

Supplementary Information

Evidence for the I-shaped dimers of a plant chloroplast F_0F_1 -ATP synthase in response to changes in ionic strength

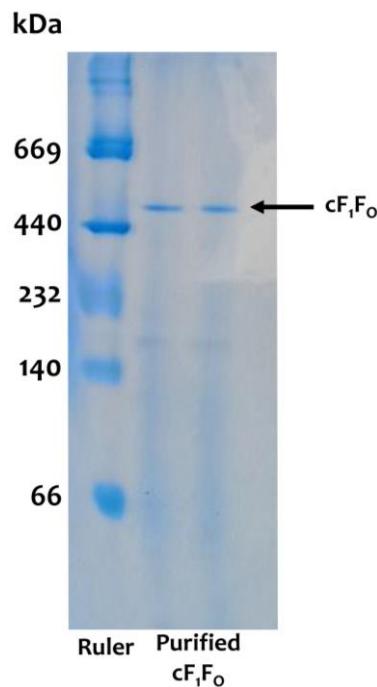


Figure S1. Characterization of the purified cF_0F_1 by BN-PAGE (6 – 18 %). MW-ruler: 66, 140, 232, 440, 669 kDa. Purified samples of cF_0F_1 showed the band between 440 and 669 kDa. The expected molecular weight of the cF_0F_1 complex is ~595 kDa.

