

Review

Not peer-reviewed version

Leveraging Data from Spontaneous Animal Disease Models to Streamline Pharmaceutical Research: A Tale of Two Species

[Jonathan P Mochel](#)^{*} and [Karin Allenspach](#)

Posted Date: 26 May 2023

doi: 10.20944/preprints202305.1827.v1

Keywords: One Health; Comparative Research; Reverse Translation



Preprints.org is a free multidiscipline platform providing preprint service that is dedicated to making early versions of research outputs permanently available and citable. Preprints posted at Preprints.org appear in Web of Science, Crossref, Google Scholar, Scilit, Europe PMC.

Copyright: This is an open access article distributed under the Creative Commons Attribution License which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Review

Leveraging Data from Spontaneous Animal Disease Models to Streamline Pharmaceutical Research: A Tale of Two Species

Jonathan P Mochel * and Karin Allenspach

Iowa State University College of Veterinary Medicine, Ames, IA, U.S.A; allek@iastate.edu

* Correspondence: jmochel@iastate.edu; Tel.: + (515)-294-7424

Abstract: According to data from the U.S FDA, the likelihood of a new drug progressing from Phase I clinical trials to market approval is only 8% (DiMasi et al., 2016). Furthermore, a significant attrition rate of 35% occurs during Phase II studies due to the failure to demonstrate clinical efficacy (Arrowsmith, 2011). These statistics underscore the urgent need to reassess the current research and development (R&D) paradigm. To address this challenge, a promising approach called reverse translational pharmacology (RTP) has emerged, aiming to enhance the predictive value of preclinical models used in biomedical research by leveraging animal models that spontaneously develop analogous diseases to humans (Schneider et al., 2018). The rationale behind RTP is that knowledge gained from clinical trials in canines can subsequently inform and guide human clinical trials, leading to improved outcomes for both species. Additionally, this approach capitalizes on the opportunity to stimulate veterinary drug development by leveraging pharmacokinetic (PK), efficacy, and safety data obtained from human clinical studies (Schneider et al., 2018). The application of reverse translational pharmacology holds immense potential for expediting drug development and advancing medical knowledge. By employing animal models that naturally develop diseases akin to those found in humans, researchers can enhance the predictive capacity of preclinical models, thereby mitigating the high attrition rates encountered in later stages of clinical trials. Moreover, the exchange of knowledge and data between human and veterinary medicine engenders a synergistic relationship, fostering progress in both domains (Schneider et al., 2018). As this innovative approach gains momentum, fostering collaborations between basic and clinician scientists, pharmaceutical companies, and veterinary professionals becomes increasingly imperative. By sharing resources, expertise, and findings, we can pave the way for groundbreaking discoveries and the development of safe and efficacious treatments for diverse diseases. Embracing reverse translational pharmacology offers an avenue to redefine the R&D paradigm, bringing us closer to a future where a higher proportion of drugs successfully traverse the arduous path from the laboratory to market approval, ultimately benefiting patients across species. In this brief commentary, we highlight four examples of diseases where reverse translational modeling has been used to support pharmaceutical research, including some applications developed in our research laboratory (SMART Pharmacology) at the Iowa State University College of Veterinary Medicine. In doing so, this report also seeks to provide a brief overview of the heuristic value of computational approaches in comparative research. To further illustrate the value of these *in silico* approaches, we deliberately chose to employ some features of ChatGPT to produce the present summary (OpenAI. (2023). GPT-3.5 [ChatGPT]. Retrieved from <https://openai.com>).

Keywords: one health; comparative research; reverse translation

1. Introduction

According to data from the U.S FDA, the likelihood of a new drug progressing from Phase I clinical trials to market approval is only 8% (DiMasi et al., 2016). Furthermore, a significant attrition rate of 35% occurs during Phase II studies due to the failure to demonstrate clinical efficacy (Arrowsmith, 2011). These statistics underscore the urgent need to reassess the current research and development (R&D) paradigm. To address this challenge, a promising approach called reverse translational pharmacology (RTP) has emerged, aiming to enhance the predictive value of preclinical models used in biomedical research by leveraging animal models that spontaneously develop

analogous diseases to humans (Schneider et al., 2018). The rationale behind RTP is that knowledge gained from clinical trials in canines can subsequently inform and guide human clinical trials, leading to improved outcomes for both species. Additionally, this approach capitalizes on the opportunity to stimulate veterinary drug development by leveraging pharmacokinetic (PK), efficacy, and safety data obtained from human clinical studies (Schneider et al., 2018). A few exemplary instances where reverse translational modeling has effectively supported pharmaceutical research efforts include:

1. **Canine Osteosarcoma:** Osteosarcoma, a prevalent malignant bone tumor in humans, also afflicts canines (Fan and Khanna, 2015; Simpson et al., 2017). By investigating the similarities between human and canine osteosarcoma, scientists have successfully exploited the canine model to evaluate potential therapeutic candidates. These investigations have not only contributed to elucidating disease progression and identifying novel targets but have also facilitated the assessment of drug efficacy (Vancil et al., 2012).

2. **Feline Immunodeficiency Virus (FIV):** FIV is a viral infection in cats that closely resembles HIV infection in humans. By utilizing this naturally occurring feline model, researchers have leveraged data to investigate the pathogenesis of the virus, evaluate antiviral therapies, and gain insights into viral evasion mechanisms and immune responses (Elder et al., 2010). The findings from these investigations have significantly contributed to our understanding of HIV/AIDS, resulting in the development of potential treatments and preventive strategies for both feline and human patients (Yamamoto et al., 2010; Sahay and Yamamoto, 2018).

3. **Spontaneous Canine Epilepsy:** Canine epilepsy exhibits similarities to human epilepsy in terms of clinical manifestations, seizure types, and response to antiepileptic drugs. Through the study of dogs naturally affected by epilepsy, scientists have achieved a deeper understanding of the underlying mechanisms and identified potential therapeutic targets (Patterson et al., 2005). These discoveries have not only improved the management of epilepsy in canines but have also accelerated the development of novel treatments for human epilepsy, benefiting both species (Löscher, 2013).

The application of reverse translational pharmacology holds immense potential for expediting drug development and advancing medical knowledge. By employing animal models that naturally develop diseases akin to those found in humans, researchers can enhance the predictive capacity of preclinical models, thereby mitigating the high attrition rates encountered in later stages of clinical trials. Moreover, the exchange of knowledge and data between human and veterinary medicine engenders a synergistic relationship, fostering progress in both domains (Schneider et al., 2018).

As this innovative approach gains momentum, fostering collaborations between basic and clinician scientists, pharmaceutical companies, and veterinary professionals becomes increasingly imperative. By sharing resources, expertise, and findings, we can pave the way for groundbreaking discoveries and the development of safe and efficacious treatments for diverse diseases. Embracing reverse translational pharmacology offers an avenue to redefine the R&D paradigm, bringing us closer to a future where a higher proportion of drugs successfully traverse the arduous path from the laboratory to market approval, ultimately benefiting patients across species.

