left atrial appendage closure guided by 3D computed tomography printing technology: A case control study. *J. Cardiovasc. Comput. Tomogr.* **2019**, *13*, 336-339.
27. Goitein, O.; Fink, N.; Guettar, V.; Beinart, R.; Brodov, Y.; Konen, E.; Goitein, D.; Segni, E.D.; Grupper, A.; Glikson, M. Printed MDCT 3D models for prediction of left atrial appendage (LAA) occlude device size: A feasibility study. *EuroIntervention* **2017**, *13*, e1076-e1079.
28. Torres, I.O.; De Luccia, N. A simulator for training in endovascular aneurysm repair: The use of three dimensional printers. *Eur. J. Vasc. Endovasc. Surg.* **2017**, *54*, 247-253.
29. Karkkainen, J.M.; Sandri, G.; Tenorio, E.R.; Alexander, A.; Bjellum, K.; Matsumoto, J.; Morris, J.; Mendes, B.C.; DeMartino, R.R.; Oderich, G.S. Simulation of endovascular aortic repair using 3D printed abdominal aortic aneurysm model and fluid pump. *Cardiovasc. Intervent. Radiol.* **2019**, *42*, 1627-1634.

30. Kaufmann, R.; Zech, C.J.; Takes, M.; Brantner, P.; Thieringer, F.; Dentschmann, M.; Hergan, K.; Scharinger, B.; Hecht, S.; Rezar, R.; et al. Vascular 3D printing with a novel biological tissue mimicking resin for patient-specific procedure simulations in interventional radiology: A feasibility study. *J. Digit. Imaging* **2022**, *35*, 9–20.
31. Abdullah, K.A.; McEntee, M.F.; Reed, W.; Kench, P.L. Development of an organ-specific insert phantom generated using a 3D printer for investigations of cardiac computed tomographic protocols. *J. Med. Radiat. Sci.* **2018**, *65*, 175–183.
32. Morup, S.D.; Stowe, J.; Precht, H.; Gervig, M.H.; Foley, S. Design of a 3D printed coronary artery model for CT optimization. *Radiography* **2022**, *28*, 426–432.
33. Sun, Z.; Ng, C.K.C.; Wong, Y.H.; Yeong, C.H. 3D-printed coronary plaques to simulate high calcification in the coronary arteries for investigation of blooming artifacts. *Biomolecules* **2021**, *11*, 1307.
34. Sun, Z.; Ng, C.K.; Squelch, A. Synchrotron radiation computed tomography assessment of calcified plaques and coronary stenosis with different slice thicknesses and beam energies on 3D printed coronary models. *Quant. Imaging Med. Surg.* **2019**, *9*, 6–22.
35. Sun, Z. 3D printed coronary models offer new opportunities for developing optimal coronary CT angiography protocols in imaging coronary stents. *Quant. Imaging Med. Surg.* **2019**, *9*, 1350–1355.
36. Sun, Z.; Jansen, S. Personalized 3D printed coronary models in coronary stenting. *Quant. Imaging Med. Surg.* **2019**, *9*, 1356–1367.
37. Sommer, K.N.; Lyer, V.; Kumamaru, K.K.; Rava, R.A.; Ionita, C.N. Method to simulate distal flow resistance in coronary arteries in 3D printed patient specific coronary models. *3D. Print. Med.* **2020**, *6*, 19.
38. Wu, C.; Squelch, A.; Sun, Z. Assessment of optimization of computed tomography angiography protocols for follow-up type B aortic dissection patients by using a 3D-printed model. *J. 3D. Print. Med.* **2022**, *6*, 117–127.
39. Aldosari, S.; Jansen, S.; Sun, Z. Optimization of computed tomography pulmonary angiography protocols using 3D printed model with simulation of pulmonary embolism. *Quant. Imaging Med. Surg.* **2019**, *9*, 53–62.
40. Aldosari, S.; Jansen, S.; Sun, Z. Patient-specific 3D printed pulmonary artery model with simulation of peripheral pulmonary embolism for developing optimal computed tomography pulmonary angiography protocols. *Quant. Imaging Med. Surg.* **2019**, *9*, 75–85.
41. Cui, H.; Miao, S.; Esworthy, T.; Zhou, X.; Lee, S.J.; Liu, C.; Yu, Z.; Fisher, J.P.; Mohiuddin, M.; Zhang, L.G. 3D bioprinting for cardiovascular regeneration and pharmacology. *Adv. Drug. Deliv. Rev.* **2018**, *132*, 252–269.
42. Alonzo, M.; Anilkumar, S.; Roman, B.; Tasnim, N.; Joddar, B. 3D bioprinting of cardiac tissue and cardiac stem cell therapy. *Transl. Res.* **2019**, *211*, 64–83.