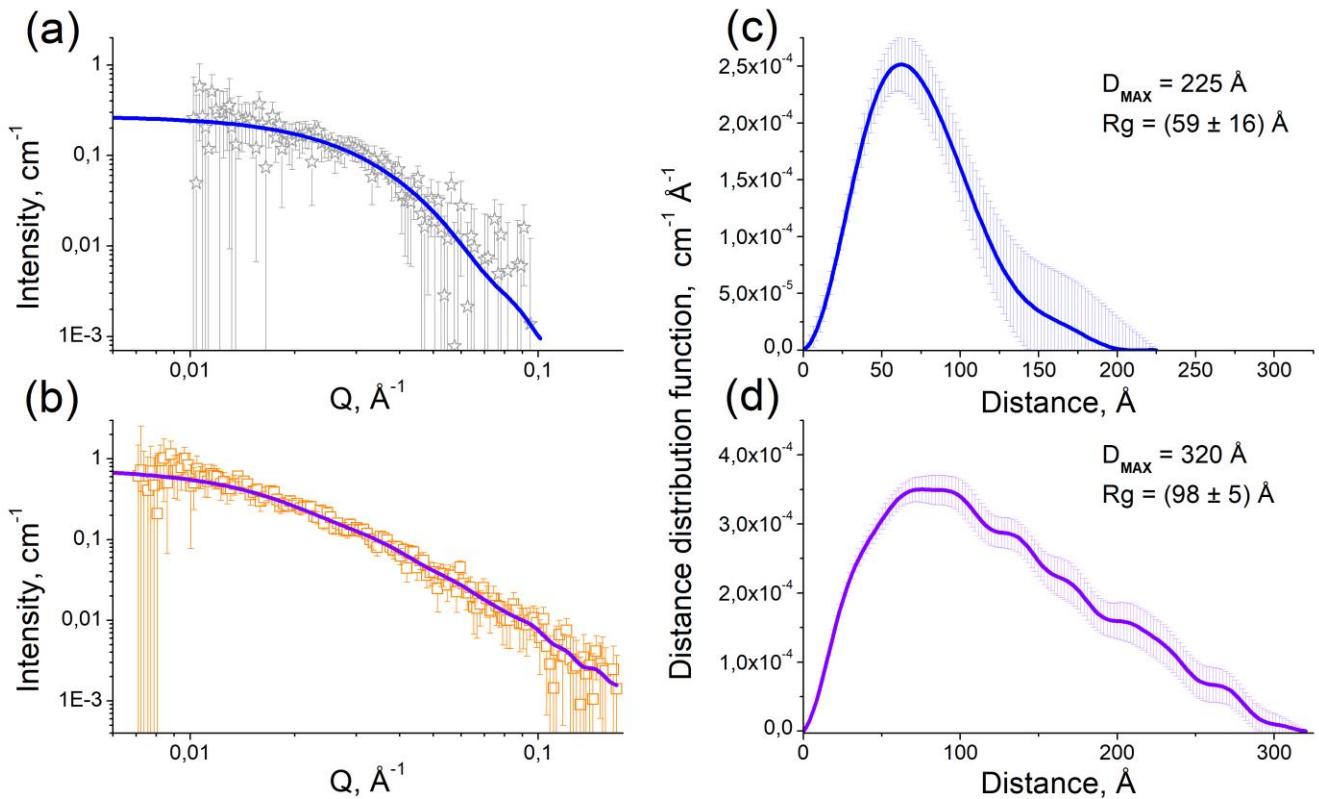


Figure S2. SANS characterization of cFoF₁: (a) SANS experimental data for AEX-purified cFoF₁ in H₂O-buffer (hollow grey stars) and a regularized fit (blue line); (b) SANS experimental data for cFoF₁ at 300 mM NaCl in 93% D₂O-buffer (hollow orange squares) and a regularized fit (purple line); (c) Pair-distance distribution function P(r) for cFoF₁ in H₂O-buffer; (d) Pair-distance distribution function P(r) for cFoF₁ at 300 mM NaCl in 93% D₂O-buffer.

