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Communication

Suzuki-Miyaura Reaction in the Presence of *N*-Acetylcysteamine Thioesters Enables Rapid Synthesis of Biomimetic Polyketide Thioester Surrogates for Biosynthetic Studies

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Abstract: Biomimetic *N*-acetylcysteamine thioesters are essential for the study of polyketide synthases, non-ribosomal peptide synthetases and fatty acid synthases. The chemistry for their preparation is however limited by their specific functionalization and their susceptibility to undesired side reactions. This is especially detrimental to transition metal-catalyzed reactions. Here we report a method for the rapid preparation of *N*-acetylcysteamine (SNAC) 7-hydroxy-2-enethioates, which are suitable for the study of various enzymatic domains of megasynthase enzymes, particularly oxygen heterocycle-forming cyclase domains. The method is based on a one-pot sequence of hydroboration and Suzuki-Miyaura reaction. Optimization of the reaction conditions made it possible to suppress potential side reactions and to introduce the highly functionalized SNAC methacrylate unit in high yield. The versatility of the sequence was demonstrated on a dienal precursor, which was subjected to Brown crotylation followed by the hydroboration-Suzuki-Miyaura reaction sequence and deprotection, finally giving a complex polyketide SNAC thioester. Backbone extension by six carbons and a terminal SNAC enethioate was achieved, introducing an *E*-configured double bond and two adjacent stereocenters in a highly selective manner. The presented method allows for the synthesis of the target motif in significantly fewer steps and with higher overall yield than previously described approaches, while maintaining higher flexibility and control over the stereogenic elements. It is also the first reported example of a transition metal-catalyzed cross-coupling reaction in the presence of an SNAC thioester.

Keywords: Suzuki-Miyaura reaction; biomimetic thioesters; polyketide synthases; enzymes; cyclases

1. Introduction

Thioesters are an important functional group in many biosynthetic systems. They often serve to link biosynthetic acyl intermediates to carrier thiols, which can be free molecules such as coenzyme A (CoA) or proteins. Important systems working with protein-bound metabolites are so-called megasynthase enzymes like fatty acid synthases, polyketide synthases (PKS) and non-ribosomal peptide synthetases (NRPS) and their hybrids^{1,2}. They are responsible for the formation of polyketide and peptide natural products, including some of the most important small-molecule drugs in clinical use, such as erythromycin, rapamycin or epothilone. The availability of suitable substrate surrogates is essential for the functional study of these biosynthetic systems. *N*-Acetylcysteamine (SNAC) thioesters are of particular importance for this as they effectively mimic protein attachment of the substrate via the 4'-phosphopantetheine arm and thus allow simplified studies with active enzymes (Figure 1A)^{3–10}.

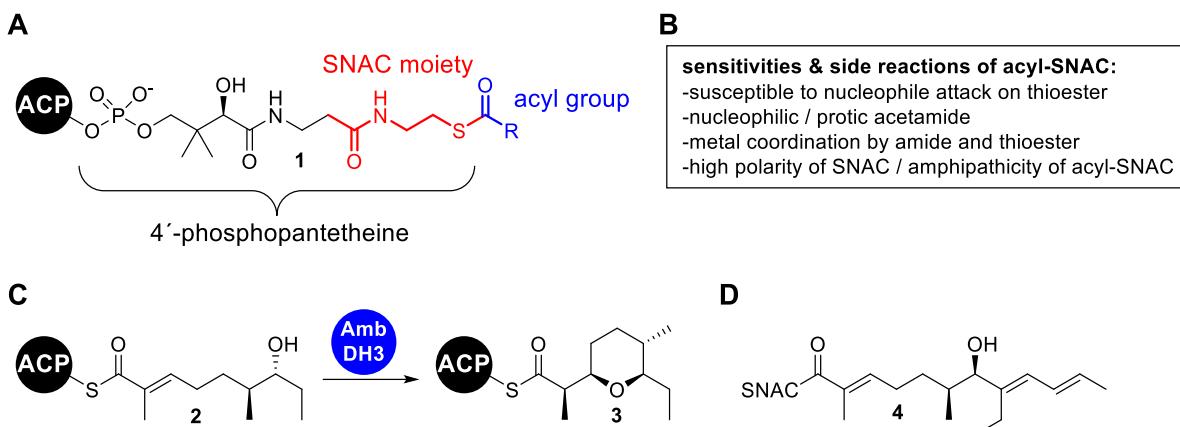


Figure 1. (A) Structure of the 4'-phosphopantetheine prosthetic group of acyl carrier proteins and the partial structure that is mimicked by SNAC. (B) Sensitivities & side reactions of acyl-SNACs. (C) IMOMA cyclases catalyze intramolecular oxa-Michael addition to oxygen heterocycles. The natural reaction of AmbDH3 is shown as an example. (D) Structure of the target compound required for our biosynthetic studies.