43. Zhang, J.; Wehrle, E.; Rubert, M.; Muller, R. 3D bioprinting of human tissues: Biofabrication, bioinks and bioreactors. *Int. J. Mol. Sci.* **2021**, *22*, 3971.
44. Wang, Z.; Lee, S.J.; Cheng, H.J.; Yoo, J.J.; Atala, A. 3D bioprinted functional and contractile cardiac tissue constructs. *Acta. Biomater.* **2018**, *70*, 48–56.
45. Xia, Z.; Jin, S.; Ye, K. Tissue and organ 3D bioprinting. *SLAS. Technol.* **2018**, *23*, 301–314.
46. Kato, B.; Wisser, G.; Agrawal, D.K.; Wood, T.; Thankam, F.G. 3D bioprinting of cardiac tissue: Current challenges and perspectives. *J. Mater. Sci. Mater. Med.* **2021**, *32*, 54.
47. Chessa, M.; Van De Bruaene, A.; Farooqi, K.; Valverde, I.; Jung, C.; Votta, E.; Sturla, F.; Paul Diller, G.; Brida, M.; Sun, Z.; et al. Three-dimensional printing, holograms, computational modelling, and artificial intelligence for adult congenital heart disease care: An exciting future. *Eur. Heart J.* **2022**, *43*, 2672–2684.
48. Jain, P.; Kathuria, H.; Dubey, N. Advances in 3D printing of tissues/organs for regenerative medicine and in-vitro models. *Biomaterials.* **2022**, *287*, 121639.
49. Hoy, S.; Frisbee, J. Common postoperative heart transplant complications. *Crit. Care. Nurs. Q.* **2018**, *41*, 383–388.
50. Award, M.A.; Shah, A.; Griffith, B.P. Current status and outcomes in heart transplantation: a narrative review. *Rev. Cardiovasc. Med.* **2022**, *23*, 11.
51. Shi, W.Y.; Smith, J.A. Role of coronary artery bypass surgery in acute myocardial infarction, in: Prim. Angioplasty, Springer Singapore, **2018**, pp. 211–221.
52. Kwon, Y.W.; Yang, H.M.; Cho, H.J. Cell therapy on myocardial infarction. *Int. J. Stem. Cells.* **2010**, *3*, 8–15.
53. Das, S.; Nam, H.; Jang, J. 3D bioprinting of stem cell-laden cardiac patch: a promising alternative for myocardial repair. *APL. Bioeng.* **2021**, *5*, 031508.
54. Vukicevic, M.; Mosadegh, B.; Min, J.K.; Little, S.H. 3D printing and its future directions. *JACC. Cardiovasc. Imaging.* **2017**, *10*, 171–134.
55. Khanna, A.; Ayna, B.; Undieh, A.A.; Yang, Y.P.; Huang, N.F. Advances in three-dimensional bioprinted stem cell-based tissue engineering for cardiovascular regeneration. *J. Mol. Cell. Cardiol.* **2022**, *169*, 13–27.

56. Liu, N.; Ye, X.; Yao, B.; Zhao, M.; Wu, P.; Liu, G.; Zhuang, D.; Jiang, H.; Chen, X.; He, Y.; et al. Advances in 3D bioprinting technology for cardiac tissue engineering and regeneration. *Bioact. Mater.* **2020**, *6*, 1388-1401.
57. Liu, J.; He, J.; Liu, J.; Ma, X.; Chen, Q.; Lawrence, N.; Zhu, W.; Xu, Y.; Chen, S. rapid 3D bioprinting of in vitro cardiac tissue models using human embryonic stem cell-derived cardiomyocytes. *Bioprinting.* **2019**, *13*, 1-6.
58. Muffari, S.H.; Mahmood, F.; Qureshi, B.Q.; Yunus, R.; Quraishi, I.; Baribeau, V.; Sharkey, A.; Matyal, R.; Khabbaz, K. Three-dimensional printing of patient-specific heart valves: separating facts from fiction and myth from reality. *J. Cardiovasc. Vasc. Anesth.* **2022**, *36*, (8 Pt A), 2643-2655.
59. Gaetani, R.; Rizzitelli, G.; Chimenti, I. et al. Cardiospheres and tissue engineering for myocardial regeneration: potential for clinical applications. *J. Cell. Mol. Med.* **2010**, *14*, 1071-1077.
60. Gaetani, R.; Doevendans, R.A.; Metz, C.H.; et al. Cardiac tissue engineering using tissue printing technology and human cardiac progenitor cells. *Biomaterials.* **2012**, *33*, 1782-1790.