In this brief commentary, we highlight four examples of diseases where reverse translational modeling has been used to support pharmaceutical research, including some applications developed in our research laboratory (*SMART Pharmacology*) at the *Iowa State University College of Veterinary Medicine*. In doing so, this report also seeks to provide a brief overview of the heuristic value of computational approaches in comparative research. To further illustrate the value of these *in silico* approaches, we deliberately chose to employ some features of *ChatGPT* to produce the present summary (OpenAI. (2023). GPT-3.5 [ChatGPT]. Retrieved from <https://openai.com>).

2. Cancer Research

About six million dogs are diagnosed with cancer each year (Jacob, 2016), and more than half of dogs above the age of ten will develop cancers, such as lymphoma (Mochel et al., 2019), osteosarcoma (Fan and Khanna, 2015), melanoma (Atherton et al., 2016) or chemodectoma (Coto et al., 2021), among others (Jacob, 2016). As emphasized multiple times by the National Cancer Institute, mice often fail to adequately represent crucial features of human cancer, including genomic instability and tumor

heterogeneity (Gordon et al., 2009). To address this challenge, researchers have turned to canine clinical trials, capitalizing on the high genetic homology between dogs and humans to advance cancer therapies.

One remarkable example of successful reverse translational pharmacology is the study of interleukin-12 (IL-12) in canine clinical trials for malignant melanoma. The substantial genetic similarity between canine and human IL-12 prompted researchers to investigate its pharmacokinetic (PK), pharmacodynamic (PD), efficacy, and toxicity profiles in dogs with melanoma (Paolini et al., 2015). The results of the study demonstrated that high levels of IL-12 could be safely administered subcutaneously to dogs with malignant melanoma, leading to both systemic immunological and clinical activity (Paolini et al., 2015). These findings played a pivotal role in guiding the decision to proceed with a Phase I clinical trial in humans.

Moreover, preliminary studies focusing on IL-12 gene electrotransfer in dogs with melanoma have yielded promising results for the treatment of spontaneous canine tumors (Lamprecht et al., 2015). By harnessing the unique opportunity provided by naturally occurring melanoma in dogs, researchers have gained valuable insights into the therapeutic potential of IL-12 gene therapy, which may ultimately be applicable to human patients as well.

Muscle-Invasive Bladder Cancer (MIBC) is associated with an extremely poor survival rate of about 50% at 5 years follow-up, which, importantly, has not improved over the last 10 years (Patel et al., 2020). Two important factors that contribute to poor treatment outcomes are (1) the phenotypic and molecular heterogeneity of MIBC, which limits the translational value of some rodent models to evaluate therapeutic drug efficacy, and (2) the time it takes to test novel drugs and combinations of drugs in clinical trials for MIBC. As opposed to mice, canine MIBC constitutes an ideal preclinical study population to evaluate novel therapeutic options for patients with bladder cancer. In fact, canine MIBC presents with very similar molecular features, tumor heterogeneity and subtypes, as well as clinical and metastatic behavior as humans (Knapp et al., 2014). Although clinical trials in dogs with MIBC can be performed in significantly shorter timeframes than in people, so far, only very few large-scale trials using the canine model have been performed. Validation of predictive ex vivo assays to evaluate the potential efficacy of novel treatment modalities before formal testing in canine clinical trials will therefore enhance translational efforts for the development of novel therapeutics. A promising technology for the preclinical screening of candidate drug efficacy lies in the culture of patient-derived tumor organoids (PDOs) (Elbadawy et al., 2019; Mastri et al., 2021; Xie and Song, 2023). Compared with 2D cell lines, 3D PDOs better reflect the underlying biology of the tumor and are therefore considered more robust models to accurately predict clinical response to treatment compared with molecular testing alone. We therefore propose that future therapeutic leads for MIBC be screened ex vivo using canine organoids to select the most promising drug candidates. Subsequently, these novel therapeutics could be tested in vivo in dogs with bladder cancer prior to clinical testing in human patients with MIBC.

The importance of these canine clinical trials in advancing cancer therapies cannot be overstated. Through reverse translational pharmacology, researchers have been able to overcome some of the limitations of traditional preclinical models, ultimately improving the chances of success in human clinical trials. The insights gained from canine studies have the potential to transform the drug development pathway, providing a more accurate representation of disease processes and treatment responses. Altogether, the utilization of canine clinical trials has emerged as a powerful tool in advancing cancer therapies. By leveraging the genetic similarities between dogs and humans, researchers have successfully evaluated the safety, efficacy, and pharmacological properties of novel treatments, including immunotherapies (Park et al., 2016; Mason et al., 2021; Ammons et al., 2023). The findings from these reverse translational studies have not only guided the design of human clinical trials but also provided valuable insights into the treatment of spontaneous canine tumors. As we continue to explore the potential of reverse translational pharmacology, collaborations between basic and clinical scientists, pharmaceutical companies, and veterinary professionals will be pivotal in translating these discoveries into tangible benefits for both human and canine patients. A future step in that direction would be to generalize computational approaches for characterizing the

pharmacokinetics and pharmacodynamics of anticancer drugs used alone or in combination, as previously described in the human literature (van Hasselt and Iyengar, 2018; Zhu, 2018; Aghamiri et al., 2022; Lam et al., 2022), including data originating from our group (Schneider et al., 2018; Vaghi et al., 2020).

3. Inflammatory Bowel Disease

Inflammatory Bowel Disease (IBD) is a chronic and prevalent disorder characterized by inflammation in the gastrointestinal tract. It affects both humans and dogs, and interestingly, dogs with IBD share remarkable similarities with human patients in terms of clinical symptoms and molecular pathways involved in the disease (Jergens, 1999; Cerquetella et al., 2010; Honnefer et al., 2014; Atherly et al., 2019; Allenspach and Mochel 2021). This parallelism presents an excellent opportunity for the application of reverse translational modeling to enhance our understanding and treatment of IBD. The application of reverse translational modeling in IBD research has several advantages. Firstly, by using dogs as a disease model, researchers can gain insights into the pathophysiology of IBD in a more clinically relevant context. Dogs naturally develop spontaneous cases of IBD, allowing for the investigation of disease progression and response to treatment in a manner that closely mirrors human patients. This provides a valuable opportunity to evaluate the efficacy and safety of potential therapeutics and refine treatment protocols before translating them to human clinical trials.