Acylated SNACs contain an acetamide and a thioester as conserved reactive functional groups that afford them with problematic properties (Figure 1B)^{7,11}. The thioester can undergo side reactions with external or internal nucleophiles, resulting in irreversible loss of substance. Due to its polarity, the acetamide can cause problems during substance purification and can, as a nucleophilic/protic group, cause undesired side reactions. The functionalization distance between acetamide and thioester carries the risk that they act as a chelate ligand and interact with metals. The synthesis of SNAC thioester surrogates of late-stage biosynthetic intermediates is as challenging as the synthesis of natural products of similar structural complexity but, for the above-mentioned reasons, has the challenge of an additional problematic functional group. A useful strategy to overcome this problem would be to introduce the SNAC moiety at a late stage of synthesis along with a larger fraction of the polyketide moiety.

Improving the specific methodology for the synthesis of complex polyketide-SNAC thioesters is therefore of great interest to the biosynthetic research community. Transition metal-mediated reactions are well suited for late-stage attachment in the convergent synthesis of complex biosynthetic thioester surrogates but have only very rarely been described in the presence of SNAC thioesters. To the best of our knowledge, the literature currently only contains a report about olefin cross metathesis between SNAC-acrylates and hydroxyolefins catalyzed by the 2nd generation Grubbs catalyst¹².

The Suzuki-Miyaura reaction (SMR) is a highly versatile Pd-catalyzed cross-coupling reaction. It allows couplings between halides and non-toxic boronic acid derivatives under relatively mild conditions (Figure 2).^{13,14} In addition to *sp*²-*sp*² bond formations, it is now possible to carry out couplings between *sp*² and *sp*³ centers as well as between two *sp*³ centers. Two aspects of the SMR could be problematic when applied to SNAC thioesters. On the one hand, the use of base is necessary to accelerate the essential group transfer from the boronic acid to the Pd during the catalytic cycle (step 3). Moreover, Pd can also insert into the C-S bond of the thioester instead of the C-halide bond (step 1).¹⁵ This reactivity is so pronounced that it forms the basis of the Liebeskind-Srogl reaction, a modification of the SMR for the direct synthesis of ketones from thioesters.^{16,17}

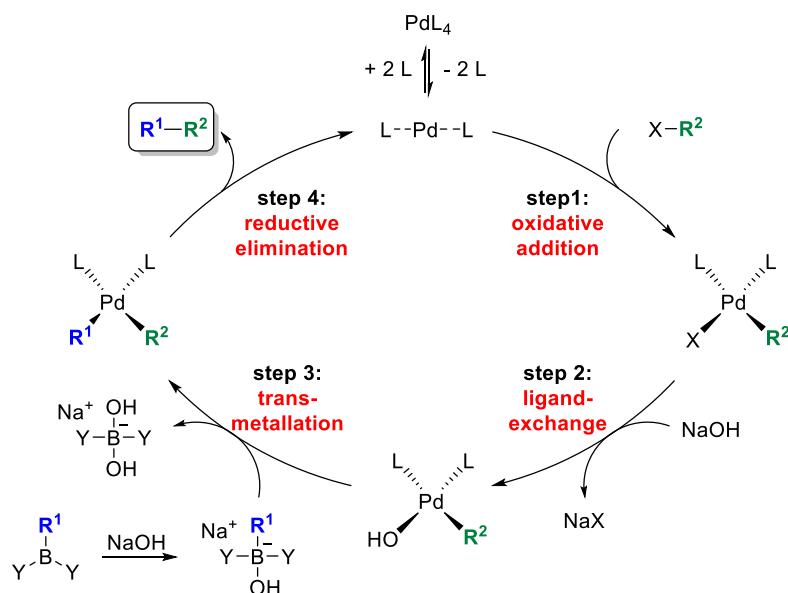


Figure 2. Mechanism of the SMR.

Among the diverse enzymatic PKS domains, cyclases that form saturated oxygen heterocycles by intramolecular oxa-Michael addition (IMOMA) stand out for the synthetical value of this transformation (Figure 1C).^{18,19} It has been shown that they catalyze ring formation with exceptional stereoselectivity and therefore represent a potential new type of biocatalyst.²⁰⁻²⁹ For the study of such enzymes, SNAC-7-hydroxy-2-enethioates are required as substrate surrogates. The synthetic methodology for the selective installation of this structural motif is however not well developed, making the generation of precursor libraries a difficult task. The multi-step routes described in the literature are either highly elaborate, are not stereoselective or lack flexibility and are therefore narrow in their applicability.²⁰⁻²³ For example, the synthesis of the SNAC surrogate of **2** in stereochemical pure form was accomplished in eight steps and required multiple purification procedures.^{21,22} A lack of convergence furthermore makes it necessary to carry out the largest part of this sequence from different starter building blocks to access derivatives with variations in the eastern part of the molecule. Other reported routes are shorter, but also less flexible due to the choice of larger starting building blocks or the choice of the introduction reaction for the SNAC thioester. Olefin cross-metathesis for example is only possible with SNAC-acrylthioates and not with SNAC-methacrylthioates. Therefore, we set out to develop a flexible, straightforward and broadly applicable method for the preparation of SNAC-7-hydroxy-2-enethioates.