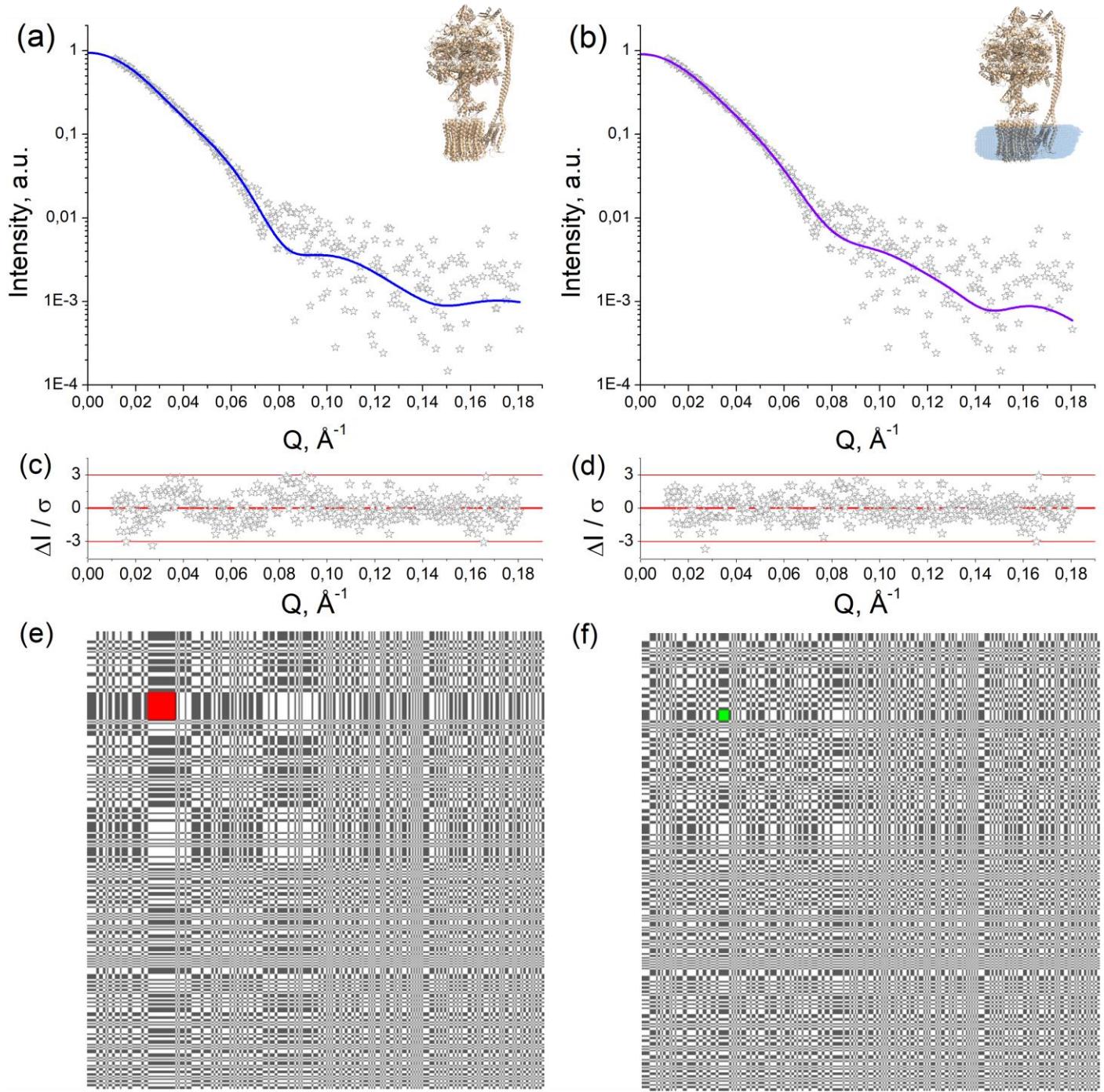


Figure S3. Influence of a detergent belt on quality of SAXS data approximation: (a) SAXS experimental data for cFoF1 (hollow grey stars) and an approximation (blue line, $\chi^2 = 1.35$) obtained by CRYSTAL for the model of cFoF1 without a detergent belt (PDB ID: 6FKF); (b) SAXS experimental data for cFoF1 the same as in panel (a) (hollow grey stars) and an approximation (blue line, $\chi^2 = 1.15$) obtained by CRYSTAL for model of protein with detergent belt built by MEMPROT; (c) Relative residues of the fit shown in panel (a); (d) Relative residues of the fit shown in panel (b); (e) Correlation map (P-value 0.000156) for the fit shown in panel (a); (f) Correlation map (P-value 0.745484) for the fit shown in panel (b).

(a)

# model	HDOCK		PDBePISA			D_{m-m} , Å	SAXS data approximation for cF ₀ F ₁					
	Docking score	Confidence score	interface area, Å ²	ΔG , kcal/mol	N_{HB}	N_{SB}	150 mM NaCl	250 mM NaCl	300 mM NaCl	350 mM NaCl	450 mM NaCl	
1	-154.82	0.5241	1910,5	-0,3	13	7	168	$\chi^2 = 1.41$ $\alpha_2 = 65 \pm 2 \%$	$\chi^2 = 1.13$ $\alpha_2 = 18 \pm 2 \%$	$\chi^2 = 1.22$ $\alpha_2 = 39 \pm 2 \%$	$\chi^2 = 1.07$ $\alpha_2 = 30 \pm 2 \%$	$\chi^2 = 1.34$ $\alpha_2 = 60 \pm 2 \%$
2	-152.11	0.5105	1046,9	1,7	6	6	159	$\chi^2 = 1.41$ $\alpha_2 = 62 \pm 2 \%$	$\chi^2 = 1.13$ $\alpha_2 = 17 \pm 2 \%$	$\chi^2 = 1.23$ $\alpha_2 = 37 \pm 2 \%$	$\chi^2 = 1.07$ $\alpha_2 = 28 \pm 2 \%$	$\chi^2 = 1.34$ $\alpha_2 = 57 \pm 2 \%$
3	-141.80	0.4591	1888,2	0,6	13	5	166	$\chi^2 = 1.42$ $\alpha_2 = 64 \pm 2 \%$	$\chi^2 = 1.13$ $\alpha_2 = 18 \pm 2 \%$	$\chi^2 = 1.22$ $\alpha_2 = 39 \pm 2 \%$	$\chi^2 = 1.08$ $\alpha_2 = 29 \pm 2 \%$	$\chi^2 = 1.35$ $\alpha_2 = 59 \pm 2 \%$