61. Gaetani, R.; Feyen, D.A.; Verhage, V. et al. Epicardial application of cardiac progenitor cells in a 3D-printed gelatin/hyaluronic acid patch preserves cardiac function after myocardial infarction. *Biomaterials.* **2015**, *61*, 339-348.
62. Ozbolat, I.T.; Moncal, K.K.; Gudapati, H. Evaluation of bioprinter technologies. *Addit. Manuf.* **2017**, *3*, 179-200.
63. Li, X.; Li, B.; Pei, B.; Chen, J.; Zhoud, D.; Peng, J.; Zhang, X.; Jia, W.; Xu, T. Inkjet bioprinting of biomaterials. *Chem Rev.* **2020**, *122*, 10596-636.
64. Mandrycky, C.; Wang, Z.; Kim, K.; Kim, D. 3D bioprinting for engineering complex tissues. *Biotechnol. Adv.* **2016**, *4*(4), 422-34.
65. Christensen, K.; Xu, C.; Chai, W.; Zhang, Z.; Fu, J.; Huang, Y. Freeform inkjet printing of cellular structures with bifurcations. *Biotechnol. Bioeng.* **2015**, *112*(5), 1047-55.
66. Schnerberg, J.; De Lorenzi, F.; Theek, B.; Blaeser, A.; Rommel, D.; Kuehne, A.J.; Kiessling, F.; Fischer, H. Engineering biofunctional in vitro vessel models using a multilayer bioprinting technique. *Sci. Rep.* **2018**, *8*(1):10430-13.
67. Saunders R.E.; Derby, B. Inkjet printing biomaterials for tissue engineering: bioprinting. *Int. Mater. Rev.* **2014**, *59*(8):430-48.
68. Shafiee, A.; Ghadiri, E.; Ramesh, H.; Kengla, C.; Kassis, J.; Calvert, P.; Williams, D.; Khademhosseini, A.; Narayan, R.; Forgacs, G.; Atala, A. Physics of bioprinting. *Appl. Phys. Rev.* **2019**, *6*(2):21315.
69. Chang, C.C.; Boland, E.D.; Williams, S.K.; Hoying, J.B. Direct-write bioprinting three-dimensional biohybrid systems for future regenerative therapies. *J. Biomed. Mater. Res. B. Appl. Biomater.* **2011**, *98B*(1),160-70.
70. Rider, P.; Kačarević, Ž.P.; Alkildani, S.; Retnasingh, S.; Barbeck, M. Bioprinting of tissue engineering scaffolds. *J. Tissue. Eng.* **2018**, *9*, 2041731418802090.
71. Chang, R.; Nam J, Sun W. Effects of dispensing pressure and nozzle diameter on cell survival from solid freeform fabrication-based direct cell writing. *Tissue. Eng Part. A.* **2008**, *14*(1), 41-8.
72. Zhang, Y.; Kumarwew, P.; Lv, S.; Xiong, D.; Zhao, H.; Cai, Z.; Zhao, X. Recent advances in 3D bioprinting of vascularized tissues. *Mater. Des.* **2021**, *199*, 109398.
73. Xiong, R.; Zhang, Z.; Chai, W.; Huang, Y.; Chrisey, D.B. Freeform drop-on-demand laser printing of 3D alginate and cellular constructs. *Biofabrication.* **2015**, *7*(4), 045011.
74. Häneke, T.; Sahara, M. Progress in bioengineering strategies for heart regenerative medicine. *Int. J. Mol. Sci.* **2022**, *23*(7), 3482.
75. Mills, R.J.; Hudson, J.E. Bioengineering adult human heart tissue: How close are we? *APL. Bioengineering.* **2019**, *3*(1), 010901.
76. Takahashi, K.; Tanabe, K.; Ohnuki, M.; Narita, M.; Ichisaka, T.; Tomoda, K.; Yamanaka, S. Induction of pluripotent stem cells from adult human fibroblasts by defined factors. *Cell.* **2007**, *131*(5), 861-872.
77. Goldfracht, I.; Protze, S.; Shiti, A.; Setter, N.; Gruber, A.; Shaheen, N.; Nartiss, Y.; Keller, G.; Gepstein, L. Generating ring-shaped engineered heart tissues from ventricular and atrial human pluripotent stem cell-derived cardiomyocytes. *Nat. Commun.* **2020**, *11*(1), 75.
78. Bremner, S.B.; Gaffney, K.S.; Sniadecki, N.J.; Mack, D.L. A Change of Heart: Human cardiac tissue engineering as a platform for drug development. *Curr. Cardiol. Rep.* **2022**, *24*(5), 473-486.