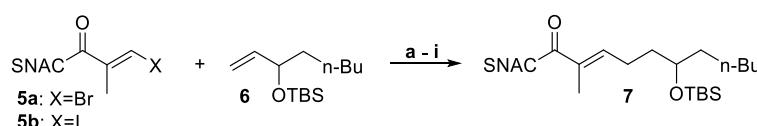
As a solution, we turned to a sequence of hydroboration and SMR to assemble the backbone and directly introduce the SNAC moiety. The specific challenge was to effectively perform the SMR in the presence of the SNAC thioester, which has not been achieved before to the best of our knowledge. The versatility of the method should be shown on the example of the synthesis of **4** (Figure 1D). This compound was on the one hand specifically required for our enzymatic studies on new IMOMA cyclases. On the other hand, it represents a particularly challenging substrate during the preparation of which various detrimental side reactions could occur and thus a reasonable benchmark.

2. Results and Discussion

Thioester-halides are rare substrates in SMRs. The literature however contains an example of coupling reactions of simple 4-bromothiophenols with 4-tolyl-boronic acid in which the bulky acyl unit of the thioesters served as a protecting group for the thiol¹⁵. We used the reported conditions for the synthesis of an ethylenoate sensitive towards hydrolysis and base described by Suzuki *et al.* as the starting point for our studies³⁰. These were

carried out using the SNAC (*E*)-3-bromo-2-methylprop-2-enethioate **5** and the OTBS-protected 3-hydroxyolefin **6** (1.0 equiv. of **5**, 1.1 equiv. 9-BBN, 5 mol% $\text{PdCl}_2(\text{dppf})$ und 2.0 equiv. K_2CO_3). Fortunately, a basic coupling reactivity was observed. The yields of the reactions however varied hardly reproducibly over a wide range and showed a strong dependence on even small variations of the amounts of thioester, alkene, borane and Pd catalyst. This suggests that several side reactions might proceed at rates similar to the desired pathway. We therefore carried out a systematic optimization study (**Table 1**).

Table 1. Optimization of the conditions for the coupling of SNAC thioester halides **5a/5b** and TBS-protected olefin **6**.



Entry	X	Base	Additive	Temperature [°C]	Isolated yield [%]
a	Br	K_2CO_3	-	50	54
b	Br	K_2CO_3	$\text{P}(o\text{-furyl})_3$	50	23
c	Br	K_2CO_3	AsPh_3	50	55
d	Br	Cs_2CO_3	-	50	55
e	Br	K_2CO_3	-	20	13
f	I	K_2CO_3	-	50	55
g	I	Cs_2CO_3	AsPh_3	50	34
h	I	Cs_2CO_3	AsPh_3	65	-
i	I	Cs_2CO_3	AsPh_3	20	78

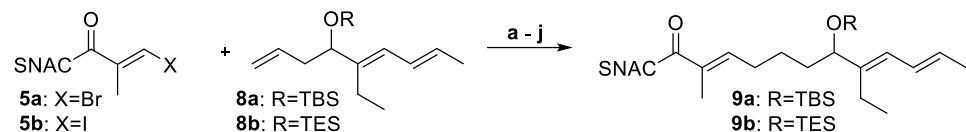
General reaction conditions: 1. **6** (1.5 eq., 1 M in THF), 9-BBN (1.5 eq., 0.5 M in THF), 0 °C to 20 °C, o.n.; 2. DMF (total 0.2 M), **5** (1.0 eq.), base (2.0 eq.), PdCl_2dppf (5 mol%), additive (5 mol%), reaction control *via* TLC. Reaction scale: 90–100 μmol .

For this, we varied the individual reaction parameters. Since we assumed that side reactions of the 3-bromoacryl thioate **5a** were be a particular problem, we worked with an excess of 1.5 equiv. of alkene **6** and 9-BBN. Different thioester halides (Br and I), bases (K_2CO_3 and Cs_2CO_3), additives ($\text{P}(o\text{-furyl})_3$ and AsPh_3) and temperatures (20 °C, 50 °C and 65 °C) were tested. All reactions were carried out on a scale of 90–100 μmol of **5a/5b** and compared based on the isolated yield after column chromatography. The yields in the basic experiment with an excess of 1.5 equiv. of **6** and 9-BBN (entry a) were fortunately stable upon repetition in a range slightly above 50%. The variation of individual parameters did not lead to a marked increase of this value, whereas the addition of $\text{P}(o\text{-furyl})_3$ and the decrease of the reaction temperature even significantly reduced the yield (entries b and e). Fortunately, the combined change of several parameters led to a significantly improved result (entry i). Using 3-iodoacryl thioate **5b**, Cs_2CO_3 , AsPh_3 and carrying out the reaction at room temperature resulted in a yield of 78%.