A recent collaborative study between Iowa State University and the University of Navarra employed a network systems approach based on Boolean equations to model IBD (Balbas-Martinez et al., 2017, 2018). This mathematical modeling technique allows for the construction of a network of interactions between different molecular components involved in the disease. These models can capture the intricate interactions between multiple molecular components and provide predictions about the effects of perturbations or interventions. By integrating experimental data and computational simulations, researchers can optimize treatment strategies, identify key molecular targets, and minimize the potential risks associated with clinical trials. By simulating these interactions, researchers can gain insights into the underlying mechanisms and identify potential therapeutic targets. In this study, the focus was on the purinergic receptor P2X7, which has been implicated in the inflammatory response in IBD. By using the model, we aimed to simulate the effect of a non-competitive antagonist of the P2X7 receptor and determine the optimal dose for an upcoming clinical trial in dogs diagnosed with IBD. This approach holds promise for refining the dosing strategy and increasing the chances of success in clinical trials.

There is currently no cure for IBD and many therapeutic interventions developed over the past three decades have consistently failed to demonstrate evidence of efficacy in canine clinical trials (Makielski et al., 2019). There is, therefore, a *critical need* to develop robust drug screening tools to accelerate the availability of effective and safe therapeutic strategies for management of canine IBD. Our research group has focused on the development of canine intestinal organoids to enhance our understanding of disease pathogenesis and to screen novel therapeutic drug candidates (Ambrosini et al., 2020; Kopper et al., 2021; Gabriel et al., 2022a, 2022b).

Organoids (2017 *Nature* Method of the Year and 2018 *Science* Breakthrough of the Year) are sampled from individuals with different genotypes, environmental risk factors or drug sensitivity profiles, thereby reflecting the diversity of the disease background across many demographics. The biologic potential of 3D organoids is derived from the method's ability to harness hard-wired cellular programming within higher order cellular tissue organization to *more predictably evaluate drug activity* in vitro compared with conventional 2D systems (Fatehullah et al., 2016). Recent research from the WSAVA emphasized that *cellular* and *architectural* changes of the *epithelium* were the most *important determinants of disease pathogenesis* in IBD (Day et al., 2008; Allenspach et al., 2019). Therefore, developing methods focusing on the epithelial cell layer, such as organoids, will provide key information for understanding disease mechanisms in canine IBD. A promising development in this regard derives from our recent findings demonstrating that stem cell-derived intestinal organoids can be replicated ex vivo in dogs with IBD (Mochel et al., 2017; Chandra et al., 2019; Sahoo et al.,

2022). This is a significant step toward the development of a completely new animal disease model for veterinary research, with the added benefit that data generated in dogs can be used to streamline similar clinical trials in human patients with IBD (Hong et al., 2017). This research is part of a broader 5-year research collaboration agreement (RCA) with the Office of New Animal Drug Evaluation (ONADE) of the FDA. This RCA is targeted at characterizing the intestinal permeability of a range of compounds in 3D canine organoids. To reach that goal, we first had to establish successful culture of organoids in a confluent monolayer using transwells. Our laboratory has just completed a first series of passive permeability assays in large intestinal organoids with metoprolol, propranolol, atenolol, and FITC-dextran, and benchmarked our results vs. conventional 2D systems, such as Caco-2 cells (Sahoo et al., 2023). Predictions of apparent permeability (Papp) parameters derived by the FDA were consistent with previously published literature. With successful completion of this first milestone, we are now evaluating the predictive performances of 3D organoids to model active drug transport.

The application of reverse translational modeling in the study of IBD holds significant promise for advancing our understanding and treatment of this complex disease. By leveraging the similarities between dogs and humans in terms of clinical presentation and molecular pathways, scientists can gain valuable insights into disease mechanisms and develop more effective therapeutic strategies. The integration of mathematical modeling techniques further enhances our ability to simulate and predict the effects of interventions, optimizing the design and outcomes of clinical trials. Through collaborative efforts and innovative approaches, reverse translational modeling in IBD research offers great potential for improving patient outcomes in both veterinary and human medicine.

4. Ocular Diseases

The ocular surface plays a critical role in maintaining the health and integrity of the eye. It is a complex and dynamic system consisting of the secreted tear film, lacrimal gland(s), eyelids, meibomian glands, cornea, conjunctiva, sclera, and nasolacrimal drainage apparatus. In a recent review by our group (Sebbag and Mochel, 2020), we have highlighted how the canine ocular surface closely resembles the human ocular surface in terms of anatomical structure, tear film composition, and immune responses. Dogs naturally develop a range of ocular surface disorders, such as dry eye syndrome, keratoconjunctivitis sicca, and corneal ulcers, which share similarities with their human counterparts. By studying these conditions in dogs, clinician scientists can gain insights into the underlying mechanisms, pathology, and potential therapeutic interventions. Specifically, several studies originating from our lab have investigated various aspects of ocular health and tear composition in different animal models, providing valuable insights into ophthalmic research. In a seminal study, Sebbag et al. (2018) evaluated the impact of flow rate, collection devices, and extraction methods on tear concentrations following oral administration of doxycycline in dogs and cats. They found that different collection methods influenced tear concentration measurements. In another study, Sebbag et al. (2019) investigated the kinetics of fluorescein in the tear film of beagle dogs after eye drop instillation. They examined the role of droplet size and its impact on tear film dynamics, suggesting that droplet size may not significantly affect tear clearance. Histamine-induced conjunctivitis and breakdown of the blood-tear barrier in dogs were later explored as a model for ocular pharmacology and therapeutics by Sebbag et al. (2019). The study provided insights into the mechanisms underlying ocular inflammation and suggested potential therapeutic strategies. Uhl et al. (2019) focused on cats and described the clinical features of cats with aqueous tear deficiency. The retrospective case series highlighted the diagnostic and management challenges associated with this condition in feline patients. Other studies investigated tear collection methods and their impact on tear composition. Bertram et al. (2021) examined the influence of Schirmer strip wetness on tear volume and total protein content in canine tears. The authors found that tear volume absorbed and recovered by the Schirmer strip, as well as total protein content, were influenced by the wetness of the strip. At last, Terhaar et al. (2021) focused on horses and evaluated serum albumin and total protein concentration in the tear film of horses with healthy or diseased eyes. Their findings indicated differences in tear protein composition between healthy and diseased eyes in horses.

Collectively, these studies demonstrate the diverse research conducted in the field of comparative ophthalmology, utilizing different animal models to investigate tear composition, ocular diseases, drug delivery, and tear collection methods. These findings contribute to our understanding of ocular health and provide valuable insights for the development of ophthalmic therapies and diagnostic approaches.

