Disclaimer/Publisher's Note: The statements, opinions, and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions, or products referred to in the content.

Review

Drugs of the Kallikrein-Kinin System: An Overview

François Marceau 1,*

- ¹ Axe Microbiologie-Infectiologie et Immunologie, Research Center, CHU de Québec-Université Laval, Québec, QC, G1V 4G2, Canada
- * Correspondence: francois.marceau@crchudequebec.ulaval.ca (F.M.)

Abstract: The kallikrein-kinin system consists of the two kininogen substrates, present in blood plasma, and of two serine proteases, the plasma and tissue kallikreins. The action of the latter on kininogens produce small peptides, short lived but endowed by powerful pharmacologic actions on blood vessels and other tissues. Many recent and exciting therapeutic developments in the field are briefly summarized. Notably, various novel strategies are being clinically developed to inhibit the formation of bradykinin or block its receptors in the management of hereditary angioedema. The interventions include orally bioavailable drugs, biotechnological proteins, and gene therapy. Several other medical indications are currently investigated. Harnessing controlled kinin formation is also of potential therapeutic interest as shown by the clinical development of recombinant tissue kallikrein for ischemic stroke and renal disease. Biomarkers of a kinin-mediated disorders, frequently implicating edemas, include the consumption of kininogen(s), plasma kallikrein activity, and the detection of circulating kinin metabolites such as fragments BK₁₋₅ and BK₂₋₉. Based on this, some opportunities to clinically apply the underexploited drugs of the kallikrein-kinin system are briefly reviewed. This personal perspective is offered by an observer of, and a participant in drug characterization during the last 4 decades.

Keywords: kallikrein-kinin system; kininogens; bradykinin; B1 receptor; B2 receptor; hereditary angioedema

1. The kallikrein kinin systems: formation and clearance of kinins

Both protective and pathogenic effects are mediated by two largely separate kallikrein-kinin systems (KKS; abbreviations are defined in Table 1) via the formation of small and unstable peptides, the kinins (Fig. 1, schematic representation). Thus, vascular effects (vasodilation, increased microvascular permeability), inflammatory manifestations (edema, pain, increased local blood flow), smooth muscle contraction, and epithelial cell stimulation are potentially initiated by kinins [1]. The nonapeptide bradykinin (BK; H-Arg¹-Pro²-Pro³-Gly⁴-Phe⁵-Ser⁶-Pro⁻-Phe⁶-Argゥ-OH) is the reference kinin sequence found in domain 4 of two circulating proteins, the high molecular weight and low molecular weight kininogens (HK, LK; about 20 and 80% molar proportions, respectively). The hepatic synthesis of both kininogen forms is based on the alternative splicing of a single gene product, *KNG1*.

HK (110 kDa), circulating in a complexed form with prekallikrein and factor XI, is part of the contact system (Fig. 1) along with coagulation factor XII (FXII, Hageman factor). When exposed to negatively charge surfaces, such as the basal membrane of denuded vascular endothelial cells, all these components assemble into a tetramolecular complex with ensuing proteolytic reactions: the mutual activation of FXII and prekallirein into their proteolytically active forms factor XIIa (FXIIa) and plasma kallikrein, respectively, the cleavage of HK releasing BK and the cleavage of factor XI that initiates the intrinsic coagulation pathway [2]. The contact system is tightly controlled by a circulating serpin inhibitor, C1-esterase inhibitor (C1INH) that is also part of the complement cascade. FXIIa and plasma kallikrein are irreversibly inhibited by C1INH [3]. Blood clots are cleared by the fibrinolytic system which is connected to the contact system (Fig. 1): plasmin, the fibrinolytic enzyme, activates FXII into FXIIa to a certain extent, indirectly promoting BK production via

secondarily activated plasma kallikrein. C1INH is a secondary inhibitor of plasmin [3]. Whether HK is directly cleaved by additional proteases has been suggested, but not well established in whole blood where endogenous inhibitors are present: plasmin and the complement associated protease MASP-1 may directly release BK from HK [4,5]. There is no evidence of BK release when platelets or neutrophils are activated in human whole blood [6], casting a doubt about previously suggested activation pathways demonstrated using purified components of the contact system.

Table 1. List of abbreviations.

Abbreviation	Standing for	Corresponding gene
ACE	angiotensin-I converting enzyme	ACE
Arg-CP	arginine carboxypeptidase	
B1R	bradykinin B1 receptor	BDKRB1
B2R	bradykinin B2 receptor	BDKRB2
BK	bradykinin	
C1INH	C1-esterase inhibitor	SERPING1
FXII	coagulation factor XII	F12
FXIIa	activated factor XII	
HAE	hereditary angioedema	
HAE-C1INH	HAE caused by C1INH	
HAE-CHINH	haplodeficiency	
HK	high molecular weight kininogen	KNG1
KKS	kallikrein-kinin system	
KLK-1	tissue kallikrein	KLK1
LK	low molecular weight kininogen	KNG1
Lys-BK	kallidin	
mAb	therapeutic monoclonal antibody	
MASP-1	mannan-binding lectin associated	MASP1
	serine protease 1	WASFI
NPA	non-peptide antagonist	
tPA	tissue plasminogen activator	PLAT

Tissue kallikrein (KLK-1; kallidinogenase) is a member of a family of 15 secreted proteases encoded on human chromosome locus 19q13.4 [7]. These serine proteases assume different, often uncertain physiological functions. Only KLK-1 is a relevant kininogenase in this family. This was verified with two KLKs normally found in the prostate: they release no kinins from purified HK (KLK-3), or very little (KLK-2 is 1000-fold less active than KLK-1 in this respect) [8]. KLK-1 releases the biologically active decapeptide Lys-BK (= kallidin) from both forms of kininogen, but mostly from the more abundant LK. KLK-1 is widely expressed and abundant in the kidney, pancreas, salivary glands, lungs, blood vessels and other tissues; its secretion and activation, via the removal of a N-terminal sequence, are not well understood. KLK-1 is regulated by its own irreversible inhibitor, kallistatin (SERPINA4 gene product). The previously claimed direct agonist effect of KLK-1 on human BK B2 receptor (B2R) has been disproved using the pure recombinant enzyme in its active form [9].

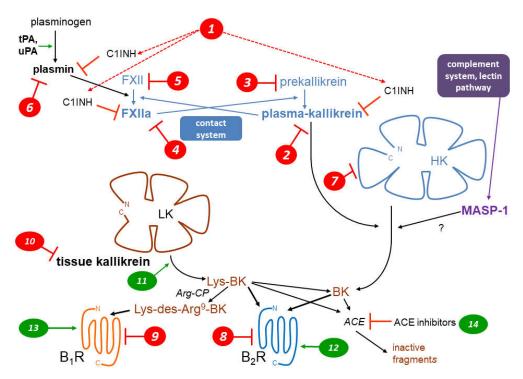


Figure 1. Schematic representation of the KKS, featuring the 2 validated pathways of kinin generation, that of plasma kallikrein (part of the contact system) releasing bradykinin (BK) from high molecular weight kininogen (HK) and that mediated by secreted tissue kallikrein (KLK-1), generating Lys-BK mainly from low molecular weight kininogen (LK). A possible alternate BK generating pathway is indicated (purple). Two G protein coupled receptors (B1R, B2R) mediate the cellular effects of kinins. Two types of metallopeptidases that hydrolyze kinins are indicated (Arg-CP, ACE). Numerical markers indicate the mode of action of inhibitory (red) or stimulatory drugs (green) of the KKS and are referred to in Tables 2-4 and main text. See Table 1 for abbreviations. Modified from [10, 11].

Kinins are inherently unstable, with a half-life well under 1 min [10], and metabolized by several metallopeptidases. A careful in vivo study showed that angiotensin converting enzyme (ACE, kininase II) is the dominant BK-inactivation pathway in rats, followed by aminopeptidase P [12]. Both these peptidases inactivate BK, initially producing the fragments BK₁₋₇ and BK₂₋₉, respectively. The fragment BK₁₋₅ is the relatively stable product of a second cycle of BK₁₋₇ cleavage by ACE. Arginine carboxypeptidases (Arg-CPs) (plasma carboxypeptidase N, cell membrane carboxypeptidase M; producing BK₁₋₈ = des-Arg⁹-BK from BK) represent only a minor metabolic pathway in vivo [13] but connects the KKS with the pharmacological profile of the kinin B1 receptor (B1R; see below). Biomarkers of kinin-mediated disorders include the consumption of kininogen(s) and the detection of circulating kinin metabolites such as fragments BK₁₋₅ and BK₂₋₉, and the detection of plasma kallikrein activity, for instance using the synthetic substrate based on the C-terminal BK sequence HD-Pro-Phe-Arg-*pNA*. These assays are technically challenging, but one or more of them have been applied to hereditary angioedema (HAE), either during attacks or in remission [14, 15], and to other edematous conditions such as ascites secondary to liver cirrhosis [16] and chronic urticaria [17, 18] and to animal models of sepsis and sickle cell disease [19,20].