Side products were regularly detected in the low-yielding reactions that could not be isolated and fully analyzed. According to TLC, these were highly polar compounds whose migration behavior suggests that they are derived from SNAC. We assume that a major part of this is the homocoupling product of the thioester acrylates **5a/5b** and the 2-(*N*-acetamidyl)-ethylketone resulting after C–S bond insertion of the Pd, a side reaction described previously for low-functionalized thioesters.¹⁵ The yield improvement observed in the optimization study would be consistent with the suppression of these side reactions. Cs_2CO_3 is much better soluble in DMF than K_2CO_3 leading to a much higher effective concentration of carbonate. This should significantly improve the activation of the *ate* complex for alkyl group transfer to the Pd (step 3 in **Figure 2**) and accelerate the heterocoupling reaction. The iodoacrylate is more reactive towards Pd insertion than the bromoacrylate, thus favoring this productive reaction (step 1) compared to insertion of the Pd

into the C-S bond. This selectivity is expected to be even more pronounced at room temperature than at 50 °C. The addition of AsPh₃ supports these effects by accelerating both, the formation of the active Pd(0) from the Pd(II) species and the transmetallation due to its lower σ-donor effect than PPh₃.^{31,32}

Table 2. Optimization of the conditions for the coupling of **5** and protected trienes **8a/8b**, varying protecting group, halogenide, base, additives and temperature.

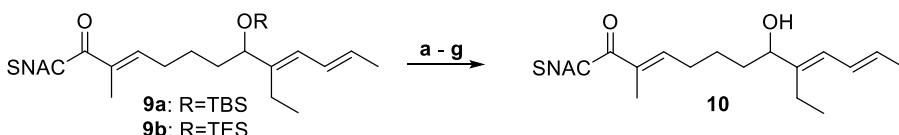


Entry	X	PG	Base	Additive	Temperature [° C]	Isolated yield [%]
a	Br	TBS	2 eq. K ₂ CO ₃	-	50	27
b	Br	TBS	3 eq. K ₃ PO ₄	-	50	17
c	Br	TES	2 eq. K ₂ CO ₃	-	50	25
d	Br	TES	3 eq. K ₃ PO ₄	-	50	12
e	Br	TES	2 eq. K ₂ CO ₃	-	20	15
f	I	TES	2 eq. K ₂ CO ₃	-	50	49
g	Br	TES	2 eq. Cs ₂ CO ₃	-	50	80
h	Br	TES	2 eq. K ₂ CO ₃	AsPh ₃	50	77
i	I	TES	2 eq. K ₂ CO ₃	-	20	63
j	I	TES	2 eq. Cs ₂ CO ₃	AsPh ₃	20	87

Reaction conditions: 1. 8 (1.5 eq., 1 M in THF), 9-BBN (1.5 eq., 0.5 M in THF), 0 °C to 20 °C, o.n.; 2. DMF (total 0.2 M), **5** (1.0 eq.), Base (2.0 eq.), PdCl₂dppf (5 mol%), additive (5 mol%), reaction control *via* TLC.

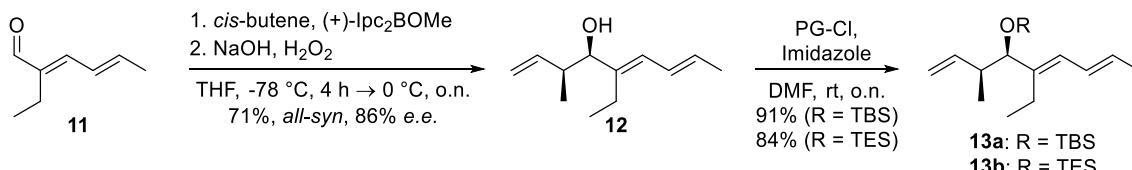
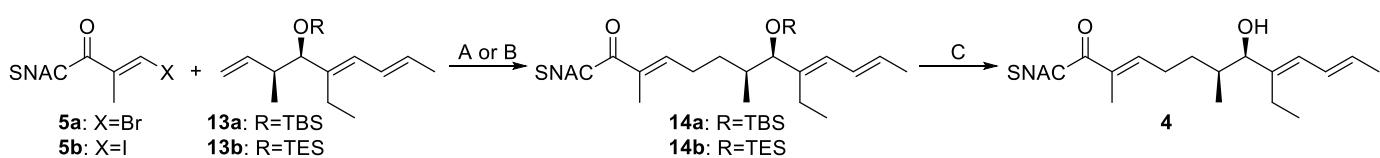
We shifted to the coupling between the thioester **5** with the olefins **8a** and **8b** that resemble the sensitive 5-hydroxy-tri-1,3,7-ene in target molecule **4** (Table 2). The higher degree of functionalization makes them more susceptible to side reactions during the introduction and removal of the protecting group and during the coupling cascade. Beside screening the same thioester halides (**5a/5b**) as in Table 1, a broader panel of bases (Cs₂CO₃, K₂CO₃ and K₃PO₄) and hydroxyl protection groups (TBS and TES) on the olefinic coupling partner were examined. Due to the superiority in the previous optimization, only AsPh₃ was applied as an additive and only 20 °C and 50 °C were tested as reaction temperature.