5. Congestive Heart Failure

Congestive heart failure (CHF) is a complex condition that affects both humans and dogs, and its development often involves the overactivation of the renin-angiotensin-aldosterone (RAA) pathway (Watkins et al., 1976; Roig et al., 2000; Güder et al., 2007; Mochel and Danhof, 2015). Understanding the dynamics of RAA biomarkers and their response to treatment is crucial for optimizing therapeutic strategies. In a seminal study by our group, the potential effect of chronobiology and food intake on the pharmacodynamic of angiotensin-converting enzyme (ACE) inhibitors such as benazepril was further discussed (Mochel et al., 2013a, 2014). This study revealed that the timing of food intake plays a significant role in synchronizing the RAA pathway in dogs, suggesting that the administration of ACE inhibitors should be adjusted based on the time of food intake. This finding is of particular importance as it highlights the need for individualized dosing regimens that take into account the influence of food on drug absorption and the subsequent modulation of RAA biomarkers.

Contrary to previous assumptions that ACE activity alone could reliably predict RAA activity, Mochel et al. (2013b, 2013c, 2015) established that ACE activity alone is not sufficient to gauge the overall activity of the RAA pathway in dogs. This finding aligns with previous studies conducted in human patients with CHF, underscoring the complexity and multifaceted nature of RAA regulation (Van de Wal et al., 2006). The use of a semi-mechanistic PK/PD model in this other study represents a significant advancement in our understanding of the interplay between food intake, drug administration, and RAA activity in dogs. PK/PD models allow for the integration of pharmacokinetic data (drug concentration profiles) with pharmacodynamic measurements (biomarker responses) to provide insights into the relationship between drug exposure and its effects on the biological system. Little is known about the optimal dosing of the ACEI benazepril in dogs with CHF. Seminal studies (Toutain et al., 2000; King et al., 2003) had initially suggested that the pharmacological action of benazepril was relatively independent of doses > 0.25 mg/kg P.O, thereby providing a rationale for the current European label dose of 0.25 mg/kg P.O q24h in dogs with cardiovascular diseases. However, most of these earlier studies on benazepril pharmacodynamics relied on measures of ACE activity, which is a sub-optimal endpoint to characterize the effect of benazepril on the RAAS (Mochel and Fink., 2013). Quantitative systems pharmacology (QSP) modeling is an established framework for characterizing the effect of therapeutic drugs on complex biological systems, such as the renin-angiotensin cascade (van der Graaf and Benson, 2011). Importantly, by simulating virtual clinical trials from such a model, one can directly and quantitatively predict the effects of various hypothetical dosing schedules for therapeutic optimization purposes. By employing this modeling approach, scientists can now simulate different scenarios and optimize dosing strategies to achieve desired therapeutic outcomes. In the most recent publication from our group, this approach was used to develop a software implementation of the benazeprilat-RAAS QSP model, which is capable of rapidly simulating clinical trials for optimal dosing of ACEI at the bedside (Schneider et al., 2023). Using this simulation engine, we could predict and quantify the effect of varying dosing schedules of benazepril on the classical and alternative arm of the RAAS on top of diurnal variations. By developing an easy-to-use simulation interface for our model, we are now able to make a first prediction of the optimal dose/time of benazepril administration in dogs in support of future investigations in patients with CHF.

Altogether, these research studies sheds light on the intricate dynamics of the RAA pathway in dogs and its modulation by food intake. The findings emphasize the importance of adjusting dosing regimens of ACE inhibitors according to the timing of food intake in order to optimize therapeutic outcomes. Furthermore, the study challenges the notion that ACE activity alone can serve as a reliable

predictor of RAA activity in dogs, reinforcing the complexity of RAA regulation in CHF. These insights, gained through the application of semi-mechanistic PK/PD modeling, contribute to the development of tailored treatment strategies for CHF in both veterinary and human medicine. One such example of our search for new therapeutic drugs in the field involves our work on the pharmacokinetics and pharmacodynamics of the angiotensin receptor–neprilysin inhibitor (ARNi) LCZ696 (Mochel et al., 2019).

6. Conclusions

Reverse translational pharmacology represents a promising approach to enhance preclinical models and facilitate the development of effective therapeutics for both human and veterinary use. By utilizing animal models that spontaneously develop diseases analogous to those in humans, researchers can improve the predictability of preclinical studies, leading to better characterization of drug efficacy and safety.

To achieve success in reverse translational pharmacology, collaboration and cross-disciplinary communication are crucial. Quantitative scientists, human health specialists, and animal health experts need to work together to exchange knowledge, share insights, and collectively design studies that address the specific needs of both human and veterinary medicine. This collaborative approach allows for the identification of shared molecular targets, evaluation of therapeutic interventions, and the establishment of parallel drug development programs for human and veterinary species.

Additionally, regulatory incentives play a vital role in promoting the adoption of reverse translational pharmacology. Regulatory agencies can provide guidance and support for the development of parallel drug development programs, acknowledging the value of veterinary studies in advancing human medicine. By recognizing the potential benefits of reverse translational modeling, regulators can encourage researchers and pharmaceutical companies to invest in this approach, leading to improved drug development strategies and better outcomes for both human and veterinary patients.

In summary, reverse translational pharmacology holds great promise in refining the drug development process by leveraging the similarities between human and veterinary diseases. By fostering collaboration, promoting regulatory incentives, and emphasizing the importance of cross-disciplinary approaches, researchers can unlock the full potential of reverse translational pharmacology, ultimately improving the selection and efficacy of therapeutics for a wide range of diseases in both humans and veterinary species.

References

- Aghamiri** SS, Amin R, Helikar T. Recent applications of quantitative systems pharmacology and machine learning models across diseases. *J Pharmacokinet Pharmacodyn*. 2022 Feb;49(1):19-37. doi: 10.1007/s10928-021-09790-9. Epub 2021 Oct 20. PMID: 34671863; PMCID: PMC8528185.
- Allenspach** KA, Mochel JP, Du Y, Priestnall SL, Moore F, Slayter M, Rodrigues A, Ackermann M, Krockenberger M, Mansell J; WSAVA GI Standardization Working Group; Luckschander N, Wang C, Suchodolski J, Berghoff N, Jergens AE. Correlating Gastrointestinal Histopathologic Changes to Clinical Disease Activity in Dogs With Idiopathic Inflammatory Bowel Disease. *Vet Pathol*. 2019 May;56(3):435-443. doi: 10.1177/0300985818813090. Epub 2018 Dec 18. PMID: 30563436.
- Allenspach** K, Mochel JP. Current diagnostics for chronic enteropathies in dogs. *Vet Clin Pathol*. 2022 Feb;50 Suppl 1:18-28. doi: 10.1111/vcp.13068. Epub 2021 Oct 26. PMID: 34699081.
- Ambrosini** YM, Park Y, Jergens AE, Shin W, Min S, Atherly T, Borcharding DC, Jang J, Allenspach K, Mochel JP, Kim HJ. Recapitulation of the accessible interface of biopsy-derived canine intestinal organoids to study epithelial-luminal interactions. *PLoS One*. 2020 Apr 17;15(4):e0231423. doi: 10.1371/journal.pone.0231423. PMID: 32302323; PMCID: PMC7164685.
- Ammons** DT, MacDonald CR, Chow L, Repasky EA, Dow S. Chronic adrenergic stress and generation of myeloid-derived suppressor cells: Implications for cancer immunotherapy in dogs. *Vet Comp Oncol*. 2023 Jun;21(2):159-165. doi: 10.1111/vco.12891. Epub 2023 Mar 23. PMID: 36876492.
- Arrowsmith** J. Trial watch: Phase II failures: 2008-2010. *Nat Rev Drug Discov*. 2011 May;10(5):328-9. doi: 10.1038/nrd3439. PMID: 21532551.