2. Kinin receptors

Before the era of molecular biology, the number and identity of kinin receptor subtypes in each mammalian species was uncertain. Historically, the first proposed kinin receptor subtype, the B1R, was discovered as the one mediating contraction in the isolated rabbit aorta and it possessed a typical order of potency for agonists and a first class of peptide antagonists [21]. Native kinins (BK and Lys-BK) from which the Arg⁹ residue has been removed by Arg-CPs (des-Arg⁹-BK, Lys-des-Arg⁹-BK, respectively) are the optimal agonists of the B1R, even if this kinin metabolic pathway is not

prominent. Only Lys-des-Arg⁹-BK, also called des-Arg¹⁰-kallidin, has a subnanomolar affinity for the human (and rabbit) B1R [1]; this optimal agonist is presumably generated from Lys-BK (kallidin), itself derived from the cleavage of kininogens by tissue kallikrein (Fig. 1). These reactions occur outside of the contact system. Early peptide antagonists, such as [Leu⁸]des-Arg⁹-BK, consolidated the pharmacological identity of the B1R; the other pharmacologic systems, directly responsive to BK and Lys-BK but insensitive to the des-Arg⁹ metabolites, were said to possess the B2R. The first B2R antagonists were discovered in the early 1980's; they featured a constrained peptide backbone and were more or less protected from peptidases. Icatibant (Hoe 140; D-Arg-[Hyp³, Thi⁵, D-Tic⁷, Oic⁸]BK) is the success story among these early drugs [1,22] (Table 2). Selected modern, nonpeptide antagonists (NPAs) of both kinin receptor subtypes are presented in Fig. 2. It is very typical that BK receptor antagonists exhibit species-dependent alterations of affinity and competitive behavior for their pharmacological targets [1]; thus, clinically developed antagonists have gone through a structural optimization process for the human forms of the B1R or B2R [1,23].

The receptor classification was confirmed by the discovery of a kinin receptor locus: in human chromosome region 14q32, genes encoding the G protein coupled receptors B2R and B1R, respectively termed BDKRB2 and BDKRB1, are in found tandem [24]; a similar organization is found in the genome of other mammals. While the expression of both genes is regulated, the B2R generally accounts for the in vivo effects of kinins in healthy laboratory animals: this receptor subtype is constitutively expressed in many cell types, including vascular endothelial cells, smooth muscle cells, some epithelia, sensory neurons, and other cell types [1]. The B1R, initially discovered in rabbit isolated blood vessels maintained in vitro for several hours, is not generally detectable in healthy animals. The paradox was resolved when the B1R was found to be expressed following tissue trauma, inflammation (such as the injection of bacterial lipopolysaccharide to animals) under the control of inflammatory cytokines (e.g., interleukin-1, tumor necrosis factor- α , interferon- γ) and signaling (e.g., mitogen activated protein kinases, NF-κB, Jak/Stat) [24-27]. While B1R and B2R are structurally related, only the latter is phosphorylated, internalized, and recycled following agonist stimulation [1,24]. Thus, the B1R is potentially important in sustained inflammatory states. It does not follow that B1R should be systematically blocked in tissue injury situations: for instance, the development of an adaptative collateral circulation is mediated by this receptor subtype following an arterial occlusion in a rodent model [28].

Both kinin receptor subtypes are coupled mainly to protein G_q and calcium signaling, leading typically to smooth muscle contraction, but also prominently to vascular endothelial cell stimulation, including calcium-dependent prostanoid and nitric oxide release and fluid leakage secondary to gap formation between endothelial cells [1,24].

Table 2. Inhibiting the kallikrein-kinin system for treating or preventing attacks of hereditary angioedema.

Type of agent mode of action marker in Fig. 1	Drug or intervention	Development status	Ref.
Parenteral replacement of C1INH 1	various C1INH concentrates natural or recombinant	approved, widely, used	29
Gene therapy to increase the	BMN 311 HAE	clinical trials	30
endogenous synthesis of C1INH 1	OTL-105 HAE	preclinical	31
Kunitz-domain based peptide inhibitor of plasma kallikrein 2	ecallantide	approved	32
Creall real and inhibitors of	berotralstat (BCX7353)	approved	33
Small molecule inhibitors of plasma kallikrein 2	sebetralstat (KVD-900)	clinical trials	34
piasina kamkrein 2	ATN-249, ATN-111	clinical trials	35
Anti-plasma kallikrein mAb 2	lanadelumab	approved	36
Anti-piasma kamkiem mad z	STAR-0215	clinical trials	37
Transfer of a gene encoding an anti-plasma kallikrein mAb 2	RegenxBio undisclosed	preclinical	38
Antisense suppressor of hepatic plasma prekallikrein production 3	donidalorsen (PKK-L Rx)	clinical trials	39
Gene therapy to disrupt hepatic plasma prekallikrein production 3	NTLA-2002	clinical trials	40
Small molecule inhibitor of factor XIIa 4	KV998086	preclinical	41
Anti-factor XII mAb 4	Garadacimab (CSL312)	clinical trials	42
Small interfering RNA targeting	ALN-F12	preclinical, halted?	43
factor XII mRNA 5	ARC-F12	preclinical, halted?	44
Plasmin/tPA inhibitor 6	tranexamic acid	approved, 2 nd line prophylactic agent	45
	peptide icatibant	approved	46
Bradykinin B2R antagonists 8	NPA deucrictibant (PHA- 022121, PHA-121)		23, 47

3. Pharmacological inhibition of the KKS: hereditary angioedema (HAE)

Fig. 1 is a schematic representation of the kinin-generating pathways and their receptors; numerical markers indicate the mode of action of the numerous drugs of the KKS. Earlier achievements, such as the early peptide receptor antagonists, are reviewed elsewhere [48]. The present emphasis is on drugs that are currently in use, have reached clinical development (successfully or not), or are in preclinical development.

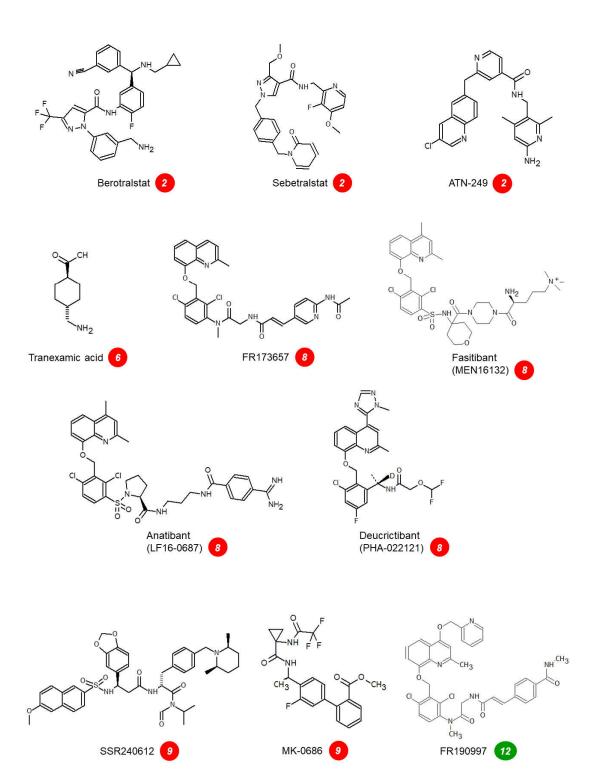


Figure 2. Structure of the small molecule drugs cited in tables 2-4 (except for KV998086 and BI1026706, currently undisclosed). The *numerical markers* indicate the mode of action as in Fig. 1. The structure of the antagonists of kinin receptors is optimized for the human forms of these receptors. Note the structural similarities of the B2R antagonists (marker 8); only deucrictibant is developed as an orally administered drug in this class.