79. Quadri, F.; Soman, S.S.; Vijayavenkataraman, S. Progress in cardiovascular bioprinting. *Artif. Organs.* **2021**, *45*(7), 652-664.
80. Marchianò, S.; Bertero, A.; Murry, C.E. Learn from your elders: developmental biology lessons to guide maturation of stem cell-derived cardiomyocytes. *Pediatr. Cardiol.* **2019**, *40*(7), 1367-1387.

81. Sedlakova, V.; McTiernan, C.; Cortes, D.; Suuronen, E.J.; Alarcon, E.I. 3D bioprinted cardiac tissues and devices for tissue maturation. *Cells. Tissues. Organs*. **2022**, 211(4), 406-419.
82. Tomasina, C.; Bodet, T.; Mota, C.; Moroni, L.; Camarero-Espinosa, S. Bioprinting vasculature: materials, cells and emergent techniques. *Materials*. **2019**, 12(17), 2701.
83. Seymour, A.J.; Westerfield, A.D.; Cornelius, V.C.; Skylar-Scott, M.A.; Heilshorn, S. Bioprinted microvasculature: progressing from structure to function. *Biofabrication*. **2022**, 14(1), 022002.
84. Wang, Z.; Wang, L.; Li, T.; Liu, S.; Guo, B.; Huang, W.; Wu, Y. 3D bioprinting in cardiac tissue engineering. *Theranostics*. **2021**, 11(16), 7948-7969.
85. Barrs, R.W.; Jia, J.; Silver, S.E.; Yost, M.; Mei, Y. Biomaterials for bioprinting microvasculature. *Chem. Rev.* **2020**, 120(19), 10887-10949.
86. Ferrini, A.; Stevens, M.M.; Sattler, S.; Rosenthal, N. Toward regeneration of the heart: bioengineering strategies for immunomodulation. *Front. Cardiovasc. Med.* **2019**, 6, 26.
87. Roche, C.D.; Brereton, R.J.; Ashton, A.W.; Jackson, C.; Gentile, C. Current challenges in three-dimensional bioprinting heart tissues for cardiac surgery. *Eur. J. Cardiothorac. Surg.* **2020**, 58(3), 500-510.
88. Tillman, B.; Hardin-Young, J.; Shannon, W.; Russell, A.J.; Parenteau, N.L. Meeting the need for regenerative therapies: translation-focused analysis of U.S. regenerative medicine opportunities in cardiovascular and peripheral vascular medicine using detailed incidence data. *Tissue. Eng. Part. B. Rev.* **2021**, 19(2), 99-115.
89. Fazal, F.; Raghav, S.; Callanan, A.; Koutsos, V.; Radacsi, N. Recent advancements in the bioprinting of vascular grafts. *Biofabrication*. **2021**, 13(3), 32003.
90. Wenger, R.; Giraud, M. 3D printing applied to tissue engineered vascular grafts. *Appl. Sci (Basel)*. **2018**, 8(12), 2631.
91. Klinkert, P.; Post, P.N.; Breslau, P.J.; Bockel, J.H. Saphenous vein versus PTFE for above-knee femoropopliteal bypass. A review of the literature. *Eur. J. Vasc. Endovasc. Surg.* **2004**, 27(4), 357-62.
92. Catto, V.; Fare, S.; Freddi, G.; Tani, M.C. Vascular tissue engineering: recent advances in small diameter blood vessel regeneration. *ISRN. Vasc. Med.* **2014**, 2014, 1-27.
93. Syedain, Z.H.; Prunty, A.; Li, J.; Tranquillo, R.T. Evaluation of the probe burst test as a measure of strength for a biologically-engineered vascular graft. *J. Mech. Behav. Biomed. Mater.* **2021**, 119, 104527.
94. Pensalfini, M.; Meneghello, S.; Lintas, V.; Bircher, K.; Ehret, A.E.; Mazzza, E. The suture retention test, revisited. *J. Mech. Behav. Biomed. Mater.* **2018**, 77, 711-7.
95. Deo, K.A.; Singh, K.A.; Peak, C.W.; Algae, D.L.; Gaharwar, A.K. Bioprinting 101: design, fabrication, and evaluation of cell-laden 3D bioprinted scaffolds. *Tissue. Eng. Part A*. **2020**, 26, 318-38.
96. Xu, C.; Christensen, K.; Zhang, Z.; Huang, Y.; Fu, J.; Markwald, R.R. Predictive compensation-enabled horizontal inkjet printing of alginate tubular constructs. *Manuf. Lett.* **2013**, 1(1), 28-32.
97. Xu, C.; Chai, W.; Huang, Y.; Markwald, R.R. Scaffold-free inkjet printing of three-dimensional zigzag cellular tubes. *Biotechno Bioeng.* **2012**, 109(12), 3152-60.