- Atherly** T, Rossi G, White R, Seo YJ, Wang C, Ackermann M, Breuer M, Allenspach K, Mochel JP, Jergens AE. Glucocorticoid and dietary effects on mucosal microbiota in canine inflammatory bowel disease. *PLoS One*. 2019 Dec 30;14(12):e0226780. doi: 10.1371/journal.pone.0226780. PMID: 31887117; PMCID: PMC6936794.
- Atherton** MJ, Morris JS, McDermott MR, Lichty BD. Cancer immunology and canine malignant melanoma: A comparative review. *Vet Immunol Immunopathol*. 2016 Jan;169:15-26. doi: 10.1016/j.vetimm.2015.11.003. Epub 2015 Dec 1. PMID: 26827834.
- Balbas-Martinez** V, Allenspach K, Kingsbury D, Jergens AE, Troconiz I, Mochel JP. One Health: Translational and Reverse Translational Modeling of Inflammatory Bowel Disease using an advanced Boolean Network. *Population Approach Group in Europe Meeting*. Budapest (2017).
- Balbas-Martinez** V, Ruiz-Cerdá L, Irurzun-Arana I, González-García I, Vermeulen A, Gómez-Mantilla JD, Trocóniz IF. A systems pharmacology model for inflammatory bowel disease. *PLoS One*. 2018 Mar 7;13(3):e0192949. doi: 10.1371/journal.pone.0192949. PMID: 29513758; PMCID: PMC5841748.
- Bertram** M, Allbaugh RA, Mochel JP, Peraza J, Page L, Sebbag L. Influence of Schirmer strip wetness on volume absorbed, volume recovered, and total protein content in canine tears. *Vet Ophthalmol*. 2021 Jul;24(4):425-428. doi: 10.1111/vop.12876. Epub 2021 Mar 15. PMID: 33720492.
- Cerquetella** M, Spaterna A, Laus F, Tesei B, Rossi G, Antonelli E, Villanacci V, Bassotti G. Inflammatory bowel disease in the dog: differences and similarities with humans. *World J Gastroenterol*. 2010 Mar 7;16(9):1050-6. doi: 10.3748/wjg.v16.i9.1050. PMID: 20205273; PMCID: PMC2835779.
- Chandra** L, Borcharding DC, Kingsbury D, Atherly T, Ambrosini YM, Bourgois-Mochel A, Yuan W, Kimber M, Qi Y, Wang Q, Wannemuehler M, Ellinwood NM, Snella E, Martin M, Skala M, Meyerholz D, Estes M, Fernandez-Zapico ME, Jergens AE, Mochel JP, Allenspach K. Derivation of adult canine intestinal organoids for translational research in gastroenterology. *BMC Biol*. 2019 Apr 11;17(1):33. doi: 10.1186/s12915-019-0652-6. PMID: 30975131; PMCID: PMC6460554.
- Coto** GM, Musser ML, Tropf MA, Ward JL, Seo YJ, Mochel JP, Johannes CM. A Multi-Institutional Retrospective Analysis of Toceranib Phosphate for Presumed or Confirmed Canine Aortic Body Chemodectomas. *Front Vet Sci*. 2021 Feb 5;8:635057. doi: 10.3389/fvets.2021.635057. PMID: 33614771; PMCID: PMC7892462.
- Day** MJ, Bilzer T, Mansell J, Wilcock B, Hall EJ, Jergens A, Minami T, Willard M, Washabau R; World Small Animal Veterinary Association Gastrointestinal Standardization Group. Histopathological standards for the diagnosis of gastrointestinal inflammation in endoscopic biopsy samples from the dog and cat: a report from the World Small Animal Veterinary Association Gastrointestinal Standardization Group. *J Comp Pathol*. 2008 Feb-Apr;138 Suppl 1:S1-43. doi: 10.1016/j.jcpa.2008.01.001. Epub 2008 Mar 11. PMID: 18336828.
- DiMasi** JA, Grabowski HG, Hansen RW. Innovation in the pharmaceutical industry: New estimates of R&D costs. *J Health Econ*. 2016 May;47:20-33. doi: 10.1016/j.jhealeco.2016.01.012. Epub 2016 Feb 12. PMID: 26928437.
- Elbadawy** M, Usui T, Mori T, Tsunedomi R, Hazama S, Nabeta R, Uchide T, Fukushima R, Yoshida T, Shibutani M, Tanaka T, Masuda S, Okada R, Ichikawa R, Omatsu T, Mizutani T, Katayama Y, Noguchi S, Iwai S, Nakagawa T, Shinohara Y, Kaneda M, Yamawaki H, Sasaki K. Establishment of a novel experimental model for muscle-invasive bladder cancer using a dog bladder cancer organoid culture. *Cancer Sci*. 2019 Sep;110(9):2806-2821. doi: 10.1111/cas.14118. Epub 2019 Jul 23. PMID: 31254429; PMCID: PMC6726682.
- Elder** JH, Lin YC, Fink E, Grant CK. Feline immunodeficiency virus (FIV) as a model for study of lentivirus infections: parallels with HIV. *Curr HIV Res*. 2010 Jan;8(1):73-80. doi: 10.2174/157016210790416389. PMID: 20210782; PMCID: PMC2853889.
- Fan** TM, Khanna C. Comparative Aspects of Osteosarcoma Pathogenesis in Humans and Dogs. *Vet Sci*. 2015 Aug 17;2(3):210-230. doi: 10.3390/vetsci2030210. PMID: 29061942; PMCID: PMC5644632.
- Fatehullah** A, Tan SH, Barker N. Organoids as an in vitro model of human development and disease. *Nat Cell Biol*. 2016 Mar;18(3):246-54. doi: 10.1038/ncb3312. PMID: 26911908.
- Gabriel** V, Zdyrski C, Sahoo DK, Dao K, Bourgois-Mochel A, Kopper J, Zeng XL, Estes MK, Mochel JP, Allenspach K. Standardization and Maintenance of 3D Canine Hepatic and Intestinal Organoid Cultures for Use in Biomedical Research. *J Vis Exp*. 2022 Jan 31;(179). doi: 10.3791/63515. PMID: 35156656.
- Gabriel** V, Zdyrski C, Sahoo DK, Dao K, Bourgois-Mochel A, Atherly T, Martinez MN, Volpe DA, Kopper J, Allenspach K, Mochel JP. Canine Intestinal Organoids in a Dual-Chamber Permeable Support System. *J Vis Exp*. 2022 Mar 2;(181). doi: 10.3791/63612. PMID: 35311824.
- Gordon** I, Paoloni M, Mazcko C, Khanna C. The Comparative Oncology Trials Consortium: using spontaneously occurring cancers in dogs to inform the cancer drug development pathway. *PLoS Med*. 2009 Oct;6(10):e1000161. doi: 10.1371/journal.pmed.1000161. Epub 2009 Oct 13. PMID: 19823573; PMCID: PMC2753665.
- Güder** G, Bauersachs J, Frantz S, Weismann D, Allolio B, Ertl G, Angermann CE, Störk S. Complementary and incremental mortality risk prediction by cortisol and aldosterone in chronic heart failure. *Circulation*. 2007 3;115(13):1754-61. doi: 10.1161/CIRCULATIONAHA.106.653964. Epub 2007 Mar 19. PMID: 17372171.
- Hong** SN, Dunn JC, Stelzner M, Martín MG. Concise Review: The Potential Use of Intestinal Stem Cells to Treat Patients with Intestinal Failure. *Stem Cells Transl Med*. 2017 Feb;6(2):666-676. doi: 10.5966/sctm.2016-0153. Epub 2016 Sep 16. PMID: 28191783; PMCID: PMC5442796.