The therapeutic showcase of the KKS is presently hereditary angioedema (HAE), a rare disease most often caused by the haplodeficiency of C1INH (numerous mutations transmitted in an autosomal dominant manner are known in the corresponding gene *SERPING1*) [49]. HAE is characterized by recurrent episodes (attacks) of swelling due to fluid extravasation; limbs, the

orofacial and genital areas, and the intestine can be affected. Attacks may be life-threatening (suffocation), painful and incapacitating. The physiopathology of HAE and its management have been recently reviewed [49-51]. While C1INH inhibits multiple proteases in the contact, fibrinolytic and complement systems, bradykinin is believed to be the ultimate mediator of HAE-C1INH attacks. Variants of six additional causal genes very rarely cause HAE independently from *SERPING1*: *PLG*, *F12*, *KNG1*, *HS3ST6*, *ANGPT1*, *MYOF*. The 3 first ones belong to the connected contact or fibrinolytic systems, also plausibly leading to the unregulated production of kinins, while the physiopathology of HAE caused by the exceedingly rare variants of the last 3 genes is essentially unknown. There are certainly other causal genes not currently identified in HAE patients with normal C1INH levels [52]. Age, sex, and hormonal status modulate the frequency and intensity of HAE attacks, the facilitatory effect of estrogens being particularly well documented.

Let us limit ourselves here to a few remarks concerning the therapy of HAE, an admittedly crowded market for such a rare disease. Drugs and biotechnological treatments are used or proposed for attack prevention (prophylaxis), to abort attacks ("on demand" treatments), or both. While numerous HAE therapies that affect the KKS are approved or under development (Table 2), the number of validated molecular target is limited. The parenteral administration of C1INH, or gene therapy to increase the hepatic biosynthesis of normal C1INH, is physiologically sound for HAE-C1INH (and proven by multiple clinical trials for C1INH concentrates). Of note, this intervention also works in many HAE patients with normal C1INH levels. The heart of the contact system is also targeted in HAE (Fig. 1, Table 2): plasma kallikrein or its proenzyme prekallikrein, FXIIa or its proenzyme FXII can be suppressed or pharmacologically inhibited by several pharmacological or biotechnological interventions. The proof of concept for a further level of intervention on the contact system has been recently reported in a preclinical study: the mAb 3E8 targets domain 6 of HK, thus inhibiting the assembly of the trimolecular complex HK-prekallikrein-factor XI (mode of action 7 in Fig. 1). In transgenic mice that express human HK, mAb 3E8 inhibits dextran sulfate-induced BK formation and FXII activation [53].

On the effector side, the BK B2R antagonists inhibit the vascular manifestations of HAE (Table 2, Fig. 1). The injectable and rapidly cleared peptide antagonist icatibant is widely used to abort HAE attacks. The nonpeptide B2R antagonist deucrictibant [23] (Fig. 2) is orally bioavailable, more potent, and longer lived than icatibant in vivo; it is currently developed for on demand treatment of HAE attacks (a potentially convenient substitute to subcutaneous icatibant, Table 2). Chronically administered deucrictibant will also be tested for prophylaxis. Both icatibant and deucrictibant are competitive and reversible antagonists at the human B2R [23]; the affinity and/or competitive behavior of these drugs varies for B2Rs from other mammalian species. Kinin B1R antagonists have not been tested for HAE, and the rationale for them may be weak, considering that the human form of the B1R is not effectively coupled to the contact system (see above).

There is clear evidence of fibrinolytic system activation during HAE attacks: the D-dimers that are fibrin degradation products are elevated in blood during attacks, but without thrombotic risks [54]. Whether fibrinolysis initiates the attacks is a complex question that has been reviewed [55]. Let us add two remarks to the debate. Indicating a particular vulnerability, adding recombinant tPA (10 nM) to human whole blood or citrated plasma from HAE-C1INH patients selectively increased the early BK production in vitro (measured by enzyme immunoassay and corroborated with bioassay) [6,56]. In addition, in a survey of patients concerning the circumstances preceding HAE attacks, most identified triggering factors could be related to limited coagulation followed by fibrinolysis: physical exertion, mechanical trauma, infection, menstruation, dental procedures, etc. [57]. Mental stress is one of the top patient-identified triggering factor of HAE attacks [57]. In well controlled clinical psychology studies in healthy volunteers, mental stress activates tPA secretion and the turnover of fibrin (detected by D-dimer formation) [58], further supporting the case for fibrinolysis-induced attacks.

Oral tranexamic acid, an inhibitor of plasmin and tissue plasminogen activator, is approved as a second line prophylactic treatment of HAE. However, it lacks validation through extensive clinical trials for this indication, does not normalize the patient lives as well as more recent therapies and does not work well in some patients [45].

Table 3. Selected additional indications for the inhibition of the KKS.

	Drug		
Indication	mode of action marker in Fig.	Status	Ref.
ACE-inhibitor induced acquired angioedema	C1INH concentrate 1	ineffective in a small clinical trial	59
Idiopathic pulmonary fibrosis, interstitial lung disease	mAb garadacimab (CSL312) 4	clinical trials	60
Thrombosis prevention	ALN-F12 5	preclinical	61
COVID-19 pneumonia	peptide B2R antagonist icatibant <mark>8</mark>	ineffective in a clinical trial	62
Cerebral edema/head trauma	B2R NPA anatibant (LF16- 0687) <mark>8</mark>	ineffective in an interrupted clinical trial	63
Osteoarthritis pain	B2R NPA fasitibant (MEN16132) <mark>\$</mark>	largely ineffective in a clinical trial	64
Diabetic macula edema	B1R NPA BI1026706 8	ineffective in a clinical trial	65
Glioma	intracerebroventricular icatibant <mark>8</mark>	preclinical	66
ACE-inhibitor induced acquired angioedema	icatibant 8	ineffective in a clinical trial	67
Diarrheal states induced by dextran sulfate	oral icatibant, oral FR173657 8	preclinical	68, 69
Pancreatitis	icatibant, FR173657 8	preclinical	70, 71
Circulatory complications of burns	icatibant 8	preclinical	72, 73
Inflammatory edema of various causes	FR173657, icatibant 8	preclinical	74, 75
Breast cancer	B1R NPA SSR240612, etc. 8	preclinical, in vitro	76
Inflammatory pain	B1R NPA MK-0686, SS240612	ineffective in clinical trials	77
Inflammatory bowel disease	B1R NPA SSR240612 9	preclinical	78
Airway disease	mAb DX-2300 10	preclinical	79
Aortic aneurysm expansion	mAb DX-2300, etc. 10	preclinical	80

4. Other application of KKS inhibitors

Table 3 presents a selection of therapeutic ongoing or terminated therapeutic projects that exploited inhibitors of the KKS. Some comments are offered here concerning specific indications. Pain is one of the cardinal signs of inflammation; despite good preclinical evidence, the clinical development of sophisticated and orally bioavailable B1R antagonists has failed due to their lack of efficacy in phase 2 trials (Table 3, Fig. 2) [77]. Fasitibant, a B2R antagonist injected in an intraarticular manner, as also failed to relieve pain associated with knee osteoarthritis (Fig. 2) [64]. These trials were a major disappointment, but preclinical research continues for a specific form of pain associated with breast cancer and its chemotherapy; the blockade of either or both kinin receptor subtypes exert favorable effects on the associated mechanical and cold allodynia [81]. The badly underdeveloped clinical research concerning the B1R as a druggable target could benefit from the repurposing of exquisitely specific drugs that have passed successfully clinical phase 1 development (Fig. 2), for instance for the prevention of COVID-19 complications [82].

Gliomas, including the highly malignant glioblastoma multiforme, overexpress the B2R in a manner correlated to clinical aggressivity; it is coupled to several pro-tumoral signaling systems [83].

Inhibiting B2R may be of therapeutic value in this condition, based on a preclinical study that also shows the difficulty of reaching the tumor through the blood-brain barrier with the presently available B2R antagonists (intracerebroventricular icatibant was used in animals) [66]. Interestingly, the B1R is also often present in glioma, but rather supports tumor growth, based on pharmacological blockade or results obtained *BDKRB1* knock out mice [66].

ACE inhibitors (enalapril, ramipril, many others) constitute an important class of antihypertensive drugs with benefits on several organs. ACE is a carboxydipeptidase that activates the formation of the pressor hormone angiotensin II from its precursor angiotensin I, but also mediates the most important inactivation pathway for kinins such as BK and Lys-des-Arg9-BK. A certain fraction of the beneficial effects of ACE inhibitors in human patients seems to derive from the potentiation of the vasodilator effect of kinins (mode of action 14 in Fig. 1) [84]. Many drugs are associated with acquired angioedema, an unpredictable and potentially life-threatening side effect. The angioedema induced by ACE inhibitors is hypothesized by several investigators to be BKdependent [85]; for instance, increased blood concentration of the BK metabolite BK₁₋₅ was measured during such episodes [86]. However, such attacks did not respond to administration of icatibant [67] or of a C1INH concentrate [59] in clinical settings that are comparable to HAE attacks for which these interventions are therapeutic. Possible explanations include the lack of involvement of the contact system in kinin generation, a prominent role for the kinin B1R or, simply, the lack of developing fluid extravasation when the therapies were initiated relatively late during such unexpected episodes. Of note, thrombolytic therapy for ischemic stroke based on a plasminogen activator such as tPA is sometimes associated with brain edema, angioedema, and hypotension: these are serious side effects presumably mediated by excessive BK formation, due to the connection between fibrinolysis and the contact system [4]. A beneficial effect of icatibant has been reported in such episodes [87,88].