Compared to the basic experiments (entries a and c, Table 2), a decrease in yield was observed when the reaction was carried out at 20 °C instead of 50 °C or when K₃PO₄ was employed as a base (entries a–e). In contrast to the experiments with the simpler coupling partner **6** (Table 1), the change of one or two reaction parameters already led to an increase of the yield up to 80% (entries f–i). When these measures were combined and the reaction was carried out at 20 °C, a further increase to 87% yield was achieved (entry j). The TES group is expected to be easier removable than the TBS group (*vide infra*). As it showed to be stable under the conditions tested and as both protecting groups gave similar yields in the comparable entries a–d, the optimization in entries f–j was conducted with this silyl protection group. The results summarized in Table 2 are in agreement with those observed in Table 1 and confirm the conclusions / interpretations drawn from them.

Table 3. Testing conditions for silyl ether deprotection.

Entry	PG	Reagent	Conditions	Result
a	TBS	PPTS	DMSO, 50 °C, o.n.	Decomposition
b	TBS	TBAF	THF, 0 °C, 1 h	No reaction
c	TBS	TBAF	THF, 0 → 20 °C, o.n.	Decomposition
d	TES	TBAF	THF, 0 °C, 1 h	Decomposition
e	TBS	HF*pyridine	THF, 0 °C, 3 h	Decomposition
f	TBS	HF*pyridine, pyridine	THF, 0 → 20 °C, 3 h	51%
g	TES	HF*pyridine, pyridine	THF, 0 → 20 °C, 3 h	81%

Numerous side reactions are conceivable during the deprotection of silyl ethers **9a** and **9b**, such as eliminations, intramolecular oxa-Michael additions or interference with the thioester. The slightly acidic conditions in the presence of PPTS resulted in elimination of the alcohol / silyl ether (entry a, **Table**). With TBAF, the formation of the desired product was also not observed in any case. No reaction of the TBS ether **9a** was found after 1 h at 0 °C (entry b). Decomposition occurred for the TBS ether **9b** after overnight reaction at 20 °C and for the TES ether already after 1 h at 0 °C (entries c and d). Standard HF*pyridine treatment also resulted in decomposition (entry e). A successful procedure was finally adopted from a protocol previously reported by Carreira *et al.*, which relied on using a premixed stock solution of HF*pyridine in THF supplemented with additional pyridine at 0 °C³³. Deprotection was successful for both silyl ethers and led to the desired alcohol **10** in pure form after column chromatography (entries f and g). The reactions were continuously monitored by TLC and stopped before noticeable decomposition occurred. The yield for the TES ether was significantly better than that of the TBS ether, suggesting the former to be the preferable protecting group for the synthesis of **4**.

**Scheme 1.** Synthesis of protected hydroxytrienes **13a**/**13b** from aldehyde **11**.**Table 4.** Two-pot-three-step reaction sequence to thioester **4**.

Entry	X	PG	Conditions	Coupling yield [%]	Deprotection yield [%]	Overall yield [%]
a	Br	TBS	A	30	53	16
b	Br	TES	A	51	86	44
c	I	TES	B	74	81	60

Reaction conditions: A. **13** (1.0 eq., 1 M in THF), 9-BBN (1.0 eq., 0.5 M in THF), 0 °C to 20 °C, o.n.; 2. DMF (total 0.2 M), **5** (1.5 eq.), K₂CO₃ (2.0 eq.), PdCl₂dppf (5 mol%), 50 °C, reaction control *via* TLC; B. **13** (1.5 eq., 1 M in THF), 9-BBN (1.5 eq., 0.5 M in THF), 0 °C to 20 °C, o.n.; 2. DMF (total 0.2 M), **5** (1.0 eq.), Cs₂CO₃ (2.0 eq.), PdCl₂dppf (5 mol%), AsPh₃ (5 mol%), 20 °C, reaction control *via* TLC; C. **14** (10.0 mg, 1.0 eq.), 110 μL of HF-containing stock solution (1 part HF*pyridine, 2 parts pyridine, 8 parts THF).