- Honneffer** JB, Minamoto Y, Suchodolski JS. Microbiota alterations in acute and chronic gastrointestinal inflammation of cats and dogs. *World J Gastroenterol*. 2014 Nov 28;20(44):16489-97. doi: 10.3748/wjg.v20.i44.16489. PMID: 25469017; PMCID: PMC4248192.
- Jacob** JA. Researchers Turn to Canine Clinical Trials to Advance Cancer Therapies. *JAMA*. 2016 Apr 19;315(15):1550-2. doi: 10.1001/jama.2016.0082. PMID: 27027696.
- Jergens** AE. Inflammatory bowel disease. Current perspectives. *Vet Clin North Am Small Anim Pract*. 1999 Mar;29(2):501-21, vii. PMID: 10202800.
- King** JN, Maurer M, Toutain PL. Pharmacokinetic/pharmacodynamic modelling of the disposition and effect of benazepril and benazeprilat in cats. *J Vet Pharmacol Ther*. 2003 Jun;26(3):213-24. doi: 10.1046/j.1365-2885.2003.00468.x. PMID: 12755906.
- Knapp** DW, Ramos-Vara JA, Moore GE, Dhawan D, Bonney PL, Young KE. Urinary bladder cancer in dogs, a naturally occurring model for cancer biology and drug development. *ILAR J*. 2014;55(1):100-18. doi: 10.1093/ilar/ilu018. PMID: 24936033.
- Kopper** JJ, Iennarella-Servantez C, Jergens AE, Sahoo DK, Guillot E, Bourgois-Mochel A, Martinez MN, Allenspach K, Mochel JP. Harnessing the Biology of Canine Intestinal Organoids to Heighten Understanding of Inflammatory Bowel Disease Pathogenesis and Accelerate Drug Discovery: A One Health Approach. *Front Toxicol*. 2021 Nov 10;3:773953. doi: 10.3389/ftox.2021.773953. PMID: 35295115; PMCID: PMC8915821.
- Lamprecht** U, Kamensek U, Stimac M, Sersa G, Tozon N, Bosnjak M, Brozic A, de Sá Oliveira GG, Nakagawa T, Saeki K, Cemazar M. Gene Electrotransfer of Canine Interleukin 12 into Canine Melanoma Cell Lines. *J Membr Biol*. 2015 Oct;248(5):909-17. doi: 10.1007/s00232-015-9800-2. Epub 2015 Apr 4. PMID: 25840833.
- Lam** I, Pilla Reddy V, Ball K, Arends RH, Mac Gabhann F. Development of and insights from systems pharmacology models of antibody-drug conjugates. *CPT Pharmacometrics Syst Pharmacol*. 2022 Aug;11(8):967-990. doi: 10.1002/psp4.12833. Epub 2022 Jul 3. PMID: 35712824; PMCID: PMC9381915.
- Löscher** W. Animal models of epilepsy for the development of antiepileptogenic and disease-modifying drugs. A comparison of the pharmacology of kindling and post-status epilepticus models of temporal lobe epilepsy. *Epilepsy Res*. 2002 Jun;50(1-2):105-23. doi: 10.1016/s0920-1211(02)00073-6. PMID: 12151122.
- Makielski** K, Cullen J, O'Connor A, Jergens AE. Narrative review of therapies for chronic enteropathies in dogs and cats. *J Vet Intern Med*. 2019 Jan;33(1):11-22. doi: 10.1111/jvim.15345. Epub 2018 Dec 6. Review. PubMed PMID: 30523666; PubMed Central PMCID: PMC6335544.
- Mason** NJ, Chester N, Xiong A, Rotolo A, Wu Y, Yoshimoto S, Glassman P, Gulendran G, Siegel DL. Development of a fully canine anti-canine CTLA4 monoclonal antibody for comparative translational research in dogs with spontaneous tumors. *MAbs*. 2021 Jan-Dec;13(1):2004638. doi: 10.1080/19420862.2021.2004638. PMID: 34856888; PMCID: PMC8726733.
- Masters** AK, Berger DJ, Ware WA, Langenfeld NR, Coetzee JF, Mochel JPM, Ward JL. Effects of short-term anti-inflammatory glucocorticoid treatment on clinicopathologic, echocardiographic, and hemodynamic variables in systemically healthy dogs. *Am J Vet Res*. 2018 Apr;79(4):411-423. doi: 10.2460/ajvr.79.4.411. PMID: 29583045.
- Mastri** M, Ramakrishnan S, Shah SD, Karasik E, Gillard BM, Moser MT, Farmer BK, Azabdaftari G, Chatta GS, Woloszynska A, Eng KH, Foster BA, Huss WJ. Patient derived models of bladder cancer enrich the signal of the tumor cell transcriptome facilitating the analysis of the tumor cell compartment. *Am J Clin Exp Urol*. 2021 Dec 15;9(6):416-434. PMID: 34993263; PMCID: PMC8727788.
- Mochel** JP, Fink M, Peyrou M, Desevaux C, Deurinck M, Giraudel JM, Danhof M. Chronobiology of the renin-angiotensin-aldosterone system in dogs: relation to blood pressure and renal physiology. *Chronobiol Int*. 2013 Nov;30(9):1144-59. doi: 10.3109/07420528.2013.807275. Epub 2013 Aug 9. PMID: 23931032.
- Mochel** JP, Peyrou M, Fink M, Strehlau G, Mohamed R, Giraudel JM, Ploeger B, Danhof M. Capturing the dynamics of systemic Renin-Angiotensin-Aldosterone System (RAAS) peptides heightens the understanding of the effect of benazepril in dogs. *J Vet Pharmacol Ther*. 2013 Apr;36(2):174-80. doi: 10.1111/j.1365-2885.2012.01406.x. Epub 2012 May 8. PMID: 22568394.
- Mochel** J, Fink M. Response to Letter from Atkins et al. *J Vet Pharmacol Ther*. 2013 Apr. DOI: 10.1111/jvp.12017.
- Mochel** JP, Fink M, Bon C, Peyrou M, Bieth B, Desevaux C, Deurinck M, Giraudel JM, Danhof M. Influence of feeding schedules on the chronobiology of renin activity, urinary electrolytes and blood pressure in dogs. *Chronobiol Int*. 2014 Jun;31(5):715-30. doi: 10.3109/07420528.2014.897711. Epub 2014 Mar 21. PMID: 24654920.
- Mochel** JP, Fink M, Peyrou M, Soubret A, Giraudel JM, Danhof M. Pharmacokinetic/Pharmacodynamic Modeling of Renin-Angiotensin Aldosterone Biomarkers Following Angiotensin-Converting Enzyme (ACE) Inhibition Therapy with Benazepril in Dogs. *Pharm Res*. 2015 Jun;32(6):1931-46. doi: 10.1007/s11095-014-1587-9. Epub 2014 Dec 2. PMID: 25446774.
- Mochel** JP, Danhof M. Chronobiology and Pharmacologic Modulation of the Renin-Angiotensin-Aldosterone System in Dogs: What Have We Learned? *Rev Physiol Biochem Pharmacol*. 2015;169:43-69. doi: 10.1007/112_2015_27. PMID: 26428686.
- Mochel** JP, Jergens AE, Kingsbury D, Kim HJ, Martín MG, Allenspach K. Intestinal Stem Cells to Advance Drug Development, Precision, and Regenerative Medicine: A Paradigm Shift in Translational Research. *AAPS J*. 2017 Dec 12;20(1):17. doi: 10.1208/s12248-017-0178-1. PMID: 29234895; PMCID: PMC6044282.