An efficient monoclonal antibody that blocks the enzymatic action of tissue kallikrein, DX-2300, has been developed and shown of potential interest in preclinical research (Table 3). Whether this physiological regulator of circulation and renal function could be inhibited in vivo without major side effects remain to be seen; however, DX-2300 is a powerful tool to selectively block the quantitatively important KLK-1-induced kinin formation that is independent from the contact system [6].

5. Therapeutic value of KKS stimulation?

Kinin generation via the contact system can be chronically blocked without apparent harm, as implied above when discussing the therapy of hereditary angioedema. The alternate kallikrein-kinin system, mediated by tissue kallikrein, appears to be adaptative in various ways. Tissue kallikrein promotes reparative neovascularization following experimental ischemia and protects the heart in animal models of pathologies [89,90]. This enzyme, produced in a regulated manner in the kidney, is released in urine and protects from sodium overload and salt-sensitive hypertension [91]. Tissue kallikrein also participates to flow-dependent vasodilation, a local circulatory adaptative mechanism [92]. So, why not consider the parenteral administration of tissue kallikrein in therapeutics? In China, active KLK-1 purified from human urine has reached clinical use for acute ischemic stroke (Table 4). When added to standard thrombolytic therapy, parenteral tissue kallikrein improved the neurological recovery in a significant manner [93]. A pharmaceutically refined recombinant tissue kallikrein, DM199, is being clinically developed for cerebrovascular and renal indications [94,95].

Is intravenous infusion of kinins feasible in intensive care situations where the stimulation of vascular B2Rs has been proposed to have therapeutic value, such as myocardial infarction and ischemic stroke? A preclinical project addressed this possibility by characterizing peptides that are both prodrugs and "soft drugs" [96-98] (Table 4). For instance, BK-Arg (BK sequence prolonged with an Arg residue) has virtually no affinity for the B2R, but releases BK in vivo following the action of Arg-CPs in the circulation, with typical vasodilator responses [97]; thus BK-Arg is a prodrug. This peptide is also a soft drug, because its active form, BK, is rapidly cleared with minimal extra-vascular effects. Other activating peptidases and agonist designs have been explored.

Type of agent			
mode of action marker in Fig. 1	Drug	status	Ref.
Activated tissue kallikrein	purified from human	approved in China,	93
injection for cerebrovascular or	urine: Kailikang	stroke	93
renal indications 11	recombinant: DM199	clinical trials	94,95
B2R agonist to open the blood-	labradimil, others	clinical trials of labradimil failed	99, 100
brain barrier: adjuvant to brain			
tumor chemotherapy 12			
Combined B1R and B2R	co-administered peptide agonists or kinin heterodimer	preclinical	101
agonists to open the blood-brain			
barrier: adjuvant to brain tumor			
chemotherapy 12, 13	neteroanner		
Peptide pro-drugs that release	bradykinin-Arg,		
bradykinin via the action of	D-Arg-bradykinin-Arg-	preclinical	96-98
peptidases 12	Arg, others		
B2R NPA for breast cancer 12	FR-190997 and analogs	preclinical, in vitro	102
Attenuation of Alzheimer's	[Hyp³,Thi⁵,NChg²,Thi8]-BK		103
disease development, B2R		preclinical, animal	
agonist 12		model	
	enalapril, ramipril, many others	a fraction of	
ACE inhibition 14		therapeutic effects	84
		mediated by kinins	

Table 4. Therapeutic stimulation of kinin formation or of kinin receptor signaling.

6. Conclusions

In a certain sense, the medicinal chemistry related to the KKS has reached maturity, with the development of modern drugs, injectable biotechnological proteins, and advanced gene therapy projects (Tables 2-4). In addition to C1INH replacement therapy, HAE has been the focus of intense drug development efforts based on a limited number of validated targets (plasma kallikrein, FXIIa and their respective zymogens, the B2R). A recent effort to transition to oral therapies is also noted. Concerning other indications, efficacy in various animal models has led to disappointing clinical outcomes, as in many other therapeutic areas. The existence of orally bioavailable drugs that have at least passed clinical phase 1 development (B1R and B2R antagonists, plasma kallikrein inhibitors; Fig. 2) could facilitate their repurposing for additional therapeutic indications. For instance, there is evidence for both neuroprotective and detrimental effects of kinins, mediated by either B1R or B2R, in various neurological disorders [103,104]. Let us hope that the underexploited pharmacopeia overviewed in the present paper finds novel clinical applications.

Funding: This research received no external funding.

Conflicts of Interest: The author served as a consultant for Pharvaris B.V. and received research funds from Shire/Takeda, DiaMedica and Pharvaris B.V. The funders had no role in the writing of the manuscript, or in the decision to publish the results.

References

- Leeb-Lundberg, L.M.; Marceau, F.; Müller-Esterl, W.; Pettibone, D.J.; Zuraw, B.L. International union of pharmacology. XLV. Classification of the kinin receptor family: from molecular mechanisms to pathophysiological consequences. *Pharmacol, Rev.* 2005, 57, 27-77. doi: 10.1124/pr.57.1.2
- 2. Kaplan, A.P.; Joseph, K.; Ghebrehiwet, B. The complex role of kininogens in hereditary angioedema. *Front. Allergy* **2022**, *3*, 952753. doi: 10.3389/falgy.2022.952753

- 3. Kaplan, A.P. Enzymatic pathways in the pathogenesis of hereditary angioedema: the role of C1 inhibitor therapy. *J. Allergy Clin. Immunol.* **2010**, *126*, 918-925. doi: 10.1016/j.jaci.2010.08.012
- Gauberti, M.; Potzeha, F.; Vivien, D.; Martinez de Lizarrondo, S. Impact of Bradykinin Generation During Thrombolysis in Ischemic Stroke. *Front. Med. (Lausanne)* 2018, 5, 195. doi: 10.3389/fmed.2018.00195
- Dobó, J.; Major, B.; Kékesi, K.A.; Szabó, I.; Megyeri, M.; Hajela, K.; Juhász, G.; Závodszky, P.; Gál, P. Cleavage of kininogen and subsequent bradykinin release by the complement component: mannose-binding lectin-associated serine protease (MASP)-1. PLoS One 2011, 6, e20036. doi: 10.1371/journal.pone.0020036
- Charest-Morin, X.; Hébert, J.; Rivard, G.É.; Bonnefoy, A.; Wagner, E.; Marceau, F. Comparing Pathways of Bradykinin Formation in Whole Blood From Healthy Volunteers and Patients With Hereditary Angioedema Due to C1 Inhibitor Deficiency. Front. Immunol. 2018, 9, 2183. doi: 10.3389/fimmu.2018.02183
- 7. Yousef, G.M.; Chang, A.; Scorilas, A.; Diamandis, E.P. Genomic organization of the human kallikrein gene family on chromosome 19q13.3-q13.4. *Biochem. Biophys. Res. Commun.* **2000**, 276, 125-133. doi: 10.1006/bbrc.2000.3448
- 8. Deperthes, D.; Marceau, F.; Frenette, G.; Lazure, C.; Tremblay, R.R.; Dubé, J.Y. Human kallikrein hK2 has low kininogenase activity while prostate-specific antigen (hK3) has none. *Biochim. Biophys. Acta* **1997**, *1343*, 102-106. doi: 10.1016/s0167-4838(97)00135-0
- 9. Charest-Morin, X.; Raghavan, A.; Charles, M.L.; Kolodka, T.; Bouthillier, J.; Jean, M.; Robbins, M.S.; Marceau, F. Pharmacological effects of recombinant human tissue kallikrein on bradykinin B2 receptors. *Pharmacol. Res. Perspect.* **2015**, *3*, e00119. doi: 10.1002/prp2.119
- 10. Marceau; F.; Rivard, G.E.; Gauthier, J.M.; Binkley, K.E.; Bonnefoy, A.; Boccon-Gibod, I.; Bouillet, L.; Picard, M.; Levesque, G.; Elfassy, H.L.; et al. Measurement of Bradykinin Formation and Degradation in Blood Plasma: Relevance for Acquired Angioedema Associated With Angiotensin Converting Enzyme Inhibition and for Hereditary Angioedema Due to Factor XII or Plasminogen Gene Variants. Front Med (Lausanne) 2020, 7, 358. doi: 10.3389/fmed.2020.00358
- 11. Marceau, F.; Bachelard, H.; Charest-Morin, X.; Hébert, J.; Rivard, G.E. In vitro modeling of bradykinin-mediated angioedema states. *Pharmaceuticals (Basel)* **2020**, *13*, 201. doi: 10.3390/ph13090201
- 12. Fryer, R.M.; Segreti, J.; Banfor, P.N.; Widomski, D.L.; Backes, B.J.; Lin, C.W.; Ballaron, S.J.; Cox, B.F.; Trevillyan, J.M.; Reinhart, G.A.; et al. Effect of bradykinin metabolism inhibitors on evoked hypotension in rats: rank efficacy of enzymes associated with bradykinin-mediated angioedema. *Br. J. Pharmacol.* **2008**, *153*, 947-955. doi: 10.1038/sj.bjp.0707641
- 13. Ishida, H.; Scicli, A.G.; Carretero, O.A. Contributions of various rat plasma peptidases to kinin hydrolysis. *J. Pharmacol. Exp. Ther.*, **1989**, 251, 817–820.
- Defendi F, Charignon D, Ghannam A, Baroso R, Csopaki F, Allegret-Cadet M, Ponard D, Favier B, Cichon S, Nicolie B, et al. National Reference Centre for Angioedema CREAK. Enzymatic assays for the diagnosis of bradykinin-dependent angioedema. *PLoS One* 2013, 8, e70140. doi: 10.1371/journal.pone.0070140
- 15. Marceau, F.; Rivard, G.E.; Hébert, J.; Gauthier, J.; Bachelard, H.; Gangnus, T.; Burckhardt, B.B. Picomolar Sensitivity Analysis of Multiple Bradykinin-Related Peptides in the Blood Plasma of Patients With Hereditary Angioedema in Remission: A Pilot Study. *Front. Allergy* **2022**, *3*, 837463. doi: 10.3389/falgy.2022.837463