98. Gudapati, H.; Dey, M.; Ozbolat, I. A comprehensive review on droplet-based bioprinting: past, present and future. *Biomaterials*. **2016**, 102, 20-42.
99. Hinton, T.J.; Jallerat, Q.; Palchesko, R.N.; Park, J.H.; Grodzicki, M.S.; Shue, H.; Ramadan MH, Hudson AR, Feinberg AW. Three-dimensional printing of complex biological structures by freeform reversible embedding of suspended hydrogels. *Sci. Adv.* **2015**, 1(9), e1500758.
100. Dikyol, C.; Altunbek, M.; Bartolo, P.; Koc, B. Multimaterial bioprinting approaches and their implementations for vascular and vascularized tissues. *Bioprinting*. **2021**, 24, e00159.
101. Hong, S.; Kim, J.S.; Jung, B.; Won, C.; Hwang, C. Coaxial bioprinting of cell-laden vascular constructs using a gelatin-tyramine bioink. *Biomater. Sci.* **2019**, 7(11), 4578-87.
102. Mohan, T.S.; Datta, P.; Nesaei, S.; Ozbolat, V.; Ozbolat, I.T. 3D coaxial bioprinting: process mechanisms, bioinks and applications. *Prog. Biomed. Eng (Bristol)*. **2022**, 4(2), 22003.
103. Milojević, M.; Vihar, B.; Banović, L.; Miško, M.; Gradišnik, L.; Zidarič, T.; Maver, U. Core/shell printing scaffolds for tissue engineering of tubular structures. *J. Vis. Exp.* **2019**, 151, e59951.
104. Wang, Y.; Kankala, R.K.; Zhu, K.; Wang, S.; Zhang, Y.S.; Chen, A.Z. Coaxial extrusion of tubular tissue constructs using a gelatin/GelMA blend bioink. *ACS. Biomater. Sci. Eng.* **2019**, 5(10), 5514-24.
105. Liu, W.; Zhong, Z.; Hu, N.; Zhou, Y.; Maggio, L.; Miri, A.K.; Fragasso, A.; Jin, Z.; Khademhosseine, A.; Zhang, Y.S. Coaxial extrusion bioprinting of 3D microfibrillar constructs with cell-favourable gelatin methacryloyl microenvironments. *Biofabrication*. **2018**, 10(2):024102.

106. Jia, W.; Gungor-Ozkerim, P.S.; Zhang, Y.S.; Yey, K.; Zhu, K.; Liu, W.; Pi, Q.; Byambaa, B.; Dokmeci, M.R.; Shin, S.R.; Khademhosseini, A. Direct 3D bioprinting of perfusable vascular constructs using a blend bioink. *Biomaterials*. **2016**, 106, 58-68.
107. Gao, Q.; Liu, Z.; Lin, Z.; Qiu, J.; Liu, Y.; Liu, A.; Wang, Y.; Xiang, M.; Chen, B.; Fu, J.; He, Y. 3D bioprinting of vessel-like structures with multilevel fluidic channels. *ACS. Biomater. Sci. Eng.* **2017**, 3(3), 399-408.
108. Jin, Q.; Zhang, G.; Xu, L.; Jin, G.; Tang, L.; Ju, J.; Zhao, W.; Hou, R. Nanofiber electrospinning combined with rotary bioprinting for fabricating small-diameter vessels with endothelium and smooth muscle. *Compos. B. Eng.* **2022**, 234(1), 109691.
109. Xu, L.; Varkey, M.; Jorgensen, A.; Ju, J.; Jin, Q.; Park, J.H.; Zhang, G.; Ke, D.; Zhao, W.; Hou, R.; Atala, A. Bioprinting small diameter blood vessel constructs with an endothelial and smooth muscle cell bilayer in a single step. *Biofabrication*. **2020**, 12(4), 045012.
110. Kang, H.W.; Lee, S.J.; Ko, I.K.; Kengla, C.; Yoo, J.J.; Atala, A. A 3D bioprinting system to produce human-scale tissue constructs with structural integrity. *Nat. Biotechnol.* **2016**, 34, 312-9.
111. Gold, K.A.; Saha, B.; Pandian, N.K.; Walther, B.K.; Palma, J.A.; Jo, J.; Cooke, J.P.; Jain, A.; Gahawar, A.K. 3D bioprinted multicellular vascular models. *Adv. Health Mater.* **2021**, 10(21), e2101141.