4	-139.19	0.4462	1776,5	-4,3	10	6	203	$\chi^2 = 1.29$ $\alpha_2 = 73 \pm 2 \%$	$\chi^2 = 1.11$ $\alpha_2 = 21 \pm 2 \%$	$\chi^2 = 1.25$ $\alpha_2 = 43 \pm 2 \%$	$\chi^2 = 1.00$ $\alpha_2 = 35 \pm 2 \%$	$\chi^2 = 1.20$ $\alpha_2 = 68 \pm 2 \%$
5	-139.18	0.4461	1218,3	4,8	17	4	206	$\chi^2 = 1.26$ $\alpha_2 = 77 \pm 2 \%$	$\chi^2 = 1.10$ $\alpha_2 = 23 \pm 2 \%$	$\chi^2 = 1.28$ $\alpha_2 = 44 \pm 3 \%$	$\chi^2 = 0.96$ $\alpha_2 = 38 \pm 2 \%$	$\chi^2 = 1.17$ $\alpha_2 = 71 \pm 2 \%$
6	-138.84	0.4444	1225,8	4,2	13	3	207	$\chi^2 = 1.26$ $\alpha_2 = 77 \pm 2 \%$	$\chi^2 = 1.10$ $\alpha_2 = 23 \pm 2 \%$	$\chi^2 = 1.28$ $\alpha_2 = 44 \pm 3 \%$	$\chi^2 = 0.95$ $\alpha_2 = 38 \pm 2 \%$	$\chi^2 = 1.17$ $\alpha_2 = 71 \pm 2 \%$
7	-137.62	0.4384	1347,6	2	9	8	206	$\chi^2 = 1.25$ $\alpha_2 = 76 \pm 2 \%$	$\chi^2 = 1.10$ $\alpha_2 = 23 \pm 2 \%$	$\chi^2 = 1.27$ $\alpha_2 = 44 \pm 3 \%$	$\chi^2 = 0.96$ $\alpha_2 = 37 \pm 2 \%$	$\chi^2 = 1.16$ $\alpha_2 = 71 \pm 2 \%$
8	-131.93	0.4106	1033,1	-4,1	8	0	194	$\chi^2 = 1.25$ $\alpha_2 = 75 \pm 2 \%$	$\chi^2 = 1.09$ $\alpha_2 = 23 \pm 2 \%$	$\chi^2 = 1.27$ $\alpha_2 = 44 \pm 3 \%$	$\chi^2 = 0.96$ $\alpha_2 = 37 \pm 2 \%$	$\chi^2 = 1.16$ $\alpha_2 = 70 \pm 2 \%$
9	-129.08	0.3969	1219,3	-0,8	10	6	209	$\chi^2 = 1.28$ $\alpha_2 = 75 \pm 2 \%$	$\chi^2 = 1.10$ $\alpha_2 = 22 \pm 2 \%$	$\chi^2 = 1.27$ $\alpha_2 = 44 \pm 3 \%$	$\chi^2 = 0.98$ $\alpha_2 = 36 \pm 2 \%$	$\chi^2 = 1.19$ $\alpha_2 = 70 \pm 2 \%$
10	-127.96	0.3916	1033,9	2,3	9	5	151	$\chi^2 = 1.36$ $\alpha_2 = 62 \pm 2 \%$	$\chi^2 = 1.12$ $\alpha_2 = 17 \pm 2 \%$	$\chi^2 = 1.24$ $\alpha_2 = 36 \pm 2 \%$	$\chi^2 = 1.05$ $\alpha_2 = 28 \pm 2 \%$	$\chi^2 = 1.29$ $\alpha_2 = 57 \pm 2 \%$