The reaction sequences to **4** were carried out starting from aldehyde **11**. Brown crotylation first afforded the highly sensitive hydroxytriene **12** in 71% yield with an *e.e.* of 86%, which was immediately transformed into the isolatable TBS and TES ethers **13a** and **13b** in 91% and 84% yield (Scheme 1). This was followed by the established one-pot-two-step-cascade of hydroboration and SMR to give **14a** and **14b**, which were deprotected to give 7-hydroxy-2-ene-SNAC thioate **4** in overall yields of 16–60% (Table). These results confirm on the one hand that TES is the preferable protecting group compared to TBS (entries a and b). On the other hand, they show the positive effect of optimizing the SMR conditions, which led to a yield improvement from 51% to 74% in the coupling step (entries b and c).

In total, product **4** was obtained in four synthetic operations from aldehyde **11** with an overall yield of 36% in high stereoisomeric purity. This represents a significant improvement over previously described routes to similar compounds, which either required significant more steps and gave lower overall yields (8 steps, 10% overall yield for the SNAC thioester analog of **2**). Other routes gave 7-hydroxy-2-ene SNAC thioates in five steps from TBS-protected 1,5-hexanediol with a total yield of 23%. The latter however only gave access to racemic products, which were also not branched in the α -position, and did not offer the flexibility in backbone installation that the presented method does.

5. Conclusions

The SMR-based coupling method presented here is compatible with the presence of SNAC thioesters and can be used in the future for the flexible and efficient preparation of substrate surrogates for studies of IMOMA cyclases. Such 2-ene thioates can also be used for studies of other catalytic megasynthase domains that act on similar functionalization patterns as present at C-1–C-6 in **4**. The method should also be of interest for the synthesis of precursors of non-enzymatic IMOMA reactions. It has been shown for chemically-catalyzed IMOMA reactions that *cis*-THP stereoselectivity can be more reliably achieved with enethioates than with enoates so that the former are attractive precursors.

Supplementary Materials: The following supporting information can be downloaded at: www.mdpi.com/xxx/s1, The Supplementary Materials contain detailed synthetic procedures and analytical data including ^1H and ^{13}C NMR spectra.

Author Contributions: Conceptualization, S.D. and F.H.; investigation, S.D. and L.S.; resources, F.H.; writing—original draft preparation, F.H.; writing—review and editing, S.D., L.S. and F.H.; supervision, F.H.; project administration, F.H.; funding acquisition, F.H. All authors have read and agreed to the published version of the manuscript.

Funding: This research was funded by the Deutsche Forschungsgemeinschaft (DFG), grant number HA 5841/5-1.

Data Availability Statement: The data presented in this study are available on request from the corresponding author (Prof. Frank Hahn).

Acknowledgments: We thank Central Analytics of the Department of Chemistry as well as the North Bavarian NMR Centre (NBNC) at the University of Bayreuth.

Conflicts of Interest: The authors declare no conflict of interest.

References

- (1) Weissman, K. J.; Müller, R. Protein–Protein Interactions in Multienzyme Megasynthetases. *ChemBioChem* **2008**, *9* (6), 826–848. <https://doi.org/10.1002/cbic.200700751>.
- (2) Grininger, M. Enzymology of Assembly Line Synthesis by Modular Polyketide Synthases. *Nat. Chem. Biol.* **2023**, *19* (4), 401–415. <https://doi.org/10.1038/s41589-023-01277-7>.
- (3) Ge, H.-M.; Huang, T.; Rudolf, J. D.; Lohman, J. R.; Huang, S.-X.; Guo, X.; Shen, B. Enediyne Polyketide Synthases Stereoselectively Reduce the β -Ketoacyl Intermediates to β -d-Hydroxyacyl Intermediates in Enediyne Core Biosynthesis. *Org. Lett.* **2014**, *16* (15), 3958–3961. <https://doi.org/10.1021/o1501767v>.

(4) Sahner, J. H.; Sucipto, H.; Wenzel, S. C.; Groh, M.; Hartmann, R. W.; Müller, R. Advanced Mutasynthesis Studies on the Natural α -Pyrone Antibiotic Myxopyronin from *Myxococcus Fulvus*. *ChemBioChem* **2015**, *16* (6), 946–953. <https://doi.org/10.1002/cbic.201402666>.

(5) Pinto, A.; Wang, M.; Horsman, M.; Boddy, C. N. 6-Deoxyerythronolide B Synthase Thioesterase-Catalyzed Macrocyclization Is Highly Stereoselective. *Org. Lett.* **2012**, *14* (9), 2278–2281. <https://doi.org/10.1021/ol300707j>.

(6) Hansen, D. A.; Rath, C. M.; Eisman, E. B.; Narayan, A. R. H.; Kittendorf, J. D.; Mortison, J. D.; Yoon, Y. J.; Sherman, D. H. Biocatalytic Synthesis of Pikromycin, Methymycin, Neomethymycin, Novamethymycin, and Ketomethymycin. *J. Am. Chem. Soc.* **2013**, *135* (30), 11232–11238. <https://doi.org/10.1021/ja404134f>.