112. Zhang, Y.S.; Khademhosseini, A. Vascular tissue engineering: the role of 3D bioprinting. In: Walpoth BH, Bergmeister H, Bowlin GL, Kong D, Rotmans JI, Zilla P, editors. *Tissue-engineered vascular grafts*. Springer Cham; **2020**, p. 321-38.
113. Krishnamoorthy, S.; Wadnap, S.; Noorani, B.; Xu, H.; Xu, C. Investigation of gelatin methacrylate working curves in dynamic optical projection stereolithography of vascular-like constructs. *Eur. Polym. J.* **2020**, 124, 109487.
114. Thomas, A.; Orellano, I.; Lam, T.; Noichl, B.; Geiger, M.; Amler, A.; Kreuder, A.; Palmer, C.; Duda, G.; Lauster, R.; Kloeke, L. Vascular bioprinting with enzymatically degradable bioinks via multi-material projection-based stereolithography. *Acta. Biomater.* **2020**, 117, 121-32.
115. Virani, S.S.; Alonso, A.; Aparicio, H.J.; Benjamin, E.J.; Bittencourt, M.S.; Callaway, C.W.; et al. Heart disease and stroke statistics—2021 Update: A report from the American Heart Association. *Circulation*. **2021**, 143(8).
116. Sanz-Garcia, A.; Oliver-de-la-Cruz, J.; Mirabet, V.; Gandía, C.; Villagrasa, A.; Sodupe, E.; et al. Heart valve tissue engineering: how far is the bedside from the bench? *Expert. Rev. Mol. Med.* **2015**, 17, e16.
117. Butcher, J.T.; Mahler, G.J.; Hockaday, L.A. Aortic valve disease and treatment: The need for naturally engineered solutions. *Adv. Drug. Deliv. Rev.* **2011**, 63(4), 242–68.
118. Goldbarg, S.H.; Elmariah, S.; Miller, M.A.; Fuster, V.; York, N.; York, N. Insights into degenerative aortic valve disease. *J. Am. Coll. Cardiol.* **2007**, 50(13), 1205-1213.
119. Hammermeister, K.; Sethi, G.K.; Henderson, W.G.; Grover, F.L.; Oprian, C.; Rahimtoola, S.H. Outcomes 15 years after valve replacement with a mechanical versus a bioprosthetic valve: final report of the Veterans Affairs randomized trial. *J. Am. Coll. Cardiol.* **2000**, 36(4), 1152–8.
120. Head, S.J.; Çelik, M.; Kappetein, A.P. Mechanical versus bioprosthetic aortic valve replacement. *Eur. Heart. J.* **2017**, 38(28), 2183–91.
121. Kostyunin, A.E.; Yuzhalin, A.E.; Rezvova, M.A.; Ovcharenko, E.A.; Glushkova, T.V.; Kutikhin, A.G. Degeneration of bioprosthetic heart valves: update 2020. *J. Am. Heart. Assoc.* **2020**, 9(19), e018506.
122. Dvir, D.; Bourguignon, T.; Otto, C.M.; Hahn, R.T.; Rosenhek, R.; Webb, J.G.; et al. Standardized definition of structural valve degeneration for surgical and transcatheter bioprosthetic aortic valves. *Circulation*. **2018**, 137(4), 388–99.
123. Capodanno, D.; Petronio, A.S.; Prendergast, B.; Eltchaninoff, H.; Vahanian, A.; Modine, T.; et al. Standardized definitions of structural deterioration and valve failure in assessing long-term durability of transcatheter and surgical aortic bioprosthetic valves: a consensus statement from the European Association of Percutaneous Cardiovascular Interventions (EAPCI) endorsed by the European Society of Cardiology (ESC) and the European Association for Cardio-Thoracic Surgery (EACTS). *Eur. Heart. J.* **2017**, 38(45), 3382–90.
124. Sathananthan, J.; Lauck, S.; Polderman, J.; Yu, M.; Stephenson, A.; Sathananthan, G.; et al. Ten year follow-up of high-risk patients treated during the early experience with transcatheter aortic valve replacement. *Catheter. Cardiovasc. Interv.* **2021**, 97(3), E431–7.
125. Jana, S.; Tefft, B.J.; Jana, S.; Tefft, B.J.; Spoon, D.B.; Simari, R.D. Scaffolds for tissue engineering of cardiac valves. *Acta. Biomaterialia*. **2014**, 17, 2877-2893.
126. Mela, P. Subject- and leaflet-specific remodeling of polymeric heart valves for in situ tissue engineering: Challenges towards Clinical translation. *JACC. Basic. Transl. Sci.* **2020**, 5(1), 32–4.