(b)

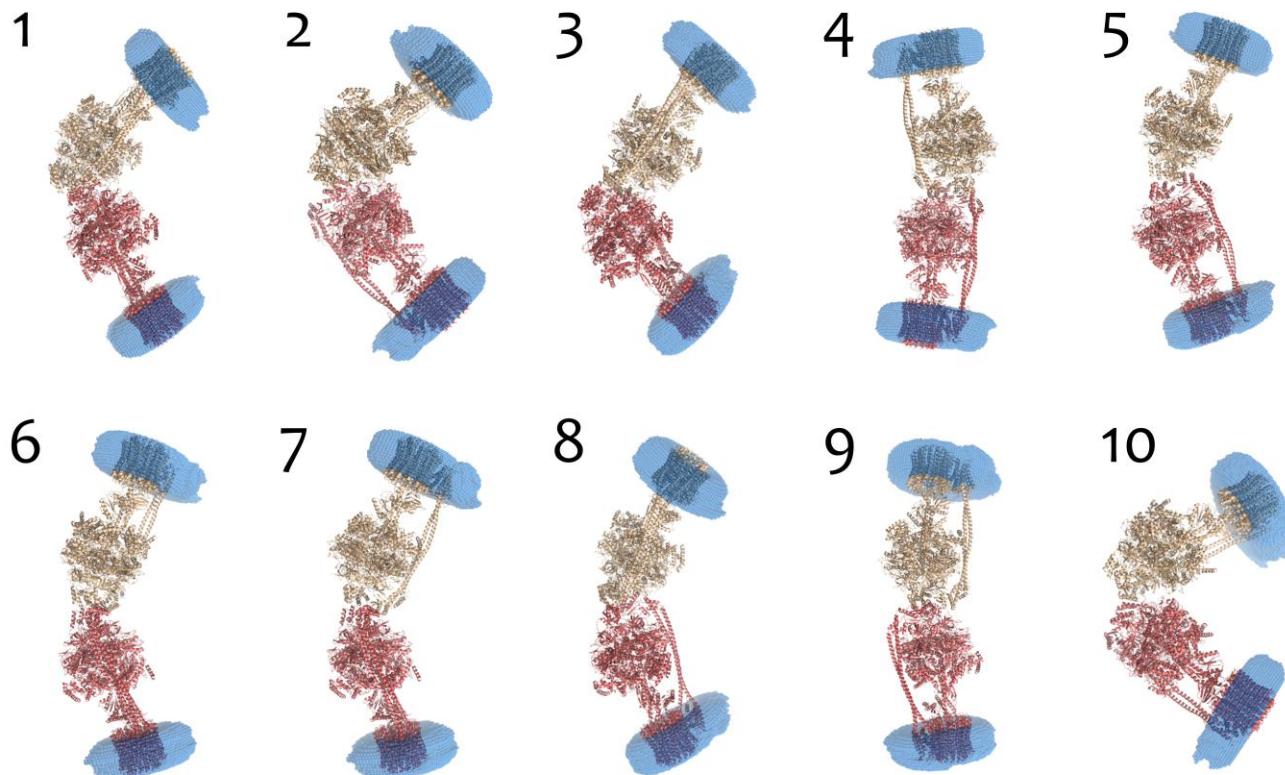


Figure S4. Models of F₁/F₁-interface dimers of cF₀F₁ based on the top 10 HDOCK predictions: (a) Table of scoring parameters and SAXS data treatment parameters obtained for the top 10 HDOCK predictions (see

section *Macromolecular docking* in Materials and Methods). Table columns are: internal HDOCK docking quality estimations (Docking score and Confidence score), parameters of macromolecular interfaces estimated using the PDBePISA web-server (interface area, Gibbs energy ΔG , numbers of hydrogen bonds N(HB) and salt bridges N(SB)), distances D_{m-m} between centers of mass of monomers in the dimers of cFoF₁, and parameters of approximations of SAXS data obtained at 150, 250, 300, 350, and 450 mM NaCl (χ^2 values and volume fractions of dimers α_2 obtained by the fit in program OLIGOMER); **(b)** Representations of cFoF₁ dimers based on the top 10 HDOCK predictions. The models contain detergent belts obtained by the program MEMPROT.