(7) Franke, J.; Hertweck, C. Biomimetic Thioesters as Probes for Enzymatic Assembly Lines: Synthesis, Applications, and Challenges. *Cell Chem. Biol.* **2016**, *23* (10), 1179–1192. <https://doi.org/10.1016/j.chembiol.2016.08.014>.

(8) Hahn, F.; Kandziora, N.; Friedrich, S.; Leadlay, P. F. Synthesis of Complex Intermediates for the Study of a Dehydratase from Borrelidin Biosynthesis. *Beilstein J. Org. Chem.* **2014**, *10* (1), 634–640. <https://doi.org/10.3762/bjoc.10.55>.

(9) Berkhan, G.; Merten, C.; Holec, C.; Hahn, F. The Interplay between a Multifunctional Dehydratase Domain and a C-Methyltransferase Effects Olefin Shift in Ambruticin Biosynthesis. *Angew. Chem. Int. Ed.* **2016**, *55* (43), 13589–13592. <https://doi.org/10.1002/anie.201607827>.

(10) Schröder, M.; Roß, T.; Hemmerling, F.; Hahn, F. Studying a Bottleneck of Multimodular Polyketide Synthase Processing: The Polyketide Structure-Dependent Performance of Ketoreductase Domains. *ACS Chem. Biol.* **2022**, *17* (5), 1030–1037. <https://doi.org/10.1021/acschembio.2c00047>.

(11) Wunderlich, J.; Roß, T.; Schröder, M.; Hahn, F. Step-Economic Synthesis of Biomimetic β -Ketopolyene Thioesters and Demonstration of Their Usefulness in Enzymatic Biosynthesis Studies. *Org. Lett.* **2020**, *22* (13), 4955–4959. <https://doi.org/10.1021/acs.orglett.0c01348>.

(12) Sundaram, S.; Kim, H. J.; Bauer, R.; Thongkongkaew, T.; Heine, D.; Hertweck, C. On-Line Polyketide Cyclization into Diverse Medium-Sized Lactones by a Specialized Ketosynthase Domain. *Angew. Chem. Int. Ed.* **2018**, *57* (35), 11223–11227. <https://doi.org/10.1002/anie.201804991>.

(13) Hooshmand, S. E.; Heidari, B.; Sedghi, R.; Varma, R. S. Recent Advances in the Suzuki–Miyaura Cross-Coupling Reaction Using Efficient Catalysts in Eco-Friendly Media. *Green Chem.* **2019**, *21* (3), 381–405. <https://doi.org/10.1039/C8GC02860E>.

(14) Miyaura, Norio.; Suzuki, Akira. Palladium-Catalyzed Cross-Coupling Reactions of Organoboron Compounds. *Chem. Rev.* **1995**, *95* (7), 2457–2483. <https://doi.org/10.1021/cr00039a007>.

(15) Zeysing, B.; Gosch, C.; Terfort, A. Protecting Groups for Thiols Suitable for Suzuki Conditions. *Org. Lett.* **2000**, *2* (13), 1843–1845. <https://doi.org/10.1021/ol0058902>.

(16) Liebeskind, L. S.; Srogl, J. Thiol Ester–Boronic Acid Coupling. A Mechanistically Unprecedented and General Ketone Synthesis. *J. Am. Chem. Soc.* **2000**, *122* (45), 11260–11261. <https://doi.org/10.1021/ja005613q>.

(17) Cheng, H.-G.; Chen, H.; Liu, Y.; Zhou, Q. The Liebeskind–Srogl Cross-Coupling Reaction and Its Synthetic Applications. *Asian J. Org. Chem.* **2018**, *7* (3), 490–508. <https://doi.org/10.1002/ajoc.201700651>.

(18) Meng, S.; Tang, G.-L.; Pan, H.-X. Enzymatic Formation of Oxygen-Containing Heterocycles in Natural Product Biosynthesis. *ChemBioChem* **2018**, *19* (19), 2002–2022. <https://doi.org/10.1002/cbic.201800225>.

(19) Hemmerling, F.; Hahn, F. Biosynthesis of Oxygen and Nitrogen-Containing Heterocycles in Polyketides. *Beilstein J. Org. Chem.* **2016**, *12* (1), 1512–1550. <https://doi.org/10.3762/bjoc.12.148>.

(20) Pöplau, P.; Frank, S.; Morinaka, B. I.; Piel, J. An Enzymatic Domain for the Formation of Cyclic Ethers in Complex Polyketides. *Angew. Chem. Int. Ed.* **2013**, *52* (50), 13215–13218. <https://doi.org/10.1002/anie.201307406>.