- Cugno, M.; Salerno, F.; Nussberger, J.; Bottasso, B.; Lorenzano, E.; Agostoni, A. Bradykinin in the ascitic fluid of patients with liver cirrhosis. Clin Sci (Lond) 2001, 101, 651-657.
- 17. Hofman, Z.L.M.; van den Elzen, M.T.; Kuijpers, J.; de Maat, S.; Hack, C.E.; Knulst, A.C.; Röckmann, H.; Maas, C. Evidence for bradykinin release in chronic spontaneous urticaria. *Clin. Exp. Allergy*. **2020**, *50*, 343-351. doi: 10.1111/cea.13558
- 18. Mostmans, Y.; De Smedt, K.; Richert, B.; Elieh Ali Komi, D.; Maurer, M.; Michel, O. Markers for the involvement of endothelial cells and the coagulation system in chronic urticaria: A systematic review. *Allergy* **2021**, *76*, 2998-3016. doi: 10.1111/all.14828
- Barratt-Due, A.; Johansen, H.T.; Sokolov, A.; Thorgersen, E.B.; Hellerud, B.C.; Reubsaet, J.L.; Seip, K.F.; Tønnessen, T.I.; Lindstad, J.K.;, Pharo, A.; et al. The role of bradykinin and the effect of the bradykinin receptor antagonist icatibant in porcine sepsis. *Shock* 2011, 36, 517-523. doi: 10.1097/SHK.0b013e3182336a34
- Sparkenbaugh, E.M.; Kasztan, M.; Henderson, M.W.; Ellsworth, P.; Davis, P.R.; Wilson, K.J.; Reeves, B.; Key, N.S.; Strickland, S.; McCrae, K.; et al. High molecular weight kininogen contributes to early mortality and kidney dysfunction in a mouse model of sickle cell disease. *J. Thromb. Haemost.* 2020, 18, 2329-2340. doi: 10.1111/jth.14972
- 21. Regoli, D.; Barabé, J. Pharmacology of bradykinin and related kinins. Pharmacol. Rev. 1980, 32, 1-46.
- 22. Hock, F.J.; Wirth, K.; Albus, U.; Linz, W.; Gerhards, H.J.; Wiemer, G.; Henke, S.; Breipohl, G.; König, W.; Knolle, J.; et al. Hoe 140 a new potent and long acting bradykinin-antagonist: in vitro studies. *Br. J. Pharmacol.* **1991**, 102, 769-773. doi: 10.1111/j.1476-5381.1991.tb12248.x
- 23. Lesage, A.; Marceau, F.; Gibson, C.; Loenders, B.; Katzer, W.; Ambrosi, H.D.; Saupe, J.; Faussner, A.; Pardali, E.; Knolle, J. In vitro pharmacological profile of PHA-022121, a small molecule bradykinin B2 receptor antagonist in clinical development. *Int. Immunopharmacol.* 2022, 105, 108523. doi: 10.1016/j.intimp.2022.108523
- 24. Marceau. F.; Bachelard. H.; Bouthillier, J.; Fortin, J.P.; Morissette, G.; Bawolak, M.T.; Charest-Morin, X.; Gera, L. Bradykinin receptors: Agonists, antagonists, expression, signaling, and adaptation to sustained stimulation. *Int. Immunopharmacol.* **2020**, *82*, 106305. doi: 10.1016/j.intimp.2020.106305
- 25. Larrivée, J.-F.; Bachvarov, D.R.; Houle, F.; Landry, J.; Huot, J.; Marceau, F. Role of the mitogenactivated protein kinases in the expression of the kinin B1 receptors induced by tissue injury. *J. Immunol.* **1998**, *160*, 1419-1426.
- 26. Moreau, M.E.; Bawolak, M.T.; Morissette, G.; Adam, A.; Marceau, F. Role of nuclear factor-kappaB and protein kinase C signaling in the expression of the kinin B1 receptor in human vascular smooth muscle cells. *Mol. Pharmacol.* **2007**, *71*, 949-956. doi: 10.1124/mol.106.030684
- Koumbadinga, G.A.; Désormeaux, A.; Adam, A.; Marceau, F. Effect of interferon-γ on inflammatory cytokine-induced bradykinin B1 receptor expression in human vascular cells. *Eur. J. Pharmacol.* 2010, 647, 117-125. doi: 10.1016/j.ejphar.2010.08.008
- 28. Emanueli, C.; Bonaria Salis, M.; Stacca, T.; Pintus, G.; Kirchmair, R.; Isner, J.M.; Pinna, A.; Gaspa, L.; Regoli, D.; Cayla, C.; et al. Targeting kinin B1 receptor for therapeutic neovascularization. *Circulation* **2002**, *105*, 360-366. doi: 10.1161/hc0302.102142
- 29. Longhurst, H.; Farkas, H. Biological therapy in hereditary angioedema: transformation of a rare disease. *Expert Opin. Biol. Ther.* **2020**, *20*, 493-501. doi: 10.1080/14712598.2020.1724280
- 30. Biomarin. https://www.biomarin.com/our-treatments/pipeline/bmn-331-for-hae/ [last accessed 1 May 2023]
- 31. Pharming Group N.V. https://www.pharming.com/pipeline [last accessed 1 May 2023]