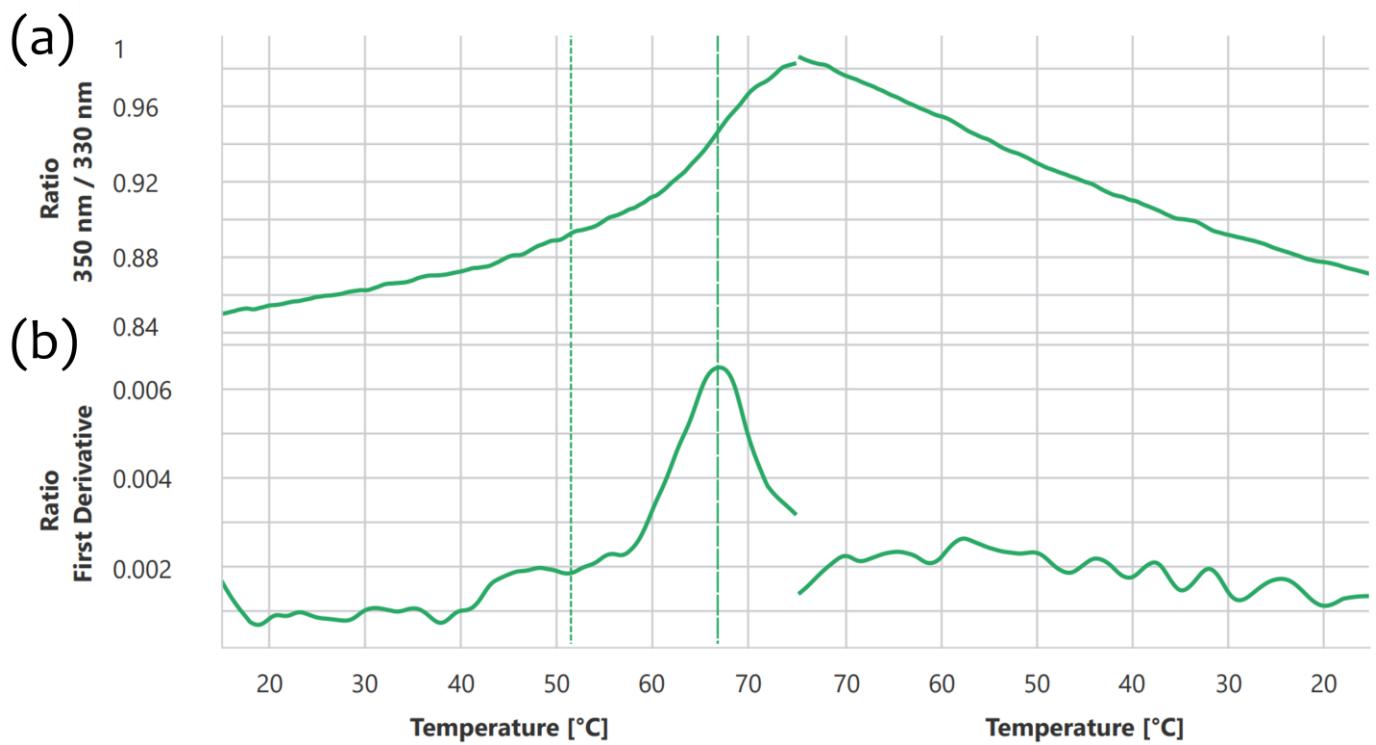


Figure S5. Thermal unfolding and refolding graphs measured by nanoDSF method. Vertical short dash line corresponds to T_{onset} , dash line shows melting temperature T_m : (a) Ratio of fluorescence intensity at 330 nm and 350 nm for temperature range from 15 °C to 75 °C; (b) First derivative of the 330 nm/350 nm fluorescence intensity ratio.

Table S1. SAXS experimental details and data evaluation summary.**(a) Sample details¹**

	ATPs AEX	ATPs 150 mM NaCl	ATPs 250 mM NaCl	ATPs 300 mM NaCl	ATPs 350 mM NaCl	ATPs 450 mM NaCl
Description of sequence	ATP synthase complex from <i>Spinacia Oleracea</i> (full structure: 6FKF), one complex contains: 3 α (UniProt ID P06450), 3 β (UniProt ID P00825), γ (UniProt ID P05435), delta (UniProt ID P11402), ϵ (UniProt ID P00833), a (UniProt ID P06451), b (UniProt ID P06453), b' (UniProt ID P31853), 14 c (UniProt ID P69447).					
ϵ_{280} (M ⁻¹ cm ⁻¹)	α : 26820, β : 17880, γ : 10430, δ : 5960, ϵ : 11000, a: 42400, b: 9970, b': 4470, c: 1490, full complex: 196 790					
Partial specific volume	0.7286 cm ³ /g					
Complex volume	722670 Å ³					
Molecular mass	597.24 kDa					
Sample concentration	5.0 mg/ml					
Solvent composition	30 mM HEPES (pH 8.0), 2 mM MgCl ₂ , 0.04% <i>tpcc</i> - α -M, ~300 mM NaCl 150 mM NaCl, 250 mM NaCl, 300 mM NaCl, 350 mM NaCl, 450 mM NaCl,					

(b) SAS data collection parameters

Instrument	Rigaku MicroMax-007 HF (MIPT, Dolgoprudny, Russia)
Wavelength	1.5406 Å
Beam geometry	Size (FWHM beam diameter at sample position): 0.3 mm; Sample-to-detector distance: 2.0 m
Sample configuration	Glass capillaries with diameters of ~1.5 mm
Q-measurement range	0.007 – 0.215 Å ⁻¹
Q-scaling method	Calibration standard: silver behenate powder
Exposure time	5400 sec for each sample
Sample temperature	20 °C

(c) Software employed for SAS data reduction, analysis and interpretation

Data averaging and subtraction, Guinier analysis	PRIMUSqt from ATSAS
Calculation of ϵ from sequence	ProtParam: https://web.expasy.org/protparam/
Calculation of volume from chemical composition	Peptide Property Calculator: http://biotools.nubic.northwestern.edu/proteincalc.html
$P(r)$ analysis	GNOM from ATSAS
Atomic structure modelling	HDOCK web-server; MEMPROT 2.2; CRYSTAL 2.0 (command line mode)
Molecular graphics	PyMOL 1.9.x