127. Vesely, I. Heart Valve Tissue Engineering. *Circ. Res.* **2005**, 97(8), 743–55.
128. Wissing, T.B.; Bonito, V.; Bouten, C.V.C.; Smits, A.I.P.M. Biomaterial-driven in situ cardiovascular tissue engineering—a multi-disciplinary perspective. *NPJ. Regen. Med.* **2017**, 2(1), 1–19.
129. Butcher, J.T. The root problem of heart valve engineering. *Sci. Transl. Med.* **2018**, 10(440), eaat5850.
130. Zhuang, R.Z.; Lock, R.; Liu, B.; Vunjak-Novakovic, G. Opportunities and challenges in cardiac tissue engineering from an analysis of two decades of advances. *Nat. Biomed. Eng.* **2022**, 6(4), 327–38.
131. Hockaday, L.A.; Kang, K.H.; Colangelo, N.W.; Cheung, P.Y.C.; Duan, B.; Malone, E.; et al. Rapid 3D printing of anatomically accurate and mechanically heterogeneous aortic valve hydrogel scaffolds. *Biofabrication*. **2012**, 4(3), 035005.
132. Duan, B.; Hockaday, L.A.; Kang, K.H.; Butcher, J.T. 3D Bioprinting of heterogeneous aortic valve conduits with alginate/gelatin hydrogels. *J. Biomed. Mater. Res.* **2013**, 101A(5), 1255–64.
133. Duan, B.; Kapetanovic, E.; Hockaday, L.A.; Butcher, J.T. Three-dimensional printed trileaflet valve conduits using biological hydrogels and human valve interstitial cells. *Acta. Biomater.* **2014**, 10(5), 1836–46.
134. Ma, N.; Cheung, D.Y.; Butcher, J.T. Incorporating nanocrystalline cellulose into a multifunctional hydrogel for heart valve tissue engineering applications. *J. Biomed. Mater. Res.* **2022**, 110(1), 76–91.
135. Kang, L.H.; Armstrong, P.A.; Lee, L.J.; Duan, B.; Kang, K.H.; Butcher, J.T. Optimizing photo-encapsulation viability of heart valve cell types in 3D printable composite hydrogels. *Ann. Biomed. Eng.* **2017**, 45(2), 360–77.
136. Wu, S.; Duan, B.; Liu, P.; Zhang, C.; Qin, X.; Butcher, J.T. Fabrication of aligned nanofiber polymer yarn networks for anisotropic soft tissue scaffolds. *ACS. Appl. Mater. Interfaces*. **2016**, 8(26), 16950–60.
137. Sodian, R.; Loebe, M.; Hein, A.; Martin, D.P.; Hoerstrup, S.P.; Potapov, E.V.; et al. Application of stereolithography for scaffold fabrication for tissue engineered heart valves: ASAIO. *J.* **2002**, 48(1), 12–6.
138. Akpek, A. Analysis of biocompatibility characteristics of stereolithography applied three dimensional (3D) bioprinted artificial heart valves. *J. Faculty. Eng. Architec. Gazi. Uni.* **2018**, 3, 929–938.
139. Lee, A.; Hudson, A.R.; Shiwardski, D.J.; Tashman, J.W.; Hinton, T.J.; Yerneni, S.; et al. 3D bioprinting of collagen to rebuild components of the human heart. *Science*. **2019**, 365(6452), 482–7.
140. Maxson, E.L.; Young, M.D.; Noble, C.; Go, J.L.; Heidari, B.; Khorramirouz, R.; et al. In vivo remodeling of a 3D-Bioprinted tissue engineered heart valve scaffold. *Bioprinting*. **2019**, 16, e00059.
141. Noble, C.; Maxson, E.L.; Lerman, A.; Young, M.D. Mechanical and finite element evaluation of a bioprinted scaffold following recellularization in a rat subcutaneous model. *J. Mech. Behav. Biomed. Mater.* **2020**, 102, 103519.
142. Zengin, A.; Castro, J.; Habibovic, P.; Van Rijt, S. Injectable, self-healing mesoporous silica nanocomposite hydrogels with improved mechanical properties. *Nanoscale*. **2021**, 13(2), 1144–54.
143. Eivazzadeh-Keihan, R.; Radinekiyan, F.; Aliabadi, H.A.M.; Sukhtezari, S.; Tahmasebi, B.; Maleki, A.; et al. Chitosan hydrogel/silk fibroin/Mg (OH) 2 nanobiocomposite as a novel scaffold with antimicrobial activity and improved mechanical properties. *Sci. Rep.* **2021**, 11(1), 650.
144. Sarriannidis, S.O.; Rey, J.M.; Dobre, O.; González-García, C.; Dalby, M.J.; Salmeron-Sanchez, M. A tough act to follow: Collagen hydrogel modifications to improve mechanical and growth factor loading capabilities. *Mater. Today. Bio.* **2021**, 10, 100098.