- 32. Duffey H, Firszt R. Management of acute attacks of hereditary angioedema: role of ecallantide. *J. Blood Med.* **2015**, *6*, 115-123. doi: 10.2147/JBM.S66825
- 33. Ahuja, M.; Dorr, A.; Bode, E.; Boulton, A.P.R.; Buckland, M.; Chee, S.; Dalley, C.; Denman, S.; Ekbote, A.; Elkhalifa, S.; et al. Berotralstat for the prophylaxis of hereditary angioedema-Real-world evidence data from the United Kingdom. *Allergy* **2023**, *in press*. doi: 10.1111/all.15641
- 34. Aygören-Pürsün, E.; Zanichelli, A.; Cohn, D.M.; Cancian, M.; Hakl, R.; Kinaciyan, T.; Magerl, M.; Martinez-Saguer, I.; Stobiecki, M.; Farkas, H., et al. An investigational oral plasma kallikrein inhibitor for on-demand treatment of hereditary angioedema: a two-part, randomised, double-blind, placebo-controlled, crossover phase 2 trial. *Lancet* 2023, 401, 458-469. doi: 10.1016/S0140-6736(22)02406-0
- 35. Kalfus, I.; Offman, E.; McDonald, A. Pharmacokinetics, safety, and potency of ATN-249, a novel oral plasma kallikrein inhibitor for hereditary angioedema. *Allergy Asthma Clin. Immunol.* **2019**, *15*(*suppl* 4), 45. [abstract]
- 36. Riedl, M.A.; Maurer, M.; Bernstein, J.A.; Banerji, A.; Longhurst, H.J.; Li, H.H.; Lu, P.; Hao, J.; Juethner, S.; Lumry, W.R.; et al. Lanadelumab demonstrates rapid and sustained prevention of hereditary angioedema attacks. *Allergy* **2020**, *75*, 2879-2887. doi: 10.1111/all.14416
- 37. Astria Therapeutics. https://astriatx.com/our-science/scientific-presentations-and-publications/ [last accessed 26 April 2023]
- 38. REGENXBIO Inc. https://ir.regenxbio.com/news-releases/news-release-details/regenxbio-reports-continued-progress-across-programs-year-end-0/ [last accessed 1 May 2023]
- 39. Ferrone, J.D.; Bhattacharjee, G.; Revenko, A.S.; Zanardi, T.A.; Warren, M.S.; Derosier, F.J.; Viney, N.J.; Pham, N.C.; Kaeser, G.E.; Baker, B.F.; et al. IONIS-PKKRx a Novel Antisense Inhibitor of Prekallikrein and Bradykinin Production. *Nucleic Acid Ther.* **2019**, 29, 82-91. doi: 10.1089/nat.2018.0754
- 40. Intellia Therapeutics, Inc. https://www.intelliatx.com/our-science/publications-and-presentations/ [last accessed 26 April 2023]
- 41. KalVista Pharmaceuticals. https://www.kalvista.com/products-pipeline/factor-xiia [last accessed 26 April 2023]
- 42. Craig, T.J.; Reshef, A.; Li, H.H.; Jacobs, J.S.; Bernstein, J.A.; Farkas, H.; Yang, W.H.; Stroes, E.S.G.; Ohsawa, I.; Tachdjian, R.; et al. Efficacy and safety of garadacimab, a factor XIIa inhibitor for hereditary angioedema prevention (VANGUARD): a global, multicentre, randomised, double-blind, placebo-controlled, phase 3 trial. *Lancet* 2023, 401, 1079-1090. doi: 10.1016/S0140-6736(23)00350-1
- 43. Liu, J.; Qin, J.; Borodovsky, A.; Racie, T.; Castoreno, A.; Schlegel, M.; Maier, M.A.; Zimmerman, T.; Fitzgerald, K.; Butler, J.; Akinc, A. An investigational RNAi therapeutic targeting Factor XII (ALN-F12) for the treatment of hereditary angioedema. *RNA* **2019**, *25*, 255-263. doi: 10.1261/rna.068916.118
- Arrowhead Pharmaceuticals, Inc. http://ir.arrowheadpharma.com/news-releases/news-releasedetails/arrowheadpharmaceuticals-presents-new-data-arc-f12-and-arc-lpa [Last accessed 1 May 2023].
- 45. Wintenberger, C.; Boccon-Gibod, I.; Launay, D.; Fain, O.; Kanny, G.; Jeandel, P.Y.; Martin, L.; Gompel, A.; Bouillet, L. Tranexamic acid as maintenance treatment for non-histaminergic angioedema: analysis of efficacy and safety in 37 patients. *Clin. Exp. Immunol.* **2014**, *178*, 112-117. doi: 10.1111/cei.12379
- 46. Maurer, M.; Aberer, W.; Caballero, T.; Bouillet, L.; Grumach, A.S.; Botha, J.; Andresen, I.; Longhurst, H.J.; IOS Study Group. The Icatibant Outcome Survey: 10 years of experience with icatibant for patients with hereditary angioedema. Clin. Exp. Allergy 2022, 52, 1048-1058. doi: 10.1111/cea.14206

- 47. Maurer, M.; Anderson, J.; Aygören-Pürsün, E.; Bouillet, L.; Baeza, M.L.; Chapdelaine, H.; et al. Efficacy And Safety of Bradykinin B2 Receptor Inhibition With Oral PHVS416 In Treating Hereditary Angioedema Attacks: Results Of RAPIDe-1 Phase 2 Trial. *J. Allergy Clin. Immunol.* 2023, 151, Supplement AB134. doi: 10.1016/j.jaci.2022.12.419
- 48. Marceau, F.; Regoli, D. Bradykinin receptor ligands: therapeutic perspectives. *Nat. Rev. Drug Discov.* **2004**, *3*, 845-852. doi: 10.1038/nrd1522
- 49. Petersen, R.S.; Fijen, L.M.; Levi, M.; Cohn, D.M. Hereditary Angioedema: The Clinical Picture of Excessive Contact Activation. *Semin. Thromb. Hemost.* **2022**, in press. doi: 10.1055/s-0042-1758820
- 50. Valerieva, A.; Longhurst, H.J. Treatment of hereditary angioedema-single or multiple pathways to the rescue. Front. Allergy **2022**, 3, 952233. doi: 10.3389/falgy.2022.952233
- 51. Maurer, M; Magerl, M.; Betschel, S.; Aberer, W.; Ansotegui, I.J.; Aygören-Pürsün, E.; Banerji, A,; Bara, N.A.; Boccon-Gibod, I.; Bork. K.; et al. The international WAO/EAACI guideline for the management of hereditary angioedema The 2021 revision and update. *World Allergy Organ J.* 2022, 15, 100627. doi: 10.1016/j.waojou.2022.100627
- Bork, K.; Wulff, K.; Witzke, G.; Staubach, P.; Hardt, J.; Meinke, P. Gene Mutations Linked to Hereditary Angioedema in Solitary Angioedema Patients With Normal C1 Inhibitor. *J. Allergy Clin. Immunol. Pract.* 2023, 12, in press, S2213-2198(23)00177-0. doi: 10.1016/j.jaip.2023.01.051
- 53. Chen ZL, Singh PK, Horn K, Calvano MR, Kaneki S, McCrae KR, Strickland S, Norris EH. Anti-HK antibody inhibits the plasma contact system by blocking prekallikrein and factor XI activation in vivo. *Blood Adv.* **2023**, *7*, 1156-1167. doi: 10.1182/bloodadvances.2021006485
- 54. Reshef, A.; Zanichelli, A.; Longhurst, H.; Relan, A.; Hack, C.E. Elevated D-dimers in attacks of hereditary angioedema are not associated with increased thrombotic risk. *Allergy* **2015**, *70*, 506-513. doi: 10.1111/all.12587
- 55. Maas, C. Plasminflammation-An Emerging Pathway to Bradykinin Production. *Front. Immunol.* **2019**, 10, 2046. doi: 10.3389/fimmu.2019.02046
- 56. Marceau, F.; Bachelard, H.; Rivard, G.É; Hébert, J. Increased fibrinolysis-induced bradykinin formation in hereditary angioedema confirmed using stored plasma and biotechnological inhibitors. *BMC Res. Notes* **2019**, *12*, 291. doi: 10.1186/s13104-019-4335-8
- Zotter, Z.; Csuka, D.; Szabó, E.; Czaller, I.; Nébenführer, Z.; Temesszentandrási, G.; Fust, G.; Varga,
 L.; Farkas, H. The influence of trigger factors on hereditary angioedema due to C1-inhibitor deficiency. Orphanet J. Rare Dis. 2014, 9: 44. doi: 10.1186/1750-1172-9-44
- 58. von Känel, R. Acute mental stress and hemostasis: When physiology becomes vascular harm. *Thromb Res.* **2015**, 135 Suppl 1, *S52-S55*. doi: 10.1016/S0049-3848(15)50444-1
- 59. Strassen, U.; Bas, M.; Wirth, M.; Wirth, M.; Gröger, M.; Stelter, K.; Volkenstein, S.; Kehl, V.; Kojda, G.; Hoffmann, T.K.; et al. Efficacy of human C1 esterase inhibitor concentrate for treatment of ACE-inhibitor induced angioedema. *Am. J. Emerg. Med.* **2023**, 64, 121-128. doi: 10.1016/j.ajem.2022.12.001
- 60. CSL, Limited. https://www.csl.com/research-and-development/product-pipeline [last accessed 1 May 2023]
- 61. Liu, J.; Cooley, B.C.; Akinc, A.; Butler, J.; Borodovsky, A. Knockdown of liver-derived factor XII by GalNAc-siRNA ALN-F12 prevents thrombosis in mice without impacting hemostatic function. *Thromb. Res.* **2020**, *196*, 200-205. doi: 10.1016/j.thromres.2020.08.040
- 62. I-SPY COVID Consortium. Report of the first seven agents in the I-SPY COVID trial: a phase 2, open label, adaptive platform randomised controlled trial. *EClinicalMedicine* **2023**, *58*, 101889. doi: 10.1016/j.eclinm.2023.101889