(21) Berkhan, G.; Hahn, F. A Dehydratase Domain in Ambruticin Biosynthesis Displays Additional Activity as a Pyran-Forming Cyclase. *Angew. Chem. Int. Ed.* **2014**, *53* (51), 14240–14244. <https://doi.org/10.1002/anie.201407979>.

(22) Hollmann, T.; Berkhan, G.; Wagner, L.; Sung, K. H.; Kolb, S.; Geise, H.; Hahn, F. Biocatalysts from Biosynthetic Pathways: Enabling Stereoselective, Enzymatic Cycloether Formation on a Gram Scale. *ACS Catal.* **2020**, *10* (9), 4973–4982. <https://doi.org/10.1021/acscatal.9b05071>.

(23) Wagner, L.; Stang, J.; Derra, S.; Hollmann, T.; Hahn, F. Towards Understanding Oxygen Heterocycle-Forming Biocatalysts: A Selectivity Study of the Pyran Synthase PedPS7. *Org. Biomol. Chem.* **2022**, *20* (48), 9645–9649. <https://doi.org/10.1039/D2OB02064E>.

(24) Sung, K. H.; Berkhan, G.; Hollmann, T.; Wagner, L.; Blankenfeldt, W.; Hahn, F. Insights into the Dual Activity of a Bifunctional Dehydratase-Cyclase Domain. *Angew. Chem. Int. Ed.* **2018**, *57* (1), 343–347. <https://doi.org/10.1002/anie.201707774>.

(25) Wagner, L.; Roß, T.; Hollmann, T.; Hahn, F. Cross-Linking of a Polyketide Synthase Domain Leads to a Recyclable Biocatalyst for Chiral Oxygen Heterocycle Synthesis. *RSC Adv.* **2021**, *11* (33), 20248–20251. <https://doi.org/10.1039/D1RA03692K>.

(26) Wagner, D. T.; Zhang, Z.; Meoded, R. A.; Cepeda, A. J.; Piel, J.; Keatinge-Clay, A. T. Structural and Functional Studies of a Pyran Synthase Domain from a Trans-Acyltransferase Assembly Line. *ACS Chem. Biol.* **2018**, *13* (4), 975–983. <https://doi.org/10.1021/acschembio.8b00049>.

(27) Ueoka, R.; Uria, A. R.; Reiter, S.; Mori, T.; Karbaum, P.; Peters, E. E.; Helfrich, E. J. N.; Morinaka, B. I.; Gugger, M.; Takeyama, H.; Matsunaga, S.; Piel, J. Metabolic and Evolutionary Origin of Actin-Binding Polyketides from Diverse Organisms. *Nat. Chem. Biol.* **2015**, *11* (9), 705–712. <https://doi.org/10.1038/nchembio.1870>.

(28) Luhavaya, H.; Dias, M. V. B.; Williams, S. R.; Hong, H.; de Oliveira, L. G.; Leadlay, P. F. Enzymology of Pyran Ring A Formation in Salinomycin Biosynthesis. *Angew. Chem. Int. Ed.* **2015**, *54* (46), 13622–13625. <https://doi.org/10.1002/anie.201507090>.

(29) Woo, A. J.; Strohl, W. R.; Priestley, N. D. Nonactin Biosynthesis: The Product of NonS Catalyzes the Formation of the Furan Ring of Nonactic Acid. *Antimicrob. Agents Chemother.* **1999**, *43* (7), 1662–1668.

(30) Miyaura, N.; Ishiyama, T.; Sasaki, H.; Ishikawa, M.; Sato, M.; Suzuki, A. Palladium-Catalyzed Inter- and Intramolecular Cross-Coupling Reactions of B-Alkyl-9-Borabicyclo[3.3.1]Nonane Derivatives with 1-Halo-1-Alkenes or Haloarenes. Syntheses of Functionalized Alkenes, Arenes, and Cycloalkenes via a Hydroboration-Coupling Sequence. *J. Am. Chem. Soc.* **1989**, *111* (1), 314–321. <https://doi.org/10.1021/ja00183a048>.

(31) Farina, V.; Krishnan, B. Large Rate Accelerations in the Stille Reaction with Tri-2-Furylphosphine and Triphenylarsine as Palladium Ligands: Mechanistic and Synthetic Implications. *J. Am. Chem. Soc.* **1991**, *113* (25), 9585–9595. <https://doi.org/10.1021/ja00025a025>.

(32) Chishiro, A.; Konishi, M.; Inaba, R.; Yumura, T.; Imoto, H.; Naka, K. Tertiary Arsine Ligands for the Stille Coupling Reaction. *Dalton Trans.* **2021**, *51* (1), 95–103. <https://doi.org/10.1039/D1DT02955J>.

(33) Carreira, E. M.; Du Bois, J. (+)-Zaragozic Acid C: Synthesis and Related Studies. *J. Am. Chem. Soc.* **1995**, *117* (31), 8106–8125. <https://doi.org/10.1021/ja00136a008>.