(d) Structural parameters

	ATPs AEX	ATPs 150 mM NaCl	ATPs 250 mM NaCl	ATPs 300 mM NaCl	ATPs 350 mM NaCl	ATPs 450 mM NaCl
	I(0) (a.u.)	1.03 ± 0.02	1.35 ± 0.04	1.00 ± 0.03	1.02 ± 0.03	1.39 ± 0.05
Rg (Å)	76.5 ± 2.2	104.7 ± 5.0	82.5 ± 3.7	90.3 ± 5.1	110.3 ± 7.8	129.9 ± 5.4
D _{max} (Å)	285.0	402.16	333.5	382.0	453.0	465.0
Q-range (Å ⁻¹)	0.0085 – 0.1355	0.0065 – 0.1355	0.0065 – 0.1355	0.0065 – 0.1355	0.0065 – 0.1355	0.0065 – 0.1355
Q _{min} D _{max} / π	0.77	0.83	0.69	0.79	0.93	0.96
Total quality estimate (GNOM)	0.6756	0.6556	0.6460	0.5497	0.6647	0.6468

Guinier analysis	I(0) (a.u.)	1.05 ± 0.03	1.33 ± 0.03	0.97 ± 0.02	0.97 ± 0.03	1.30 ± 0.03	1.54 ± 0.07
	Rg (Å)	75.9 ± 3.0	96.0 ± 2.9	73.7 ± 1.7	78.3 ± 3.1	88.7 ± 2.8	107.7 ± 6.8
	Q Rg – range	0.64 – 1.29	0.72 – 1.30	0.48 – 1.29	0.51 – 1.29	0.58 – 1.29	0.70 – 1.29

(e) Atomistic modelling

	ATPs AEX	ATPs 150 mM NaCl	ATPs 250 mM NaCl	ATPs 300 mM NaCl	ATPs 350 mM NaCl	ATPs 450 mM NaCl
Q-range (Å ⁻¹)	0.011 – 0.1805	0.0075 – 0.1805	0.0070 – 0.1805	0.0065 – 0.1805	0.0065 – 0.1805	0.0065 – 0.1805
Method	CRYSTAL 2.0 (command line mode)					
Any measures of model precision	Protein model (PDB ID: 6FKF) without detergent belt					
Solvation shell ΔQ	0.020 e / Å ³					
Ra	1.640 Å					
Volume	757369 Å ³					
χ ² value	1.347					
Method	MEMPROT; CRYSTAL 2.0 (command line mode)					
Any measures of model precision	Protein model (PDB ID: 6FKF) with pseudo-atomic detergent belt obtained using MEMPROT. Parameters of the detergent belt: adaptive shape algorithm type 2 (MBJP), a = 32.4 Å, b = 7.0 Å, t = 6.5 Å, e = 1.0, #mol = 517.					
Solvation shell ΔQ	0.075 e / Å ³					
Ra	1.400 Å					
Volume	980265 Å ³					
χ ² value	1.149					
Method	OLIGOMER					
Any measures of model precision	Experimental data were fitted by a set of curves calculated using CRYSTAL 2.0 (command line mode) for monomers and dimers of cFoF1. Calculations of a set of form-factors were performed for the models of monomeric and dimeric cFoF1 containing detergent belts obtained by MEMPROT using the optimal parameters of the monomeric fit obtained for the AEX-purified protein: Ra = 1.4 Å, ΔQ(shell) = 0.075 e / Å ³ , monomer volume = 980265 Å ³ .					
Volume fraction of the cFoF1 dimers		0.733 ± 0.022	0.214 ± 0.022	0.431 ± 0.024	0.350 ± 0.020	0.680 ± 0.020
χ ² value		1.29	1.11	1.25	1.00	1.20

(f) Data and model deposition IDs *

	ATPs AEX	ATPs 150 mM NaCl	ATPs 250 mM NaCl	ATPs 300 mM NaCl	ATPs 350 mM NaCl	ATPs 450 mM NaCl
	SASxxxx (draft ID: 4860)	SASxxxx (draft ID: 4861)	SASxxxx (draft ID: 4862)	SASxxxx (draft ID: 4863)	SASxxxx (draft ID: 4864)	SASxxxx (draft ID: 4865)

* These values were calculated from the protein sequence without taking into account possible ligands (ADP, ATP, etc.) or detergent molecules.