- Mochel JP**, Teng CH, Peyrou M, Giraudel J, Danhof M, Rigel DF. Sacubitril/valsartan (LCZ696) significantly reduces aldosterone and increases cGMP circulating levels in a canine model of RAAS activation. *Eur J Pharm Sci.* 2019 Feb 1;128:103-111. doi: 10.1016/j.ejps.2018.11.037. Epub 2018 Nov 30. PMID: 30508581.
- Paoloni M**, Khanna C. Translation of new cancer treatments from pet dogs to humans. *Nat Rev Cancer.* 2008 Feb;8(2):147-56. doi: 10.1038/nrc2273. PMID: 18202698.
- Paoloni M**, Mazcko C, Selting K, Lana S, Barber L, Phillips J, Skorupski K, Vail D, Wilson H, Biller B, Avery A, Kiupel M, LeBlanc A, Bernhardt A, Brunkhorst B, Tighe R, Khanna C. Defining the Pharmacodynamic Profile and Therapeutic Index of NHS-IL12 Immunocytokine in Dogs with Malignant Melanoma. *PLoS One.* 2015 Jun 19;10(6):e0129954. doi: 10.1371/journal.pone.0129954. PMID: 26091536; PMCID: PMC4474860.
- Park JS**, Withers SS, Modiano JF, Kent MS, Chen M, Luna JL, Culp WTN, Sparger EE, Rebhun RB, Monjazebe AM, Murphy WJ, Canter RJ. Canine cancer immunotherapy studies: linking mouse and human. *J Immunother Cancer.* 2016 Dec 20;4:97. doi: 10.1186/s40425-016-0200-7. PMID: 28031824; PMCID: PMC5171656.
- Patel VG**, Oh WK, Galsky MD. Treatment of muscle-invasive and advanced bladder cancer in 2020. *CA Cancer J Clin.* 2020 Sep;70(5):404-423. doi: 10.3322/caac.21631. Epub 2020 Aug 7. PMID: 32767764.
- Patterson EE**, Armstrong PJ, O'Brien DP, Roberts MC, Johnson GS, Mickelson JR. Clinical description and mode of inheritance of idiopathic epilepsy in English springer spaniels. *J Am Vet Med Assoc.* 2005 Jan 1;226(1):54-8. doi: 10.2460/javma.2005.226.54. PMID: 15646572.
- Roig E**, Perez-Villa F, Morales M, Jiménez W, Orús J, Heras M, Sanz G. Clinical implications of increased plasma angiotensin II despite ACE inhibitor therapy in patients with congestive heart failure. *Eur Heart J.* 2000 Jan;21(1):53-7. doi: 10.1053/euhj.1999.1740. PMID: 10610744.
- Sahay B**, Yamamoto JK. Lessons Learned in Developing a Commercial FIV Vaccine: The Immunity Required for an Effective HIV-1 Vaccine. *Viruses.* 2018 May 22;10(5):277. doi: 10.3390/v10050277. PMID: 29789450; PMCID: PMC5977270.
- Sahoo DK**, Borcharding DC, Chandra L, Jergens AE, Atherly T, Bourgois-Mochel A, Ellinwood NM, Snella E, Severin AJ, Martin M, Allenspach K, Mochel JP. Differential Transcriptomic Profiles Following Stimulation with Lipopolysaccharide in Intestinal Organoids from Dogs with Inflammatory Bowel Disease and Intestinal Mast Cell Tumor. *Cancers (Basel).* 2022 Jul 20;14(14):3525. doi: 10.3390/cancers14143525. PMID: 35884586; PMCID: PMC9322748.
- Sahoo DK**, Martinez MN, Dao K, Gabriel V, Zdyrski C, Jergens AE, Atherly T, Iennarella-Servantez CA, Burns LE, Schunk D, Volpe DA, Allenspach K, Mochel JP. Canine Intestinal Organoids as a Novel In Vitro Model of Intestinal Drug Permeability: A Proof-of-Concept Study. *Cells.* 2023 Apr 27;12(9):1269. doi: 10.3390/cells12091269. PMID: 37174669; PMCID: PMC10177590.
- Schneider B**, Balbas-Martinez V, Jergens AE, Troconiz IF, Allenspach K, Mochel JP. Model-Based Reverse Translation Between Veterinary and Human Medicine: The One Health Initiative. *CPT Pharmacometrics Syst Pharmacol.* 2018 Feb;7(2):65-68. doi: 10.1002/psp4.12262. Epub 2017 Nov 27. PMID: 29178333; PMCID: PMC5824107.
- Schneider BK**, Ward J, Sotillo S, Garelli-Paar C, Guillot E, Prikazsky M, Mochel JP. Breakthrough: a first-in-class virtual simulator for dose optimization of ACE inhibitors in translational cardiovascular medicine. *Sci Rep.* 2023 Feb 26;13(1):3300. doi: 10.1038/s41598-023-30453-x. PMID: 36843132; PMCID: PMC9968717.
- Sebbag L**, Harrington DM, Mochel JP. Tear fluid collection in dogs and cats using ophthalmic sponges. *Vet Ophthalmol.* 2018 May;21(3):249-254. doi: 10.1111/vop.12502. Epub 2017 Aug 27. PMID: 28845579.
- Sebbag L**, Showman L, McDowell EM, Perera A, Mochel JP. Impact of Flow Rate, Collection Devices, and Extraction Methods on Tear Concentrations Following Oral Administration of Doxycycline in Dogs and Cats. *J Ocul Pharmacol Ther.* 2018 Jul/Aug;34(6):452-459. doi: 10.1089/jop.2018.0008. Epub 2018 Apr 30. PMID: 29708819; PMCID: PMC6088255.
- Sebbag L**, Allbaugh RA, Weaver A, Seo YJ, Mochel JP. Histamine-Induced Conjunctivitis and Breakdown of Blood-Tear Barrier in Dogs: A Model for Ocular Pharmacology and Therapeutics. *Front Pharmacol.* 2019 Jul 9;10:752. doi: 10.3389/fphar.2019.00752. PMID: 31354477; PMCID: PMC6629934.
- Sebbag L**, Kirner NS, Allbaugh RA, Reis A, Mochel JP. Kinetics of Fluorescein in Tear Film After Eye Drop Instillation in Beagle Dogs: Does Size Really Matter? *Front Vet Sci.* 2019 Dec 19;6:457. doi: 10.3389/fvets.2019.00457. PMID: 31921915; PMCID: PMC6930880.
- Sebbag L**, Mochel JP. An eye on the dog as the scientist's best friend for translational research in ophthalmology: Focus on the ocular surface. *Med Res Rev.* 2020 Nov;40(6):2566-2604. doi: 10.1002/med.21716. Epub 2020 Jul 31. PMID: 32735080.
- Simpson S**, Dunning MD, de Brot S, Grau-Roma L, Mongan NP, Rutland CS. Comparative review of human and canine osteosarcoma: morphology, epidemiology, prognosis, treatment and genetics. *Acta Vet Scand.* 2017 Oct 24;59(1):71. doi: 10.1186/s13028-017-0341-9. PMID: 29065898; PMCID: PMC5655853.
- Terhaar HM**, Allbaugh RA, Mochel JP, Sebbag L. Serum albumin and total protein concentration in the tear film of horses with healthy or diseased eyes. *Vet Ophthalmol.* 2021 Jan;24(1):20-27. doi: 10.1111/vop.12822. Epub 2020 Sep 12. PMID: 32920954.