- 63. Shakur, H.; Andrews, P.; Asser, T.; Balica, L.; Boeriu, C.; Quintero, J.D.; Dewan, Y.; Druwé, P.; Fletcher, O.; Frost, C.; et al. The BRAIN TRIAL: a randomised, placebo controlled trial of a Bradykinin B2 receptor antagonist (Anatibant) in patients with traumatic brain injury. *Trials* **2009**, *10*, 109. doi: 10.1186/1745-6215-10-109
- 64. Werner CG, Pavelka K, Nizzardo A, Rossi C, Scartoni S, Contini MP, di Molfetta S, Bertolotti M, Capriati A, Maggi CA. A Double-Blind, Randomized, Controlled, Four parallel Arm, Dose-Finding Study to Evaluate the Efficacy, Safety, Tolerability, and Pharmacokinetics of Single Intra-Articular (IA) Injections of Fasitibant in Patients with Symptomatic OA of the Knee [abstract]. *Arthritis Rheumatol.* 2015, 67 (suppl 10). https://acrabstracts.org/abstract/a-double-blind-randomized-controlled-four-parallel-arm-dose-finding-study-to-evaluate-the-efficacy-safety-tolerability-and-pharmacokinetics-of-single-intra-articular-ia-injections-of-fas/. [Last Accessed 1 May 2023].
- 65. Lang, G.E.; Tadayoni, R.; Tang, W.; Barth, C.; Weiss-Haljiti, C.; Chong, V. BI 1026706 Study Group. Bradykinin 1 Receptor Antagonist BI1026706 Does Not Reduce Central Retinal Thickness in Center-Involved Diabetic Macular Edema. *Transl. Vis. Sci. Technol.* **2020**, *9*, 25. doi: 10.1167/tvst.9.4.25. PMID: 32818112; PMCID: PMC7396196.
- Nicoletti, N,F.; Sénécal, J.;, da Silva, V.D.; Roxo, M.R.; Ferreira, N.P.; de Morais, R.L.T.; Pesquero, J.B.;,
 Campos, M.M.; Couture, R.; Morrone, F.B. Primary Role for Kinin B1 and B2 Receptors in Glioma
 Proliferation. *Mol. Neurobiol.* 2017, 54, 7869-7882. doi: 10.1007/s12035-016-0265-9
- 67. Straka, B.T.; Ramirez, C.E.; Byrd, J.B.; Stone, E.; Woodard-Grice, A.; Nian, H.; Yu, C.; Banerji, A.; Brown, N.J. Effect of bradykinin receptor antagonism on ACE inhibitor-associated angioedema. *J. Allergy Clin. Immunol.* **2017**, *140*, 242-248.e2. doi: 10.1016/j.jaci.2016.09.051
- 68. Arai, Y.; Takanashi, H.; Kitagawa, H.; Wirth, K.J.; Okayasu, I. Effect of icatibant, a bradykinin B2 receptor antagonist, on the development of experimental ulcerative colitis in mice. *Dig. Dis. Sci.* **1999**, 44, 845-851.
- 69. Kamata, K.; Hayashi, I.; Mizuguchi, Y.; Arai, K.; Saeki, T.; Ohno, T.; Saigenji, K.; Majima, M. Suppression of dextran sulfate sodium-induced colitis in kininogen-deficient rats and non-peptide B2 receptor antagonist-treated rats. Jpn. J. Pharmacol. 2002, 90, 59-66. doi: 10.1254/jjp.90.59
- 70. Hirata, M.; Hayashi, I.; Yoshimura, K.; Ishii, K.; Soma, K.; Ohwada, T.; Kakita, A.; Majima, M. Blockade of bradykinin B2 receptor suppresses acute pancreatitis induced by obstruction of the pancreaticobiliary duct in rats. *Br. J. Pharmacol.* **2002**, *135*, 29-36. doi: 10.1254/jjp.90.59
- 71. Kanbe, T.; Naruse, S.; Kitagawa, M.; Nakae, Y.; Hayakawa, T. Effects of a bradykinin receptor antagonist (HOE140) on taurocholate-induced acute pancreatitis in rats. *Pancreas* **1996**, *13*, 283-288. doi: 10.1097/00006676-199610000-00011
- 72. Jonkam, C.C.; Enkhbaatar, P.; Nakano, Y.; Boehm, T.; Wang, J.; Nussberger, J.; Esechie, A.; Traber, L.D.; Herndon, D.; Traber, D.L. Effects of the bradykinin B2 receptor antagonist icatibant on microvascular permeability after thermal injury in sheep. *Shock* 2007, 28, 704-709. doi: 10.1097/shk.0b013e3180536124
- 73. Chen, M.M.; O'Halloran, E.B.; Ippolito, J.A.; Choudhry, M.A.; Kovacs, .J. Alcohol potentiates postburn remote organ damage through shifts in fluid compartments mediated by bradykinin. *Shock* **2015**, *43*, 80-84. doi: 10.1097/SHK.0000000000000000055
- 74. Griesbacher, T.; Legat, F.J. Effects of the non-peptide B2 receptor antagonist FR173657 in models of visceral and cutaneous inflammation. *Inflamm. Res.* **2000**, *49*, 535-540. doi: 10.1007/s000110050628
- 75. Wirth, K.J.; Alpermann, H.G.; Satoh, R.; Inazu, M. The bradykinin antagonist Hoe 140 inhibits carrageenan- and thermically induced paw oedema in rats. *Agents Actions Suppl.* **1992**, *38*, 428-431.

- 76. Dubuc, C.; Savard, M.; Bovenzi, V.; Lessard, A.; Côté, J.; Neugebauer, W.; Geha, S.; Chemtob, S.; Gobeil, F. Jr. Antitumor activity of cell-penetrant kinin B1 receptor antagonists in human triplenegative breast cancer cells. *J. Cell. Physiol.* 2019, 234, 2851-2865. doi: 10.1002/jcp.27103
- 77. Fincham, C.I.; Bressan, A.; Paris, M.; Rossi, C.; Fattori, D. Bradykinin receptor antagonists a review of the patent literature 2005-2008. *Expert Opin. Ther. Pat.* **2009**, *19*, 919-941. doi: 10.1517/13543770902994389
- 78. Hara, D.B.; Leite, D.F.; Fernandes, E.S.; Passos, G.F.; Guimarães, A.O.; Pesquero, J.B.; Campos, M.M.; Calixto, J.B. The relevance of kinin B1 receptor upregulation in a mouse model of colitis. *Br. J. Pharmacol.* **2008**, *154*, 1276-1286. doi: 10.1038/bjp.2008.212
- 79. Sexton, D.J.; Chen, T.; Martik, D.; Kuzmic, P.; Kuang, G.; Chen, J.; Nixon, A.E.; Zuraw, B.L.; Forteza, R.M.; Abraham, W.M.; et al. Specific inhibition of tissue kallikrein 1 with a human monoclonal antibody reveals a potential role in airway diseases. *Biochem J.* **2009**, *422*, 383-392. doi: 10.1042/BJ20090010
- 80. Moran, C.S.; Biros, E.; Krishna, S.M.; Morton, S.K.; Sexton, D.J.; Golledge, J. Kallikrein-1 Blockade Inhibits Aortic Expansion in a Mouse Model and Reduces Prostaglandin E2 Secretion From Human Aortic Aneurysm Explants. *J. Am. Heart Assoc.* **2021**, *10*, e019372. doi: 10.1161/JAHA.120.019372
- 81. Brusco, I.; Becker, G.; Palma, T.V.; Pillat, M.M.; Scussel, R.; Steiner, B.T.; Sampaio, T.B.; Ardisson-Araújo, D.M.P.; de Andrade, C.M.; Oliveira, M.S.; et al. Kinin B1 and B2 receptors mediate cancer pain associated with both the tumor and oncology therapy using aromatase inhibitors. *Sci. Rep.* **2023**, 13, 4418. doi: 10.1038/s41598-023-31535-6
- 82. Mendes, G.M.M.; Do Nascimento, I.J.B.; Marazzi-Diniz, P.H.; Da Silveira, I..B; Itaborahy, M.F.; Viana, L.E.; Silva, F.A.; Santana, M.F.; Pinto, R.A. Dutra, B.G.; et al. The des-Arg⁹-bradykinin/B₁R axis: Hepatic damage in COVID-19. *Front, Physiol.* **2022**, *13*, 1080837. doi: 10.3389/fphys.2022.1080837
- 83. Yang, Y.; Wang, J.; Shi, F.; Shan, A.; Xu, S.; Lv, W. BDKRB2 is a novel EMT-related biomarker and predicts poor survival in glioma. *Aging (Albany NY)*, **2021**, *13*, 7499-7516. doi: 10.18632/aging.202614
- 84. Gainer, J.V.; Morrow, J.D.; Loveland, A.; King, D.J.; Brown, N.J. Effect of bradykinin-receptor blockade on the response to angiotensin-converting-enzyme inhibitor in normotensive and hypertensive subjects. *N. Engl. J. Med.* **1998**, 339, 1285-1292. doi: 10.1056/NEJM199810293391804
- 85. Hoover, T.; Lippmann, M.; Grouzmann, E.; Marceau, F.; Herscu, P. Angiotensin converting enzyme inhibitor induced angio-oedema: a review of the pathophysiology and risk factors. *Clin. Exp. Allergy* **2010**, *40*, 50-61. doi: 10.1111/j.1365-2222.2009.03323.x
- 86. Hubers, S.A.; Kohm, K.; Wei, S.; Yu, C.; Nian, H.; Grabert, R.; Sexton, D.J.; Brown, N.J. Endogenous bradykinin and B1-B5 during angiotensin-converting enzyme inhibitor-associated angioedema. *J. Allergy Clin. Immunol.* **2018**, *142*, 1636-1639.e5. doi: 10.1016/j.jaci.2018.06.037
- 87. Cheong, E;, Dodd, L;, Smith, W; Kleinig, T. Icatibant as a Potential Treatment of Life-Threatening Alteplase-Induced Angioedema. *J. Stroke Cerebrovasc. Dis.* **2018**, 27, e36-e37. doi: 10.1016/j.jstrokecerebrovasdis.2017.09.039
- 88. Brown, E.; Campana, C.; Zimmerman, J.; Brooks, S. Icatibant for the treatment of orolingual angioedema following the administration of tissue plasminogen activator. *Am. J. Emerg. Med.* **2018**, 36, 1125.e1-1125.e2. doi: 10.1016/j.ajem.2018.03.018
- 89. Stone, O.A.; Richer, C.; Emanueli, C.; van Weel, V.; Quax, P.H.; Katare, R.; Kraenkel, N.; Campagnolo, P.; Barcelos, L.S.; Siragusa, M.; et al. Critical role of tissue kallikrein in vessel formation and maturation: implications for therapeutic revascularization. Arterioscler. Thromb. Vasc. Biol. 2009, 29, 657-664. doi: 10.1161/ATVBAHA.108.182139