145. Bas, O.; Lucarotti, S.; Angella, D.D.; Castro, N.J.; Meinert, C.; Wunner, F.M.; et al. Rational design and fabrication of multiphasic soft network composites for tissue engineering articular cartilage: A numerical model-based approach. *Chem. Engn. J.* **2018**, 340, 15–23.
146. Visser, J.; Melchels, F.P.W.; Jeon, J.E.; Van Bussel, E.M.; Kimpton, L.S.; Byrne, H.M.; et al. Reinforcement of hydrogels using three-dimensionally printed microfibres. *Nat. Commun.* **2015**, 6, 1–10.
147. Liu, N.; Ye, X.; Yao, B.; Zhao, M.; Wu, P.; Liu, G.; et al. Advances in 3D bioprinting technology for cardiac tissue engineering and regeneration. *Bioact Mater.* **2021**, 6(5), 1388–401.
148. Montero, P.; Flandes-Iparraguirre, M.; Musquiz, S.; Pérez Araluce, M.; Plano, D.; Sanmartín, C.; et al. Cells, Materials, and Fabrication Processes for Cardiac Tissue Engineering. *Front. Bioeng. Biotechnol.* **2020**, 8, 955.
149. Bejleri, D.; Streeter, B.W.; Nachlas, A.L.Y.; Brown, M.E.; Gaetani, R.; Christman, K.L.; et al. A Bioprinted Cardiac patch composed of cardiac-specific extracellular matrix and progenitor cells for heart repair. *Adv. Healthc. Mater.* **2018**, 7(23), 1800672.
150. Zhu, K.; Shin, S.R.; van Kempen, T.; Li, Y.; Ponraj, V.; Nasajpour, A.; et al. Gold nanocomposite bioink for printing 3D cardiac constructs. *Adv. Funct. Mater.* **2017**, 27(12), 1605352.
151. Erdem, A.; Darabi, M.A.; Nasiri, R.; Sangabathuni, S.; Ertas, Y.N.; Alem, H.; et al. 3D Bioprinting of oxygenated cell-laden gelatin methacryloyl constructs. *Adv. Healthc. Mater.* **2020**, 9(15), 1901794.

152. Ahrens, J.H.; Uzel, S.G.M.; Skylar-Scott, M.; Mata, M.M.; Lu, A.; Kroll, K.T.; et al. Programming cellular alignment in engineered cardiac tissue via bioprinting anisotropic organ building blocks. *Adv. Mater.* **2022**, *34*(26), 2200217.
153. Ong, C.S.; Fukunishi, T.; Zhang, H.; Huang, C.Y.; Nashed, A.; Blazeski, A.; et al. Biomaterial-free three-dimensional bioprinting of cardiac tissue using human induced pluripotent stem cell derived cardiomyocytes. *Sci. Rep.* **2017**, *7*(1), 4566.
154. Yeung, E.; Fukunishi, T.; Bai, Y.; Bedja, D.; Pitaktong, I.; Mattson, G.; et al. Cardiac regeneration using human-induced pluripotent stem cell-derived biomaterial-free 3D-bioprinted cardiac patch in vivo. *J. Tissue. Eng. Regen. Med.* **2019**, *13*(11), 2031–9.
155. Asulin, M.; Michael, I.; Shapira, A.; Dvir, T. One-step 3D printing of heart patches with built-in electronics for performance regulation. *Adv. Sci.* **2021**, *8*(9).
156. Edri, R.; Gal, I.; Noor, N.; Harel, T.; Fleischer, S.; Adadi, N.; et al. Personalized hydrogels for engineering diverse fully autologous tissue implants. *Adv. Mater.* **2019**, *31*(1), 1803895.
157. Noor, N.; Shapira, A.; Edri, R.; Gal, I.; Wertheim, L.; Dvir, T. 3D Printing of personalized thick and perfusable cardiac patches and hearts. *Adv. Sci (Weinh)*. **2019**, *6*(11), 1900344.
158. Kupfer, M.E.; Lin, W.H.; Ravikumar, V.; Qiu, K.; Wang, L.; Gao, L.; et al. In Situ expansion, differentiation, and electromechanical coupling of human cardiac muscle in a 3D bioprinted, chambered organoid. *Circ. Res.* **2020**, *127*(2), 207–24.
159. Zhang, Z.; Wu, C.; Dai, C.; Shi, Q.; Fang, G.; Xie, D.; et al. A multi-axis robot-based bioprinting system supporting natural cell function preservation and cardiac tissue fabrication. *Bioact. Mater.* **2022**, *18*, 138–50.

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.