Toutain PL, Lefebvre HP, King JN. Benazeprilat disposition and effect in dogs revisited with a pharmacokinetic/pharmacodynamic modeling approach. *J Pharmacol Exp Ther*. 2000 Mar;292(3):1087-93. PMID: 10688627.

Uhl LK, Saito A, Iwashita H, Maggs DJ, Mochel JP, Sebbag L. Clinical features of cats with aqueous tear deficiency: a retrospective case series of 10 patients (17 eyes). *J Feline Med Surg*. 2019 Oct;21(10):944-950. doi: 10.1177/1098612X18810867. Epub 2018 Nov 12. PMID: 30417738.

Vaghi C, Rodallec A, Fanciullino R, Ciccolini J, Mochel JP, Mastri M, Poignard C, Ebos JML, Benzekry S. Population modeling of tumor growth curves and the reduced Gompertz model improve prediction of the age of experimental tumors. *PLoS Comput Biol*. 2020 Feb 25;16(2):e1007178. doi: 10.1371/journal.pcbi.1007178. PMID: 32097421; PMCID: PMC7059968.

van de Wal RM, Plokker HW, Lok DJ, Boomsma F, van der Horst FA, van Veldhuisen DJ, van Gilst WH, Voors AA. Determinants of increased angiotensin II levels in severe chronic heart failure patients despite ACE inhibition. *Int J Cardiol*. 2006 Jan 26;106(3):367-72. doi: 10.1016/j.ijcard.2005.02.016. PMID: 16337046.

van der Graaf PH, Benson N. Systems pharmacology: bridging systems biology and pharmacokinetics-pharmacodynamics (PKPD) in drug discovery and development. *Pharm Res*. 2011 Jul;28(7):1460-4. doi: 10.1007/s11095-011-0467-9. Epub 2011 May 11. PMID: 21560018.

van Hasselt JGC, Iyengar R. Systems Pharmacology: Defining the Interactions of Drug Combinations. *Annu Rev Pharmacol Toxicol*. 2019 Jan 6;59:21-40. doi: 10.1146/annurev-pharmtox-010818-021511. Epub 2018 Sep 27. PMID: 30260737.

Vancil JM, Henry CJ, Milner RJ, McCoig AM, Lattimer JC, Villamil JA, McCaw DL, Bryan JN. Use of samarium Sm 153 lexidronam for the treatment of dogs with primary tumors of the skull: 20 cases (1986-2006). *J Am Vet Med Assoc*. 2012 Jun 1;240(11):1310-5. doi: 10.2460/javma.240.11.1310. PMID: 22607597.

Watkins L Jr, Burton JA, Haber E, Cant JR, Smith FW, Barger AC. The renin- angiotensin-aldosterone system in congestive failure in conscious dogs. *J Clin Invest*. 1976 Jun;57(6):1606-17. doi: 10.1172/JCI108431. PMID: 180056; PMCID: PMC436820.

Withrow SJ, Powers BE, Straw RC, Wilkins RM. Comparative aspects of osteosarcoma. Dog versus man. *Clin Orthop Relat Res*. 1991 Sep;(270):159-68. PMID: 1884536.

Xie X, Li X, Song W. Tumor organoid biobank-new platform for medical research. *Sci Rep*. 2023 Feb 1;13(1):1819. doi: 10.1038/s41598-023-29065-2. PMID: 36725963; PMCID: PMC9892604.

Yamamoto JK, Sanou MP, Abbott JR, Coleman JK. Feline immunodeficiency virus model for designing HIV/AIDS vaccines. *Curr HIV Res*. 2010 Jan;8(1):14-25. doi: 10.2174/157016210790416361. PMID: 20210778; PMCID: PMC3721975.

Zhu AZ. Quantitative translational modeling to facilitate preclinical to clinical efficacy & toxicity translation in oncology. *Future Sci OA*. 2018 Apr 23;4(5):FSO306. doi: 10.4155/fsoa-2017-0152. PMID: 29796306; PMCID: PMC5961452.

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.