- 90. Koch, M.; Spillmann, F.; Dendorfer, A.; Westermann, D.; Altmann, C.; Sahabi, M.; Linthout, S.V.; Bader, M.; Walther, T.; Schultheiss, H.P.; et al. Cardiac function and remodeling is attenuated in transgenic rats expressing the human kallikrein-1 gene after myocardial infarction. Eur. J. Pharmacol. 2006, 550, 143-148. doi:10.1016/j.ejphar.2006.08.054
- 91. Katori, M.; Majima, M. Renal (tissue) kallikrein-kinin system in the kidney and novel potential drugs for salt-sensitive hypertension. *Prog. Drug Res.* **2014**, *69*, 59-109. doi: 10.1007/978-3-319-06683-7_4
- 92. Bergaya, S.; Meneton, P.; Bloch-Faure, M.; Mathieu, E.; Alhenc-Gelas, F.; Lévy, B.I.; Boulanger, C.M. Decreased flow-dependent dilation in carotid arteries of tissue kallikrein-knockout mice. *Circ. Res.* **2001**, *88*, 593-599. doi: 10.1161/01.res.88.6.593
- 93. Wu, J.; Wu, J.; Wang, L.; Liu, J. Urinary Kallidinogenase plus rt-PA Intravenous Thrombolysis for Acute Ischemic Stroke: A Systematic Review and Meta-Analysis of Randomized Controlled Trials. *Comput. Math. Methods Med.* **2022**, 2022, 1500669. doi: 10.1155/2022/1500669
- 94. Alexander-Curtis, M.; Pauls, R.; Chao, J.; Volpi, J.J.; Bath, P.M.; Verdoorn, T.A. Human tissue kallikrein in the treatment of acute ischemic stroke. *Ther. Adv. Neurol. Disord.* **2019**, *12*, 1756286418821918. doi: 10.1177/1756286418821918
- 95. Shehjar, F.; Maktabi, B.; Rahman, Z.A.; Bahader, G.A.; James, A.W.; Naqvi, A.; Mahajan, R.; Shah, Z.A. Stroke: Molecular mechanisms and therapies: Update on recent developments. *Neurochem. Int.* **2023**, *162*, 105458. doi: 10.1016/j.neuint.2022.105458
- 96. Charest-Morin, X.; Roy, C.; Fortin, E.J.; Bouthillier, J.; Marceau, F. Pharmacological evidence of bradykinin regeneration from extended sequences that behave as peptidase-activated B2 receptor agonists. *Front. Pharmacol.* **2014**, *5*, 32. doi: 10.3389/fphar.2014.00032
- 97. Jean, M.; Gera, L.; Charest-Morin, X.; Marceau, F.; Bachelard, H. In Vivo Effects of Bradykinin B2 Receptor Agonists with Varying Susceptibility to Peptidases. *Front. Pharmacol.* **2016**, *6*, 306. doi: 10.3389/fphar.2015.00306
- 98. Bachelard, H.; Charest-Morin, X.; Marceau, F. D-Arg⁰-Bradykinin-Arg-Arg, a Latent Vasoactive Bradykinin B2 Receptor Agonist Metabolically Activated by Carboxypeptidases. *Front. Pharmacol.* **2018**, *9*, 273. doi: 10.3389/fphar.2018.00273
- 99. Prados, M.D.; Schold, S.C.; Fine, H.A.; Jaeckle, K.; Hochberg, F.; Mechtler, L.; Fetell, M.R.; Phuphanich, S.; Feun, L.; Janus, T.J.; et al. A randomized, double-blind, placebo-controlled, phase 2 study of RMP-7 in combination with carboplatin administered intravenously for the treatment of recurrent malignant glioma. *Neuro. Oncol.* **2003**, *5*, 96–103. doi: 10.1093/neuonc/5.2.96
- 100. Warren, K.; Jakacki, R.; Widemann, B.; Aikin, A.; Libucha, M.; Packer, R.; Vezina, G.; Reaman, G.; Shaw, D.; Krailo, M.; et al. Phase II trial of intravenous lobradimil and carboplatin in childhood brain tumors: A report from the Children's Oncology Group. *Cancer Chemother. Pharmacol.* **2006**, *58*, 343–347. doi: 10.1007/s00280-005-0172-7
- 101. Côté, J.; Savard, M.; Neugebauer, W.; Fortin, D.; Lepage, M.; Gobeil, F. Dual kinin B1 and B2 receptor activation provides enhanced blood-brain barrier permeability and anticancer drug delivery into brain tumors. *Cancer Biol. Ther.* **2013**, *14*, 806-811. doi: 10.4161/cbt.25327
- 102. Rassias, G.; Leonardi, S.; Rigopoulou, D.; Vachlioti, E.; Afratis, K.; Piperigkou, Z.; Koutsakis, C.; Karamanos, N.K.; Gavras, H.; Papaioannou, D. Potent antiproliferative activity of bradykinin B2 receptor selective agonist FR-190997 and analogue structures thereof: A paradox resolved? *Eur. J. Med. Chem.* 2021, 210, 112948. doi: 10.1016/j.ejmech.2020.112948
- 103. Nunes, M.A.; Toricelli, M.; Schöwe, N.M.; Malerba, H.N.; Dong-Creste, K.E.; Farah, D.M.A.T.; De Angelis, K. Irigoyen, M,C,; Gobeil, F.; Araujo Viel, T.; et al. Kinin B2 Receptor Activation Prevents the

- Evolution of Alzheimer's Disease Pathological Characteristics in a Transgenic Mouse Model. *Pharmaceuticals (Basel)* **2020**, *13*, 288. doi: 10.3390/ph13100288
- 104. Nokkari, A.; Abou-El-Hassan, H.; Mechref, Y.; Mondello, S.; Kindy, M.S.; Jaffa, A.A.; Kobeissy, F. Implication of the Kallikrein-Kinin system in neurological disorders: Quest for potential biomarkers and mechanisms. *Prog. Neurobiol.* **2018**, *165-167*, 26-50. doi: 10.1016/j.pneurobio.2018.01.003